ZOONOSES MONITORING

Hungary

TRENDS AND SOURCES OF ZOONOSES AND ZOONOTIC AGENTS IN FOODSTUFFS, ANIMALS AND FEEDINGSTUFFS

including information on foodborne outbreaks, antimicrobial resistance in zoonotic and indicator bacteria and some pathogenic microbiological agents

IN 2020

PREFACE

This report is submitted to the European Commission in accordance with Article 9 of Council Directive 2003/99/EC*. The information has also been forwarded to the European Food Safety Authority (EFSA).

The report contains information on trends and sources of zoonoses and zoonotic agents in Hungary during the year 2020.

The information covers the occurrence of these diseases and agents in animals, foodstuffs and in some cases also in feedingstuffs. In addition the report includes data on antimicrobial resistance in some zoonotic agents and indicator bacteria as well as information on epidemiological investigations of foodborne outbreaks.
Complementary data on susceptible animal populations in the country is also given. The information given covers both zoonoses that are important for the public health in the whole European Union as well as zoonoses, which are relevant on the basis of the national epidemiological situation.
The report describes the monitoring systems in place and the prevention and control strategies applied in the country. For some zoonoses this monitoring is based on legal requirements laid down by the European Union legislation, while for the other zoonoses national approaches are applied.

The report presents the results of the examinations carried out in the reporting year. A national evaluation of the epidemiological situation, with special reference to trends and sources of zoonotic infections, is given. Whenever possible, the relevance of findings in foodstuffs and animals to zoonoses cases in humans is evaluated.
The information covered by this report is used in the annual European Union Summary Reports on zoonoses and antimicrobial resistance that are published each year by EFSA.

The national report contains two parts: tables summarising data reported in the Data Collection Framework and the related text forms. The text forms were sent by email as pdf files and they are incorporated at the end of the report.

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* Directive 2003/ 99/ EC of the European Parliament and of the Council of 12 December 2003 on the monitoring of zoonoses and zoonotic agents, amending Decision 90/ 424/ EEC and repealing Council Directive 92/ 117/ EEC, OJ L 325, 17.11.2003, p. 31

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ANIMAL POPULATION TABLES

Table Susceptible animal population

Animal species	Category of animals	Population animal
Cattle (bovine animals)	Cattle (bovine animals)	991,503
Leporidae	Rabbits - farmed	1,200,000
Pigs	Pigs	2,850,200
Small ruminants	Goats	33,823
	Sheep	920,682
Solipeds, domestic	Solipeds, domestic	60,000

DISEASE STATUS TABLES

Table Bovine brucellosis in countries and regions that do not receive Community co-financing for eradication programme

Region	Zoonotic agent	Number of animals serologically tested under investigations of suspect cases	Number of suspended herds under investigations of suspect cases	Number of seropositive animals under investigations of suspect cases	Number of animals positive to BST under investigations of suspect cases	Number of animals positive in microbiological testing under investigations of suspect cases	Number of herds with status officially free	Number of infected herds	Total number of animals	Number of herds tested under surveillance	Number of animals tested under surveillance	Total number of herds	Number of infected herds tested under surveillance	Number of herds tested under surveillance by bulk milk	Number of animals or pools tested under surveillance by bulk milk	Number of infected herds tested under surveillance by bulk milk	Number of notified abortions whatever cause under investigations of suspect cases	Number of isolations of Brucella abortus under investigations of suspect cases	Number of abortions due to Brucella infection under investigations of suspect cases	Number of animals tested in microbiological and/or molecular-biology testing under investigations of suspect cases
HUNGARY	Brucella	0	0	0	0	0	15,338	0	991,503	12,617	460,712	15,341	0	84	5,075	0	766	0	0	0

Table Ovine or Caprine brucellosis in countries and regions that do not receive Community co-financing for eradication programme

Region	Zoonotic agent	Number of animals serologically tested under investigations of suspect cases	Number of suspended herds under investigations of suspect cases	Number of seropositive animals under investigations of suspect cases	Number of animals positive in microbiological testing under investigations of suspect cases	Number of herds with status officially free	Number of infected herds	Total number of animals	Number of herds tested under surveillance	Number of animals tested under surveillance	Total number of herds	Number of infected herds tested under surveillance	Number of animals tested in microbiological and/or molecular-biology testing under investigations of suspect cases
HUNGARY	Brucella	0	0	0	0	9,398	0	954,505	2,484	47,146	9,398	0	0

DISEASE STATUS TABLES

Table Bovine tuberculosis in countries and regions that do not receive Community co-financing for eradication programme

Region	Zoonotic agent	Number of herds with status officially free	Number of infected herds	Total number of animals	Interval between routine tuberculin tests	Number of animals tested with tuberculin routine testing	Number of tuberculin tests carried out before the introduction into the herds	Number of animals with suspicious lesions of tuberculosis examined and submitted to histopathological and bacteriological and/or molecular-biology examinations	Number of animals detected positive in bacteriological and/or molecular-biology examination	Total number of herds
HUNGARY	Mycobacterium bovis	15,338	4	991,503	1	711,985	37,805	625	17	15,341

PREVALENCE TABLES

Table Campylobacter:CAMPYLOBACTER in food

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Meat from bovine animals - fresh - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 10272-1:2017 Campylobacter	103	0	Campylobacter	0
	Meat from bovine animals - fresh - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 10272-1:2017 Campylobacter	104	1	Campylobacter, unspecified sp.	1
	Meat from broilers (Gallus gallus) - carcase - chilled - Slaughterhouse - Not Available - food sample - neck skin - Surveillance - based on Regulation 2073 - Official, based on Regulation 2019/627 - Objective sampling	slaughter animal batch	10	Gram	N_A	ISO 10272-2:2017 Campylobacter	206	35	Campylobacter	35
	Meat from broilers (Gallus gallus) - fresh - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 10272-1:2017 Campylobacter	356	117	Campylobacter, unspecified sp.	117
	Meat from broilers (Gallus gallus) - fresh - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 10272-1:2017 Campylobacter	280	99	Campylobacter, unspecified sp.	99
	Meat from duck - fresh - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 10272-1:2017 Campylobacter	24	6	Campylobacter, unspecified sp.	6
	Meat from duck - fresh - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 10272-1:2017 Campylobacter	93	27	Campylobacter, unspecified sp.	27
	Meat from geese - fresh - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 10272-1:2017 Campylobacter	6	3	Campylobacter, unspecified sp.	3
	Meat from geese - fresh - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 10272-1:2017 Campylobacter	6	0	Campylobacter	0
	Meat from pig - fresh - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 10272-1:2017 Campylobacter	101	4	Campylobacter, unspecified sp.	4
	Meat from pig - fresh - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 10272-1:2017 Campylobacter	102	1	Campylobacter, unspecified sp.	1
	Meat from turkey - fresh - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 10272-1:2017 Campylobacter	275	76	Campylobacter, unspecified sp.	76
	Meat from turkey - fresh - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 10272-1:2017 Campylobacter	1	0	Campylobacter	0
	Milk, cows' - raw milk - intended for direct human consumption - Farm - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Millilitre	N_A	ISO 10272-1:2017 Campylobacter	92	2	Campylobacter, unspecified sp.	2

Table COXIELLA in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sampling Details	Method	Total units tested	Total units positive	N of clinical affected herds	Zoonoses	N of units positive
Not Available	Cattle (bovine animals) - Farm - Not Available - animal sample - blood - Clinical investigations - Industry sampling - Suspect sampling	animal	N_A	Complement fixation test (CFT)	2	0		Coxiella	0
	Cattle (bovine animals) - Farm - Not Available - animal sample - blood - Clinical investigations - Industry sampling - Suspect sampling	animal	N_A	Enzyme-linked immunosorbent assay (ELISA)	495	179		Coxiella burnetii	179
	Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling	animal	N_A	Immuno Histo Chemistry (IHC)	10	1		Coxiella burnetii	1
	Cattle (bovine animals) - Farm - Not Available - animal sample - placental swab - Clinical investigations - Industry sampling - Suspect sampling	animal	N_A	PCR	4	4		Coxiella burnetii	4
	Goats - Farm - Not Available - animal sample - blood - Clinical investigations - Industry sampling - Suspect sampling	animal	N_A	Enzyme-linked immunosorbent assay (ELISA)	5	0		Coxiella	0
	Goats - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling	animal	N_A	Immuno Histo Chemistry (IHC)	1	0		Coxiella	0
	Sheep - Farm - Not Available - animal sample - blood - Clinical investigations - Industry sampling - Suspect sampling	animal	N_A	Enzyme-linked immunosorbent assay (ELISA)	26	5		Coxiella burnetii	5
	Sheep - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling	animal	N_A	Immuno Histo Chemistry (IHC)	11	0		Coxiella	0

Table Cronobacter:CRONOBACTER in food

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Follow-on formulae - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 22964:2017 Cronobacter	72	0	Cronobacter	0
	Infant formula - dried - intended for infants below 6 months - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 22964:2017 Cronobacter	37	0	Cronobacter	0

Table Echinococcus:ECHINOCOCCUS in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling Details	Method	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
HUNGARY	Cattle (bovine animals) - Slaughterhouse - Not Available - Not Available - Surveillance - Official sampling - Suspect sampling	N_A	Not Available	animal	1	1	Echinococcus granulosus	1
	Foxes - Unspecified - Not Available - Not Available - Unspecified - Private sampling - Convenient sampling	N_A	Not Available	animal	2	0	Echinococcus	0
	Pigs - Slaughterhouse - Not Available - Not Available - Surveillance - Official sampling - Suspect sampling	N_A	Not Available	animal	32	1	Echinococcus granulosus	1

Table Escherichia coli:ESCHERICHIA COLI in food

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Cereals and meals - flour/meal or finely ground powder - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4)	5	0	Shiga toxin-producing Escherichia coli (STEC)	0
	Cereals and meals - flour/meal or finely ground powder - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4)	21	2	STEC other than O157 O26 O103 O111 O145	2
	Fruits - non-pre-cut - Farm - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4)	25	0	Shiga toxin-producing Escherichia coli (STEC)	0
	Meat from bovine animals - fresh - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4)	73	0	Shiga toxin-producing Escherichia coli (STEC)	0
	Meat from bovine animals - meat products - raw but intended to be eaten cooked - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4)	42	0	Shiga toxin-producing Escherichia coli (STEC)	0
	Meat from bovine animals and pig - minced meat - intended to be eaten cooked - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4)	47	1	STEC other than O157 O26 O103 O111 O145	1
	Milk, cows' - raw milk - Farm - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Millilitre	N_A	ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4)	11	0	Shiga toxin-producing Escherichia coli (STEC)	0
	Seeds, sprouted - ready-to-eat - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4)	11	0	Shiga toxin-producing Escherichia coli (STEC)	0
	Seeds, sprouted - ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4)	45	0	Shiga toxin-producing Escherichia coli (STEC)	0
	Vegetables - non-pre-cut - Farm - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4)	35	0	Shiga toxin-producing Escherichia coli (STEC)	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Vegetables - products - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4)	16	0	Shiga toxin-producing Escherichia coli (STEC)	0
	Vegetables - products - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4)	22	0	Shiga toxin-producing Escherichia coli (STEC)	0

Table FLAVIVIRUS in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Vaccination status	Sampling Details	Method	Total units tested	Total units positive	Zoonoses		N of units positive
Not Available	Birds - wild - Hospital or medical care facility - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	animal	Unknown	N_A	PCR	8	0		West Nile virus	0
	Birds - wild - Natural habitat - Not Available - Not Available - Clinical investigations - Official sampling - Suspect sampling	animal	Unknown	N_A	Immuno Histo Chemistry (IHC)	1	0		West Nile virus	0
	Solipeds, domestic - Farm - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	animal	Unknown	N_A	PCR	8	0		West Nile virus	0
	Solipeds, domestic - Farm - Not Available - Not Available - Clinical investigations - Official sampling - Suspect sampling	animal	Unknown	N_A	PCR	1	0		West Nile virus	0
Csongrád	Solipeds, domestic - Farm - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	animal	Unknown	N_A	IgM-capture ELISA (MAC-ELISA)	164	1		West Nile virus	1

Table Listeria:LISTERIA in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling Details	Method	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Cattle (bovine animals) - Farm - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	N_A	Not Available	animal	3	3	Listeria	3
	Sheep - Farm - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	N_A	Not Available	animal	6	4	Listeria	4

