European Food Safety Authority

# ZOONOSES MONITORING

## Romania

# TRENDS AND SOURCES OF ZOONOSES AND ZOONOTIC AGENTS IN FOODSTUFFS, ANIMALS AND FEEDINGSTUFFS

including information on foodborne outbreaks, antimicrobial resistance in zoonotic and indicator bacteria and some pathogenic microbiological agents

## IN 2018

# PREFACE

This report is submitted to the European Commission in accordance with Article 9 of Council Directive 2003/99/ EC*. The information has also been forwarded to the European Food Safety Authority (EFSA).

The report contains information on trends and sources of zoonoses and zoonotic agents in Romania during the year 2018.

The information covers the occurrence of these diseases and agents in animals, foodstuffs and in some cases also in feedingstuffs. In addition the report includes data on antimicrobial resistance in some zoonotic agents and indicator bacteria as well as information on epidemiological investigations of foodborne outbreaks.
Complementary data on susceptible animal populations in the country is also given. The information given covers both zoonoses that are important for the public health in the whole European Union as well as zoonoses, which are relevant on the basis of the national epidemiological situation.
The report describes the monitoring systems in place and the prevention and control strategies applied in the country. For some zoonoses this monitoring is based on legal requirements laid down by the European Union legislation, while for the other zoonoses national approaches are applied.

The report presents the results of the examinations carried out in the reporting year. A national evaluation of the epidemiological situation, with special reference to trends and sources of zoonotic infections, is given. Whenever possible, the relevance of findings in foodstuffs and animals to zoonoses cases in humans is evaluated.
The information covered by this report is used in the annual European Union Summary Reports on zoonoses and antimicrobial resistance that are published each year by EFSA.

The national report contains two parts: tables summarising data reported in the Data Collection Framework and the related text forms. The text forms were sent by email as pdf files and they are incorporated at the end of the report.

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* Directive 2003/ 99/ EC of the European Parliament and of the Council of 12 December 2003 on the monitoring of zoonoses and zoonotic agents, amending Decision 90/ 424/ EEC and repealing Council Directive 92/ 117/ EEC, OJ L 325, 17.11.2003, p. 31

List of Contents

# ANIMAL POPULATION TABLES

## Table Susceptible animal population

| | | Population | | | |
|---|---|---|---|---|---|
| **Animal species** | **Category of animals** | **holding** | **animal** | **slaughter animal (heads)** | **herd/flock** |
| Cattle (bovine animals) | Cattle (bovine animals) | 419,668 | 1,994,917 | 247,654 | |
| Gallus gallus (fowl) | Gallus gallus (fowl) - breeding flocks, unspecified | 42 | 3,395,149 | | 704 |
| | Gallus gallus (fowl) - broilers - before slaughter | 279 | 301,087,135 | 261,690,492 | 12,529 |
| | Gallus gallus (fowl) - laying hens | 248 | 14,430,338 | | 819 |
| Goats | Goats | 77,983 | 1,660,972 | 30,086 | |
| | Goats - animals over 1 year | | 1,658,523 | | |
| | Goats - animals under 1 year | | 2,449 | | |
| Pigs | Pigs | 454,423 | 1,920,166 | 4,834,380 | |
| Sheep | Sheep | 158,704 | 11,845,182 | 935,631 | |
| | Sheep - animals over 1 year | | 11,824,740 | | |
| | Sheep - animals under 1 year (lambs) | | 20,442 | | |
| Solipeds, domestic | Solipeds, domestic | 280,861 | 338,026 | 29,958 | |
| Turkeys | Turkeys | 17 | 1,382,719 | 1,450,320 | 260 |

## DISEASE STATUS TABLES

### Table Bovine brucellosis in countries and regions that do not receive Community co-financing for eradication programme

| Region | Number of animals serologicall y tested under investigatio ns of suspect cases | Number of suspended herds under investigatio ns of suspect cases | Number of seropositiv e animals under investigatio ns of suspect cases | Number of animals positive to BST under investigatio ns of suspect cases | Number of animals positive in microbiolog ical testing under investigatio ns of suspect cases | Number of herds with status officially free | Number of infected herds | Total number of animals | Number of herds tested under surveillance | Number of animals tested under surveillance | Total number of herds | Number of infected herds tested under surveillance | Number of herds tested under surveillance by bulk milk | Number of animals or pools tested under surveillance by bulk milk | Number of infected herds tested under surveillance by bulk milk | Number of notified abortions whatever cause under investigatio ns of suspect cases | Number of isolations of Brucella abortus under investigatio ns of suspect cases | Number of abortions due to Brucella infection under investigatio ns of suspect cases | Number of animals tested by microbiolog y under investigatio ns of suspect cases |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| ROMANIA | 59 | 1 | 3 | 0 | 0 | 466,709 | 0 | 1,904,116 | 466,321 | 1,161,513 | 469,200 | 0 | 320 | 56,728 | 0 | 36 | 0 | 0 | 2 |
| Bihor | 14 | 0 | 0 | 0 | 0 | 14,391 | 0 | 71,030 | 14,391 | 44,159 | 14,391 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 |
| Bistriţa-Năsăud | 0 | 0 | 0 | 0 | 0 | 14,465 | 0 | 75,845 | 14,437 | 48,105 | 14,465 | 0 | 28 | 3,858 | 0 | 1 | 0 | 0 | 0 |
| Cluj | 0 | 0 | 0 | 0 | 0 | 9,957 | 0 | 62,571 | 9,924 | 37,545 | 9,957 | 0 | 33 | 2,859 | 0 | 4 | 0 | 0 | 0 |
| Maramureş | 0 | 0 | 0 | 0 | 0 | 37,542 | 0 | 79,003 | 37,537 | 54,376 | 37,542 | 0 | 5 | 854 | 0 | 0 | 0 | 0 | 0 |
| Satu Mare | 3 | 0 | 0 | 0 | 0 | 7,587 | 0 | 43,150 | 7,587 | 24,969 | 7,587 | 0 | 0 | 0 | 0 | 3 | 0 | 0 | 0 |
| Sălaj | 1 | 0 | 0 | 0 | 0 | 7,139 | 0 | 29,837 | 7,139 | 20,001 | 7,139 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Alba | 0 | 0 | 0 | 0 | 0 | 13,452 | 0 | 67,497 | 13,422 | 35,879 | 13,452 | 0 | 49 | 5,526 | 0 | 0 | 0 | 0 | 0 |
| Braşov | 1 | 0 | 1 | 0 | 0 | 9,405 | 0 | 66,216 | 9,405 | 43,983 | 9,405 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 |
| Covasna | 0 | 0 | 0 | 0 | 0 | 5,277 | 0 | 45,561 | 5,277 | 30,225 | 5,277 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Harghita | 0 | 0 | 0 | 0 | 0 | 15,123 | 0 | 86,990 | 15,123 | 57,162 | 15,123 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 |
| Mureş | 0 | 0 | 0 | 0 | 0 | 8,271 | 0 | 75,977 | 8,271 | 47,466 | 8,271 | 0 | 0 | 0 | 0 | 2 | 0 | 0 | 0 |
| Sibiu | 15 | 0 | 0 | 0 | 0 | 5,035 | 0 | 47,948 | 5,028 | 26,717 | 5,035 | 0 | 7 | 590 | 0 | 3 | 0 | 0 | 0 |
| Bacău | 0 | 0 | 0 | 0 | 0 | 17,486 | 0 | 47,231 | 17,333 | 33,652 | 17,486 | 0 | 6 | 2,000 | 0 | 0 | 0 | 0 | 0 |
| Botoşani | 0 | 0 | 0 | 0 | 0 | 25,536 | 0 | 102,290 | 25,536 | 57,787 | 25,536 | 0 | 10 | 473 | 0 | 3 | 0 | 0 | 0 |
| Iaşi | 1 | 0 | 1 | 0 | 0 | 23,857 | 0 | 70,009 | 23,842 | 30,973 | 23,857 | 0 | 15 | 3,154 | 0 | 0 | 0 | 0 | 0 |
| Neamţ | 1 | 0 | 0 | 0 | 0 | 19,924 | 0 | 62,111 | 19,912 | 38,080 | 19,929 | 0 | 12 | 2,535 | 0 | 0 | 0 | 0 | 0 |
| Suceava | 0 | 0 | 0 | 0 | 0 | 32,854 | 0 | 116,971 | 32,854 | 75,083 | 32,854 | 0 | 0 | 0 | 0 | 15 | 0 | 0 | 0 |
| Vaslui | 0 | 0 | 0 | 0 | 0 | 17,085 | 0 | 40,170 | 17,085 | 25,129 | 18,045 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Brăila | 0 | 0 | 0 | 0 | 0 | 13,642 | 0 | 36,127 | 13,642 | 20,997 | 13,642 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Buzău | 0 | 0 | 0 | 0 | 0 | 12,823 | 0 | 55,669 | 12,814 | 27,877 | 12,823 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Constanţa | 0 | 0 | 0 | 0 | 0 | 4,061 | 0 | 30,494 | 4,054 | 15,141 | 4,061 | 0 | 7 | 1,271 | 0 | 0 | 0 | 0 | 0 |
| Galaţi | 0 | 0 | 0 | 0 | 0 | 8,964 | 0 | 32,771 | 8,959 | 13,871 | 8,964 | 0 | 5 | 581 | 0 | 0 | 0 | 0 | 0 |
| Tulcea | 0 | 0 | 0 | 0 | 0 | 2,024 | 0 | 39,332 | 2,024 | 18,620 | 2,740 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Vrancea | 0 | 0 | 0 | 0 | 0 | 10,039 | 0 | 36,190 | 10,036 | 21,325 | 10,039 | 0 | 3 | 720 | 0 | 0 | 0 | 0 | 0 |
| Argeş | 14 | 0 | 0 | 0 | 0 | 16,041 | 0 | 39,228 | 16,039 | 34,693 | 16,041 | 0 | 2 | 998 | 0 | 0 | 0 | 0 | 0 |
| Călăraşi | 0 | 0 | 0 | 0 | 0 | 3,217 | 0 | 18,746 | 3,217 | 11,888 | 3,217 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Dâmboviţa | 0 | 0 | 0 | 0 | 0 | 7,306 | 0 | 23,815 | 7,300 | 16,300 | 7,308 | 0 | 8 | 560 | 0 | 0 | 0 | 0 | 0 |
| Giurgiu | 0 | 0 | 0 | 0 | 0 | 4,193 | 0 | 13,484 | 4,186 | 5,880 | 4,433 | 0 | 7 | 4,781 | 0 | 0 | 0 | 0 | 0 |
| Ialomiţa | 0 | 0 | 0 | 0 | 0 | 5,655 | 0 | 26,231 | 5,645 | 10,569 | 5,655 | 0 | 10 | 2,608 | 0 | 1 | 0 | 0 | 0 |
| Prahova | 6 | 0 | 0 | 0 | 0 | 10,875 | 0 | 24,140 | 10,871 | 17,543 | 10,875 | 0 | 4 | 2,222 | 0 | 0 | 0 | 0 | 0 |
| Teleorman | 0 | 0 | 0 | 0 | 0 | 7,010 | 0 | 27,054 | 7,010 | 19,305 | 7,010 | 0 | 7 | 1,501 | 0 | 0 | 0 | 0 | 0 |
| Bucureşti | 0 | 0 | 0 | 0 | 0 | 23 | 0 | 165 | 23 | 105 | 23 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Ilfov | 0 | 1 | 1 | 0 | 0 | 831 | 0 | 5,187 | 819 | 1,573 | 898 | 0 | 12 | 1,538 | 0 | 0 | 0 | 0 | 1 |

| Region | Number of animals serologically tested under investigations of suspect cases | Number of suspended herds under investigations of suspect cases | Number of seropositive animals under investigations of suspect cases | Number of animals positive to BST under investigations of suspect cases | Number of animals positive in microbiological testing under investigations of suspect cases | Number of herds with status officially free | Number of infected herds | Total number of animals | Number of herds tested under surveillance | Number of animals tested under surveillance | Total number of herds | Number of infected herds tested under surveillance | Number of herds tested under surveillance by bulk milk | Number of animals or pools tested under surveillance by bulk milk | Number of infected herds tested under surveillance by bulk milk | Number of notified abortions whatever cause under investigations of suspect cases | Number of isolations of Brucella abortus under investigations of suspect cases | Number of abortions due to Brucella infection under investigations of suspect cases | Number of animals tested by microbiology under investigations of suspect cases |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| Dolj | 0 | 0 | 0 | 0 | 0 | 8,107 | 0 | 26,766 | 8,102 | 18,367 | 8,107 | 0 | 5 | 886 | 0 | 0 | 0 | 0 | 0 |
| Gorj | 0 | 0 | 0 | 0 | 0 | 10,233 | 0 | 34,412 | 10,233 | 22,209 | 10,233 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Mehedinţi | 0 | 0 | 0 | 0 | 0 | 8,893 | 0 | 30,924 | 8,893 | 20,767 | 8,893 | 0 | 5 | 110 | 0 | 0 | 0 | 0 | 0 |
| Olt | 0 | 0 | 0 | 0 | 0 | 8,242 | 0 | 23,314 | 8,242 | 15,597 | 8,721 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Vâlcea | 0 | 0 | 0 | 0 | 0 | 13,673 | 0 | 37,406 | 13,673 | 20,644 | 13,673 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Arad | 0 | 0 | 0 | 0 | 0 | 6,332 | 0 | 56,671 | 6,332 | 33,509 | 6,332 | 0 | 23 | 7,115 | 0 | 0 | 0 | 0 | 0 |
| Caraş-Severin | 3 | 0 | 0 | 0 | 0 | 6,680 | 0 | 17,935 | 6,680 | 17,935 | 6,680 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 |
| Hunedoara | 0 | 0 | 0 | 0 | 0 | 9,081 | 0 | 41,552 | 9,074 | 25,135 | 9,081 | 0 | 7 | 2,757 | 0 | 1 | 0 | 0 | 0 |
| Timiş | 0 | 0 | 0 | 0 | 0 | 4,378 | 0 | 36,096 | 4,350 | 20,372 | 4,400 | 0 | 50 | 7,231 | 0 | 0 | 0 | 0 | 0 |

## Table Ovine or Caprine brucellosis in countries and regions that do not receive Community co-financing for eradication programme

| Region | Number of animals serologically tested under investigations of suspect cases | Number of suspended herds under investigations of suspect cases | Number of seropositive animals under investigations of suspect cases | Number of animals positive in microbiological testing under investigations of suspect cases | Number of herds with status officially free | Number of infected herds | Total number of animals | Number of herds tested under surveillance | Number of animals tested under surveillance | Total number of herds | Number of infected herds tested under surveillance | Number of animals tested by microbiology under investigations of suspect cases |
|---|---|---|---|---|---|---|---|---|---|---|---|---|
| ROMANIA | 8 | 0 | 0 | 0 | 256,078 | 0 | 12,678,577 | 228,090 | 908,385 | 257,292 | 0 | 1 |
| Bihor | 0 | 0 | 0 | 0 | 2,757 | 0 | 460,794 | 2,757 | 32,327 | 2,757 | 0 | 0 |
| Bistriţa-Năsăud | 0 | 0 | 0 | 0 | 8,829 | 0 | 436,845 | 8,829 | 31,247 | 8,829 | 0 | 0 |
| Cluj | 0 | 0 | 0 | 0 | 4,872 | 0 | 558,783 | 4,872 | 44,286 | 4,872 | 0 | 0 |
| Maramureş | 0 | 0 | 0 | 0 | 35,532 | 0 | 281,101 | 35,532 | 22,972 | 35,532 | 0 | 0 |
| Satu Mare | 0 | 0 | 0 | 0 | 1,569 | 0 | 302,908 | 1,569 | 18,532 | 1,569 | 0 | 0 |
| Sălaj | 0 | 0 | 0 | 0 | 2,215 | 0 | 371,494 | 2,215 | 23,981 | 2,215 | 0 | 0 |
| Alba | 0 | 0 | 0 | 0 | 7,308 | 0 | 400,072 | 7,308 | 25,948 | 7,308 | 0 | 0 |
| Braşov | 0 | 0 | 0 | 0 | 7,218 | 0 | 519,446 | 7,218 | 33,217 | 7,218 | 0 | 0 |
| Covasna | 0 | 0 | 0 | 0 | 6,767 | 0 | 216,894 | 6,767 | 16,411 | 6,767 | 0 | 0 |
| Harghita | 0 | 0 | 0 | 0 | 13,281 | 0 | 257,649 | 2,578 | 16,755 | 13,281 | 0 | 0 |
| Mureş | 0 | 0 | 0 | 0 | 9,664 | 0 | 488,933 | 9,664 | 36,192 | 9,664 | 0 | 0 |
| Sibiu | 0 | 0 | 0 | 0 | 5,065 | 0 | 595,606 | 5,065 | 39,721 | 5,065 | 0 | 0 |
| Bacău | 0 | 0 | 0 | 0 | 7,138 | 0 | 339,632 | 7,138 | 18,697 | 7,138 | 0 | 0 |
| Botoşani | 0 | 0 | 0 | 0 | 5,899 | 0 | 269,522 | 5,899 | 19,813 | 5,899 | 0 | 0 |
| Iaşi | 0 | 0 | 0 | 0 | 5,942 | 0 | 364,650 | 5,942 | 23,195 | 5,942 | 0 | 0 |
| Neamţ | 0 | 0 | 0 | 0 | 8,243 | 0 | 188,720 | 8,243 | 12,019 | 8,243 | 0 | 0 |
| Suceava | 0 | 0 | 0 | 0 | 4,322 | 0 | 251,592 | 4,322 | 17,620 | 4,322 | 0 | 0 |
| Vaslui | 0 | 0 | 0 | 0 | 6,551 | 0 | 303,559 | 6,551 | 26,440 | 6,552 | 0 | 0 |
| Brăila | 0 | 0 | 0 | 0 | 3,244 | 0 | 335,827 | 235 | 22,663 | 3,244 | 0 | 0 |
| Buzău | 0 | 0 | 0 | 0 | 13,267 | 0 | 360,834 | 13,267 | 22,346 | 13,267 | 0 | 0 |
| Constanţa | 0 | 0 | 0 | 0 | 4,311 | 0 | 529,657 | 4,311 | 39,804 | 4,311 | 0 | 0 |
| Galaţi | 0 | 0 | 0 | 0 | 6,342 | 0 | 359,473 | 6,342 | 21,949 | 6,342 | 0 | 0 |
| Tulcea | 1 | 0 | 0 | 0 | 2,040 | 0 | 274,169 | 2,040 | 29,511 | 3,236 | 0 | 1 |
| Vrancea | 0 | 0 | 0 | 0 | 6,157 | 0 | 199,984 | 6,157 | 14,375 | 6,157 | 0 | 0 |
| Argeş | 7 | 0 | 0 | 0 | 7,178 | 0 | 253,176 | 7,178 | 16,173 | 7,178 | 0 | 0 |
| Călăraşi | 0 | 0 | 0 | 0 | 5,025 | 0 | 182,803 | 5,025 | 10,125 | 5,025 | 0 | 0 |
| Dâmboviţa | 0 | 0 | 0 | 0 | 2,257 | 0 | 84,902 | 133 | 5,365 | 2,257 | 0 | 0 |
| Giurgiu | 0 | 0 | 0 | 0 | 2,757 | 0 | 86,531 | 2,389 | 3,767 | 2,757 | 0 | 0 |
| Ialomiţa | 0 | 0 | 0 | 0 | 5,779 | 0 | 199,176 | 5,779 | 13,619 | 5,779 | 0 | 0 |
| Prahova | 0 | 0 | 0 | 0 | 8,692 | 0 | 253,277 | 1,496 | 17,599 | 8,692 | 0 | 0 |
| Teleorman | 0 | 0 | 0 | 0 | 4,599 | 0 | 24,165 | 4,599 | 14,902 | 4,599 | 0 | 0 |
| Bucureşti | 0 | 0 | 0 | 0 | 24 | 0 | 896 | 24 | 89 | 24 | 0 | 0 |
| Ilfov | 0 | 0 | 0 | 0 | 153 | 0 | 29,480 | 153 | 2,068 | 170 | 0 | 0 |
| Dolj | 0 | 0 | 0 | 0 | 7,767 | 0 | 315,339 | 7,767 | 28,005 | 7,767 | 0 | 0 |
| Gorj | 0 | 0 | 0 | 0 | 2,320 | 0 | 149,486 | 2,320 | 2,576 | 2,320 | 0 | 0 |
| Mehedinţi | 0 | 0 | 0 | 0 | 5,182 | 0 | 216,836 | 5,182 | 10,648 | 5,182 | 0 | 0 |

| Region | Number of animals serologically tested under investigations of suspect cases | Number of suspended herds under investigations of suspect cases | Number of seropositive animals under investigations of suspect cases | Number of animals positive in microbiological testing under investigations of suspect cases | Number of herds with status officially free | Number of infected herds | Total number of animals | Number of herds tested under surveillance | Number of animals tested under surveillance | Total number of herds | Number of infected herds tested under surveillance | Number of animals tested by microbiology under investigations of suspect cases |
|---|---|---|---|---|---|---|---|---|---|---|---|---|
| Olt | 0 | 0 | 0 | 0 | 5,135 | 0 | 203,196 | 5,135 | 14,740 | 5,135 | 0 | 0 |
| Vâlcea | 0 | 0 | 0 | 0 | 3,826 | 0 | 174,083 | 3,826 | 8,866 | 3,826 | 0 | 0 |
| Arad | 0 | 0 | 0 | 0 | 2,452 | 0 | 568,516 | 2,042 | 51,274 | 2,452 | 0 | 0 |
| Caraş-Severin | 0 | 0 | 0 | 0 | 3,969 | 0 | 240,362 | 3,969 | 19,882 | 3,969 | 0 | 0 |
| Hunedoara | 0 | 0 | 0 | 0 | 7,262 | 0 | 280,566 | 3,084 | 21,399 | 7,262 | 0 | 0 |
| Timiş | 0 | 0 | 0 | 0 | 3,168 | 0 | 747,643 | 3,168 | 57,267 | 3,168 | 0 | 0 |

## DISEASE STATUS TABLES

### Table Bovine tuberculosis in countries and regions that do not receive Community co-financing for eradication programme

| Region | Number of herds with status officially free | Number of infected herds | Total number of animals | Interval between routine tuberculin tests | Number of animals tested with tuberculin routine testing | Number of tuberculin tests carried out before the introduction into the herds | Number of animals with suspicious lesions of tuberculosis examined and submitted to histopathological and bacteriological examinations | Number of animals detected positive in bacteriological examination | Total number of herds |
|---|---|---|---|---|---|---|---|---|---|
| ROMANIA | 456,439 | 34 | 1,923,647 | 12 | 1,809,883 | 0 | 423 | 262 | 457,655 |
| Bihor | 14,378 | 13 | 70,808 | 12 | 62,230 | 0 | 24 | 38 | 14,391 |
| Bistriţa-Năsăud | 14,463 | 2 | 75,845 | 12 | 75,126 | 0 | 36 | 1 | 14,465 |
| Cluj | 9,957 | 0 | 62,751 | 12 | 60,641 | 0 | 0 | 0 | 9,957 |
| Maramureş | 37,541 | 1 | 79,003 | 12 | 76,754 | 0 | 31 | 16 | 37,542 |
| Satu Mare | 7,579 | 5 | 43,150 | 12 | 41,203 | 0 | 20 | 5 | 7,584 |
| Sălaj | 7,135 | 4 | 29,837 | 12 | 28,617 | 0 | 4 | 7 | 7,139 |
| Alba | 13,451 | 1 | 67,497 | 12 | 67,497 | 0 | 1 | 3 | 13,452 |
| Braşov | 9,405 | 0 | 66,216 | 12 | 60,978 | 0 | 20 | 0 | 9,405 |
| Covasna | 5,277 | 0 | 45,561 | 12 | 41,069 | 0 | 7 | 0 | 5,277 |
| Harghita | 15,123 | 0 | 86,990 | 12 | 79,628 | 0 | 1 | 0 | 15,123 |
| Mureş | 8,271 | 1 | 75,977 | 12 | 73,149 | 0 | 15 | 2 | 8,271 |
| Sibiu | 5,035 | 0 | 47,948 | 12 | 46,301 | 0 | 0 | 0 | 5,035 |
| Bacău | 17,486 | 0 | 47,231 | 12 | 43,033 | 0 | 0 | 0 | 17,486 |
| Botoşani | 25,536 | 0 | 102,290 | 12 | 96,559 | 0 | 0 | 0 | 25,536 |
| Iaşi | 23,857 | 0 | 70,009 | 12 | 68,700 | 0 | 42 | 0 | 23,857 |
| Neamţ | 19,926 | 0 | 62,111 | 12 | 62,790 | 0 | 0 | 0 | 19,927 |
| Suceava | 32,854 | 0 | 116,971 | 12 | 116,971 | 0 | 0 | 1 | 32,854 |
| Vaslui | 17,893 | 0 | 40,170 | 12 | 37,446 | 0 | 0 | 0 | 18,045 |
| Brăila | 13,640 | 2 | 36,127 | 12 | 35,049 | 0 | 173 | 156 | 13,642 |
| Buzău | 1,280 | 0 | 55,669 | 12 | 47,115 | 0 | 0 | 0 | 1,280 |
| Constanţa | 4,058 | 3 | 30,494 | 12 | 35,418 | 0 | 40 | 10 | 4,061 |
| Galaţi | 8,964 | 0 | 32,771 | 12 | 30,814 | 0 | 0 | 0 | 8,964 |
| Tulcea | 2,017 | 0 | 39,332 | 12 | 24,232 | 0 | 0 | 0 | 2,740 |
| Vrancea | 10,039 | 0 | 36,190 | 12 | 36,924 | 0 | 0 | 0 | 10,039 |
| Argeş | 16,042 | 0 | 46,678 | 12 | 45,013 | 0 | 0 | 0 | 16,042 |
| Călăraşi | 3,217 | 0 | 18,746 | 12 | 17,504 | 0 | 0 | 5 | 3,217 |
| Dâmboviţa | 7,308 | 0 | 23,815 | 12 | 23,490 | 0 | 0 | 0 | 7,308 |
| Giurgiu | 4,193 | 0 | 13,484 | 12 | 10,654 | 0 | 0 | 0 | 4,433 |
| Ialomiţa | 5,655 | 0 | 26,231 | 12 | 19,992 | 0 | 5 | 10 | 5,655 |
| Prahova | 10,875 | 0 | 30,582 | 12 | 27,880 | 0 | 0 | 1 | 10,875 |
| Teleorman | 7,011 | 1 | 33,450 | 12 | 27,195 | 0 | 2 | 2 | 7,012 |

| Region | Number of herds with status officially free | Number of infected herds | Total number of animals | Interval between routine tuberculin tests | Number of animals tested with tuberculin routine testing | Number of tuberculin tests carried out before the introduction into the herds | Number of animals with suspicious lesions of tuberculosis examined and submitted to histopathological and bacteriological examinations | Number of animals detected positive in bacteriological examination | Total number of herds |
|---|---|---|---|---|---|---|---|---|---|
| Bucureşti | 23 | 0 | 165 | 12 | 146 | 0 | 0 | 0 | 23 |
| Ilfov | 831 | 0 | 5,187 | 12 | 4,600 | 0 | 0 | 0 | 898 |
| Dolj | 8,107 | 0 | 26,766 | 12 | 26,776 | 0 | 0 | 0 | 8,107 |
| Gorj | 10,233 | 0 | 34,412 | 12 | 33,311 | 0 | 0 | 0 | 10,233 |
| Mehedinţi | 8,893 | 0 | 23,960 | 12 | 19,539 | 0 | 0 | 0 | 8,893 |
| Olt | 8,720 | 0 | 23,314 | 12 | 21,508 | 0 | 0 | 0 | 8,721 |
| Vâlcea | 13,673 | 0 | 37,406 | 12 | 32,246 | 0 | 0 | 0 | 13,673 |
| Arad | 6,332 | 0 | 56,671 | 12 | 55,139 | 0 | 0 | 2 | 6,332 |
| Caraş-Severin | 6,680 | 0 | 24,184 | 12 | 24,184 | 0 | 0 | 2 | 6,680 |
| Hunedoara | 9,081 | 0 | 41,552 | 12 | 37,593 | 0 | 1 | 0 | 9,081 |
| Timiş | 4,400 | 1 | 36,096 | 12 | 34,869 | 0 | 1 | 1 | 4,400 |

## PREVALENCE TABLES

### Table Brucella:BRUCELLA in animal

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| ROMANIA | Bison - farmed - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | N_A | Complement fixation test (CFT) | animal | 38 | 0 | Brucella | 0 |
| | Bison - farmed - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | N_A | Rose Bengal plate test (RBT)/Buffered Brucella antigen test (BBAT) | animal | 38 | 0 | Brucella | 0 |
| | Bison - farmed - Farm - Romania - animal sample - blood - Unspecified - Official sampling - Objective sampling | N_A | Rose Bengal plate test (RBT)/Buffered Brucella antigen test (BBAT) | animal | 269 | 0 | Brucella | 0 |
| | Buffalos - zoo animal - Zoo - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | N_A | Complement fixation test (CFT) | animal | 6 | 0 | Brucella | 0 |
| | Buffalos - zoo animal - Zoo - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | N_A | Rose Bengal plate test (RBT)/Buffered Brucella antigen test (BBAT) | animal | 6 | 0 | Brucella | 0 |
| | Camels - zoo animals - Zoo - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | N_A | Complement fixation test (CFT) | animal | 12 | 0 | Brucella | 0 |
| | Camels - zoo animals - Zoo - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | N_A | Rose Bengal plate test (RBT)/Buffered Brucella antigen test (BBAT) | animal | 12 | 0 | Brucella | 0 |
| | Dogs - Unspecified - Romania - animal sample - blood - Unspecified - Official sampling - Objective sampling | backyards | Complement fixation test (CFT) | animal | 3 | 3 | Brucella canis | 3 |
| | Dogs - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | backyards | Microbiological tests | animal | 1 | 1 | Brucella canis | 1 |
| | Dogs - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | backyards | PCR | animal | 1 | 1 | Brucella canis | 1 |
| | Lamas - zoo animal - Zoo - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | N_A | Complement fixation test (CFT) | animal | 5 | 0 | Brucella | 0 |
| | Lamas - zoo animal - Zoo - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | N_A | Rose Bengal plate test (RBT)/Buffered Brucella antigen test (BBAT) | animal | 5 | 0 | Brucella | 0 |
| | Mouflons - zoo animal - Zoo - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | muflon | Complement fixation test (CFT) | animal | 5 | 0 | Brucella | 0 |
| | Mouflons - zoo animal - Zoo - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | muflon | Rose Bengal plate test (RBT)/Buffered Brucella antigen test (BBAT) | animal | 5 | 0 | Brucella | 0 |
| | Other ruminants - wild - Zoo - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | yak | Complement fixation test (CFT) | animal | 2 | 0 | Brucella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| ROMANIA | Other ruminants - wild - Zoo - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | yak | Rose Bengal plate test (RBT)/Buffered Brucella antigen test (BBAT) | animal | 2 | 0 | Brucella | 0 |
| | Other ruminants - zoo animals - Farm - Romania - animal sample - blood - Unspecified - Official sampling - Objective sampling | zebu | Complement fixation test (CFT) | animal | 1 | 0 | Brucella | 0 |
| | Other ruminants - zoo animals - Farm - Romania - animal sample - blood - Unspecified - Official sampling - Objective sampling | zebu | Rose Bengal plate test (RBT)/Buffered Brucella antigen test (BBAT) | animal | 1 | 0 | Brucella | 0 |
| | Other ruminants - zoo animals - Zoo - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | watusi | Complement fixation test (CFT) | animal | 1 | 0 | Brucella | 0 |
| | | zebu | Complement fixation test (CFT) | animal | 2 | 0 | Brucella | 0 |
| | Other ruminants - zoo animals - Zoo - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | watusi | Rose Bengal plate test (RBT)/Buffered Brucella antigen test (BBAT) | animal | 1 | 0 | Brucella | 0 |
| | | zebu | Rose Bengal plate test (RBT)/Buffered Brucella antigen test (BBAT) | animal | 2 | 0 | Brucella | 0 |
| | Pigs - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | N_A | Complement fixation test (CFT) | animal | 19859 | 0 | Brucella | 0 |
| | Pigs - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | N_A | Rose Bengal plate test (RBT)/Buffered Brucella antigen test (BBAT) | animal | 19895 | 0 | Brucella | 0 |
| | Pigs - Farm - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Out of 132 samples, 131 were semen samples. | Microbiological tests | animal | 132 | 0 | Brucella | 0 |
| | Pigs - Unspecified - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | backyards | Complement fixation test (CFT) | animal | 17401 | 6 | Brucella suis | 6 |
| | Pigs - Unspecified - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | backyards | Rose Bengal plate test (RBT)/Buffered Brucella antigen test (BBAT) | animal | 17698 | 6 | Brucella suis | 6 |
| | Pigs - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Out of the 6 CFT positive samples, only 2 animals were tested by bacteriological methods. The others were samples from dead animals or abortions. | Microbiological tests | animal | 6 | 0 | Brucella | 0 |
| | Wild boars - Natural habitat - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | N_A | Complement fixation test (CFT) | animal | 816 | 4 | Brucella suis | 4 |
| | Wild boars - Natural habitat - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | N_A | Rose Bengal plate test (RBT)/Buffered Brucella antigen test (BBAT) | animal | 816 | 4 | Brucella suis | 4 |
| | Wild boars - Natural habitat - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Microbiological tests | animal | 521 | 7 | Brucella suis | 7 |
| | Wild boars - Natural habitat - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | PCR | animal | 7 | 7 | Brucella suis | 7 |

## Table Calicivirus:CALICIVIRUS in food

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Fruits - non-pre-cut - chilled - Border inspection activities - Albania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Reverse-transcription PCR (RT-PCR) | 2 | 0 | Calicivirus | 0 |
| | Fruits - non-pre-cut - chilled - Border inspection activities - Turkey - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Reverse-transcription PCR (RT-PCR) | 11 | 0 | Calicivirus | 0 |
| | Fruits - non-pre-cut - chilled - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Reverse-transcription PCR (RT-PCR) | 2 | 0 | Calicivirus | 0 |
| | Fruits - non-pre-cut - frozen - Border inspection activities - Serbia - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Reverse-transcription PCR (RT-PCR) | 2 | 0 | Calicivirus | 0 |
| | Fruits - non-pre-cut - frozen - Border inspection activities - Turkey - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Reverse-transcription PCR (RT-PCR) | 1 | 0 | Calicivirus | 0 |
| | Fruits - non-pre-cut - frozen - Processing plant - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Reverse-transcription PCR (RT-PCR) | 2 | 0 | Calicivirus | 0 |

## Table Campylobacter:CAMPYLOBACTER in animal

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| ROMANIA | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 838 | 741 | Campylobacter coli | 416 |
| | | | | | | | Campylobacter jejuni | 338 |
| | Turkeys - fattening flocks - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 18 | 16 | Campylobacter coli | 9 |
| | | | | | | | Campylobacter jejuni | 7 |
| Bihor | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 6 | 5 | Campylobacter coli | 4 |
| | | | | | | | Campylobacter jejuni | 1 |
| Cluj | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 42 | 33 | Campylobacter coli | 25 |
| | | | | | | | Campylobacter jejuni | 8 |
| Maramureş | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 7 | 7 | Campylobacter coli | 6 |
| | | | | | | | Campylobacter jejuni | 1 |
| Satu Mare | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 29 | 25 | Campylobacter coli | 15 |
| | | | | | | | Campylobacter jejuni | 10 |
| Sălaj | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 6 | 6 | Campylobacter coli | 4 |
| | | | | | | | Campylobacter jejuni | 2 |
| Alba | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 33 | 17 | Campylobacter coli | 4 |
| | | | | | | | Campylobacter jejuni | 13 |
| Braşov | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 58 | 50 | Campylobacter coli | 43 |
| | | | | | | | Campylobacter jejuni | 8 |
| | Turkeys - fattening flocks - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 13 | 11 | Campylobacter coli | 5 |
| | | | | | | | Campylobacter jejuni | 6 |
| Harghita | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 6 | 2 | Campylobacter coli | 0 |
| | | | | | | | Campylobacter jejuni | 2 |
| Mureş | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 13 | 12 | Campylobacter coli | 4 |
| | | | | | | | Campylobacter jejuni | 10 |
| | Turkeys - fattening flocks - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 5 | 5 | Campylobacter coli | 4 |
| | | | | | | | Campylobacter jejuni | 1 |
| Sibiu | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 16 | 13 | Campylobacter coli | 7 |
| | | | | | | | Campylobacter jejuni | 6 |
| Bacău | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 90 | 87 | Campylobacter coli | 53 |
| | | | | | | | Campylobacter jejuni | 34 |
| Botoşani | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 20 | 19 | Campylobacter coli | 11 |
| | | | | | | | Campylobacter jejuni | 9 |
| Iaşi | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 43 | 42 | Campylobacter coli | 18 |
| | | | | | | | Campylobacter jejuni | 24 |
| Neamţ | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 12 | 11 | Campylobacter coli | 2 |
| | | | | | | | Campylobacter jejuni | 9 |
| Suceava | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 1 | 1 | Campylobacter coli | 0 |
| | | | | | | | Campylobacter jejuni | 1 |
| Vaslui | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 38 | 37 | Campylobacter coli | 22 |
| | | | | | | | Campylobacter jejuni | 17 |
| Brăila | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 7 | 4 | Campylobacter coli | 3 |
| | | | | | | | Campylobacter jejuni | 1 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Buzău | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 67 | 60 | Campylobacter coli | 21 |
| | | | | | | | Campylobacter jejuni | 42 |
| Constanţa | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 11 | 10 | Campylobacter coli | 6 |
| | | | | | | | Campylobacter jejuni | 4 |
| Galaţi | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 16 | 11 | Campylobacter coli | 10 |
| | | | | | | | Campylobacter jejuni | 1 |
| Vrancea | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 10 | 10 | Campylobacter coli | 6 |
| | | | | | | | Campylobacter jejuni | 4 |
| Argeş | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 18 | 17 | Campylobacter coli | 8 |
| | | | | | | | Campylobacter jejuni | 9 |
| Călăraşi | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 109 | 101 | Campylobacter coli | 70 |
| | | | | | | | Campylobacter jejuni | 31 |
| Dâmboviţa | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 10 | 10 | Campylobacter coli | 6 |
| | | | | | | | Campylobacter jejuni | 4 |
| Giurgiu | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 20 | 17 | Campylobacter coli | 7 |
| | | | | | | | Campylobacter jejuni | 13 |
| Ialomiţa | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 30 | 30 | Campylobacter coli | 18 |
| | | | | | | | Campylobacter jejuni | 13 |
| Prahova | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 36 | 33 | Campylobacter coli | 12 |
| | | | | | | | Campylobacter jejuni | 21 |
| Teleorman | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 4 | 4 | Campylobacter coli | 1 |
| | | | | | | | Campylobacter jejuni | 3 |
| Dolj | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 4 | 4 | Campylobacter coli | 1 |
| | | | | | | | Campylobacter jejuni | 3 |
| Gorj | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 6 | 6 | Campylobacter coli | 1 |
| | | | | | | | Campylobacter jejuni | 5 |
| Vâlcea | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 22 | 18 | Campylobacter coli | 1 |
| | | | | | | | Campylobacter jejuni | 17 |
| Arad | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 1 | 1 | Campylobacter coli | 0 |
| | | | | | | | Campylobacter jejuni | 1 |
| Caraş-Severin | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 27 | 19 | Campylobacter coli | 11 |
| | | | | | | | Campylobacter jejuni | 8 |
| Hunedoara | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 17 | 17 | Campylobacter coli | 16 |
| | | | | | | | Campylobacter jejuni | 1 |
| Timiş | Gallus gallus (fowl) - broilers - Slaughterhouse - Romania - animal sample - caecum - Monitoring - Official sampling - Objective sampling | N_A | ISO 10272-1:2017 Campylobacter | slaughter animal batch | 3 | 2 | Campylobacter coli | 0 |
| | | | | | | | Campylobacter jejuni | 2 |

## Table Campylobacter:CAMPYLOBACTER in food

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Meat from broilers (Gallus gallus) - carcase - chilled - Slaughterhouse - Romania - food sample - neck skin - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | The indigenous origin | Enumeration method | 1700 | 750 | Campylobacter, unspecified sp. | 750 |
| | Meat from broilers (Gallus gallus) - carcase - chilled - Slaughterhouse - Romania - food sample - neck skin - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | The indigenous origin | Enumeration method | 130 | 76 | Campylobacter, unspecified sp. | 76 |
| | Meat from broilers (Gallus gallus) - fresh - Retail - Romania - food sample - meat - Surveillance - HACCP and own check - Other | batch (food/feed) | 10 | Gram | The indigenous origin | Enumeration method | 10 | 0 | Campylobacter | 0 |
| | Meat from broilers (Gallus gallus) - fresh - Retail - Romania - food sample - meat - Surveillance - HACCP and own check - Other | batch (food/feed) | 25 | Gram | The indigenous origin | Detection method presence in x g | 1 | 1 | Campylobacter, unspecified sp. | 1 |
| | Meat from broilers (Gallus gallus) - fresh - Slaughterhouse - Romania - food sample - meat - Surveillance - HACCP and own check - Other | batch (food/feed) | 10 | Gram | The indigenous origin | Enumeration method | 5 | 0 | Campylobacter | 0 |
| | Meat from broilers (Gallus gallus) - fresh - Slaughterhouse - Romania - food sample - meat - Surveillance - HACCP and own check - Other | batch (food/feed) | 25 | Gram | The indigenous origin | Detection method presence in x g | 76 | 9 | Campylobacter, unspecified sp. | 9 |

## Table COXIELLA in animal

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sampling Details | Method | Total units tested | Total units positive | N of clinical affected herds | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|
| ROMANIA | Cattle (bovine animals) - Unspecified - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | animal | backyards | Indirect ELISA (I-ELISA) | 5 | 0 | | Coxiella burnetii | 0 |
| | Goats - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | animal | N_A | Indirect ELISA (I-ELISA) | 20 | 0 | | Coxiella burnetii | 0 |
| | Goats - Unspecified - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | animal | backyards | Indirect ELISA (I-ELISA) | 14 | 10 | | Coxiella burnetii | 10 |
| | Goats - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | backyards | PCR | 1 | 0 | | Coxiella burnetii | 0 |
| | Sheep - Unspecified - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | animal | backyards | Indirect ELISA (I-ELISA) | 7 | 0 | | Coxiella burnetii | 0 |

## Table Cronobacter:CRONOBACTER in food

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Foodstuffs intended for special nutritional uses - dried dietary foods for special medical purposes intended for infants below 6 months - Retail - European Union - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 10 | 0 | Cronobacter | 0 |
| | Foodstuffs intended for special nutritional uses - dried dietary foods for special medical purposes intended for infants below 6 months - Retail - European Union - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 8 | 0 | Cronobacter | 0 |

## Table Echinococcus:ECHINOCOCCUS in animal

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| ROMANIA | Cattle (bovine animals) - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | PCR | animal | 10 | 10 | Echinococcus granulosus | 10 |
| | Cattle (bovine animals) - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | PCR | animal | 12 | 12 | Echinococcus granulosus | 12 |
| | Deer - Natural habitat - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Detection method of microorganisms | animal | 2 | 0 | Echinococcus | 0 |
| | Dogs - Farm - Romania - animal sample - faeces - Surveillance - Official sampling - Objective sampling | N_A | Detection method of microorganisms | animal | 34 | 0 | Echinococcus | 0 |
| | Dogs - Unspecified - Romania - animal sample - faeces - Surveillance - Industry sampling - Objective sampling | N_A | Detection method of microorganisms | animal | 1 | 0 | Echinococcus | 0 |
| | Pigs - Farm - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Detection method of microorganisms | animal | 12 | 0 | Echinococcus | 0 |
| | Pigs - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Detection method of microorganisms | animal | 3 | 0 | Echinococcus | 0 |
| | Pigs - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | PCR | animal | 2 | 2 | Echinococcus granulosus | 2 |
| | Sheep - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Detection method of microorganisms | animal | 4 | 0 | Echinococcus | 0 |
| | Sheep - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | PCR | animal | 1 | 1 | Echinococcus granulosus | 1 |
| | Sheep - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Detection method of microorganisms | animal | 11 | 2 | Echinococcus | 2 |
| Bihor | Sheep - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Suspect sampling | N_A | Detection method of microorganisms | animal | 3 | 0 | Echinococcus | 0 |
| Alba | Dogs - Unspecified - Romania - animal sample - faeces - Surveillance - Industry sampling - Objective sampling | N_A | Detection method of microorganisms | animal | 1 | 0 | Echinococcus | 0 |
| Sibiu | Cattle (bovine animals) - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | PCR | animal | 1 | 1 | Echinococcus granulosus | 1 |
| Bacău | Sheep - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Detection method of microorganisms | animal | 1 | 0 | Echinococcus | 0 |
| | Sheep - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Detection method of microorganisms | animal | 2 | 2 | Echinococcus | 2 |
| Neamț | Deer - Natural habitat - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Detection method of microorganisms | animal | 2 | 0 | Echinococcus | 0 |
| | Dogs - Farm - Romania - animal sample - faeces - Surveillance - Official sampling - Objective sampling | N_A | Detection method of microorganisms | animal | 34 | 0 | Echinococcus | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Neamţ | Sheep - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Detection method of microorganisms | animal | 2 | 0 | Echinococcus | 0 |
| Brăila | Cattle (bovine animals) - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | PCR | animal | 10 | 10 | Echinococcus granulosus | 10 |
| | Sheep - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Detection method of microorganisms | animal | 3 | 0 | Echinococcus | 0 |
| | Sheep - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | PCR | animal | 1 | 1 | Echinococcus granulosus | 1 |
| Tulcea | Sheep - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Detection method of microorganisms | animal | 4 | 0 | Echinococcus | 0 |
| Vrancea | Pigs - Farm - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Detection method of microorganisms | animal | 11 | 0 | Echinococcus | 0 |
| Argeş | Cattle (bovine animals) - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | PCR | animal | 4 | 4 | Echinococcus granulosus | 4 |
| Dolj | Pigs - Farm - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Detection method of microorganisms | animal | 1 | 0 | Echinococcus | 0 |
| | Pigs - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Detection method of microorganisms | animal | 3 | 0 | Echinococcus | 0 |
| | Pigs - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | PCR | animal | 2 | 2 | Echinococcus granulosus | 2 |
| Gorj | Cattle (bovine animals) - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | PCR | animal | 2 | 2 | Echinococcus granulosus | 2 |
| Vâlcea | Cattle (bovine animals) - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | PCR | animal | 4 | 4 | Echinococcus granulosus | 4 |
| Hunedoara | Cattle (bovine animals) - Unspecified - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | PCR | animal | 1 | 1 | Echinococcus granulosus | 1 |