Table Listeria:LISTERIA in food

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Bakery products - bread - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	29	0	detection	Listeria monocytogenes	29	0
	Bakery products - bread - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	57	0	detection	Listeria monocytogenes	57	0
	Bakery products - bread - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	37	0	detection	Listeria monocytogenes	37	0
	Bakery products - cakes - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	34	0	detection	Listeria monocytogenes	34	0
	Bakery products - cakes - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	9	0	detection	Listeria monocytogenes	9	0
	Bakery products - cakes - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	34	0	detection	Listeria monocytogenes	34	0
	Bakery products - pastry - biscuits - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Bakery products - pastry - biscuits - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	2	0	detection	Listeria monocytogenes	2	0
	Cereals and meals - flakes - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	10	0	detection	Listeria monocytogenes	10	0
	Cereals and meals - flakes - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	86	0	detection	Listeria monocytogenes	86	0
	Cereals and meals - flour/meal or finely ground powder - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	22	0	detection	Listeria monocytogenes	22	0
	Cereals and meals - flour/meal or finely ground powder - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	12	0	detection	Listeria monocytogenes	12	0
	Cereals and meals - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	2	0	detection	Listeria monocytogenes	2	0
	Cereals and meals - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	6	0	detection	Listeria monocytogenes	6	0
	Cheeses made from cows' milk - curd - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	6	0	detection	Listeria monocytogenes	6	0
	Cheeses made from cows' milk - curd - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	50	0	detection	Listeria monocytogenes	50	0
	Cheeses made from cows' milk - fresh - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	9	0	detection	Listeria monocytogenes	9	0
	Cheeses made from cows' milk - fresh - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	16	0	detection	Listeria monocytogenes	16	0
	Cheeses made from cows' milk - hard - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	16	0	detection	Listeria monocytogenes	16	0
	Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	12	0	detection	Listeria monocytogenes	12	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	98	0	detection	Listeria monocytogenes	98	0
	Cheeses made from cows' milk - unspecified - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	12	0	detection	Listeria monocytogenes	12	0
	Cheeses made from cows' milk - unspecified - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	6	0	detection	Listeria monocytogenes	6	0
	Cheeses made from cows' milk - unspecified - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	4	0	detection	Listeria monocytogenes	4	0
	Cheeses made from goats' milk - unspecified - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	2	0	detection	Listeria monocytogenes	2	0
	Cheeses made from sheep's milk - fresh - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	2	0	detection	Listeria monocytogenes	2	0
	Cheeses made from sheep's milk - fresh - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	83	0	detection	Listeria monocytogenes	83	0
	Cheeses, made from mixed milk from cows, sheep and/or goats - fresh - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Cheeses, made from mixed milk from cows, sheep and/or goats - soft and semi-soft - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Chocolate - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	3	0	detection	Listeria monocytogenes	3	0
	Chocolate - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	22	0	detection	Listeria monocytogenes	22	0
	Chocolate - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	110	0	detection	Listeria monocytogenes	110	0
	Cocoa and cocoa preparations, coffee and tea - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	2	0	detection	Listeria monocytogenes	2	0
	Cocoa and cocoa preparations, coffee and tea - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	27	0	detection	Listeria monocytogenes	27	0
	Confectionery products and pastes - soft candy - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Confectionery products and pastes - soft candy - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	8	0	detection	Listeria monocytogenes	8	0
	Dairy products (excluding cheeses) - butter - made from pasteurised milk - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	12	0	detection	Listeria monocytogenes	12	0
	Dairy products (excluding cheeses) - butter - made from pasteurised milk - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	34	0	detection	Listeria monocytogenes	34	0
	Dairy products (excluding cheeses) - cream - made from pasteurised milk - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	3	0	detection	Listeria monocytogenes	3	0
	Dairy products (excluding cheeses) - cream - made from pasteurised milk - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	4	0	detection	Listeria monocytogenes	4	0
	Dairy products (excluding cheeses) - dairy desserts - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	27	0	detection	Listeria monocytogenes	27	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Dairy products (excluding cheeses) - dairy desserts - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	103	0	detection	Listeria monocytogenes	103	0
	Dairy products (excluding cheeses) - fermented dairy products - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	27	0	detection	Listeria monocytogenes	27	0
	Dairy products (excluding cheeses) - fermented dairy products - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	90	0	detection	Listeria monocytogenes	90	0
	Dairy products (excluding cheeses) - ice-cream - made from pasteurised milk - Border Control Posts - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	5	0	detection	Listeria monocytogenes	5	0
	Dairy products (excluding cheeses) - ice-cream - made from pasteurised milk - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	104	0	detection	Listeria monocytogenes	104	0
	Dairy products (excluding cheeses) - ice-cream - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	98	0	detection	Listeria monocytogenes	98	0
	Dairy products (excluding cheeses) - ice-cream - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	157	0	detection	Listeria monocytogenes	157	0
	Dairy products (excluding cheeses) - milk powder and whey powder - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	10	0	detection	Listeria monocytogenes	10	0
	Dairy products (excluding cheeses) - milk powder and whey powder - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	49	0	detection	Listeria monocytogenes	49	0
	Dairy products (excluding cheeses) - milk-based drinks - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	64	0	detection	Listeria monocytogenes	64	0
	Dairy products (excluding cheeses) - milk-based drinks - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	35	0	detection	Listeria monocytogenes	35	0
	Fish - canned - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Fish - canned - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	85	0	detection	Listeria monocytogenes	85	0
	Fish - marinated - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	26	0	detection	Listeria monocytogenes	26	0
	Fish - smoked - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	28	0	detection	Listeria monocytogenes	28	0
	Fishery products, unspecified - cooked - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	20	2	<=100	Listeria monocytogenes	20	2
								>100	Listeria monocytogenes	20	0
	Fishery products, unspecified - cooked - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	20	2	detection	Listeria monocytogenes	20	2
	Fishery products, unspecified - non-ready-to-eat - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	20	0	detection	Listeria monocytogenes	20	0
	Fishery products, unspecified - raw - frozen - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	14	0	detection	Listeria monocytogenes	14	0
	Fishery products, unspecified - raw - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	5	0	detection	Listeria monocytogenes	5	0
	Follow-on formulae - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	26	0	detection	Listeria monocytogenes	26	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Foodstuffs intended for special nutritional uses - processed cereal-based food for infants and young children - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	47	0	detection	Listeria monocytogenes	47	0
	Foodstuffs intended for special nutritional uses - ready-to-eat meal for infants and young children - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	7	0	detection	Listeria monocytogenes	7	0
	Foodstuffs intended for special nutritional uses - ready-to-eat meal for infants and young children - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	4	0	detection	Listeria monocytogenes	4	0
	Fruits - edible part - Farm - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	26	0	detection	Listeria monocytogenes	26	0
	Fruits - edible part - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	43	0	detection	Listeria monocytogenes	43	0
	Fruits - pre-cut - frozen - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Fruits - pre-cut - frozen - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	2	0	detection	Listeria monocytogenes	2	0
	Fruits - pre-cut - frozen - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	42	0	detection	Listeria monocytogenes	42	0
	Fruits - products - canned - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Fruits - products - canned - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	5	0	detection	Listeria monocytogenes	5	0
	Fruits - products - dried - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Fruits - products - dried - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	7	0	detection	Listeria monocytogenes	7	0
	Meat from bovine animals - meat products - cooked, ready-to-eat - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	6	0	detection	Listeria monocytogenes	6	0
	Meat from bovine animals - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Meat from bovine animals - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	10	0	detection	Listeria monocytogenes	10	0
	Meat from bovine animals - meat products - fermented sausages - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Meat from bovine animals - meat products - raw and intended to be eaten raw - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Meat from bovine animals - meat products - raw but intended to be eaten cooked - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	4	0	detection	Listeria monocytogenes	4	0
	Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	9	0	detection	Listeria monocytogenes	9	0
	Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	46	0	detection	Listeria monocytogenes	46	0
	Meat from duck - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	5	0	detection	Listeria monocytogenes	5	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Meat from geese - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	5	0	detection	Listeria monocytogenes	5	0
	Meat from horse - meat products - fermented sausages - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Meat from pig - meat products - cooked, ready-to-eat - Border Control Posts - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	5	0	detection	Listeria monocytogenes	5	0
	Meat from pig - meat products - cooked, ready-to-eat - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	18	0	detection	Listeria monocytogenes	18	0
	Meat from pig - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	42	1	<=100	Listeria monocytogenes	42	1
								>100	Listeria monocytogenes	42	0
	Meat from pig - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	42	1	detection	Listeria monocytogenes	42	1
	Meat from pig - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	67	0	detection	Listeria monocytogenes	67	0
	Meat from pig - meat products - fermented sausages - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	60	5	<=100	Listeria monocytogenes	60	5
								>100	Listeria monocytogenes	60	0
	Meat from pig - meat products - fermented sausages - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	60	5	detection	Listeria monocytogenes	60	5
	Meat from pig - meat products - fermented sausages - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	156	2	<=100	Listeria monocytogenes	156	2
								>100	Listeria monocytogenes	156	0
	Meat from pig - meat products - fermented sausages - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	156	2	detection	Listeria monocytogenes	156	2
	Meat from pig - meat products - raw and intended to be eaten raw - Border Control Posts - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	2	0	detection	Listeria monocytogenes	2	0
	Meat from pig - meat products - raw and intended to be eaten raw - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	42	0	detection	Listeria monocytogenes	42	0
	Meat from pig - meat products - raw and intended to be eaten raw - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	29	0	detection	Listeria monocytogenes	29	0
	Meat from poultry, unspecified - meat products - cooked, ready-to-eat - frozen - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	3	0	detection	Listeria monocytogenes	3	0
	Meat from poultry, unspecified - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	17	0	detection	Listeria monocytogenes	17	0
	Meat from poultry, unspecified - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	86	0	detection	Listeria monocytogenes	86	0
	Meat from turkey - meat products - cooked, ready-to-eat - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	5	0	detection	Listeria monocytogenes	5	0
	Meat from turkey - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	12	1	<=100	Listeria monocytogenes	12	1
								>100	Listeria monocytogenes	12	0
	Meat from turkey - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	12	1	detection	Listeria monocytogenes	12	1
	Meat from turkey - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	72	0	detection	Listeria monocytogenes	72	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Meat from turkey - meat products - raw and intended to be eaten raw - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Meat from turkey - meat products - raw and intended to be eaten raw - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	2	0	detection	Listeria monocytogenes	2	0
	Meat from wild game - land mammals - meat products - fermented sausages - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Meat, mixed meat - meat products - fermented sausages - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Meat, mixed meat - meat products - fermented sausages - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	3	0	detection	Listeria monocytogenes	3	0
	Meat, mixed meat - meat products - raw but intended to be eaten cooked - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	3	0	detection	Listeria monocytogenes	3	0
	Meat, mixed meat - meat products - ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Milk, cows' - raw milk - intended for direct human consumption - Farm - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	108	4	<=100	Listeria monocytogenes	108	4
								>100	Listeria monocytogenes	108	0
	Milk, cows' - raw milk - intended for direct human consumption - Farm - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	108	4	detection	Listeria monocytogenes	108	4
	Milk, cows' - raw milk - intended for direct human consumption - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	9	0	detection	Listeria monocytogenes	9	0
	Milk, cows' - raw milk - intended for direct human consumption - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Milk, cows' - UHT milk - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	8	0	detection	Listeria monocytogenes	8	0
	Milk, cows' - UHT milk - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	27	0	detection	Listeria monocytogenes	27	0
	Milk, goats' - raw milk - intended for direct human consumption - Farm - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	3	0	detection	Listeria monocytogenes	3	0
	Nuts and nut products - dried - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	6	0	detection	Listeria monocytogenes	6	0
	Nuts and nut products - dried - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	50	0	detection	Listeria monocytogenes	50	0
	Other processed food products and prepared dishes - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	157	0	detection	Listeria monocytogenes	157	0
	Other processed food products and prepared dishes - fish and seafood based dishes - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Other processed food products and prepared dishes - fish and seafood based dishes - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Other processed food products and prepared dishes - fish and seafood based dishes - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Other processed food products and prepared dishes - meat based dishes - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	33	0	detection	Listeria monocytogenes	33	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Other processed food products and prepared dishes - meat based dishes - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	2	0	detection	Listeria monocytogenes	2	0
	Other processed food products and prepared dishes - meat based dishes - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	64	0	detection	Listeria monocytogenes	64	0
	Other processed food products and prepared dishes - pasta based dishes - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	30	0	detection	Listeria monocytogenes	30	0
	Other processed food products and prepared dishes - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	2	0	detection	Listeria monocytogenes	2	0
	Other processed food products and prepared dishes - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	9	0	detection	Listeria monocytogenes	9	0
	Other processed food products and prepared dishes - sandwiches - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	45	0	detection	Listeria monocytogenes	45	0
	Other processed food products and prepared dishes - sandwiches - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	3	0	detection	Listeria monocytogenes	3	0
	Other processed food products and prepared dishes - sandwiches - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	65	1	<=100	Listeria monocytogenes	65	1
								>100	Listeria monocytogenes	65	0
	Other processed food products and prepared dishes - sandwiches - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	65	1	detection	Listeria monocytogenes	65	1
	Other processed food products and prepared dishes - sushi - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	11	0	detection	Listeria monocytogenes	11	0
	Other processed food products and prepared dishes - sushi - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	10	0	detection	Listeria monocytogenes	10	0
	Other processed food products and prepared dishes - unspecified - ready-to-eat foods - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	27	1	<=100	Listeria monocytogenes	27	1
								>100	Listeria monocytogenes	27	0
	Other processed food products and prepared dishes - unspecified - ready-to-eat foods - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	27	1	detection	Listeria monocytogenes	27	1
	Ready-to-eat salads - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	49	0	detection	Listeria monocytogenes	49	0
	Ready-to-eat salads - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	2	1	<=100	Listeria monocytogenes	2	1
								>100	Listeria monocytogenes	2	0
	Ready-to-eat salads - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	2	1	detection	Listeria monocytogenes	2	1
	Ready-to-eat salads - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	128	1	<=100	Listeria monocytogenes	128	1
								>100	Listeria monocytogenes	128	0
	Ready-to-eat salads - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	128	1	detection	Listeria monocytogenes	128	1
	Seeds, sprouted - ready-to-eat - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	12	1	<=100	Listeria monocytogenes	12	1
								>100	Listeria monocytogenes	12	0
	Seeds, sprouted - ready-to-eat - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	12	1	detection	Listeria monocytogenes	12	1
	Seeds, sprouted - ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	44	0	detection	Listeria monocytogenes	44	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Soups - ready-to-eat - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	74	0	detection	Listeria monocytogenes	74	0
	Soups - ready-to-eat - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Soups - ready-to-eat - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	3	0	detection	Listeria monocytogenes	3	0
	Spices and herbs - dried - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	5	0	detection	Listeria monocytogenes	5	0
	Spices and herbs - dried - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	62	0	detection	Listeria monocytogenes	62	0
	Vegetables - non-pre-cut - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	1	0	detection	Listeria monocytogenes	1	0
	Vegetables - non-pre-cut - Farm - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	36	0	detection	Listeria monocytogenes	36	0
	Vegetables - non-pre-cut - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	8	0	detection	Listeria monocytogenes	8	0
	Vegetables - non-pre-cut - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	36	0	detection	Listeria monocytogenes	36	0
	Vegetables - pre-cut - frozen vegetables - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	12	2	<=100	Listeria monocytogenes	12	2
								>100	Listeria monocytogenes	12	0
	Vegetables - pre-cut - frozen vegetables - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	12	2	detection	Listeria monocytogenes	12	2
	Vegetables - pre-cut - frozen vegetables - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	98	16	<=100	Listeria monocytogenes	98	16
								>100	Listeria monocytogenes	98	0
	Vegetables - pre-cut - frozen vegetables - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	98	16	detection	Listeria monocytogenes	98	16
	Vegetables - pre-cut - ready-to-eat - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	33	7	<=100	Listeria monocytogenes	33	7
								>100	Listeria monocytogenes	33	0
	Vegetables - pre-cut - ready-to-eat - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	33	7	detection	Listeria monocytogenes	33	7
	Vegetables - pre-cut - ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	25	8	<=100	Listeria monocytogenes	25	8
								>100	Listeria monocytogenes	25	0
	Vegetables - pre-cut - ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	25	8	detection	Listeria monocytogenes	25	8

Table Lyssavirus:LYSSAVIRUS in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling Details	Method	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Badgers - Natural habitat - Not Available - Not Available - Monitoring - passive - Official sampling - Suspect sampling	N_A	Indirect Immunofluorescent Antibody test (IFAT)	animal	12	0	Rabies virus	0
	Bats - wild - Natural habitat - Not Available - Not Available - Monitoring - passive - Official sampling - Suspect sampling	N_A	Indirect Immunofluorescent Antibody test (IFAT)	animal	8	0	Lyssavirus	0
	Cats - Veterinary clinics - Not Available - Not Available - Monitoring - passive - Official sampling - Suspect sampling	N_A	Indirect Immunofluorescent Antibody test (IFAT)	animal	317	0	Rabies virus	0
	Cattle (bovine animals) - Farm - Not Available - Not Available - Monitoring - passive - Official sampling - Suspect sampling	N_A	Indirect Immunofluorescent Antibody test (IFAT)	animal	54	0	Rabies virus	0
	Deer - wild - roe deer - Natural habitat - Not Available - Not Available - Monitoring - passive - Official sampling - Suspect sampling	N_A	Indirect Immunofluorescent Antibody test (IFAT)	animal	19	0	Rabies virus	0
	Dogs - Veterinary clinics - Not Available - Not Available - Monitoring - passive - Official sampling - Suspect sampling	N_A	Indirect Immunofluorescent Antibody test (IFAT)	animal	128	0	Rabies virus	0
	Foxes - wild - Hunting - Not Available - Not Available - Monitoring - active - Official sampling - Objective sampling	N_A	Indirect Immunofluorescent Antibody test (IFAT)	animal	1645	0	Rabies virus	0
	Foxes - wild - Natural habitat - Not Available - Not Available - Monitoring - passive - Official sampling - Suspect sampling	N_A	Indirect Immunofluorescent Antibody test (IFAT)	animal	244	0	Rabies virus	0
	Jackals - wild - Hunting - Not Available - Not Available - Monitoring - active - Official sampling - Objective sampling	N_A	Indirect Immunofluorescent Antibody test (IFAT)	animal	143	0	Rabies virus	0
	Jackals - wild - Natural habitat - Not Available - Not Available - Monitoring - passive - Official sampling - Suspect sampling	N_A	Indirect Immunofluorescent Antibody test (IFAT)	animal	5	0	Rabies virus	0
	Sheep - Farm - Not Available - Not Available - Monitoring - passive - Official sampling - Suspect sampling	N_A	Indirect Immunofluorescent Antibody test (IFAT)	animal	28	0	Rabies virus	0
	Solipeds, domestic - Farm - Not Available - Not Available - Monitoring - passive - Official sampling - Suspect sampling	N_A	Indirect Immunofluorescent Antibody test (IFAT)	animal	6	0	Rabies virus	0
	Wild boars - Natural habitat - Not Available - Not Available - Monitoring - passive - Official sampling - Suspect sampling	N_A	Indirect Immunofluorescent Antibody test (IFAT)	animal	9	0	Rabies virus	0
	Wolves - Natural habitat - Not Available - Not Available - Monitoring - passive - Official sampling - Suspect sampling	N_A	Indirect Immunofluorescent Antibody test (IFAT)	animal	1	0	Rabies virus	0

Table Mycobacterium:MYCOBACTERIUM in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling Details	Method	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Badgers - wild - Natural habitat - Not Available - Not Available - Clinical investigations - Official sampling - Selective sampling	N_A	Not Available	animal	1	0	Mycobacterium	0
	Badgers - wild - Natural habitat - Not Available - Not Available - Monitoring - Official sampling - Selective sampling	N_A	Not Available	animal	1	0	Mycobacterium	0
	Cattle (bovine animals) - Slaughterhouse - Not Available - Not Available - Clinical investigations - Official sampling - Suspect sampling	N_A	Not Available	animal	608	115	Mycobacterium avium	5
							Mycobacterium avium subsp. paratuberculosis	18
							Mycobacterium spp., unspecified	92
	Cattle (bovine animals) - Slaughterhouse - Not Available - Not Available - Monitoring - Official sampling - Selective sampling	N_A	Not Available	animal	99	18	Mycobacterium avium subsp. paratuberculosis	5
							Mycobacterium spp., unspecified	13
	Deer - wild - red deer - Hunting - Not Available - Not Available - Monitoring - Official sampling - Selective sampling	N_A	Not Available	animal	14	5	Mycobacterium	2
							Mycobacterium avium	1
							Mycobacterium caprae	2
	Deer - wild - roe deer - Hunting - Not Available - Not Available - Monitoring - Official sampling - Selective sampling	N_A	Not Available	animal	1	0	Mycobacterium	0
	Foxes - wild - Natural habitat - Not Available - Not Available - Clinical investigations - Official sampling - Suspect sampling	N_A	Not Available	animal	3	0	Mycobacterium	0
	Giraffes - zoo animal - Zoo - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	N_A	Not Available	animal	1	1	Mycobacterium intracellulare	1
	Pigs - Slaughterhouse - Not Available - Not Available - Clinical investigations - Official sampling - Suspect sampling	N_A	Not Available	animal	23	9	Mycobacterium avium	8
							Mycobacterium avium subsp. avium	1
	Wild boars - wild - Hunting - Not Available - Not Available - Monitoring - Official sampling - Selective sampling	N_A	Not Available	animal	237	95	Mycobacterium avium	1
							Mycobacterium avium subsp. avium	2
							Mycobacterium caprae	37
							Mycobacterium spp., unspecified	55

Table Salmonella:SALMONELLA in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	N of flocks under control programme	Target verification	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Birds - zoo animal - Farm - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	animal		N_A	N_A	Not Available	1	0	Salmonella	0
	Cattle (bovine animals) - Farm - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	animal		N_A	N_A	Not Available	50	0	Salmonella	0
	Ducks - Farm - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	herd/flock		N_A	N_A	Not Available	286	7	Salmonella	7
	Gallus gallus (fowl) - breeding flocks, unspecified - adult - Farm - Not Available - Not Available - Control and eradication programmes - Official and industry sampling - Census	herd/flock	727	Y	N_A	Not Available	727	18	Salmonella Agona	1
									Salmonella Anatum	1
									Salmonella Bredeney	2
									Salmonella Enteritidis	2
									Salmonella Infantis	1
									Salmonella Newport	2
									Salmonella Stanley	9
	Gallus gallus (fowl) - broilers - before slaughter - Farm - Not Available - Not Available - Control and eradication programmes - Official and industry sampling - Census	herd/flock	7655	Y	N_A	Not Available	7655	35	Salmonella Enteritidis	2
									Salmonella Kedougou	17
									Salmonella Kottbus	1
									Salmonella Other serovars	8
									Salmonella Senftenberg	1
									Salmonella Szentes	1
									Salmonella Tennessee	1
									Salmonella Typhimurium	2
									Salmonella Typhimurium, monophasic	2
	Gallus gallus (fowl) - laying hens - adult - Farm - Not Available - Not Available - Control and eradication programmes - Official and industry sampling - Census	herd/flock	860	Y	N_A	Not Available	860	27	Salmonella Agona	1
									Salmonella Bovismorbificans	1
									Salmonella Chester	2
									Salmonella Enteritidis	10
									Salmonella Kedougou	4
									Salmonella Kentucky	3
									Salmonella Lille	1
									Salmonella Other serovars	1
									Salmonella Rissen	2
									Salmonella Typhimurium, monophasic	2
	Geese - Farm - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	herd/flock		N_A	N_A	Not Available	81	34	Salmonella	34
	Monkeys - Zoo - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	animal		N_A	N_A	Not Available	3	0	Salmonella	0
	Pheasants - Farm - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	herd/flock		N_A	N_A	Not Available	48	0	Salmonella	0
	Pigeons - Farm - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	herd/flock		N_A	N_A	Not Available	3	0	Salmonella	0
	Pigs - Farm - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	herd/flock		N_A	N_A	Not Available	1	0	Salmonella	0
	Quails - laying hens - Farm - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	herd/flock		N_A	N_A	Not Available	1	0	Salmonella	0
	Turkeys - breeding flocks, unspecified - adult - Farm - Not Available - Not Available - Control and eradication programmes - Official and industry sampling - Census	herd/flock	156	Y	N_A	Not Available	156	2	Salmonella Hadar	2
	Turkeys - fattening flocks - before slaughter - Farm - Not Available - Not Available - Control and eradication programmes - Official and industry sampling - Census	herd/flock	2170	Y	N_A	Not Available	2170	313	Salmonella Agona	16
									Salmonella Anatum	2
									Salmonella Bredeney	139
									Salmonella Enteritidis	4
									Salmonella Hadar	34
									Salmonella Infantis	65

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	N of flocks under control programme	Target verification	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Turkeys - fattening flocks - before slaughter - Farm - Not Available - Not Available - Control and eradication programmes - Official and industry sampling - Census	herd/flock	2170	Y	N_A	Not Available	2170	313	Salmonella Kentucky	15
									Salmonella Newport	15
									Salmonella Other serovars	8
									Salmonella Tennessee	1
									Salmonella Typhimurium	14