## Table Escherichia coli:ESCHERICHIA COLI in food

| Area of sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | total units tested | total units positive | Zoonoses | ANTH | VTX | AG | N units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Cheeses made from cows' milk - fresh - made from raw or low heat-treated milk - Processing plant - Romania - food sample - milk - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 8 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Cheeses made from cows' milk - fresh - made from raw or low heat-treated milk - Processing plant - Romania - food sample - milk - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 1 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Romania - food sample - milk - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 1 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Cheeses made from goats' milk - hard - made from pasteurised milk - Processing plant - Romania - food sample - milk - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 2 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Dairy products (excluding cheeses) - yoghurt - Processing plant - Romania - food sample - milk - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 3 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Meat from bovine animals - carcase - chilled - Slaughterhouse - Romania - food sample - carcase swabs - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 400 | Square centimetre | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 2 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Meat from bovine animals - fresh - chilled - Slaughterhouse - Romania - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 1 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Meat from bovine animals - minced meat - intended to be eaten cooked - Processing plant - Romania - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 1 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Meat from bovine animals - minced meat - intended to be eaten cooked - Retail - Romania - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 1 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Meat from bovine animals - minced meat - intended to be eaten cooked - Slaughterhouse - Romania - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 1 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Meat from broilers (Gallus gallus) - fresh - chilled - Slaughterhouse - Romania - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 4 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Meat from sheep - carcase - chilled - Slaughterhouse - Romania - food sample - carcase swabs - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 400 | Square centimetre | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 1 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |

| Area of sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | total units tested | total units positive | Zoonoses | ANTH | VTX | AG | N units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Meat from sheep - fresh - chilled - Processing plant - Romania - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 1 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Meat from sheep - fresh - chilled - Slaughterhouse - Romania - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 1 | 1 | VTEC other than O157 O26 O103 O111 O145 | H-antigen unknown | VT2, gene identified, subtype unspecified ;VT1, gene identified, subtype unspecified | eae negative | 1 |
| | Meat, mixed meat - meat preparation - intended to be eaten cooked - Cutting plant - Romania - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 1 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Milk, cows' - raw milk - Farm - Romania - food sample - milk - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Millilitre | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 1 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Milk, cows' - raw milk - Farm - Romania - food sample - milk - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Millilitre | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 1 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Milk, cows' - raw milk - Processing plant - Romania - food sample - milk - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Millilitre | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 8 | 1 | VTEC other than O157 O26 O103 O111 O145 | H-antigen unknown | VT2, gene identified, subtype unspecified | eae negative | 1 |
| | Milk, cows' - UHT milk - Catering - Romania - food sample - milk - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Millilitre | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 5 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Milk, goats' - raw milk - Processing plant - Romania - food sample - milk - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Millilitre | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 1 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Other processed food products and prepared dishes - Catering - Romania - food sample - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 1 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Seeds, sprouted - ready-to-eat - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 1 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Seeds, sprouted - ready-to-eat - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 3 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Seeds, sprouted - ready-to-eat - Retail - Hungary - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 3 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |

| Area of sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | total units tested | total units positive | Zoonoses | ANTH | VTX | AG | N units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Seeds, sprouted - ready-to-eat - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 11 | 0 | Verocytotoxigenic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |

## Table FLAVIVIRUS in animal

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Vaccination status | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| ROMANIA | Gallus gallus (fowl) - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Census | animal | No | N_A | IgG ELISA | 21 | 8 | | West Nile virus | 8 |
| | Solipeds, domestic - horses - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Census | animal | No | N_A | IgG ELISA | 1 | 1 | | West Nile virus | 1 |
| | Solipeds, domestic - horses - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Census | animal | No | N_A | IgM-capture ELISA (MAC-ELISA) | 235 | 13 | | West Nile virus | 13 |
| | Solipeds, domestic - horses - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Suspect sampling | animal | No | N_A | IgG ELISA | 2 | 2 | | West Nile virus | 2 |
| | Solipeds, domestic - horses - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Suspect sampling | animal | No | N_A | IgM-capture ELISA (MAC-ELISA) | 9 | 5 | | West Nile virus | 5 |
| Covasna | Solipeds, domestic - horses - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Suspect sampling | animal | No | N_A | IgG ELISA | 2 | 2 | | West Nile virus | 2 |
| | Solipeds, domestic - horses - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Suspect sampling | animal | No | N_A | IgM-capture ELISA (MAC-ELISA) | 2 | 1 | | West Nile virus | 1 |
| Harghita | Solipeds, domestic - horses - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Suspect sampling | animal | No | N_A | IgM-capture ELISA (MAC-ELISA) | 2 | 0 | | West Nile virus | 0 |
| Sibiu | Gallus gallus (fowl) - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Census | animal | No | N_A | IgG ELISA | 12 | 6 | | West Nile virus | 6 |
| | Solipeds, domestic - horses - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Census | animal | No | N_A | IgG ELISA | 1 | 1 | | West Nile virus | 1 |
| | Solipeds, domestic - horses - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Census | animal | No | N_A | IgM-capture ELISA (MAC-ELISA) | 3 | 1 | | West Nile virus | 1 |
| Brăila | Solipeds, domestic - horses - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Census | animal | No | N_A | IgM-capture ELISA (MAC-ELISA) | 151 | 10 | | West Nile virus | 10 |
| Constanţa | Solipeds, domestic - horses - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Census | animal | No | N_A | IgM-capture ELISA (MAC-ELISA) | 45 | 0 | | West Nile virus | 0 |
| Ilfov | Solipeds, domestic - horses - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Census | animal | No | N_A | IgM-capture ELISA (MAC-ELISA) | 20 | 0 | | West Nile virus | 0 |
| | Solipeds, domestic - horses - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Suspect sampling | animal | No | N_A | IgM-capture ELISA (MAC-ELISA) | 5 | 4 | | West Nile virus | 4 |
| Dolj | Gallus gallus (fowl) - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Census | animal | No | N_A | IgG ELISA | 9 | 2 | | West Nile virus | 2 |
| | Solipeds, domestic - horses - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Census | animal | No | N_A | IgM-capture ELISA (MAC-ELISA) | 16 | 2 | | West Nile virus | 2 |

## Table Hepatitis virus:HEPATITIS VIRUS in food

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Fruits - non-pre-cut - chilled - Border inspection activities - Albania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Reverse-transcription PCR (RT-PCR) | 2 | 0 | Hepatitis virus | 0 |
| | Fruits - non-pre-cut - chilled - Border inspection activities - Turkey - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Reverse-transcription PCR (RT-PCR) | 11 | 0 | Hepatitis virus | 0 |
| | Fruits - non-pre-cut - chilled - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Reverse-transcription PCR (RT-PCR) | 2 | 0 | Hepatitis virus | 0 |
| | Fruits - non-pre-cut - frozen - Border inspection activities - Serbia - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Reverse-transcription PCR (RT-PCR) | 2 | 0 | Hepatitis virus | 0 |
| | Fruits - non-pre-cut - frozen - Border inspection activities - Turkey - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Reverse-transcription PCR (RT-PCR) | 1 | 0 | Hepatitis virus | 0 |
| | Fruits - non-pre-cut - frozen - Processing plant - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Reverse-transcription PCR (RT-PCR) | 2 | 0 | Hepatitis virus | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Border inspection activities - Non European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Other | single (food/feed) | 10 | Gram | Products from third country trade. | 18 | 0 | <= 100 | Histamine | 0 | 0 |
| | | | | | | | | >100 TO <= 200 | Histamine | 0 | 0 |
| | | | | | | | | >200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Conservation facilities - European Union - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products from intra-Community trade. Sample taken from storage space | 18 | 0 | <= 100 | Histamine | 0 | 0 |
| | | | | | | | | >100 TO <= 200 | Histamine | 0 | 0 |
| | | | | | | | | >200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Conservation facilities - European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products from intra-Community trade. Sample taken from storage space | 90 | 0 | <= 100 | Histamine | 0 | 0 |
| | | | | | | | | >100 TO <= 200 | Histamine | 0 | 0 |
| | | | | | | | | >200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Conservation facilities - Non European Union - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products from third country trade. Sample taken from storage space | 18 | 0 | <= 100 | Histamine | 0 | 0 |
| | | | | | | | | >100 TO <= 200 | Histamine | 0 | 0 |
| | | | | | | | | >200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Conservation facilities - Non European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products from third country trade. Sample taken from storage space | 63 | 0 | <= 100 | Histamine | 0 | 0 |
| | | | | | | | | >100 TO <= 200 | Histamine | 0 | 0 |
| | | | | | | | | >200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products of indigenous origin. Sample taken from storage space | 18 | 3 | <= 100 | Histamine | 0 | 13 |
| | | | | | | | | >100 TO <= 200 | Histamine | 0 | 2 |
| | | | | | | | | >200 | Histamine | 0 | 3 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products of indigenous origin. Sample taken from storage space | 19 | 0 | <= 100 | Histamine | 0 | 0 |
| | | | | | | | | >100 TO <= 200 | Histamine | 0 | 0 |
| | | | | | | | | >200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Processing plant - European Union - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products from intra-Community trade. | 9 | 0 | <= 100 | Histamine | 0 | 0 |
| | | | | | | | | >100 TO <= 200 | Histamine | 0 | 0 |
| | | | | | | | | >200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Processing plant - European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products from intra-Community trade. | 81 | 0 | <= 100 | Histamine | 0 | 0 |
| | | | | | | | | >100 TO <= 200 | Histamine | 0 | 0 |
| | | | | | | | | >200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Processing plant - Non European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products from third country trade. | 18 | 0 | <= 100 | Histamine | 0 | 0 |
| | | | | | | | | >100 TO <= 200 | Histamine | 0 | 0 |
| | | | | | | | | >200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 10 | Gram | Products of indigenous origin. | 18 | 0 | <= 100 | Histamine | 0 | 0 |
| | | | | | | | | >100 TO <= 200 | Histamine | 0 | 0 |
| | | | | | | | | >200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products of indigenous origin. | 9 | 0 | <= 100 | Histamine | 0 | 0 |
| | | | | | | | | >100 TO <= 200 | Histamine | 0 | 0 |
| | | | | | | | | >200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Retail - European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products from intra-Community trade. | 423 | 0 | <= 100 | Histamine | 0 | 0 |
| | | | | | | | | >100 TO <= 200 | Histamine | 0 | 0 |
| | | | | | | | | >200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Retail - Non European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products from third country trade. | 91 | 0 | <= 100 | Histamine | 0 | 0 |
| | | | | | | | | >100 TO <= 200 | Histamine | 0 | 0 |
| | | | | | | | | >200 | Histamine | 0 | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products of indigenous origin. | 153 | 0 | <= 100 | Histamine | 0 | 0 |
| | | | | | | | | >100 TO <= 200 | Histamine | 0 | 0 |
| | | | | | | | | >200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - which have undergone enzyme maturation treatment in brine - Border inspection activities - Non European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Other | single (food/feed) | 10 | Gram | Products from third country trade. | 18 | 0 | > 400 | Histamine | 0 | 0 |
| | | | | | | | | >200 TO <= 400 | Histamine | 0 | 0 |
| | | | | | | | | <=200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - which have undergone enzyme maturation treatment in brine - Border inspection activities - Non European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 10 | Gram | Products from third country trade. | 18 | 0 | > 400 | Histamine | 0 | 0 |
| | | | | | | | | >200 TO <= 400 | Histamine | 0 | 0 |
| | | | | | | | | <=200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - which have undergone enzyme maturation treatment in brine - Conservation facilities - European Union - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products from intra-Community trade. Sample taken from storage space | 27 | 0 | > 400 | Histamine | 0 | 0 |
| | | | | | | | | >200 TO <= 400 | Histamine | 0 | 0 |
| | | | | | | | | <=200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - which have undergone enzyme maturation treatment in brine - Conservation facilities - European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products from intra-Community trade. Sample taken from storage space | 27 | 0 | > 400 | Histamine | 0 | 0 |
| | | | | | | | | >200 TO <= 400 | Histamine | 0 | 0 |
| | | | | | | | | <=200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - which have undergone enzyme maturation treatment in brine - Conservation facilities - Non European Union - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products from third country trade. Sample taken from storage space | 117 | 0 | > 400 | Histamine | 0 | 0 |
| | | | | | | | | >200 TO <= 400 | Histamine | 0 | 0 |
| | | | | | | | | <=200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - which have undergone enzyme maturation treatment in brine - Conservation facilities - Non European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products from third country trade. Sample taken from storage space | 18 | 0 | > 400 | Histamine | 0 | 0 |
| | | | | | | | | >200 TO <= 400 | Histamine | 0 | 0 |
| | | | | | | | | <=200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - which have undergone enzyme maturation treatment in brine - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products of indigenous origin. Sample taken from storage space | 9 | 0 | > 400 | Histamine | 0 | 0 |
| | | | | | | | | >200 TO <= 400 | Histamine | 0 | 0 |
| | | | | | | | | <=200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - which have undergone enzyme maturation treatment in brine - Processing plant - European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products from intra-Community trade. | 18 | 0 | > 400 | Histamine | 0 | 0 |
| | | | | | | | | >200 TO <= 400 | Histamine | 0 | 0 |
| | | | | | | | | <=200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - which have undergone enzyme maturation treatment in brine - Processing plant - Non European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products from third country trade. | 18 | 0 | > 400 | Histamine | 0 | 0 |
| | | | | | | | | >200 TO <= 400 | Histamine | 0 | 0 |
| | | | | | | | | <=200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - which have undergone enzyme maturation treatment in brine - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products of indigenous origin. | 27 | 0 | > 400 | Histamine | 0 | 0 |
| | | | | | | | | >200 TO <= 400 | Histamine | 0 | 0 |
| | | | | | | | | <=200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - which have undergone enzyme maturation treatment in brine - Retail - European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products from intra-Community trade. | 27 | 0 | > 400 | Histamine | 0 | 0 |
| | | | | | | | | >200 TO <= 400 | Histamine | 0 | 0 |
| | | | | | | | | <=200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - which have undergone enzyme maturation treatment in brine - Retail - Non European Union - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 10 | Gram | Products from third country trade. | 9 | 0 | > 400 | Histamine | 0 | 0 |
| | | | | | | | | >200 TO <= 400 | Histamine | 0 | 0 |
| | | | | | | | | <=200 | Histamine | 0 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - which have undergone enzyme maturation treatment in brine - Retail - Non European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products from third country trade. | 99 | 5 | > 400 | Histamine | 0 | 5 |
| | | | | | | | | >200 TO <= 400 | Histamine | 0 | 0 |
| | | | | | | | | <=200 | Histamine | 0 | 94 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - which have undergone enzyme maturation treatment in brine - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products of indigenous origin. | 54 | 0 | > 400 | Histamine | 0 | 0 |
| | | | | | | | | >200 TO <= 400 | Histamine | 0 | 0 |
| | | | | | | | | <=200 | Histamine | 0 | 0 |

## Table Listeria:LISTERIA in animal

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| ROMANIA | All animals - farmed - Farm - Not Available - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 2 | 0 | Listeria monocytogenes | 0 |
| | Bears - Zoo - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Buffalos - Zoo - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Cats - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Microbiological tests | animal | 33 | 2 | Listeria monocytogenes | 1 |
| | | | | | | | Listeria welshimeri | 1 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 7 | 0 | Listeria monocytogenes | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | N_A | Microbiological tests | animal | 31 | 0 | Listeria monocytogenes | 0 |
| | Chinchillas - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Deer - Veterinary clinics - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Dogs - Farm - Not Available - animal sample - brain - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Goats - mixed herds - Farm - Not Available - animal sample - brain - Clinical investigations - Industry sampling - Suspect sampling | N_A | Microbiological tests | animal | 9 | 0 | Listeria monocytogenes | 0 |
| | Goats - mixed herds - Farm - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | backyards | Microbiological tests | animal | 2 | 2 | Listeria innocua | 2 |
| | Goats - mixed herds - Farm - Not Available - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 5 | 0 | Listeria monocytogenes | 0 |
| | Goats - mixed herds - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 14 | 0 | Listeria monocytogenes | 0 |
| | Goats - Zoo - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Minks - Farm - Not Available - animal sample - brain - Clinical investigations - Industry sampling - Suspect sampling | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | backyards | Microbiological tests | animal | 31 | 1 | Listeria monocytogenes | 1 |
| | | N_A | Microbiological tests | animal | 1 | 1 | Listeria innocua | 1 |
| | Rabbits - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 58 | 8 | Listeria innocua | 4 |
| | | | | | | | Listeria monocytogenes | 4 |
| | Sheep - Farm - Not Available - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 2 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 44 | 0 | Listeria monocytogenes | 0 |
| Bihor | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 3 | 1 | Listeria monocytogenes | 1 |
| Bistriţa-Năsăud | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 3 | 0 | Listeria monocytogenes | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Goats - mixed herds - Farm - Not Available - animal sample - brain - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 4 | 0 | Listeria monocytogenes | 0 |
| Cluj | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 2 | 0 | Listeria monocytogenes | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 4 | 0 | Listeria monocytogenes | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Cluj | Cattle (bovine animals) - Farm - Not Available - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Deer - Veterinary clinics - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Goats - mixed herds - Farm - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 1 | 1 | Listeria monocytogenes | 1 |
| | | N_A | Microbiological tests | animal | 1 | 1 | Listeria innocua | 1 |
| | Sheep - Farm - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 9 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| Alba | All animals - farmed - Farm - Not Available - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 2 | 0 | Listeria monocytogenes | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 3 | 0 | Listeria monocytogenes | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 3 | 0 | Listeria monocytogenes | 0 |
| | | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Goats - mixed herds - Farm - Not Available - animal sample - brain - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 3 | 0 | Listeria monocytogenes | 0 |
| | Goats - mixed herds - Farm - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | | backyards | Microbiological tests | animal | 2 | 2 | Listeria innocua | 2 |
| | Goats - mixed herds - Farm - Not Available - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 2 | 0 | Listeria monocytogenes | 0 |
| | Goats - mixed herds - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 25 | 3 | Listeria innocua | 3 |
| | Sheep - Farm - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 2 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 10 | 0 | Listeria monocytogenes | 0 |
| Braşov | Minks - Farm - Not Available - animal sample - brain - Clinical investigations - Industry sampling - Suspect sampling | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| Covasna | Sheep - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 2 | 0 | Listeria monocytogenes | 0 |
| Harghita | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 2 | 1 | Listeria monocytogenes | 1 |
| Mureş | Bears - Zoo - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 5 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 5 | 0 | Listeria monocytogenes | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Mureş | Sheep - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| Sibiu | Sheep - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 2 | 0 | Listeria monocytogenes | 0 |
| Bacău | Pigs - Farm - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | N_A | Microbiological tests | animal | 1 | 1 | Listeria innocua | 1 |
| | | | | | | | Listeria monocytogenes | 1 |
| Botoşani | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 3 | 0 | Listeria monocytogenes | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 2 | 0 | Listeria monocytogenes | 0 |
| | | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Goats - mixed herds - Farm - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 3 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 2 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 3 | 0 | Listeria monocytogenes | 0 |
| Iaşi | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Dogs - Farm - Not Available - animal sample - brain - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Pigs - Farm - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 28 | 0 | Listeria monocytogenes | 0 |
| Neamţ | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 2 | 2 | Listeria monocytogenes | 2 |
| Vaslui | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| Brăila | Buffalos - Zoo - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 2 | 0 | Listeria monocytogenes | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 2 | 0 | Listeria monocytogenes | 0 |
| | Goats - mixed herds - Farm - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 2 | 0 | Listeria monocytogenes | 0 |
| | Goats - mixed herds - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Goats - Zoo - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 2 | 0 | Listeria monocytogenes | 0 |
| Buzău | Goats - mixed herds - Farm - Not Available - animal sample - brain - Clinical investigations - Industry sampling - Suspect sampling | N_A | Microbiological tests | animal | 4 | 0 | Listeria monocytogenes | 0 |
| Constanţa | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Microbiological tests | animal | 2 | 1 | Listeria welshimeri | 1 |
| Galaţi | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 4 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| Argeş | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| Călăraşi | Sheep - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 2 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| Dâmboviţa | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Dâmboviţa | Cattle (bovine animals) - Farm - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 7 | 0 | Listeria monocytogenes | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 6 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 9 | 0 | Listeria monocytogenes | 0 |
| Giurgiu | Cattle (bovine animals) - Farm - Not Available - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Goats - mixed herds - Farm - Not Available - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 2 | 0 | Listeria monocytogenes | 0 |
| Ialomiţa | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Goats - mixed herds - Farm - Not Available - animal sample - brain - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| Prahova | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | | N_A | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| Teleorman | Sheep - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| Ilfov | Cattle (bovine animals) - Farm - Not Available - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Goats - mixed herds - Farm - Not Available - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| Dolj | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| Olt | Chinchillas - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Goats - mixed herds - Farm - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 2 | 0 | Listeria monocytogenes | 0 |
| Vâlcea | Cats - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Goats - mixed herds - Farm - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Goats - mixed herds - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 2 | 0 | Listeria monocytogenes | 0 |
| | Rabbits - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| Hunedoara | Cattle (bovine animals) - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| Timiş | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - brain - Surveillance - Official sampling - Suspect sampling | backyards | Microbiological tests | animal | 3 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - foetus/stillbirth - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 4 | 0 | Listeria monocytogenes | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Clinical investigations - Industry sampling - Suspect sampling | animal from backyards | Microbiological tests | animal | 1 | 0 | Listeria monocytogenes | 0 |

## Table Listeria:LISTERIA in food

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Bakery products - desserts - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 15 | 0 | detection | Listeria monocytogenes | 15 | 0 |
| | Bakery products - desserts - Processing plant - European Union - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 3 | 0 | detection | Listeria monocytogenes | 3 | 0 |
| | Bakery products - desserts - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 248 | 0 | detection | Listeria monocytogenes | 248 | 0 |
| | Bakery products - desserts - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 60 | 0 | <= 100 | Listeria monocytogenes | 25 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 25 | 0 |
| | Bakery products - desserts - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 60 | 0 | detection | Listeria monocytogenes | 35 | 0 |
| | Bakery products - desserts - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 15 | 0 | detection | Listeria monocytogenes | 15 | 0 |
| | Bakery products - desserts - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 25 | Gram | N_A | 10 | 0 | detection | Listeria monocytogenes | 10 | 0 |
| | Bakery products - desserts - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 190 | 0 | <= 100 | Listeria monocytogenes | 135 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 135 | 0 |
| | Bakery products - desserts - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 190 | 0 | detection | Listeria monocytogenes | 55 | 0 |
| | Bakery products - desserts - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 10 | 0 | detection | Listeria monocytogenes | 10 | 0 |
| | Bakery products - pastry - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 8 | 0 | detection | Listeria monocytogenes | 8 | 0 |
| | Bakery products - pastry - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 25 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Bakery products - pastry - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 25 | 0 | detection | Listeria monocytogenes | 20 | 0 |
| | Bakery products - pastry - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Bakery products - pastry - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 32 | 0 | <= 100 | Listeria monocytogenes | 30 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 30 | 0 |
| | Bakery products - pastry - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 32 | 0 | detection | Listeria monocytogenes | 2 | 0 |
| | Bakery products - pastry - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 60 | 0 | detection | Listeria monocytogenes | 60 | 0 |
| | Bakery products - pastry - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 10 | 0 | detection | Listeria monocytogenes | 10 | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 6 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 6 | 0 | detection | Listeria monocytogenes | 1 | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 35 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 10 | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Cheeses made from cows' milk - hard - made from pasteurised milk - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 35 | 0 | detection | Listeria monocytogenes | 25 | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 1761 | 0 | <= 100 | Listeria monocytogenes | 100 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 100 | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 1761 | 0 | detection | Listeria monocytogenes | 1,661 | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 95 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 95 | 0 | detection | Listeria monocytogenes | 85 | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 10 | Gram | N_A | 4 | 0 | <= 100 | Listeria monocytogenes | 4 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 4 | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 56 | 0 | <= 100 | Listeria monocytogenes | 20 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 20 | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 56 | 0 | detection | Listeria monocytogenes | 36 | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 60 | 0 | <= 100 | Listeria monocytogenes | 20 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 20 | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 60 | 0 | detection | Listeria monocytogenes | 40 | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 150 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 150 | 0 | detection | Listeria monocytogenes | 145 | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 10 | Gram | N_A | 15 | 0 | <= 100 | Listeria monocytogenes | 15 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 15 | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 20 | 0 | detection | Listeria monocytogenes | 20 | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 18 | 0 | detection | Listeria monocytogenes | 18 | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 100 | 8 | <= 100 | Listeria monocytogenes - molecular serogroup IVb | 100 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes - molecular serogroup IVb | 100 | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 100 | 8 | detection | Listeria monocytogenes - molecular serogroup IVb | 100 | 8 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 15 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 15 | 0 | detection | Listeria monocytogenes | 10 | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 1415 | 4 | <= 100 | Listeria monocytogenes - molecular serogroup IIa | 20 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes - molecular serogroup IIa | 20 | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 1415 | 4 | detection | Listeria monocytogenes - molecular serogroup IIa | 1,395 | 4 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 20 | 0 | detection | Listeria monocytogenes | 20 | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 81 | 0 | <= 100 | Listeria monocytogenes | 20 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 20 | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 81 | 0 | detection | Listeria monocytogenes | 61 | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 315 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 315 | 0 | detection | Listeria monocytogenes | 305 | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 131 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 131 | 0 | detection | Listeria monocytogenes | 126 | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 169 | 0 | detection | Listeria monocytogenes | 169 | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 95 | 0 | detection | Listeria monocytogenes | 95 | 0 |
| | Cheeses made from goats' milk - hard - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 70 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from goats' milk - hard - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 70 | 0 | detection | Listeria monocytogenes | 65 | 0 |
| | Cheeses made from goats' milk - hard - made from pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from goats' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from goats' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from goats' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Suspect sampling | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from sheep's milk - hard - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 95 | 0 | detection | Listeria monocytogenes | 95 | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Cheeses made from sheep's milk - hard - made from pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Suspect sampling | single (food/feed) | 25 | Gram | N_A | 1 | 0 | detection | Listeria monocytogenes | 1 | 0 |
| | Cheeses made from sheep's milk - hard - made from raw or low heat-treated milk - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from sheep's milk - hard - made from raw or low heat-treated milk - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 10 | Gram | N_A | 1 | 0 | <= 100 | Listeria monocytogenes | 1 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 1 | 0 |
| | Cheeses made from sheep's milk - hard - made from raw or low heat-treated milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from sheep's milk - hard - made from raw or low heat-treated milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 15 | 0 | detection | Listeria monocytogenes | 15 | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 15 | 0 | detection | Listeria monocytogenes | 15 | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 10 | 0 | detection | Listeria monocytogenes | 10 | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 106 | 0 | detection | Listeria monocytogenes | 106 | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 20 | 0 | detection | Listeria monocytogenes | 20 | 0 |
| | Dairy products (excluding cheeses) - butter - made from pasteurised milk - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 10 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| | Dairy products (excluding cheeses) - butter - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 132 | 0 | detection | Listeria monocytogenes | 132 | 0 |
| | Dairy products (excluding cheeses) - butter - made from pasteurised milk - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 10 | Gram | N_A | 1 | 0 | <= 100 | Listeria monocytogenes | 1 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 1 | 0 |
| | Dairy products (excluding cheeses) - butter - made from pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Dairy products (excluding cheeses) - cream - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 99 | 0 | <= 100 | Listeria monocytogenes | 99 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 99 | 0 |
| | Dairy products (excluding cheeses) - cream - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 99 | 0 | detection | Listeria monocytogenes | 99 | 0 |
| | Dairy products (excluding cheeses) - cream - made from raw or low heat-treated milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Dairy products (excluding cheeses) - dairy products, not specified - ready-to-eat - made from pasteurised milk - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 30 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| | Dairy products (excluding cheeses) - dairy products, not specified - ready-to-eat - made from pasteurised milk - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 30 | 0 | detection | Listeria monocytogenes | 20 | 0 |
| | Dairy products (excluding cheeses) - dairy products, not specified - ready-to-eat - made from pasteurised milk - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Dairy products (excluding cheeses) - dairy products, not specified - ready-to-eat - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 3377 | 0 | <= 100 | Listeria monocytogenes | 290 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 290 | 0 |
| | Dairy products (excluding cheeses) - dairy products, not specified - ready-to-eat - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 3377 | 0 | detection | Listeria monocytogenes | 3,087 | 0 |
| | Dairy products (excluding cheeses) - dairy products, not specified - ready-to-eat - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 25 | Gram | N_A | 25 | 0 | detection | Listeria monocytogenes | 25 | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Dairy products (excluding cheeses) - dairy products, not specified - ready-to-eat - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Suspect sampling | single (food/feed) | 10 | Gram | N_A | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Dairy products (excluding cheeses) - dairy products, not specified - ready-to-eat - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 10 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| | Dairy products (excluding cheeses) - dairy products, not specified - ready-to-eat - made from pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 55 | 0 | detection | Listeria monocytogenes | 55 | 0 |
| | Dairy products (excluding cheeses) - dairy products, not specified - ready-to-eat - made from pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 100 | 0 | detection | Listeria monocytogenes | 100 | 0 |
| | Dairy products (excluding cheeses) - dairy products, not specified - ready-to-eat - made from raw or low heat-treated milk - Packing centre - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Suspect sampling | single (food/feed) | 25 | Gram | N_A | 1 | 0 | detection | Listeria monocytogenes | 1 | 0 |
| | Dairy products (excluding cheeses) - dairy products, not specified - ready-to-eat - made from raw or low heat-treated milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 20 | 0 | detection | Listeria monocytogenes | 20 | 0 |
| | Dairy products (excluding cheeses) - ice-cream - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Dairy products (excluding cheeses) - ice-cream - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 35 | 0 | detection | Listeria monocytogenes | 35 | 0 |
| | Dairy products (excluding cheeses) - ice-cream - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 25 | Gram | N_A | 25 | 0 | detection | Listeria monocytogenes | 25 | 0 |
| | Dairy products (excluding cheeses) - ice-cream - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 1 | 0 | detection | Listeria monocytogenes | 1 | 0 |
| | Dairy products (excluding cheeses) - ice-cream - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 10 | Gram | N_A | 1 | 0 | <= 100 | Listeria monocytogenes | 1 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 1 | 0 |
| | Dairy products (excluding cheeses) - ice-cream - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Dairy products (excluding cheeses) - ice-cream - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Egg products - non-ready-to-eat - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 15 | 0 | detection | Listeria monocytogenes | 15 | 0 |
| | Fishery products, unspecified - non-ready-to-eat - Catering - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 80 | 0 | <= 100 | Listeria monocytogenes | 25 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 25 | 0 |
| | Fishery products, unspecified - non-ready-to-eat - Catering - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 80 | 0 | detection | Listeria monocytogenes | 55 | 0 |
| | Fishery products, unspecified - non-ready-to-eat - Catering - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 65 | 0 | <= 100 | Listeria monocytogenes | 55 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 55 | 0 |
| | Fishery products, unspecified - non-ready-to-eat - Catering - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 65 | 0 | detection | Listeria monocytogenes | 10 | 0 |
| | Fishery products, unspecified - non-ready-to-eat - Processing plant - Non European Union - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Fishery products, unspecified - non-ready-to-eat - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 156 | 0 | <= 100 | Listeria monocytogenes | 26 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 26 | 0 |
| | Fishery products, unspecified - non-ready-to-eat - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 156 | 0 | detection | Listeria monocytogenes | 130 | 0 |
| | Fishery products, unspecified - non-ready-to-eat - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Fishery products, unspecified - non-ready-to-eat - Processing plant - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 20 | 0 | <= 100 | Listeria monocytogenes | 20 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 20 | 0 |
| | Fishery products, unspecified - non-ready-to-eat - Retail - European Union - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Fishery products, unspecified - non-ready-to-eat - Retail - Non European Union - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Fishery products, unspecified - non-ready-to-eat - Retail - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 52 | 0 | <= 100 | Listeria monocytogenes | 15 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 15 | 0 |
| | Fishery products, unspecified - non-ready-to-eat - Retail - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 52 | 0 | detection | Listeria monocytogenes | 37 | 0 |
| | Fishery products, unspecified - non-ready-to-eat - Retail - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 70 | 0 | detection | Listeria monocytogenes | 70 | 0 |
| | Fishery products, unspecified - raw - Conservation facilities - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 2 | 0 | <= 100 | Listeria monocytogenes | 2 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 2 | 0 |
| | Fishery products, unspecified - raw - Farm - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 1 | 0 | detection | Listeria monocytogenes | 1 | 0 |
| | Fishery products, unspecified - raw - Retail - European Union - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 3 | 0 | detection | Listeria monocytogenes | 3 | 0 |
| | Fishery products, unspecified - raw - Retail - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 62 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| | Fishery products, unspecified - raw - Retail - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 62 | 0 | detection | Listeria monocytogenes | 52 | 0 |
| | Fishery products, unspecified - ready-to-eat - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 5 | 1 | detection | Listeria monocytogenes - molecular serogroup IIa | 5 | 1 |
| | Fishery products, unspecified - ready-to-eat - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 75 | 6 | detection | Listeria monocytogenes - molecular serogroup IIa | 75 | 6 |
| | Fishery products, unspecified - ready-to-eat - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 10 | Gram | N_A | 35 | 0 | <= 100 | Listeria monocytogenes | 35 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 35 | 0 |
| | Fishery products, unspecified - ready-to-eat - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 25 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Fishery products, unspecified - ready-to-eat - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 25 | 0 | detection | Listeria monocytogenes | 20 | 0 |
| | Fishery products, unspecified - ready-to-eat - Retail - Non European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 10 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| | Fishery products, unspecified - ready-to-eat - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 111 | 2 | <= 100 | Listeria monocytogenes - molecular serogroup IVa | 30 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes - molecular serogroup IVa | 30 | 0 |
| | Fishery products, unspecified - ready-to-eat - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 111 | 2 | detection | Listeria monocytogenes - molecular serogroup IVa | 81 | 2 |
| | Fishery products, unspecified - ready-to-eat - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 60 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Fishery products, unspecified - ready-to-eat - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 60 | 0 | detection | Listeria monocytogenes | 55 | 0 |
| | Foodstuffs intended for special nutritional uses - dietary foods for special medical purposes - Conservation facilities - European Union - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 25 | Gram | N_A | 10 | 0 | detection | Listeria monocytogenes | 10 | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Foodstuffs intended for special nutritional uses - dietary foods for special medical purposes - Processing plant - European Union - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 25 | Gram | N_A | 7 | 0 | detection | Listeria monocytogenes | 7 | 0 |
| | Foodstuffs intended for special nutritional uses - dried dietary foods for special medical purposes intended for infants below 6 months - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 100 | 0 | <= 100 | Listeria monocytogenes | 100 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 100 | 0 |
| | Fruits and vegetables - non-pre-cut - Retail - European Union - food sample - Monitoring - Official sampling - Suspect sampling | batch (food/feed) | 10 | Gram | Frozen fruits. Samples taken as a result of multi-country outbreak of Listeria monocytogenes | 10 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| | Meat from bovine animals - fresh - Catering - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 30 | 0 | <= 100 | Listeria monocytogenes | 30 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 30 | 0 |
| | Meat from bovine animals - fresh - Cutting plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 6 | 0 | detection | Listeria monocytogenes | 6 | 0 |
| | Meat from bovine animals - fresh - Packing centre - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 16 | 0 | detection | Listeria monocytogenes | 16 | 0 |
| | Meat from bovine animals - fresh - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 90 | 12 | detection | Listeria monocytogenes - molecular serogroup IIa | 90 | 2 |
| | | | | | | | | | Listeria monocytogenes - molecular serogroup IIb | 90 | 1 |
| | | | | | | | | | Listeria monocytogenes - molecular serogroup IVb | 90 | 9 |
| | Meat from bovine animals - fresh - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | 20 | 8 | detection | Listeria monocytogenes - molecular serogroup IIa | 20 | 8 |
| | Meat from bovine animals - fresh - Slaughterhouse - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 50 | 0 | detection | Listeria monocytogenes | 50 | 0 |
| | Meat from bovine animals - meat preparation - intended to be eaten cooked - Catering - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 10 | 0 | detection | Listeria monocytogenes | 10 | 0 |
| | Meat from bovine animals - meat preparation - intended to be eaten cooked - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Meat from bovine animals - meat products - cooked, ready-to-eat - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 100 | 0 | <= 100 | Listeria monocytogenes | 55 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 55 | 0 |
| | Meat from bovine animals - meat products - cooked, ready-to-eat - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 100 | 0 | detection | Listeria monocytogenes | 45 | 0 |
| | Meat from bovine animals - meat products - cooked, ready-to-eat - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 215 | 0 | <= 100 | Listeria monocytogenes | 205 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 205 | 0 |
| | Meat from bovine animals - meat products - cooked, ready-to-eat - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 215 | 0 | detection | Listeria monocytogenes | 10 | 0 |
| | Meat from bovine animals - meat products - cooked, ready-to-eat - Packing centre - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 15 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| | Meat from bovine animals - meat products - cooked, ready-to-eat - Packing centre - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 15 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Meat from bovine animals - meat products - cooked, ready-to-eat - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Meat from bovine animals - minced meat - intended to be eaten cooked - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 25 | 0 | detection | Listeria monocytogenes | 25 | 0 |
| | Meat from bovine animals - minced meat - intended to be eaten cooked - Retail - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Meat from broilers (Gallus gallus) - fresh - Catering - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 115 | 0 | <= 100 | Listeria monocytogenes | 115 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 115 | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Meat from broilers (Gallus gallus) - fresh - Conservation facilities - European Union - food sample - Surveillance - HACCP and own check - Other | batch (food/feed) | 25 | Gram | N_A | 1 | 0 | detection | Listeria monocytogenes | 1 | 0 |
| | Meat from broilers (Gallus gallus) - fresh - Conservation facilities - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 10 | 0 | detection | Listeria monocytogenes | 10 | 0 |
| | Meat from broilers (Gallus gallus) - fresh - Cutting plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Meat from broilers (Gallus gallus) - fresh - Packing centre - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Meat from broilers (Gallus gallus) - fresh - Packing centre - Romania - food sample - Surveillance - HACCP and own check - Other | batch (food/feed) | 25 | Gram | N_A | 3 | 0 | detection | Listeria monocytogenes | 3 | 0 |
| | Meat from broilers (Gallus gallus) - fresh - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 15 | 1 | detection | Listeria monocytogenes - molecular serogroup IIa | 15 | 1 |
| | Meat from broilers (Gallus gallus) - fresh - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Other | batch (food/feed) | 25 | Gram | N_A | 10 | 0 | detection | Listeria monocytogenes | 10 | 0 |
| | Meat from broilers (Gallus gallus) - fresh - Retail - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 7 | 2 | <= 100 | Listeria monocytogenes - molecular serogroup IIa | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes - molecular serogroup IIa | 5 | 0 |
| | Meat from broilers (Gallus gallus) - fresh - Retail - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 7 | 2 | detection | Listeria monocytogenes - molecular serogroup IIa | 2 | 2 |
| | Meat from broilers (Gallus gallus) - fresh - Retail - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Meat from broilers (Gallus gallus) - fresh - Slaughterhouse - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 35 | 0 | <= 100 | Listeria monocytogenes | 17 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 17 | 0 |
| | Meat from broilers (Gallus gallus) - fresh - Slaughterhouse - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 35 | 0 | detection | Listeria monocytogenes | 18 | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 810 | 0 | <= 100 | Listeria monocytogenes | 485 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 485 | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 810 | 0 | detection | Listeria monocytogenes | 325 | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Suspect sampling | single (food/feed) | 10 | Gram | N_A | 1 | 0 | <= 100 | Listeria monocytogenes | 1 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 1 | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 1100 | 0 | <= 100 | Listeria monocytogenes | 1,060 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 1,060 | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 1100 | 0 | detection | Listeria monocytogenes | 40 | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 10 | 0 | detection | Listeria monocytogenes | 10 | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Packing centre - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 10 | 0 | detection | Listeria monocytogenes | 10 | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Packing centre - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 15 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Packing centre - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 15 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 75 | 0 | detection | Listeria monocytogenes | 75 | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 135 | 0 | <= 100 | Listeria monocytogenes | 115 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 115 | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 135 | 0 | detection | Listeria monocytogenes | 20 | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 90 | 0 | detection | Listeria monocytogenes | 90 | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 25 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 25 | 0 | detection | Listeria monocytogenes | 15 | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 25 | Gram | N_A | 1 | 0 | detection | Listeria monocytogenes | 1 | 0 |
| | Meat from broilers (Gallus gallus) - mechanically separated meat (MSM) - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 20 | 0 | detection | Listeria monocytogenes | 20 | 0 |
| | Meat from other animal species or not specified - meat preparation - intended to be eaten cooked - Conservation facilities - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 10 | 0 | detection | Listeria monocytogenes | 10 | 0 |
| | Meat from other animal species or not specified - meat preparation - intended to be eaten cooked - Cutting plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 25 | 0 | detection | Listeria monocytogenes | 25 | 0 |
| | Meat from other animal species or not specified - meat preparation - intended to be eaten cooked - Cutting plant - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Meat from other animal species or not specified - meat preparation - intended to be eaten cooked - Packing centre - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 26 | 0 | <= 100 | Listeria monocytogenes | 1 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 1 | 0 |
| | Meat from other animal species or not specified - meat preparation - intended to be eaten cooked - Packing centre - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 26 | 0 | detection | Listeria monocytogenes | 25 | 0 |
| | Meat from other animal species or not specified - meat preparation - intended to be eaten cooked - Packing centre - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Meat from other animal species or not specified - meat preparation - intended to be eaten cooked - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 255 | 0 | detection | Listeria monocytogenes | 255 | 0 |
| | Meat from other animal species or not specified - meat preparation - intended to be eaten cooked - Processing plant - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 15 | 0 | <= 100 | Listeria monocytogenes | 15 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 15 | 0 |
| | Meat from other animal species or not specified - meat preparation - intended to be eaten cooked - Retail - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 8 | 0 | <= 100 | Listeria monocytogenes | 2 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 2 | 0 |
| | Meat from other animal species or not specified - meat preparation - intended to be eaten cooked - Retail - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 8 | 0 | detection | Listeria monocytogenes | 6 | 0 |
| | Meat from other animal species or not specified - meat products - raw and intended to be eaten raw - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 4 | 0 | detection | Listeria monocytogenes | 4 | 0 |
| | Meat from other animal species or not specified - meat products - raw and intended to be eaten raw - Cutting plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 15 | 0 | detection | Listeria monocytogenes | 15 | 0 |
| | Meat from other animal species or not specified - meat products - raw and intended to be eaten raw - Cutting plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 35 | 0 | detection | Listeria monocytogenes | 35 | 0 |
| | Meat from other animal species or not specified - meat products - raw and intended to be eaten raw - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 331 | 1 | <= 100 | Listeria monocytogenes - molecular serogroup IIa | 15 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes - molecular serogroup IIa | 15 | 0 |
| | Meat from other animal species or not specified - meat products - raw and intended to be eaten raw - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 331 | 1 | detection | Listeria monocytogenes - molecular serogroup IIa | 316 | 1 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Meat from other animal species or not specified - meat products - raw and intended to be eaten raw - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 30 | 6 | detection | Listeria monocytogenes - molecular serogroup IIa | 30 | 1 |
| | | | | | | | | | Listeria monocytogenes - molecular serogroup IVb | 30 | 5 |
| | Meat from other animal species or not specified - meat products - raw and intended to be eaten raw - Retail - European Union - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 25 | 0 | detection | Listeria monocytogenes | 25 | 0 |
| | Meat from other animal species or not specified - meat products - raw and intended to be eaten raw - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 49 | 0 | <= 100 | Listeria monocytogenes | 44 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 44 | 0 |
| | Meat from other animal species or not specified - meat products - raw and intended to be eaten raw - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 49 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Meat from other animal species or not specified - meat products - raw and intended to be eaten raw - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 60 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| | Meat from other animal species or not specified - meat products - raw and intended to be eaten raw - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 60 | 0 | detection | Listeria monocytogenes | 50 | 0 |
| | Meat from other animal species or not specified - minced meat - intended to be eaten cooked - Conservation facilities - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 50 | 0 | detection | Listeria monocytogenes | 50 | 0 |
| | Meat from other animal species or not specified - minced meat - intended to be eaten cooked - Packing centre - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 11 | 0 | detection | Listeria monocytogenes | 11 | 0 |
| | Meat from other animal species or not specified - minced meat - intended to be eaten cooked - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 100 | 0 | detection | Listeria monocytogenes | 100 | 0 |
| | Meat from other animal species or not specified - minced meat - intended to be eaten cooked - Retail - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 25 | 0 | detection | Listeria monocytogenes | 25 | 0 |
| | Meat from other animal species or not specified - minced meat - intended to be eaten cooked - Retail - Romania - food sample - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Meat from pig - fresh - Conservation facilities - European Union - food sample - Surveillance - HACCP and own check - Other | batch (food/feed) | 10 | Gram | N_A | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Meat from pig - fresh - Cutting plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Meat from pig - fresh - Packing centre - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 20 | 0 | detection | Listeria monocytogenes | 20 | 0 |
| | Meat from pig - fresh - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 157 | 15 | <= 100 | Listeria monocytogenes - molecular serogroup IIa | 40 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes - molecular serogroup IIa | 40 | 0 |
| | Meat from pig - fresh - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 157 | 15 | detection | Listeria monocytogenes - molecular serogroup IIa | 117 | 15 |
| | Meat from pig - fresh - Processing plant - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Meat from pig - fresh - Retail - European Union - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 1 | 0 | detection | Listeria monocytogenes | 1 | 0 |
| | Meat from pig - fresh - Retail - European Union - food sample - Surveillance - HACCP and own check - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | 4 | 1 | detection | Listeria monocytogenes - molecular serogroup IIb | 4 | 1 |
| | Meat from pig - fresh - Retail - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Meat from pig - fresh - Slaughterhouse - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 198 | 0 | <= 100 | Listeria monocytogenes | 63 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 63 | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Meat from pig - fresh - Slaughterhouse - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 198 | 0 | detection | Listeria monocytogenes | 135 | 0 |
| | Meat from pig - meat preparation - intended to be eaten cooked - Cutting plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Meat from pig - meat preparation - intended to be eaten cooked - Packing centre - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Meat from pig - meat preparation - intended to be eaten cooked - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 48 | 0 | <= 100 | Listeria monocytogenes | 3 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 3 | 0 |
| | Meat from pig - meat preparation - intended to be eaten cooked - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 48 | 0 | detection | Listeria monocytogenes | 45 | 0 |
| | Meat from pig - meat preparation - intended to be eaten cooked - Processing plant - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Meat from pig - meat preparation - intended to be eaten cooked - Retail - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Meat from pig - meat preparation - intended to be eaten cooked - Retail - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 25 | 0 | <= 100 | Listeria monocytogenes | 15 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 15 | 0 |
| | Meat from pig - meat preparation - intended to be eaten cooked - Retail - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 25 | 0 | detection | Listeria monocytogenes | 10 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 530 | 0 | <= 100 | Listeria monocytogenes | 410 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 410 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 530 | 0 | detection | Listeria monocytogenes | 120 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 590 | 0 | <= 100 | Listeria monocytogenes | 590 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 590 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 25 | Gram | N_A | 10 | 0 | detection | Listeria monocytogenes | 10 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Conservation facilities - European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 10 | 0 | detection | Listeria monocytogenes | 10 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 45 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 45 | 0 | detection | Listeria monocytogenes | 40 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 25 | 0 | <= 100 | Listeria monocytogenes | 15 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 15 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 25 | 0 | detection | Listeria monocytogenes | 10 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Cutting plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 215 | 0 | <= 100 | Listeria monocytogenes | 215 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 215 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Cutting plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 25 | Gram | N_A | 21 | 0 | detection | Listeria monocytogenes | 21 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Cutting plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 70 | 0 | <= 100 | Listeria monocytogenes | 20 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 20 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Cutting plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 70 | 0 | detection | Listeria monocytogenes | 50 | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Meat from pig - meat products - cooked, ready-to-eat - Packing centre - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 255 | 10 | detection | Listeria monocytogenes - molecular serogroup IIa | 255 | 5 |
| | | | | | | | | | Listeria monocytogenes - molecular serogroup IIc | 255 | 5 |
| | Meat from pig - meat products - cooked, ready-to-eat - Packing centre - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 355 | 0 | detection | Listeria monocytogenes | 355 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 1741 | 22 | detection | Listeria monocytogenes - molecular serogroup IIa | 1,741 | 8 |
| | | | | | | | | | Listeria monocytogenes - molecular serogroup IIc | 1,741 | 13 |
| | | | | | | | | | Listeria monocytogenes - molecular serogroup IVa | 1,741 | 1 |
| | Meat from pig - meat products - cooked, ready-to-eat - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 10 | Gram | N_A | 195 | 0 | <= 100 | Listeria monocytogenes | 145 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 145 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 25 | Gram | N_A | 195 | 0 | detection | Listeria monocytogenes | 50 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 365 | 0 | <= 100 | Listeria monocytogenes | 50 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 50 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 365 | 0 | detection | Listeria monocytogenes | 315 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 140 | 0 | detection | Listeria monocytogenes | 140 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 25 | Gram | N_A | 10 | 0 | detection | Listeria monocytogenes | 10 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 10 | Gram | N_A | 1 | 0 | <= 100 | Listeria monocytogenes | 1 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 1 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 588 | 0 | <= 100 | Listeria monocytogenes | 169 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 169 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 588 | 0 | detection | Listeria monocytogenes | 419 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 25 | Gram | N_A | 30 | 0 | detection | Listeria monocytogenes | 30 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 735 | 0 | <= 100 | Listeria monocytogenes | 390 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 390 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 735 | 0 | detection | Listeria monocytogenes | 345 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Slaughterhouse - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 50 | 0 | detection | Listeria monocytogenes | 50 | 0 |
| | Meat from pig - minced meat - intended to be eaten cooked - Packing centre - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 2 | 0 | <= 100 | Listeria monocytogenes | 2 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 2 | 0 |
| | Meat from pig - minced meat - intended to be eaten cooked - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 20 | 5 | detection | Listeria monocytogenes - molecular serogroup IIa | 20 | 5 |
| | Meat from pig - minced meat - intended to be eaten cooked - Retail - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 10 | Gram | N_A | 70 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Meat from pig - minced meat - intended to be eaten cooked - Retail - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 70 | 0 | detection | Listeria monocytogenes | 65 | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Meat from pig - offal - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | 25 | 6 | detection | Listeria monocytogenes - molecular serogroup IIa | 25 | 1 |
| | | | | | | | | | Listeria monocytogenes - molecular serogroup IIc | 25 | 3 |
| | | | | | | | | | Listeria monocytogenes - molecular serogroup IVa | 25 | 1 |
| | | | | | | | | | Listeria monocytogenes - molecular serogroup IVb | 25 | 1 |
| | Meat from wild game - land mammals - meat products - cooked, ready-to-eat - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 20 | 3 | <= 100 | Listeria monocytogenes - molecular serogroup IIa | 20 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes - molecular serogroup IIa | 20 | 3 |
| | Milk, cows' - pasteurised milk - Packing centre - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 25 | Millilitre | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Milk, cows' - pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Millilitre | N_A | 236 | 0 | detection | Listeria monocytogenes | 236 | 0 |
| | Milk, cows' - pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 10 | Millilitre | N_A | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Milk, cows' - pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Millilitre | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Milk, cows' - pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Millilitre | N_A | 120 | 0 | detection | Listeria monocytogenes | 120 | 0 |
| | Milk, cows' - raw milk - intended for direct human consumption - Automatic distribution system for raw milk - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 10 | Millilitre | N_A | 3 | 0 | <= 100 | Listeria monocytogenes | 3 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 3 | 0 |
| | Milk, cows' - raw milk - intended for direct human consumption - Farm - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Millilitre | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Milk, cows' - raw milk for manufacture - intended for manufacture of pasteurised/UHT products - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Millilitre | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Milk, cows' - raw milk for manufacture - intended for manufacture of raw or low heat-treated products - Farm - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Millilitre | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Other food of non-animal origin - Conservation facilities - European Union - food sample - Monitoring - HACCP and own check - Suspect sampling | batch (food/feed) | 25 | Gram | Frozen vegetables.Samples taken as a result of multi-country outbreak of Listeria monocytogenes | 45 | 0 | detection | Listeria monocytogenes | 45 | 0 |
| | Other food of non-animal origin - Conservation facilities - Hungary - food sample - Monitoring - HACCP and own check - Suspect sampling | batch (food/feed) | 25 | Gram | Frozen vegetables.Samples taken as a result of multi-country outbreak of Listeria monocytogenes | 130 | 65 | detection | Listeria monocytogenes - molecular serogroup IIa | 130 | 65 |
| | Other food of non-animal origin - Processing plant - Belgium - food sample - Monitoring - HACCP and own check - Suspect sampling | batch (food/feed) | 25 | Gram | Frozen vegetables.Samples taken as a result of multi-country outbreak of Listeria monocytogenes | 15 | 0 | detection | Listeria monocytogenes | 15 | 0 |
| | Other food of non-animal origin - Processing plant - Romania - food sample - Monitoring - HACCP and own check - Suspect sampling | batch (food/feed) | 25 | Gram | Frozen vegetables.Samples taken as a result of multi-country outbreak of Listeria monocytogenes | 26 | 3 | detection | Listeria monocytogenes - molecular serogroup IIa | 26 | 2 |
| | | | | | | | | | Listeria monocytogenes - molecular serogroup IIc | 26 | 1 |
| | Other food of non-animal origin - Retail - European Union - food sample - Monitoring - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | Frozen vegetables.Samples taken as a result of multi-country outbreak of Listeria monocytogenes | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Other food of non-animal origin - Retail - Hungary - food sample - Monitoring - Official sampling - Suspect sampling | batch (food/feed) | 10 | Gram | Frozen vegetables.Samples taken as a result of multi-country outbreak of Listeria monocytogenes | 22 | 0 | <= 100 | Listeria monocytogenes | 22 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 22 | 0 |
| | Other food of non-animal origin - Retail - Poland - food sample - Monitoring - Official sampling - Suspect sampling | batch (food/feed) | 10 | Gram | Frozen vegetables.Samples taken as a result of multi-country outbreak of Listeria monocytogenes | 8 | 0 | <= 100 | Listeria monocytogenes | 3 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 3 | 0 |
| | Other food of non-animal origin - Retail - Poland - food sample - Monitoring - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | Frozen vegetables.Samples taken as a result of multi-country outbreak of Listeria monocytogenes | 8 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Other food of non-animal origin - Retail - Romania - food sample - Monitoring - Official sampling - Suspect sampling | batch (food/feed) | 10 | Gram | Frozen vegetables.Samples taken as a result of multi-country outbreak of Listeria monocytogenes | 20 | 0 | <= 100 | Listeria monocytogenes | 15 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 15 | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Other food of non-animal origin - Retail - Romania - food sample - Monitoring - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | Frozen vegetables.Samples taken as a result of multi-country outbreak of Listeria monocytogenes | 20 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Other processed food products and prepared dishes - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 4568 | 5 | <= 100 | Listeria monocytogenes - molecular serogroup IIc | 1,505 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes - molecular serogroup IIc | 1,505 | 0 |
| | Other processed food products and prepared dishes - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 4568 | 5 | detection | Listeria monocytogenes - molecular serogroup IIc | 3,063 | 5 |
| | Other processed food products and prepared dishes - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 10 | Gram | N_A | 50 | 0 | <= 100 | Listeria monocytogenes | 50 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 50 | 0 |
| | Other processed food products and prepared dishes - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 4763 | 0 | <= 100 | Listeria monocytogenes | 2,705 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 2,705 | 0 |
| | Other processed food products and prepared dishes - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 4763 | 0 | detection | Listeria monocytogenes | 2,058 | 0 |
| | Other processed food products and prepared dishes - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 25 | Gram | N_A | 2 | 0 | detection | Listeria monocytogenes | 2 | 0 |
| | Other processed food products and prepared dishes - Conservation facilities - Non European Union - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Other processed food products and prepared dishes - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 15 | 0 | detection | Listeria monocytogenes | 15 | 0 |
| | Other processed food products and prepared dishes - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 25 | Gram | N_A | 40 | 0 | detection | Listeria monocytogenes | 40 | 0 |
| | Other processed food products and prepared dishes - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Other processed food products and prepared dishes - fish and seafood based dishes - Processing plant - Non European Union - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 55 | 0 | detection | Listeria monocytogenes | 55 | 0 |
| | Other processed food products and prepared dishes - Hospital or medical care facility - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 230 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| | Other processed food products and prepared dishes - Hospital or medical care facility - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 230 | 0 | detection | Listeria monocytogenes | 220 | 0 |
| | Other processed food products and prepared dishes - Hospital or medical care facility - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Other processed food products and prepared dishes - Packing centre - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 50 | 0 | <= 100 | Listeria monocytogenes | 20 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 20 | 0 |
| | Other processed food products and prepared dishes - Packing centre - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 50 | 0 | detection | Listeria monocytogenes | 30 | 0 |
| | Other processed food products and prepared dishes - Packing centre - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 10 | Gram | N_A | 20 | 0 | <= 100 | Listeria monocytogenes | 20 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 20 | 0 |
| | Other processed food products and prepared dishes - Packing centre - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 25 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Other processed food products and prepared dishes - Packing centre - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 25 | 0 | detection | Listeria monocytogenes | 20 | 0 |
| | Other processed food products and prepared dishes - Packing centre - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 25 | Gram | N_A | 1 | 0 | detection | Listeria monocytogenes | 1 | 0 |
| | Other processed food products and prepared dishes - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 1308 | 0 | <= 100 | Listeria monocytogenes | 481 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 481 | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Other processed food products and prepared dishes - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 1308 | 0 | detection | Listeria monocytogenes | 1,268 | 0 |
| | Other processed food products and prepared dishes - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 182 | 0 | <= 100 | Listeria monocytogenes | 30 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 30 | 0 |
| | Other processed food products and prepared dishes - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 182 | 0 | detection | Listeria monocytogenes | 152 | 0 |
| | Other processed food products and prepared dishes - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 2499 | 0 | <= 100 | Listeria monocytogenes | 1,292 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 1,292 | 0 |
| | Other processed food products and prepared dishes - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 2499 | 0 | detection | Listeria monocytogenes | 1,207 | 0 |
| | Other processed food products and prepared dishes - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 10 | Gram | N_A | 100 | 0 | <= 100 | Listeria monocytogenes | 25 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 25 | 0 |
| | Other processed food products and prepared dishes - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 25 | Gram | N_A | 100 | 0 | detection | Listeria monocytogenes | 75 | 0 |
| | Other processed food products and prepared dishes - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Suspect sampling | single (food/feed) | 25 | Gram | N_A | 75 | 0 | detection | Listeria monocytogenes | 75 | 0 |
| | Other processed food products and prepared dishes - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 750 | 0 | <= 100 | Listeria monocytogenes | 580 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 580 | 0 |
| | Other processed food products and prepared dishes - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 750 | 0 | detection | Listeria monocytogenes | 170 | 0 |
| | Other processed food products and prepared dishes - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 25 | Gram | N_A | 15 | 0 | detection | Listeria monocytogenes | 15 | 0 |
| | Other processed food products and prepared dishes - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | 1926 | 0 | <= 100 | Listeria monocytogenes | 1,242 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes | 1,242 | 0 |
| | Other processed food products and prepared dishes - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | 1926 | 0 | detection | Listeria monocytogenes | 684 | 0 |
| | Other processed food products and prepared dishes - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Other | single (food/feed) | 25 | Gram | N_A | 45 | 0 | detection | Listeria monocytogenes | 45 | 0 |
| | Other processed food products and prepared dishes - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | 1775 | 5 | <= 100 | Listeria monocytogenes - molecular serogroup IVb | 1,265 | 0 |
| | | | | | | | | >100 | Listeria monocytogenes - molecular serogroup IVb | 1,265 | 0 |
| | Other processed food products and prepared dishes - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | 1775 | 5 | detection | Listeria monocytogenes - molecular serogroup IVb | 605 | 5 |
| | Other processed food products and prepared dishes - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 25 | Gram | N_A | 46 | 0 | detection | Listeria monocytogenes | 46 | 0 |
| | Other products of animal origin - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | 8 | 0 | detection | Listeria monocytogenes | 8 | 0 |
| | Snails - Processing plant - European Union - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Snails cooked meat | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Snails - Processing plant - Non European Union - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Snails cooked meat | 11 | 0 | detection | Listeria monocytogenes | 11 | 0 |
| | Snails - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Snails cooked meat | 78 | 2 | detection | Listeria monocytogenes - molecular serogroup IIb | 78 | 2 |