Table Salmonella:SALMONELLA in food

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Bakery products - bread - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	30	0	Salmonella	0
	Bakery products - bread - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	71	0	Salmonella	0
	Bakery products - bread - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	87	0	Salmonella	0
	Bakery products - cakes - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	37	0	Salmonella	0
	Bakery products - cakes - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	12	0	Salmonella	0
	Bakery products - cakes - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	36	0	Salmonella	0
	Bakery products - pastry - biscuits - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	4	0	Salmonella	0
	Bakery products - pastry - biscuits - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	5	0	Salmonella	0
	Beverages, non-alcoholic - soft drinks - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	101	0	Salmonella	0
	Cereals and meals - flakes - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	9	0	Salmonella	0
	Cereals and meals - flakes - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	87	0	Salmonella	0
	Cereals and meals - flour/meal or finely ground powder - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	50	1	Salmonella enterica, subspecies salamae	1
	Cereals and meals - flour/meal or finely ground powder - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	50	0	Salmonella	0
	Cereals and meals - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	7	0	Salmonella	0
	Cheeses made from cows' milk - curd - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	0	Salmonella	0
	Cheeses made from cows' milk - curd - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	50	0	Salmonella	0
	Cheeses made from cows' milk - fresh - made from pasteurised milk - Farm - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Cheeses made from cows' milk - fresh - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	8	0	Salmonella	0
	Cheeses made from cows' milk - fresh - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	16	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Cheeses made from cows' milk - hard - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	13	0	Salmonella	0
	Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Farm - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	11	0	Salmonella	0
	Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	60	0	Salmonella	0
	Cheeses made from cows' milk - unspecified - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	11	0	Salmonella	0
	Cheeses made from cows' milk - unspecified - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Cheeses made from cows' milk - unspecified - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	4	0	Salmonella	0
	Cheeses made from goats' milk - Farm - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Cheeses made from goats' milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Cheeses made from sheep's milk - fresh - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	0	Salmonella	0
	Cheeses made from sheep's milk - fresh - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	85	0	Salmonella	0
	Cheeses, made from mixed milk from cows, sheep and/or goats - fresh - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Cheeses, made from mixed milk from cows, sheep and/or goats - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Chocolate - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	0	Salmonella	0
	Chocolate - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	23	0	Salmonella	0
	Chocolate - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	113	0	Salmonella	0
	Cocoa and cocoa preparations, coffee and tea - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Cocoa and cocoa preparations, coffee and tea - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	19	0	Salmonella	0
	Cocoa and cocoa preparations, coffee and tea - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	81	0	Salmonella	0
	Coconut - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Coconut - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	79	0	Salmonella	0
	Confectionery products and pastes - soft candy - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	0	Salmonella	0
	Confectionery products and pastes - soft candy - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	8	0	Salmonella	0
	Crustaceans - unspecified - raw - Border Control Posts - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	5	0	Salmonella	0
	Dairy products (excluding cheeses) - butter - made from pasteurised milk - Farm - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Dairy products (excluding cheeses) - cream - made from pasteurised milk - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Dairy products (excluding cheeses) - dairy desserts - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	39	0	Salmonella	0
	Dairy products (excluding cheeses) - dairy desserts - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	158	0	Salmonella	0
	Dairy products (excluding cheeses) - fermented dairy products - Farm - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Dairy products (excluding cheeses) - fermented dairy products - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	10	0	Salmonella	0
	Dairy products (excluding cheeses) - fermented dairy products - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	43	0	Salmonella	0
	Dairy products (excluding cheeses) - ice-cream - made from pasteurised milk - Border Control Posts - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	6	0	Salmonella	0
	Dairy products (excluding cheeses) - ice-cream - made from pasteurised milk - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	116	0	Salmonella	0
	Dairy products (excluding cheeses) - ice-cream - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	98	0	Salmonella	0
	Dairy products (excluding cheeses) - ice-cream - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	157	0	Salmonella	0
	Dairy products (excluding cheeses) - milk powder and whey powder - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	10	0	Salmonella	0
	Dairy products (excluding cheeses) - milk powder and whey powder - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	50	0	Salmonella	0
	Dairy products (excluding cheeses) - milk-based drinks - Farm - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	0	Salmonella	0
	Dairy products (excluding cheeses) - milk-based drinks - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Dairy products (excluding cheeses) - milk-based drinks - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	5	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Egg products - dried - Border Control Posts - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	8	0	Salmonella	0
	Egg products - dried - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Egg products - dried - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	24	0	Salmonella	0
	Egg products - dried - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Egg products - liquid - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	0	Salmonella	0
	Egg products - liquid - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	30	1	Salmonella Thompson	1
	Egg products - liquid - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	4	0	Salmonella	0
	Eggs - table eggs - shell - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Eggs - table eggs - shell - Packing centre - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	18	0	Salmonella	0
	Eggs - table eggs - shell - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	176	0	Salmonella	0
	Eggs - table eggs - yolk - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Eggs - table eggs - yolk - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	29	0	Salmonella	0
	Eggs - table eggs - yolk - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	316	0	Salmonella	0
	Fish - canned - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Fish - canned - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	88	0	Salmonella	0
	Fish - marinated - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	26	0	Salmonella	0
	Fish - raw - Border Control Posts - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	8	0	Salmonella	0
	Fish - raw - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	25	0	Salmonella	0
	Fish - raw - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	97	0	Salmonella	0
	Fish - smoked - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	29	0	Salmonella	0
	Fishery products, unspecified - cooked - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	19	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Fishery products, unspecified - non-ready-to-eat - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Fishery products, unspecified - non-ready-to-eat - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	21	0	Salmonella	0
	Fishery products, unspecified - raw - frozen - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	15	0	Salmonella	0
	Fishery products, unspecified - raw - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	6	0	Salmonella	0
	Follow-on formulae - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	89	0	Salmonella	0
	Foodstuffs intended for special nutritional uses - processed cereal-based food for infants and young children - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	71	0	Salmonella	0
	Foodstuffs intended for special nutritional uses - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	0	Salmonella	0
	Foodstuffs intended for special nutritional uses - ready-to-eat meal for infants and young children - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	6	0	Salmonella	0
	Foodstuffs intended for special nutritional uses - ready-to-eat meal for infants and young children - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	4	0	Salmonella	0
	Foodstuffs intended for special nutritional uses - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	11	0	Salmonella	0
	Fruits - edible part - Farm - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	26	1	Salmonella Abony	1
	Fruits - edible part - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	46	0	Salmonella	0
	Fruits - pre-cut - frozen - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Fruits - pre-cut - frozen - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Fruits - pre-cut - frozen - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	32	0	Salmonella	0
	Fruits - products - canned - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Fruits - products - canned - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	7	0	Salmonella	0
	Fruits - products - dried - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Fruits - products - dried - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	8	0	Salmonella	0
	Juice - vegetable juice - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Meat from bovine animals - carcase - Processing plant - Not Available - food sample - carcase swabs - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	400	Square centimetre	N_A	ISO 6579-1:2017 Salmonella	120	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Meat from bovine animals - fresh - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	104	1	Salmonella Mbandaka	1
	Meat from bovine animals - fresh - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	105	0	Salmonella	0
	Meat from bovine animals - meat preparation - intended to be eaten cooked - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Meat from bovine animals - meat products - cooked, ready-to-eat - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	6	0	Salmonella	0
	Meat from bovine animals - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat from bovine animals - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	9	0	Salmonella	0
	Meat from bovine animals - meat products - fermented sausages - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat from bovine animals - meat products - raw and intended to be eaten raw - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat from bovine animals - meat products - raw but intended to be eaten cooked - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	0	Salmonella	0
	Meat from bovine animals - meat products - raw but intended to be eaten cooked - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	46	1	Salmonella Typhimurium	1
	Meat from bovine animals - minced meat - intended to be eaten cooked - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	ISO 6579-1:2017 Salmonella	21	0	Salmonella	0
	Meat from bovine animals - minced meat - intended to be eaten cooked - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	ISO 6579-1:2017 Salmonella	74	0	Salmonella	0
	Meat from broilers (Gallus gallus) - carcase - Processing plant - Not Available - food sample - neck skin - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	312	34	Salmonella Hadar	5
									Salmonella Infantis	27
									Salmonella Kentucky	2
	Meat from broilers (Gallus gallus) - fresh - Border Control Posts - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	51	4	Salmonella Infantis	4
	Meat from broilers (Gallus gallus) - fresh - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat from broilers (Gallus gallus) - fresh - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	276	39	Salmonella Agona	1
									Salmonella Enteritidis	1
									Salmonella group O:7	1
									Salmonella Infantis	34
									Salmonella Kentucky	2
	Meat from broilers (Gallus gallus) - fresh - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	165	33	Salmonella Agona	1
									Salmonella Enteritidis	1
									Salmonella Infantis	28
									Salmonella Kentucky	2
									Salmonella Thompson	1
	Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	1	Salmonella Infantis	1

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	29	6	Salmonella Infantis	6
	Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	12	0	Salmonella	0
	Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	50	0	Salmonella	0
	Meat from duck - fresh - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	22	2	Salmonella Typhimurium	2
	Meat from duck - fresh - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	94	16	Salmonella Enteritidis	1
									Salmonella Hadar	8
									Salmonella Indiana	1
									Salmonella Infantis	1
									Salmonella Kottbus	2
									Salmonella Typhimurium	3
	Meat from duck - meat preparation - intended to be eaten cooked - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat from duck - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat from duck - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	5	0	Salmonella	0
	Meat from duck - meat products - non-ready-to-eat - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	7	0	Salmonella	0
	Meat from duck - meat products - non-ready-to-eat - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	1	Salmonella Hadar	1
	Meat from duck - meat products - raw but intended to be eaten cooked - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Meat from geese - carcase - Processing plant - Not Available - food sample - carcase swabs - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Meat from geese - fresh - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	6	0	Salmonella	0
	Meat from geese - fresh - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	6	0	Salmonella	0
	Meat from geese - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	5	0	Salmonella	0
	Meat from horse - meat products - fermented sausages - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Meat from pig - carcase - Processing plant - Not Available - food sample - carcase swabs - Surveillance - based on Regulation 2073 - Official, based on Regulation 2019/627 - Objective sampling	single (food/feed)	400	Square centimetre	N_A	ISO 6579-1:2017 Salmonella	2131	13	Salmonella Derby	6
									Salmonella Enteritidis	1
									Salmonella group O:7	2
									Salmonella Newport	1
									Salmonella Typhimurium	1
									Salmonella Typhimurium, monophasic	2

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Meat from pig - fresh - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	101	2	Salmonella Derby	2
	Meat from pig - fresh - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	124	2	Salmonella Typhimurium	1
									Salmonella Typhimurium, monophasic	1
	Meat from pig - meat preparation - intended to be eaten cooked - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Meat from pig - meat preparation - intended to be eaten cooked - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	ISO 6579-1:2017 Salmonella	43	1	Salmonella Derby	1
	Meat from pig - meat preparation - intended to be eaten cooked - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	ISO 6579-1:2017 Salmonella	82	1	Salmonella Give	1
	Meat from pig - meat products - cooked, ready-to-eat - Border Control Posts - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	5	0	Salmonella	0
	Meat from pig - meat products - cooked, ready-to-eat - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	37	0	Salmonella	0
	Meat from pig - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	63	0	Salmonella	0
	Meat from pig - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	82	0	Salmonella	0
	Meat from pig - meat products - fermented sausages - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat from pig - meat products - fermented sausages - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	96	0	Salmonella	0
	Meat from pig - meat products - fermented sausages - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	166	1	Salmonella Derby	1
	Meat from pig - meat products - raw and intended to be eaten raw - Border Control Posts - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Meat from pig - meat products - raw and intended to be eaten raw - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	49	0	Salmonella	0
	Meat from pig - meat products - raw and intended to be eaten raw - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	31	0	Salmonella	0
	Meat from pig - meat products - raw but intended to be eaten cooked - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Meat from pig - meat products - raw but intended to be eaten cooked - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	0	Salmonella	0
	Meat from pig - minced meat - intended to be eaten cooked - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	ISO 6579-1:2017 Salmonella	52	3	Salmonella Derby	1
									Salmonella group O:7	1
									Salmonella Saintpaul	1
	Meat from pig - minced meat - intended to be eaten cooked - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	ISO 6579-1:2017 Salmonella	170	7	Salmonella Derby	2
									Salmonella Typhimurium	1
									Salmonella Typhimurium, monophasic	4

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Meat from poultry, unspecified - fresh - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Meat from poultry, unspecified - meat preparation - intended to be eaten cooked - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat from poultry, unspecified - meat products - cooked, ready-to-eat - frozen - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat from poultry, unspecified - meat products - cooked, ready-to-eat - frozen - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	4	0	Salmonella	0
	Meat from poultry, unspecified - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	18	0	Salmonella	0
	Meat from poultry, unspecified - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	89	0	Salmonella	0
	Meat from poultry, unspecified - meat products - raw but intended to be eaten cooked - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat from poultry, unspecified - meat products - raw but intended to be eaten cooked - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	7	0	Salmonella	0
	Meat from rabbit - fresh - Processing plant - Not Available - food sample - blood - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	9	0	Salmonella	0
	Meat from rabbit - fresh - Retail - Not Available - food sample - blood - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat from turkey - carcase - Processing plant - Not Available - food sample - neck skin - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	257	20	Salmonella Anatum	1
									Salmonella Bredeney	3
									Salmonella Hadar	4
									Salmonella Infantis	2
									Salmonella Kentucky	7
									Salmonella Newport	1
									Salmonella Senftenberg	2
	Meat from turkey - fresh - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	107	4	Salmonella Infantis	1
									Salmonella Kentucky	1
									Salmonella Newport	1
									Salmonella Senftenberg	1
	Meat from turkey - fresh - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	84	9	Salmonella Bredeney	2
									Salmonella Kentucky	3
									Salmonella Newport	2
									Salmonella Stanley	1
									Salmonella Thompson	1
	Meat from turkey - meat preparation - intended to be eaten cooked - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat from turkey - meat preparation - intended to be eaten cooked - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	7	1	Salmonella Hadar	1
	Meat from turkey - meat products - cooked, ready-to-eat - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	6	0	Salmonella	0
	Meat from turkey - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	14	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Meat from turkey - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	72	0	Salmonella	0
	Meat from turkey - meat products - raw and intended to be eaten raw - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat from turkey - meat products - raw and intended to be eaten raw - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	6	0	Salmonella	0
	Meat from turkey - meat products - raw but intended to be eaten cooked - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat from turkey - meat products - raw but intended to be eaten cooked - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	5	0	Salmonella	0
	Meat from turkey - minced meat - intended to be eaten cooked - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	7	0	Salmonella	0
	Meat from turkey - minced meat - intended to be eaten cooked - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	42	6	Salmonella Agona	1
									Salmonella Bredeney	2
									Salmonella Infantis	1
									Salmonella Kentucky	1
									Salmonella Typhimurium, monophasic	1
	Meat from wild game - birds - fresh - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat from wild game - birds - fresh - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	11	1	Salmonella Typhimurium	1
	Meat from wild game - birds - fresh - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	71	4	Salmonella Infantis	3
									Salmonella Thompson	1
	Meat from wild game - land mammals - fresh - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat from wild game - land mammals - fresh - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	37	1	Salmonella Coeln	1
	Meat from wild game - land mammals - fresh - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	74	2	Salmonella Enteritidis	2
	Meat from wild game - land mammals - meat products - fermented sausages - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat from wild game - land mammals - meat products - fermented sausages - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	4	0	Salmonella	0
	Meat from wild game - land mammals - meat products - raw but intended to be eaten cooked - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat, mixed meat - meat products - fermented sausages - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat, mixed meat - meat products - fermented sausages - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	4	0	Salmonella	0
	Meat, mixed meat - meat products - raw but intended to be eaten cooked - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Meat, mixed meat - meat products - raw but intended to be eaten cooked - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	0	Salmonella	0
	Meat, mixed meat - meat products - ready-to-eat - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Meat, mixed meat - minced meat - intended to be eaten cooked - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	10	Gram	N_A	ISO 6579-1:2017 Salmonella	4	0	Salmonella	0
	Meat, red meat (meat from bovines, pigs, goats, sheep, horses, donkeys, bison and water buffalos) - fresh - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Meat, red meat (meat from bovines, pigs, goats, sheep, horses, donkeys, bison and water buffalos) - meat products - stabilized products, unspecified - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Meat, red meat (meat from bovines, pigs, goats, sheep, horses, donkeys, bison and water buffalos) - meat products - stabilized products, unspecified - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	25	0	Salmonella	0
	Milk, cows' - raw milk - Farm - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	92	0	Salmonella	0
	Milk, cows' - raw milk - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Milk, cows' - raw milk - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Milk, cows' - UHT milk - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	19	0	Salmonella	0
	Milk, goats' - raw milk - Farm - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	0	Salmonella	0
	Nuts and nut products - dried - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Nuts and nut products - dried - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	11	0	Salmonella	0
	Nuts and nut products - dried - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	135	0	Salmonella	0
	Other processed food products and prepared dishes - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	165	0	Salmonella	0
	Other processed food products and prepared dishes - chips, crisps, fries and dough-based analogues - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	7	0	Salmonella	0
	Other processed food products and prepared dishes - fish and seafood based dishes - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Other processed food products and prepared dishes - fish and seafood based dishes - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Other processed food products and prepared dishes - fish and seafood based dishes - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Other processed food products and prepared dishes - meat based dishes - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	36	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Other processed food products and prepared dishes - meat based dishes - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	0	Salmonella	0
	Other processed food products and prepared dishes - meat based dishes - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	64	0	Salmonella	0
	Other processed food products and prepared dishes - pasta - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Other processed food products and prepared dishes - pasta - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	9	0	Salmonella	0
	Other processed food products and prepared dishes - pasta - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	213	2	Salmonella Enteritidis	1
									Salmonella Mbandaka	1
	Other processed food products and prepared dishes - pasta based dishes - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	30	0	Salmonella	0
	Other processed food products and prepared dishes - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	0	Salmonella	0
	Other processed food products and prepared dishes - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	6	0	Salmonella	0
	Other processed food products and prepared dishes - sandwiches - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	43	0	Salmonella	0
	Other processed food products and prepared dishes - sandwiches - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	0	Salmonella	0
	Other processed food products and prepared dishes - sandwiches - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	65	0	Salmonella	0
	Other processed food products and prepared dishes - sushi - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Other processed food products and prepared dishes - sushi - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Other processed food products and prepared dishes - unspecified - ready-to-eat foods - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	29	0	Salmonella	0
	Other products of animal origin - gelatin and collagen - Border Control Posts - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	15	0	Salmonella	0
	Other products of animal origin - gelatin and collagen - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	39	0	Salmonella	0
	Ready-to-eat salads - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	51	0	Salmonella	0
	Ready-to-eat salads - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Ready-to-eat salads - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	128	0	Salmonella	0
	Seeds, sprouted - ready-to-eat - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	12	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Seeds, sprouted - ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	45	0	Salmonella	0
	Soups - dehydrated - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Soups - dehydrated - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	56	0	Salmonella	0
	Soups - ready-to-eat - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	82	0	Salmonella	0
	Soups - ready-to-eat - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Soups - ready-to-eat - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	0	Salmonella	0
	Spices and herbs - Border Control Posts - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Spices and herbs - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	25	0	Salmonella	0
	Spices and herbs - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	183	0	Salmonella	0
	Vegetables - non-pre-cut - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	0	Salmonella	0
	Vegetables - non-pre-cut - Farm - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	35	0	Salmonella	0
	Vegetables - non-pre-cut - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	11	0	Salmonella	0
	Vegetables - non-pre-cut - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	57	0	Salmonella	0
	Vegetables - pre-cut - frozen vegetables - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	11	0	Salmonella	0
	Vegetables - pre-cut - frozen vegetables - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	28	0	Salmonella	0
	Vegetables - pre-cut - non-ready-to-eat - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Vegetables - pre-cut - non-ready-to-eat - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Vegetables - pre-cut - ready-to-eat - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	33	0	Salmonella	0
	Vegetables - pre-cut - ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	25	0	Salmonella	0
	Vegetables - products - Catering - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Vegetables - products - dried - Processing plant - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	5	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Vegetables - products - dried - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	1	0	Salmonella	0
	Vegetables - products - Retail - Not Available - food sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	14	0	Salmonella	0

Table Salmonella:SALMONELLA in feed

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Sampling Details	Method	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Compound feedingstuffs for cattle - Feed mill - Not Available - feed sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	5	0	Salmonella	0
	Compound feedingstuffs for pigs - final product - Feed mill - Not Available - feed sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	6	0	Salmonella	0
	Compound feedingstuffs for poultry (non specified) - final product - Feed mill - Not Available - feed sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	21	0	Salmonella	0
	Compound feedingstuffs for poultry, broilers - final product - Feed mill - Not Available - feed sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	25	0	Salmonella	0
	Compound feedingstuffs for poultry, laying hens - final product - Feed mill - Not Available - feed sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	14	0	Salmonella	0
	Feed material of cereal grain origin - maize derived - Feed mill - Not Available - feed sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Feed material of cereal grain origin - wheat derived - Feed mill - Not Available - feed sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	0	Salmonella	0
	Feed material of land animal origin - meat meal - Feed mill - Not Available - feed sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	6	0	Salmonella	0
	Feed material of marine animal origin - fish meal - Feed mill - Not Available - feed sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	2	0	Salmonella	0
	Feed material of oil seed or fruit origin - rape seed derived - Feed mill - Not Available - feed sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	3	0	Salmonella	0
	Feed material of oil seed or fruit origin - soya (bean) derived - Feed mill - Not Available - feed sample - Monitoring - Official sampling - Objective sampling	single (food/feed)	25	Gram	N_A	ISO 6579-1:2017 Salmonella	24	6	Salmonella	6
			26	Gram	N_A	ISO 6579-1:2017 Salmonella	6	6	Salmonella Senftenberg	6

Table Toxoplasma:TOXOPLASMA in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling Details	Method	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Sheep - Farm - Not Available - Not Available - Clinical investigations - Industry sampling - Convenient sampling	N_A	Immuno Histo Chemistry (IHC)	animal	1	0	Toxoplasma	0
	Sheep - Farm - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	N_A	Complement fixation test (CFT)	animal	4	0	Toxoplasma	0
	Sheep - Farm - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	N_A	Immuno Histo Chemistry (IHC)	animal	1	0	Toxoplasma	0

Table Trichinella:TRICHINELLA in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling Details	Method	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Foxes - Hunting - Not Available - animal sample - organ/tissue - Monitoring - Official sampling - Objective sampling	N_A	Not Available	animal	273	0	Trichinella	0
	Foxes - Unspecified - Not Available - animal sample - organ/tissue - Unspecified - Private sampling - Convenient sampling	N_A	Not Available	animal	1	0	Trichinella	0
	Jackals - Hunting - Not Available - animal sample - organ/tissue - Monitoring - Official sampling - Objective sampling	N_A	Not Available	animal	3	0	Trichinella	0
	Pigs - breeding animals - not raised under controlled housing conditions - Slaughterhouse - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Census	N_A	Not Available	animal	1061660	0	Trichinella	0
	Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Census	N_A	Not Available	animal	3640066	0	Trichinella	0
	Rats - Natural habitat - Not Available - animal sample - organ/tissue - Unspecified - Private sampling - Convenient sampling	N_A	Not Available	animal	3	0	Trichinella	0
	Solipeds, domestic - Slaughterhouse - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Census	N_A	Not Available	animal	221	0	Trichinella	0
	Wild boars - Hunting - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Census	N_A	Not Available	animal	46295	3	Trichinella spiralis	2
							Trichinella, unspecified sp.	1

Table Yersinia:YERSINIA in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling Details	Method	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	All animals - zoo animals - Zoo - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	N_A	Not Available	animal	1	1	Yersinia pseudotuberculosis	1
	Birds - wild - Conservation facilities - Not Available - Not Available - Clinical investigations - Industry sampling - Convenient sampling	N_A	Not Available	animal	1	1	Yersinia	1
	Cattle (bovine animals) - Farm - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	N_A	Not Available	animal	1	1	Yersinia pseudotuberculosis	1
	Pigs - Farm - Not Available - Not Available - Clinical investigations - Industry sampling - Suspect sampling	N_A	Not Available	animal	5	5	Yersinia	5

FOODBORNE OUTBREAKS TABLES

Foodborne Outbreaks: summarized data

when numbers referring to cases, hospitalized people and deaths are reported as unknown, they will be not included in the sum calculation

		Outbreak strenght	Strong				Weak			
Causative agent	Food vehicle		N outbreaks	N human cases	N hospitalized	N deaths	N outbreaks	N human cases	N hospitalized	N deaths
Bacillus cereus	Mixed food						1	10	0	0
Microorganisms	Pig meat and products thereof						1	29	0	0
	Broiler meat (Gallus gallus) and products thereof		1	15	5	0				
	Other foods						1	14	0	0
Salmonella Enteritidis	Pig meat and products thereof		1	21	6	0				
	Sweets and chocolate		2	78	9	0				
Unknown	Sweets and chocolate						1	43	0	0
	Other foods						1	15	0	0
	Mixed food						2	52	1	0

Strong Foodborne Outbreaks: detailed data

Causative agent	H	AG	VT	Other Causative Agent	FBO nat. code	Outbreak type	Food vehicle	More food vehicle info	Nature of evidence	Setting	Place of origin of problem	Origin of food vehicle	Contributory factors	Comment	N outbreaks	N human cases	N hosp.	N deaths
Microorganisms	unk	Not Available	Not Available	Not Available	Étbi_6	General	Broiler meat (Gallus gallus) and products thereof	N_A	Descriptive epidemiological evidence	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Hungary	Inadequate heat treatment	N_A	1	15	5	0
Salmonella Enteritidis	unk	Not Available	Not Available	Not Available	Étbi_5	General	Sweets and chocolate	N_A	Detection of causative agent in food vehicle or its component - Detection of indistinguishable causative agent in humans	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Hungary	Inadequate heat treatment	N_A	1	72	7	0
					Étbi_7	General	Pig meat and products thereof	N_A	Detection of causative agent in food vehicle or its component - Detection of indistinguishable causative agent in humans	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Hungary	Storage time/temperature abuse	N_A	1	21	6	0
					Étbi_8	General	Sweets and chocolate	N_A	Detection of causative agent in food vehicle or its component - Detection of indistinguishable causative agent in humans	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Hungary	Cross-contamination	N_A	1	6	2	0

Weak Foodborne Outbreaks: detailed data

Causative agent	H	AG	VT	Other Causative Agent	FBO nat. code	Outbreak type	Food vehicle	More food vehicle info	Nature of evidence	Setting	Place of origin of problem	Origin of food vehicle	Contributory factors	Comment	N outbreaks	N human cases	N hosp.	N deaths
Bacillus cereus	unk	Not Available	Not Available	Not Available	Étbi_2	General	Mixed food	N_A	Unknown	School or kindergarten	School or kindergarten	Hungary	Unknown	N_A	1	10	0	0
Microorganisms	unk	Not Available	Not Available	Not Available	Étbi_11	General	Pig meat and products thereof	N_A	Unknown	School or kindergarten	School or kindergarten	Hungary	Unknown	N_A	1	29	0	0
					Étbi_9	General	Other foods	N_A	Unknown	Canteen or workplace catering	Canteen or workplace catering	Hungary	Unknown	N_A	1	14	0	0
Unknown	unk	Not Available	Not Available	Not Available	Étbi_1	General	Mixed food	N_A	Unknown	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Hungary	Unknown	N_A	1	38	1	0
					Étbi_10	General	Other foods	N_A	Unknown	Residential institution (nursing home or prison or boarding school)	Residential institution (nursing home or prison or boarding school)	Hungary	Storage time/temperature abuse	N_A	1	15	0	0
					Étbi_3	General	Sweets and chocolate	N_A	Unknown	School or kindergarten	Retail	Hungary	Unknown	N_A	1	43	0	0
					Étbi_4	General	Mixed food	N_A	Unknown	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Hungary	Unknown	N_A	1	14	0	0

ANTIMICROBIAL RESISTANCE TABLES FOR CAMPYLOBACTER

Table Antimicrobial susceptibility testing of Campylobacter jejuni in Gallus gallus (fowl) - broilers

Sampling Stage: Slaughterhouse | Sampling Type: animal sample - caecum | Sampling Context: Monitoring

Sampler: Official sampling | Sampling Strategy: Objective sampling | Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling details:

	AM substance	Ciprofloxacin	Erythromycin	Gentamicin	Nalidixic acid	Streptomycin	Tetracycline
	ECOFF	**0.5**	**4**	**2**	**16**	**4**	**1**
	Lowest limit	**0.12**	**1**	**0.12**	**1**	**0.25**	**0.5**
	Highest limit	**16**	**128**	**16**	**64**	**16**	**64**
	N of tested isolates	**170**	**170**	**170**	**170**	**170**	**170**
MIC	**N of resistant isolates**	**156**	**0**	**0**	**156**	**23**	**102**
<=0.125		14		9			
<=0.25						2	
0.25				46			
<=0.5							67
0.5				98		5	
<=1			170				
1				17		58	1
2					1	80	
4		5			13	2	
8		115					2
16		31					
>16		5				23	
32					3		14
64					23		43
>64					130		43

Table Antimicrobial susceptibility testing of Campylobacter jejuni in Turkeys - fattening flocks

Sampling Stage: Slaughterhouse | Sampling Type: animal sample - caecum | Sampling Context: Monitoring

Sampler: Official sampling | Sampling Strategy: Objective sampling | Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling details:

	AM substance	Ciprofloxacin	Erythromycin	Gentamicin	Nalidixic acid	Streptomycin	Tetracycline
	ECOFF	**0.5**	**4**	**2**	**16**	**4**	**1**
	Lowest limit	**0.12**	**1**	**0.12**	**1**	**0.25**	**0.5**
	Highest limit	**16**	**128**	**16**	**64**	**16**	**64**
	N of tested isolates	**170**	**170**	**170**	**170**	**170**	**170**
MIC	**N of resistant isolates**	**150**	**0**	**0**	**149**	**16**	**98**
<=0.125		20		10			
0.25				70			
<=0.5							72
0.5				88		8	
<=1			169				
1				2		84	
2		1	1		7	61	1
4		11			11	1	
8		110			2		1
16		20			1		7
>16		8				16	
32					4		23
64					26		36
>64					119		30

ANTIMICROBIAL RESISTANCE TABLES FOR SALMONELLA

Table Antimicrobial susceptibility testing of Salmonella Abony in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1													
2								1							
<=4											1				
4													1		
<=8						1									
8			1												
16												1			

Table Antimicrobial susceptibility testing of Salmonella Agona in Meat from turkey - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							1								
<=0.03										2					
0.03							1								
<=0.25				2										2	2
<=0.5					2				2						
<=1		2													
<=2													2		
2								2							
<=4											2				
<=8						2									
8			2												
16												2			

Table Antimicrobial susceptibility testing of Salmonella Agona in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**
MIC	**N of resistant isolates**	**1**	**0**	**0**	**0**	**1**	**2**	**0**	**0**	**0**	**2**	**1**	**2**	**0**	**0**
<=0.015							3								
<=0.03										9					
0.03							4								
<=0.25				9										2	8
0.25							2								
<=0.5					9				9						
0.5														7	1
<=1		7						4							
<=2													7		
2		1						5							
<=4											7				
<=8						8						2			
8			9												
16												6			
>64		1											2		
>128						1					2				
>1024												1			

Table Antimicrobial susceptibility testing of Salmonella Anatum in Meat from turkey - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1													
<=2													1		
2								1							
<=4											1				
4			1												
<=8						1						1			

Table Antimicrobial susceptibility testing of Salmonella Anatum in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**
MIC	**N of resistant isolates**	**2**	**0**	**0**	**0**	**0**	**2**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.03										2					
<=0.25				2										2	2
<=0.5					1				2						
0.5							2								
<=1								2							
1					1										
<=2													2		
<=8						2									
8			2								2				
16												2			
>64		2													

Table Antimicrobial susceptibility testing of Salmonella Anatum in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**1**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**1**	**0**	**1**	**1**	**0**
<=0.03										1					
<=0.25				1											1
<=0.5					1				1						
0.5							1								
2								1						1	
<=8												1			
16			1			1									
>64		1											1		
>128											1				

Table Antimicrobial susceptibility testing of Salmonella Bovismorbificans in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: environmental sample - dust

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON pnl2

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	**Not Available**	**Not Available**	**Negative/Absent**	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Not Available**
	Ceftazidime synergy test	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Negative/Absent**	**Not Available**	**Not Available**	**Not Available**	**Not Available**
	ECOFF	**0.125**	**0.5**	**0.5**	**8**	**2**	**2**	**0.06**	**0.5**	**0.125**	**32**
	Lowest limit	**0.064**	**0.25**	**0.064**	**0.5**	**0.25**	**0.125**	**0.015**	**0.12**	**0.03**	**0.5**
	Highest limit	**32**	**64**	**64**	**64**	**128**	**128**	**2**	**16**	**16**	**64**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**1**	**1**	**1**	**1**	**1**	**1**	**0**	**0**	**0**	**0**
0.03								1			
0.064										1	
0.25									1		
0.5		1									
8											1
16			1	1		1	1				
64					1						

Table Antimicrobial susceptibility testing of Salmonella Bovismorbificans in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: environmental sample - dust

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**1**	**0**	**1**	**1**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.03										1					
0.03							1								
<=0.25														1	1
<=0.5									1						
<=2													1		
2								1							
<=4											1				
4			1												
>4				1											
<=8						1									
>8					1										
64												1			
>64		1													

Table Antimicrobial susceptibility testing of Salmonella Braenderup in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	1	0	0	0	0	0	0	0	0
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
0.5							1								
<=1		1						1							
<=2													1		
<=8						1						1			
8			1								1				

Table Antimicrobial susceptibility testing of Salmonella Bredeney in Meat from turkey - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**12**	**12**	**12**	**12**	**12**	**12**	**12**	**12**	**12**	**12**	**12**	**12**	**12**	**12**
MIC	**N of resistant isolates**	**11**	**1**	**0**	**0**	**1**	**12**	**0**	**1**	**0**	**12**	**2**	**12**	**5**	**1**
<=0.03										12					
0.12							5								
<=0.25				12										1	10
0.25							1								
<=0.5					11				9						
0.5							5							6	1
<=1		1						1							
1					1				2						
2								11						5	
4			6												
<=8						6						10			
8			1				1		1						
16			4			5									
32			1												
>32															1
64													3		
>64		11											9		
128						1					6				
>128											6				
>1024												2			

Table Antimicrobial susceptibility testing of Salmonella Bredeney in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**50**	**50**	**50**	**50**	**50**	**50**	**50**	**50**	**50**	**50**	**50**	**50**	**50**	**50**
MIC	**N of resistant isolates**	**48**	**4**	**0**	**0**	**0**	**50**	**0**	**0**	**0**	**50**	**4**	**48**	**30**	**15**
<=0.03										50					
0.12							10								
<=0.25				48											34
0.25							6								
<=0.5					48				48						
0.5				2			34							15	1
<=1		1						2							
1					2				2					5	
2		1						48						30	
4			14										2		
<=8						17						45			
8			3												
16			29			33						1			
32		5	4												
>32															15
64													3		
>64		43											45		
128											9				
>128											41				
>1024												4			

Table Antimicrobial susceptibility testing of Salmonella Bredeney in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**
MIC	**N of resistant isolates**	**2**	**0**	**0**	**0**	**0**	**2**	**0**	**0**	**0**	**2**	**0**	**2**	**1**	**0**
<=0.03										2					
<=0.25				1											2
<=0.5					2				2						
0.5				1			2								
1														1	
2								2						1	
<=8												2			
16			2			2									
>64	2												2		
>128											2				

Table Antimicrobial susceptibility testing of Salmonella Bredeney in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**1**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**1**	**0**	**1**	**0**	**0**
<=0.03										1					
0.12							1								
<=0.25				1											
<=0.5					1				1						
0.5														1	1
<=1								1							
4			1												
<=8						1									
16												1			
>64		1											1		
>128											1				

Table Antimicrobial susceptibility testing of Salmonella Chester in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.03										2					
0.03							2								
<=0.25				2										2	2
<=0.5					2				2						
<=1								1							
<=2													2		
2		2						1							
<=4											2				
<=8						2						1			
8			2												
16												1			

Table Antimicrobial susceptibility testing of Salmonella Chester in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.03										1					
0.03							1								
<=0.25				1										1	1
<=0.5					1				1						
<=1		1													
<=2													1		
2								1							
<=4											1				
<=8						1									
16			1									1			

Table Antimicrobial susceptibility testing of Salmonella Coeln in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							1								
<=0.03										2					
0.03							1								
<=0.25				2										2	2
<=0.5					2				2						
<=1		2													
<=2													2		
2								2							
<=4											2				
4			2												
<=8						2									
32												2			

Table Antimicrobial susceptibility testing of Salmonella Coeln in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.03										1					
0.03							1								
<=0.25				1										1	1
<=0.5					1				1						
<=1		1													
<=2													1		
2								1							
<=4											1				
4			1												
<=8						1									
16												1			

Table Antimicrobial susceptibility testing of Salmonella enterica, subspecies salamae in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1								1							
<=2													1		
<=4											1				
4		1													
<=8						1									
8			1												
16												1			

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=2													1		
2		1													
<=4											1				
4								1							
<=8						1									
8			1												
32												1			

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							2								
<=0.03										4					
0.03							2								
<=0.25				4										4	3
<=0.5					4				4						
0.5															1
<=1		3						1							
<=2													4		
2		1						3							
<=4											4				
4			3												
<=8						4						2			
8			1												
16												2			

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**1**	**0**	**0**	**1**	**0**	**0**	**0**	**0**
<=0.015							2								
<=0.03										3					
0.12							1								
<=0.25				3										3	3
<=0.5					3				3						
<=1		2						1							
<=2													3		
2		1						1							
<=4											2				
4			2												
<=8						3									
8			1					1							
16												2			
32												1			
>128											1				

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**
MIC	**N of resistant isolates**	**1**	**0**	**0**	**0**	**0**	**1**	**1**	**0**	**0**	**1**	**0**	**0**	**0**	**0**
<=0.03										4					
0.03							3								
<=0.25				4										3	3
<=0.5					4				4						
0.5							1							1	1
<=1		2						2							
<=2													3		
2		1						1							
<=4											3				
4			2					1					1		
<=8						4									
8			1												
16			1									3			
64												1			
>64		1													
>128											1				

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							2								
<=0.03										2					
<=0.25				1										2	2
<=0.5					2				2						
0.5				1											
<=1								1							
<=2													1		
2		2						1							
<=4											2				
4			1										1		
<=8						2									
8			1												
32												2			

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON pnl2

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	**Not Available**	**Not Available**	**Negative/Absent**	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Not Available**
	Ceftazidime synergy test	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Negative/Absent**	**Not Available**	**Not Available**	**Not Available**	**Not Available**
	ECOFF	**0.125**	**0.5**	**0.5**	**8**	**2**	**2**	**0.06**	**0.5**	**0.125**	**32**
	Lowest limit	**0.064**	**0.25**	**0.064**	**0.5**	**0.25**	**0.125**	**0.015**	**0.12**	**0.03**	**0.5**
	Highest limit	**32**	**64**	**64**	**64**	**128**	**128**	**2**	**16**	**16**	**64**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**1**	**1**	**1**	**1**	**1**	**1**	**0**	**0**	**0**	**0**
<=0.015								1			
<=0.03										1	
0.25		1							1		
4											1
8				1							
16			1			1	1				
32					1						

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**
MIC	**N of resistant isolates**	**1**	**0**	**1**	**1**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							1								
<=0.03										3					
0.03							2								
<=0.25				2										3	3
<=0.5					2				3						
<=1		1						1							
<=2													3		
2		1						2							
<=4											3				
4			1												
>4				1											
<=8						3						3			
8			2												
>8					1										
>64		1													

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.03										1					
0.03							1								
<=0.25				1										1	1
<=0.5					1				1						
<=1								1							
<=2													1		
2		1													
<=4											1				
4			1												
<=8						1									
16												1			

Table Antimicrobial susceptibility testing of Salmonella Hadar in Meat from turkey - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**3**	**0**	**0**	**0**	**1**	**0**	**3**	**0**	**0**
<=0.03										3					
<=0.25				3										2	3
<=0.5					3				3						
0.5							3							1	
<=1		2													
2		1						3							
4			3												
<=8						3						2			
16											2	1			
32											1				
64													3		

Table Antimicrobial susceptibility testing of Salmonella Hadar in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**7**	**0**	**0**	**0**	**4**	**0**	**6**	**1**	**0**
<=0.03										7					
<=0.25				6											7
0.25							1								
<=0.5					7				7						
0.5				1			4							5	
<=1		4						3							
1							2							1	
<=2													1		
2		1						4						1	
4		2	3												
<=8						7						5			
8			2												
16			2								3	1			
32											2	1			
64													3		
>64													3		
>128											2				

Table Antimicrobial susceptibility testing of Salmonella Hadar in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**
<=0.03										1					
<=0.25				1											1
<=0.5					1				1						
0.5							1								
1														1	
2		1						1							
4			1												
<=8						1									
16											1	1			
64													1		

Table Antimicrobial susceptibility testing of Salmonella Hadar in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**24**	**24**	**24**	**24**	**24**	**24**	**24**	**24**	**24**	**24**	**24**	**24**	**24**	**24**
MIC	**N of resistant isolates**	**1**	**0**	**0**	**0**	**0**	**24**	**0**	**0**	**0**	**12**	**3**	**24**	**1**	**0**
<=0.03										24					
<=0.25				22										4	23
0.25							2								
<=0.5					24				22						
0.5				2			13							9	1
<=1		10						10							
1							9		2					10	
2		6						14						1	
4		7	11												
<=8						21						4			
8			10												
16			3			3					12	16			
32											10	1	2		
64		1											13		
>64													9		
>128											2				
>1024												3			

Table Antimicrobial susceptibility testing of Salmonella Hadar in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
0.5							1								
<=1		1													
2								1							
4			1												
<=8						1						1			
16											1				
64													1		

Table Antimicrobial susceptibility testing of Salmonella Hadar in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	8	8	8	8	8	8	8	8	8	8	8	8	8	8
MIC	N of resistant isolates	0	0	0	0	0	8	0	0	0	1	1	8	0	0
<=0.03										8					
<=0.25				8										5	8
0.25							2								
<=0.5					7				7						
0.5							6							2	
<=1		7						2							
1					1				1					1	
2								6							
4		1	7												
<=8						8						5			
8			1								1				
16											6	1			
32												1			
64													7		
>64													1		
>128											1				
>1024												1			

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from turkey - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**3**	**0**	**0**	**0**	**3**	**4**	**4**	**1**	**0**
<=0.03										4					
0.03							1								
<=0.25				4											4
<=0.5					3				4						
0.5							1							1	
<=1								1							
1					1		2							2	
2		3						3						1	
<=4											1				
4		1													
<=8						2									
8			2												
16			2			2									
64													1		
>64													3		
>128											3				
>1024												4			