## Table Listeria:LISTERIA in feed

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Silage - Farm - Romania - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 33 | 0 | Listeria monocytogenes | 0 |
| | Silage - Farm - Romania - feed sample - Surveillance - HACCP and own check - Other | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 4 | 0 | Listeria monocytogenes | 0 |
| | Silage - Farm - Romania - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 9 | 0 | Listeria monocytogenes | 0 |

## Table Lyssavirus:LYSSAVIRUS in animal

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| ROMANIA | Badgers - wild - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| | Bears - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 66 | 0 | Lyssavirus | 0 |
| | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 49 | 1 | Lyssavirus | 1 |
| | Deer - farmed - fallow deer - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 7 | 0 | Lyssavirus | 0 |
| | Deer - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 131 | 1 | Lyssavirus | 1 |
| | Ferrets - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 4 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 2590 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 485 | 1 | Lyssavirus | 1 |
| | Goats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 23 | 0 | Lyssavirus | 0 |
| | Jackals - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 5 | 0 | Lyssavirus | 0 |
| | Pigs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| | Rats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Sheep - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 240 | 0 | Lyssavirus | 0 |
| | Solipeds, domestic - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Wild boars - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Wild cat (Felis silvestris) - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Wolves - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| Bihor | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 10 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 9 | 0 | Lyssavirus | 0 |
| | Sheep - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 9 | 0 | Lyssavirus | 0 |
| Bistriţa-Năsăud | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 7 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 63 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 23 | 0 | Lyssavirus | 0 |
| | Goats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| Cluj | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 74 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 20 | 0 | Lyssavirus | 0 |
| Maramureş | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 9 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 90 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 14 | 0 | Lyssavirus | 0 |
| Satu Mare | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 66 | 0 | Lyssavirus | 0 |
| | Foxes - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| Sălaj | Badgers - wild - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 4 | 0 | Lyssavirus | 0 |
| | Deer - farmed - fallow deer - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 5 | 0 | Lyssavirus | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Sălaj | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 49 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 9 | 0 | Lyssavirus | 0 |
| | Goats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| | Sheep - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 7 | 0 | Lyssavirus | 0 |
| Alba | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 142 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| | Goats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 5 | 0 | Lyssavirus | 0 |
| | Sheep - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 23 | 0 | Lyssavirus | 0 |
| Braşov | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 55 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| Covasna | Bears - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 6 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| | Sheep - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Wolves - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| Mureş | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 4 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 211 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 8 | 0 | Lyssavirus | 0 |
| | Sheep - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 5 | 0 | Lyssavirus | 0 |
| Sibiu | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 153 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 5 | 0 | Lyssavirus | 0 |
| | Sheep - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 5 | 0 | Lyssavirus | 0 |
| | Wolves - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| Botoşani | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Ferrets - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 36 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 5 | 0 | Lyssavirus | 0 |
| | Sheep - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 4 | 0 | Lyssavirus | 0 |
| Iaşi | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 6 | 0 | Lyssavirus | 0 |
| | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 1 | Lyssavirus | 1 |
| | Deer - farmed - fallow deer - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 8 | 1 | Lyssavirus | 1 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 40 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 138 | 0 | Lyssavirus | 0 |
| | Sheep - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| Neamţ | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Deer - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 7 | 0 | Lyssavirus | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Neamţ | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 80 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| | Sheep - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| Suceava | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 4 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 208 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 7 | 0 | Lyssavirus | 0 |
| Vaslui | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 25 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 1 | Lyssavirus | 1 |
| Buzău | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 32 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 9 | 0 | Lyssavirus | 0 |
| Constanţa | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 51 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 13 | 0 | Lyssavirus | 0 |
| Galaţi | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 5 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 63 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| Tulcea | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 103 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| Vrancea | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 4 | 0 | Lyssavirus | 0 |
| | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 92 | 0 | Lyssavirus | 0 |
| | Solipeds, domestic - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| Argeş | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 36 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Sheep - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| Călăraşi | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 47 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| Dâmboviţa | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 33 | 0 | Lyssavirus | 0 |
| Giurgiu | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 68 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Pigs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| Ialomiţa | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| | Ferrets - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Sheep - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 10 | 0 | Lyssavirus | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Prahova | Bears - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 7 | 0 | Lyssavirus | 0 |
| | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 89 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 5 | 0 | Lyssavirus | 0 |
| | Sheep - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Wild cat (Felis silvestris) - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| Teleorman | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 16 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 12 | 0 | Lyssavirus | 0 |
| Bucureşti | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Rats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| Ilfov | Badgers - wild - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 5 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Wild boars - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| Dolj | Badgers - wild - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 14 | 0 | Lyssavirus | 0 |
| | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 21 | 0 | Lyssavirus | 0 |
| | Ferrets - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 65 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 37 | 0 | Lyssavirus | 0 |
| | Jackals - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 5 | 0 | Lyssavirus | 0 |
| Gorj | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| | Deer - farmed - fallow deer - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Deer - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 6 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 43 | 0 | Lyssavirus | 0 |
| | Goats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 7 | 0 | Lyssavirus | 0 |
| | Sheep - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 5 | 0 | Lyssavirus | 0 |
| Olt | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 40 | 0 | Lyssavirus | 0 |
| Vâlcea | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 94 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| Arad | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 85 | 0 | Lyssavirus | 0 |
| Caraş-Severin | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 164 | 0 | Lyssavirus | 0 |
| Hunedoara | Cats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 14 | 0 | Lyssavirus | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Hunedoara | Deer - farmed - fallow deer - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 3 | 0 | Lyssavirus | 0 |
| | Deer - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Dogs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 8 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 170 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 9 | 0 | Lyssavirus | 0 |
| | Goats - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 7 | 0 | Lyssavirus | 0 |
| | Pigs - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 2 | 0 | Lyssavirus | 0 |
| | Sheep - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 130 | 0 | Lyssavirus | 0 |
| Timiş | Cattle (bovine animals) - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Monitoring - Official sampling - Objective sampling | N_A | Not Available | animal | 134 | 0 | Lyssavirus | 0 |
| | Foxes - Hunting - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 6 | 0 | Lyssavirus | 0 |
| | Sheep - Unspecified - Romania - animal sample - brain - Surveillance - Official sampling - Suspect sampling | N_A | Not Available | animal | 35 | 0 | Lyssavirus | 0 |

## Table Salmonella:SALMONELLA in animal

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Gallus gallus (fowl) - breeding flocks, unspecified - adult - Farm - Not Available - environmental sample - Control and eradication programmes - Industry sampling - Census | herd/flock | 414 | N | N_A | Not Available | 414 | 11 | Salmonella Amsterdam | 3 |
| | | | | | | | | | Salmonella Enteritidis | 1 |
| | | | | | | | | | Salmonella Infantis | 3 |
| | | | | | | | | | Salmonella Livingstone | 1 |
| | | | | | | | | | Salmonella Senftenberg | 2 |
| | | | | | | | | | Salmonella Taksony | 1 |
| | Gallus gallus (fowl) - breeding flocks, unspecified - adult - Farm - Not Available - environmental sample - Control and eradication programmes - Official and industry sampling - Census | herd/flock | 414 | Y | N_A | Not Available | 414 | 38 | Salmonella Amsterdam | 5 |
| | | | | | | | | | Salmonella Enteritidis | 2 |
| | | | | | | | | | Salmonella Infantis | 4 |
| | | | | | | | | | Salmonella Liverpool | 2 |
| | | | | | | | | | Salmonella Livingstone | 1 |
| | | | | | | | | | Salmonella Mbandaka | 5 |
| | | | | | | | | | Salmonella Senftenberg | 5 |
| | | | | | | | | | Salmonella Taksony | 14 |
| | Gallus gallus (fowl) - breeding flocks, unspecified - adult - Farm - Not Available - environmental sample - Control and eradication programmes - Official sampling - Census | herd/flock | 414 | N | N_A | Not Available | 414 | 27 | Salmonella Amsterdam | 2 |
| | | | | | | | | | Salmonella Enteritidis | 1 |
| | | | | | | | | | Salmonella Infantis | 1 |
| | | | | | | | | | Salmonella Liverpool | 2 |
| | | | | | | | | | Salmonella Mbandaka | 5 |
| | | | | | | | | | Salmonella Senftenberg | 3 |
| | | | | | | | | | Salmonella Taksony | 13 |
| | Gallus gallus (fowl) - broilers - before slaughter - Farm - Not Available - environmental sample - boot swabs - Control and eradication programmes - Industry sampling - Census | herd/flock | 12529 | N | N_A | Not Available | 12189 | 649 | Salmonella | 1 |
| | | | | | | | | | Salmonella Agona | 29 |
| | | | | | | | | | Salmonella Bovismorbificans | 4 |
| | | | | | | | | | Salmonella Bredeney | 5 |
| | | | | | | | | | Salmonella Hadar | 8 |
| | | | | | | | | | Salmonella Infantis | 276 |
| | | | | | | | | | Salmonella Kedougou | 2 |
| | | | | | | | | | Salmonella Kentucky | 40 |
| | | | | | | | | | Salmonella Kottbus | 2 |
| | | | | | | | | | Salmonella Liverpool | 30 |
| | | | | | | | | | Salmonella Livingstone | 25 |
| | | | | | | | | | Salmonella Mbandaka | 30 |
| | | | | | | | | | Salmonella Montevideo | 12 |
| | | | | | | | | | Salmonella Newport | 8 |
| | | | | | | | | | Salmonella Orion | 14 |
| | | | | | | | | | Salmonella Senftenberg | 59 |
| | | | | | | | | | Salmonella Taksony | 41 |
| | | | | | | | | | Salmonella Tennessee | 60 |
| | | | | | | | | | Salmonella Uganda | 3 |
| | Gallus gallus (fowl) - broilers - before slaughter - Farm - Not Available - environmental sample - boot swabs - Control and eradication programmes - Official and industry sampling - Census | herd/flock | 12529 | Y | N_A | Not Available | 12529 | 703 | Salmonella | 1 |
| | | | | | | | | | Salmonella Agona | 29 |
| | | | | | | | | | Salmonella Albany | 1 |
| | | | | | | | | | Salmonella Bovismorbificans | 4 |
| | | | | | | | | | Salmonella Bredeney | 6 |
| | | | | | | | | | Salmonella Enteritidis | 3 |
| | | | | | | | | | Salmonella Hadar | 8 |
| | | | | | | | | | Salmonella Infantis | 299 |
| | | | | | | | | | Salmonella Kedougou | 2 |
| | | | | | | | | | Salmonella Kentucky | 44 |
| | | | | | | | | | Salmonella Kottbus | 2 |
| | | | | | | | | | Salmonella Liverpool | 30 |
| | | | | | | | | | Salmonella Livingstone | 37 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Gallus gallus (fowl) - broilers - before slaughter - Farm - Not Available - environmental sample - boot swabs - Control and eradication programmes - Official and industry sampling - Census | herd/flock | 12529 | Y | N_A | Not Available | 12529 | 703 | Salmonella Mbandaka | 30 |
| | | | | | | | | | Salmonella Montevideo | 12 |
| | | | | | | | | | Salmonella Newport | 8 |
| | | | | | | | | | Salmonella Orion | 16 |
| | | | | | | | | | Salmonella Senftenberg | 62 |
| | | | | | | | | | Salmonella Taksony | 41 |
| | | | | | | | | | Salmonella Tennessee | 65 |
| | | | | | | | | | Salmonella Uganda | 3 |
| | Gallus gallus (fowl) - broilers - before slaughter - Farm - Not Available - environmental sample - boot swabs - Control and eradication programmes - Official sampling - Objective sampling | herd/flock | 12529 | N | N_A | Not Available | 340 | 54 | Salmonella Albany | 1 |
| | | | | | | | | | Salmonella Bredeney | 1 |
| | | | | | | | | | Salmonella Enteritidis | 3 |
| | | | | | | | | | Salmonella Infantis | 23 |
| | | | | | | | | | Salmonella Kentucky | 4 |
| | | | | | | | | | Salmonella Livingstone | 12 |
| | | | | | | | | | Salmonella Orion | 2 |
| | | | | | | | | | Salmonella Senftenberg | 3 |
| | | | | | | | | | Salmonella Tennessee | 5 |
| | Gallus gallus (fowl) - laying hens - adult - Farm - Not Available - environmental sample - Control and eradication programmes - Industry sampling - Census | herd/flock | 818 | N | N_A | Not Available | 818 | 25 | Salmonella Cubana | 1 |
| | | | | | | | | | Salmonella Enteritidis | 2 |
| | | | | | | | | | Salmonella Infantis | 5 |
| | | | | | | | | | Salmonella Kentucky | 1 |
| | | | | | | | | | Salmonella Kottbus | 10 |
| | | | | | | | | | Salmonella Livingstone | 1 |
| | | | | | | | | | Salmonella Montevideo | 2 |
| | | | | | | | | | Salmonella Taksony | 2 |
| | | | | | | | | | Salmonella Tennessee | 1 |
| | Gallus gallus (fowl) - laying hens - adult - Farm - Not Available - environmental sample - Control and eradication programmes - Official and industry sampling - Census | herd/flock | 818 | Y | N_A | Not Available | 818 | 100 | Salmonella Amsterdam | 2 |
| | | | | | | | | | Salmonella Cubana | 1 |
| | | | | | | | | | Salmonella Enteritidis | 21 |
| | | | | | | | | | Salmonella Infantis | 5 |
| | | | | | | | | | Salmonella Kentucky | 1 |
| | | | | | | | | | Salmonella Kottbus | 47 |
| | | | | | | | | | Salmonella Liverpool | 2 |
| | | | | | | | | | Salmonella Livingstone | 1 |
| | | | | | | | | | Salmonella Mbandaka | 4 |
| | | | | | | | | | Salmonella Montevideo | 2 |
| | | | | | | | | | Salmonella Senftenberg | 2 |
| | | | | | | | | | Salmonella Taksony | 11 |
| | | | | | | | | | Salmonella Tennessee | 1 |
| | Gallus gallus (fowl) - laying hens - adult - Farm - Not Available - environmental sample - Control and eradication programmes - Official sampling - Objective sampling | herd/flock | 818 | N | N_A | Not Available | 717 | 75 | Salmonella Amsterdam | 2 |
| | | | | | | | | | Salmonella Enteritidis | 19 |
| | | | | | | | | | Salmonella Kottbus | 37 |
| | | | | | | | | | Salmonella Liverpool | 2 |
| | | | | | | | | | Salmonella Mbandaka | 4 |
| | | | | | | | | | Salmonella Senftenberg | 2 |
| | | | | | | | | | Salmonella Taksony | 9 |
| | Turkeys - fattening flocks - before slaughter - Farm - Not Available - environmental sample - boot swabs - Control and eradication programmes - Industry sampling - Census | herd/flock | 260 | N | N_A | Not Available | 260 | 2 | Salmonella Hadar | 1 |
| | | | | | | | | | Salmonella Typhimurium | 1 |
| | Turkeys - fattening flocks - before slaughter - Farm - Not Available - environmental sample - boot swabs - Control and eradication programmes - Official and industry sampling - Census | herd/flock | 260 | Y | N_A | Not Available | 260 | 2 | Salmonella Hadar | 1 |
| | | | | | | | | | Salmonella Typhimurium | 1 |
| | Turkeys - fattening flocks - before slaughter - Farm - Not Available - environmental sample - boot swabs - Control and eradication programmes - Official sampling - Objective sampling | herd/flock | 260 | N | N_A | Not Available | 38 | 0 | Salmonella | 0 |
| ROMANIA | All animals - unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards, samples taken from peacock | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| ROMANIA | All animals - unspecified - Zoo - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | Samples taken from owl | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | All animals - wild - Natural habitat - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 5 | 0 | Salmonella | 0 |
| | All animals - zoo animals - Zoo - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Buffalos - Natural habitat - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Canary - Farm - Not Available - animal sample - organ/tissue - Unspecified - Official sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 1 | Salmonella Infantis | 1 |
| | Cats - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 5 | 0 | Salmonella | 0 |
| | Cats - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - faeces - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - faeces - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 5 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards, farm | Detection method of microorganisms | 8 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - foetus/stillbirth - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards, farm | Detection method of microorganisms | 18 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards, farm | Detection method of microorganisms | 19 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Unspecified - Official sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 1 | Salmonella Infantis | 1 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - Surveillance - Industry sampling - Objective sampling | animal | | N_A | sample type - semen | Detection method of microorganisms | 44 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| ROMANIA | Cattle (bovine animals) - Zoo - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Crows - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Dogs - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 7 | 0 | Salmonella | 0 |
| | Dogs - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| | Dogs - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 4 | 0 | Salmonella | 0 |
| | Dogs - Veterinary clinics - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Ducks - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards, farm | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| | Ducks - Zoo - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Foxes - Farm - Not Available - animal sample - organ/tissue - Unspecified - Official sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - faeces - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 7 | 6 | Salmonella Enteritidis | 4 |
| | | | | | | | | | Salmonella Typhimurium | 2 |
| | Goats - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 12 | 0 | Salmonella | 0 |
| | | | | | N_A | Detection method of microorganisms | 4 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - foetus/stillbirth - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 10 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 17 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| ROMANIA | Goats - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards, farm | Detection method of microorganisms | 18 | 0 | Salmonella | 0 |
| | Goats - Natural habitat - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Goats - Zoo - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Gulls - Natural habitat - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Lion - Zoo - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Monkeys - Zoo - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Mouflons - Natural habitat - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Ostriches - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Otter - Natural habitat - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Parrots - Farm - Not Available - animal sample - organ/tissue - Unspecified - Official sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 1 | Salmonella Infantis | 1 |
| | Parrots - Zoo - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Pheasants - Farm - Not Available - animal sample - organ/tissue - Unspecified - Official sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 1 | Salmonella Infantis | 1 |
| | Pigeons - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Pigeons - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Pigeons - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 9 | 0 | Salmonella | 0 |
| | | | | | samples taken on request | Detection method of microorganisms | 1 | 1 | Salmonella Typhimurium | 1 |
| | Pigs - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 66 | 5 | Salmonella Typhimurium, monophasic | 5 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| ROMANIA | Pigs - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 28 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 4 | 0 | Salmonella | 0 |
| | | | | | N_A | Detection method of microorganisms | 195 | 5 | Salmonella Typhimurium | 4 |
| | | | | | | | | | Salmonella Typhimurium, monophasic | 1 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards, farm | Detection method of microorganisms | 348 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 306 | 6 | Salmonella Bredeney | 1 |
| | | | | | | | | | Salmonella Derby | 1 |
| | | | | | | | | | Salmonella Enteritidis | 1 |
| | | | | | | | | | Salmonella Kedougou | 1 |
| | | | | | | | | | Salmonella Typhimurium, monophasic | 2 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Unspecified - Official sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 8 | 2 | Salmonella Enteritidis | 2 |
| | Pigs - Farm - Not Available - animal sample - Unspecified - Industry sampling - Objective sampling | animal | | N_A | sample type - semen | Detection method of microorganisms | 13 | 0 | Salmonella | 0 |
| | Pigs - Natural habitat - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | | | | | Salmonella Fillmore | Detection method of microorganisms | 49 | 2 | Salmonella Other serovars | 2 |
| | | | | | samples taken on request | Detection method of microorganisms | 110 | 21 | Salmonella Kentucky | 17 |
| | | | | | | | | | Salmonella Livingstone | 2 |
| | | | | | | | | | Salmonella Montevideo | 1 |
| | | | | | | | | | Salmonella Newport | 1 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 169 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | farm, backyards | Detection method of microorganisms | 15 | 0 | Salmonella | 0 |
| | | | | | N_A | Detection method of microorganisms | 19 | 1 | Salmonella Gallinarum biovar Gallinarum | 1 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 2772 | 0 | Salmonella | 0 |
| | | | | | samples taken on request | Detection method of microorganisms | 83 | 3 | Salmonella Bredeney | 1 |
| | | | | | | | | | Salmonella Gallinarum biovar Pullorum | 1 |
| | | | | | | | | | Salmonella Kentucky | 1 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| ROMANIA | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request, backyards | Detection method of microorganisms | 79 | 2 | Salmonella Enteritidis | 2 |
| | | | | | samples taken on request, farm | Detection method of microorganisms | 146 | 11 | Salmonella Gallinarum biovar Gallinarum | 11 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 144 | 0 | Salmonella | 0 |
| | | | | | samples taken on request | Detection method of microorganisms | 430 | 1 | Salmonella Rissen | 1 |
| | Quails - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 5 | 3 | Salmonella Typhimurium | 3 |
| | Quails - Farm - Not Available - animal sample - organ/tissue - Unspecified - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Rabbits - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Rabbits - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Rabbits - Natural habitat - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Rabbits - Veterinary clinics - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 62 | 28 | Salmonella Abortusovis | 28 |
| | Sheep - Farm - Not Available - animal sample - foetus/stillbirth - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 13 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 8 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 63 | 0 | Salmonella | 0 |
| | | | | | N_A | Detection method of microorganisms | 1 | 1 | Salmonella Enteritidis | 1 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards, samples taken on requests | Detection method of microorganisms | 48 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| ROMANIA | Solipeds, domestic - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Swans - Natural habitat - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 4 | 0 | Salmonella | 0 |
| | Turkeys - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 10 | 2 | Salmonella Kentucky | 2 |
| | | | | | | | | | Salmonella Livingstone | 2 |
| | Turkeys - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 5 | 0 | Salmonella | 0 |
| | Turtles - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 1 | Salmonella Brezany | 1 |
| | Wild boars - Natural habitat - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 5 | 0 | Salmonella | 0 |
| | Wild boars - Natural habitat - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| Bihor | Pigs - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| Bistriţa-Năsăud | All animals - unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards, samples taken from peacock | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Cats - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards, samples taken on requests | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| | Dogs - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards, samples taken on requests | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Mouflons - Natural habitat - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 7 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Bistriţa-Năsăud | Pigs - Farm - Not Available - animal sample - Unspecified - Industry sampling - Objective sampling | animal | | N_A | sample type - semen | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request, backyards | Detection method of microorganisms | 10 | 0 | Salmonella | 0 |
| | Rabbits - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 8 | 3 | Salmonella Abortusovis | 3 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 8 | 0 | Salmonella | 0 |
| | Turkeys - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Wild boars - Natural habitat - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| Cluj | Sheep - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 9 | 5 | Salmonella Abortusovis | 5 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 1 | Salmonella Enteritidis | 1 |
| | Turtles - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 1 | Salmonella Brezany | 1 |
| Maramureş | Goats - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Pigeons - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards, samples taken on requests | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| | | | | | samples taken on request | Detection method of microorganisms | 1 | 1 | Salmonella Bredeney | 1 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 41 | 7 | Salmonella Gallinarum biovar Gallinarum | 7 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Maramureş | Sheep - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 12 | 11 | Salmonella Abortusovis | 11 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| Satu Mare | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards, farm | Detection method of microorganisms | 9 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 5 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 143 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 50 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 7 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 28 | 0 | Salmonella | 0 |
| Sălaj | Goats - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 11 | 6 | Salmonella Abortusovis | 6 |
| Alba | Cattle (bovine animals) - Farm - Not Available - animal sample - faeces - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - faeces - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 4 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | backyards, farm | Detection method of microorganisms | 19 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Alba | Sheep - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 8 | 0 | Salmonella | 0 |
| Braşov | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 12 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 4 | 0 | Salmonella | 0 |
| Mureş | All animals - unspecified - Zoo - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | Samples taken from owl | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards, farm | Detection method of microorganisms | 6 | 0 | Salmonella | 0 |
| | Crows - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Dogs - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - foetus/stillbirth - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Monkeys - Zoo - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Parrots - Zoo - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | Salmonella Fillmore | Detection method of microorganisms | 49 | 2 | Salmonella Other serovars | 2 |
| | | | | | samples taken on request | Detection method of microorganisms | 49 | 16 | Salmonella Kentucky | 12 |
| | | | | | | | | | Salmonella Livingstone | 2 |
| | | | | | | | | | Salmonella Montevideo | 1 |
| | | | | | | | | | Salmonella Newport | 1 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - foetus/stillbirth - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 6 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 9 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Mureş | Turkeys - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 10 | 2 | Salmonella Kentucky | 2 |
| | | | | | | | | | Salmonella Livingstone | 2 |
| Bacău | Canary - Farm - Not Available - animal sample - organ/tissue - Unspecified - Official sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 1 | Salmonella Infantis | 1 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Unspecified - Official sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 1 | Salmonella Infantis | 1 |
| | Foxes - Farm - Not Available - animal sample - organ/tissue - Unspecified - Official sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Parrots - Farm - Not Available - animal sample - organ/tissue - Unspecified - Official sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 1 | Salmonella Infantis | 1 |
| | Pheasants - Farm - Not Available - animal sample - organ/tissue - Unspecified - Official sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 1 | Salmonella Infantis | 1 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Unspecified - Official sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 8 | 2 | Salmonella Enteritidis | 2 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Official sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 430 | 1 | Salmonella Rissen | 1 |
| | Quails - Farm - Not Available - animal sample - organ/tissue - Unspecified - Official sampling - Objective sampling | animal | | N_A | backyards, samples taken from peacock, on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| Botoşani | Cattle (bovine animals) - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - foetus/stillbirth - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| | Pigeons - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 1 | Salmonella Typhimurium | 1 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 16 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| Iaşi | Cattle (bovine animals) - Farm - Not Available - animal sample - faeces - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 5 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Iaşi | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 6 | 0 | Salmonella | 0 |
| | Dogs - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 5 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 7 | 0 | Salmonella | 0 |
| | Goats - Natural habitat - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 28 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 117 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 8 | 0 | Salmonella | 0 |
| | Rabbits - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 19 | 0 | Salmonella | 0 |
| | Wild boars - Natural habitat - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| Neamţ | Goats - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 9 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| | Rabbits - Natural habitat - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Neamţ | Sheep - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 4 | 0 | Salmonella | 0 |
| Suceava | Cattle (bovine animals) - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 14 | 3 | Salmonella Abortusovis | 3 |
| Brăila | All animals - zoo animals - Zoo - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | Samples taken from nutria | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request, farm, backyards | Detection method of microorganisms | 4 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Zoo - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Dogs - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Dogs - Veterinary clinics - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Ducks - Zoo - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Goats - Zoo - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Lion - Zoo - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Ostriches - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 4 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Brăila | Pigs - Farm - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 195 | 5 | Salmonella Typhimurium | 4 |
| | | | | | | | | | Salmonella Typhimurium, monophasic | 1 |
| | Pigs - Natural habitat - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 155 | 0 | Salmonella | 0 |
| | Quails - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 5 | 3 | Salmonella Typhimurium | 3 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards, samples taken on request | Detection method of microorganisms | 7 | 0 | Salmonella | 0 |
| Buzău | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 2288 | 0 | Salmonella | 0 |
| Constanţa | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 49 | 0 | Salmonella | 0 |
| Galaţi | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 22 | 0 | Salmonella | 0 |
| Tulcea | All animals - wild - Natural habitat - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 5 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Ducks - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| | Gulls - Natural habitat - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Otter - Natural habitat - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Pigeons - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 60 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Tulcea | Pigs - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 25 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 19 | 1 | Salmonella Gallinarum biovar Gallinarum | 1 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 5 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 10 | 0 | Salmonella | 0 |
| | Swans - Natural habitat - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 4 | 0 | Salmonella | 0 |
| | Wild boars - Natural habitat - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 4 | 0 | Salmonella | 0 |
| Vrancea | Pigs - Farm - Not Available - animal sample - organ/tissue - Unspecified - Official sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 37 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Official sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 144 | 0 | Salmonella | 0 |
| Călăraşi | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 74 | 1 | Salmonella Enteritidis | 1 |
| Dâmboviţa | Buffalos - Natural habitat - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 7 | 6 | Salmonella Enteritidis | 4 |
| | | | | | | | | | Salmonella Typhimurium | 2 |
| | Goats - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards, farm | Detection method of microorganisms | 5 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards, samples taken on request | Detection method of microorganisms | 5 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 209 | 3 | Salmonella Bredeney | 1 |
| | | | | | | | | | Salmonella Enteritidis | 1 |
| | | | | | | | | | Salmonella Gallinarum biovar Gallinarum | 1 |
| | Solipeds, domestic - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| Ialomiţa | Cattle (bovine animals) - Farm - Not Available - animal sample - foetus/stillbirth - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Ialomița | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Surveillance - Industry sampling - Objective sampling | animal | | N_A | N_A | Detection method of microorganisms | 88 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| Prahova | Goats - Farm - Not Available - animal sample - foetus/stillbirth - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request, backyards | Detection method of microorganisms | 16 | 0 | Salmonella | 0 |
| | Rabbits - Veterinary clinics - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - foetus/stillbirth - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| Dolj | All animals - unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards, samples taken from peacock | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Cats - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 5 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Cattle (bovine animals) - Farm - Not Available - animal sample - Surveillance - Industry sampling - Objective sampling | animal | | N_A | sample type - semen | Detection method of microorganisms | 44 | 0 | Salmonella | 0 |
| | Dogs - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 6 | 0 | Salmonella | 0 |
| | Ducks - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Dolj | Goats - Farm - Not Available - animal sample - foetus/stillbirth - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards, farm | Detection method of microorganisms | 4 | 0 | Salmonella | 0 |
| | Pigeons - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 7 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards, samples taken on requests | Detection method of microorganisms | 37 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - Unspecified - Industry sampling - Objective sampling | animal | | N_A | sample type - semen | Detection method of microorganisms | 12 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 2 | 2 | Salmonella Kentucky | 2 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 46 | 1 | Salmonella Gallinarum biovar Pullorum | 1 |
| | | | | | samples taken on request, farm, backyards | Detection method of microorganisms | 93 | 2 | Salmonella Gallinarum biovar Gallinarum | 2 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 9 | 0 | Salmonella | 0 |
| | Turkeys - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 4 | 0 | Salmonella | 0 |
| Olt | Goats - Farm - Not Available - animal sample - foetus/stillbirth - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 66 | 5 | Salmonella Typhimurium, monophasic | 5 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| | | | | | samples taken on request | Detection method of microorganisms | 80 | 4 | Salmonella Enteritidis | 1 |
| | | | | | | | | | Salmonella Kedougou | 1 |
| | | | | | | | | | Salmonella Typhimurium, monophasic | 2 |
| | Poultry, unspecified - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 10 | 3 | Salmonella Kentucky | 3 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 32 | 1 | Salmonella Kentucky | 1 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under control programme | Target verification | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Olt | Sheep - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| Vâlcea | Cattle (bovine animals) - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - foetus/stillbirth - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 12 | 1 | Salmonella Gallinarum biovar Gallinarum | 1 |
| Hunedoara | Poultry, unspecified - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 8 | 0 | Salmonella | 0 |
| Timiş | Cattle (bovine animals) - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 3 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - foetus/stillbirth - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Goats - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Pigeons - Farm - Not Available - animal sample - faeces - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 1 | 0 | Salmonella | 0 |
| | Pigeons - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 2 | 0 | Salmonella | 0 |
| | Pigs - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 65 | 1 | Salmonella Derby | 1 |
| | Sheep - Farm - Not Available - animal sample - foetus/stillbirth - Unspecified - Industry sampling - Objective sampling | animal | | N_A | backyards | Detection method of microorganisms | 4 | 0 | Salmonella | 0 |
| | Sheep - Farm - Not Available - animal sample - organ/tissue - Unspecified - Industry sampling - Objective sampling | animal | | N_A | samples taken on request | Detection method of microorganisms | 6 | 0 | Salmonella | 0 |