Table Antimicrobial susceptibility testing of Salmonella Infantis in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON pnl2

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	**Not Available**	**Not Available**	**Negative/Absent**	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Not Available**
	Ceftazidime synergy test	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Negative/Absent**	**Not Available**	**Not Available**	**Not Available**	**Not Available**
	ECOFF	**0.125**	**0.5**	**0.5**	**8**	**2**	**2**	**0.06**	**0.5**	**0.125**	**32**
	Lowest limit	**0.064**	**0.25**	**0.064**	**0.5**	**0.25**	**0.125**	**0.015**	**0.12**	**0.03**	**0.5**
	Highest limit	**32**	**64**	**64**	**64**	**128**	**128**	**2**	**16**	**16**	**64**
	N of tested isolates	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**
MIC	**N of resistant isolates**	**3**	**1**	**0**	**3**	**3**	**1**	**0**	**0**	**0**	**2**
<=0.015								3			
<=0.03										3	
<=0.125									2		
0.25		2		2					1		
0.5		1	2	1							
1			1								
2							2				
4						3	1				
16					3						
32											1
64											2

Table Antimicrobial susceptibility testing of Salmonella Infantis in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**132**	**132**	**132**	**132**	**132**	**132**	**132**	**132**	**132**	**132**	**132**	**132**	**132**	**132**
MIC	**N of resistant isolates**	**17**	**3**	**1**	**3**	**0**	**132**	**1**	**1**	**0**	**132**	**99**	**102**	**6**	**0**
<=0.03										130					
0.064										2					
0.12							1								
<=0.25				115										16	126
0.25							38								
<=0.5					108				130						
0.5				16			68							62	6
<=1	34							27							
1				1	18		21		1					48	
<=2			2										18		
2	44				3		4	104						6	
4	33	23			3								12		
<=8						91						7			
8	4	58						1							
16		46				41						21			
32		3							1			5			
64											2		33		
>64	17												69		
128											1				
>128											129				
1024												2			
>1024												97			

Table Antimicrobial susceptibility testing of Salmonella Infantis in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**
MIC	**N of resistant isolates**	**8**	**0**	**0**	**0**	**0**	**38**	**0**	**0**	**0**	**37**	**33**	**36**	**3**	**0**
<=0.03										38					
0.12							1								
<=0.25				37										2	38
0.25							4								
<=0.5					32				38						
0.5				1			23							12	
<=1		6						12							
1					6		9							21	
<=2													1		
2		10						26						3	
4		14	8				1						1		
<=8						22						2			
8			11												
16			19			16					1	2			
32												1			
64													11		
>64		8											25		
>128											37				
>1024												33			

Table Antimicrobial susceptibility testing of Salmonella Infantis in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**
MIC	**N of resistant isolates**	**0**	**1**	**0**	**0**	**0**	**7**	**0**	**0**	**0**	**7**	**6**	**7**	**0**	**0**
<=0.03										7					
<=0.25				5											5
<=0.5					5				7						
0.5				2			5								2
<=1								6							
1					2		2							7	
2		3						1							
4		4													
<=8						5									
8			2												
16			4			2						1			
32			1												
>64													7		
>128											7				
>1024												6			

Table Antimicrobial susceptibility testing of Salmonella Infantis in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	1	0	0	0	1	1	1	0	0
<=0.03										1					
<=0.25				1											1
<=0.5					1				1						
1							1							1	
2								1							
4		1													
16			1			1									
>64													1		
>128											1				
>1024												1			

Table Antimicrobial susceptibility testing of Salmonella Infantis in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							1								
<=0.03										1					
<=0.25				1											1
<=0.5					1				1						
0.5														1	
<=1		1													
<=2													1		
2								1							
<=4											1				
<=8						1									
8			1												
16												1			

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**100**	**100**	**100**	**100**	**100**	**100**	**100**	**100**	**100**	**100**	**100**	**100**	**100**	**100**
MIC	**N of resistant isolates**	**22**	**0**	**0**	**0**	**0**	**97**	**0**	**0**	**0**	**96**	**69**	**71**	**3**	**0**
<=0.015							3								
<=0.03										100					
<=0.25				91										17	96
0.25							21								
<=0.5					87				99						
0.5				9			61							37	3
<=1		20						50							
1					13		12		1					43	
<=2													17		
2		29					1	50						3	1
<=4											3				
4		29	23				2						11		
<=8						71						10			
8			36										1		
16		1	41			29					1	15			
32												6			
64		1											21		
>64		20											50		
128											1				
>128											95				
1024												3			
>1024												66			

Table Antimicrobial susceptibility testing of Salmonella Kedougou in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							1								
<=0.03										4					
0.03							3								
<=0.25				4										4	4
<=0.5					4				4						
<=1		4						1							
<=2													4		
2								3							
<=4											4				
4			2												
<=8						4									
8			2												
16												1			
32												3			

Table Antimicrobial susceptibility testing of Salmonella Kentucky in Meat from turkey - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**
MIC	**N of resistant isolates**	**7**	**0**	**0**	**0**	**1**	**7**	**0**	**3**	**0**	**7**	**5**	**5**	**0**	**0**
<=0.03										7					
<=0.25				6										2	7
<=0.5					1				4						
0.5				1										5	
<=1								4							
1					6										
<=2													2		
2								3							
4			4												
<=8						6						2			
8			3				7		1						
16									2						
64													5		
>64		7													
128						1									
>128											7				
>1024												5			

Table Antimicrobial susceptibility testing of Salmonella Kentucky in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**15**	**15**	**15**	**15**	**15**	**15**	**15**	**15**	**15**	**15**	**15**	**15**	**15**	**15**
MIC	**N of resistant isolates**	**15**	**0**	**0**	**0**	**0**	**15**	**0**	**1**	**0**	**15**	**0**	**0**	**0**	**0**
<=0.03										15					
<=0.25				15										12	15
<=0.5					6				14						
0.5														3	
<=1								3							
1					8										
<=2													15		
2					1			12							
4			9												
<=8						15						8			
8			6				14								
>8							1								
16												7			
32									1						
>64		15													
>128											15				

Table Antimicrobial susceptibility testing of Salmonella Kentucky in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**14**	**14**	**14**	**14**	**14**	**14**	**14**	**14**	**14**	**14**	**14**	**14**	**14**	**14**
MIC	**N of resistant isolates**	**13**	**0**	**0**	**0**	**0**	**14**	**0**	**8**	**0**	**14**	**13**	**13**	**0**	**0**
<=0.03										14					
<=0.25				14											13
<=0.5					1				6						
0.5														11	1
<=1								7							
1					13		1							3	
<=2													1		
2		1						7							
4			3												
<=8						14						1			
8			11				12		1						
>8							1								
16									7						
32											1				
64													6		
>64		13											7		
>128											13				
>1024												13			

Table Antimicrobial susceptibility testing of Salmonella Kentucky in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	2	0	0	0	0	2	0	0	0	2	2	2	0	0
<=0.03										2					
<=0.25				2										1	2
<=0.5					1				2						
0.5														1	
<=1								1							
1					1										
<=2			1												
2								1							
4			1												
<=8						2									
8							2								
64													1		
>64		2											1		
>128											2				
>1024												2			

Table Antimicrobial susceptibility testing of Salmonella Kentucky in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**
MIC	**N of resistant isolates**	**4**	**0**	**0**	**0**	**0**	**4**	**0**	**0**	**0**	**4**	**0**	**0**	**0**	**0**
<=0.03										4					
<=0.25				4										4	4
<=0.5					3				3						
<=1								2							
1					1				1						
<=2													4		
2								2							
4			3												
<=8						4						3			
8			1				4								
16												1			
>64		4													
>128											4				

Table Antimicrobial susceptibility testing of Salmonella Kottbus in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**
<=0.03										1					
<=0.25				1										1	1
0.25							1								
<=0.5					1				1						
<=1								1							
<=2													1		
2	1														
4		1													
<=8						1						1			
>128											1				

Table Antimicrobial susceptibility testing of Salmonella Livingstone in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1						1			
8			1												

Table Antimicrobial susceptibility testing of Salmonella Montevideo in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: environmental sample - dust

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									
8			1												
32												1			

Table Antimicrobial susceptibility testing of Salmonella Newport in Meat from turkey - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**9**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**
<=0.03										9					
<=0.25				9										8	9
0.25							1								
<=0.5					9				8						
0.5							8							1	
<=1		8						7							
1									1						
<=2													9		
2		1						2							
4			5												
<=8						9						1			
8			4												
16											8	7			
32											1	1			

Table Antimicrobial susceptibility testing of Salmonella Newport in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**	**9**
MIC	**N of resistant isolates**	**8**	**0**	**0**	**0**	**0**	**9**	**0**	**0**	**0**	**1**	**0**	**8**	**0**	**0**
<=0.03										9					
<=0.25				9											8
0.25							1								
<=0.5					8				9						
0.5							7							7	1
<=1								4							
1					1		1							2	
<=2													1		
2		1						5							
4			2												
<=8						9						1			
8			6												
16			1								8	4			
32											1	4			
64													2		
>64		8											6		

Table Antimicrobial susceptibility testing of Salmonella Oranienburg in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										2					
0.03							1								
<=0.25				2										2	2
<=0.5					2				2						
<=1		2													
<=2													2		
2								2							
<=4											2				
<=8						2						2			
8			2												

Table Antimicrobial susceptibility testing of Salmonella Oranienburg in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.03										1					
0.03							1								
<=0.25				1										1	1
<=0.5					1				1						
<=1		1													
<=2													1		
2								1							
<=4											1				
<=8						1						1			
8			1												

Table Antimicrobial susceptibility testing of Salmonella Rissen in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							2								
<=0.03										2					
<=0.25				2										2	2
<=0.5					2				2						
<=1		2						2							
<=2													2		
<=4											2				
4			2												
<=8						2									
64												2			

Table Antimicrobial susceptibility testing of Salmonella Senftenberg in Meat from turkey - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1													
<=2													1		
2								1							
<=4											1				
4			1												
<=8						1						1			

Table Antimicrobial susceptibility testing of Salmonella Senftenberg in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1													
<=2													1		
2								1							
<=4											1				
4			1												
<=8						1									
16												1			

Table Antimicrobial susceptibility testing of Salmonella Senftenberg in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1						1			
8			1												

Table Antimicrobial susceptibility testing of Salmonella Senftenberg in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
0.5							1								
<=1		1						1							
<=2													1		
4			1												
<=8						1						1			
32											1				

Table Antimicrobial susceptibility testing of Salmonella spp., unspecified in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**1**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**
<=0.03										1					
<=0.25				1											1
<=0.5					1				1						
0.5							1							1	
<=1								1							
<=8						1									
8			1								1				
32												1			
64													1		
>64		1													

Table Antimicrobial susceptibility testing of Salmonella spp., unspecified in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	1	0	0	0	1	0	0	0	0
<=0.03										1					
<=0.25				1											1
<=0.5									1						
0.5														1	
1					1		1								
<=2													1		
2	1							1							
<=8												1			
16			1			1									
>128											1				

Table Antimicrobial susceptibility testing of Salmonella Stanley in Meat from turkey - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**6**	**6**	**6**	**6**	**6**	**6**	**6**	**6**	**6**	**6**	**6**	**6**	**6**	**6**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**6**	**0**	**0**	**0**	**6**	**0**	**0**	**0**	**0**
<=0.03										6					
<=0.25				6										3	6
0.25							5								
<=0.5					6				4						
0.5							1							3	
<=1		2						6							
<=2													6		
2		3							2						
4		1	2												
<=8						6									
8			4												
16												4			
64												1			
128												1			
>128											6				

Table Antimicrobial susceptibility testing of Salmonella Stanley in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**5**	**5**	**5**	**5**	**5**	**5**	**5**	**5**	**5**	**5**	**5**	**5**	**5**	**5**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**5**	**0**	**0**	**0**	**5**	**0**	**0**	**0**	**0**
<=0.03										5					
<=0.25				5										1	5
0.25							5								
<=0.5					5				5						
0.5														4	
<=1		5						4							
<=2													5		
2								1							
4			5												
<=8						5									
16												4			
32												1			
>128											5				

Table Antimicrobial susceptibility testing of Salmonella Szentes in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							2								
<=0.03										2					
<=0.25				2										2	2
<=0.5					2				2						
<=1		1													
<=2													2		
2		1						2							
<=4											2				
<=8						2									
8			2												
16												2			

Table Antimicrobial susceptibility testing of Salmonella Tennessee in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							1								
<=0.03										1					
<=0.25				1											1
<=0.5					1				1						
0.5														1	
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									
8			1												
16												1			

Table Antimicrobial susceptibility testing of Salmonella Tennessee in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1						1			
8			1												

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
4			1												
<=8						1						1			

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
<=0.015							1								
<=0.03										2					
0.03							1								
<=0.25				2										1	2
<=0.5					2				1						
0.5														1	
1									1						
<=2													2		
2		2						2							
<=4											2				
<=8						2						1			
8			2												
16												1			

Table Antimicrobial susceptibility testing of Salmonella Typhimurium, monophasic in Meat from turkey - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**
<=0.03										1					
0.03							1								
<=0.25				1										1	1
<=0.5					1				1						
<=1		1													
2								1							
<=4											1				
4			1												
<=8						1						1			
>64													1		

Table Antimicrobial susceptibility testing of Salmonella Typhimurium, monophasic in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**
MIC	**N of resistant isolates**	**1**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**1**	**1**	**0**	**0**
<=0.015							2								
<=0.03										3					
0.03							1								
<=0.25				3										3	3
<=0.5					3				3						
<=1		2						1							
<=2													2		
2								2							
<=4											3				
4			2												
<=8						3									
8			1												
16												2			
>64		1											1		
>1024												1			

Table Antimicrobial susceptibility testing of Salmonella Typhimurium, monophasic in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**1**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**1**	**1**	**0**	**0**
<=0.03										1					
0.03							1								
<=0.25				1										1	1
<=0.5					1				1						
<=1								1							
<=4											1				
<=8						1									
8			1												
>64		1											1		
>1024												1			

Table Antimicrobial susceptibility testing of Salmonella Typhimurium, monophasic in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**1**	**1**	**0**	**0**
<=0.015							1								
<=0.03										1					
<=0.25														1	1
<=0.5					1										
0.5				1											
<=1								1							
2		1													
<=4											1				
<=8						1									
8			1												
16									1						
>64													1		
>1024												1			

ANTIMICROBIAL RESISTANCE TABLES FOR INDICATOR ESCHERICHIA COLI

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method:

Country of Origin: Romania

Sampling Details:

Ceftazidime synergy test	Cefotaxime synergy test	MIC	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
			ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
			Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
			Highest limit	32	64	64	64	128	128	2	16	16	64
			N of tested isolates	4	4	4	4	4	4	4	4	4	4
			N of resistant isolates	4	4	1	1	3	1	1	0	0	0
Not Available	Not Available	<=0.015								3			
		<=0.03										4	
		<=0.125									2		
		0.12								1			
		0.25		1							2		
		0.5		1				1					
		1			1								
		2						2					1
		4		1			2						1
		8		1			1						1
		16			3			1					1
		>64					1						
	Positive/Present	<=0.064				3							

			AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
			ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
			Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
			Highest limit	32	64	64	64	128	128	2	16	16	64
			N of tested isolates	4	4	4	4	4	4	4	4	4	4
Ceftazidime synergy test	**Cefotaxime synergy test**	**MIC**	**N of resistant isolates**	4	4	1	1	3	1	1	0	0	0
Not Available	Negative/Absent	8				1							
Positive/Present	Not Available	<=0.125							2				
Negative/Absent	Not Available	<=0.125							1				
		16							1				

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.25**	**0.5**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**64**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**	**4**
MIC	**N of resistant isolates**	**4**	**1**	**4**	**3**	**2**	**4**	**0**	**2**	**0**	**4**	**3**	**2**	**0**	**2**
<=0.03										4					
<=0.25														4	
0.25							1								
<=0.5					1				2						
0.5															1
<=1								4							
1				1											1
<=2													2		
2					2										
4			1												
>4				3											
<=8						2						1			
8			2				1								
>8					1		2								
16									1						
32						1			1						
>32															2
64			1			1					1				
>64		4											2		
>128											3				
>1024												3			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method:

Country of Origin: Hungary

Sampling Details:

Ceftazidime synergy test	Cefotaxime synergy test	MIC	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
			ECOFF	**0.125**	**0.25**	**0.25**	**8**	**0.5**	**0.5**	**0.06**	**0.5**	**0.125**	**32**
			Lowest limit	**0.064**	**0.25**	**0.064**	**0.5**	**0.25**	**0.12**	**0.015**	**0.12**	**0.03**	**0.5**
			Highest limit	**32**	**64**	**64**	**64**	**128**	**128**	**2**	**16**	**16**	**64**
			N of tested isolates	**142**	**142**	**142**	**142**	**142**	**142**	**142**	**142**	**142**	**142**
			N of resistant isolates	**131**	**142**	**61**	**64**	**138**	**61**	**2**	**0**	**0**	**0**
Not Available	Not Available	<=0.015								65			
		<=0.03										142	
		0.03								39			
		<=0.064		1									
		0.064								36			
		<=0.125									115		
		0.12		10						2			
		0.25		52							27		
		0.5		16				4					
		1		4	4			22					
		2		15	13		5	29					12
		4		28	14		45	4					68
		8		16	45		28	43					54
		16			40		3	35					7
		32			21		8	5					1
		64			4		40						
		>64			1		13						

Ceftazidime synergy test	Cefotaxime synergy test	MIC	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
			ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
			Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
			Highest limit	32	64	64	64	128	128	2	16	16	64
			N of tested isolates	142	142	142	142	142	142	142	142	142	142
			N of resistant isolates	131	142	61	64	138	61	2	0	0	0
Not Available	Positive/Present	<=0.064				72							
		0.12				8							
		0.25				1							
	Negative/Absent	2				1							
		4				39							
		8				19							
		16				2							
Positive/Present	Not Available	<=0.125							56				
		0.25							17				
		0.5							1				
Negative/Absent	Not Available	<=0.125							3				
		0.25							4				
		1							1				
		2							3				
		4							31				
		8							25				
		16							1				

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.25**	**0.5**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**64**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**142**	**142**	**142**	**142**	**142**	**142**	**142**	**142**	**142**	**142**	**142**	**142**	**142**	**142**
MIC	**N of resistant isolates**	**142**	**2**	**142**	**138**	**24**	**130**	**0**	**46**	**0**	**114**	**87**	**76**	**1**	**21**
<=0.015							11								
<=0.03										142					
0.03							1								
0.12							2								
<=0.25														130	94
0.25							17								
<=0.5					4				84						
0.5							16							11	25
<=1								140							
1				4	22		9		12						2
<=2			2										64		
2				15	29		4	2						1	
<=4											12				
4			83	10	5		11						2		
>4				113											
<=8						117						53			
8			52		29		51				11				
>8					53		20								
16			3			1			10		5	2			
32									8		1		1		
>32									28						21

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.25**	**0.5**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**64**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**142**	**142**	**142**	**142**	**142**	**142**	**142**	**142**	**142**	**142**	**142**	**142**	**142**	**142**
MIC	**N of resistant isolates**	**142**	**2**	**142**	**138**	**24**	**130**	**0**	**46**	**0**	**114**	**87**	**76**	**1**	**21**
64		3	2			1					13		30		
>64		139											45		
128						22					18				
>128						1					82				
1024												1			
>1024												86			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON pnl2

Analytical Method:

Country of Origin: Hungary

Sampling Details:

Ceftazidime synergy test	Cefotaxime synergy test	MIC	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
			ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
			Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
			Highest limit	32	64	64	64	128	128	2	16	16	64
			N of tested isolates	2	2	2	2	2	2	2	2	2	2
			N of resistant isolates	1	2	2	2	2	2	0	0	0	0
Not Available	Not Available	<=0.03										2	
		<=0.064		1									
		0.064								2			
		<=0.125									1		
		0.25		1							1		
		1			1								
		4			1			1					1
		8						1					1
		16					1						
		64					1						
	Negative/Absent	1				1							
		4				1							
Negative/Absent	Not Available	2							1				
		4							1				

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.25**	**0.5**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**64**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**
MIC	**N of resistant isolates**	**87**	**1**	**2**	**2**	**25**	**151**	**0**	**15**	**0**	**140**	**59**	**64**	**0**	**35**
<=0.015							18								
<=0.03										170					
0.03							1								
0.12							1								
<=0.25				168										154	112
0.25							35								
<=0.5					168				133						
0.5							15							15	19
<=1		6						167							
1							22		20					1	2
<=2			16										100		
2		38		1	1		4	3	2						2
<=4											20				
4		36	70				16						6		
>4				1											
<=8						138						97			
8		3	70				41		2		6				
>8					1		17								
16			13			7			3		4	12			
32		1	1			6			4		3	2	1		
>32									6						35

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.25**	**0.5**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**64**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**
MIC	**N of resistant isolates**	**87**	**1**	**2**	**2**	**25**	**151**	**0**	**15**	**0**	**140**	**59**	**64**	**0**	**35**
64						4					7		18		
>64		86											45		
128						11					29				
>128						4					101				
1024												1			
>1024												58			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method:

Country of Origin: Hungary

Sampling Details:

Ceftazidime synergy test	Cefotaxime synergy test	MIC	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
			ECOFF	**0.125**	**0.25**	**0.25**	**8**	**0.5**	**0.5**	**0.06**	**0.5**	**0.125**	**32**
			Lowest limit	**0.064**	**0.25**	**0.064**	**0.5**	**0.25**	**0.12**	**0.015**	**0.12**	**0.03**	**0.5**
			Highest limit	**32**	**64**	**64**	**64**	**128**	**128**	**2**	**16**	**16**	**64**
			N of tested isolates	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**
			N of resistant isolates	**103**	**121**	**63**	**64**	**116**	**63**	**4**	**0**	**0**	**0**
Not Available	Not Available	<=0.015								44			
		<=0.03										119	
		0.03								42			
		<=0.064		4									
		0.064								31		2	
		<=0.125									83		
		0.12		14						4			
		0.25		45							37		
		0.5		6				5			1		
		1		2	2			14					
		2		10	13		4	24					8
		4		19	10		30	7					59
		8		18	37		23	45					49
		16		2	33		6	22					5
		32		1	14		4	3					
		64			6		49	1					
		>64			6		5						

			AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
			ECOFF	**0.125**	**0.25**	**0.25**	**8**	**0.5**	**0.5**	**0.06**	**0.5**	**0.125**	**32**
			Lowest limit	**0.064**	**0.25**	**0.064**	**0.5**	**0.25**	**0.12**	**0.015**	**0.12**	**0.03**	**0.5**
			Highest limit	**32**	**64**	**64**	**64**	**128**	**128**	**2**	**16**	**16**	**64**
			N of tested isolates	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**
Ceftazidime synergy test	**Cefotaxime synergy test**	**MIC**	**N of resistant isolates**	**103**	**121**	**63**	**64**	**116**	**63**	**4**	**0**	**0**	**0**
Not Available	Positive/Present	<=0.064				53							
		0.12				5							
		4				1							
	Negative/Absent	0.5				1							
		1				4							
		2				3							
		4				34							
		8				19							
		16				1							
Positive/Present	Not Available	<=0.125							34				
		0.25							13				
		0.5							1				
Negative/Absent	Not Available	<=0.125							5				
		0.25							5				
		1							3				
		2							8				
		4							25				
		8							26				
		64							1				

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.25**	**0.5**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**64**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**
MIC	**N of resistant isolates**	**121**	**4**	**121**	**116**	**20**	**104**	**0**	**32**	**0**	**97**	**63**	**63**	**0**	**21**
<=0.015							15								
<=0.03										120					
0.03							2								
0.064										1					
<=0.25														100	84
0.25							13								
<=0.5					5				82						
0.5							7							19	15
<=1								117							
1				1	19		2		6					2	1
<=2			7										56		
2				12	23		5	4	1						
<=4											20				
4			59	12	11		8						2		
>4				96											
<=8						98						56			
8			47		26		52		1		2				
>8					37		17								
16			4			3			4		2	2			
32			1			1			7		1		1		
>32									20						21

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.25**	**0.5**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**64**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**	**121**
MIC	**N of resistant isolates**	**121**	**4**	**121**	**116**	**20**	**104**	**0**	**32**	**0**	**97**	**63**	**63**	**0**	**21**
64		3	3			2					10		32		
>64		118											30		
128						16					10				
>128						1					76				
1024												2			
>1024												61			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Turkeys - fattening flocks

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.25**	**0.5**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**64**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**
MIC	**N of resistant isolates**	**93**	**5**	**0**	**0**	**42**	**99**	**3**	**5**	**0**	**66**	**65**	**116**	**0**	**38**
<=0.015							65								
<=0.03										170					
0.03							4								
0.064							2								
0.12							1								
<=0.25				170										160	91
0.25							32								
<=0.5					170				151						
0.5							21							10	39
<=1		2						167							
1							3		14						2
<=2			19										54		
2		32					4								
<=4											72				
4		40	97				3	2							
<=8						120						77			
8		3	46				21	1	2		21				
>8							14								
16			3			8					11	25			
32			4			3			1		2	3	3		
>32									2						38

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.25**	**0.5**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**64**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**	**170**
MIC	**N of resistant isolates**	**93**	**5**	**0**	**0**	**42**	**99**	**3**	**5**	**0**	**66**	**65**	**116**	**0**	**38**
64		1	1			7					7		32		
>64		92											81		
128						13					11				
>128						19					46				
>1024												65			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Turkeys - fattening flocks

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method:

Country of Origin: Hungary

Sampling Details:

Ceftazidime synergy test	Cefotaxime synergy test	MIC	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
			ECOFF	**0.125**	**0.25**	**0.25**	**8**	**0.5**	**0.5**	**0.06**	**0.5**	**0.125**	**32**
			Lowest limit	**0.064**	**0.25**	**0.064**	**0.5**	**0.25**	**0.12**	**0.015**	**0.12**	**0.03**	**0.5**
			Highest limit	**32**	**64**	**64**	**64**	**128**	**128**	**2**	**16**	**16**	**64**
			N of tested isolates	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**
			N of resistant isolates	**42**	**51**	**16**	**16**	**46**	**16**	**0**	**0**	**0**	**0**
Not Available	Not Available	<=0.015								35			
		<=0.03										50	
		0.03								13			
		0.064								3		1	
		<=0.125									38		
		0.12		9									
		0.25		7							13		
		0.5						5					
		1		5	2			10					
		2		12	3		2	9					1
		4		6	2		19	3					20
		8		6	14		14	14					25
		16		6	3			8					5
		32			12		7	2					
		64			10		8						
		>64			5		1						

Ceftazidime synergy test	Cefotaxime synergy test	MIC	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
			ECOFF	**0.125**	**0.25**	**0.25**	**8**	**0.5**	**0.5**	**0.06**	**0.5**	**0.125**	**32**
			Lowest limit	**0.064**	**0.25**	**0.064**	**0.5**	**0.25**	**0.12**	**0.015**	**0.12**	**0.03**	**0.5**
			Highest limit	**32**	**64**	**64**	**64**	**128**	**128**	**2**	**16**	**16**	**64**
			N of tested isolates	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**
			N of resistant isolates	**42**	**51**	**16**	**16**	**46**	**16**	**0**	**0**	**0**	**0**
Not Available	Positive/Present	<=0.064				30							
		0.12				4							
	Negative/Absent	0.25				1							
		1				4							
		2				1							
		4				9							
		8				2							
Positive/Present	Not Available	<=0.125							16				
		0.25							11				
Negative/Absent	Not Available	<=0.125							4				
		0.25							3				
		0.5							1				
		1							1				
		2							4				
		4							4				
		8							7				

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Turkeys - fattening flocks

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method:

Country of Origin: Hungary

Sampling Details:

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.25**	**0.5**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**64**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**
MIC	**N of resistant isolates**	**51**	**4**	**51**	**46**	**25**	**48**	**0**	**5**	**0**	**40**	**27**	**44**	**0**	**14**
<=0.015							3								
<=0.03										51					
<=0.25														34	30
0.25							11								
<=0.5					5				39						
0.5							10							15	7
<=1								50							
1				1	11				7					2	
<=2			1										6		
2				3	8			1							
<=4											3				
4			8	6	5		1						1		
>4				41											
<=8						26						22			
8			36		12		13				4				
>8					10		13								
16			2						1		4	2			
32									4				3		
>32															14
64		3	4			1					2		19		
>64		48											22		

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.25**	**0.5**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**64**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**	**51**
MIC	**N of resistant isolates**	**51**	**4**	**51**	**46**	**25**	**48**	**0**	**5**	**0**	**40**	**27**	**44**	**0**	**14**
128						8					3				
>128						16					35				
>1024												27			

OTHER ANTIMICROBIAL RESISTANCE TABLES

Specific monitoring of ESBL-/AmpC-/carbapenemase-producing bacteria and specific monitoring of carbapenemase-producing bacteria, in the absence of isolate detected

Programme Code	Matrix Detailed	Zoonotic Agent Detailed	Sampling Strategy	Sampling Stage	Sampling Details	Sampling Context	Sampler	Sample Type	Sampling Unit Type	Sample Origin	Comment	Total Units Tested	Total Units Positive
CARBAMON	Gallus gallus (fowl) - broilers	Escherichia coli, non-pathogenic, unspecified	Objective sampling	Slaughterhouse	N_A	Monitoring	Official sampling	animal sample - caecum	slaughter animal batch	Hungary	N_A	300	0
	Meat from broilers (Gallus gallus) - fresh	Escherichia coli, non-pathogenic, unspecified	Objective sampling	Retail	sampOrigs are variable	Monitoring	Official sampling	food sample - meat	batch (food/feed)	Unknown	N_A	300	0
	Turkeys - fattening flocks	Escherichia coli, non-pathogenic, unspecified	Objective sampling	Slaughterhouse	N_A	Monitoring	Official sampling	animal sample - caecum	slaughter animal batch	Hungary	N_A	300	0

Specific monitoring of ESBL-/AmpC-/carbapenemase-producing bacteria and specific monitoring of carbapenemase-producing bacteria, in the absence of isolate detected

Latest Transmission set

Table Name	Last submitted dataset transmission date
Animal Population	22-Jul-2022
Disease Status	22-Jul-2022
Food Borne Outbreaks	22-Jul-2022
Prevalence	26-Jul-2022

Institutions and Laboratories involved in zoonoses monitoring and reporting

National Food Chain Safety Office:
Veterinary Diagnostic Directorate, Animal Health and Animal Welfare Directorate, Food Chain Safety Laboratory Directorate, Food and Feed Safety Directorate

Short description of the institutions and laboratories involved in data collection and reporting

Animal population

1. Sources of information and the date(s) (months, years) the information relates to[(a)]

National animal identification and registration database, National reports and the Central Statistical Office, other central databases (herd register system, etc.)

2. Definitions used for different types of animals, herds, flocks and holdings as well as the production types covered

We report the population data by animal.

(a): National identification and registration system(s), source of reported statistics (Eurostat, others)
(b): Link to website with density maps if available, tables with number of herds and flocks according to geographical area

General evaluation*: Mycobacterium

1. History of the disease and/or infection in the country[(a)]

Hungary is officially free from bovine tuberculosis.

3. Any recent specific action in the Member State or suggested for the European Union[(b)]

All farm workers have to be checked (including also screening for TBC) by the competent public health authority for their compliance with the rules set for persons dealing with animals and food intended for human consumption. The documents proving their compliance are subject to on farm checks performed by the veterinary service**.**

* **For each zoonotic agent**
(a): Epidemiological evaluation (trends and sources) over time until recent/current situation for the different relevant matrixes (food, feed, animal). If relevant: the official "disease status" to be specified for the whole country and/or specific regions within the country
(b): If applicable

General evaluation*: Brucella

1. History of the disease and/or infection in the country[(a)]

.The last time bovine brucellosis occurred in Hungary was in 1985. Ovine and caprine brucellosis has never occured in Hungary. Laboratory tests occasionally showed B. suis bv. 2 of pig abortions and of wild boars.

* **For each zoonotic agent**
(a): Epidemiological evaluation (trends and sources) over time until recent/current situation for the different relevant matrixes (food, feed, animal). If relevant: the official "disease status" to be specified for the whole country and/or specific regions within the country
(b): If applicable

General evaluation*: Salmonella

1. History of the disease and/or infection in the country[(a)]

In 1992 the Veterinary Science Committee of the Hungarian Academy of Sciences has established its Salmonella Subcommittee with the main aim to support the work of the Hungarian Ministry of Agriculture and Rural Development in the control of Salmonella with regards to poultry flocks. After the accession the EC regulations became directly applicable in Hungary as well. From that time EC regulations are followed. The implementation of these regulations is regulated by national Decree 180/2009. (XII. 29.) of Ministry of Agriculture. Due to the control programs, salmonella prevalence decreased significally in the last decades. EU prevalence targets were reached first in 2012. In the last years the prevalence remained under the limit in all types of poultry, with the exception of Gallus gallus breeders in 2013 (1,1%) and in Gallus gallus layers in 2016 (2,48%). Relevant salmonella serotypes in turkey breeders have not been observed for years, unti 2016, when mST was found at one farm. **In 2020 the EU targets were achieved in all production types again.**

3. Any recent specific action in the Member State or suggested for the European Union[(b)]

Vaccination is not compulsory in flocks of Gallus gallus and Meleagris gallopavo. The rules of using vaccination and treatment are laid down in Commission Regulation (EC) No 200/2010 and in Regulation (EC) No 2160/2003 of the European Parliament and of the Council.

* **For each zoonotic agent**
(a): Epidemiological evaluation (trends and sources) over time until recent/current situation for the different relevant matrixes (food, feed, animal). If relevant: the official "disease status" to be specified for the whole country and/or specific regions within the country
(b): If applicable

General evaluation*: Yersinia

4. Additional information

Diagnostic methods: bacteriological examination and PCR

* **For each zoonotic agent**
(a): Epidemiological evaluation (trends and sources) over time until recent/current situation for the different relevant matrixes (food, feed, animal). If relevant: the official "disease status" to be specified for the whole country and/or specific regions within the country
(b): If applicable

General evaluation*: Trichinella

1. History of the disease and/or infection in the country[(a)]

In Hungary, mandatory testing for ***Trichinella*** spp. is in place since 1960. Slaughtered susceptible animals intended to be placed on the market are subject to mandatory testing for ***Trichinella*** spp.

2. Evaluation of status, trends and relevance as a source for humans

Trichinellosis was a significant zoonotic disease in Hungary in the 1950s and 1960s. Due to the introduction of control strategies, the average annual incidence of human trichinellosis decreased to 0-0.7 cases per 100,000 for the early 1990s. The decrease of incidence observed in humans is similar to that of prevalence seen in swine at slaughterhouses. In the past decade, the annual incidence of autochtonous human cases dropped to 0 case per 100,000, and the parasite was not detected in swine. Nevertheless, ***Trichinella*** spp. are present in the Hungarian wildlife. The prevalence of infection was low (0.01%) in wild boars and relatively high in red foxes (2.1%) between 2009 and 2018. The dominant species was *Trichinella britovi* in both wild boars (51% of the isolates) and red foxes (79% of the isolates). *Trichinella spiralis* was found less frequently in wild boars and foxes. *Trichinella pseudospiralis* was also detected in three wild boars. The prevalence of trichinellosis was high (9.1%) in golden jackals. In this host, *T. spiralis* is the dominant species. As jackals migrate for long distances, they may play a significant role in the long distance spread of *T. spiralis* from the surrounding endemic countries. There was no correlation between environmental parameters in the home range of foxes and wild boars and the *T. spiralis* larval counts, but there was a positive correlation between the boundary zone of Hungary and *T. spiralis* infection ($P<0.0001$; odds ratio: 24.1). These results indicate that the distribution of *T. spiralis* in the Hungarian wildlife is determined by the transborder transmission of the parasite from the surrounding endemic countries. Based on the statistical analysis, non-agricultural areas (forests, scrubs, herbaceous vegetation and pastures) and the mean annual temperature were the major determinants of the spatial distribution of *T. britovi* in Hungary. The positive relationship with non-agricultural areas can be explained by the generalist feeding behaviour including scavenging of foxes in these areas. The negative relationship with the mean annual temperature can be attributed to the slower decomposition of wildlife carcasses favouring a longer survival of *T. britovi* larvae in the host carrion and to the increase of scavenging of foxes.

3. Any recent specific action in the Member State or suggested for the European Union[(b)]

Mandatory testing during meat inspection in all susceptible species (swine, horse, nutria, wild boar).

* **For each zoonotic agent**
(a): Epidemiological evaluation (trends and sources) over time until recent/current situation for the different relevant matrixes (food, feed, animal). If relevant: the official "disease status" to be specified for the whole country and/or specific regions within the country
(b): If applicable

General evaluation*: Echinococcus

1. History of the disease and/or infection in the country[(a)]

Cystic echinococcosis caused by Echinococcus granulosus was a significant zoonosis in Hungary in the 1960s and 1970s. Due to the introduction of integrated control strategies, the average annual incidence of human cystic echinococcosis decreased to 0.08-0.2 case per 100,000 population for the early 1990s. The decrease of incidence observed in humans is almost parallel with that of overall prevalence seen in swine, sheep and cattle at slaughterhouses. Echinococcus multilocularis was not detected in man or animals in Hungary until 2002. The parasite was detected in foxes and in humans for the first time in 2002 and 2008, respectively. In the past decade, 16 alveolar echinococcosis cases were described including autochtonous cases. Feedbacks from ruminant slaughterhouses showed an evidence of cystic echinococcus infections in cattle in 2020 and in 2021 as well.

2. Evaluation of status, trends and relevance as a source for humans

In the past decade, the annual incidence of cystic echinococcosis was 0.05-0.1 case per 100,000 human population. Echinococcosis was confirmed in 119 animals between 2015 and 2020. *Echinococcus intermedius* (n = 76), *Echinococcus granulosus* sensu stricto (n = 7), and *Echinococcus multilocularis* (n = 3) was identified in swine. In cattle, only *E. granulosus* s. s. (n = 24) was detected. *E. granulosus* s. s. (n = 7) was the dominant species in sheep; nevertheless, *E. intermedius* was also identified in two animals. Cysts were fertile in nine sheep (100%), 30 swine (64%), and three cattle (14%) indicating that all three species play a role in some extent in the epidemiology of cystic echinococcosis in Hungary. Based on the number of animals killed in the slaughterhouses involved in the present study, the rate of infection was 0.012% in sheep, 0.007% in cattle, and 0.001% in swine. Animals with hydatidosis originated from family farms
E. multilocularis was first detected in red foxes (Vulpes vulpes) in Hungary in the northern border area in 2002. Between 2002 and 2004, the parasite was described in 7 northern counties with low overall prevalence (8.7%) in foxes. In the study carried out between 2009 and 2010, *E. multilocularis* was detected in foxes of 16 out of the 19 Hungarian counties and in the suburban areas of the capital,

Budapest. The prevalence of infection was significantly higher in the north-western half (16.2%) than in the south-eastern half (4.2%) of the country. The multilocus microsatellite analysis of the isolates indicate that Hungary should be considered as a peripheral area of a single European focus, where the dispersal movement of foxes resulted in the spreading of *E. multilocularis* within a time period short enough to avoid a substantial genetic drift. Out of 3265 foxes, *Echinococcus multilocularis* infection was detected in 247 animals (prevalence: 7.6%) between 2009 and 2019. Out of 15 golden jackals, one individual was infected with *E. multilocularis* (prevalence: 6.7%). The prevalence of infection was almost four times higher in the north-western half of the country than in the south-eastern half. In some northern areas, the prevalence was 35-40%. Based on statistical analysis, mean annual temperature and annual precipitation were the major determinants of the spatial distribution of *E. multilocularis* in Hungary. It can be attributed to the sensitivity of *E. multilocularis* eggs to high temperatures and desiccation.