## Table Salmonella:SALMONELLA in food

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Bakery products - cakes - Catering - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 1 | Salmonella Enteritidis | 1 |
| | Bakery products - cakes - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 75 | 0 | Salmonella | 0 |
| | Bakery products - cakes - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 595 | 0 | Salmonella | 0 |
| | Bakery products - cakes - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Bakery products - pastry - Catering - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 25 | 0 | Salmonella | 0 |
| | Bakery products - pastry - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 112 | 0 | Salmonella | 0 |
| | Bakery products - pastry - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 66 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 524 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 275 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 174 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 335 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 4 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 250 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 61 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 557 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 440 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 960 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 170 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 58 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 115 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 3 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 490 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Suspect sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 30 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 120 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 968 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1265 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Cheeses made from goats' milk - hard - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 20 | 0 | Salmonella | 0 |
| | Cheeses made from goats' milk - hard - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Cheeses made from goats' milk - hard - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Cheeses made from goats' milk - hard - made from raw or low heat-treated milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 40 | 0 | Salmonella | 0 |
| | Cheeses made from goats' milk - hard - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 50 | 0 | Salmonella | 0 |
| | Cheeses made from goats' milk - hard - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 25 | 0 | Salmonella | 0 |
| | Cheeses made from goats' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 20 | 0 | Salmonella | 0 |
| | Cheeses made from goats' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Cheeses made from goats' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - Official sampling - Selective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Cheeses made from goats' milk - soft and semi-soft - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 276 | 0 | Salmonella | 0 |
| | Cheeses made from goats' milk - soft and semi-soft - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 85 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Cheeses made from sheep's milk - hard - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 30 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - hard - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 45 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - hard - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 6 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - hard - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 55 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - hard - made from raw or low heat-treated milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 56 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - hard - made from raw or low heat-treated milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 180 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - hard - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 303 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - hard - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 135 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - hard - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 20 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 71 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 125 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 464 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 48 | 0 | Salmonella | 0 |
| | Cheeses, made from unspecified milk or other animal milk - fresh - made from raw or low heat-treated milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 61 | 0 | Salmonella | 0 |
| | Cheeses, made from unspecified milk or other animal milk - fresh - made from raw or low heat-treated milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Cheeses, made from unspecified milk or other animal milk - fresh - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 561 | 0 | Salmonella | 0 |
| | Cheeses, made from unspecified milk or other animal milk - fresh - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 205 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Cheeses, made from unspecified milk or other animal milk - hard - made from raw or low heat-treated milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 127 | 0 | Salmonella | 0 |
| | Cheeses, made from unspecified milk or other animal milk - hard - made from raw or low heat-treated milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 25 | 0 | Salmonella | 0 |
| | Cheeses, made from unspecified milk or other animal milk - hard - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 233 | 0 | Salmonella | 0 |
| | Cheeses, made from unspecified milk or other animal milk - hard - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 26 | 0 | Salmonella | 0 |
| | Cheeses, made from unspecified milk or other animal milk - hard - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1 | 1 | Salmonella Enteritidis | 1 |
| | Crustaceans - unspecified - cooked - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Crustaceans - unspecified - cooked - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Crustaceans - unspecified - cooked - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Crustaceans - unspecified - cooked - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Crustaceans - unspecified - raw - frozen - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Crustaceans - unspecified - raw - frozen - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Crustaceans - unspecified - raw - frozen - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Crustaceans - unspecified - raw - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Meat of boiled snails. Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 11 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - butter - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 175 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - butter - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - butter - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 11 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - butter - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - butter - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - butter - made from raw or low heat-treated milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 36 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Dairy products (excluding cheeses) - butter - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - cream - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 82 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - cream - made from pasteurised milk - Processing plant - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - cream - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - cream - made from raw or low heat-treated milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 50 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - cream - made from raw or low heat-treated milk - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 25 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - cream - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 6 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - cream - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 40 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - ice-cream - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 190 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - ice-cream - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - ice-cream - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 170 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - ice-cream - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 120 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - ice-cream - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - milk powder and whey powder - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 81 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - milk powder and whey powder - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 35 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - milk powder and whey powder - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 84 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - milk powder and whey powder - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 100 | 0 | Salmonella | 0 |
| | Egg products - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 380 | 31 | Salmonella Enteritidis | 17 |
| | | | | | | | | | Salmonella Livingstone | 14 |
| | Egg products - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 2 | 0 | Salmonella | 0 |
| | Egg products - Processing plant - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 35 | 0 | Salmonella | 0 |
| | Egg products - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Eggs - Farm - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the production units (farm) | Detection method presence in x g | 27 | 2 | Salmonella Gloucester | 2 |
| | Eggs - Packing centre - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the production units | Detection method presence in x g | 23 | 0 | Salmonella | 0 |
| | Eggs - Packing centre - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the production units | Detection method presence in x g | 2 | 0 | Salmonella | 0 |
| | Eggs - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 6 | 0 | Salmonella | 0 |
| | Eggs - table eggs - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 2 | 0 | Salmonella | 0 |
| | Eggs - table eggs - Farm - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the production units (farm) | Detection method presence in x g | 525 | 0 | Salmonella | 0 |
| | Eggs - table eggs - Farm - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the production units (farm) | Detection method presence in x g | 64 | 0 | Salmonella | 0 |
| | Eggs - table eggs - Packing centre - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the production units | Detection method presence in x g | 1838 | 0 | Salmonella | 0 |
| | Eggs - table eggs - Packing centre - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the production units | Detection method presence in x g | 242 | 0 | Salmonella | 0 |
| | Eggs - table eggs - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the production units | Detection method presence in x g | 28 | 0 | Salmonella | 0 |
| | Eggs - table eggs - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 158 | 0 | Salmonella | 0 |
| | Eggs - table eggs - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 324 | 0 | Salmonella | 0 |
| | Fishery products, unspecified - non-ready-to-eat - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 6 | 0 | Salmonella | 0 |
| | Fishery products, unspecified - non-ready-to-eat - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 88 | 0 | Salmonella | 0 |
| | Fishery products, unspecified - non-ready-to-eat - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 6 | 0 | Salmonella | 0 |
| | Fishery products, unspecified - raw - chilled - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 3 | 0 | Salmonella | 0 |
| | Fishery products, unspecified - raw - chilled - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1 | 0 | Salmonella | 0 |
| | Fishery products, unspecified - raw - chilled - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 74 | 0 | Salmonella | 0 |
| | Fishery products, unspecified - raw - chilled - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Fishery products, unspecified - smoked - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 19 | 0 | Salmonella | 0 |
| | Fishery products, unspecified - smoked - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 22 | 0 | Salmonella | 0 |
| | Fruits - pre-cut - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 20 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Fruits - pre-cut - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Fruits - pre-cut - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 70 | 0 | Salmonella | 0 |
| | Fruits - pre-cut - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 40 | 0 | Salmonella | 0 |
| | Juice - fruit juice - unpasteurised - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Juice - fruit juice - unpasteurised - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 45 | 0 | Salmonella | 0 |
| | Juice - fruit juice - unpasteurised - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 115 | 0 | Salmonella | 0 |
| | Juice - fruit juice - unpasteurised - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 95 | 0 | Salmonella | 0 |
| | Juice - fruit juice - unpasteurised - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 355 | 0 | Salmonella | 0 |
| | Juice - fruit juice - unpasteurised - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 645 | 0 | Salmonella | 0 |
| | Juice - fruit juice - unpasteurised - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Selective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Juice - vegetable juice - unpasteurised - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Juice - vegetable juice - unpasteurised - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Juice - vegetable juice - unpasteurised - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 35 | 0 | Salmonella | 0 |
| | Juice - vegetable juice - unpasteurised - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 65 | 0 | Salmonella | 0 |
| | Meat from bovine animals - carcase - Slaughterhouse - Not Available - food sample - carcase swabs - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 400 | Square centimetre | N_A | Detection method presence in x g | 5514 | 0 | Salmonella | 0 |
| | Meat from bovine animals - carcase - Slaughterhouse - Not Available - food sample - carcase swabs - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 400 | Square centimetre | N_A | Detection method presence in x g | 1524 | 1 | Salmonella Typhimurium | 1 |
| | Meat from bovine animals - fresh - Cutting plant - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1015 | 0 | Salmonella | 0 |
| | Meat from bovine animals - fresh - Cutting plant - Not Available - food sample - meat - Surveillance - HACCP and own check - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1 | 0 | Salmonella | 0 |
| | Meat from bovine animals - fresh - Processing plant - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 119 | 0 | Salmonella | 0 |
| | Meat from bovine animals - fresh - Retail - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 169 | 0 | Salmonella | 0 |
| | Meat from bovine animals - fresh - Slaughterhouse - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1817 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Meat from bovine animals - meat preparation - intended to be eaten cooked - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 70 | 0 | Salmonella | 0 |
| | Meat from bovine animals - meat preparation - intended to be eaten cooked - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 25 | 0 | Salmonella | 0 |
| | Meat from bovine animals - meat preparation - intended to be eaten cooked - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 180 | 0 | Salmonella | 0 |
| | Meat from bovine animals - meat preparation - intended to be eaten cooked - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 20 | 0 | Salmonella | 0 |
| | Meat from bovine animals - meat preparation - intended to be eaten cooked - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 50 | 0 | Salmonella | 0 |
| | Meat from bovine animals - meat preparation - intended to be eaten cooked - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Meat from bovine animals - meat preparation - intended to be eaten raw - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Meat from bovine animals - meat products - cooked, ready-to-eat - Catering - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Meat from bovine animals - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Meat from bovine animals - minced meat - intended to be eaten cooked - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 75 | 0 | Salmonella | 0 |
| | Meat from bovine animals - minced meat - intended to be eaten cooked - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 270 | 0 | Salmonella | 0 |
| | Meat from bovine animals - minced meat - intended to be eaten cooked - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 40 | 0 | Salmonella | 0 |
| | Meat from bovine animals - minced meat - intended to be eaten cooked - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 609 | 0 | Salmonella | 0 |
| | Meat from bovine animals - minced meat - intended to be eaten cooked - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 60 | 0 | Salmonella | 0 |
| | Meat from bovine animals - minced meat - intended to be eaten raw - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - carcase - chilled - Slaughterhouse - Not Available - food sample - neck skin - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 2441 | 32 | Salmonella Derby | 3 |
| | | | | | | | | | Salmonella Hadar | 12 |
| | | | | | | | | | Salmonella Infantis | 14 |
| | | | | | | | | | Salmonella Kottbus | 1 |
| | | | | | | | | | Salmonella Liverpool | 1 |
| | | | | | | | | | Salmonella Tennessee | 1 |
| | Meat from broilers (Gallus gallus) - carcase - chilled - Slaughterhouse - Not Available - food sample - neck skin - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 697 | 3 | Salmonella Hadar | 1 |
| | | | | | | | | | Salmonella Infantis | 2 |
| | Meat from broilers (Gallus gallus) - fresh - Catering - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 8 | Salmonella Enteritidis | 4 |
| | | | | | | | | | Salmonella Infantis | 4 |
| | Meat from broilers (Gallus gallus) - fresh - Conservation facilities - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 30 | 21 | Salmonella Bredeney | 1 |
| | | | | | | | | | Salmonella Infantis | 18 |
| | | | | | | | | | Salmonella Kentucky | 2 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Meat from broilers (Gallus gallus) - fresh - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 948 | 35 | Salmonella Bredeney | 1 |
| | | | | | | | | | Salmonella Enteritidis | 4 |
| | | | | | | | | | Salmonella Hadar | 2 |
| | | | | | | | | | Salmonella Infantis | 25 |
| | | | | | | | | | Salmonella Kentucky | 2 |
| | | | | | | | | | Salmonella Newport | 1 |
| | Meat from broilers (Gallus gallus) - fresh - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 45 | 4 | Salmonella Infantis | 4 |
| | Meat from broilers (Gallus gallus) - fresh - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 256 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - fresh - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - fresh - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 556 | 5 | Salmonella Infantis | 5 |
| | Meat from broilers (Gallus gallus) - fresh - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1043 | 10 | Salmonella Infantis | 10 |
| | Meat from broilers (Gallus gallus) - fresh - Slaughterhouse - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 596 | 7 | Salmonella Infantis | 7 |
| | Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 145 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 565 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 82 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 145 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 60 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - Slaughterhouse - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 220 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Catering - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 175 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Catering - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 25 | 10 | Salmonella Infantis | 10 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 45 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 25 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 26 | 5 | Salmonella Infantis | 5 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Surveillance - HACCP and own check - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 85 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat products - raw but intended to be eaten cooked - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat products - raw but intended to be eaten cooked - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat products - raw but intended to be eaten cooked - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 30 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat products - raw but intended to be eaten cooked - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 70 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat products - raw but intended to be eaten cooked - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 80 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat products - raw but intended to be eaten cooked - Slaughterhouse - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - mechanically separated meat (MSM) - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 110 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - mechanically separated meat (MSM) - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 100 | 3 | Salmonella Infantis | 3 |
| | Meat from broilers (Gallus gallus) - mechanically separated meat (MSM) - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - mechanically separated meat (MSM) - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 160 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - mechanically separated meat (MSM) - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 85 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - mechanically separated meat (MSM) - Slaughterhouse - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 495 | 2 | Salmonella Infantis | 2 |
| | Meat from broilers (Gallus gallus) - minced meat - intended to be eaten cooked - Conservation facilities - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 15 | 4 | Salmonella Infantis | 4 |
| | Meat from broilers (Gallus gallus) - minced meat - intended to be eaten cooked - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 90 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - minced meat - intended to be eaten cooked - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 75 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - minced meat - intended to be eaten cooked - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 400 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - minced meat - intended to be eaten cooked - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 38 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Meat from broilers (Gallus gallus) - minced meat - intended to be eaten cooked - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 240 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - minced meat - intended to be eaten cooked - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 140 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - minced meat - intended to be eaten cooked - Slaughterhouse - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 185 | 9 | Salmonella Infantis | 9 |
| | Meat from duck - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Meat from duck - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Meat from horse - carcase - Slaughterhouse - Not Available - food sample - carcase swabs - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 400 | Square centimetre | N_A | Detection method presence in x g | 330 | 0 | Salmonella | 0 |
| | Meat from horse - carcase - Slaughterhouse - Not Available - food sample - carcase swabs - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 400 | Square centimetre | N_A | Detection method presence in x g | 110 | 0 | Salmonella | 0 |
| | Meat from horse - fresh - Slaughterhouse - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 20 | 0 | Salmonella | 0 |
| | Meat from horse - minced meat - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 25 | 0 | Salmonella | 0 |
| | Meat from horse - minced meat - Slaughterhouse - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 35 | 0 | Salmonella | 0 |
| | Meat from other animal species or not specified - meat products - pâté - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 68 | 0 | Salmonella | 0 |
| | Meat from other animal species or not specified - meat products - pâté - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1 | 0 | Salmonella | 0 |
| | Meat from pig - carcase - Slaughterhouse - Not Available - food sample - carcase swabs - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 400 | Square centimetre | N_A | Detection method presence in x g | 4246 | 0 | Salmonella | 0 |
| | Meat from pig - carcase - Slaughterhouse - Not Available - food sample - carcase swabs - Surveillance - based on Regulation 2073 - Official, based on Regulation 854/2004 - Objective sampling | single (food/feed) | 400 | Square centimetre | N_A | Detection method presence in x g | 2388 | 2 | Salmonella Typhimurium | 2 |
| | Meat from pig - carcase - Slaughterhouse - Not Available - food sample - carcase swabs - Surveillance - based on Regulation 2073 - Official, based on Regulation 854/2004 - Selective sampling | single (food/feed) | 400 | Square centimetre | N_A | Detection method presence in x g | 25 | 0 | Salmonella | 0 |
| | Meat from pig - fresh - Conservation facilities - Hungary - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 25 | 5 | Salmonella Goldcoast | 5 |
| | Meat from pig - fresh - Cutting plant - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1859 | 5 | Salmonella Brandenburg | 5 |
| | Meat from pig - fresh - Processing plant - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 447 | 0 | Salmonella | 0 |
| | Meat from pig - fresh - Retail - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 402 | 17 | Salmonella Muenchen | 5 |
| | | | | | | | | | Salmonella Rissen | 5 |
| | | | | | | | | | Salmonella Typhimurium | 7 |
| | Meat from pig - fresh - Retail - Not Available - food sample - meat - Surveillance - HACCP and own check - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Meat from pig - fresh - Slaughterhouse - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 200 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Meat from pig - meat preparation - intended to be eaten cooked - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 250 | 2 | Salmonella Bredeney | 2 |
| | Meat from pig - meat preparation - intended to be eaten cooked - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 200 | 2 | Salmonella Rissen | 2 |
| | Meat from pig - meat preparation - intended to be eaten cooked - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 2512 | 0 | Salmonella | 0 |
| | Meat from pig - meat preparation - intended to be eaten cooked - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 125 | 0 | Salmonella | 0 |
| | Meat from pig - meat preparation - intended to be eaten cooked - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 444 | 0 | Salmonella | 0 |
| | Meat from pig - meat preparation - intended to be eaten cooked - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 225 | 0 | Salmonella | 0 |
| | Meat from pig - meat preparation - intended to be eaten raw - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 65 | 0 | Salmonella | 0 |
| | Meat from pig - meat preparation - intended to be eaten raw - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 20 | 0 | Salmonella | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Catering - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 20 | 0 | Salmonella | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1502 | 1 | Salmonella Gloucester | 1 |
| | Meat from pig - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 170 | 0 | Salmonella | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 306 | 0 | Salmonella | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 77 | 0 | Salmonella | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Meat from pig - meat products - raw but intended to be eaten cooked - Cutting plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 15 | 8 | Salmonella Derby | 6 |
| | | | | | | | | | Salmonella Rissen | 1 |
| | | | | | | | | | Salmonella Typhimurium | 1 |
| | Meat from pig - meat products - raw but intended to be eaten cooked - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 308 | 0 | Salmonella | 0 |
| | Meat from pig - meat products - raw but intended to be eaten cooked - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 95 | 0 | Salmonella | 0 |
| | Meat from pig - meat products - raw but intended to be eaten cooked - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 20 | 0 | Salmonella | 0 |
| | Meat from pig - mechanically separated meat (MSM) - Processing plant - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Meat from pig - minced meat - intended to be eaten cooked - Catering - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Meat from pig - minced meat - intended to be eaten cooked - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 1822 | 5 | Salmonella Infantis | 2 |
| | | | | | | | | | Salmonella Typhimurium | 3 |
| | Meat from pig - minced meat - intended to be eaten cooked - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 780 | 5 | Salmonella Derby | 1 |
| | | | | | | | | | Salmonella Give | 1 |
| | | | | | | | | | Salmonella Infantis | 2 |
| | | | | | | | | | Salmonella Rissen | 1 |
| | Meat from pig - minced meat - intended to be eaten cooked - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 2724 | 0 | Salmonella | 0 |
| | Meat from pig - minced meat - intended to be eaten cooked - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 510 | 0 | Salmonella | 0 |
| | Meat from pig - minced meat - intended to be eaten cooked - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 1292 | 8 | Salmonella Bredeney | 1 |
| | | | | | | | | | Salmonella Derby | 3 |
| | | | | | | | | | Salmonella Muenchen | 1 |
| | | | | | | | | | Salmonella Rissen | 3 |
| | Meat from pig - minced meat - intended to be eaten cooked - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 605 | 2 | Salmonella Bredeney | 2 |
| | Meat from pig - minced meat - intended to be eaten raw - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 81 | 0 | Salmonella | 0 |
| | Meat from pig - minced meat - intended to be eaten raw - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 25 | 0 | Salmonella | 0 |
| | Meat from pig - minced meat - intended to be eaten raw - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 110 | 0 | Salmonella | 0 |
| | Meat from pig - minced meat - intended to be eaten raw - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 40 | 0 | Salmonella | 0 |
| | Meat from pig - offal - liver - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 2 | 0 | Salmonella | 0 |
| | Meat from pig - offal - liver - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 3 | 2 | Salmonella Derby | 2 |
| | Meat from pig - offal - liver - Slaughterhouse - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1 | 0 | Salmonella | 0 |
| | Meat from pig - offal - liver - Slaughterhouse - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 20 | 3 | Salmonella Rissen | 3 |
| | Meat from pig - offal - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1 | 1 | Salmonella Derby | 1 |
| | Meat from pig - offal - Slaughterhouse - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 39 | 3 | Salmonella Rissen | 3 |
| | Meat from poultry, unspecified - meat preparation - Processing plant - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 35 | 0 | Salmonella | 0 |
| | Meat from poultry, unspecified - meat preparation - Processing plant - Not Available - food sample - meat - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Meat from poultry, unspecified - meat products - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 2 | 0 | Salmonella | 0 |
| | Meat from poultry, unspecified - meat products - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Meat from poultry, unspecified - meat products - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 85 | 0 | Salmonella | 0 |
| | Meat from poultry, unspecified - offal - liver - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 12 | 0 | Salmonella | 0 |
| | Meat from poultry, unspecified - offal - liver - Slaughterhouse - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 310 | 3 | Salmonella Infantis | 3 |
| | Meat from poultry, unspecified - offal - liver - Slaughterhouse - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Meat from poultry, unspecified - offal - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 1 | Salmonella Infantis | 1 |
| | Meat from poultry, unspecified - offal - Slaughterhouse - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 325 | 6 | Salmonella Infantis | 6 |
| | Meat from poultry, unspecified - offal - Slaughterhouse - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Meat from poultry, unspecified - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Meat from sheep - carcase - Slaughterhouse - Not Available - food sample - carcase swabs - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 400 | Square centimetre | N_A | Detection method presence in x g | 1787 | 0 | Salmonella | 0 |
| | Meat from sheep - carcase - Slaughterhouse - Not Available - food sample - carcase swabs - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 400 | Square centimetre | N_A | Detection method presence in x g | 493 | 0 | Salmonella | 0 |
| | Meat from sheep - fresh - Cutting plant - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 56 | 0 | Salmonella | 0 |
| | Meat from sheep - fresh - Processing plant - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 50 | 0 | Salmonella | 0 |
| | Meat from sheep - fresh - Retail - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 147 | 0 | Salmonella | 0 |
| | Meat from sheep - fresh - Slaughterhouse - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Meat from sheep - minced meat - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 55 | 0 | Salmonella | 0 |
| | Meat from sheep - minced meat - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Meat from sheep - minced meat - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 30 | 0 | Salmonella | 0 |
| | Meat from sheep - offal - Slaughterhouse - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1 | 1 | Salmonella Typhimurium | 1 |
| | Meat from turkey - carcase - chilled - Slaughterhouse - Not Available - food sample - neck skin - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 85 | 13 | Salmonella Hadar | 13 |
| | Meat from turkey - carcase - chilled - Slaughterhouse - Not Available - food sample - neck skin - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 40 | 0 | Salmonella | 0 |
| | Meat from turkey - fresh - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 10 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Meat from turkey - fresh - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 35 | 0 | Salmonella | 0 |
| | Meat from turkey - fresh - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 6 | 0 | Salmonella | 0 |
| | Meat from turkey - fresh - Slaughterhouse - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Meat from turkey - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 35 | 2 | Salmonella Typhimurium | 2 |
| | Meat from turkey - mechanically separated meat (MSM) - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Meat from turkey - minced meat - intended to be eaten cooked - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Meat from wild game - birds - meat products - Processing plant - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 25 | 0 | Salmonella | 0 |
| | Meat from wild game - birds - meat products - Processing plant - Not Available - food sample - meat - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 25 | 0 | Salmonella | 0 |
| | Meat from wild game - birds - Processing plant - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Meat from wild game - birds - Slaughterhouse - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 30 | 0 | Salmonella | 0 |
| | Meat from wild game - land mammals - meat products - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 20 | 0 | Salmonella | 0 |
| | Meat from wild game - land mammals - Processing plant - Not Available - food sample - meat - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat preparation - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 1701 | 25 | Salmonella Brandenburg | 1 |
| | | | | | | | | | Salmonella Infantis | 1 |
| | | | | | | | | | Salmonella Litchfield | 2 |
| | | | | | | | | | Salmonella Paratyphi B | 1 |
| | | | | | | | | | Salmonella Rissen | 5 |
| | | | | | | | | | Salmonella Typhimurium | 15 |
| | Meat, mixed meat - meat preparation - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Suspect sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 1 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat preparation - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 965 | 10 | Salmonella Derby | 5 |
| | | | | | | | | | Salmonella Gloucester | 2 |
| | | | | | | | | | Salmonella Typhimurium | 3 |
| | Meat, mixed meat - meat preparation - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Selective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat preparation - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 3483 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat preparation - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 270 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat preparation - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 1077 | 2 | Salmonella Enteritidis | 1 |
| | | | | | | | | | Salmonella Goldcoast | 1 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Meat, mixed meat - meat preparation - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 590 | 2 | Salmonella Typhimurium | 2 |
| | Meat, mixed meat - meat products - cooked, ready-to-eat - Catering - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat products - cooked, ready-to-eat - Cutting plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 205 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 3426 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat products - cooked, ready-to-eat - Processing plant - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 196 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 457 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 75 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat products - cooked, ready-to-eat - Retail - Not Available - food sample - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 3 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat products - raw and intended to be eaten raw - Cutting plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 310 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat products - raw and intended to be eaten raw - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 260 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat products - raw and intended to be eaten raw - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 188 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat products - raw and intended to be eaten raw - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 240 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat products - raw and intended to be eaten raw - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 181 | 2 | Salmonella Typhimurium | 2 |
| | Meat, mixed meat - meat products - raw but intended to be eaten cooked - Cutting plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 530 | 1 | Salmonella Rissen | 1 |
| | Meat, mixed meat - meat products - raw but intended to be eaten cooked - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 470 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat products - raw but intended to be eaten cooked - Processing plant - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 3 | Salmonella Rissen | 3 |
| | Meat, mixed meat - meat products - raw but intended to be eaten cooked - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 50 | 0 | Salmonella | 0 |
| | Meat, mixed meat - minced meat - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 615 | 0 | Salmonella | 0 |
| | Meat, mixed meat - minced meat - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 300 | 2 | Salmonella Kottbus | 2 |
| | Meat, mixed meat - minced meat - Cutting plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Meat, mixed meat - minced meat - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 2831 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Meat, mixed meat - minced meat - Processing plant - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 260 | 0 | Salmonella | 0 |
| | Meat, mixed meat - minced meat - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 839 | 0 | Salmonella | 0 |
| | Meat, mixed meat - minced meat - Retail - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 10 | Gram | N_A | Detection method presence in x g | 290 | 0 | Salmonella | 0 |
| | Milk, cows' - pasteurised milk - Catering - Not Available - food sample - milk - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Millilitre | N_A | Detection method presence in x g | 6 | 0 | Salmonella | 0 |
| | Milk, cows' - pasteurised milk - Processing plant - Not Available - food sample - milk - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Millilitre | N_A | Detection method presence in x g | 70 | 0 | Salmonella | 0 |
| | Milk, cows' - pasteurised milk - Retail - Not Available - food sample - milk - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Millilitre | N_A | Detection method presence in x g | 6 | 0 | Salmonella | 0 |
| | Milk, cows' - raw milk - intended for direct human consumption - Catering - Not Available - food sample - milk - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Millilitre | N_A | Detection method presence in x g | 1 | 0 | Salmonella | 0 |
| | Milk, cows' - raw milk - intended for direct human consumption - Retail - Not Available - food sample - milk - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Millilitre | N_A | Detection method presence in x g | 1 | 0 | Salmonella | 0 |
| | Molluscan shellfish - cooked - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Molluscan shellfish - cooked - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 205 | 0 | Salmonella | 0 |
| | Molluscan shellfish - raw - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 30 | 0 | Salmonella | 0 |
| | Molluscan shellfish - raw - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Other food - Catering - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 23 | 0 | Salmonella | 0 |
| | Other food - Catering - Not Available - food sample - Surveillance - HACCP and own check - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 2 | Salmonella Enteritidis | 2 |
| | Other food - Catering - Not Available - food sample - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 7 | 0 | Salmonella | 0 |
| | Other food - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 436 | 0 | Salmonella | 0 |
| | Other food - Processing plant - Not Available - food sample - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Other food - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 338 | 0 | Salmonella | 0 |
| | Other food - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 226 | 0 | Salmonella | 0 |
| | Other food - Retail - Not Available - food sample - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Other food - Slaughterhouse - Not Available - food sample - meat - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 10 | 2 | Salmonella Typhimurium | 2 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Other processed food products and prepared dishes - Catering - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 805 | 10 | Salmonella Enteritidis | 10 |
| | Other processed food products and prepared dishes - Catering - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - fish and seafood based dishes - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - Hospital or medical care facility - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 38 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 45 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - Processing plant - Not Available - food sample - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 25 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 967 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 85 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - Retail - Not Available - food sample - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - unspecified - Catering - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 537 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - unspecified - Catering - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 40 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - unspecified - Catering - Not Available - food sample - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - unspecified - containing raw egg - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - unspecified - containing raw egg - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - unspecified - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 133 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - unspecified - Processing plant - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 85 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - unspecified - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 312 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - unspecified - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - unspecified - Retail - Not Available - food sample - Surveillance - Official sampling - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 3 | 0 | Salmonella | 0 |
| | Other products of animal origin - gelatin and collagen - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 10 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Other products of animal origin - gelatin and collagen - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 45 | 0 | Salmonella | 0 |
| | Other products of animal origin - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 200 | 0 | Salmonella | 0 |
| | Other products of animal origin - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 24 | 0 | Salmonella | 0 |
| | Other products of animal origin - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 2 | 0 | Salmonella | 0 |
| | Other products of animal origin - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 2 | 0 | Salmonella | 0 |
| | Seeds, sprouted - ready-to-eat - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 75 | 0 | Salmonella | 0 |
| | Seeds, sprouted - ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 80 | 0 | Salmonella | 0 |
| | Snails - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Meat of boiled snails. Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 87 | 0 | Salmonella | 0 |
| | Snails - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Meat of boiled snails. Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Spices and herbs - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 191 | 0 | Salmonella | 0 |
| | Spices and herbs - Processing plant - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 17 | 0 | Salmonella | 0 |
| | Spices and herbs - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 21 | 0 | Salmonella | 0 |
| | Vegetables - pre-cut - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 70 | 0 | Salmonella | 0 |
| | Vegetables - pre-cut - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 490 | 0 | Salmonella | 0 |
| | Vegetables - pre-cut - Catering - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Selective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Vegetables - pre-cut - frozen vegetables - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Vegetables - pre-cut - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 36 | 0 | Salmonella | 0 |
| | Vegetables - pre-cut - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | Products held for sale within the storage facilities of the processing units | Detection method presence in x g | 40 | 0 | Salmonella | 0 |
| | Vegetables - pre-cut - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 225 | 0 | Salmonella | 0 |
| | Vegetables - pre-cut - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 390 | 0 | Salmonella | 0 |
| | Vegetables - Processing plant - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 6 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Vegetables - products - dried - Retail - Not Available - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |

## Table Salmonella:SALMONELLA in feed

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Compound feedingstuffs for cattle - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 16 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for cattle - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 20 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for cattle - final product - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 55 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for cattle - final product - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 64 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for cattle - final product - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 35 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for cattle - process control - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for cattle - process control - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for fish - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 6 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for pigs - final product - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 180 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for pigs - final product - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 333 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for pigs - final product - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 260 | 1 | Salmonella Typhimurium | 1 |
| | Compound feedingstuffs for pigs - process control - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 118 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for pigs - process control - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 60 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for poultry (non specified) - final product - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 59 | 5 | Salmonella Livingstone | 5 |
| | Compound feedingstuffs for poultry (non specified) - final product - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 277 | 1 | Salmonella Montevideo | 1 |
| | Compound feedingstuffs for poultry (non specified) - final product - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 130 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for poultry (non specified) - process control - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 37 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for poultry (non specified) - process control - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 110 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for poultry (non specified) - process control - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 37 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for poultry, breeders - final product - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 63 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Compound feedingstuffs for poultry, breeders - final product - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 1 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for poultry, broilers - final product - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 318 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for poultry, broilers - final product - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 34 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for poultry, broilers - final product - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 22 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for poultry, broilers - process control - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 6 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for poultry, broilers - process control - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 33 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for poultry, broilers - process control - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for poultry, laying hens - final product - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 139 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for poultry, laying hens - final product - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for poultry, laying hens - final product - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 70 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for poultry, laying hens - final product - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 100 | 1 | Salmonella Typhimurium | 1 |
| | Compound feedingstuffs for poultry, laying hens - process control - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for poultry, laying hens - process control - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Feed material of cereal grain origin - barley derived - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 6 | 0 | Salmonella | 0 |
| | Feed material of cereal grain origin - barley derived - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Feed material of cereal grain origin - barley derived - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Feed material of cereal grain origin - maize derived - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 44 | 0 | Salmonella | 0 |
| | Feed material of cereal grain origin - maize derived - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 16 | 0 | Salmonella | 0 |
| | Feed material of cereal grain origin - maize derived - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 152 | 0 | Salmonella | 0 |
| | Feed material of cereal grain origin - wheat derived - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 18 | 0 | Salmonella | 0 |
| | Feed material of cereal grain origin - wheat derived - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 36 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Feed material of cereal grain origin - wheat derived - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Feed material of cereal grain origin - wheat derived - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 115 | 0 | Salmonella | 0 |
| | Feed material of land animal origin - animal fat - Processing plant - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 470 | 0 | Salmonella | 0 |
| | Feed material of land animal origin - blood meal - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 57 | 0 | Salmonella | 0 |
| | Feed material of land animal origin - dairy products - whey powder - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Feed material of land animal origin - dairy products - whey powder - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Feed material of land animal origin - dairy products - whey powder - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 80 | 0 | Salmonella | 0 |
| | Feed material of land animal origin - feather meal - Processing plant - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 736 | 0 | Salmonella | 0 |
| | Feed material of land animal origin - meat and bone meal - Processing plant - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 51 | 0 | Salmonella | 0 |
| | Feed material of land animal origin - meat meal - Processing plant - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 140 | 0 | Salmonella | 0 |
| | Feed material of land animal origin - poultry offal meal - Processing plant - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 681 | 0 | Salmonella | 0 |
| | Feed material of marine animal origin - fish meal - Processing plant - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 25 | 0 | Salmonella | 0 |
| | Feed material of marine animal origin - fish meal - Processing plant - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 15 | 0 | Salmonella | 0 |
| | Feed material of oil seed or fruit origin - linseed derived - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Feed material of oil seed or fruit origin - other oil seeds derived - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Feed material of oil seed or fruit origin - other oil seeds derived - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 6 | 0 | Salmonella | 0 |
| | Feed material of oil seed or fruit origin - other oil seeds derived - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 121 | 0 | Salmonella | 0 |
| | Feed material of oil seed or fruit origin - soya (bean) derived - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 2 | 0 | Salmonella | 0 |
| | Feed material of oil seed or fruit origin - soya (bean) derived - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 152 | 0 | Salmonella | 0 |
| | Feed material of oil seed or fruit origin - soya (bean) derived - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Suspect sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Feed material of oil seed or fruit origin - soya (bean) derived - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 286 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Feed material of oil seed or fruit origin - sunflower seed derived - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 10 | 0 | Salmonella | 0 |
| | Feed material of oil seed or fruit origin - sunflower seed derived - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 62 | 0 | Salmonella | 0 |
| | Feed material of oil seed or fruit origin - sunflower seed derived - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 110 | 0 | Salmonella | 0 |
| | Other feed material - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 1 | Salmonella Typhimurium | 1 |
| | Other feed material - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 182 | 0 | Salmonella | 0 |
| | Other feed material - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 105 | 0 | Salmonella | 0 |
| | Other feed material - forages and roughages - Farm - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 6 | 0 | Salmonella | 0 |
| | Other feed material - forages and roughages - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 12 | 0 | Salmonella | 0 |
| | Other feed material - legume seeds and similar products - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Other feed material - legume seeds and similar products - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |
| | Pet food - dog snacks (pig ears, chewing bones) - Processing plant - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 265 | 0 | Salmonella | 0 |
| | Pet food - dog snacks (pig ears, chewing bones) - Processing plant - Not Available - feed sample - Surveillance - HACCP and own check - Selective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 50 | 0 | Salmonella | 0 |
| | Pet food - dog snacks (pig ears, chewing bones) - Retail - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 20 | 0 | Salmonella | 0 |
| | Premixtures - Feed mill - Not Available - feed sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 11 | 0 | Salmonella | 0 |
| | Premixtures - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Detection method presence in x g | 5 | 0 | Salmonella | 0 |

## Table Staphylococcal enterotoxins:STAPHYLOCOCCAL ENTEROTOXINS in food

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Cheeses made from cows' milk - hard - made from pasteurised milk - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 1 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Conservation facilities - European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 10 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 40 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 10 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 15 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 295 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 45 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 150 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 35 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 45 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 80 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 115 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Catering - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 1 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Suspect sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 5 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 40 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from goats' milk - hard - made from pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 25 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from goats' milk - hard - made from raw or low heat-treated milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 5 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from goats' milk - hard - made from raw or low heat-treated milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 5 | 0 | Staphylococcal enterotoxins | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Sampling Details | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|---|---|
| Not Available | Cheeses made from goats' milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 5 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from sheep's milk - hard - made from pasteurised milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 45 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from sheep's milk - hard - made from raw or low heat-treated milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 80 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from pasteurised milk - Conservation facilities - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 5 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Farm - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 5 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 5 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 73 | 2 | Staphylococcal enterotoxins - Enterotoxin C | 2 |
| | | | | | | | | | Staphylococcal enterotoxins - Enterotoxin H | 2 |
| | Dairy products (excluding cheeses) - milk powder and whey powder - Processing plant - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 10 | 0 | Staphylococcal enterotoxins | 0 |
| | Dairy products (excluding cheeses) - milk powder and whey powder - Retail - European Union - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 5 | 0 | Staphylococcal enterotoxins | 0 |
| | Dairy products (excluding cheeses) - milk powder and whey powder - Retail - Romania - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | N_A | Microbiological special tests | 45 | 0 | Staphylococcal enterotoxins | 0 |
| | Dairy products, unspecified - Processing plant - Romania - food sample - Surveillance - HACCP and own check - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Microbiological special tests | 1 | 0 | Staphylococcal enterotoxins | 0 |
| | Dairy products, unspecified - Retail - Romania - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | N_A | Microbiological special tests | 5 | 0 | Staphylococcal enterotoxins | 0 |

## Table Toxoplasma:TOXOPLASMA in animal

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| ROMANIA | Cats - pet animals - Veterinary clinics - Romania - animal sample - blood - Clinical investigations - Private sampling - Not specified | N_A | Enzyme-linked immunosorbent assay (ELISA) | animal | 4 | 2 | Toxoplasma gondii | 2 |
| | Goats - mixed herds - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | N_A | Enzyme-linked immunosorbent assay (ELISA) | animal | 29 | 6 | Toxoplasma gondii | 6 |
| | Sheep - mixed herds - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | N_A | Enzyme-linked immunosorbent assay (ELISA) | animal | 7 | 4 | Toxoplasma gondii | 4 |
| Harghita | Sheep - mixed herds - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | Backyard | Enzyme-linked immunosorbent assay (ELISA) | animal | 7 | 4 | Toxoplasma gondii | 4 |
| Tulcea | Goats - mixed herds - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | N_A | Enzyme-linked immunosorbent assay (ELISA) | animal | 9 | 3 | Toxoplasma gondii | 3 |
| Bucureşti | Cats - pet animals - Veterinary clinics - Romania - animal sample - blood - Clinical investigations - Private sampling - Not specified | N_A | Enzyme-linked immunosorbent assay (ELISA) | animal | 4 | 2 | Toxoplasma gondii | 2 |
| Hunedoara | Goats - mixed herds - Farm - Romania - animal sample - blood - Surveillance - Official sampling - Objective sampling | N_A | Enzyme-linked immunosorbent assay (ELISA) | animal | 20 | 3 | Toxoplasma gondii | 3 |

## Table Trichinella:TRICHINELLA in animal

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| ROMANIA | Bears - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 16 | 4 | Trichinella britovi | 1 |
| | | | | | | | Trichinella spiralis | 2 |
| | | | | | | | Trichinella, unspecified sp. | 1 |
| | Pigs - breeding animals - raised under controlled housing conditions - sows and boars - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 6950 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 252310 | 134 | Trichinella britovi | 28 |
| | | | | | | | Trichinella spiralis | 81 |
| | | | | | | | Trichinella, unspecified sp. | 25 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 4582070 | 0 | Trichinella | 0 |
| | Solipeds, domestic - horses - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Horses from backyards | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 13871 | 0 | Trichinella | 0 |
| | | Horses from breeders (farms) | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 16087 | 0 | Trichinella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| ROMANIA | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 16925 | 166 | Trichinella britovi | 96 |
| | | | | | | | Trichinella spiralis | 47 |
| | | | | | | | Trichinella, unspecified sp. | 23 |
| Bihor | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 8213 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 14791 | 0 | Trichinella | 0 |
| | Solipeds, domestic - horses - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Horses from backyards | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 20 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 562 | 1 | Trichinella britovi | 1 |
| Bistriţa-Năsăud | Bears - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 2 | 0 | Trichinella | 0 |
| | Pigs - breeding animals - raised under controlled housing conditions - sows and boars - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 79 | 0 | Trichinella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Bistriţa-Năsăud | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 13449 | 8 | Trichinella spiralis | 8 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 38487 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 362 | 0 | Trichinella | 0 |
| Cluj | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 12179 | 21 | Trichinella spiralis | 16 |
| | | | | | | | Trichinella, unspecified sp. | 5 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 6576 | 0 | Trichinella | 0 |
| | Solipeds, domestic - horses - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Horses from breeders (farms) | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 96 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 267 | 5 | Trichinella britovi | 2 |
| | | | | | | | Trichinella spiralis | 2 |
| | | | | | | | Trichinella, unspecified sp. | 1 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Maramureş | Pigs - breeding animals - raised under controlled housing conditions - sows and boars - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 742 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 1762 | 1 | Trichinella britovi | 1 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 299139 | 0 | Trichinella | 0 |
| | Solipeds, domestic - horses - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Horses from backyards | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 6431 | 0 | Trichinella | 0 |
| | | Horses from breeders (farms) | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 254 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 616 | 28 | Trichinella britovi | 17 |
| | | | | | | | Trichinella spiralis | 3 |
| | | | | | | | Trichinella, unspecified sp. | 8 |
| Satu Mare | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 4929 | 26 | Trichinella britovi | 2 |
| | | | | | | | Trichinella spiralis | 7 |
| | | | | | | | Trichinella, unspecified sp. | 17 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Satu Mare | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 100712 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 1 | 1 | Trichinella, unspecified sp. | 1 |
| Sălaj | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 1986 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 516 | 1 | Trichinella britovi | 1 |
| Alba | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 15075 | 3 | Trichinella spiralis | 3 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 80203 | 0 | Trichinella | 0 |
| | Solipeds, domestic - horses - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Horses from backyards | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 1154 | 0 | Trichinella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Alba | Solipeds, domestic - horses - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Horses from breeders (farms) | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 1121 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 813 | 1 | Trichinella britovi | 1 |
| Braşov | Pigs - breeding animals - raised under controlled housing conditions - sows and boars - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 4 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 12705 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 73239 | 0 | Trichinella | 0 |
| | Solipeds, domestic - horses - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Horses from backyards | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 191 | 0 | Trichinella | 0 |
| | | Horses from breeders (farms) | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 8389 | 0 | Trichinella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Braşov | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 19 | 0 | Trichinella | 0 |
| Covasna | Bears - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 9 | 1 | Trichinella spiralis | 1 |
| | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 4665 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 64981 | 0 | Trichinella | 0 |
| | Solipeds, domestic - horses - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Horses from backyards | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 423 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 93 | 0 | Trichinella | 0 |
| Harghita | Bears - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 2 | 2 | Trichinella spiralis | 1 |
| | | | | | | | Trichinella, unspecified sp. | 1 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Harghita | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 5321 | 5 | Trichinella spiralis | 5 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 2 | 2 | Trichinella britovi | 1 |
| | | | | | | | Trichinella, unspecified sp. | 1 |
| Mureş | Pigs - breeding animals - raised under controlled housing conditions - sows and boars - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 540 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 19889 | 10 | Trichinella spiralis | 10 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 55463 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 144 | 0 | Trichinella | 0 |
| Sibiu | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 3060 | 1 | Trichinella britovi | 1 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Sibiu | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 40674 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 501 | 4 | Trichinella britovi | 1 |
| | | | | | | | Trichinella spiralis | 3 |
| Bacău | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 490 | 6 | Trichinella spiralis | 5 |
| | | | | | | | Trichinella, unspecified sp. | 1 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 82835 | 0 | Trichinella | 0 |
| | Solipeds, domestic - horses - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Horses from backyards | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 72 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 2426 | 17 | Trichinella britovi | 9 |
| | | | | | | | Trichinella spiralis | 5 |
| | | | | | | | Trichinella, unspecified sp. | 3 |
| Botoşani | Pigs - breeding animals - raised under controlled housing conditions - sows and boars - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 24 | 0 | Trichinella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Botoşani | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 16310 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 75903 | 0 | Trichinella | 0 |
| | Solipeds, domestic - horses - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Horses from backyards | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 160 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 25 | 1 | Trichinella britovi | 1 |
| Iaşi | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 192 | 8 | Trichinella spiralis | 8 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 363074 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 366 | 1 | Trichinella britovi | 1 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Neamț | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 404 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 38057 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 40 | 0 | Trichinella | 0 |
| Suceava | Bears - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 3 | 1 | Trichinella britovi | 1 |
| | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 53 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 1951 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 803 | 18 | Trichinella britovi | 18 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Vaslui | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 9287 | 1 | Trichinella, unspecified sp. | 1 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 3206 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 265 | 1 | Trichinella britovi | 1 |
| Brăila | Pigs - breeding animals - raised under controlled housing conditions - sows and boars - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 80 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 1216 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 195154 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 2 | 2 | Trichinella spiralis | 2 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Buzău | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 4775 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 34408 | 0 | Trichinella | 0 |
| | Solipeds, domestic - horses - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Horses from breeders (farms) | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 142 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 1252 | 5 | Trichinella britovi | 3 |
| | | | | | | | Trichinella spiralis | 2 |
| Constanţa | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 4552 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 59962 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 1 | 1 | Trichinella britovi | 1 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Galaţi | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 2950 | 20 | Trichinella britovi | 11 |
| | | | | | | | Trichinella spiralis | 9 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 22119 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 465 | 1 | Trichinella spiralis | 1 |
| Tulcea | Pigs - breeding animals - raised under controlled housing conditions - sows and boars - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 900 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 543 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 39542 | 0 | Trichinella | 0 |
| | Solipeds, domestic - horses - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Horses from backyards | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 27 | 0 | Trichinella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Tulcea | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 1 | 1 | Trichinella britovi | 1 |
| Vrancea | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 438 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 151370 | 0 | Trichinella | 0 |
| | Solipeds, domestic - horses - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Horses from backyards | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 2 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 75 | 10 | Trichinella britovi | 8 |
| | | | | | | | Trichinella spiralis | 2 |
| Argeş | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 576 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 163723 | 0 | Trichinella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Călăraşi | Pigs - breeding animals - raised under controlled housing conditions - sows and boars - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 2637 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 1371 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 127552 | 0 | Trichinella | 0 |
| | Solipeds, domestic - horses - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Horses from backyards | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 2528 | 0 | Trichinella | 0 |
| | | Horses from breeders (farms) | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 2593 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 681 | 2 | Trichinella britovi | 1 |
| | | | | | | | Trichinella spiralis | 1 |
| Dâmboviţa | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 821 | 1 | Trichinella spiralis | 1 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Dâmboviţa | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 393214 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 542 | 5 | Trichinella britovi | 2 |
| | | | | | | | Trichinella spiralis | 3 |
| Giurgiu | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 2746 | 2 | Trichinella spiralis | 2 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 2 | 2 | Trichinella spiralis | 1 |
| | | | | | | | Trichinella, unspecified sp. | 1 |
| Ialomiţa | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 5361 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 141668 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 372 | 0 | Trichinella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Prahova | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 12022 | 7 | Trichinella britovi | 6 |
| | | | | | | | Trichinella spiralis | 1 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 142528 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 1835 | 16 | Trichinella britovi | 10 |
| | | | | | | | Trichinella spiralis | 6 |
| Teleorman | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 11605 | 3 | Trichinella spiralis | 3 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 4051 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 675 | 5 | Trichinella britovi | 2 |
| | | | | | | | Trichinella spiralis | 3 |
| Bucureşti - Ilfov | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 6855 | 0 | Trichinella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Bucureşti - Ilfov | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 89025 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 2 | 0 | Trichinella | 0 |
| Bucureşti | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 122 | 0 | Trichinella | 0 |
| Dolj | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 9109 | 2 | Trichinella britovi | 2 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 4448 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 17 | 0 | Trichinella | 0 |
| Gorj | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 5193 | 0 | Trichinella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Gorj | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 12902 | 0 | Trichinella | 0 |
| Mehedinţi | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 5935 | 2 | Trichinella britovi | 1 |
| | | | | | | | Trichinella spiralis | 1 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 1960 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 277 | 0 | Trichinella | 0 |
| Olt | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 5596 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 10901 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 740 | 17 | Trichinella britovi | 8 |
| | | | | | | | Trichinella spiralis | 7 |
| | | | | | | | Trichinella, unspecified sp. | 2 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Vâlcea | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 1320 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 90784 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 380 | 0 | Trichinella | 0 |
| Arad | Pigs - breeding animals - raised under controlled housing conditions - sows and boars - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 1404 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 3750 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 53299 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 314 | 14 | Trichinella britovi | 4 |
| | | | | | | | Trichinella spiralis | 4 |
| | | | | | | | Trichinella, unspecified sp. | 6 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Caraş-Severin | Pigs - breeding animals - raised under controlled housing conditions - sows and boars - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 540 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 15886 | 5 | Trichinella britovi | 3 |
| | | | | | | | Trichinella spiralis | 2 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 168516 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 118 | 0 | Trichinella | 0 |
| Hunedoara | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 13545 | 2 | Trichinella britovi | 1 |
| | | | | | | | Trichinella, unspecified sp. | 1 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 2500 | 0 | Trichinella | 0 |
| | Solipeds, domestic - horses - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Horses from backyards | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 140 | 0 | Trichinella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling Details | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|---|---|---|---|---|---|---|---|---|
| Hunedoara | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 167 | 3 | Trichinella britovi | 2 |
| | | | | | | | Trichinella spiralis | 1 |
| Timiş | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Fattening pigs from backyards and free-range pigs not raised under controlled housing conditions | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 6054 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 1333153 | 0 | Trichinella | 0 |
| | Solipeds, domestic - horses - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | Horses from backyards | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 2723 | 0 | Trichinella | 0 |
| | | Horses from breeders (farms) | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 3492 | 0 | Trichinella | 0 |
| | Wild boars - wild - Slaughterhouse - Romania - animal sample - organ/tissue - Surveillance - Official sampling - Objective sampling | N_A | Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test | animal | 1186 | 1 | Trichinella spiralis | 1 |