* **For each zoonotic agent**
(a): Epidemiological evaluation (trends and sources) over time until recent/current situation for the different relevant matrixes (food, feed, animal). If relevant: the official "disease status" to be specified for the whole country and/or specific regions within the country
(b): If applicable

General evaluation*: Rabies

1. History of the disease and/or infection in the country[(a)]

At the beginning of the twentieth century, the urban rabies was present in Hungary (affecting domestic animals) and was transmitted to humans mainly by dogs. Therefore, in the 1930's strict animal health regulations were introduced. These measures included nationwide mandatory regular vaccination of dogs over three months of age. During World War II, epidemiological actions were hindered, which resulted in a re-emergence of urban rabies in 1946-47. As a result of the re-introduction of regulatory measures as well as mandatory preventive vaccination, urban rabies became sporadic in Hungary. The register of the annual vaccination of dogs shows that around 1.1-1.5 million dogs are vaccinated every year Preventive vaccination of cats against rabies is recommended but not mandatory and special epidemiological aspects are to be considered. Sylvatic rabies reached the North-Eastern part of Hungary in the year 1954. Until 1966 cases remained sporadic (a total of 97 foxes, 16 badgers and wild cats confirmed positive for rabies). In the same timeframe, 35 dogs and 96 domestic cats were confirmed positive for the disease. In 1967, sylvatic rabies crossed the Danube and by 1971 the whole country was infected. At this time, intensive attempts were executed in order to reduce the number of foxes, with minimum results. These actions were suspended in 1987. Between 1988 and 1996 around 1000 rabies cases in foxes were diagnosed per year. Oral vaccination of foxes was introduced in Hungary in 1992. From that year, the rabies cases in foxes decreased year by year, as the vaccination zone was extended from the western part of the country to the whole territory of Hungary (2005-2007). The efficacy of the oral immunization of foxes can be demonstrated by the drastic decrease in the number of rabies cases in the country. In the calendar years 2005 only 9, in 2006 only 3, in 2007 only 4, in 2008 only 7 and in 2009 only 2 positive cases could be detected for the whole territory of the country. In 2010 Lyssaviruses were detected in 11 cases in Hungary: 1 dog, 9 foxes (RABV) and 1 bat (EBLV-1). In 2011 and 2012 no rabies cases were diagnosed in domestic animals or wildlife (except 3 bats, EBLV-1). In September 2013 rabies was diagnosed in a red fox originating from Bács-Kiskun county, a territory that had not been vaccinated since 2008. In 2013, 24 cases were detected within 3,5

months. An emergency ring vaccination was implemented in autumn 2013. In 2014, 23 cases were detected while vaccination area was extended to the north up to highway M3 (E71) and in this area a double baiting density was applied (40 baits/km2). After 3 consecutive campaigns in the infected area, no further cases were found. The epidemic concerned 3 counties (Bács-Kiskun, Pest, Jász-Nagykun-Szolnok), and 47 cases were found in total, of which 4 in domestic animals (2 cattle, 1 goat, 1 dog) and 43 in wild animals (1 roe deer, 42 foxes). In 2015, vaccination area was further extended to the north and no rabies cases were diagnosed in domestic animals or wildlife. Only one bat (EBLV-1, Pest county) and one fox was found positive (Békés county, vaccine induced case, confirmed by the EURL as well.)
In February 2016 rabies was confirmed in the laboratory of the Veterinary Diagnostic Directorate (NRL for rabies) of the National Food Chain Safety Office of Hungary, from a red fox originating from Borsod-Abaúj-Zemplén county (north-eastern part of Hungary), within the vaccination area. The virus strain isolated from the sample was different from the strain detected during the 2013-2014 rabies epidemic in Hungary. In March 2017, in the same area, a red fox showing neurological symptoms was found positive. The strain was identical to the one isolated in 2016. Two weeks later, in a nearby farm, rabies was confirmed in two goats.
During the years 2018-2020, no classical rabies cases were detected in Hungary (only one case of EBLV-1 in bats, Pest county, January 2018). Last human rabies case in Hungary occurred in 1994.

2. Evaluation of status, trends and relevance as a source for humans

After the rabies epidemic in 2013-2014, the territory under oral vaccination campaigns was extended respectively. In 2015 there were no rabies cases in Hungary. (1 bat, EBLV-1 and 1 vaccine induced case in fox). In February 2016, a rabid fox was found in Borsod-Abaúj-Zemplén county. In the same area, in March 2017 a rabid fox was detected and two goats were infected too. No rabies cases were found in Hungary since March 2017. According to EU legislation, Hungary is a country free of rabies and is listed in the Part I of Annex III to Commission Implementing Regulation (EU) 2021/620. However, due to the presence of the disease in neighbouring countries (Ukraine, Romania), measures against the re-introduction of the disease are continued: yearly vaccination of dogs is still mandatory and oral vaccination of foxes is continued along the eastern and southern country border.

3. Any recent specific action in the Member State or suggested for the European Union[(b)]

In order to eradicate rabies from Hungary and to protect public health, regulatory measures on domestic animals are in place. Regular preventive vaccination of dogs is mandatory two times between 3 months of age and under 1 year of age with monovalent vaccine. After 1 year of age vaccination shall be repeated annually. Stray dogs are removed from public areas and are vaccinated against the disease.
Oral vaccination of foxes is performed twice yearly in a specific part of Hungary's territory. The vaccination area is regularly reviewed by the central authority and adapted to the epidemiological situation.
An awareness campaign was started in 2016, by the central veterinary authority, in order to increase reporting of foxes found dead or animals showing symptoms suspicious for rabies. Leaflets have been produced and distributed, explaining the importance of rabies and describing the symptoms of the disease and the way of reporting the suspicion of the disease to the veterinary services. A website has been developed as well, specifically dedicated for rabies, in order to provide information about the disease to the public (veszettsegmentesites.hu). In 2017-2020, the awareness activities were continued by broadcasting TV and internet spots and billboards along public roads. In 2020, also a mini-series of educational videos about rabies have been produced and published.

Golden jackals are a species of concern in some areas of Hungary. Sampling and laboratory testing of golden jackals was within the framework of monitoring of effectiveness of OV was not eligible for co-financing until 2016. We welcome the decision of the Commission to finance the testing of this species as of 2016.

* **For each zoonotic agent** (a): Epidemiological evaluation (trends and sources) over time until recent/current situation for the different relevant matrixes (food, feed, animal). If relevant: the official “disease status” to be specified for the whole country and/or specific regions within the country (b): If applicable

Description of Monitoring/Surveillance/Control programmes system*: Mycobacterium

1. Monitoring/Surveillance/Control programmes system[(a)]

Sampling strategy
Post mortem inspections: According to the meat inspection rules in force in Hungary, based on a tradition of at least a century, each animal for slaughter is to be checked individually ante and post mortem. Technical methods applied at meat inspection is suitable to detect even the slightest tuberculotic lesions. The legal provisions for tuberculosis require that the organs, together with the lymphnodes belonging to them, shall be sent to the National Food Chain Safety Office, Veterinary Diagnostic Directorate for further laboratory examination, if during post mortem inspection of a slaughtered animal the tuberculotic lesions are revealed. In case of animals ordered to be slaughtered for establishing the reason for unclarified positive or inconclusive reactions during intradermal tuberculin testing, a set of lymph nodes belonging to several organs and systems, shall be sent to the National Food Chain Safety Office, Veterinary Diagnostic Directorate. Intradermal tuberculin testing, together with the post mortem control program, the compulsory intradermal tuberculin testing with a yearly interval of the whole Hungarian cattle population (older than 12 monthsshas been maintained and executed.

Methods of sampling (description of sampling techniques)
The rules of taking samples are the followings: All samples taken from animals with a large body (cattle, swine) must include the organs showing signs of the disease and the adjacent lymphatic glands, in case of birds and smaller animals the sample must be an entire carcassl; For the purpose of detecting the presence of mycobacteria from the feedingstuffs, litter, soil etc. 20-50 gramm samples must be taken, 20 gramm samples from faeces, 50cm3 from urine and 1 litre from drinking water. The samples must be sent to the VDD with a view to carry out tests to detect tuberculosis and confirm the presence of mycobacteria.

Case definition
An animal is considered a positive case, if the presence of bovine tuberculosis is confirmed by the isolation of M. bovis, M. caprae or M. tuberculosis from its lymph node(s) or parenchymatous organs on laboratory examination. Suspension or withdrawal of the free status of a herd is based upon the analysis of the results of the intradermal tuberculin tests (if necessary, repeated and completed by simultaneous testing), post mortem examinations and laboratory tests.The officially tuberculosis -free status of the herd have to be withdrawn if the presence of tuberculosis is confirmed by the isolation of M. bovis, M. caprae or M. tuberculosis on laboratory examination.

2. Measures in place[(b)]

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The control program/strategies in place
The whole cattle population is continuously monitored for bovine tuberculosis on a yearly basis by the intradermal tuberculine tests and by post-mortem inspections.

Recent actions taken to control the zoonoses

New national legislation entered into force at the end of 2019.
New guidelines have been issued by the National Food Chain Safety Office about how to carry out the tuberculin test in cattle herds taking into consideration the fals positive or interference reactions as well as the data collection, and reporting by the regional authorities. Veterinarians who perform intradermal tuberculine tests are required to take a course with both theoretical and practical elements.

Guidances made by National Food Chain Safety Office (how to carry out tuberculin tests; guide on non-specific reactions; guide for sending animals to diagnostical slaughter, guide on registrations, etc.) https://portal.nebih.gov.hu/-/nebih-utmutatok-a-szarvasmarha-gumokorrol

Measures in case of the positive findings or single cases
When an animal is considered to be a positive reactor in the intradermal tests, it is removed from the herd and slaughtered. The post-mortem, laboratory and epidemiological examinations shall be carried out. The status of the herd will remain suspended until all laboratory examinations have been completed. If the presence of tuberculosis is not confirmed, the suspension of the officially tuberculosis-free status may be lifted following a test of all animals over six weeks of age with negative results at least 60 days after the removal of the reactor animal. The officially tuberculosis -free status of the herd have to be withdrawn if the presence of tuberculosis is confirmed by the isolation of M. bovis. M- caprae or M. tuberculosis on laboratory examination.The district chief veterinarian may withdraw the tuberculosis-free status of the herdif the conditions for retention of the officially free status are not complied with, or classical lesions of tuberculosis are seen at post-mortem examination,an epidemiological enquiry establishes the likelihood of infection,or it is deemed necessary to control of bovine tuberculosis in the herd for any other reason.

3. Notification system in place to the national competent authority(c)

Yes. Bovine tuberculosis is compulsory notifiable by Decree No 113/2008 (VIII. 30.) of the Ministry of Agriculture and Rural Development (MARD) on notification of animal diseases. The detailed rules regarding bovine tuberculosis are laid down by the Decree No. 56/2019 (XII. 12.) of the Minister of Agriculture.

4. Results of investigations and national evaluation of the situation, the trends (d) and sources of infection(e)

Hungary is officially free of bovine tuberculosis (2021/620 IR). In some years sporadic cases (M caprae) are reported.

* **For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**
(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.
(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.
(c): Mandatory: Yes/No.
(d): Minimum five years.
(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

Description of Monitoring/Surveillance/Control programmes system*: Brucella abortus

1. Monitoring/Surveillance/Control programmes system(a)

Sampling strategy
Yearly blood sampling of the Hungarian cattle population, as well as case-by-case testing of animals moved from one
herd to another, a system of checking abortions and irregular parturition has been maintained.

Frequency of the sampling
The whole cattle population in Hungary is subject to regular checks. Investigation of abortion and related cases is the key point of the
system. Yearly testing of bovine herds had taken place on animals over 24 months until the application of the AHL. After 21st April, 2021 all animals over 12 months are tested in all bovine herd. .

Methods of sampling (description of sampling techniques)
Blood or milk samples are taken at farm. In case of abortion, the aborted fetus, its chorions and a blood sample from the aborted
cattle shall be sent to the laboratory.

Diagnostic methods
The test used for screening is I-ELISA. In case of positive result, CFT (complement fixation test) and RBT (rose bengal test) is performed.

Case definition
An animal is considered to be infected with B. abortus, when:
- it shows clinical signs of the disease or pathological lesions can be detected
; and the disease can be detected by bacteriological or repeated serological tests
-if it shows clinical signs or pathological lesions and originates from an infected herd
- shows no clinical signs but the disease agent is detected
-shows no clinical signs but repeated serological tests have detected brucellosis without doubt

2. Measures in place[(b)]

Vaccination policy
Preventive vaccination against B. abortus is prohibited in the whole territory of Hungary.

Control program/mechanisms
Recent actions taken to control the zoonoses
Continuous monitoring of bovine herds and investigation of aborted fetuses as well as pre-movement checks are continued. A 3-year eradication programme required by the AHL and related acts is submitted to the Commission in May, 2021.

Measures in case of the positive findings or single cases
Movement restriction is maintained in the infected herd, the status of the herd is suspended or withdrawn.
Infected animals are to be killed as soon as possible but not later than five days. An eradication plan must be made or if the number of seropositive animals is more than 5% in the herd, the whole herd must be eradicated.

3. Notification system in place to the national competent authority[(c)]

Yes. Investigation of cases of abortion is compulsory. In case of abortion or irregular parturition, the veterinarian in charge has to send a set of samples for further laboratory examination. Until thorough clarification of the case, the animal is kept separated and, if necessary, repeatedly tested.

4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]

Since 1985 B. abortus has not been isolated.

* **For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

Description of Monitoring/Surveillance/Control programmes system*: Brucella melitensis

1. Monitoring/Surveillance/Control programmes system[(a)]

Sampling strategy

Frequency of the sampling
Approximately 5% of the ovine and caprine population has been sampled and tested for B. melitensis until the application of the AHL.

Type of specimen taken
Blood

Methods of sampling (description of sampling techniques)
Blood samples are taken at farm.

Case definition
See above, same as for cattle

2. Measures in place[(b)]

Vaccination policy
Vaccination or treatment is banned in Hungary.

Control program/mechanisms
The control program/strategies in place
Hungary is free of B. melitensis. However, monitoring of ovine and caprine populations is continuously done.

Measures in case of the positive findings or single cases

Movement restriction is maintained in the infected herd, The status of the herd is suspended or withdrawn.
Infected animals are to be killed as soon as possible but not later than five days. An eradication plan must be made or if the number of seropositive animals is more than 5% in the herd, the whole herd must be eradicated.

3. Notification system in place to the national competent authority[(c)]

Yes. Ovine and caprine brucellosis (B. melitensis) are compulsorily notifiable since 1 January 1982.

4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]

No evidence of infection with B. melitensis was ever found. Hungary is officially free of the disease.

* **For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

Description of Monitoring/Surveillance/Control programmes system*: Trichinella in animal - Pigs - food sample

1. Monitoring/Surveillance/Control programmes system[(a)]

Sampling strategy
Trichinella sampling and testing is mandatory for all pigs intended to be placed on the market.

Frequency of the sampling
Every slaughtered animal is sampled

Type of specimen taken
Diaphragm muscle

Methods of sampling (description of sampling techniques)
Methods specified in Regulation 1375/2015/EU

Case definition
Animal with one or more Trichinella larva in the official examination.

Diagnostic/analytical methods used
Artificial digestion method of collective samples.

2. Measures in place[(b)]

Vaccination policy
None.

Control program/mechanisms
The control program/strategies in place

See above.

Measures in case of the positive findings or single cases
Positive cases are considered not to be eligible for human consumption.

* **For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

Description of Monitoring/Surveillance/Control programmes system*: Trichinella in animal - Solipeds, domestic - horses - food sample

1. Monitoring/Surveillance/Control programmes system[(a)]

Sampling strategy
Trichinella testing is mandatory, all animal is sampled.

Frequency of the sampling
Every slaughtered animal is sampled

Type of specimen taken
Diaphragm muscle

Methods of sampling (description of sampling techniques)
1375/2015/EU regulation

Case definition
Animal with one or more Trichinella larva in the official examination

Diagnostic/analytical methods used
Artificial digestion method of collective samples

2. Measures in place[(b)]

Vaccination policy
None.

Measures in case of the positive findings or single cases
Positive cases are considered not to be eligible for human consumption.

4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]

Trichinella infection has never been detected in horses in Hungary.

* **For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

Description of Monitoring/Surveillance/Control programmes system*: Lyssavirus (rabies) in animal - All animals - wild - animal sample

1. Monitoring/Surveillance/Control programmes system[(a)]

Sampling strategy
1. Passive monitoring (surveillance) of dead foxes and other indicator animals (suspect animals as well as road kills) in the whole territory of the country
2. Monitoring to control the effectiveness of oral vaccinationin the vaccinated area.

Frequency of the sampling
1. Passive monitoring (surveillance): during the whole year
2. Monitoring of oral vaccination (ORV): In the dedicated period of the year in a definite number: sampling period starts 30 day after the completion of each vaccination campaign and the minimum number of foxes to be sampled is 4 foxes/100 km2/year (2 foxes/100 km2/campaign)

Type of specimen taken
1. Passive monitoring (surveillance): whole carcass or head in case of large animals. Brain tissue is taken in the laboratory with specific regard to the predilection sites.
2. Active monitoring of ORV: Whole fox and jackal carcasses are submitted to the veterinary authority by hunters in the framework of monitoring of OV. Brain tissue is taken in the laboratory with specific regard to the predilection sites to rule out infection. Transversal tooth section is performed to detect presence of tetracycline, and ELISA test is carried out to detect antibodies from blood samples.

Methods of sampling (description of sampling techniques)
Whole carcasses of healthy shot foxes, suspect foxes or suspect individuals of other species are submitted to the laboratory. Brain tissue sample is taken in the laboratory from all categories. Mandible and blood sample is taken in the laboratory from foxes shot in the framework of monitoring effectiveness of OV.

Case definition
Rabid animal: an animal in which, with laboratory examinations, rabies had been confirmed undoubtedly. Suspect animals: 1. animals showing clinical signs of rabies; 2. animals not showing clinical signs of rabies but injured by a rabid or a rabies suspected animal; 3. all wild mammals that are showing abnormal behavior or attack humans. Potentially rabies infected animal: animals that had possibly had contacted rabid or suspect animals within 90 days. Animals with a concern for rabies: 1. all mammals not showing clinical signs or abnormal behavior and not in contact with rabid or suspect animals, but attacking or injuring humans, 2. as well as dogs without a valid rabies vaccination.

Diagnostic/analytical methods used
Direct immunfluorescence (fluorescent antibody test -FAT) of brain imprints with a monovalent anti-nucleocapside conjugate is the primary diagnostic test applied. Furthermore, PCR and isolation of the virus in neuroblastoma cells cultures is performed in some cases (eg. from animals causing human

injury). All FAT positive results are confirmed by 1. qRT-PCR (Picard-Meyer et. al., 2004.) with Rotor-Gene SYBR-Green RT-PCR kit QIAGEN 2. RTCIT (OIE Manual Chapter 2.1.13. Rabies (NB: Version adopted in May 2013) with N2A cells and Fujirebio monoclonal globulins (FDI) The inconclusive results are examined beside these above mentioned methods with 3. IHC ("in house" developed) 4. MIT (OIE Manual Chapter 2.1.13. Rabies (NB: Version adopted in May 2013) 5. RT-PCR (Heaton et. al., 1997) 6. Sequencing (Sanger et. al., 1977)
In the framework of the monitoring of efficiency of the oral immunization of foxes the following tests are performed: - direct immunfluorescence (fluorescent antibody test -FAT) of imprints of the brain - Transversal tooth section – test for the presence of tetracycline - serological (ELISA) test to detect antibodies

2. Measures in place[(b)]

Vaccination policy
Vaccination area in 2020: South and East border zone of Hungary, 41.970 km2 (50 km zone along the borders to countries that are not free from rabies). Two vaccination campaigns per year (April and October)

Control program/mechanisms
The control program/strategies in place
Decree No 81/2002 of the MARD on the animal health issues of the protection against zoonotic diseases
Decree No 164/2008 of the MARD on detailed rules of the protection against rabies

Recent actions taken to control the zoonoses

Measures in case of the positive findings or single cases
Tracing human contacts, animal contacts. Vaccination of cats and farm animals upon the decision of the veterinary authority. Emergency ring vaccination of wild foxes.

3. Notification system in place to the national competent authority[(c)]

Yes. Rabies is a notifiable disease in Hungary according to Decree No 113/2008 of Ministry of Agriculture and Rural Development (MARD) on the order of the notification of animal diseases Moreover, rules regarding the notification of rabies suspected animals are detailed in the Decree No 164/2008 of the MARD on detailed rules of the protection against rabies.