# FOODBORNE OUTBREAKS TABLES

## Foodborne Outbreaks: summarized data

| | | Outbreak strenght | Strong | | | Weak | | | |
|---|---|---|---|---|---|---|---|---|---|
| Causative agent | Food vehicle | N outbreaks | N human cases | N hospitalized | N deaths | N outbreaks | N human cases | N hospitalized | N deaths |
| Clostridium botulinum | Other foods | 1 | 2 | 2 | 1 | | | | |
| Clostridium botulinum toxins | Pig meat and products thereof | 1 | 2 | 1 | 0 | | | | |
| Enterococcus | Tap water, including well water | 1 | 4 | 4 | 0 | | | | |
| Escherichia coli | Cheese | 1 | 7 | 2 | 0 | | | | |
| | Buffet meals | 2 | 11 | 8 | 0 | | | | |
| Listeria monocytogenes | Buffet meals | 1 | 19 | 0 | 0 | | | | |
| Salmonella | Other foods | 1 | 15 | 14 | 0 | | | | |
| | Buffet meals | 2 | 235 | 144 | 0 | | | | |
| Salmonella Enteritidis | Other foods | 1 | 217 | 0 | 0 | | | | |
| | Buffet meals | 1 | 19 | 19 | 0 | | | | |
| Salmonella Infantis | Buffet meals | 1 | 8 | 4 | 0 | | | | |
| Staphylococcal enterotoxins | Cheese | 1 | 8 | 5 | 0 | | | | |
| Staphylococcus | Buffet meals | 2 | 37 | 37 | 0 | | | | |
| Staphylococcus aureus | Other, mixed or unspecified poultry meat and products thereof | 1 | 5 | 1 | 0 | | | | |
| | Buffet meals | 1 | 2 | 1 | 0 | | | | |
| Trichinella britovi | Other or mixed red meat and products thereof | 1 | 4 | 4 | 0 | | | | |
| Trichinella spiralis | Pig meat and products thereof | 5 | 60 | 60 | 0 | | | | |
| Unknown | Cheese | | | | | 1 | 30 | 0 | 0 |
| | Other foods | | | | | 1 | 3 | 3 | 0 |
| | Buffet meals | | | | | 3 | 50 | 15 | 0 |

## Strong Foodborne Outbreaks: detailed data

| Causative agent | Other Causative Agent | FBO nat. code | Outbreak type | Food vehicle | More food vehicle info | Nature of evidence | Setting | Place of origin of problem | Origin of food vehicle | Contributory factors | Comment | N outbreaks | N human cases | N hosp. | N deaths |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| Clostridium botulinum | Not Available | N_A | Unknown | Other foods | Pig meat products and fried fish | Descriptive environmental evidence | Household | Household | Romania | Other contributory factor | Detection of botulinum neurotoxin clostridium in sample was negative. The toxin was not identified in food but in the blood serum of patients harvested in the hospital. No evidence could be taken to correlate illness and if the causative agent had a food or tellurium vehicle or other food preserved and consumed by the two human cases | 1 | 2 | 2 | 1 |
| Clostridium botulinum toxins | Not Available | N_A | Unknown | Pig meat and products thereof | Pig meat products (pork ham) | Detection of causative agent in food vehicle or its component - Detection of indistinguishable causative agent in humans | Household | Household | Romania | Other contributory factor | Detection of botulinum neurotoxin clostridium in sample and human case - positive for genotype B | 1 | 2 | 1 | 0 |
| Enterococcus | Escherichia coli;Enterococcus | N_A | General | Tap water, including well water | Water (own source) used in culinary preparations | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent | Restaurant or Cafe or Pub or Bar or Hotel or Catering service | Restaurant or Cafe or Pub or Bar or Hotel or Catering service | Romania | Water treatment failure | The source of contamination was drinking water (own source) used in food preparation and workplace hygiene (present Intestinal enterococcus, Coliform bacteria, E. Coli). No pathogens were identified in the raw materials used in the preparation of menus. | 1 | 4 | 4 | 0 |
| Escherichia coli | Escherichia coli;Staphylococcus | N_A | General | Buffet meals | Mixed foodstuffs: prepared meals and snacks | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent | Restaurant or Cafe or Pub or Bar or Hotel or Catering service | Restaurant or Cafe or Pub or Bar or Hotel or Catering service | Romania | Infected food handler;Cross-contamination | The causative agent was not identified in humans cases. The case was classified based on clinical and epidemiological data according to surveillance methodology. | 1 | 8 | 8 | 0 |
| | Not Available | N_A | General | Cheese | Cheeses made from cows' milk - unspecified | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent | Others | Household | Romania | Other contributory factor | Real-time PCR screening for genes detection stx1, eae: positive. Real-time PCR screening for detection of genes associated with O145 and O26: positive but without bacterial isolation.The case was classified based on clinical and epidemiological data according to surveillance methodology. | 1 | 7 | 2 | 0 |

| Causative agent | Other Causative Agent | FBO nat. code | Outbreak type | Food vehicle | More food vehicle info | Nature of evidence | Setting | Place of origin of problem | Origin of food vehicle | Contributory factors | Comment | N outbreaks | N human cases | N hosp. | N deaths |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| Escherichia coli | Not Available | N_A | Unknown | Buffet meals | Food dishes: Grilled chicken breast with potatoes, fresh cow's cheese, donuts with sour cream | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent | Restaurant or Cafe or Pub or Bar or Hotel or Catering service | Restaurant or Cafe or Pub or Bar or Hotel or Catering service | Romania | Storage time/temperature abuse;Other contributory factor | The causative agent was not identified in humans cases. The case was classified based on clinical and epidemiological data according to surveillance methodology. | 1 | 3 | 0 | 0 |
| Listeria monocytogenes | Staphylococcus aureus;Escherichia coli;Salmonella Infantis | N_A | General | Buffet meals | Food dishes: fresh chicken meat, pressed ham, meat chicken products redy to eat, cheeses made from cows' milk | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent | Restaurant or Cafe or Pub or Bar or Hotel or Catering service | Restaurant or Cafe or Pub or Bar or Hotel or Catering service | Romania | Storage time/temperature abuse;Other contributory factor | In food dishes were identified: Listeria monocytogenes (in pressed ham and meat chicken products ready to eat); Salmonella Infantis (in fresh chicken meat), Staphylococcus aureus (in pressed ham and cheeses); E.coli in cheeses. The causative agent was not identified in humans cases. The case was classified based on clinical and epidemiological data according to surveillance methodology. | 1 | 19 | 0 | 0 |
| Salmonella | Not Available | N_A | General | Other foods | A traditional product from wheat served at funeral ceremony. | Detection of causative agent in food chain or its environment - Detection of indistinguishable causative agent in humans | Restaurant or Cafe or Pub or Bar or Hotel or Catering service | Household | Romania | Infected food handler | The human cases involved participated at a meal organized after a funeral ceremony. The product was prepared in the house by the organizer of the event, the person who showed clinical signs and from the biological samples collected , has been identified Salmonella group D. Salmonella has also been identified in the hospitalized cases. | 1 | 15 | 14 | 0 |
| | | | | Buffet meals | Food dishes (aperitif, meat, vegetables, meat soup) | Detection of causative agent in food vehicle or its component - Detection of indistinguishable causative agent in humans | Others | Restaurant or Cafe or Pub or Bar or Hotel or Catering service | Romania | Infected food handler | The causative agent ( Salmonella group D) was identified in food samples and humans. | 1 | 9 | 9 | 0 |
| | Staphylococcus;Salmonella | N_A | General | Buffet meals | Mayonnaise sauce, pizza, mini crispy, sandwich, crispy, shaorma served with mayonnaise and bechamel sauce | Detection of causative agent in food vehicle or its component - Detection of indistinguishable causative agent in humans | Take-away or fast-food outlet | Take-away or fast-food outlet | Romania | Infected food handler;Cross-contamination | Pathogenic infections (Salmonella group D si Staphylococcal coagulase positive) have been identified in the raw materials used in food preparation and human cases. | 1 | 226 | 135 | 0 |

| Causative agent | Other Causative Agent | FBO nat. code | Outbreak type | Food vehicle | More food vehicle info | Nature of evidence | Setting | Place of origin of problem | Origin of food vehicle | Contributory factors | Comment | N outbreaks | N human cases | N hosp. | N deaths |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| Salmonella Enteritidis | Not Available | N_A | General | Other foods | Fresh poultry meat (chicken legs), Fresh cow cheese, Sour cream, Milk pasteurized, Cheese pastries, Cookies | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent | Canteen or workplace catering | Canteen or workplace catering | Romania | Cross-contamination | The case was classified based on clinical and epidemiological data according to surveillance methodology. | 1 | 217 | 0 | 0 |
| | | | | Buffet meals | Prepared meals and snacks | Detection of causative agent in food chain or its environment - Detection of indistinguishable causative agent in humans | Restaurant or Cafe or Pub or Bar or Hotel or Catering service | Restaurant or Cafe or Pub or Bar or Hotel or Catering service | Romania | Infected food handler | The source was represented by the human carrier (the causative agent has been identified in humans no food vehicle was incriminated). | 1 | 19 | 19 | 0 |
| Salmonella Infantis | Not Available | N_A | General | Buffet meals | Cooking food (chicken with rice) | Detection of causative agent in food vehicle or its component - Detection of indistinguishable causative agent in humans | Canteen or workplace catering | Canteen or workplace catering | Romania | Other contributory factor;Cross-contamination | Pathogenic germs were identified in the raw materials used in the preparation (Salmonella in chicken breast and Enterobacteriaceae in rice). The causative agent was identified in humans as well. | 1 | 8 | 4 | 0 |
| Staphylococcal enterotoxins | Staphylococcal enterotoxins - Enterotoxin H;Staphylococcal enterotoxins - Enterotoxin C | N_A | General | Cheese | Cheeses made from sheep's milk - unspecified | Detection of causative agent in food vehicle or its component - Detection of indistinguishable causative agent in humans | Household | Farm | Romania | Infected food handler | Detection of staphylococcal enterotoxin coding genes (positive for types H, C, I) by multiplex PCR in the analyzed samples. The results obtained from laboratory tests identified Staphylococcus aureus in food samples, in sanitary tests and in humans (nasal and pharyngeal exudates). | 1 | 8 | 5 | 0 |
| Staphylococcus | Not Available | N_A | General | Buffet meals | Food dishes (cheese pie and spinach, peppers stuffed with cheese) | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent | Restaurant or Cafe or Pub or Bar or Hotel or Catering service | Restaurant or Cafe or Pub or Bar or Hotel or Catering service | Romania | Unknown | No pathogenic germs were identified in the raw materials used in the preparation. The causal agent was identified in the served aperitif (prepared meals). The case was classified based on clinical and epidemiological data according to surveillance methodology. | 1 | 24 | 24 | 0 |
| | | | | | Prepared meals and snacks | Descriptive environmental evidence;Descriptive environmental evidence | Restaurant or Cafe or Pub or Bar or Hotel or Catering service | Restaurant or Cafe or Pub or Bar or Hotel or Catering service | Romania | Infected food handler | The source was represented by the human carrier (the causative agent has been identified in humans no food vehicle was incriminated).The case was classified based on clinical and epidemiological data according to surveillance methodology. | 1 | 13 | 13 | 0 |

| Causative agent | Other Causative Agent | FBO nat. code | Outbreak type | Food vehicle | More food vehicle info | Nature of evidence | Setting | Place of origin of problem | Origin of food vehicle | Contributory factors | Comment | N outbreaks | N human cases | N hosp. | N deaths |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| Staphylococcus aureus | Not Available | N_A | General | Other, mixed or unspecified poultry meat and products thereof | Cooked poultry meat, boiled eggs | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent | Take-away or fast-food outlet | Take-away or fast-food outlet | Non European Union | Infected food handler | The pathogen was isolated in prepared chicken breast and boiled eggs. The case was classified based on clinical and epidemiological data according to surveillance methodology. | 1 | 5 | 1 | 0 |
| | | | | Buffet meals | Grilled pork, gratinated potatoes, cabbage salad | Descriptive environmental evidence | Canteen or workplace catering | Canteen or workplace catering | Romania | Infected food handler | The source was represented by the human carrier (the causative agent has been identified in humans no food vehicle was incriminated).The case was classified based on clinical and epidemiological data according to surveillance methodology. | 1 | 2 | 1 | 0 |
| Trichinella britovi | Not Available | N_A | Unknown | Other or mixed red meat and products thereof | Wild boar meat | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent | Household | Others | Romania | Inadequate heat treatment | Consumption of infected wild boar meat (without sanitary - veterinary examination).The case was classified based on clinical and epidemiological data according to surveillance methodology. | 1 | 4 | 4 | 0 |
| Trichinella spiralis | Not Available | N_A | General | Pig meat and products thereof | Pork and products (sausages and other products) | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent | Household | Household | Romania | Inadequate heat treatment | Consumption of infected pigs from households (without sanitary - veterinary examination).The case was classified based on clinical and epidemiological data according to surveillance methodology. | 5 | 60 | 60 | 0 |

## Weak Foodborne Outbreaks: detailed data

| Causative agent | Other Causative Agent | FBO nat. code | Outbreak type | Food vehicle | More food vehicle info | Nature of evidence | Setting | Place of origin of problem | Origin of food vehicle | Contributory factors | Comment | N outbreaks | N human cases | N hosp. | N deaths |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| Unknown | Not Available | N_A | General | Cheese | Cheeses made from cows' milk - unspecified | Descriptive environmental evidence;Descriptive environmental evidence | Canteen or workplace catering | Canteen or workplace catering | Romania | Other contributory factor;Cross-contamination | The case was classified based on clinical and epidemiological data according to surveillance methodology. | 1 | 30 | 0 | 0 |
| | | | | Buffet meals | Cheese, salami, tomatoes, cucumbers, pressed ham | Descriptive environmental evidence;Descriptive environmental evidence | Household | Household | Romania | Other contributory factor;Cross-contamination | The case was classified based on clinical and epidemiological data according to surveillance methodology. | 1 | 8 | 7 | 0 |
| | | | | | Food dishes (meat pork pruducts, cheese, salami, sausages, meatballs, ham) | Descriptive environmental evidence;Descriptive environmental evidence | Household | Household | Romania | Other contributory factor | The case was classified based on clinical and epidemiological data according to surveillance methodology. | 1 | 8 | 8 | 0 |
| | | | | | Prepared meals and snacks | Descriptive environmental evidence;Descriptive environmental evidence | Canteen or workplace catering | Canteen or workplace catering | Romania | Other contributory factor;Cross-contamination | The case was classified based on clinical and epidemiological data according to surveillance methodology. | 1 | 34 | 0 | 0 |
| | | | Unknown | Other foods | Homemade foods and chocolate raisins | Descriptive environmental evidence;Descriptive environmental evidence | Household | Household | Romania | Other contributory factor;Cross-contamination | The only common food was raisins in chocolate (from the supermarket). The case was classified based on clinical and epidemiological data according to surveillance methodology. | 1 | 3 | 3 | 0 |

## ANTIMICROBIAL RESISTANCE TABLES FOR CAMPYLOBACTER

### Table Antimicrobial susceptibility testing of Campylobacter coli in Meat from broilers (Gallus gallus) - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Surveillance - based on Regulation 2073

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Romania

Sampling details:

| | AM substance | Ciprofloxacin | Erythromycin | Gentamicin | Nalidixic acid | Streptomycin | Tetracycline |
|---|---|---|---|---|---|---|---|
| | **ECOFF** | **0.5** | **8** | **2** | **16** | **4** | **2** |
| | **Lowest limit** | **0.12** | **1** | **0.12** | **1** | **0.25** | **0.5** |
| | **Highest limit** | **16** | **128** | **16** | **64** | **16** | **64** |
| | **N of tested isolates** | **2** | **2** | **2** | **2** | **2** | **2** |
| **MIC** | **N of resistant isolates** | **2** | **0** | **0** | **2** | **0** | **1** |
| 0.25 | | | | 1 | | | |
| <=0.5 | | | | | | | 1 |
| 0.5 | | | | 1 | | | |
| <=1 | | | 2 | | | | |
| 1 | | | | | | 1 | |
| 4 | | | | | | 1 | |
| 8 | | 1 | | | | | |
| >16 | | 1 | | | | | |
| 64 | | | | | 1 | | 1 |
| >64 | | | | | 1 | | |

## Table Antimicrobial susceptibility testing of Campylobacter coli in Meat from broilers (Gallus gallus) - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Surveillance - based on Regulation 2073

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Romania

Sampling details:

| | AM substance | Ciprofloxacin | Erythromycin | Gentamicin | Nalidixic acid | Streptomycin | Tetracycline |
|---|---|---|---|---|---|---|---|
| | **ECOFF** | **0.5** | **8** | **2** | **16** | **4** | **2** |
| | **Lowest limit** | **0.12** | **1** | **0.12** | **1** | **0.25** | **0.5** |
| | **Highest limit** | **16** | **128** | **16** | **64** | **16** | **64** |
| | **N of tested isolates** | **11** | **11** | **11** | **11** | **11** | **11** |
| **MIC** | **N of resistant isolates** | **11** | **0** | **0** | **11** | **0** | **6** |
| 0.25 | | | | 1 | | | |
| <=0.5 | | | | | | | 5 |
| 0.5 | | | | 10 | | | |
| <=1 | | | 11 | | | | |
| 1 | | | | | | 1 | |
| 2 | | | | | | 10 | |
| 8 | | 1 | | | | | |
| 16 | | 5 | | | | | |
| >16 | | 5 | | | | | |
| 64 | | | | | | | 2 |
| >64 | | | | | 11 | | 4 |

## Table Antimicrobial susceptibility testing of Campylobacter coli in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Slaughterhouse

Sampling Type: food sample - meat

Sampling Context: Surveillance - based on Regulation 2073

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Romania

Sampling details:

| | AM substance | Ciprofloxacin | Erythromycin | Gentamicin | Nalidixic acid | Streptomycin | Tetracycline |
|---|---|---|---|---|---|---|---|
| | **ECOFF** | **0.5** | **8** | **2** | **16** | **4** | **2** |
| | **Lowest limit** | **0.12** | **1** | **0.12** | **1** | **0.25** | **0.5** |
| | **Highest limit** | **16** | **128** | **16** | **64** | **16** | **64** |
| | **N of tested isolates** | **6** | **6** | **6** | **6** | **6** | **6** |
| **MIC** | **N of resistant isolates** | **6** | **0** | **0** | **6** | **0** | **1** |
| <=0.5 | | | | | | | 5 |
| 0.5 | | | | 6 | | | |
| <=1 | | | 6 | | | | |
| 2 | | | | | | 6 | |
| 16 | | 3 | | | | | |
| >16 | | 3 | | | | | |
| 64 | | | | | | | 1 |
| >64 | | | | | 6 | | |

## Table Antimicrobial susceptibility testing of Campylobacter jejuni in Meat from broilers (Gallus gallus) - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Surveillance - based on Regulation 2073

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Romania

Sampling details:

| | AM substance | Ciprofloxacin | Erythromycin | Gentamicin | Nalidixic acid | Streptomycin | Tetracycline |
|---|---|---|---|---|---|---|---|
| | **ECOFF** | **0.5** | **4** | **2** | **16** | **4** | **1** |
| | **Lowest limit** | **0.12** | **1** | **0.12** | **1** | **0.25** | **0.5** |
| | **Highest limit** | **16** | **128** | **16** | **64** | **16** | **64** |
| | **N of tested isolates** | **2** | **2** | **2** | **2** | **2** | **2** |
| **MIC** | **N of resistant isolates** | **1** | **0** | **0** | **1** | **0** | **2** |
| <=0.12 | | 1 | | | | | |
| 0.25 | | | | 2 | | | |
| <=1 | | | 2 | | | | |
| 1 | | | | | | 2 | |
| 8 | | 1 | | | 1 | | |
| 64 | | | | | | | 1 |
| >64 | | | | | 1 | | 1 |

## Table Antimicrobial susceptibility testing of Campylobacter jejuni in Meat from broilers (Gallus gallus) - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Surveillance - based on Regulation 2073

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Romania

Sampling details:

| | AM substance | Ciprofloxacin | Erythromycin | Gentamicin | Nalidixic acid | Streptomycin | Tetracycline |
|---|---|---|---|---|---|---|---|
| | **ECOFF** | **0.5** | **4** | **2** | **16** | **4** | **1** |
| | **Lowest limit** | **0.12** | **1** | **0.12** | **1** | **0.25** | **0.5** |
| | **Highest limit** | **16** | **128** | **16** | **64** | **16** | **64** |
| | **N of tested isolates** | **18** | **18** | **18** | **18** | **18** | **18** |
| **MIC** | **N of resistant isolates** | **6** | **0** | **0** | **6** | **0** | **7** |
| <=0.12 | | 9 | | 4 | | | |
| <=0.25 | | | | | | 3 | |
| 0.25 | | | | 12 | | | |
| <=0.5 | | | | | | | 8 |
| 0.5 | | 3 | | 2 | | 1 | |
| <=1 | | | 18 | | | | |
| 1 | | | | | | 10 | 3 |
| 2 | | | | | 5 | 4 | |
| 4 | | | | | 3 | | |
| 8 | | 1 | | | 4 | | |
| 16 | | 5 | | | | | |
| 32 | | | | | | | 1 |
| 64 | | | | | 3 | | 6 |
| >64 | | | | | 3 | | |

## Table Antimicrobial susceptibility testing of Campylobacter jejuni in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Slaughterhouse

Sampling Type: food sample - meat

Sampling Context: Surveillance - based on Regulation 2073

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Romania

Sampling details:

| | AM substance | Ciprofloxacin | Erythromycin | Gentamicin | Nalidixic acid | Streptomycin | Tetracycline |
|---|---|---|---|---|---|---|---|
| | **ECOFF** | **0.5** | **4** | **2** | **16** | **4** | **1** |
| | **Lowest limit** | **0.12** | **1** | **0.12** | **1** | **0.25** | **0.5** |
| | **Highest limit** | **16** | **128** | **16** | **64** | **16** | **64** |
| | **N of tested isolates** | **3** | **3** | **3** | **3** | **3** | **3** |
| **MIC** | **N of resistant isolates** | **2** | **0** | **0** | **2** | **0** | **0** |
| <=0.12 | | 1 | | | | | |
| 0.25 | | | | 3 | | | |
| <=0.5 | | | | | | | 3 |
| <=1 | | | 3 | | | | |
| 1 | | | | | | 3 | |
| 2 | | | | | 1 | | |
| 8 | | 2 | | | | | |
| 64 | | | | | 2 | | |

## Table Antimicrobial susceptibility testing of Campylobacter jejuni in Gallus gallus (fowl) - broilers

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling details:

| | AM substance | Ciprofloxacin | Erythromycin | Gentamicin | Nalidixic acid | Streptomycin | Tetracycline |
|---|---|---|---|---|---|---|---|
| | **ECOFF** | **0.5** | **4** | **2** | **16** | **4** | **1** |
| | **Lowest limit** | **0.12** | **1** | **0.12** | **1** | **0.25** | **0.5** |
| | **Highest limit** | **16** | **128** | **16** | **64** | **16** | **64** |
| | **N of tested isolates** | **338** | **338** | **338** | **338** | **338** | **338** |
| **MIC** | **N of resistant isolates** | **290** | **3** | **0** | **290** | **28** | **210** |
| <=0.12 | | 44 | | 22 | | | |
| <=0.25 | | | | | | 1 | |
| 0.25 | | 4 | | 79 | | | |
| <=0.5 | | | | | | | 125 |
| 0.5 | | | | 179 | | 24 | |
| <=1 | | | 334 | | | | |
| 1 | | | | 58 | | 140 | 3 |
| 2 | | 2 | 1 | | 3 | 138 | |
| 4 | | 11 | | | 30 | 7 | |
| 8 | | 165 | | | 15 | 1 | 1 |
| 16 | | 80 | | | | 3 | 10 |
| >16 | | 32 | | | | 24 | |
| 32 | | | | | 20 | | 12 |
| 64 | | | | | 62 | | 69 |
| >64 | | | | | 208 | | 118 |
| >128 | | | 3 | | | | |

## Table Antimicrobial susceptibility testing of Campylobacter jejuni in Turkeys - fattening flocks

Sampling Stage: Slaughterhouse | Sampling Type: animal sample - caecum | Sampling Context: Monitoring

Sampler: Official sampling | Sampling Strategy: Objective sampling | Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling details:

| | AM substance | Ciprofloxacin | Erythromycin | Gentamicin | Nalidixic acid | Streptomycin | Tetracycline |
|---|---|---|---|---|---|---|---|
| | **ECOFF** | **0.5** | **4** | **2** | **16** | **4** | **1** |
| | **Lowest limit** | **0.12** | **1** | **0.12** | **1** | **0.25** | **0.5** |
| | **Highest limit** | **16** | **128** | **16** | **64** | **16** | **64** |
| | **N of tested isolates** | **7** | **7** | **7** | **7** | **7** | **7** |
| **MIC** | **N of resistant isolates** | **4** | **0** | **0** | **4** | **0** | **4** |
| <=0.12 | | 3 | | | | | |
| <=0.5 | | | | | | | 3 |
| 0.5 | | | | 6 | | | |
| <=1 | | | 7 | | | | |
| 1 | | | | 1 | | 1 | |
| 2 | | | | | | 5 | |
| 4 | | | | | 3 | 1 | |
| 8 | | 3 | | | | | |
| 16 | | 1 | | | | | |
| 64 | | | | | | | 1 |
| >64 | | | | | 4 | | 3 |

## ANTIMICROBIAL RESISTANCE TABLES FOR SALMONELLA

### Table Antimicrobial susceptibility testing of Salmonella Agona in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **1** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 5 | | | | | |
| 0.03 | | | | | | | 5 | | | | | | | | |
| <=0.25 | | | | 5 | | | | | | | | | | 4 | 2 |
| <=0.5 | | | | | 5 | | | | 5 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 2 |
| <=1 | | 4 | | | | | | 5 | | | | | | | |
| 1 | | | | | | | | | | | | | | | 1 |
| <=2 | | | | | | | | | | | | | 5 | | |
| 2 | | 1 | | | | | | | | | | | | | |
| <=4 | | | | | | | | | | | 4 | | | | |
| <=8 | | | | | | 5 | | | | | | | | | |
| 8 | | | 5 | | | | | | | | 1 | | | | |
| 32 | | | | | | | | | | | | 2 | | | |
| 64 | | | | | | | | | | | | 2 | | | |
| >1024 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Albany in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Amsterdam in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| 0.03 | | | | | | | 1 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | 1 | | | |
| 8 | | | 1 | | | | | | | | | | | | |

## Table Antimicrobial susceptibility testing of Salmonella Bovismorbificans in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **4** | **4** | **4** | **4** | **4** | **4** | **4** | **4** | **4** | **4** | **4** | **4** | **4** | **4** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.015 | | | | | | | 4 | | | | | | | | |
| <=0.03 | | | | | | | | | | 4 | | | | | |
| <=0.25 | | | | 4 | | | | | | | | | | 4 | 4 |
| <=0.5 | | | | | 4 | | | | 4 | | | | | | |
| <=1 | | 4 | | | | | | 4 | | | | | | | |
| <=2 | | | | | | | | | | | | | 4 | | |
| <=4 | | | | | | | | | | | 4 | | | | |
| 4 | | | 4 | | | | | | | | | | | | |
| <=8 | | | | | | 4 | | | | | | 1 | | | |
| 16 | | | | | | | | | | | | 2 | | | |
| 32 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Bredeney in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **1** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| 0.25 | | | | | | | 1 | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=8 | | | | | | 1 | | | | | | 1 | | | |
| 8 | | | 1 | | | | | | | | 1 | | | | |

## Table Antimicrobial susceptibility testing of Salmonella Bredeney in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **5** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 4 | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 5 | | | | | | | | | | 4 | 4 |
| 0.25 | | | | | | | 5 | | | | | | | | |
| <=0.5 | | | | | 5 | | | | 5 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 1 |
| <=1 | | 4 | | | | | | 2 | | | | | | | |
| <=2 | | | | | | | | | | | | | 5 | | |
| 2 | | 1 | | | | | | 3 | | | | | | | |
| <=8 | | | | | | 5 | | | | | | 2 | | | |
| 8 | | | 5 | | | | | | | | 5 | | | | |
| 16 | | | | | | | | | | | | 1 | | | |
| 32 | | | | | | | | | | | | 2 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Cubana in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | 1 | | | | | | 1 | | | | | | | |
| <=8 | | | | | | 1 | | | | | | 1 | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | 1 | | | | |

## Table Antimicrobial susceptibility testing of Salmonella Derby in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.015 | | | | | | | 3 | | | | | | | | |
| <=0.03 | | | | | | | | | | 3 | | | | | |
| <=0.25 | | | | 3 | | | | | | | | | | 3 | 3 |
| <=0.5 | | | | | 3 | | | | 3 | | | | | | |
| <=1 | | 3 | | | | | | | | | | | | | |
| <=2 | | | | | | | | | | | | | 3 | | |
| 2 | | | | | | | | 3 | | | | | | | |
| <=4 | | | | | | | | | | | 3 | | | | |
| <=8 | | | | | | 3 | | | | | | | | | |
| 8 | | | 3 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 3 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **3** | **0** | **0** | **0** | **3** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| 0.12 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 3 | | | | | | | | | | 3 | 3 |
| 0.25 | | | | | | | 2 | | | | | | | | |
| <=0.5 | | | | | 3 | | | | 3 | | | | | | |
| <=1 | | 2 | | | | | | 2 | | | | | | | |
| <=2 | | | | | | | | | | | | | 3 | | |
| 2 | | 1 | | | | | | 1 | | | | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 3 | | | | | | 1 | | | |
| 8 | | | 2 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |
| 64 | | | | | | | | | | | | 1 | | | |
| 128 | | | | | | | | | | | 1 | | | | |
| >128 | | | | | | | | | | | 2 | | | | |

## Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **7** | **7** | **7** | **7** | **7** | **7** | **7** | **7** | **7** | **7** | **7** | **7** | **7** | **7** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.015 | | | | | | | 2 | | | | | | | | |
| <=0.03 | | | | | | | | | | 6 | | | | | |
| 0.03 | | | | | | | 5 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 7 | | | | | | | | | | 7 | 7 |
| <=0.5 | | | | | 7 | | | | 7 | | | | | | |
| <=1 | | | | | | | | 2 | | | | | | | |
| <=2 | | | | | | | | | | | | | 7 | | |
| 2 | | 7 | | | | | | 5 | | | | | | | |
| <=4 | | | | | | | | | | | 7 | | | | |
| 4 | | | 3 | | | | | | | | | | | | |
| <=8 | | | | | | 7 | | | | | | | | | |
| 8 | | | 4 | | | | | | | | | | | | |
| 64 | | | | | | | | | | | | 6 | | | |
| 128 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | 1 | | | | | | 1 | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Hadar in Meat from turkey - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **13** | **5** | **0** | **0** | **0** | **0** | **13** | **0** | **0** |
| <=0.03 | | | | | | | | | | 13 | | | | | |
| <=0.25 | | | | 13 | | | | | | | | | | | 13 |
| <=0.5 | | | | | 13 | | | | 5 | | | | | | |
| 0.5 | | | | | | | 13 | | | | | | | 13 | |
| <=1 | | 10 | | | | | | | | | | | | | |
| 1 | | | | | | | | | 8 | | | | | | |
| 2 | | 3 | | | | | | 8 | | | | | | | |
| 4 | | | 5 | | | | | 5 | | | | | | | |
| <=8 | | | | | | 13 | | | | | | | | | |
| 8 | | | 8 | | | | | | | | 6 | | | | |
| 16 | | | | | | | | | | | 7 | | | | |
| 32 | | | | | | | | | | | | 13 | | | |
| 64 | | | | | | | | | | | | | 6 | | |
| >64 | | | | | | | | | | | | | 7 | | |

## Table Antimicrobial susceptibility testing of Salmonella Hadar in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **2** | **0** | **0** | **0** | **1** | **1** | **2** | **0** | **0** |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | 2 | 2 |
| 0.25 | | | | | | | 1 | | | | | | | | |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | | |
| <=1 | | 2 | | | | | | 2 | | | | | | | |
| 4 | | | 2 | | | | | | | | | | | | |
| <=8 | | | | | | 2 | | | | | | | | | |
| 16 | | | | | | | | | | | 1 | | | | |
| 32 | | | | | | | | | | | 1 | 1 | 1 | | |
| 64 | | | | | | | | | | | | | 1 | | |
| >1024 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Hadar in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **1** | **0** | **0** | **0** | **0** | **0** | **1** | **0** | **0** |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | | |
| <=1 | | 1 | | | | | | | | | | | | | |
| 2 | | | | | | | | 1 | | | | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 16 | | | | | | | | | | | 1 | | | | |
| 32 | | | | | | | | | | | | 1 | | | |
| 64 | | | | | | | | | | | | | 1 | | |

## Table Antimicrobial susceptibility testing of Salmonella Hadar in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | | | | | | | 1 | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Hadar in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 12 | 12 | 12 | 12 | 12 | 12 | 12 | 12 | 12 | 12 | 12 | 12 | 12 | 12 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 7 | | | | | | | | |
| <=0.03 | | | | | | | | | | 12 | | | | | |
| 0.03 | | | | | | | 5 | | | | | | | | |
| <=0.25 | | | | 12 | | | | | | | | | | 11 | 12 |
| <=0.5 | | | | | 12 | | | | 12 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | 6 | | | | | | | | | | | | | |
| <=2 | | | | | | | | | | | | | 12 | | |
| 2 | | 6 | | | | | | 12 | | | | | | | |
| <=4 | | | | | | | | | | | 12 | | | | |
| 4 | | | 5 | | | | | | | | | | | | |
| <=8 | | | | | | 12 | | | | | | | | | |
| 8 | | | 7 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 12 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Infantis in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **22** | **22** | **22** | **22** | **22** | **22** | **22** | **22** | **22** | **22** | **22** | **22** | **22** | **22** |
| **MIC** | **N of resistant isolates** | **1** | **0** | **0** | **0** | **0** | **21** | **0** | **0** | **0** | **21** | **18** | **18** | **1** | **3** |
| <=0.03 | | | | | | | | | | 22 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 21 | | | | | | | | | | 1 | 18 |
| <=0.5 | | | | | 11 | | | | 22 | | | | | | |
| 0.5 | | | | 1 | | | 4 | | | | | | | 8 | 1 |
| <=1 | | | | | | | | 7 | | | | | | | |
| 1 | | | | | 11 | | 16 | | | | | | | 12 | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | 4 | | | | | 1 | 15 | | | | | | 1 | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 4 | | 17 | | | | | | | | | | | 2 | | |
| <=8 | | | | | | 11 | | | | | | | | | |
| 8 | | | 15 | | | | | | | | | | 1 | | |
| 16 | | | 7 | | | 11 | | | | | | | | | |
| 32 | | | | | | | | | | | | 4 | | | |
| >32 | | | | | | | | | | | | | | | 3 |
| >64 | | 1 | | | | | | | | | | | 18 | | |
| 128 | | | | | | | | | | | 1 | | | | |
| >128 | | | | | | | | | | | 20 | | | | |
| 1024 | | | | | | | | | | | | 1 | | | |
| >1024 | | | | | | | | | | | | 17 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Infantis in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **42** | **42** | **42** | **42** | **42** | **42** | **42** | **42** | **42** | **42** | **42** | **42** | **42** | **42** |
| **MIC** | **N of resistant isolates** | **3** | **0** | **0** | **0** | **2** | **42** | **0** | **0** | **0** | **42** | **38** | **38** | **7** | **15** |
| <=0.03 | | | | | | | | | | 40 | | | | | |
| 0.064 | | | | | | | | | | 2 | | | | | |
| <=0.25 | | | | 34 | | | | | | | | | | | 20 |
| <=0.5 | | | | | 33 | | | | 42 | | | | | | |
| 0.5 | | | | 8 | | | 18 | | | | | | | 9 | 7 |
| <=1 | | | | | | | | 27 | | | | | | | |
| 1 | | | | | 8 | | 23 | | | | | | | 26 | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | 13 | | | 1 | | | 15 | | | | | | 7 | |
| 4 | | 26 | | | | | 1 | | | | | | 3 | | |
| <=8 | | | | | | 22 | | | | | | | | | |
| 8 | | | 27 | | | | | | | | | | | | |
| 16 | | | 15 | | | 18 | | | | | | | | | |
| 32 | | | | | | 2 | | | | | | 3 | | | |
| >32 | | | | | | | | | | | | | | | 15 |
| 64 | | | | | | | | | | | | 1 | | | |
| >64 | | 3 | | | | | | | | | | | 38 | | |
| >128 | | | | | | | | | | | 42 | | | | |
| 1024 | | | | | | | | | | | | 4 | | | |
| >1024 | | | | | | | | | | | | 34 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Infantis in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **3** | **0** | **0** | **0** | **3** | **3** | **3** | **0** | **0** |
| <=0.03 | | | | | | | | | | 3 | | | | | |
| <=0.25 | | | | 3 | | | | | | | | | | | 2 |
| <=0.5 | | | | | 2 | | | | 3 | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | 1 | 1 |
| 1 | | | | | 1 | | 2 | | | | | | | 2 | |
| 2 | | 3 | | | | | | 3 | | | | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 2 | | | | | | | | | |
| 8 | | | 2 | | | | | | | | | | | | |
| 16 | | | | | | 1 | | | | | | | | | |
| >64 | | | | | | | | | | | | | 3 | | |
| >128 | | | | | | | | | | | 3 | | | | |
| 1024 | | | | | | | | | | | | 1 | | | |
| >1024 | | | | | | | | | | | | 2 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **2** | **0** | **0** | **0** | **2** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 2 | 1 |
| 1 | | | | | | | 2 | | | | | | | | |
| <=2 | | | | | | | | | | | | | 2 | | |
| 2 | | 1 | | | | | | 2 | | | | | | | |
| 4 | | 1 | | | | | | | | | | | | | |
| <=8 | | | | | | 2 | | | | | | | | | |
| 16 | | | 2 | | | | | | | | | | | | |
| 64 | | | | | | | | | | | | 2 | | | |
| >128 | | | | | | | | | | | 2 | | | | |

## Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 14 | 14 | 14 | 14 | 14 | 14 | 14 | 14 | 14 | 14 | 14 | 14 | 14 | 14 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 14 | 0 | 0 | 0 | 14 | 14 | 14 | 0 | 7 |
| <=0.03 | | | | | | | | | | 14 | | | | | |
| <=0.25 | | | | 14 | | | | | | | | | | | 4 |
| <=0.5 | | | | | 2 | | | | 14 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 9 | 3 |
| <=1 | 2 | | | | | | | 14 | | | | | | | |
| 1 | | | | | 12 | | 14 | | | | | | | 5 | |
| 2 | 8 | | | | | | | | | | | | | | |
| 4 | 4 | | | | | | | | | | | | | | |
| <=8 | | | | | | 14 | | | | | | | | | |
| 8 | | | 13 | | | | | | | | | | | | |
| 16 | | | 1 | | | | | | | | | | | | |
| >32 | | | | | | | | | | | | | | | 7 |
| >64 | | | | | | | | | | | | | 14 | | |
| >128 | | | | | | | | | | | 14 | | | | |
| >1024 | | | | | | | | | | | | 14 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Kedougou in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | 2 | |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| 0.5 | | | | | | | | | | | | | | | 2 |
| <=1 | | | | | | | | 2 | | | | | | | |
| <=2 | | | | | | | | | | | | | 2 | | |
| 2 | | 2 | | | | | | | | | | | | | |
| <=4 | | | | | | | | | | | 2 | | | | |
| 4 | | | 2 | | | | | | | | | | | | |
| <=8 | | | | | | 2 | | | | | | | | | |
| 32 | | | | | | | | | | | | 2 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Kentucky in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **4** | **4** | **4** | **4** | **4** | **4** | **4** | **4** | **4** | **4** | **4** | **4** | **4** | **4** |
| **MIC** | **N of resistant isolates** | **4** | **0** | **0** | **0** | **0** | **4** | **0** | **2** | **0** | **4** | **2** | **2** | **0** | **0** |
| <=0.03 | | | | | | | | | | 4 | | | | | |
| <=0.25 | | | | 4 | | | | | | | | | | 1 | 3 |
| <=0.5 | | | | | | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 2 | 1 |
| <=1 | | | | | | | | 4 | | | | | | | |
| 1 | | | | | 4 | | | | 1 | | | | | 1 | |
| <=2 | | | | | | | | | | | | | 2 | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 4 | | | | | | | | | |
| 8 | | | 3 | | | | 3 | | | | | | | | |
| >8 | | | | | | | 1 | | | | | | | | |
| 16 | | | | | | | | | 1 | | | 1 | | | |
| 32 | | | | | | | | | 1 | | | 1 | | | |
| >64 | | 4 | | | | | | | | | | | 2 | | |
| >128 | | | | | | | | | | | 4 | | | | |
| >1024 | | | | | | | | | | | | 2 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Kentucky in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** |
| **MIC** | **N of resistant isolates** | **5** | **1** | **0** | **0** | **0** | **6** | **0** | **3** | **0** | **6** | **4** | **3** | **0** | **1** |
| <=0.03 | | | | | | | | | | 6 | | | | | |
| <=0.25 | | | | 6 | | | | | | | | | | 2 | 4 |
| <=0.5 | | | | | 1 | | | | 2 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 3 | 1 |
| <=1 | | | | | | | | 4 | | | | | | | |
| 1 | | | | | 5 | | | | 1 | | | | | 1 | |
| <=2 | | | | | | | | | | | | | 3 | | |
| 2 | | | | | | | | 2 | | | | | | | |
| 4 | | 1 | 3 | | | | 1 | | | | | | | | |
| <=8 | | | | | | 6 | | | | | | | | | |
| 8 | | | 2 | | | | 5 | | | | | | | | |
| 16 | | | | | | | | | 2 | | | 2 | | | |
| 32 | | | 1 | | | | | | 1 | | | | | | |
| >32 | | | | | | | | | | | | | | | 1 |
| 64 | | | | | | | | | | | | | 1 | | |
| >64 | | 5 | | | | | | | | | | | 2 | | |
| >128 | | | | | | | | | | | 6 | | | | |
| 512 | | | | | | | | | | | | 1 | | | |
| 1024 | | | | | | | | | | | | 1 | | | |
| >1024 | | | | | | | | | | | | 2 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Kentucky in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **1** | **0** | **0** | **0** | **0** | **1** | **0** | **1** | **0** | **1** | **1** | **1** | **0** | **0** |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| 0.5 | | | | | | | | | | | | | | 1 | |
| 1 | | | | | 1 | | | | | | | | | | |
| 2 | | | | | | | | 1 | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| >8 | | | | | | | 1 | | | | | | | | |
| 16 | | | | | | | | | 1 | | | | | | |
| >64 | | 1 | | | | | | | | | | | 1 | | |
| >128 | | | | | | | | | | | 1 | | | | |
| >1024 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Kottbus in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | 2 | 2 |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| <=1 | | 2 | | | | | | 2 | | | | | | | |
| <=2 | | | | | | | | | | | | | 2 | | |
| <=4 | | | | | | | | | | | 2 | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 2 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 2 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Kottbus in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 13 | | | | | |
| 0.03 | | | | | | | 12 | | | | | | | | |
| <=0.25 | | | | 13 | | | | | | | | | | 7 | 13 |
| <=0.5 | | | | | 13 | | | | 13 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 6 | |
| <=1 | | 10 | | | | | | 2 | | | | | | | |
| <=2 | | | | | | | | | | | | | 13 | | |
| 2 | | 3 | | | | | | 11 | | | | | | | |
| <=4 | | | | | | | | | | | 13 | | | | |
| 4 | | | 2 | | | | | | | | | | | | |
| <=8 | | | | | | 13 | | | | | | 1 | | | |
| 8 | | | 11 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |
| 32 | | | | | | | | | | | | 11 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Kottbus in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **7** | **7** | **7** | **7** | **7** | **7** | **7** | **7** | **7** | **7** | **7** | **7** | **7** | **7** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 7 | | | | | |
| 0.03 | | | | | | | 6 | | | | | | | | |
| <=0.25 | | | | 7 | | | | | | | | | | 3 | 7 |
| <=0.5 | | | | | 7 | | | | 7 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 4 | |
| <=1 | | 4 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 7 | | |
| 2 | | 3 | | | | | | 6 | | | | | | | |
| <=4 | | | | | | | | | | | 7 | | | | |
| <=8 | | | | | | 7 | | | | | | 2 | | | |
| 8 | | | 7 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |
| 32 | | | | | | | | | | | | 4 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Kottbus in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | | | | | | | 1 | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Liverpool in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** |
| **MIC** | **N of resistant isolates** | **6** | **0** | **0** | **0** | **2** | **6** | **0** | **0** | **0** | **2** | **0** | **2** | **0** | **2** |
| <=0.03 | | | | | | | | | | 6 | | | | | |
| <=0.25 | | | | 6 | | | | | | | | | | | 3 |
| <=0.5 | | | | | 5 | | | | 6 | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | 6 | 1 |
| <=1 | | | | | | | | 6 | | | | | | | |
| 1 | | | | | 1 | | 5 | | | | | | | | |
| <=2 | | | | | | | | | | | | | 3 | | |
| 4 | | | | | | | | | | | | | 1 | | |
| <=8 | | | | | | 3 | | | | | | | | | |
| 8 | | | 6 | | | | | | | | | | | | |
| 16 | | | | | | 1 | | | | | 4 | 1 | | | |
| 32 | | | | | | | | | | | 2 | 3 | | | |
| >32 | | | | | | | | | | | | | | | 2 |
| 64 | | | | | | | | | | | | 1 | | | |
| >64 | | 6 | | | | | | | | | | | 2 | | |
| 128 | | | | | | | | | | | | 1 | | | |
| >128 | | | | | | 2 | | | | | | | | | |

## Table Antimicrobial susceptibility testing of Salmonella Liverpool in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **1** | **0** | **0** | **0** | **1** | **0** | **0** | **0** | **0** | **0** | **1** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 1 |
| <=1 | | | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| >64 | | 1 | | | | | | | | | | | | | |
| >128 | | | | | | 1 | | | | | | | | | |
| 1024 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Liverpool in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **1** | **0** | **0** | **0** | **0** | **1** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | | | | | | | 1 | | | | | | | |
| 1 | | | | | | | 1 | | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 16 | | | | | | | | | | | 1 | | | | |
| 32 | | | | | | | | | | | | 1 | | | |
| >64 | | 1 | | | | | | | | | | | | | |

## Table Antimicrobial susceptibility testing of Salmonella Livingstone in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **12** | **12** | **12** | **12** | **12** | **12** | **12** | **12** | **12** | **12** | **12** | **12** | **12** | **12** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **12** | **0** | **0** | **0** | **12** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 12 | | | | | |
| <=0.25 | | | | 12 | | | | | | | | | | 11 | 1 |
| <=0.5 | | | | | 12 | | | | 11 | | | | | | |
| 0.5 | | | | | | | 12 | | | | | | | 1 | 11 |
| <=1 | | 3 | | | | | | 10 | | | | | | | |
| 1 | | | | | | | | | 1 | | | | | | |
| <=2 | | | | | | | | | | | | | 11 | | |
| 2 | | 9 | | | | | | 2 | | | | | | | |
| 4 | | | | | | | | | | | | | 1 | | |
| <=8 | | | | | | 12 | | | | | | | | | |
| 8 | | | 12 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | 12 | 11 | | | |
| 64 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Livingstone in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON pnl2

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Cefepime | Cefotaxim | Cefotaxime + Clavulanic acid | Cefoxitin | Ceftazidim | Ceftazidime + Clavulanic acid | Ertapenem | Imipenem | Meropenem | Temocillin |
|---|---|---|---|---|---|---|---|---|---|---|---|
| | **Cefotaxime synergy test** | **Not Available** | **Not Available** | **Positive/Present** | **Not Available** | **Not Available** | **Not Available** | **Not Available** | **Not Available** | **Not Available** | **Not Available** |
| | **Ceftazidime synergy test** | **Not Available** | **Not Available** | **Not Available** | **Not Available** | **Not Available** | **Positive/Present** | **Not Available** | **Not Available** | **Not Available** | **Not Available** |
| | **ECOFF** | **0.125** | **0.5** | **0.5** | **8** | **2** | **2** | **0.06** | **1** | **0.125** | **32** |
| | **Lowest limit** | **0.06** | **0.25** | **0.06** | **0.5** | **0.25** | **0.12** | **0.015** | **0.12** | **0.03** | **0.5** |
| | **Highest limit** | **32** | **64** | **64** | **64** | **128** | **128** | **2** | **16** | **16** | **64** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **1** | **1** | **0** | **0** | **1** | **0** | **0** | **0** | **0** | **0** |
| <=0.015 | | | | | | | | 1 | | | |
| <=0.03 | | | | | | | | | | 1 | |
| <=0.12 | | | | | | | | | 1 | | |
| 0.12 | | | | 1 | | | | | | | |
| 0.5 | | | | | | | 1 | | | | |
| 4 | | | | | 1 | 1 | | | | | |
| 8 | | 1 | | | | | | | | | 1 |
| 32 | | | 1 | | | | | | | | |

## Table Antimicrobial susceptibility testing of Salmonella Livingstone in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** |
| **MIC** | **N of resistant isolates** | **1** | **0** | **1** | **1** | **0** | **4** | **0** | **0** | **0** | **4** | **2** | **1** | **0** | **0** |
| <=0.03 | | | | | | | | | | 5 | | | | | |
| 0.064 | | | | | | | 1 | | | | | | | | |
| 0.12 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 4 | | | | | | | | | | 3 | 5 |
| <=0.5 | | | | | 4 | | | | 5 | | | | | | |
| 0.5 | | | | | | | 3 | | | | | | | 1 | |
| <=1 | | 3 | | | | | | 4 | | | | | | | |
| 1 | | | | | | | | | | | | | | 1 | |
| <=2 | | | | | | | | | | | | | 3 | | |
| 2 | | | | | | | | 1 | | | | | | | |
| 4 | | 1 | | | 1 | | | | | | | | 1 | | |
| >4 | | | | 1 | | | | | | | | | | | |
| <=8 | | | | | | 3 | | | | | | | | | |
| 8 | | | 4 | | | | | | | | 1 | | | | |
| 16 | | | 1 | | | 2 | | | | | | 1 | | | |
| 32 | | | | | | | | | | | 2 | 2 | | | |
| >64 | | 1 | | | | | | | | | | | 1 | | |
| 128 | | | | | | | | | | | 1 | | | | |
| >128 | | | | | | | | | | | 1 | | | | |
| >1024 | | | | | | | | | | | | 2 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Livingstone in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **1** | **0** | **0** | **0** | **1** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | 1 | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Mbandaka in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **1** | **0** | **0** | **0** |
| <=0.015 | | | | | | | 4 | | | | | | | | |
| <=0.03 | | | | | | | | | | 5 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 5 | | | | | | | | | | 5 | 4 |
| <=0.5 | | | | | 5 | | | | 5 | | | | | | |
| 0.5 | | | | | | | | | | | | | | | 1 |
| <=1 | | 4 | | | | | | 5 | | | | | | | |
| <=2 | | | | | | | | | | | | | 5 | | |
| 2 | | 1 | | | | | | | | | | | | | |
| <=4 | | | | | | | | | | | 5 | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 5 | | | | | | | | | |
| 8 | | | 4 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |
| 64 | | | | | | | | | | | | 2 | | | |
| 128 | | | | | | | | | | | | 1 | | | |
| >1024 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Mbandaka in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.015 | | | | | | | 2 | | | | | | | | |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | 2 | 2 |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| <=1 | | 2 | | | | | | 2 | | | | | | | |
| <=2 | | | | | | | | | | | | | 2 | | |
| <=4 | | | | | | | | | | | 2 | | | | |
| <=8 | | | | | | 2 | | | | | | | | | |
| 8 | | | 2 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 2 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Montevideo in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** |
| **MIC** | **N of resistant isolates** | **0** | **1** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **2** | **0** | **0** | **1** |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | 2 | 1 |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| <=1 | | 2 | | | | | | 2 | | | | | | | |
| <=2 | | | | | | | | | | | | | 2 | | |
| <=4 | | | | | | | | | | | 2 | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 16 | | | | | | 1 | | | | | | | | | |
| >32 | | | | | | | | | | | | | | | 1 |
| >64 | | | 1 | | | | | | | | | | | | |
| >1024 | | | | | | | | | | | | 2 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Montevideo in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Newport in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** |
| **MIC** | **N of resistant isolates** | **2** | **0** | **0** | **0** | **0** | **2** | **0** | **0** | **0** | **0** | **0** | **2** | **0** | **0** |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | | 1 |
| 0.25 | | | | | | | 2 | | | | | | | | |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 2 | 1 |
| <=1 | | | | | | | | 2 | | | | | | | |
| 4 | | | 2 | | | | | | | | | | | | |
| <=8 | | | | | | 2 | | | | | | | | | |
| 16 | | | | | | | | | | | 2 | | | | |
| 64 | | | | | | | | | | | | 2 | | | |
| >64 | | 2 | | | | | | | | | | | 2 | | |

## Table Antimicrobial susceptibility testing of Salmonella Orion in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** | **2** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **2** | **0** | **0** | **0** | **2** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | | 2 |
| <=0.5 | | | | | | | | | 2 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 2 | |
| <=1 | | | | | | | | 2 | | | | | | | |
| 1 | | | | | 2 | | 2 | | | | | | | | |
| 2 | | 2 | | | | | | | | | | | | | |
| 4 | | | | | | | | | | | | | 2 | | |
| <=8 | | | | | | 1 | | | | | | 2 | | | |
| 8 | | | 2 | | | | | | | | | | | | |
| 16 | | | | | | 1 | | | | | | | | | |
| 32 | | | | | | | | | | | 1 | | | | |
| 64 | | | | | | | | | | | 1 | | | | |

## Table Antimicrobial susceptibility testing of Salmonella Orion in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **1** | **0** | **0** | **0** | **1** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | | | | | | | 1 | | | | | | | |
| 1 | | | | | 1 | | 1 | | | | | | | | |
| 2 | | 1 | | | | | | | | | | | | | |
| 4 | | | | | | | | | | | | | 1 | | |
| <=8 | | | | | | 1 | | | | | | 1 | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | 1 | | | | |

## Table Antimicrobial susceptibility testing of Salmonella Other serovars in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Senftenberg in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** |
| **MIC** | **N of resistant isolates** | **1** | **0** | **0** | **0** | **0** | **1** | **0** | **0** | **0** | **1** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 3 | | | | | | | | | | 2 | 1 |
| <=0.5 | | | | | 3 | | | | 3 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 2 |
| <=1 | | 2 | | | | | | 3 | | | | | | | |
| 1 | | | | | | | 1 | | | | | | | | |
| <=2 | | | | | | | | | | | | | 3 | | |
| <=4 | | | | | | | | | | | 2 | | | | |
| 4 | | | 3 | | | | | | | | | | | | |
| <=8 | | | | | | 3 | | | | | | | | | |
| 32 | | | | | | | | | | | 1 | 3 | | | |
| >64 | | 1 | | | | | | | | | | | | | |

## Table Antimicrobial susceptibility testing of Salmonella Senftenberg in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **8** | **8** | **8** | **8** | **8** | **8** | **8** | **8** | **8** | **8** | **8** | **8** | **8** | **8** |
| **MIC** | **N of resistant isolates** | **3** | **0** | **0** | **0** | **0** | **3** | **0** | **0** | **0** | **1** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 7 | | | | | |
| 0.03 | | | | | | | 5 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 8 | | | | | | | | | | 7 | 3 |
| <=0.5 | | | | | 7 | | | | 8 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 5 |
| <=1 | | 4 | | | | | | 5 | | | | | | | |
| 1 | | | | | 1 | | 3 | | | | | | | | |
| <=2 | | | | | | | | | | | | | 8 | | |
| 2 | | 1 | | | | | | 3 | | | | | | | |
| <=4 | | | | | | | | | | | 5 | | | | |
| 4 | | | 8 | | | | | | | | | | | | |
| <=8 | | | | | | 8 | | | | | | | | | |
| 16 | | | | | | | | | | | 2 | | | | |
| 32 | | | | | | | | | | | 1 | 7 | | | |
| 64 | | | | | | | | | | | | 1 | | | |
| >64 | | 3 | | | | | | | | | | | | | |

## Table Antimicrobial susceptibility testing of Salmonella Senftenberg in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Taksony in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** | **6** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **2** | **0** | **0** | **0** | **2** | **1** | **0** | **0** | **0** |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 6 | | | | | |
| 0.03 | | | | | | | 3 | | | | | | | | |
| <=0.25 | | | | 6 | | | | | | | | | | 4 | 4 |
| <=0.5 | | | | | 5 | | | | 6 | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | 2 | 2 |
| <=1 | | 4 | | | | | | 6 | | | | | | | |
| 1 | | | | | 1 | | | | | | | | | | |
| <=2 | | | | | | | | | | | | | 4 | | |
| 2 | | | | | | | 1 | | | | | | | | |
| <=4 | | | | | | | | | | | 4 | | | | |
| 4 | | 2 | 3 | | | | | | | | | | 2 | | |
| <=8 | | | | | | 4 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | 2 | | | 2 | | | | | | | | | |
| 32 | | | | | | | | | | | | 5 | | | |
| >128 | | | | | | | | | | | 2 | | | | |
| >1024 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Taksony in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **1** | **0** | **0** | **0** | **1** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 5 | | | | | |
| 0.03 | | | | | | | 4 | | | | | | | | |
| <=0.25 | | | | 5 | | | | | | | | | | 4 | 5 |
| 0.25 | | | | | | | 1 | | | | | | | | |
| <=0.5 | | | | | 5 | | | | 5 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | 5 | | | | | | 3 | | | | | | | |
| <=2 | | | | | | | | | | | | | 5 | | |
| 2 | | | | | | | | 2 | | | | | | | |
| <=4 | | | | | | | | | | | 4 | | | | |
| 4 | | | 4 | | | | | | | | | | | | |
| <=8 | | | | | | 5 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 4 | | | |
| 64 | | | | | | | | | | | | 1 | | | |
| 128 | | | | | | | | | | | 1 | | | | |

## Table Antimicrobial susceptibility testing of Salmonella Taksony in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **1** | **0** | **0** | **0** | **1** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| 0.25 | | | | | | | 1 | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |
| >128 | | | | | | | | | | | 1 | | | | |

## Table Antimicrobial susceptibility testing of Salmonella Tennessee in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** | **5** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.015 | | | | | | | 4 | | | | | | | | |
| <=0.03 | | | | | | | | | | 4 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 5 | | | | | | | | | | 2 | 4 |
| <=0.5 | | | | | 5 | | | | 5 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 3 | 1 |
| <=1 | | 5 | | | | | | 5 | | | | | | | |
| <=2 | | | | | | | | | | | | | 5 | | |
| <=4 | | | | | | | | | | | 5 | | | | |
| <=8 | | | | | | 5 | | | | | | | | | |
| 8 | | | 5 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 2 | | | |
| 64 | | | | | | | | | | | | 3 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Tennessee in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **10** | **10** | **10** | **10** | **10** | **10** | **10** | **10** | **10** | **10** | **10** | **10** | **10** | **10** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.015 | | | | | | | 4 | | | | | | | | |
| <=0.03 | | | | | | | | | | 10 | | | | | |
| 0.03 | | | | | | | 6 | | | | | | | | |
| <=0.25 | | | | 10 | | | | | | | | | | 6 | 3 |
| <=0.5 | | | | | 10 | | | | 10 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 4 | 7 |
| <=1 | | 7 | | | | | | 9 | | | | | | | |
| <=2 | | | | | | | | | | | | | 10 | | |
| 2 | | 3 | | | | | | 1 | | | | | | | |
| <=4 | | | | | | | | | | | 10 | | | | |
| <=8 | | | | | | 10 | | | | | | 1 | | | |
| 8 | | | 10 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 6 | | | |
| 64 | | | | | | | | | | | | 2 | | | |
| 128 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Tennessee in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 1 |
| <=1 | | | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | 1 | | | | | | | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 64 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Tennessee in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 1 |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** | **1** |
| **MIC** | **N of resistant isolates** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | | | | | | | 1 | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

## Table Antimicrobial susceptibility testing of Salmonella Uganda in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.5** | **2** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **256** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** | **3** |
| **MIC** | **N of resistant isolates** | **3** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **0** | **3** | **0** | **0** |
| <=0.03 | | | | | | | | | | 3 | | | | | |
| 0.03 | | | | | | | 3 | | | | | | | | |
| <=0.25 | | | | 3 | | | | | | | | | | | 3 |
| <=0.5 | | | | | 3 | | | | 3 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 3 | |
| <=1 | | | | | | | | 3 | | | | | | | |
| <=4 | | | | | | | | | | | 3 | | | | |
| 4 | | | 2 | | | | | | | | | | | | |
| <=8 | | | | | | 3 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |
| 32 | | | | | | | | | | | | 2 | | | |
| >64 | | 3 | | | | | | | | | | | 3 | | |

## ANTIMICROBIAL RESISTANCE TABLES FOR INDICATOR ESCHERICHIA COLI

### Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method:

Country of Origin: Romania

Sampling Details:

| AM substance | Cefepime | Cefotaxim | Cefotaxime + Clavulanic acid | | Cefoxitin | Ceftazidim | Ceftazidime + Clavulanic acid | | Ertapenem | Imipenem | Meropenem | Temocillin |
|---|---|---|---|---|---|---|---|---|---|---|---|---|
| Cefotaxime synergy test | Not Available | Not Available | Positive/Present | Negative/Absent | Not Available | Not Available | Not Available | | Not Available | Not Available | Not Available | Not Available |
| Ceftazidime synergy test | Not Available | Not Available | Not Available | Not Available | Not Available | Not Available | Positive/Present | Negative/Absent | Not Available | Not Available | Not Available | Not Available |
| ECOFF | 0.125 | 0.25 | 0.25 | 0.25 | 8 | 0.5 | 0.5 | 0.5 | 0.06 | 0.5 | 0.125 | 32 |
| Lowest limit | 0.06 | 0.25 | 0.064 | 0.064 | 0.5 | 0.25 | 0.12 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| Highest limit | 32 | 64 | 64 | 64 | 64 | 128 | 128 | 128 | 2 | 16 | 16 | 128 |
| N of tested isolates | 60 | 60 | 60 | 60 | 60 | 60 | 60 | 60 | 60 | 60 | 60 | 60 |
| MIC N of resistant isolates | 57 | 60 | 38 | 38 | 39 | 57 | 38 | 38 | 8 | 0 | 0 | 0 |
| <=0.015 | | | | | | | | | 10 | | | |
| <=0.03 | | | | | | | | | | | 57 | |
| 0.03 | | | | | | | | | 28 | | | |
| <=0.064 | | | 17 | | | | | | | | | |
| 0.064 | | | | | | | | | 14 | | 3 | |
| <=0.12 | | | | | | | 12 | | | 14 | | |
| 0.12 | 3 | | 5 | | | | | | 6 | | | |
| 0.25 | 25 | | | | | | 9 | 1 | 2 | 39 | | |
| 0.5 | 9 | | | | | 3 | | | | 7 | | |
| 1 | 3 | | | | | 5 | | | | | | |
| 2 | 1 | | | 3 | 1 | 4 | | 2 | | | | |

| | AM substance | Cefepime | Cefotaxim | Cefotaxime + Clavulanic acid | | Cefoxitin | Ceftazidim | Ceftazidime + Clavulanic acid | | Ertapenem | Imipenem | Meropenem | Temocillin |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **Cefotaxime synergy test** | **Not Available** | **Not Available** | **Positive/Present** | **Negative/Absent** | **Not Available** | **Not Available** | **Not Available** | | **Not Available** | **Not Available** | **Not Available** | **Not Available** |
| | **Ceftazidime synergy test** | **Not Available** | **Not Available** | **Not Available** | **Not Available** | **Not Available** | **Not Available** | **Positive/Present** | **Negative/Absent** | **Not Available** | **Not Available** | **Not Available** | **Not Available** |
| | **ECOFF** | **0.125** | **0.25** | **0.25** | **0.25** | **8** | **0.5** | **0.5** | **0.5** | **0.06** | **0.5** | **0.125** | **32** |
| | **Lowest limit** | **0.06** | **0.25** | **0.064** | **0.064** | **0.5** | **0.25** | **0.12** | **0.12** | **0.015** | **0.12** | **0.03** | **0.5** |
| | **Highest limit** | **32** | **64** | **64** | **64** | **64** | **128** | **128** | **128** | **2** | **16** | **16** | **128** |
| | **N of tested isolates** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** |
| **MIC** | **N of resistant isolates** | **57** | **60** | **38** | **38** | **39** | **57** | **38** | **38** | **8** | **0** | **0** | **0** |
| 4 | | 7 | 7 | | 10 | 7 | 6 | | 17 | | | | 17 |
| 8 | | 7 | 23 | | 20 | 13 | 18 | | 12 | | | | 30 |
| 16 | | 4 | 20 | | 4 | 1 | 22 | | 7 | | | | 12 |
| 32 | | 1 | 5 | | 1 | 4 | 2 | | | | | | 1 |
| 64 | | | 3 | | | 17 | | | | | | | |
| >64 | | | 2 | | | 17 | | | | | | | |

Sampling Details:

| AM substance | Cefepime | Cefotaxim | Cefotaxime + Clavulanic acid | | Cefoxitin | Ceftazidim | Ceftazidime + Clavulanic acid | | Ertapenem | Imipenem | Meropenem | Temocillin |
|---|---|---|---|---|---|---|---|---|---|---|---|---|
| **Cefotaxime synergy test** | **Not Available** | **Not Available** | **Positive/Present** | **Negative/Absent** | **Not Available** | **Not Available** | **Not Available** | | **Not Available** | **Not Available** | **Not Available** | **Not Available** |
| **Ceftazidime synergy test** | **Not Available** | **Not Available** | **Not Available** | **Not Available** | **Not Available** | **Not Available** | **Positive/Present** | **Negative/Absent** | **Not Available** | **Not Available** | **Not Available** | **Not Available** |
| **ECOFF** | **0.125** | **0.25** | **0.25** | **0.25** | **8** | **0.5** | **0.5** | **0.5** | **0.06** | **0.5** | **0.125** | **32** |
| **Lowest limit** | **0.06** | **0.25** | **0.064** | **0.064** | **0.5** | **0.25** | **0.12** | **0.12** | **0.015** | **0.12** | **0.03** | **0.5** |
| **Highest limit** | **32** | **64** | **64** | **64** | **64** | **128** | **128** | **128** | **2** | **16** | **16** | **128** |
| **N of tested isolates** | **39** | **39** | **39** | **39** | **39** | **39** | **39** | **39** | **39** | **39** | **39** | **39** |
| **MIC** **N of resistant isolates** | **37** | **39** | **27** | **27** | **30** | **39** | **27** | **27** | **3** | **0** | **0** | **0** |
| <=0.015 | | | | | | | | | 8 | | | |
| <=0.03 | | | | | | | | | | | 37 | |
| 0.03 | | | | | | | | | 14 | | | |
| <=0.064 | | | 10 | | | | | | | | | |
| 0.064 | | | | | | | | | 14 | | 2 | |
| <=0.12 | | | | | | | 5 | | | 7 | | |
| 0.12 | 2 | | 2 | | | | | | 1 | | | |
| 0.25 | 21 | | | | | | 6 | | 2 | 21 | | |
| 0.5 | 6 | | | | | | 1 | | | 11 | | |
| 1 | | | | 2 | | 6 | | | | | | |
| 2 | 2 | 4 | | | | 2 | | | | | | |
| 4 | 4 | 5 | | 10 | 2 | 2 | | 12 | | | | 6 |
| 8 | 4 | 13 | | 13 | 7 | 15 | | 11 | | | | 21 |
| 16 | | 14 | | 2 | 4 | 11 | | 4 | | | | 12 |
| 32 | | 2 | | | 3 | 3 | | | | | | |
| 64 | | 1 | | | 16 | | | | | | | |
| >64 | | | | | 7 | | | | | | | |

## Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.25** | **0.5** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **64** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** |
| **MIC** | **N of resistant isolates** | **60** | **0** | **60** | **55** | **20** | **48** | **2** | **29** | **0** | **47** | **45** | **43** | **0** | **22** |
| <=0.015 | | | | | | | 2 | | | | | | | | |
| <=0.03 | | | | | | | | | | 58 | | | | | |
| 0.03 | | | | | | | 8 | | | | | | | | |
| 0.064 | | | | | | | 2 | | | 1 | | | | | |
| 0.12 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | | | | | | | | | | | 36 | 27 |
| 0.25 | | | | | | | 4 | | | | | | | | |
| <=0.5 | | | | | 5 | | | | 20 | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | 24 | 10 |
| <=1 | | | | | | | | 58 | | | | | | | |
| 1 | | | | | 3 | | 2 | | 11 | | | | | | 1 |
| <=2 | | | 8 | | | | | | | | | | 15 | | |
| 2 | | | | 2 | 6 | | | | | | | | | | |
| <=4 | | | | | | | | | | | 7 | | | | |
| 4 | | | 22 | 6 | 6 | | 3 | 2 | | | | | 2 | | |
| >4 | | | | 52 | | | | | | | | | | | |
| <=8 | | | | | | 39 | | | | | | 11 | | | |
| 8 | | | 29 | | 21 | | 16 | | | | 3 | | | | |
| >8 | | | | | 19 | | 22 | | | | | | | | |
| 16 | | | 1 | | | 1 | | | 4 | | 3 | 2 | | | |
| 32 | | | | | | | | | 9 | | 1 | 2 | | | |

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.25** | **0.5** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **64** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** | **60** |
| **MIC** | **N of resistant isolates** | **60** | **0** | **60** | **55** | **20** | **48** | **2** | **29** | **0** | **47** | **45** | **43** | **0** | **22** |
| >32 | | | | | | | | | 16 | | | | | | 22 |
| 64 | | | | | | 2 | | | | | 1 | | 21 | | |
| >64 | | 60 | | | | | | | | | | | 22 | | |
| 128 | | | | | | 10 | | | | | 2 | | | | |
| >128 | | | | | | 8 | | | | | 43 | | | | |
| 1024 | | | | | | | | | | | | 1 | | | |
| >1024 | | | | | | | | | | | | 44 | | | |

Sampling Details:

| MIC | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.25** | **0.5** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **64** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **39** | **39** | **39** | **39** | **39** | **39** | **39** | **39** | **39** | **39** | **39** | **39** | **39** | **39** |
| | **N of resistant isolates** | **39** | **1** | **39** | **38** | **10** | **35** | **0** | **18** | **0** | **33** | **23** | **29** | **0** | **13** |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 38 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| 0.064 | | | | | | | 1 | | | 1 | | | | | |
| <=0.25 | | | | | | | | | | | | | | 25 | 20 |
| 0.25 | | | | | | | 4 | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 16 | | | | | | |
| 0.5 | | | | | | | 2 | | | | | | | 14 | 6 |
| <=1 | | | | | | | | 38 | | | | | | | |
| 1 | | | | | 5 | | 1 | | 4 | | | | | | |
| <=2 | | | 4 | | | | | | | | | | 9 | | |
| 2 | | | | 3 | 2 | | | 1 | 1 | | | | | | |
| <=4 | | | | | | | | | | | 2 | | | | |
| 4 | | | 7 | 6 | 5 | | 3 | | | | | | 1 | | |
| >4 | | | | 30 | | | | | | | | | | | |
| <=8 | | | | | | 27 | | | | | | 12 | | | |
| 8 | | | 25 | | 15 | | 5 | | | | 1 | | | | |
| >8 | | | | | 11 | | 20 | | | | | | | | |
| 16 | | | 2 | | | 2 | | | 3 | | 3 | 4 | | | |
| 32 | | | 1 | | | 1 | | | 2 | | | | | | |
| >32 | | | | | | | | | 13 | | | | | | 13 |
| 64 | | | | | | | | | | | 2 | | 10 | | |
| >64 | | 39 | | | | | | | | | | | 19 | | |
| 128 | | | | | | 4 | | | | | 1 | | | | |
| >128 | | | | | | 5 | | | | | 30 | | | | |
| >1024 | | | | | | | | | | | | 23 | | | |

## Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON pnl2

Analytical Method:

Country of Origin: Romania

Sampling Details:

| AM substance | Cefepime | Cefotaxim | Cefotaxime + Clavulanic acid | Cefoxitin | Ceftazidim | Ceftazidime + Clavulanic acid | | | Ertapenem | Imipenem | Meropenem | Temocillin |
|---|---|---|---|---|---|---|---|---|---|---|---|---|
| Cefotaxime synergy test | Not Available | Not Available | Positive/Present | Negative/Absent | Not Available | Not Available | | Not Available | | Not Available | Not Available | Not Available | Not Available |
| Ceftazidime synergy test | Not Available | Not Available | Not Available | Not Available | Not Available | Not Available | Not Available | Positive/Present | Negative/Absent | Not Available | Not Available | Not Available | Not Available |
| ECOFF | 0.125 | 0.25 | 0.25 | 0.25 | 8 | 0.5 | 0.5 | 0.5 | 0.5 | 0.06 | 0.5 | 0.125 | 32 |
| Lowest limit | 0.064 | 0.25 | 0.064 | 0.064 | 0.5 | 0.25 | 0.12 | 0.12 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| Highest limit | 32 | 64 | 64 | 64 | 64 | 128 | 128 | 128 | 128 | 2 | 16 | 16 | 64 |
| N of tested isolates | 8 | 8 | 8 | 8 | 8 | 8 | 8 | 8 | 8 | 8 | 8 | 8 | 8 |
| N of resistant isolates | 6 | 8 | 4 | 4 | 4 | 7 | 4 | 4 | 4 | 0 | 0 | 0 | 0 |
| MIC | | | | | | | | | | | | | |
| <=0.015 | | | | | | | | | | 4 | | | |
| <=0.03 | | | | | | | | | | | | 8 | |
| 0.03 | | | | | | | | | | 3 | | | |
| <=0.064 | | | 4 | | | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | |
| <=0.12 | | | | | | | 1 | 2 | | | 6 | | |
| 0.12 | 2 | | | | | | | | | | | | |
| 0.25 | 2 | | | | | | | 1 | | | 2 | | |
| 0.5 | 1 | | | | | 1 | | | | | | | |
| 1 | | 1 | | | | 1 | | | | | | | |
| 2 | 1 | | | 1 | | 1 | | | 1 | | | | |
| 4 | 1 | 1 | | | 2 | 1 | | | | | | | 3 |
| 8 | 1 | 1 | | 2 | 2 | 1 | | | 2 | | | | 5 |
| 16 | | 4 | | 1 | | 3 | | | 1 | | | | |

| | AM substance | Cefepime | Cefotaxim | Cefotaxime + Clavulanic acid | | Cefoxitin | Ceftazidim | Ceftazidime + Clavulanic acid | | | Ertapenem | Imipenem | Meropenem | Temocillin |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | Cefotaxime synergy test | Not Available | Not Available | Positive/Present | Negative/Absent | Not Available | Not Available | | Not Available | | Not Available | Not Available | Not Available | Not Available |
| | Ceftazidime synergy test | Not Available | Not Available | Not Available | Not Available | Not Available | Not Available | Not Available | Positive/Present | Negative/Absent | Not Available | Not Available | Not Available | Not Available |
| | ECOFF | 0.125 | 0.25 | 0.25 | 0.25 | 8 | 0.5 | 0.5 | 0.5 | 0.5 | 0.06 | 0.5 | 0.125 | 32 |
| | Lowest limit | 0.064 | 0.25 | 0.064 | 0.064 | 0.5 | 0.25 | 0.12 | 0.12 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| | Highest limit | 32 | 64 | 64 | 64 | 64 | 128 | 128 | 128 | 128 | 2 | 16 | 16 | 64 |
| | N of tested isolates | 8 | 8 | 8 | 8 | 8 | 8 | 8 | 8 | 8 | 8 | 8 | 8 | 8 |
| MIC | N of resistant isolates | 6 | 8 | 4 | 4 | 4 | 7 | 4 | 4 | 4 | 0 | 0 | 0 | 0 |
| 32 | | | | | | 1 | | | | | | | | |
| 64 | | | 1 | | | 3 | | | | | | | | |

## Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.25** | **0.5** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **64** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **170** | **170** | **170** | **170** | **170** | **170** | **170** | **170** | **170** | **170** | **170** | **170** | **170** | **170** |
| **MIC** | **N of resistant isolates** | **115** | **17** | **8** | **7** | **56** | **156** | **8** | **34** | **0** | **148** | **111** | **98** | **0** | **89** |
| <=0.015 | | | | | | | 12 | | | | | | | | |
| <=0.03 | | | | | | | | | | 169 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| 0.12 | | | | | | | 2 | | | | | | | | |
| <=0.25 | | | | 162 | | | | | | | | | | 138 | 63 |
| 0.25 | | | | | | | 19 | | | | | | | | |
| <=0.5 | | | | | 163 | | | | 77 | | | | | | |
| 0.5 | | | | | | | 9 | | | | | | | 32 | 17 |
| <=1 | | 1 | | | | | | 158 | | | | | | | |
| 1 | | | | 1 | 1 | | 13 | | 54 | | | | | | 1 |
| <=2 | | | 10 | | | | | | | | | | 61 | | |
| 2 | | 20 | | | 1 | | 9 | 4 | 5 | | | | | | |
| <=4 | | | | | | | | | | | 14 | | | | |
| 4 | | 31 | 63 | 1 | 1 | | 22 | 3 | | | | | 10 | | |
| >4 | | | | 6 | | | | | | | | | | | |
| <=8 | | | | | | 109 | | | | | | 38 | | | |
| 8 | | 3 | 71 | | 1 | | 59 | 5 | 2 | | 2 | | 1 | | |
| >8 | | | | | 3 | | 23 | | | | | | | | |
| 16 | | | 9 | | | 5 | | | 8 | | 6 | 16 | 1 | | |
| 32 | | | 10 | | | 9 | | | 13 | | | 5 | 2 | | |

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.25** | **0.5** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **64** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **170** | **170** | **170** | **170** | **170** | **170** | **170** | **170** | **170** | **170** | **170** | **170** | **170** | **170** |
| **MIC** | **N of resistant isolates** | **115** | **17** | **8** | **7** | **56** | **156** | **8** | **34** | **0** | **148** | **111** | **98** | **0** | **89** |
| >32 | | | | | | | | | 11 | | | | | | 89 |
| 64 | | 1 | 4 | | | 8 | | | | | 7 | | 35 | | |
| >64 | | 114 | 3 | | | | | | | | | | 60 | | |
| 128 | | | | | | 21 | | | | | 13 | | | | |
| >128 | | | | | | 18 | | | | | 128 | | | | |
| 1024 | | | | | | | | | | | | 8 | | | |
| >1024 | | | | | | | | | | | | 103 | | | |

## Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method:

Country of Origin: Romania

Sampling Details:

| AM substance | Cefepime | Cefotaxim | Cefotaxime + Clavulanic acid | Cefoxitin | Ceftazidim | Ceftazidime + Clavulanic acid | | | Ertapenem | Imipenem | Meropenem | Temocillin |
|---|---|---|---|---|---|---|---|---|---|---|---|---|
| Cefotaxime synergy test | Not Available | Not Available | Positive/Present | Negative/Absent | Not Available | Not Available | | Not Available | | Not Available | Not Available | Not Available | Not Available |
| Ceftazidime synergy test | Not Available | Not Available | Not Available | Not Available | Not Available | Not Available | Not Available | Positive/Present | Negative/Absent | Not Available | Not Available | Not Available | Not Available |
| ECOFF | 0.125 | 0.25 | 0.25 | 0.25 | 8 | 0.5 | 0.5 | 0.5 | 0.5 | 0.06 | 0.5 | 0.125 | 32 |
| Lowest limit | 0.064 | 0.25 | 0.064 | 0.064 | 0.5 | 0.25 | 0.12 | 0.12 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| Highest limit | 32 | 64 | 64 | 64 | 64 | 128 | 128 | 128 | 128 | 2 | 16 | 16 | 64 |
| N of tested isolates | 574 | 574 | 574 | 574 | 574 | 574 | 574 | 574 | 574 | 574 | 574 | 574 | 574 |
| N of resistant isolates | 512 | 574 | 229 | 229 | 251 | 454 | 228 | 228 | 228 | 14 | 0 | 0 | 0 |
| MIC | | | | | | | | | | | | | |
| <=0.015 | | | | | | | | | | 324 | | | |
| <=0.03 | | | | | | | | | | | | 564 | |
| 0.03 | | | | | | | | | | 163 | | | |
| <=0.064 | 9 | | 320 | | | | | | | | | | |
| 0.064 | | | | | | | | | | 73 | | 10 | |
| <=0.12 | | | | | | | 116 | 172 | | | 283 | | |
| 0.12 | 53 | | 24 | | | | | | | 12 | | | |
| <=0.25 | | | | | | 3 | | | | | | | |
| 0.25 | 136 | | 1 | | | | 4 | 41 | 11 | 1 | 272 | | |
| 0.5 | 88 | | | 2 | | 117 | | 1 | 1 | 1 | 19 | | |
| 1 | 19 | 40 | 2 | 30 | | 76 | | | 1 | | | | |
| 2 | 93 | 45 | | 11 | 18 | 64 | | | 26 | | | | 18 |
| 4 | 103 | 44 | 1 | 66 | 154 | 44 | | 1 | 62 | | | | 207 |
| 8 | 54 | 141 | 1 | 95 | 151 | 95 | | | 94 | | | | 313 |

| | AM substance | Cefepime | Cefotaxim | Cefotaxime + Clavulanic acid | | Cefoxitin | Ceftazidim | | Ceftazidime + Clavulanic acid | | Ertapenem | Imipenem | Meropenem | Temocillin |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **Cefotaxime synergy test** | **Not Available** | **Not Available** | **Positive/Present** | **Negative/Absent** | **Not Available** | **Not Available** | | **Not Available** | | **Not Available** | **Not Available** | **Not Available** | **Not Available** |
| | **Ceftazidime synergy test** | **Not Available** | **Not Available** | **Not Available** | **Not Available** | **Not Available** | **Not Available** | **Not Available** | **Positive/Present** | **Negative/Absent** | **Not Available** | **Not Available** | **Not Available** | **Not Available** |
| | **ECOFF** | **0.125** | **0.25** | **0.25** | **0.25** | **8** | **0.5** | **0.5** | **0.5** | **0.5** | **0.06** | **0.5** | **0.125** | **32** |
| | **Lowest limit** | **0.064** | **0.25** | **0.064** | **0.064** | **0.5** | **0.25** | **0.12** | **0.12** | **0.12** | **0.015** | **0.12** | **0.03** | **0.5** |
| | **Highest limit** | **32** | **64** | **64** | **64** | **64** | **128** | **128** | **128** | **128** | **2** | **16** | **16** | **64** |
| | **N of tested isolates** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** |
| **MIC** | **N of resistant isolates** | **512** | **574** | **229** | **229** | **251** | **454** | **228** | **228** | **228** | **14** | **0** | **0** | **0** |
| 16 | | 17 | 154 | | 20 | 34 | 133 | | | 40 | | | | 34 |
| 32 | | 2 | 83 | | 1 | 65 | 37 | | | 4 | | | | 2 |
| 64 | | | 49 | | | 104 | 5 | | | | | | | |
| >64 | | | 18 | | | 48 | | | | | | | | |

## Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.25** | **0.5** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **64** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** |
| **MIC** | **N of resistant isolates** | **574** | **36** | **574** | **454** | **198** | **531** | **58** | **222** | **0** | **487** | **433** | **392** | **0** | **253** |
| <=0.015 | | | | | | | 27 | | | | | | | | |
| <=0.03 | | | | | | | | | | 565 | | | | | |
| 0.03 | | | | | | | 16 | | | | | | | | |
| 0.064 | | | | | | | | | | 9 | | | | | |
| 0.12 | | | | | | | 3 | | | | | | | | |
| <=0.25 | | | | | | | | | | | | | | 450 | 238 |
| 0.25 | | | | | | | 40 | | | | | | | | |
| <=0.5 | | | | | 120 | | | | 204 | | | | | | |
| 0.5 | | | | | | | 38 | | | | | | | 121 | 77 |
| <=1 | | | | | | | | 507 | | | | | | | |
| 1 | | | | 32 | 71 | | 11 | | 131 | | | | | 3 | 6 |
| <=2 | | | 37 | | | | | | | | | | 172 | | |
| 2 | | | | 54 | 74 | | 16 | 9 | 17 | | | | | | |
| <=4 | | | | | | | | | | | 50 | | | | |
| 4 | | | 230 | 51 | 47 | | 17 | 42 | 1 | | | | 10 | | |
| >4 | | | | 437 | | | | | | | | | | | |
| <=8 | | | | | | 369 | | | | | | 97 | | | |
| 8 | | | 186 | | 78 | | 219 | 16 | 7 | | 23 | | | | |
| >8 | | | | | 184 | | 187 | | | | | | | | |
| 16 | | | 85 | | | 7 | | | 41 | | 14 | 27 | | | |
| 32 | | | 20 | | | 13 | | | 42 | | 7 | 15 | 16 | | 1 |

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.25** | **0.5** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **64** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** | **574** |
| **MIC** | **N of resistant isolates** | **574** | **36** | **574** | **454** | **198** | **531** | **58** | **222** | **0** | **487** | **433** | **392** | **0** | **253** |
| >32 | | | | | | | | | 131 | | | | | | 252 |
| 64 | | 17 | 11 | | | 37 | | | | | 8 | 2 | 173 | | |
| >64 | | 557 | 5 | | | | | | | | | | 203 | | |
| 128 | | | | | | 83 | | | | | 22 | 2 | | | |
| >128 | | | | | | 65 | | | | | 450 | | | | |
| 256 | | | | | | | | | | | | 1 | | | |
| 1024 | | | | | | | | | | | | 18 | | | |
| >1024 | | | | | | | | | | | | 412 | | | |

## Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Turkeys - fattening flocks

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.25** | **0.5** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **64** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **18** | **18** | **18** | **18** | **18** | **18** | **18** | **18** | **18** | **18** | **18** | **18** | **18** | **18** |
| **MIC** | **N of resistant isolates** | **14** | **2** | **0** | **0** | **14** | **12** | **0** | **8** | **0** | **7** | **14** | **12** | **0** | **9** |
| <=0.015 | | | | | | | 5 | | | | | | | | |
| <=0.03 | | | | | | | | | | 18 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 18 | | | | | | | | | | 14 | 7 |
| <=0.5 | | | | | 18 | | | | 4 | | | | | | |
| 0.5 | | | | | | | 2 | | | | | | | 4 | 2 |
| <=1 | | | | | | | | 17 | | | | | | | |
| 1 | | | | | | | 4 | | 4 | | | | | | |
| <=2 | | | 3 | | | | | | | | | | 6 | | |
| 2 | 2 | | | | | | | 1 | 2 | | | | | | |
| <=4 | | | | | | | | | | | 6 | | | | |
| 4 | 1 | 9 | | | | | | | | | | | | | |
| <=8 | | | | | | 4 | | | | | | 3 | | | |
| 8 | 1 | 3 | | | | | 3 | | | | 1 | | | | |
| >8 | | | | | | | 3 | | | | | | | | |
| 16 | | 1 | | | | | | | | | 4 | 1 | | | |
| 32 | | 1 | | | | 1 | | | 2 | | 1 | | 1 | | |
| >32 | | | | | | | | | 6 | | | | | | 9 |
| 64 | | 1 | | | | 4 | | | | | | | 3 | | |
| >64 | 14 | | | | | | | | | | | | 8 | | |
| 128 | | | | | | 4 | | | | | | | | | |

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.25** | **0.5** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **64** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **18** | **18** | **18** | **18** | **18** | **18** | **18** | **18** | **18** | **18** | **18** | **18** | **18** | **18** |
| **MIC** | **N of resistant isolates** | **14** | **2** | **0** | **0** | **14** | **12** | **0** | **8** | **0** | **7** | **14** | **12** | **0** | **9** |
| >128 | | | | | | 5 | | | | | 6 | | | | |
| >1024 | | | | | | | | | | | | 14 | | | |

## Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Turkeys - fattening flocks

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method:

Country of Origin: Romania

Sampling Details:

| AM substance | Cefepime | Cefotaxim | Cefotaxime + Clavulanic acid | | Cefoxitin | Ceftazidim | Ceftazidime + Clavulanic acid | | Ertapenem | Imipenem | Meropenem | Temocillin |
|---|---|---|---|---|---|---|---|---|---|---|---|---|
| **Cefotaxime synergy test** | **Not Available** | **Not Available** | **Positive/Present** | **Negative/Absent** | **Not Available** | **Not Available** | **Not Available** | | **Not Available** | **Not Available** | **Not Available** | **Not Available** |
| **Ceftazidime synergy test** | **Not Available** | **Not Available** | **Not Available** | **Not Available** | **Not Available** | **Not Available** | **Positive/Present** | **Negative/Absent** | **Not Available** | **Not Available** | **Not Available** | **Not Available** |
| **ECOFF** | **0.125** | **0.25** | **0.25** | **0.25** | **8** | **0.5** | **0.5** | **0.5** | **0.06** | **0.5** | **0.125** | **32** |
| **Lowest limit** | **0.064** | **0.25** | **0.064** | **0.064** | **0.5** | **0.25** | **0.12** | **0.12** | **0.015** | **0.12** | **0.03** | **0.5** |
| **Highest limit** | **32** | **64** | **64** | **64** | **64** | **128** | **128** | **128** | **2** | **16** | **16** | **64** |
| **N of tested isolates** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** |
| **MIC** **N of resistant isolates** | **10** | **13** | **6** | **6** | **7** | **13** | **6** | **6** | **2** | **0** | **0** | **0** |
| <=0.015 | | | | | | | | | 5 | | | |
| <=0.03 | | | | | | | | | | | 13 | |
| 0.03 | | | | | | | | | 4 | | | |
| <=0.064 | 1 | | 6 | | | | | | | | | |
| 0.064 | | | | | | | | | 2 | | | |
| <=0.12 | | | | | | | 3 | | | 1 | | |
| 0.12 | 2 | | 1 | | | | | | 2 | | | |
| 0.25 | 2 | | | | | | 3 | | | 11 | | |
| 0.5 | 2 | | | | | | 1 | | | 1 | | |
| 2 | | 1 | | 2 | 1 | | | 1 | | | | |
| 4 | 1 | 3 | | 4 | 2 | 1 | | 3 | | | | 2 |
| 8 | 3 | 3 | | | 3 | 8 | | 2 | | | | 6 |
| 16 | 2 | | | | 2 | 4 | | | | | | 5 |
| 32 | | | | | 2 | | | | | | | |

| | AM substance | Cefepime | Cefotaxim | Cefotaxime + Clavulanic acid | | Cefoxitin | Ceftazidim | Ceftazidime + Clavulanic acid | | Ertapenem | Imipenem | Meropenem | Temocillin |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | Cefotaxime synergy test | Not Available | Not Available | Positive/Present | Negative/Absent | Not Available | Not Available | Not Available | | Not Available | Not Available | Not Available | Not Available |
| | Ceftazidime synergy test | Not Available | Not Available | Not Available | Not Available | Not Available | Not Available | Positive/Present | Negative/Absent | Not Available | Not Available | Not Available | Not Available |
| | ECOFF | 0.125 | 0.25 | 0.25 | 0.25 | 8 | 0.5 | 0.5 | 0.5 | 0.06 | 0.5 | 0.125 | 32 |
| | Lowest limit | 0.064 | 0.25 | 0.064 | 0.064 | 0.5 | 0.25 | 0.12 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| | Highest limit | 32 | 64 | 64 | 64 | 64 | 128 | 128 | 128 | 2 | 16 | 16 | 64 |
| | N of tested isolates | 13 | 13 | 13 | 13 | 13 | 13 | 13 | 13 | 13 | 13 | 13 | 13 |
| MIC | N of resistant isolates | 10 | 13 | 6 | 6 | 7 | 13 | 6 | 6 | 2 | 0 | 0 | 0 |
| 64 | | | 2 | | | 3 | | | | | | | |
| >64 | | | 4 | | | | | | | | | | |

## Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Turkeys - fattening flocks

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method:

Country of Origin: Romania

Sampling Details:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.25** | **0.5** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **64** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** |
| **MIC** | **N of resistant isolates** | **13** | **0** | **13** | **13** | **12** | **13** | **1** | **4** | **0** | **9** | **13** | **12** | **0** | **12** |
| <=0.03 | | | | | | | | | | 13 | | | | | |
| <=0.25 | | | | | | | | | | | | | | 12 | 1 |
| 0.25 | | | | | | | 2 | | | | | | | | |
| <=0.5 | | | | | | | | | 5 | | | | | | |
| 0.5 | | | | | | | 2 | | | | | | | 1 | |
| <=1 | | | | | | | | 12 | | | | | | | |
| 1 | | | | | | | | | 3 | | | | | | |
| <=2 | | | 3 | | | | | | | | | | 1 | | |
| 2 | | | | 1 | | | 1 | | 1 | | | | | | |
| <=4 | | | | | | | | | | | 4 | | | | |
| 4 | | | 7 | 3 | 1 | | | 1 | | | | | | | |
| >4 | | | | 9 | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 2 | | 10 | | | | | | | | | | |
| >8 | | | | | 2 | | 8 | | | | | | | | |
| 16 | | | 1 | | | | | | 1 | | | | | | |
| 32 | | | | | | | | | | | 1 | | 1 | | |
| >32 | | | | | | | | | 3 | | | | | | 12 |
| 64 | | | | | | 4 | | | | | | | 2 | | |
| >64 | | 13 | | | | | | | | | | | 9 | | |
| 128 | | | | | | 4 | | | | | | | | | |

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | **ECOFF** | **8** | **16** | **0.25** | **0.5** | **16** | **0.064** | **2** | **2** | **0.125** | **16** | **64** | **8** | **1** | **2** |
| | **Lowest limit** | **1** | **2** | **0.25** | **0.5** | **8** | **0.015** | **1** | **0.5** | **0.03** | **4** | **8** | **2** | **0.25** | **0.25** |
| | **Highest limit** | **64** | **64** | **4** | **8** | **128** | **8** | **16** | **32** | **16** | **128** | **1024** | **64** | **8** | **32** |
| | **N of tested isolates** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** | **13** |
| **MIC** | **N of resistant isolates** | **13** | **0** | **13** | **13** | **12** | **13** | **1** | **4** | **0** | **9** | **13** | **12** | **0** | **12** |
| >128 | | | | | | 4 | | | | | 8 | | | | |
| >1024 | | | | | | | | | | | | 13 | | | |

## OTHER ANTIMICROBIAL RESISTANCE TABLES

## Specific monitoring of ESBL-/AmpC-/carbapenemase-producing bacteria and specific monitoring of carbapenemase-producing bacteria, in the absence of isolate detected

No data returned for this view. This might be because the applied filter excludes all data.