4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]

One rabies case (RABV, fox) in 2016 and one rabies case (RABV, fox) in 2017.

National evaluation of the recent situation, the trends and sources of infection
Genom sequencing was performed in all cases of the past years, identifying the closest related strains. The strain was similar to some eastern-European isolates, however, the way of introduction of rabies into the country could not be proved.

Results of the investigation
Investigations of the human contacts with positive cases
All positive cases shall be reported to the human health service according to national legislation. Decision about immunization of a person in contact with a rabies positive animal is the competence of the human health authorities.

5. Additional information

Cooperation with Ukraine to conduct oral vaccination of foxes in a 50 km wide buffer zone in Ukraine along the border since 2015. In 2019 and 2020, two vaccination campaigns in each year were performed successfully in the buffer zone.

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(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

Description of Monitoring/Surveillance/Control programmes system*: Lyssavirus (rabies) in animal - Dogs - animal sample

1. Monitoring/Surveillance/Control programmes system[(a)]

Sampling strategy
In case of dogs and other domestic animals, only suspect animals are sampled. According to legislation, animals showing symptoms of rabies (=suspect animals) are killed and tested for rabies. Animals not showing the clinical signs of rabies but contacted and injured by a rabid or a rabies suspected animal are considered as suspects as well and are killed and tested for rabies (or in certain circumstances, when they have been vaccinated against rabies earlier and that vaccination is still valid, they can be put under official observation for 90 days.) Animals that had possibly had contacted rabid or suspect animals are put under official observation for 90 days. Animals not showing clinical signs or abnormal behavior but causing human injuries, as well as dogs without a valid rabies vaccination, are put under official observation for 14 days. If the animal perishes during the time of official observation, it will be sampled and tested for rabies.

Frequency of the sampling
Passive surveillance – Sampling only in case of suspicion (see point 1.).

Type of specimen taken
Whole carcass / head / brain tissue.

Methods of sampling (description of sampling techniques)
Whole carcasses of suspect dogs or other species are submitted to the laboratory. Brain tissue sample is taken in the laboratory with specific regard to the predilection sites.

Case definition
Rabid animal: an animal in which, with laboratory examinations, rabies had been confirmed undoubtedly. Suspect animals: 1. animals showing clinical signs of rabies; 2. animals not showing clinical signs of rabies but injured by a rabid or a rabies suspected animal; 3. all wild mammals that are showing abnormal behavior or attack humans. Potentially rabies infected animal: animals that had possibly had contacted rabid or suspect animals within 90 days. Animals with a concern for rabies: 1. all mammals not showing clinical signs or abnormal behavior and not in contact with rabid or suspect animals, but attacking or injuring humans, 2. as well as dogs without a valid rabies vaccination.

Diagnostic/analytical methods used
Direct immunfluorescence (fluorescent antibody test -FAT) of brain imprints with a monovalent anti-nucleocapside conjugate is the primary diagnostic test applied. Furthermore, PCR and isolation of the virus in neuroblastoma cells cultures is performed in some cases (eg. from animals causing human injury). All FAT positive results are confirmed by 1. qRT-PCR (Picard-Meyer et. al., 2004.) with Rotor-Gene SYBR-Green RT-PCR kit QIAGEN 2. RTCIT (OIE Manual Chapter 2.1.13. Rabies (NB: Version adopted in May 2013) with N2A cells and Fujirebio monoclonal globulins (FDI) The inconclusive results are examined beside these above mentioned methods with 3. IHC ("in house" developed) 4. MIT (OIE Manual Chapter 2.1.13. Rabies (NB: Version adopted in May 2013) 5. RT-PCR (Heaton et. al., 1997) 6. Sequencing (Sanger et. al., 1977)

2. Measures in place[(b)]

Vaccination policy
Obligatory vaccination of dogs, once a year. According to national legislation in force, dogs shall be vaccinated against rabies two times between 3 months of age and under 1 year of age with monovalent vaccine. After 1 year of age vaccination shall be repeated annually. Vaccination of cats is recommended. In case of an outbreak, taking into consideration the epidemiological situation, the veterinary authority can order obligatory vaccination of cats and farm animals.

Other preventive measures than vaccination in place
In the framework of an awareness campaign, leaflets have been produced and distributed, explaining the importance of rabies and describing the symptoms of the disease and the way of reporting the suspicion of the disease to the veterinary services. A website has been developed as well, specifically dedicated for rabies, in order to provide information about the disease to the public (veszettsegmentesites.hu).

The control program/strategies in place
Decree No 81/2002 of the MARD on the animal health issues of the protection against zoonotic diseases; Decree No 164/2008 of the MARD on detailed rules of the protection against rabies

Measures in case of the positive findings or single cases
Tracing human contacts, animal contacts. Vaccination of cats and farm animals upon the decision of the veterinary authority. Emergency ring vaccination of wild foxes.

3. Notification system in place to the national competent authority[(c)]

Yes. Rabies is a notifiable disease in Hungary according to Decree No 113/2008 of Ministry of Agriculture and Rural Development (MARD) on the order of the notification of animal diseases Moreover, rules regarding the notification of rabies suspected animals are detailed in the Decree No 164/2008 of the MARD on detailed rules of the protection against rabies.

4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]

Last RABV case in dog in 2014. Two rabid goats (in the same farm, at the same time) in 2017. No classical rabies cases in the country in 2018-2020.

National evaluation of the recent situation, the trends and sources of infection
Genom sequencing was performed in all cases of the past years, identifying the closest related strains. The strain was similar to some eastern-European isolates, however, the way of introduction of rabies into the country could not be proved.

Investigations of the human contacts with positive cases

All positive cases shall be reported to the human health service according to national legislation. Decision about immunization of a person in contact with a rabies positive animal is the competence of the human health authorities

* **For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

Institutions and laboratories involved in antimicrobial resistance monitoring and reporting

- National Food Chain Safety Office, Veterinary Diagnostic Directorate (NRL for AMR)
- National Food Chain Safety Office, Food and Feed Safety Laboratory Directorate (NRL for *Salmonella)*

Short description of the institutions and laboratories involved in data collection and reporting

1. General Description of Antimicrobial Resistance Monitoring; Indicator E. coli from domestic broilers- caecal samples

1. General description of sampling design and strategy

The number of samples per animal population was planned for the specific ESBL/AmpC monitoring (300).

Sampling technique: Caecum should be cut out and placed into a plastic bag, closed properly and cooled +2 ─ +8 °C, transported within 48h.

- stage of sampling: Samples were taken at slaughterhouses.
- type of sample: Caecal samples were taken from domestic broilers.
- sampler: competent authorities

Frequency of sampling: Every month, evenly distributed over the whole year
Procedure of selection of isolates for susceptibility testing: In ESBL/AmpC/Carbapenemase monitoring program: *E. coli* isolates growing on Cefotaxim-McConkey plates were submitted to MIC-determination.
Randomized selection of 200 commensal *E. coli* isolations for general susceptibility tests.

Method used for collecting data: Along with the samples, a paper form submitted containing the data regarding sampling and origin of sample and the code of epidemiological unit (area) for identification.

2. Stratification procedure per animal population and food category

The sampling was stratified at slaughterhouse level based on the annual production data of the previous year (2019). Selection of slaughterhouses was made according to their production starting with higher throughput. So, bigger slaughterhouses produced together above the 60% of the total national production were involved into sampling. The number of samples had distributed per slaughterhouse proportionally to the annual throughput of the slaughterhouse.

3. Randomisation procedure per animal population and food category

Sampling days were definied for the availability of courier service by authority. After exclusion of epidemiological units already sampled in that year, samples were randomly collected at slaughterhouses on the day suitable for submission of samples in required time frame with even distribution by date. Samples were randomly collected from different suitable epidemiological units manually.

The random sampling was stratified at one level. The central authority (NFCSO) had distributed the sample numbers based on the annual slaughter capacity per slaughterhouses proportionally to the annual throughput of the slaughterhouse starting with the higher producer.

4. Analytical method used for detection and confirmation

Identification of bacterial isolates is performed by chromogenic media (Coliform agar purchased from Biolab Zrt.), indole and oxidase tests and when necessary additional API ID32E biochemical tests.

Selective isolation of ESBL/AmpC/Carbapenemase-producing *E. coli* is performed by MacConkey agar plates supplemented with 1 mg/L cefotaxime which is purchased from Biolab Zrt.

Selective isolation of Carbapenemase-producing *E. coli* is performed by chromID CarbaSmart (purchased from bioMerieux Hungária Kft.)

5. Laboratory methodology used for detection of antimicrobial resistance

Broth microdilution using the Thermo Scientific™Sensititre™ SWIN™ Software System.

Antimicrobals were ampicillin, azithromycin, cefotaxime, ceftazidime, chloramphenicol, ciprofloxacin, colistin, gentamicin, meropenem, nalidixic acid, sulphamethoxazole, tetracycline, tigecycline and trimethoprim (first panel). Cefepime, cefotaxime, cefotaxime+clavulanic acid, cefoxitin, ceftazidime, ceftazidime+clavulanic acid, ertapenem, imipenem, meropenem and temocillin for extended susceptibility testing (second panel).

Results were interpreted using the EFSA published epidemiological cut-off (ECOFF) values.

2. General Description of Antimicrobial Resistance Monitoring; Indicator E. coli from fattening turkeys- caecal samples

1. General description of sampling design and strategy

The number of samples per animal population was planned for the specific ESBL/AmpC monitoring (300).

Sampling technique: Caecum should be cut out and placed into a plastic bag, closed properly and cooled +2 ─ +8 °C, transported within 48h.

- stage of sampling: Samples were taken at slaughterhouses.
- type of sample: Caecal samples were taken from domestic fattening turkeys.
- sampler: competent authorities

Frequency of sampling: Every month, evenly distributed over the whole year
Procedure of selection of isolates for susceptibility testing: In ESBL/AmpC/Carbapenemase monitoring program: *E. coli* isolates growing on Cefotaxim-McConkey plates were submitted to MIC-determination.
Randomized selection of 200 commensal *E. coli* isolations for general susceptibility tests.

Method used for collecting data: Along with the samples, a paper form submitted containing the data regarding sampling and origin of sample and the code of epidemiological unit (area) for identification.

2. Stratification procedure per animal population and food category

The sampling was stratified at slaughterhouse level based on the annual production data of the previous year (2019). Selection of slaughterhouses was made according to their production starting with higher throughput. So, bigger slaughterhouses produced together above the 60% of the total national production were involved into sampling. The number of samples had distributed per slaughterhouse proportionally to the annual throughput of the slaughterhouse.

3. Randomisation procedure per animal population and food category

Sampling days were definied for the availability of courier service by authority. After exclusion of epidemiological units already sampled in that year, samples were randomly collected at slaughterhouses on the day suitable for submission of samples in required time frame with even distribution by date. Samples were randomly collected from different suitable epidemiological units manually.

The random sampling was stratified at one level. The central authority (NFCSO) had distributed the sample numbers based on the annual slaughter capacity per slaughterhouses proportionally to the annual throughput of the slaughterhouse starting with the higher producer.

4. Analytical method used for detection and confirmation

Identification of bacterial isolates is performed by chromogenic media (Coliform agar purchased from Biolab Zrt.), indole and oxidase tests and when necessary additional API ID32E biochemical tests.

Selective isolation of ESBL/AmpC/Carbapenemase-producing *E. coli* is performed by MacConkey agar plates supplemented with 1 mg/L cefotaxime which is purchased from Biolab Zrt.

Selective isolation of Carbapenemase-producing *E. coli* is performed by chromID CarbaSmart (purchased from bioMerieux Hungária Kft.)

5. Laboratory methodology used for detection of antimicrobial resistance

Broth microdilution using the Thermo Scientific™Sensititre™ SWIN™ Software System.

Antimicrobals were ampicillin, azithromycin, cefotaxime, ceftazidime, chloramphenicol, ciprofloxacin, colistin, gentamicin, meropenem, nalidixic acid, sulphamethoxazole, tetracycline, tigecycline and trimethoprim (first panel). Cefepime, cefotaxime, cefotaxime+clavulanic acid, cefoxitin, ceftazidime, ceftazidime+clavulanic acid, ertapenem, imipenem, meropenem and temocillin for extended susceptibility testing (second panel).

Results were interpreted using the EFSA published epidemiological cut-off (ECOFF) values.

3. General Description of Antimicrobial Resistance Monitoring; Indicator E. coli from domestic broilers-meat samples

1. General description of sampling design and strategy

The number of samples of each food category (broiler chicken fresh meat) was planned for the specific ESBL/AmpC monitoring (300 samples per a year).

Sampling technique:
Manual pick up of samples from the refrigerator in the shop.

- stage of sampling: Samples were taken at retail outlets.
- type of sample: Fresh meat from broilers
- sampler: competent authorities

Frequency of sampling: Every month
Procedure of selection of isolates for susceptibility testing: *E. coli* isolates growing on Cefotaxim-McConkey plate were submitted to MIC-determination.

Method used for collecting data: Along with the samples, a paper form submitted containing the data regarding sampling and origin of sample and the code of epidemiological unit (area) for identification.

2. Stratification procedure per animal population and food category

The random sampling was stratified geographically by counties based on the human population according to NUTS3 level.

3. Randomisation procedure per animal population and food category

Samples were randomly collected at retail with even distribution of the date. Retail shops were chosen randomly excluding the shops visited earlier.

4. Analytical method used for detection and confirmation

Identification of bacterial isolates is performed by chromogenic media (Coliform agar purchased from Biolab Zrt.), indole and oxidase tests and when necessary additional API ID32E biochemical tests.

Selective isolation of ESBL/AmpC/Carbapenemase-producing *E. coli* is performed by MacConkey agar plates supplemented with 1 mg/L cefotaxime which is purchased from Biolab Zrt.

Selective isolation of Carbapenemase-producing *E. coli* is performed by chromID CarbaSmart (purchased from bioMerieux Hungária Kft.)

5. Laboratory methodology used for detection of antimicrobial resistance

Broth microdilution using the Thermo Scientific™Sensititre™ SWIN™ Software System. Antimicrobals were ampicillin, azithromycin, cefotaxime, ceftazidime, chloramphenicol, ciprofloxacin, colistin, gentamicin, meropenem, nalidixic acid, sulphamethoxazole, tetracycline, tigecycline and trimethoprim (first panel). Cefepime, cefotaxime, cefotaxime+clavulanic acid, cefoxitin, ceftazidime, ceftazidime+clavulanic acid, ertapenem, imipenem, meropenem and temocillin for extended susceptibility testing (second panel).

Results were interpreted using the EFSA published epidemiological cut-off (ECOFF) values.

4. General Description of Antimicrobial Resistance Monitoring; Campylobacter jejuni from domestic broilers-caecal samples

1. General description of sampling design and strategy

The number of caecal samples per animal population was planned to achieve the required number of Campylobacter jejuni. Prevalence was estimated based on the results of 2018.

Sampling technique:
Caecum should be cut and dropped into a plastic bag, cooled +2-+8°C, transported within 48h

- stage of sampling: Samples were taken at slaughterhouses.
- type of sample: Caecal samples were taken from domestic broilers.
- sampler: competent authorities

Frequency of sampling: Every month, evenly distributed over the whole year
Procedure of selection of isolates for susceptibility testing: We used PCR method for submit the Campylobacter jejuni isolates to MIC determination.

Method used for collecting data: Along with the samples, a paper form submitted containing the data regarding sampling and origin of sample and the code of epidemiological unit (area) for identification.

2. Stratification procedure per animal population and food category

The sampling was stratified at slaughterhouse level based on the annual production data of the previous year (2019). Selection of slaughterhouses was made according to their production starting with higher throughput. So, bigger slaughterhouses produced together above the 60% of the total national production were involved into sampling. The number of samples had distributed per slaughterhouse proportionally to the annual throughput of the slaughterhouse.

3. Randomisation procedure per animal population and food category

Sampling days were definied for the availability of courier service by authority. After exclusion of epidemiological units already sampled in that year, samples were randomly collected at slaughterhouses on the day suitable for submission of samples in required time frame with even distribution by date. Samples were randomly collected from different suitable epidemiological units manually.

The random sampling was stratified at one level. The central authority (NFCSO) had distributed the sample numbers based on the annual slaughter capacity per slaughterhouses proportionally to the annual throughput of the slaughterhouse starting with the higher producer.

4. Analytical method used for detection and confirmation

Isolation of bacterial isolates is performed by CCDA agar which is purchased from Biolab Zrt., and identification of Campylobacter isolates are performed by PCR method, we used also oxidase tests for identification by biochemical tests.

5. Laboratory methodology used for detection of antimicrobial resistance

Broth microdilution using the Thermo Scientific™Sensititre™ SWIN™ Software System.
Antimicrobals were ciprofloxacin, erythromycin, gentamicin, nalidixic acid, streptomycin, tetracycline

Results were interpreted using the EFSA published epidemiological cut-off (ECOFF) values.

5. General Description of Antimicrobial Resistance Monitoring; Camplyobacter jejuni from fattening turkey- caecal samples

1. General description of sampling design and strategy

The number of caecal samples per animal population was planned to achieve the required number of Campylobacter jejuni. Prevalence was estimated based on the results of 2018.

Sampling technique:
Caecum should be cut and dropped into a plastic bag, cooled +2-+8°C, transported within 48h

- stage of sampling: Samples were taken at slaughterhouses.
- type of sample: Caecal samples were taken from fattening turkey.
- sampler: competent authorities

Frequency of sampling: Every month, evenly distributed over the whole year.
Procedure of selection of isolates for susceptibility testing: We used PCR method for submit the Campylobacter jejuni isolates to MIC determination.

Method used for collecting data: Along with the samples, a paper form submitted containing the data regarding sampling and origin of sample and the code of epidemiological unit (area) for identification.

2. Stratification procedure per animal population and food category

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Isolation of bacterial isolates is performed by CCDA agar which is purchased from Biolab Zrt. and identification of Campylobacter isolates are performed by PCR method, we used oxidase tests also for identification by biochemical test.

5. Laboratory methodology used for detection of antimicrobial resistance

Broth microdilution using the Thermo Scientific™Sensititre™ SWIN™ Software System.

Antimicrobals were ciprofloxacin, erythromycin, gentamicin, nalidixic acid, streptomycin, tetracycline.

Results were interpreted using the EFSA published epidemiological cut-off (ECOFF) values.

6. General Description of Antimicrobial Resistance Monitoring; Salmonella – domestic broilers, fattening turkeys, laying hens, domestic broiler and fattening turkey neck skin

1. General description of sampling design and strategy

Sampling technique:

- stage- and frequency of sampling, type of sample: according to regulations
- sampler: competent authorities

Procedure of selection of isolates for susceptibility testing: Randomized
Method used for collecting data: electronic datasheets

2. Stratification procedure per animal population and food category

The NRL-*Salmonella* collects Salmonella isolates for typing from the whole country according to the regulations.

3. Randomisation procedure per animal population and food category
The required No. of *Salmonella* isolates were randomly selected from the actual monthly isolates collected by the NRL-Salmonella to obtain the defined 170 isolates/year in an even distribution. All 170 isolates submitted to the NRL-AR were included into antimicrobial susceptibility testing. If the No. of collected isolates from a special matrix were lower than required, all available isolates were submitted for susceptibility testing.
4. Analytical method used for detection and confirmation
Isolates were collected and serotyped at the NRL for *Salmonella* under the Directorate of Food Chain Safety Laboratory. These isolates are confirmed on selective Rambach agar plates (purchased from VWR International Kft.) and additional agglutination tests by the NRL-AR.
5. Laboratory methodology used for detection of antimicrobial resistance
Broth microdilution using the Thermo Scientific™Sensititre™ SWIN™ Software System. Antimicrobals included in monitoring for *Salmonella sp*. were ampicillin, azithromycin, cefotaxime, ceftazidime, chloramphenicol, ciprofloxacin, colistin, gentamicin, meropenem, nalidixic acid, sulphamethoxazole, tetracycline, tigecycline and trimethoprim (first panel). Cefepime, cefotaxime, cefotaxime+clavulanic acid, cefoxitin, ceftazidime, ceftazidime+clavulanic acid, ertapenem, imipenem, meropenem and temocillin for extended susceptibility testing (second panel). Results were interpreted using the EFSA published epidemiological cut-off (ECOFF) values.