**Specific monitoring of ESBL-/AmpC-/carbapenemase-producing bacteria and specific monitoring of carbapenemase-producing bacteria, in the absence of isolate detected**

# Latest Transmission set

| Table Name | Last submitted dataset transmission date |
|---|---|
| Antimicrobial Resistance | 24-Jan-2020 |
| Animal Population | 29-Jul-2019 |
| Disease Status | 29-Jul-2019 |
| Food Borne Outbreaks | 12-Sep-2019 |
| Prevalence | 27-Nov-2019 |

## 1. Institutions and Laboratories involved in zoonoses monitoring and reporting

National Sanitary Veterinary and Food Safety Authority – central veterinary authority

County Sanitary Veterinary and Food Safety Directorates – local veterinary authority

Institute for Diagnosis and Animal Health – central animal health diagnostic institute

Institute for Hygiene and Veterinary Public Health - central food and feed diagnostic institute

## 2. Short description of the institutions and laboratories involved in data collection and reporting

The monitoring of zoonoses and zoonotic agents is made according with the Romanian National Surveillance Programme published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority (N.S.V.F.S.A.) yearly updated, which is according to the provisions of Directive 2003/99/EC (transposed into Romanian legislation by order of the N.S.V.F.S.A. no.34 /2006). The samples for zoonoses and zoonotic agents are taken by the official veterinarians from County Sanitary Veterinary and Food Safety Directorates (C.S.V.F.S.D.) and the strains isolated in animals/foodstuffs/feedingstuffs are serotyped by specific National Reference Laboratory (NRLs). The NRLs are organized within the central diagnostic institutes, which are a public institution with legal personality, designated as national reference authority in the field of animal/food safety, under the responsibility of N.S.V.F.S.A.
The central institutes collects from regional laboratories (Sanitary Veterinary and for Food Safety Laboratories – S.V.F.S.L.) and from County Sanitary Veterinary and Food Safety Directorates (C.S.V.F.S.D.) and reports to the N.S.V.F.S.A. all zoonoses and zoonotic agents data in the field of animal health, food and feed safety.

## 3. Animal population

### 1. Sources of information and the date(s) (months, years) the information relates to[(a)]

Based on statistical research on livestock and livestock production in 2018, made by the National Institute of Statistics, at the date of December, 31, 2018, compared to the same date of 2017, the livestock of sheep and goats have increased and livestock of bovine, swine and poultry has declined.

Data source are reports from the National Institute of Statistic, and from our National Data Base.

### 2. Definitions used for different types of animals, herds, flocks and holdings as well as the production types covered

Definitions used for the purposes of monitoring and eradication of zoonoses are in compliance with the definitions determined by the Regulation 178/2002, Regulation 2160/2003 and Directives: 2003/99, 64/432, 90/539.
Holding: any establishment, construction or, in the case of an open air farm, any place in which animals are held, kept or handled. The localization of the holding is based on the address and the coordinates of the geographical entity. A geographical entity is a unit of one building or a complex of buildings included grounds and territories where an animal species is or could be hold.
Flock: a single group or multiple groups of animals which share the same production unit (i.e.

using the same air-space or range area). Where housing systems are not typical, the situation is likely to be assessed on a case by case basis. Multiple groups of animals which have beak-to-beak contact (inside or outside the house) are likely to be treated as a single flock for the same epidemiological reasons.

**3. National changes of the numbers of susceptible population and trends**

According to the National Institute of Statistics in December 2018, compared to the same month of the previous year, the number of slaughtered animals and poultry increased for cattle, and decreased for sheep, goats, poultry and pigs; the carcass weight increased for cattle and decreased for sheep, goats and pigs and for poultry remain constant.

**4. Geographical distribution and size distribution of the herds, flocks and holdings[(b)]**

Animal population at the end of 2018 in Romania includes approximately 1.994.917 bovines, 4.834.380 pigs, 338.026 horses, 294.712.864 poultry, 11.845.182 sheep and 1.660.972 goats. According to Identification and Registration Service on current events recorded at agricultural holdings (incoming and outgoing animals, newly registered animals) there were 419.668 bovine holdings, 280.861 horse holdings, 236.687 small ruminant holdings, and 454.423 porcine holdings in Romania. A minor portion of holdings in Romania are specialised farms rearing one animal species only, e.g. milking cows.

Most Romanian farms are mixed establishments rearing ruminants as well as non-ruminants. Such holdings normally operate extensive rearing systems with a small share of purchased feed.

Animal population differs from species to species and from county to county.

**5. Additional information**

These statistics and numerical values may vary from other national or E.U. official sources of animal population records.

(a): National identification and registration system(s), source of reported statistics (Eurostat, others)
(b): Link to website with density maps if available, tables with number of herds and flocks according to geographical area

## 4. General evaluation*: West Nile disease

### 1. History of the disease and/or infection in the country[(a)]

At present West Nile disease is considered endemic in susceptible animal population from the entire territory of Romania.
During 2006 – 2007 research activities were implemented in the horse population in eastern and south eastern part of the country.

As a result of this research evidence was gathered that a high proportion of the horse population proved to be seropositive for West Nile virus antibodies. Following this find measures were taken in order to implement an active surveillance programme at national level in order to detect the prevalence of the disease in the horse population, by detecting IgG and IgM antibodies.

During 2009 – 2011 sufficient data was gathered in order to demonstrate that West Nile disease is endemic at least in the local horse population. As a consequence active surveillance was reduced to only two counties (Constanța and Brăila) in three localities (Esna, Polizești and Nuntași) where outbreaks were declared to O.I.E. in 2010.

### 2. Evaluation of status, trends and relevance as a source for humans

Information gathered during the active surveillance was shared with the Institute for Public Health, in order to help decision making regarding the control of the disease in humans.

During 2018, two horses with clinical signs, from Ilfov county, were tested for IgM antibodies, with positive results. Following the results, control measures were implemented and an immediate notification was issued to O.I.E. (on 27.08.2018).

### 3. Any recent specific action in the Member State or suggested for the European Union[(b)]

The Ministry of Human Health, following recommendations of experts from ECDC and WHO will elaborate, in collaboration with all administrative partners and stakeholders, an National Strategic Preparedness Plan for West Nile disease focused mainly on disease control in human population.

### 4. Additional information

Write text here please

* **For each zoonotic agent**

(a): Epidemiological evaluation (trends and sources) over time until recent/current situation for the different relevant matrixes (food, feed, animal). If relevant: the official “disease status” to be specified for the whole country and/or specific regions within the country

(b): If applicable

## 5.General evaluation*: Echinococcosis

**1. History of the disease and/or infection in the country[(a)]**

Analysis the situation after 2007 in inspected carcasses in slaughter houses shows on the decreasing of cases.The monitoring program for Echinococcosis in the dogs was introduced in the year 2007. The samples are taken from stray dogs. Were tested 19136 samples for echinococcosis, 77 were positive for Echinococcus spp.In the period 2007-2008 were tested 16784 samples from dogs for echinococcosis, 28 samples were positive for Echinococcus spp.In the year 2009 were tested 2352 samples from dogs for echinococcosis, 49 samples were positive for Echinococcus spp. In the year 2010 were tested 809 samples from dogs for echinococcosis by ELISA coproantigen test and two of them were positive for Echinococcus spp. In 2011 were tested 5262 samples from dogs by ELISA coproantigen. From them 121 samples were positive for Echinococcuss spp.. In 2012 were tested 5119 samples from dogs by ELISA coproantigen, From them 9 samples were positive for Echinococcus spp. In 2013 were tested 3267 samples from dogs by ELISA coproantigen, From them 159 samples were positive for Echinococcus spp. In 2013 it was introduced PCR technique for identification the Echinococcus granulosus species on intermediate hosts. In 2014 a total of 173 samples were examined from dogs, from which 6 were positive for Echinococcus spp. In 2015 were examined a total of 59 samples from dogs from which all were negative. In 2017 were tested 269 samples from cattle and pigs by PCR technique for identification the Echinococcus granulosus species 264 samples were positive for Echinococcus granulosus.

**2. Evaluation of status, trends and relevance as a source for humans**

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**3. Any recent specific action in the Member State or suggested for the European Union[(b)]**

**----------**

**4. Additional information**

Write text here please

* **For each zoonotic agent**

(a): Epidemiological evaluation (trends and sources) over time until recent/current situation for the different relevant matrixes (food, feed, animal). If relevant: the official "disease status" to be specified for the whole country and/or specific regions within the country

(b): If applicable

## 6.General evaluation*: Rabies

**1. History of the disease and/or infection in the country[(a)]**

In 2011 was made the oral vaccination of foxes in 16 counties (Arad, Alba, Bihor, Mures, Maramures, Bistria Nasaud, Brasov, Cluj, Covasna, Caras-Severin, Harghita, Hunedoara, Salaj, Sibiu, Satu Mare,Timis) in West and center of Romania, which is the entire territory bounded by the Carpathian Mountains. The baits distribution included Hungarian, Serbian and part of Ukrainian border. The vaccination campaigns of foxes with baits were made by air distribution (approximately 20 baits/km2) and manual distribution (approximately 25 bites/km2) in inaccessible places and areas, in the aircraft with significant populations of foxes near towns, national roads, areas considered at risk. The manual distribution was done by the managers of the hunting areas with the official vets. Air distribution was provided by a service provider under contract for each campaign. The oral vaccination of foxes was made with the baits containing the strain SAD Bern. In one bait there is one vaccination virus dose (1.8 ml) closed in aluminum-plastic blister. Round, dark brown bait is made of feed mixture attractive for foxes- strongly fish

smell. After vaccination campaigns at 45 days, we started the vaccination evaluation program. Foxes shot were brought to the laboratory by hunting managers according to Article 11 (2) and 12 of HG nr.55/2008. The laboratories worked on flow chart, each fox was controlled by FAT (for rabies diagnosis); then, tests negative was sent to the NRL, the only approved laboratory for examining sera fox rabies antibodies in this direction and the achievement test detection marker "tetracycline" the mandible. In 2012, due to political and legislative changes that took place in Romania, the legal basis for approving the oral vaccination of foxes in the whole territory was not approved until the 1st of June, 2012. Therefore, in Romania the spring vaccination campaign of foxes against rabies was not performed. In August 2012 the legal basis has been approved in order to implement the oral vaccination of foxes in the whole territory. We are currently in conflict with the company of aerial distribution of vaccinal baits. The NSVFSA makes all efforts to implement (perform) the oral vaccination campaign of foxes. The NSVFSA addressed to The Ministry of National Defence, by requesting the support for the carrying out of autumn campaign in 2012, by air distribution of antirabies vaccines, as vaccinal baits for foxes, but from legal and economic reasons, this could not be carried out. From these reasons, in the autumn of 2012, Romania failed to carry out the vaccination of foxes by manual distribution to dens of 80475 vaccinal baits (58.680 national vaccination +21.795 emergency vaccination in counties AG, DB, PH, VN ) in 41 counties. In the autumn of 2012, there has been purchased a number of 80.520 baits, of which 40 baits were sampled for testing for establishing the stability of vaccinal titre and 5 baits being kept as counter samples. Of 40 baits samples, 16 baits were tested for virus titre and stability of virus titre. In 2013, the conflict with the company of aerial distribution of vaccinal baits was resolved and the aerial vaccination was performed on the whole territory of the country of 41 counties. There have been distributed a number of 7774398 of baits in total, in two vaccination campaigned, in spring and in autumn. The spring vaccination of foxes was carried out by air distribution of baits (number of 3.846.098 baits with an approx. 20 baits/km2) and also by manual distribution (number of 57499 baits) around localities and areas difficult to reach by plane (approximately 25 baits/km2).The autumn vaccination of foxes was carried out by air distribution of a number of 3.928.300 baits and also by manual distribution ( 58.715 baits). Concerning the baits testing, a number of 580 baits were tested and a number of 350 baits were kept as counter samples. After vaccination campaigns at 45 days, we started the vaccination evaluation program. The shot foxes were brought to the laboratory by hunting managers according to Article 11 (2) and 12 of HG nr.55/2008.

The laboratories worked on flow chart, each fox was controlled by FAT (for rabies diagnosis); then, the negative tests was sent to the NRL, the only approved laboratory for examining sera fox rabies antibodies in this direction and the achievement test detection marker "tetracycline" the mandible.

If it is possible co-finance for the vaccination in cats and dogs.

In 2014-2017 the whole territory of Romania was vaccinated by baits for foxes. There were two campains per year, in spring and in autumn, by air distribution as wel as by manual distribution. In 2018 no oral rabies vaccination of foxes occuered.

**2. Evaluation of status, trends and relevance as a source for humans**

**-**

**3. Any recent specific action in the Member State or suggested for the European Union[(b)]**

**-**

**4. Additional information**

Write text here pleas

* **For each zoonotic agent**

(a): Epidemiological evaluation (trends and sources) over time until recent/current situation for the different relevant matrixes (food, feed, animal). If relevant: the official "disease status" to be specified for the whole country and/or specific regions within the country

(b): If applicable

## 7. General evaluation*: Q FEVER

### 1. History of the disease and/or infection in the country[(a)]

Q fever is a zoonotic disease caused by Coxiella burnetii, a stable bacteria that resists to heat, drying and many common disinfectants. This resistance enables the bacteria to survive for a long period in the environment. Cattle, sheep, and goats are the main reservoirs but a wide variety of other animals can be contaminated, including domestic pets. Coxiella burnetii does not usually cause clinical disease in these animals, although an increased abortion rate and fertility problems in cattle, sheep and goats are observed. The emergence of these common symptoms over a longer period of time leads finally to the diagnosis of Q fever. Organisms are excreted in milk, urine, and faeces by infected animals. Animals shed the organisms especially during parturition within the amniotic fluids and the placenta. Airborne transmission can occur in premises contaminated by placental material, birth fluids or excreta from infected animals. Airborne inhalation is an important transmission route of infection.

### 2. Evaluation of status, trends and relevance as a source for humans

Livestock farmers, dairy workers, veterinarians, slaughterhouse and meat processing plant workers, and researchers at laboratories or facilities housing susceptible animals are especially concerned and have to be informed about this disease, the possible transmission of infection and preventive measures to be respected.

### 3. Any recent specific action in the Member State or suggested for the European Union[(b)]

**-**

### 4. Additional information

Write text here please

* **For each zoonotic agent**

(a): Epidemiological evaluation (trends and sources) over time until recent/current situation for the different relevant matrixes (food, feed, animal). If relevant: the official "disease status" to be specified for the whole country and/or specific regions within the country

(b): If applicable

## 8.General evaluation*: Salmonella spp., unspecified - general evaluation

**1. History of the disease and/or infection in the country[(a)]**

Salmonella have been recognized as important pathogens, Salmonella Enteritidis and Salmonella Typhimurium have accounted for the majority of cases of human Salmonella for many years and have consistently been the most commonly implicated pathogens in general outbreaks of foodborne disease. Since 2007 in Romania was put in place the National Control Programme of S. Enteritidis, S.Typhimurium, S. Virchow, S.Infantis and S. Hadar in breeding flocks of Gallus gallus. This programme has been approved by the Commission with the Decision 2006/ 876/ EC. In 2008 in Romania the National Programme for Control of S. Enteritidis, S. Typhimurium, S. Virchow, S.Infantis and S. Hadar in breeder flocks of Gallus gallus and National Control Programme for S. Enteritidis and S. Typhimurium in laying hens of Gallus gallus was approved by the Commision with the Decision 782/2007.In 2009 in Romania the National Programme for Control of S. Enteritidis, S. Typhimurium, S. Virchow, S.Infantis and S. Hadar in breeder flocks of Gallus gallus, National Control Programme for S. Enteritidis and S. Typhimurium in laying hens of Gallus gallus and National Control programme for Salmonella Enteritidis and S. Typhimurium was approved by the Commission with the Decision 897/2008. In 2010 the National the National Programme for Control of S. Enteritidis, S. Typhimurium, S. Virchow, S.Infantis and S. Hadar in breeder flocks of Gallus gallus, National Control Programme for S. Enteritidis and S. Typhimurium in laying hens of Gallus gallus, the National Control programme for Salmonella Enteritidis and S. Typhimurium and the National Control Programme for S. Enteritidis and S. Typhimurium in turkeys were approved by the Commission with the Decision 883/2010. For Salmonella in geese, ducks, pigs, cattle, there is not a national control programme in place for these animal species.
In 2017, were identified one positive flocks of Salmonella Enteritidis on laying hens, 2 positive flocks of Salmonella in in breeders (one positive flocks of Salmonella Infantis and one positive flocks of Salmonella Typhymurium) and 2 positive flocks of Salmonella in broilers (one whit Salmonella Enteritidis and one whit Salmonella Typhymurium).
In 2018, were identified 2 positive flocks of Salmonella Enteritidis on laying hens, 3 positive flocks of Salmonella in breeders (one positive flocks of Salmonella Infantis and 2 positive flocks of Salmonella Enteritidis), 3 positive flocks of Salmonella Enteritidis in broilers and 1 positive flocks of Salmonella Typhymurium in fattening turkeys;

Salmonella spp. in animal populations without EU control programs.
In 2017 year, were detected: 20 positive cases in sheep, 16 positive cases in pigs, 1 positive cases in pigeons, 1 positive cases in polar fox, 1 positive cases in goats, 1 positive cases in wild birds.

In 2018 year, were detected 29 positive cases in sheep.
In 2018 year, were detected 18 positive cases in pigs.
In 2018 year, were detected 4 positive cases in turkeys.
In 2018 year, were detected 1 positive cases in pigeons.
In 2018 year, were detected 3 positive cases in quails.
In 2018 year, were detected 6 positive cases in goats.
In 2018 year, were detected 1 positive cases in cattle.

**2. Evaluation of status, trends and relevance as a source for humans**

The Romanian National Surveillance Programme published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority no 35/2016, yearly updated which is according with the provisions of Regulation 2005/2073/EC. In 2013, 436 strains of Salmonella spp. were isolated, from which: 219 meat from broilers and products thereof, 93 meat from pig and products thereof, 64 meat, mixed meat, 42 meat from turkey and products thereof, 10 cheeses, 6 meat

from bovine; 1 meat from sheep and 1 strain egg. In 2014, 207 strains of Salmonella spp. were isolated, in meat from poultry and products thereof, meat from pig and products thereof, meat from other species, meat, mixed meat, cheeses, egg and other food. In 2015, 256 strains of Salmonella spp. were isolated in food, from which: 14 1meat from poultry and products thereof, 72 meat from pig and products thereof, 28 meat, mixed meat,3 meat from other species, 3 cheeses, 5 strains egg and 4 other food.
In 2016, 308 strains of Salmonella spp. were isolated in food, from which: 166 meat from poultry and products thereof, 81 meat from pig and products thereof, 5 meat from bovine and products thereof, 27 meat and mixed meat, 3 meat from other species, 21 strains egg, and 5 other food.
In 2017, 183 of Salmonella spp. were isolated in food, from which: 101 meat from poultry and products thereof, 8meat from turkey and products thereof, 44 meat from pig and products thereof, 3 meat from bovine, 2 meat from sheep and products thereof, 13 meat and mixed meat, 1 strain cheeses, 7 strains eggs, 1 strain egg products, 1 strain bakery products (cakes) and 2 strains in other food category.
In 2018, 352 of Salmonella spp. were isolated in food, from which: 170 meat from poultry and products thereof, 15 meat from turkey and products thereof, 71 meat from pig and products thereof, 1 meat from bovine, 1 products thereof sheep, 47 meat and mixed meat, 1 strain cheeses, 2 strains eggs, 31 strain egg products, 1 strain bakery products (cakes) and 12 strains in other food category.The foodstuffs is considered to be an important source of infection at human cases in Romania**.** Comparison of the Salmonella sero-types found in animals, feeding stuffs, food and human helps to suggest possible sources of infection in the food chain.

**3. Any recent specific action in the Member State or suggested for the European Union[(b)]**

There are no data to provide

**4. Additional information**

Salmonella in feedingstuffs: The feeding stuffs for poultry and other animals must be free from Salmonella. The samples of feeding stuffs are sent for testing by the owners of poultry farms. Veterinary Inspection conducts random, regular inspection in feeding stuffs production plants, in particular of microbiological standards, types of internal controls used by the owners of these plants to guarantee the appropriate quality of final product. In addition, it was foreseen that within the National Plan for the official control of animal feedstuffs in the scope of the supervision of Veterinary Inspection which is approved every year, samples are going to be randomly taken from the feedstuffs production plants, holdings and trading and tested for Salmonella. Operators duties in case of detection of inappropriate microbiological quality of product 1.notifying the District Veterinary Officer on the results of sample testing and the batch of products from which they were taken; 2.secondary processing of contaminated batch, according to an indicated method, under supervision of Veterinary Inspection; 3.increasing the frequency of sampling. In 2013, 27 strains of Salmonella spp. were isolated, from which: 13 feed material of land animal origin, 10 compound feedingstuffs for poultry - laying hens, 6 compound feedingstuffs for pig. In 2014, 22 strains of Salmonella spp. were isolated, from which: 14 feed material of land animal origin, 6 compound feedingstuffs for poultry - laying hens, 2 feed material of cereal grain origin. In 2015, 8 strains of Salmonella spp. were isolated, from which: 4 feed material of land animal origin, 1 compound feedingstuffs for pig and 3 feed material of cereal grain origin. In 2016, 17 strains of Salmonella spp. were isolated in feed , from which: 6 feed material of cereal grain origin, 3 feed material of land animal origin, 5 compound feedingstuffs for poultry and 3 strains in compound feedingstuffs for pig and.
In 2017, 18 strains of Salmonella spp. were isolated in feed, from which: 11 feed material of land animal origin, 3 compound feedingstuffs for poultry, 2 feed material of cereal grain origin and 2 strains pet food - dog snacks. In 2018, 9 strains of Salmonella spp. were isolated in feed, from which: 1 compound feedingstuffs for pig, 7 compound feedingstuffs for poultry and 1 strain in other feed category

* **For each zoonotic agent**

(a):Epidemiological evaluation (trends and sources) over time until recent/current situation for the different relevant matrixes (food, feed, animal). If relevant: the official “disease status” to be specified for the whole country and/or specific regions within the country (b): If applicable

**9. General evaluation*: Salmonellosis, other species**

**1. History of the disease and/or infection in the country[(a)]**

Salmonella spp. in animal populations without EU control programs.

In 2017 year, were detected 20 positive cases in sheep.
In 2017 year, were detected 16 positive cases in pigs.
In 2017 year, were detected 2 positive cases in turkeys.
In 2017 year, were detected 1 positive cases in pigeons.
In 2017 year, were detected 1 positive cases in polar fox.
In 2017 year, were detected 1 positive cases in goats.
In 2017 year, were detected 1 positive cases in wild birds.

In 2018 year, were detected 29 positive cases in sheep.
In 2018 year, were detected 18 positive cases in pigs.
In 2018 year, were detected 4 positive cases in turkeys.
In 2018 year, were detected 1 positive cases in pigeons.
In 2018 year, were detected 3 positive cases in quails.
In 2018 year, were detected 6 positive cases in goats.
In 2018 year, were detected 1 positive cases in cattle.

**2. Evaluation of status, trends and relevance as a source for humans**

**-**

**3. Any recent specific action in the Member State or suggested for the European Union[(b)]**

**-**

**4. Additional information**ite text here please

* **For each zoonotic agent**
(a): Epidemiological evaluation (trends and sources) over time until recent/current situation for the different relevant matrixes (food, feed, animal). If relevant: the official "disease status" to be specified for the whole country and/or specific regions within the country
(b): If applicable

**10. General evaluation*: Salmonella spp, in foodstuffs**

**1. History of the disease and/or infection in the country[(a)]**

**The Institute for Hygiene and Veterinary Public Health do not have any data provided**

**2. Evaluation of status, trends and relevance as a source for humans**

The monitoring of Salmonella is a part of the Romanian National Surveillance Programme published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority no. 35/2016, yearly updated which is according with the provisions of Regulation 2005/2073/EC.

In 2013, 436 strains of Salmonella spp. were isolated in food, from which: 219 meat from broilers and products thereof, 93 meat from pig and products thereof, 64 meat, mixed meat, 42 meat from turkey and products thereof, 10 cheeses, 6 meat from bovine; 1 meat from sheep

and 1 strain egg.In 2014, 207 strains of Salmonella spp. were isolated in food, in meat from poultry and products thereof, meat from pig and products thereof, meat from bovine and products thereof, meat from other species, meat, mixed meat, cheeses, egg and other food. In 2015, 256 strains of Salmonella spp. were isolated in food, from which: 141 meat from poultry and products thereof, 72 meat from pig and products thereof, 1 meat from bovine and products thereof, 27 meat, mixed meat, 3 meat from other species, 3 cheeses, 5 strains egg and 4 other food. In 2016, 308 strains of Salmonella spp. were isolated in food, from which: 166 meat from poultry and products thereof, 81 meat from pig and products thereof, 5 meat from bovine and products thereof, 27 meat and mixed meat, 3 meat from other species, 21 strains egg, and 5 other food. In 2017, 183 of Salmonella spp. were isolated in food, from which: 101 meat from poultry and products thereof, 8 meat from turkey and products thereof, 44 meat from pig and products thereof, 3 meat from bovine, 2 meat from sheep and products thereof, 13 meat and mixed meat, 1 strain cheeses, 7 strains eggs, 1 strain egg products, 1 strain bakery products (cakes) and 2 strains in other food category.

In 2018, 352 of Salmonella spp. were isolated in food, from which: 170 meat from poultry and products thereof, 15 meat from turkey and products thereof, 71 meat from pig and products thereof, 1 meat from bovine, 1 products thereof sheep, 47 meat and mixed meat, 1 strain cheeses, 2 strains eggs, 31 strain egg products, 1 strain bakery products (cakes) and 12 strains in other food category.The foodstuffs is considered to be an important source of infection at human cases in Romania.

**3. Any recent specific action in the Member State or suggested for the European Union[(b)]**

The Institute for Hygiene and Veterinary Public Health do not have any data provided

**4. Additional information**

The Romanian National Surveillance Program approved by Order of the President of the National Sanitary Veterinary and Food Safety Authority (N.S.V.F.S.A.) also includes classification of food risk:

1. Group A: milk powder, UHT milk or otherwise sterilized, smoked or matured cheeses for more than 6 months, canned food, honey, dried fish;

2. Group B: Dried and / or matured raw meat products, semi-cooked meat products, heat-treated meat products, cream, pasteurized milk, butter, matured cheese for more than 60 days, beaten milk, yoghurt, milk products fermented, pasteurized or sterilized egg products, meat pasta;

3. Group C: Fresh meat from cattle, horses, pigs, goats, sheep, poultry and lagomorphs, minced meat and prepared meat, fresh or matured cheese less than 60 days, fresh fish, fish fillets, eggs;

4. Group D: Crude milk, unpasteurized milk products, prepared fish products, eggs, smoked fish.

* **For each zoonotic agent**

(a): Epidemiological evaluation (trends and sources) over time until recent/current situation for the different relevant matrixes (food,

feed, animal). If relevant: the official "disease status" to be specified for the whole country and/or specific regions within the country

**(b): If applicable**

## 11. General evaluation*: Listeriosis

### 1. History of the disease and/or infection in the country[(a)] -

Clinical listeriosis is mainly a ruminant disease, affecting sheep , goats and cattle. The types of specimens taken are milk, abortion material, uterus excretions and other clinical specimens e.g. lesions from brain, liver, spleen.

Investigations are initiated by the owners of the animals. Testing is performed on owner request and on clinical suspicion.
In 2016 year, were detected 4 positive cases in cattle
In 2016 year, were detected 10 positive cases in sheep
In 2017 year, were detected 3 positive cases in cattle
In 2017 year, were detected 3 positive cases in goats
In 2017 year, were detected 11 positive cases in sheep
In 2018 year, were detected 2 positive cases in cattle.
In 2018 year, were detected 2 positive cases in goats
In 2018 year, were detected 8 positive cases in sheep
In 2018 year, were detected 2 positive cases in pigs

### 2. Evaluation of status, trends and relevance as a source for humans

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### 3. Any recent specific action in the Member State or suggested for the European Union[(b)]

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### 4. Additional information

Write text here please

* **For each zoonotic agent**
(a): Epidemiological evaluation (trends and sources) over time until recent/current situation for the different relevant matrixes (food, feed, animal). If relevant: the official "disease status" to be specified for the whole country and/or specific regions within the country
(b): If applicable

## 12. General evaluation*: Trichinella spp. In animal sample (organ/tissue) - food sample

**1. History of the disease and/or infection in the country[(a)]**

Romania does not have any regions or holdings official free of trichinelosis. Trichinella spp. is detected in pigs belonging to the small holdings (individual backyards), wild boars and bears.

**2. Evaluation of status, trends and relevance as a source for humans**

The Romanian Surveillance Program is a national program, published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority (N.S.V.F.S.A.), yearly updated and the susceptibility testing of Trichinella spp. to receptive speciesis is a part of this program, according with the provisions of Regulation 2005/2075/EC, repealed by Regulation 2015/1375/EC in order to control the Trichinelosis.

In 2012, were detected a total number of 287 positive cases of Trichinella spp from which: 171 positive cases in fattening pigs from backyards (not raised under controlled housing conditions); 107 positive cases in wild boars, 9 positive cases in bears. In 2013, were detected a total number of 361 positive cases of Trichinella spp from which: 193 positive cases in fattening pigs from backyards (not raised under controlled housing conditions), 148 positive cases in wild boars, 20 positive cases in bears. In 2014, were detected a total number of 255 positive cases of Trichinella spp. from which: 141 positive cases in fattening pigs from backyards (not raised under controlled housing conditions); 88 positive cases in wild boars-wild and 26 positive cases in bears. In 2015, were detected a total number of 210 positive cases of Trichinella spp. from which: 87 positive cases in fattening pigs from backyards (not raised under controlled housing conditions); 94 positive cases in wild boars-wild and 29 positive cases in bears. In 2016, were detected a total number of 256 positive cases of Trichinella spp from which: 120 positive cases in fattening pigs from backyards (meat from pig not raised under controlled housing conditions), 31 positive cases fattening pigs from farms (meat from pig not raised under recognised controlled housing conditions), 89 positive cases in wild boars-wild and 16 positive cases in bears. In 2016, it can be observed a sligtly increase trend of positive cases in Romania compared with 2015. In 2017, were detected a total number of 250 positive cases of Trichinella spp from which:120 positive cases in fattening pigs from backyards (not raised under controlled housing conditions), 128 positive cases in wild boars-wild and 2 positive cases in bears. In the last four years there has been a stagnation in positive cases in Romania, but a different distribution of species from year to year.

In 2018, were detected a total number of 304 positive cases of Trichinella spp from which: 134 positive cases in fattening pigs from backyards (not raised under controlled housing conditions), 166 positive cases in wild boars-wild and 4 positive cases in bears.

**3. Any recent specific action in the Member State or suggested for the European Union[(b)]**

The Institute for Hygiene and Veterinary Public Health do not have any data provided

**4. Additional information**

All positive samples (larvae detected) were sent to identify the species of Trichinella to National

Reference Laboratory for Trichinella (N.R.L.) which is organized in Institute of Hygiene and Veterinary Public Health (I.H.V.P.H.). Some of them are identified by the N.R.L. for Trichinella, which is in Institute of Hygiene and Veterinary Public Health and other of the larvae were sent to EURL Parasites Roma. Comparison of the Trichinella species found in animals, food and human helps to suggest possible sources of infection in the food chain.

* **For each zoonotic agent**

(a): Epidemiological evaluation (trends and sources) over time until recent/current situation for the different relevant matrixes (food, feed, animal). If relevant: the official "disease status" to be specified for the whole country and/or specific regions within the country

(b): If applicable

**13.Description of Monitoring/Surveillance/Control programmes system*: blood serum, West Nile virus**

**12. Monitoring/Surveillance/Control programmes system[(a)]**

For 2018, the passive surveillance specifies that it is compulsory for owners or their representatives to notify to the private vets all cases of receptive animals (horses, birds) with clinical signs or found dead. Another aspect of the passive surveillance is the monitoring of relevant documents that accompany the animal transports.
The active surveillance system involved sampling from horses in three villages from two counties (Constanța and Brăila) where IgM conversions were found in 2010, and, subsequently official notification was sent to O.I.E. The surveillance system was the same as in 2016.

The sampling took place in June, August and October. In case seroconversions were found in June or August, no sampling would take place in the following sampling months. The sample size was calculated based on the entire horse population in each locality and it allowed the detection of the infection at a prevalence of 5% with a confidence of 95%. The sampling matrix was whole blood, and the testing matrix was blood serum. The test used was IgM ELISA using a commercial kit.

**2. Measures in place[(b)]**

In case of outbreaks, measures are taken in accordance with NSVFSA
- movement restrictions
- treatment of infected animals

**3. Notification system in place to the national competent authority[(c)]**

Yes

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

In 2018 evidence of West Nile virus circulation was found in animals (horses with and without clinical signs).

**5. Additional information**

* **For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.
(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.
(c): Mandatory: Yes/No.
(d): Minimum five years.
(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

## 14. Description of Monitoring/Surveillance/Control programmes system*: organ/ tissues, Echinococcosis

**1. Monitoring/Surveillance/Control programmes system[(a)]**

Testing for detection of Echinococcus is a part of post-mortem inspection of slaughtered animals. It is a visual inspection of the internal organs of the slaughtered animals accompanied by cuts of liver if is necessary. The Echinococcus is not routinely distinguished by species.
Analysis the situation after 2007 in inspected carcasses in slaughter houses shows on the decreasing of cases. The monitoring program for Echinococcosis in the dogs was introduced in the year 2007.The samples are taken from stray dogs. Were tested 19136 samples for echinococcosis, 77 were positive for Echinococcus spp. In the period 2007-2008 were tested 16784 samples from dogs for echinococcosis, 28 samples were positive for Echinococcus spp. In the year 2009 were tested 2352 samples from dogs for echinococcosis, 49 samples were positive for Echinococcus spp. In the year 2010 were tested 809 samples from dogs for echinococcosis by ELISA coproantigen test and two of them were positive for Echinococcus spp. In 2011 were tested 5262 samples from dogs by ELISA coproantigen. From them 121 samples were positive for Echinococcuss spp. In 2012 were tested 5119 samples from dogs by ELISA coproantigen, From them 9 samples were positive for Echinococcus spp. In 2013 were tested 3267 samples from dogs by ELISA coproantigen. From them 159 samples were positive for Echinococcus spp.

In 2014 a total of 173 samples were examined from dogs, from which 6 were positive for

Echinococcus spp. In 2015 were examined a total of 59 samples from dogs from which all were negative .In 2013 it was introduced PCR technique for identification the Echinococcus granulosus species on intermediate hosts.

In 2017 were tested 269 samples from cattle and pigs by PCR technique for identification the Echinococcus granulosus species. 264 samples were positive for Echinococcus granulosus.

In 2018 were tested 25 samples from cattle, sheep and pigs by PCR technique for identification the Echinococcus granulosus species .

25 samples were positive for Echinococcus granulosus"

**2. Measures in place[(b)]**

**3. Notification system in place to the national competent authority[(c)]**

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

**5. Additional information**

* **For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.
(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.
(c): Mandatory: Yes/No.
(d): Minimum five years.
(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

## 15.Description of Monitoring/Surveillance/Control programmes system*: Brain, Rabies

**1. Monitoring/Surveillance/Control programmes system[(a)]**

As a member state of the European Union, Romania had annual programes for the surveillance and control of rabies approved, in conformity with the provisions of the European Commission decisions no. 2006/876/CE, 2007/782/CE, 2008/897/CE and 2009/883/CE. Nevertheless, the programes for the anti rabic vaccination of wild foxes could not be implemented, but partially, during the period between 2007-2009, by manual administration of vaccine baits, on restricted areas. One of the causes for not applying the programe represented the impossibility of acquiring the vaccine baits due to legal obstructions found in the process of justice.

Confirmation of rabies diagnosis is established only by laboratory tests on samples taken (brain) from dogs that died or were killed due to clinical signs of disease (nervous signs)Samples for laboratory tests if suspicion of rabies - the entire bodies of the dog- are packaged properly so as to avoid any leakage of fluids.Transport is carried out in refrigerated containers, within 24 hours in winter time and 12 hours in summer time, labeled "biological samples with a high risk of contamination - WARNING RABIES". If the samples are not sent to the laboratory in time, they are frozen.

If the dog becomes ill with symptoms of rabies or dies from a rabies-like illness during the observation period, the dog should be tested for rabies.

Organs/tissues: brain samples (bulb, Ammon horn, cerebellum, cortex, brain stem)

The entire bodies of small animals or heads of large animals - are packaged properly so as to avoid any leakage of fluids. Harvesting and handling must comply with strict work protection measures and biosecurity; must wear personal protective equipment plus disposable mask, goggles, surgical gloves; are mandatory disinfection of instruments and working table used for sampling, in accordance with veterinary rules in force, and washing and disinfecting hands of the operator. Accompanying the evidence clearly indicated the origin of the animal and it owner, owner address, phone number, changes in behavior or physiological status of that animal, if has bitten or scratched other people, and identification and their residence. Transport measures are required to destroyed the bodies, destruction of consumables used in handling samples and destruction of laboratory animals (white mice) used for confirmation or denial of rabies diagnosis.

A case of dog rabies is defined as an illness characterized by acute encephalomyelitis that almost always progresses to coma or death and is laboratory confirmed

Fluorescent Antibody Test (FAT) on smears from hippocampus or medulla oblongata

All dogs over 3 months are vaccinated once a year with a rabies vaccine registered and marketed in Romania. Rabies immunization is done by organizing mass vaccination campaigns, annual autumn-winter period, followed by completing vaccination. Each vaccinated carnivorous receives a completed and signed by the empowered veterinary practitioner health book which certifies the carrying out of the vaccination against rabies, details about the vaccinated animal, owner, location, veterinarian and the vaccine used. Each health book has one series and one number.

The administration of the counties should build shelters for stray dogs, according to national legislation

The Romanian Control Programmer was a national programme for domestic and wild animals, published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority no 29/2008,for the approval of the sanitary veterinary Norm regarding the general measures of prevention and control of rabies in domestic and wild animals. The Surveillance, control, and monitoring of domestic animals and wild animals for rabies makes the objective The programme for the actions of surveillance, prevention and control of animal diseases, of those transmissible from animals to man, for protection of animals and environment which is carried out yearly by the National Sanitary Veterinary and for Food Safety Authority; this programme is suplimented, everytime it is necessary, with epidemiological and risk analysis.

Rabies Vaccination Program for stray dogs and stray cats to be Cofinancing by the UE

After rabies confirmation, the county SVFSD acts as follows: a) perform the epidemiological enquire ; b) establishes the protection and the surveillance zones ; c) issues the control plan with deadlines and responsibilities; The control measures in the protection zone include: - drawing up the epidemiological maps; - killing of carnivores which were bitted or scratched by sick animals, if they were not vaccinated against rabies, or if they have less than 21 days since first vaccination, - isolation by the rest of the animals of the vaccinated carnivores which have been bitted or scratched by the sick animal;- placement under observation of all animals from that holding for 14 days, beginning with the contact moment ; - killing of all animals from that holding, in case when they manifest clinical signs in this period of time; animals which did not manifest clinical signs of rabies, are released from observation; - interdiction of animal movement for animal which were under observation for a period of, at least 3 month. The control measures in the surveillance zones include: - a census for all dogs and cats; - vaccination of dogs and cats with inactivated vaccine; - surveillance and movement control of dogs and cats.

Rabies is a notifiable disease from local to central level, in accordance with the NSVFSA President Order no.79/2008 for the approval of the sanitary veterinary Norm on notifying animal diseases, represents the official transposition of the Council Directive 1982/894/CE regarding the notification of animal diseases. The obligativity of disease notification comes to the free practice empowered practitioners which notify the official veterinarian about the rabies suspicions in the field. Rabies suspicion is notified from the field to SVFSD, and samples are sent to the county sanitary veterinary laboratory accredited and authorized for diagnosis. The official vet responsible with animal health from CSVFSD, notifies the suspicion by a rapid communication mean to the director of Animal Health and Welfare Directorate from NSVFSA and also by using a notification report form, to NSVFSA all suspected cases of rabies. Following to laboratory confirmation of rabies, the county SVFSD and of the Bucharest Municipality, will notify, using a notification report form, to NSVFSA all confirmed cases of rabies. If rabies is confirmed in a domestic animal, the owner is also notified and a complete file issued in view of applying the control measures, if necessary. The situation concerning rabies cases is notified twice/ year to OIE, and quarterly to the European Institute for Rabies Control.

In 2010 year there were detected 46 positive cases in dogs. The vaccination against Rabies of foxes will decrease the number of cases in domestic animals, because foxes are natural virus reservoir. In 2011 were detected 40 positive cases in dogs. In 2012 were detected by FAT 49 positive cases in dogs. In 2013 were detected 38 positive cases in dogs.

The people who have been in contact with positive cases are send to hospitals for examination

and medical treatment.

There is no actual monitoring of bats-wild.

Organs/tissues: brain samples

In 2009 year there were detected by the FAT 1 positive cases in bat-wild. The sample was not submitted to the National Reference Laboratory for Rabies for characterization by geno-typing. In the years 2010, 2011 and 2012 there were no detected cases in bats-wild. In 2013, there were not positive cases in bats.

In 2015 year were detected 28 rabies cases, diagnosed by FAT .

All positive samples were sequenced in order to distinguish between wild strain and vaccine strain (27 wild strain and one vaccine strain - bovine)

In 2016 year were detected 16 rabies cases, diagnosed by FAT (4 foxes, 9 cattle, 2 cats, 1 dog) . All positive samples were sequenced in order to distinguish between wild strain and vaccine strain. All of the positive samples were caused by infection with wild strain rabies.

In 2017 year were tested 9291 animals and found only 2 positive cases (1 dog and 1 cattle). Both positive samples were caused by infection with wild strain rabies

In 2018 there were 3 FAT positive cases (one dog, one fox and one bovine). All these samples were sequenced in order to distinguish between wild strain and vaccine strain. All of the positive samples were caused by infection with wild strain rabies virus.

**2. Measures in place[(b)]**

**3. Notification system in place to the national competent authority[(c)]**

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

**5. Additional information**

* **For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.
(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.
(c): Mandatory: Yes/No.
(d): Minimum five years.
(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

## 16.Description of Monitoring/Surveillance/Control programmes system*: Q FEVER

**1. Monitoring/Surveillance/Control programmes system[(a)]**

Scope: disease surveillance

Surveillance in all types of holdings (cattle, sheep and goat)

TECHNICAL PROVISIONS

Passive surveillance
Passive surveillance for all cases of abortion, stillbirth and other reproductive symptoms with unspecified diagnosis:

1. Necropsy examination of fetuses, histopathological examination (HE/HEA/HEV, ZNM, Pappenheim, immunohistochemistry) on lymph nodes, liver, lung, kidney, myocardium and placenta samples.
2. Sampling of blood from animals (cattle, sheep, goat) with abortions, 14 – 21 days after abortion, by iELISA and CFR.
3. Data monitoring on suspicions and confirmed cases.
4. Quarterly report on suspicions, confirmations, sent to NSVFSA by each county SVFSD.

The procedure in case of confirmation is as follows:

For cattle:

1. Sampling for PCR testing, as follows:

i) Blood samples from minimum 6 cattle taking into account the ratio of 3 multiparous and 3 primiparous from the herd were abortions occurred in the previous minimum 15 days and maximum 4 months; the test used is ELISA, preferably using an antigen obtained from Coxiella isolates on ruminants.

ii) A total of 6 blood samples from cattle with reproductive symptoms such as retained placenta, metritis expressed in the last 4 months; the test used is ELISA, preferably using an antigen obtained from Coxiella isolates on ruminants;

2. A total of 6 blood samples from animals in the same herd, that do not express reproductive symptoms; the test used is ELISA, preferably using an antigen obtained from Coxiella isolates on ruminants;

For small ruminants:

1. Vaginal or placenta swabs from 2 to 6 goat or sheep that aborted in the last 8 days. Alternatively, samples from the abortion can be used (placenta, stomach content, spleen, lung, liver). The test used is PCR in order to make a differential diagnosis. 2 PCR tests will be performed on individual samples. Pooling is allowed in case more than 2 animals are tested.

2. In case only one sample for PCR testing is available, the procedure is as follows:
i) In sheep and goat herds were abortions occurred, blood sampling will be performed starting at 15 days after the abortions but no later than 3 weeks after the abortion. The minimum number of animals sampled is 10/herd, preferably those that aborted. Blood samples will be tested by ELISA, preferably using an antigen obtained from Coxiella isolates on ruminants.

ii) In sheep and goat herds were stillbirths occurred, blood sampling will be performed starting at 15 days after the birth but no later than 3 weeks after the birth. The minimum number of animals sampled is 10/herd, preferably those that had stillbirths. Blood samples will be tested by ELISA, preferably using an antigen obtained from Coxiella isolates on ruminants.

IMPLEMENTATION PROVISIONS

Passive surveillance
Passive surveillance is performed by the owners and workers in daily contact with animals, private veterinarians. They are required to report any case of disease.
The official veterinarians take samples for confirmation diagnosis and deliver them to county SVFSL or NRL in IDSA.
The suspicion is confirmed by evaluating the results of the ELISA and PCR tests.
The disease notification is made in accordance with NSVFSA Order no. 79/2008, with subsequent modifications.

**2. Measures in place[(b)]**

The following measures could be used in the prevention and control of Q fever:
Public education and information on sources of infection.
Advice to persons that present a high risk for infection, especially with preexisting cardiac valvular disease or individuals with vascular grafts and pregnant women .
Access restrictions to barns and laboratories used in housing potentially infected animals.
Quarantine of aborted animals.
Appropriately disposal of placenta, birth products, fetal membranes, and aborted fetuses.
Using of only pasteurized milk and milk products.
Infected holdings and facilities should be located away from populated areas.
Measures should be implemented to prevent airflow to other occupied areas.

**3. Notification system in place to the national competent authority[(c)]**

Yes

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

**5. Additional information**

* **For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.
(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.
(c): Mandatory: Yes/No.
(d): Minimum five years.
(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

## 17. Description of Monitoring/Surveillance/Control programmes system*: Salmonella in animal - Gallus gallus (fowl) - Farm - animal sample - Control and eradication programmes - Official and industry sampling - Census

**1. Monitoring/Surveillance/Control programmes system[(a)]**

Sampling strategy

**Broiler flocks**

The main objective of Romania National Control programme for the reduction of Salmonella Enteritidis and Salmonella Typhimurium and in broilers flocks of Gallus gallus is a reduction of the maximum percentage of positive flocks to 1 % or less. In broiler flocks all isolation of Salmonella must be reported to the Competent authority. In Romania holdings of broiler flocks where S. Enteritidis and S. Typhimurium have been isolated are given advice on Salmonella control and a visit to carry out an epidemiological enquiry as appropriate. The National Control Programme for Salmonella in broiler flocks of Gallus gallus was put in place in 01 January 2009. Starting with 01 January 2009 the National Control Programme for Salmonella in broilers was held in all holdings of broiler flocks consisting of at least 500 poultry of Gallus gallus. Broilers holdings which have between 500 and 5,000 of birds were not the subject of official testing, but they perform tests on the initiative of operators (self-control) within 3 weeks prior to depopulation and sending the birds abattoir.

Frequency of the sampling

Broiler flocks: Before slaughter at farm

Within 3 weeks prior to moving to the slaughter/depopulation

Type of specimen taken

Broiler flocks: Before slaughter at farm

Boot swabs

Methods of sampling (description of sampling techniques)

Broiler flocks: Before slaughter at farm

Broiler flocks: Before slaughter at farm Operators were required to implement the sampling programme in the Annex to EC Regulation 200/2012 (self-control sampling).Two pairs of boot sock/swabs were taken by the operator within the period of three weeks before the birds are due for slaughter. The samples were taken in sufficient time for the laboratory results to be known before the birds are transported to the slaughter house. It is important to know the Salmonella status of the flock before the first birds are slaughtered. Samples were submitted to a laboratory authorized by the Competent Authority and which applies quality assurance systems that conform to the requirements of the current EN/ISO standard. Official control: Each year at least 10% of holdings with more than 5,000 birds were selected and at least one flock on the holding were sampled by Animal Health, or other authorized agent, acting on behalf of the Competent Authority, who took an official sample. In addition, attention was given to flocks where there have been previously positive Salmonella findings in the samples taken by the operators. Particular attention was given to holdings where S. Enteritidis or S. Typhimurium has been isolated from samples. When an official sample was taken it may replace the sample required to be taken by the operator. In accordance with Regulation (EC) No. 200/2012 Annex point 1 (c) the operator of a broiler holding may make an application to the Competent Authority for a derogation not to sample all flocks on the holding. The Competent Authority will assess the application for derogation against the criteria listed in the Annex. The Competent Authority may approve the derogation if satisfied. Sampling protocol. For each flock at least two pairs of boot/sock swabs shall be taken. All boot/sock swabs must be pooled into one sample. For free range broiler flocks, samples shall only be collected in the area inside the house. Before using the boot/sock swabs, their surface shall be moistened with deionised water, or sterile water or any other diluents approved by the national reference laboratory referred to in Article 11 of Regulation (EC) No 2160/2003. The use of farm water containing antimicrobials or additional disinfectants shall be prohibited. The recommended way to moisten boot swabs shall be to pour the liquid inside before putting them on. It shall be ensured that all sections in a house are represented in the sampling in a proportionate way and that at least 100 steps are taken with each pair of boot swabs. Each pair should cover about 50 % of the area of the house. On completion of sampling the boot/sock swabs shall be carefully removed so as not to dislodge adherent material. Boot swabs may be inverted to retain material. They shall be placed in a

bag or pot and labelled to identify the flock sampled, and the date the samples were taken. According to the provisions of the Order of President on National Sanitary Veterinary and Food Safety Authority no.34/2006, transposing into Romanian legislation the Directive 2003/99/EC, all the Salmonella spp. strains isolated in foodstuffs derived from products of animal origin were compulsory tested for the antimicrobial resistance.
Diagnostic/analytical methods used
Broiler flocks: Before slaughter at farm
Bacteriological method: ISO 6579:2002/Amd1:2007

The programme regarding reduction of prevalence of Salmonella serotypes in broiler flocks populations is approved by the EC (https://ec.europa.eu/food/funding/animal-health/national-veterinary programmes_en)

**2. Measures in place[(b)]**

**Vaccination policy**
Broiler flocks
Live Salmonella vaccines are not used in the framework of national control programme where the manufacturer does not provide an appropriate method to distinguish bacteriological wild-type strains of salmonella from vaccine strains. Although vaccines against Salmonella are not currently used in broilers.
Other preventive measures than vaccination in place
Broiler flocks
According to the Romanian program of surveillance, prevention and animal disease control, of the diseases transmissible from animals to humans, animal protection and environment protection and program for surveillance and control in food safety field approved every year by N.S.V.F.S.A. President Order, feeding stuffs intended for poultry nutrition are checked in view to avoid the contamination with Salmonella
spp. Also, in conformity with the same legislation the feed stuffs are checked in view to detect the use of antibiotics. Residues examination is performed according to the Romanian annual plan for examination for residues in live animals and animal origin products. For broiler,hens, turkeys, other poultry a sample consists on one or more animals depending on the requirements of the analytical methods. For each category of poultry considered, the minimum number of samples to be taken each year must be at least equal to one per 200 tones of annual production, with a minimum of 100 samples for each group of substances if the annual production of the category of birds considered is over 5 000 tones.
Control program/mechanisms
The control program/strategies in place
Broiler flocks
According to the provisions of N.S.V.F.S.A. President Order 147/2006, Regulation 2160/2003/EC, the following measures are to be adopted in order to prevent the dissemination of Salmonella enteritidis, Salmonella typhimurium, into commercial holdings. Animals from infected flocks belonging to commercial holdings are to be kept isolated and special conditions apply for removal of these animals. No bird may leave the house concerned unless the competent authority has authorized the slaughter or/and destruction under supervision of slaughter in a slaughterhouse designated by the competent authority. All the birds in the house must be slaughtered in accordance with the provisions of the REGULATION (EC) No. 853/2004 laying down specific hygiene rules for food of animal origin in order to reduce as much as possible the risk of spreading Salmonella.
Measures in case of the positive findings or single cases
Broiler flocks: Before slaughter at farm
In case of suspicion or confirmation of Salmonella enteritidis or Salmonella typhimurium the NRL shall notify immediately the N.S.V.F.S.A. and local S.V.F.S.D. In case of suspicion of infection the local S.V.F.S.D. and the relevant authorities: prohibited the movement of broilerstake. When the broilers are confirmed for the presence of Salmonella enteritidis or Salmonella typhimurium:1. Fresh meat from broilers may be placed on the market on the condition that it meets the requirement of absence of Salmonella in 25 grams from the meat.
2. The requirement laid down in point 1 does not apply to fresh poultry meat destined for heat treatment or another treatment to eliminate salmonella in accordance with Community legislation on food hygiene.

3. The criterion laid down in point 1 does not apply to fresh poultry meat destined for industrial heat treatment or another treatment to eliminate salmonella in accordance with Community legislation on food hygiene. Competent Authority will notify the operator to clean and disinfect the building from which the infected flock originated. After depopulation of a positive flock it is mandatory to harvest official samples to verify the efficiency of disinfections . In case that the results of these samples are not properly, it is mandatory to perform once again in the house the decontamination procedures and to take again official samples for verify the efficiency of disinfections. The house will be repopulated only when the results of the testes will be properly. A flock positive for a specific serotype will be recorded only once for that serotype. Operators with a flock which is positive for S. Enteritidis or S. Typhimurium will be contacted by the Competent Authority for advice on how to reduce or eliminate the Salmonella. Advice on the control of Salmonella in broilers will be available from government experts on Salmonella control. Advice may include recommendations on management, cleaning and disinfection, pest control, biosecurity, monitoring, and the potential use of other aids in the control of Salmonella.

**3. Notification system in place to the national competent authority[(c)]**

A positive laboratory finding of Salmonella ssp in food stuff derived from poultry is followed by a notification by RASFF to all levels (central, regional and local).Then the all food chain is controlled in order to identify the origin of the contamination, if it is possible. The contaminated products are traced back and detent under restrictions, till the results of salmonella serotyping come, and depending of the type of the Salmonella we apply different measures ( general measures : effective cleaning and disinfection of the premises and equipment are carried out and monitoring too).
Target serovars of Salmonella (SE+ST) in broiler flocks are mandatory notified according to national legislation (President Order no. 79/2009 with the followed amendments). The owner is responsible for the health and welfare of the poultry on the holding, and for ensuring that a veterinarian is consulted on disease and welfare issues as appropriate. It is mandatory for each holding to have a contract with a private veterinarian who is responsible for veterinary care. A veterinarian on behalf of the the Competent Authority carries out inspections on farms for animal health and welfare reasons, to take samples for residues, and to check medicine records. Also a veterinarian on behalf of the Competent Authority visit the farms and take official samples in the framework of Salmonella NCP according with the legislation in force. It is mandatory for each county sanitary veterinary and food safety directorate (local CA) to report to the NSVFSA every month the number of samples and results of these tests for each flock. Also the Salmonella NRL has the obligation to notify immediately NSVFSA and CSVFSD each positive sample for the relevant Salmonella.

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

National evaluation of the recent situation, the trends and source of infection- The programme for the control of Salmonella Enteritidis and Salmonella Typhimurium in broiler flocks has been in operation in Romania from 2008. As a result, the number of Salmonella Enteritidis and Salmonella Typhimurium infected broiler flocks is currently below the Community target. During 2015, a totally of 11619 broiler flocks were tested for Salmonella infection and there were 39 positive flocks for Salmonella Typhimurium and Salmonella Enteritidis . The prevalence for the target serotypes in broiler flocks in 2015 was 0,3%. In 2016, totally no. of 11945 broiler flocks were tested for Salmonella infection and there were 45 positive flocks for Salmonella Enteritidis . The prevalence for the target serotypes in broiler flocks in 2016 was 0,4%. However there is one notes of an increase of Salmonella outbreaks evolution in broilers flocks in semester two of year 2015 and semester 1 of 2016. The source of infection was represented by one day old chicks with origin from intra-trade movements.
In 2017, totally no. of 12549 broiler flocks were tested for Salmonella infection and there were 2 positive flocks for Salmonella spp: 1 positive flock of Salmonella Enteritidis and 1 positive flock of Salmonella Typhimurium.
In 2018, totally no. of 12672 broiler flocks were tested for Salmonella infection and there were 3 positive flocks for Salmonella Salmonella Enteritidis.

**5. Additional information**

The programme for the control of Salmonella Enteritidis and Salmonella Typhimurium in broiler flocks has been in operation in Romania from 2008.

Between 2008 and 2018 a decrease of the positive cases was noticed.

*** For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.
(b): The control program/strategies in place, including vaccination if relevant.If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.
(c): Mandatory: Yes/No.
(d): Minimum five years.
(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

## 18.Description of Monitoring/Surveillance/Control programmes system*: Salmonella in animal - Gallus gallus (fowl) - Farm - animal sample - Control and eradication programmes - Official and industry sampling - Census

**1. Monitoring/Surveillance/Control programmes system[(a)]**

*Sampling strategy*

*Laying hens flocks*

Starting with 2008 in Romania was implemented the National Salmonella control programme in laying hens flocks of Gallus gallus.

The main objective of our programme for the reduction of Salmonella enteritidis and Salmonella typhimurium in adult laying hens of Gallus gallus shall be a reduction of the maximum 2% percentage of positive adult laying flocks according to Regulation (EC) No 2160/2003 and Regulation (EC) 517/2011. The National Control Programme for Salmonella in laying flocks will be held in all holdings of laying hens consisting of at least 350 poultry of Gallus gallus which produce eggs for human consumption. Laying hens holdings which have between 350 and 1000 of birds will not be the subject of official testing, but will perform tests on the initiative

of operators (self-control). Small flocks that are reared to supply eggs for private domestic use, or small quantities of primary product supplied directly by the producer to the final consumer, will be exempt, as permitted in Regulation (EC) No 2160/2003 Article 1.3. The National Salmonella Control Programme encompasses the following serovars of zoonotic Salmonella: Salmonella enteritidis and Salmonella typhimurium. The sampling programme will be in accordance to Regulation 2160/2003 EC and Regulation 517/2011 EC.

*Breeding flocks (separate elite, grand parent and parent flocks when necessary)*

Starting with 2007 in Romania was implemented the National Salmonella control programme in breeding flocks of Gallus gallus. The sampling frame cover all adult breeding flocks comprising at least 250 birds. Sampling at the initiative of the operator and official sampling. Operator checks:-day -old chicks,-four-week-old birds,-birds two weeks before moving to laying phase or laying unit andevery second week during the laying period. Official sampling include:-within four weeks following moving to laying phase/laying unit,-toward the end of the laying phase, not earlier than eight weeks before the end of production cycle and-during the

production, at any time sufficiently distant from sample referred above.

*Frequency of the sampling*

*Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks*

Every flock is sampled (sampling at the initiative of the operator)

*Laying hens: Day-old chicks*

Laying hens: Day-old chicks No official sampling; only samples taken by operators (self control) can consist in:

(a) One chick box liner, up to a maximum of 10, for every 500 chicks delivered from each hatchery. Samples taken on the day of arrival. (b) The carcasses of all chicks, up to a maximum of 60, from each hatchery which are dead on arrival.

*Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period*

When birds are 4 weeks old and 2 weeks before moving to laying phase/laying unit (sampling at the initiative of the operator)
*Laying hens: Rearing period*
Laying hens: Rearing period No official sampling; only samples taken by the operators (self control)
*Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period*
Every 2 weeks during the production period (sampling at the initiative of the operator)
*Laying hens: Production period*
Laying hens: Production period Monitoring by operators shall take place according to Regulation (EC) No 517/2011 Annex Point 2:
Monitoring in Laying Flocks every 15 weeks starting when the birds are 22- 26 weeks of age. Official samples: The samples will be taken under the control of the Competent Authority for Regulation 2160/2003 from each layer flock on each holding with more than 1000 birds during the period of production of eggs for human consumption as specified in 2.1 of Annex to Commission Regulation (EC) No 517/2011. Laying hens: Before slaughter at farm Other: no official samples Laying hens: At slaughter Other: no official samples
Eggs at packing centre (flock based approach) Every 3 months
*Type of specimen taken*
*Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks*
internal linings of delivery boxes, dead chicks, meconium,etc
*Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period*
Environmental sample: boot swabs or composite faeces
*Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period*
Environmental sample: boot swabs or composite faeces
*Methods of sampling (description of sampling techniques)*
*Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks*
According to the National Control Programme. Samples comprising the following from each hatchery supplying the chicks: chick box liners (one liner per 500 chicks to maximum 10 liners) and all chicks dead on arrival (up to maximum of 60).
*Laying hens: Day-old chicks*
Samples taken by the operators can consist in:(a) One chick box liner, up to a maximum of 10, for every 500 chicks delivered from each hatchery. Samples taken on the day of arrival.(b) The carcasses of all chicks, up to a maximum of 60, from each hatchery which are dead on arrival.
*Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period*
According to the requirements of the National Control Programme, mandatory sampling is required at 4 weeks old and then 2 weeks before moving to the laying phase or laying unit as follows: - A minimum of 2 pairs of boot swabs or -A composite faeces sample made up from individual 1g faeces samples selected at random from sites to represent the whole building/space available to the birds.The size of the sample required is determined by the number of birds in the building/ flock.
*Laying hens: Rearing period*
Laying hens: Rearing period The samples can consist in: a minimum 2 pairs of boot swabs per house, or composite faeces sample taken according to the Council Regulation (EC) No 517/2011
*Breeding flocks: Production period*
According to the requirements of the National Control programme, mandatory sampling is required every 2 weeks during the laying/production period as follows: - A minimum of 5 pairs of boot swabs or -A composite faeces sample made up from individual 1g faeces samples selected at random from sites to represent the whole building/space available to the birds. The size of the sample required is determined by the number of birds in the building/ flock. In addition to the sampling above, 3 sets of Official Control Samples are collected from each breeding flock as follows: a) within 4 weeks of moving to the laying accommodation, b) in the middle of the lay, and c) within the last 8 weeks of production. Other operator voluntary monitoring can include hatchery debris, fluff, boot swabs, dust samples etc.
*Laying hens: Production period*
Laying hens: Production period Samples taken by the operators and samples taken by the Official samples consist in boot swabs/faeces, and dust samples Eggs at packing centre (flock based approach) Surface of egg shells and mixture of white and yellow.
*Case definition*
*Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks*

Samples taken by operators are sent to authorized laboratory for examination. Isolates are sent to the NRL for serotyping and phage typing and priority is given to any isolate culture result Group B or Group D.A flock is an epidemiological unit. Definition of a case: A positive case is a flock, where positive result in laboratory tests for detection of Salmonella was confirmed

*Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period*

Samples taken by operators are sent to authorized laboratory for examination. Isolates sent to NRL for serotyping and phage typing (as priority if a Group B or Group D has been cultured).A flock is an epidemiological unit. A positive case is a flock, where
positive result in laboratory tests for detection of Salmonella was confirmed.

*Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period*

Samples taken by operators are sent to authorised laboratory for examination. Isolates sent to NRL for serotyping and phage typing as priority if a Group B or Group D has been cultured. Official samples taken are sent to a approved C.S.V.F.S.L or to National Reference Laboratory for culture. A flock is an epidemiological unit. A positive case is a flock, where positive result in laboratory tests for detection of Salmonella was confirmed, according to Annex, point 4 of Regulation EU no. 200/2010.

*Diagnostic/analytical methods used*

*Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks*

Bacteriological method : ISO 6579:2002/Amd1:2007

*Laying hens: Day-old chicks*

Laying hens: Day-old chicks Samples taken by operators are sent to authorized and approved laboratory for examination. Isolates are sent to the NRL for serotyping and priority is given to any isolate culture result Group B or Group D.

*Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period*

Bacteriological method: ISO 6579: 2002/ Amd : 2002

*Laying hens: Rearing period*

Laying hens: Rearing period Samples taken by operators are sent to authorized and approved laboratory for examination. Isolates are sent to the NRL for serotyping and phage typing and priority is given to any isolate culture result Group B or Group D

Romania - 2016 31

*Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period*

Bacteriological method : ISO 6579:2002/Amd1:2007

*Laying hens: Production period*

Samples taken by operators are sent to authorized laboratory for examination. Isolates are sent to the NRL for serotyping and phage typing and priority is given to any isolate culture result Group B or Group D. A flock is an epidemiological unit.

**Eggs at packing centre (flock based approach)**

Eggs at packing centre (flock based approach) Definition of a positive finding here are 2 situations:-for the matrix which are found in Regulation 2005/2073, c=0, absence in 25 grams; -for the matrix which were not found in Regulation 2005/2073, but there were in The National Surveillance Programme no 4/31.01.2008, foodstuff is considered to be positive when Salmonella spp is detected.

Laying hens: Day-old chicks

Bacteriological method :ISO 6579:2002/Amd1:2007

Laying hens: Rearing period

Bacteriological method :ISO 6579:2002/Amd1:2007

Laying hens: Production period

Bacteriological method :ISO 6579:2002/Amd1:2007

Laying hens flocks

Laying hens flocks Live Salmonella vaccines are not used in the framework of national control programme where the manufacturer does not provide an appropriate method to distinguish bacteriological wild-type strains of salmonella from vaccine strains. A large proportion of the commercial layer flocks are vaccinated with a Salmonella vaccine.

The programmes regarding reduction of prevalence of Salmonella serotypes in Laying hens flocks and b*reeding flocks* populations is approved by the EC (https://ec.europa.eu/food/funding/animal health/national-veterinary programmes_en).

**2. Measures in place[(b)]**

Vaccination policy

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

Vaccination may only be used as a preventative measure; it is not an alternative to the requirements in Annex II C of Commission

Regulation (EC) No 2160/2003 for the use of specific control methods in the framework of the National Programmes for the Control of Salmonella. There are no restrictions on the use of Salmonella vaccines which have a marketing authorization. The vaccination is not mandatory and the costs regarding purchase of vaccine doses and the vaccination are incurred by the business operators. Vaccination is performed in accordance with Regulation 1177/2006 and differentiation tests are available to distinguish vaccine strains used in live vaccines from field strains of Salmonella.

Control program/mechanisms

The control program/strategies in place

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

Starting to 2007 obligatory National control programme for Salmonella is in place, according to Regulation 2160/2003 and Regulation 200/2010. National control programme for 5 serotypes of Salmonella is in place, which cover the whole territory of Romania.

Laying hens: Day-old chicks

Laying hens: Day-old chicks Samples taken by operators are sent to authorized laboratory for examination. Isolates are sent to the NRL for serotyping and phage typing and priority is given to any isolate culture result Group B or Group D. A flock is an epidemiological unit. Definition of a case positive: A positive case is a flock, where positive result in laboratory tests for detection of Salmonella was confirmed by official sampling.

Laying hens: Rearing period

Laying hens: Rearing period Samples taken by operators are sent to authorized laboratory for examination. Isolates are sent to the

NRL for serotyping and phage typing and priority is given to any isolate culture result Group B or Group D. A flock is an epidemiological unit. Definition of a positive case: A positive case is a flock, where positive result in laboratory tests for detection of Salmonella was confirmed by official sampling.

Laying hens flocks

According to the Romanian program of surveillance, prevention and animal disease control, of the diseases transmissible from animals to humans, animal protection and environment protection and program for surveillance and control in food safety field approved every year by N.S.V.F.S.A. President Order, feeding stuffs intended for poultry nutrition are checked in view to avoid the contamination with Salmonella spp. Also, in conformity with the same legislation the feed stuffs are checked in view to detect the use of antibiotics. Residues examination is performed according to the Romanian annual plan for examination for residues in live animals and animal origin products. For broiler, hens, turkeys, other poultry a sample consists on one or more animals depending on the requirements of the analytical methods. For each category of poultry considered, the minimum number of samples to be taken each year must be at least equal to one per 200 tones of annual production, with a minimum of 100 samples for each group of substances if the annual production of the category of birds considered is over 5 000 tones.

Laying hens flocks

The control program/strategies in place The National Control Programme will be implemented throughout Romania, covering all the national territory and will cover all laying hens flocks of Gallus gallus with more than 350 birds . The administrative boundaries are the boundaries of the country. Romania is administrative divided in 42 counties. There are 42 County Sanitary Veterinary and Food Safety Directorates and 41 County Sanitary veterinary and food Safety Laboratories. Measures in the event of positive findings or single cases.

1. Eggs shall not be used for direct human consumption as table eggs unless they originate from a commercial flock of laying hens subject to Salmonella national control programme established and is not under official restriction.

2. Eggs originating from flocks with unknown health status, that are suspected of being infected or that

are infected with Salmonella serotypes for which a target for reduction has been set or which were identified as the source of infection in a specific human food-borne outbreak, may be used for human consumption only if they are treated in a manner that guarantees the destruction of all Salmonella serotypes with public health significance in accordance with Community legislation on food hygiene. Eggs originating from flocks with unknown health status, that are suspected of being infected or that are infected with Salmonella serotypes for which a target for reduction has been set or which were identified as the source of infection in a specific human food-borne outbreak, shall be: (a) considered as Class B eggs as defined in Article 2(4) of Commission Regulation (EC) No 589/2008 laying down detailed rules for implementing Council Regulation (EC) No 1234/2007 on marketing standards for eggs (1); (b) marked with the indication referred to in Article 10 of Commission Regulation (EC) No 589/2007 which clearly distinguishes them from Class A eggs prior to being placed on the market; (c) prohibited access to packaging centers unless the competent authority is satisfied with the measures to prevent possible cross-contamination of eggs from other flocks.
3. When birds from infected flocks are slaughtered or destroyed, steps are taken to reduce the risk of spreading zoonoses as soon as possible. Slaughtering shall be carried out in accordance with Community legislation on food hygiene. Products derived from such birds may be placed on the market for human consumption in accordance with Community legislation on food hygiene. If they are not destined for human consumption, this products must be used or disposed of in accordance with Regulation (EC) No 1069/2009.
4. In order to exclude false-positive initial results, the competent authority may lift the restrictions laid down in point 2 of this Part: (a) when the flock of layers is not the source of infection for humans by the consumption of eggs or egg products as a result of the epidemiological investigation of food-borne outbreaks in accordance with Article 8 of Directive 2003/99/EC; and (b) where the flock is subjected to a Salmonella national control programme and Salmonella serotypes which a target for reduction has been set, is not confirmed by the following sampling protocol carried out by the competent authority: (i) the technical specifications referred to in Article 5 of Commission Decision 2004/665/EC (seven samples); however, a sub-sample of 25 grams must be collected of each faecal material and dust sample for analysis; all samples must be analyzed separately; or (ii) bacteriological investigation of the caecal and oviducts of 300 birds; or (iii) bacteriological investigation of the shell and the content of 4 000 eggs of each flock in pools of maxi-mum 40 eggs. In addition to the sampling in point (b), the competent authority shall verify the absence of the use of antimicrobial, potentially affecting the result of the analysis of the sampling.

**3. Notification system in place to the national competent authority[(c)]**

A positive laboratory finding of Salmonella ssp in food stuff derived from poultry is followed by a notification by RASFF to all levels (central, regional and local).Then the all food chain is controlled in order to identify the origin of the contamination, if it is possible. The contaminated products are traced back and detent under restrictions, till the results of salmonella serotyping come, and depending of the type of the Salmonella we apply different measures (general measures: effective cleaning and disinfection of the premises and equipment are carried out and monitoring too).
Target serovars of Salmonella (SE+ST) in broiler flocks are mandatory notified according to national legislation (President Order no. 79/2009 with the followed amendments). The owner is responsible for the health and welfare of the poultry on the holding, and for ensuring that a veterinarian is consulted on disease and welfare issues as appropriate. It is mandatory for each holding to have a contract with a private veterinarian who is responsible for veterinary care. A veterinarian on behalf of the the Competent Authority carries out inspections on farms for animal health and welfare reasons, to take samples for residues, and to check medicine records. Also a veterinarian on behalf of the Competent Authority visit the farms and take official samples in the framework of Salmonella NCP according with the legislation in force. It is mandatory for each county sanitary veterinary and food safety directorate (local CA) to report to the NSVFSA every month the number of samples and results of these tests for each flock. Also the Salmonella NRL has the obligation to notify immediately NSVFSA and CSVFSD each positive sample for the relevant Salmonella.

On the basis of National Control Programme 5 serotypes in breeding flocks are under control. Target serovars of Salmonella (SE+ST+SI+SH+SV) in breeders are mandatory notified according to national legislation (President Order no. 79/2009 with the followed amendments). The owner is responsible for the

health and welfare of the poultry on the holding, and for ensuring that a veterinarian is consulted on disease and welfare issues as appropriate. It is mandatory for each holding to have a contract with a private veterinarian who is responsible for veterinary care. A veterinarian on behalf of the the Competent Authority carries out inspections on farms for animal health and welfare reasons, to take samples for residues, and to check medicine records. Also a veterinarian on behalf of the Competent Authority visit the farms and take official samples in the framework of Salmonella NCP according with the legislation in force. It is mandatory for each county sanitary veterinary and food safety directorate (local CA) to report to the NSVFSA every month the number of samples and results of these tests for each flock. Also the Salmonella NRL has the obligation to notify immediately NSVFSA and CSVFSD each positive sample for the relevant Salmonella.

Notification system in place A positive laboratory finding of Salmonella ssp in food stuff derived from poultry is followed by a notification by RASFF to all levels (central, regional and local).Then the all food chain is controlled in order to identify the origin of the contamination, if it is possible. The contaminated products are traced back and detent under restrictions, till the results of salmonella serotyping come, and depending of the type of the Salmonella we apply different measures ( general measures : effective cleaning and disinfection of the premises and equipment are carried out and monitoring too). Target serovars of Salmonella (SE+ST) in laying hens are mandatory notified according to national legislation (President Order no. 79/2009 with the followed amendments). The owner is responsible for the health and welfare of the poultry on the holding, and for ensuring that a veterinarian is consulted on disease and welfare issues as appropriate. It is mandatory for each holding to have a contract with a private veterinarian who is responsible for veterinary care. A veterinarian on behalf of the the Competent Authority carries out inspections on farms for animal health and welfare reasons, to take samples for residues, and to check medicine records. Also a veterinarian on behalf of the Competent Authority visit the farms and take official samples in the framework of Salmonella NCP according with the legislation in force. It is mandatory for each county sanitary veterinary and food safety directorate (local CA) to report to the NSVFSA every month the number of samples and results of these tests for each flock. Also the Salmonella NRL has the obligation to notify immediately NSVFSA and CSVFSD each positive sample for the relevant Salmonella.

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

Results of the investigation National evaluation of the recent situation, the trends and source of infection-Starting to 2008 obligatory National control programme for Salmonella is in place, according to Regulation 2160/ 2003 . As a result, the number of Salmonella Enteritidis and Salmonella Typhimurium infected laying hens flocks is currently below the Community target. During 2015, a totally of 683 laying hens flocks were tested for Salmonella and there were only 10 flocks positive for Salmonella Enteritidis . The prevalence for the target serotypes in laying hens flock in 2015 was 1,46%, which is low and below the Community target. In 2016 a totally of 617 laying hens flocks were tested for Salmonella and there were only 7 flocks positive for Salmonella Typhimurium and Salmonella Enteritidis . The prevalence for the target serotypes in laying hens flock in 2016 was 1,1%, which is low and below the Community target. History of the disease and/or infection in the county The programme for the control of Salmonella Enteritidis and Salmonella Typhimurium in laying hens has been in operation in Romania from 2008. Between 2008 and 2017 a decrease of the positive cases was noticed.
In 2017, totally no. of 1056 laying hens flocks were tested for Salmonella infection and was positive flocks for Salmonella Enteritidis.
In 2018, totally no. of 1096 laying hens flocks were tested for Salmonella infection and was 2 positive flocks for Salmonella Enteritidis

Starting to 2007 obligatory National control programme for Salmonella is in place, according to Regulation 2160/ 2003. As a result, the number of Salmonella target serovars infected breeder flocks is currently below the Community target. During 2015, a totally of 318 breeder flocks were tested for Salmonella and there were no positive flocks. The prevalence for the target serotypes in breeder flocks in 2015 was 0%, which is low and below the Community target. In 2016 a totally of 377 breeder flocks were tested for Salmonella and there were only 3 flocks positive for Salmonella Infantis. The prevalence for the target

serotypes in breeder flocks in 2016 was 0,8%, which is low and below the Community target.
In 2017, totally no. of 656 breeder flocks were tested for Salmonella infection and was 1 positive flocks for Salmonella Typhimurium and 1 positive flocks for Salmonella Infantis .
In 2018, totally no. of 704 breeder flocks were tested for Salmonella infection and was 1 positive flocks for Salmonella Infantis and 2 positive flocks for Salmonella Enteritidis.

**5. Additional information**

The programme for the control of Salmonella target serovars in breeder flocks and laying hens flocks has been in operation in Romania from 2007. Between 2008 and 2018 a decrease of the positive cases was noticed.

* **For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.
(b): The control program/strategies in place, including vaccination if relevant.If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.
(c): Mandatory: Yes/No.
(d): Minimum five years.
(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

## 19. Description of Monitoring/Surveillance/Control programmes system*: Salmonellosis, other species

**1. Monitoring/Surveillance/Control programmes system[(a)]**

There is no official monitoring system on farm level. Investigations are initiated by the owners of the animals.
Frequency of the sampling: vvoluntary sampling usually taken by a veterinarian for diagnostic purposes.
Type of specimen taken: faeces and various organs
Diagnostic/analytical methods used: OIE method or those described in SR EN ISO 6579-1:2017

**2. Measures in place[(b)]**

**3. Notification system in place to the national competent authority[(c)]**

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

**5. Additional information**

*** For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.
(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.
(c): Mandatory: Yes/No.
(d): Minimum five years.
(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

**20.Description of Monitoring/Surveillance/Control programmes system*: Salmonella spp. - Meat from bovine and products thereof - food sample**

**1. Monitoring/Surveillance/Control programmes system[(a)]**

The Romanian Surveillance Program is a national program, published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority (N.S.V.F.S.A.), yearly updated and the susceptibility testing of Salmonella from bovine meat and products thereof is a part of the program. According to the provisions of the Romanian National Surveillance Program, all food industry establishments are classified into 3 categories, based on the risk assessment provided by the official vets acting at regional/county Sanitary Veterinary and Food Safety Directorates level: category III - high risk; category II - medium risk; and category I - low risk. The samples for monitoring and testing of Salmonella are compulsory taken by the official vets acting at slaughterhouses, cutting plants, meat processing plant on the base of risk assessment of establishments.

The *control program* for Salmonella spp., according to the provisions of the Romanian National Surveillance Program, approved by Order of the N.S.V.F.S.A. no 35/2016 (with subsequent amendments and completions) and in accordance with European Union regulations, includes sampling and analysis, as follows:

A) Sampling at meat processing plant including minced meat, meat preparation, and meat products (from food producing species) placed on the market and during the period of validity for the testing of Salmonella spp., as a food safety criteria.

B) Sampling at slaughterhouses from bovine carcasses shall be sampled on the surface of carcases of this species by the non-destructive method using abrasive sponges for the testing

of Salmonella spp. as a hygiene criteria of the technological process.

The *type of specimen* taken according to the stage of sampling is:

- At slaughterhouse and cutting plant - Surface of carcass, fresh meat (muscle tissue) and offal (liver, kidney);
- At meat processing plant - Meat products, meat preparation, minced meat;
- Retail - Raw material (fresh meat) and finish products (meat products, meat preparations, minced meat).

The samples for monitoring and testing of Salmonella are taken by the official vets acting at slaughterhouses, cutting plants, meat processing plant on the base of risk assessment of establishments, as follows *frequency*:

Samples of meat from bovine *at slaughterhouses*, including carcasses surfaces, fresh meat (muscle tissue) and offal (liver, kidney) for testing of Salmonella:

- once a month (monthly) at slaughterhouses in category III;
- once a quarter (quarterly) at slaughterhouses in category II;
- once a semester (twice/year) at slaughterhouses in category I.

Samples of fresh meat from bovine *at cutting plants* for testing of Salmonella:

- once a quarter (quarterly) at cutting plants in category III;
- once a semester (twice/year) at cutting plants in category II;
- once a year (annually) at cutting plants in category I.

Samples of meat from bovine *at meat processing plant*, including meat products, minced meat and meat preparation for testing of Salmonella:

- once a quarter (quarterly) at meat processing plants in category III;
- once a semester (twice/year) at meat processing plants in category II;
- once a year (annually) at meat processing plants in category I .

From *retail* the samples of meat from bovine and products thereof for monitoring and testing of Salmonella are taken by the official vets annually and also in any case of: consumer complaints, suspicions or food borne outbreaks.

*Methods of sampling (sampling techniques):*

*At slaughterhouse and cutting plant* - According to the provisions of the Regulation 2005/2073/EC, with subsequent amendments and completions, shall be sampled in the framework of National Surveillance Program and of food bussiness operators own control programmes. Sample sites must be selected taking into account the slaughter technology used in each plant and five carcases shall be sampled at random during each sampling session. The sampling for Salmonella analyses is performed using an abrasive sponge sampling method. Areas most likely to be contaminated shall be selected. The total sampling area shall cover a

minimum of 400 cm2. For bovine meat including fresh meat (muscle tissue) and offal (liver, kidney) at slaughterhouse level and for fresh meat at cutting plant level the final sample it is obtained in the lab and consists of at least 25 grams of each product.

*At meat processing plant* - According to the provisions of the Regulation 2005/2073/EC, with subsequent amendments and completions, shall be sampled in the framework of National Surveillance Program and of food bussiness operators own control programmes. For meat from bovine, for the matrix which are found in Regulation 2005/2073 a sample consists of 5 pooled sample; and for the matrix which are not found in Regulation 2005/2073, but are mentioned in The National Surveillance Program, a tested unit consists of 1 sample. According to the provisions of the Regulation 2005/2073/EC, with subsequent amendments and completions, the food business operators of establishments producing minced meat, meat preparations or mechanically separated meat shall take samples for microbiological analysis at least once a week. The day of sampling shall be changed each week to ensure that each day of the week is covered. In the case of sampling for Salmonella analyses of minced meat, meat preparations and carcases, the frequency may be reduced to fortnightly if satisfactory results have been obtained for 30 consecutive weeks.

*At retail* - According to the provisions of the Regulation 2005/2073/EC, with subsequent amendments and completions, shall be sampled in the framework of National Surveillance Program and of food bussiness operators own control programmes.

*Diagnostic/analytical methods* used for detection and serotyping Salmonella is microbiological method: EN ISO 6579 - detection and serotyping (Kauffmann White Le Minor scheme).

*Definition of positive finding* - Bovine meat and products thereof are considered to be positive when Salmonella spp. is isolated by the microbiological method.

**2. Measures in place[(b)]**

A positive laboratory finding of Salmonella spp. it is followed by a notification to RASFF to all levels (central, regional and local). Then all the food chain it is controlled in order to identify the source of contamination. The contaminated batches of bovine meat are traced back and detent under restrictions, until the results of Salmonella serotyping is communicated and depending on the serotype of Salmonella the different measures are applied. If the sample of bovine meat is found positive for Salmonella Enteritidis and/or Salmonella Typhimurium the whole batch of bovine meat is declared unfitted for human consumption and are denaturated. If the sample of bovine meat is found positive for Salmonella spp., other than Salmonella Enteritidis and Salmonella Typhimurium, the bovine meat will admitted for human consumption only if it is undergone to an adequate heat treatment, under veterinary surveillance and if the results of the microbiological analysis of the heat treated bovine meat is found negative for Salmonella spp. If the sample of bovine meat products is found positive for Salmonella spp., the whole batch of bovine meat products are declared unfitted for human consumption and are denaturated.

**3. Notification system in place to the national competent authority[(c)]**

The laboratory that identifies the positive sample has the obligation to notify the positive result through the RASFF to the regional and central authority, and the regional authority will notify the food business operator.

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

In 2013, were isolated 6 strains of Salmonella in meat from bovine. In 2014, were isolated 9 strains of Salmonella in meat from bovine. In 2015, 1 strain of Salmonella was isolated in meat from bovine and products thereof. In 2016, were isolated 5 strains of Salmonella in meat from bovine and products thereof and in 2017, were isolated 3 strains of Salmonella in meat from bovine.

In 2018, 1 strain of Salmonella was isolated in meat from bovine.

Bovine meat is not considered to be an important source of infection at human cases in Romania.

**5. Additional information**

Comparison of the Salmonella serotypes found in animals, feeding stuffs, food and human helps to suggest possible sources of infection in the food chain.

*** For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

## 21.Description of Monitoring/Surveillance/Control programmes system*: Salmonella spp. - Meat from pig and products thereof - food sample

**1. Monitoring/Surveillance/Control programmes system[(a)]**

The Romanian Surveillance Program is a national program, published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority

(N.S.V.F.S.A.), yearly updated and the susceptibility testing of Salmonella from pig meat and products thereof is a part of the program. According to the provisions of the Romanian National Surveillance Program, all food industry establishments are classified into 3 categories, based on the risk assessment provided by the official vets acting at regional/county Sanitary Veterinary and Food Safety Directorates level: category III - high risk; category II - medium risk; and category I - low risk. The samples for monitoring and testing of Salmonella are compulsory taken by the official vets acting at slaughterhouses, cutting plants, meat processing plant on the base of risk assessment of establishments.

The *control program* for Salmonella spp., according to the provisions of the Romanian National Surveillance Program, approved by Order of the N.S.V.F.S.A. no 35/2016 (with subsequent amendments and completions) and in accordance with European Union regulations, includes sampling and analysis, as follows:

A) Sampling at meat processing plant including minced meat, meat preparation, and meat products (from food producing species) placed on the market and during the period of validity for the testing of Salmonella spp., as a *food safety criteria*.

B) Sampling at slaughterhouses from pig carcasses shall be sampled on the surface of carcases of this species by the non-destructive method using abrasive sponges for the testing of Salmonella spp. as a *hygiene criteria* of the technological process.

The *type of specimen* taken according to the stage of sampling is:

- At slaughterhouse and cutting plant - Surface of carcass, fresh meat (muscle tissue) and offal (liver, kidney);
- At meat processing plant - Meat products, meat preparation, minced meat;
- Retail - Raw material (fresh meat) and finish products (meat products, meat preparations, minced meat).

The samples for monitoring and testing of Salmonella are taken by the official vets acting at slaughterhouses, cutting plants, meat processing plant on the base of risk assessment of establishments, as follows *frequency*:

Samples of meat from pig *at slaughterhouses*, including carcasses surfaces, fresh meat (muscle tissue) and offal (liver, kidney) for testing of Salmonella:

- once a month (monthly) at slaughterhouses in category III;
- once a quarter (quarterly) at slaughterhouses in category II;
- once a semester (twice/year) at slaughterhouses in category I.

Samples of fresh meat from pig *at cutting plants* for testing of Salmonella:

- once a quarter (quarterly) at cutting plants in category III;
- once a semester (twice/year) at cutting plants in category II;
- once a year (annually) at cutting plants in category I.

Samples of meat from pig *at meat processing plant*, including meat products, minced meat and

meat preparation for testing of Salmonella:

- once a quarter (quarterly) at meat processing plants in category III;
- once a semester (twice/year) at meat processing plants in category II;
- once a year (annually) at meat processing plants in category I .

From *retail* the samples of meat from pig and products thereof for monitoring and testing of Salmonella are taken by the official vets annually and also in any case of: consumer complaints, suspicions or food borne outbreaks.

Methods of sampling (sampling techniques):

*At slaughterhouse and cutting plant* - According to the provisions of the Regulation 2005/2073/EC, with subsequent amendments and completions, shall be sampled in the framework of National Surveillance Program and of food bussiness operators own control programmes. Sample sites must be selected taking into account the slaughter technology used in each plant and five carcases shall be sampled at random during each sampling session. The sampling for Salmonella analyses is performed using an abrasive sponge sampling method. Areas most likely to be contaminated shall be selected. The total sampling area shall cover a minimum of 400 cm2. For pig meat including fresh meat (muscle tissue) and offal (liver, kidney) at slaughterhouse level and for fresh meat at cutting plant level the final sample it is obtained in the lab and consists of at least 25 grams of each product.

*At meat processing plant* - According to the provisions of the Regulation 2005/2073/EC, with subsequent amendments and completions, shall be sampled in the framework of National Surveillance Program and of food bussiness operators own control programmes. For meat from pig, for the matrix which are found in Regulation 2005/2073 a sample consists of 5 pooled sample; and for the matrix which are not found in Regulation 2005/2073, but are mentioned in The National Surveillance Program, a tested unit consists of 1 sample. According to the provisions of the Regulation 2005/2073/EC, with subsequent amendments and completions, the food business operators of establishments producing minced meat, meat preparations or mechanically separated meat shall take samples for microbiological analysis at least once a week. The day of sampling shall be changed each week to ensure that each day of the week is covered. In the case of sampling for Salmonella analyses of minced meat, meat preparations and carcases, the frequency may be reduced to fortnightly if satisfactory results have been obtained for 30 consecutive weeks.

*At retail* - According to the provisions of the Regulation 2005/2073/EC, with subsequent amendments and completions, shall be sampled in the framework of National Surveillance Program and of food bussiness operators own control programmes.

*Diagnostic/analytical methods* used for detection and serotyping Salmonella is microbiological method: EN ISO 6579 - detection and serotyping (Kauffmann White Le Minor scheme).

*Definition of positive finding* – meat from pig and products thereof are considered to be positive

when Salmonella spp. is isolated by the microbiological method.

**2. Measures in place[(b)]**

A positive laboratory finding of Salmonella spp. is followed by a notification by RASFF to all levels (central, regional and local). Then all the food chain it is controlled in order to identify the source of contamination. The contaminated batches of pig meat are traced back and detent under restrictions, until the results of Salmonella serotyping is communicate and depending on the serotype of Salmonella the different measures are applied. If the sample of pig meat was found positive for Salmonella Enteritidis and/or Salmonella Typhimurium then the whole batch of pig meat is declared unfitted for human consumption and is denaturated. If a sample of pig meat is found positive for Salmonella spp., other than Salmonella Enteritidis and Salmonella Typhimurium, the pig meat can be admitted for human consumption only if it is undergone to an adequate heat treatment, under veterinary surveillance and if the results of microbiological analysis of the pig meat heat treated are found negative for Salmonella spp.If a sample of pig meat products is found positive for Salmonella spp. the whole batch of pig meat products are declared unfitted for human consumption and is denaturated.

**3. Notification system in place to the national competent authority[(c)]**

The laboratory that identifies the positive sample has the obligation to notify the positive result through the RASFF to the regional and central authority, and the regional authority will notify the food business operator.

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

In 2013, were isolated 93 strains of Salmonella in meat from pig and products thereof. In 2014, were isolated 51 strains of Salmonella in meat from pig and products thereof. In 2015, were isolated 72 strains of Salmonella in meat from pig and products thereof. In 2016, were isolated 81 strains of Salmonella in meat from pig and products thereof. In 2017, were isolated 44 strains of Salmonella in meat from pig and products thereof. In 2018, were isolated 71 strains of Salmonella in meat from pig and products thereof.

**5. Additional information**

Comparison of the Salmonella serotypes found in animals, feeding stuffs, food and human helps to suggest possible sources of infection in the food chain.

* **For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures

are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

## 22.Description of Monitoring/Surveillance/Control programmes system*: Salmonella spp. - Meat from poultry (of broilers and turkeys) and products thereof - food sample

### 1. Monitoring/Surveillance/Control programmes system[(a)]

The Romanian Surveillance Program is a national program, published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority (N.S.V.F.S.A.), yearly updated and the susceptibility testing of Salmonella from poultry (broilers and turkeys) and products thereof is a part of the program. According to the provisions of the Romanian National Surveillance Program, all food industry establishments are classified into 3 categories, based on the risk assessment provided by the official vets acting at regional/county Sanitary Veterinary and Food Safety Directorates level: category III - high risk; category II - medium risk; and category I - low risk. The samples for monitoring and testing of Salmonella are compulsory taken by the official vets acting at slaughterhouses, cutting plants, meat processing plant on the base of risk assessment of establishments.

The *control program* for Salmonella spp., according to the provisions of the Romanian National Surveillance Program, approved by Order of the N.S.V.F.S.A. no 35/2016 (with subsequent amendments and completions) and in accordance with European Union regulations, includes sampling and analysis, as follows:

A) Sampling at meat processing plant including minced meat, meat preparation, and meat products (from food producing species) placed on the market and during the period of validity for the testing of Salmonella spp., as a *food safety criteria*.

B) Sampling at slaughterhouses from poultry carcases of broilers and turkeys shall be sampled of neck skin for the testing of Salmonella spp., as a *hygiene criteria* of the technological process.

The *type of specimen* taken according to the stage of sampling is:

- At slaughterhouse and cutting plant - Surface of carcass, fresh meat (muscle tissue) and offal;
- At meat processing plant - Meat products, meat preparation, minced meat; mechanically separated meat (MSM);
- Retail - Raw material (fresh meat) and finish products (meat products, meat preparations, minced meat).

The samples for monitoring and testing of Salmonella are taken by the official vets acting at

slaughterhouses, cutting plants, meat processing plant on the base of risk assessment of establishments, as follows *frequency*:

Samples of broilers and turkeys *at slaughterhouses*, including carcasses surfaces, fresh meat (muscle tissue) and offal, for testing of Salmonella:

- once a month (monthly) at slaughterhouses in category III;
- once a quarter (quarterly) at slaughterhouses in category II;
- once a semester (twice/year) at slaughterhouses in category I.

Samples of broilers/turkeys *at cutting plants* for testing of Salmonella:

- once a quarter (quarterly) at cutting plants in category III;
- once a semester (twice/year) at cutting plants in category II;
- once a year (annually) at cutting plants in category I.

Samples of broilers/turkeys *at meat processing plant*, including meat products, minced meat and meat preparation for testing of Salmonella:

- once a quarter (quarterly) at meat processing plants in category III;
- once a semester (twice/year) at meat processing plants in category II;
- once a year (annually) at meat processing plants in category I .

From *retail* the samples of broilers/turkeys and products thereof for monitoring and testing of Salmonella are taken by the official vets annually and also in any case of: consumer complaints, suspicions or food borne outbreaks.

Methods of sampling (sampling techniques):

*At slaughterhouse and cutting plant* - According to the provisions of Regulation 2005/2073/EC, with subsequent amendments and completions, for the Salmonella analyzes, a minimum of 15 carcass were sampled at random during each sampling session and after chilling. A piece of approximately 10 g from neck skin was obtained from each carcass. On each occasion the neck skin samples from three carcasses were pooled before examination in order to form 5 x 25 g final samples. For poultry meat including fresh meat (muscle tissue) at slaughterhouse level and at cutting plant level the final sample it is prepared in the lab and consists of at least 25 grams of each product.

*At meat processing plant* - According to the provisions of the Regulation 2005/2073/EC, with subsequent amendments and completions, shall be sampled in the framework of National Surveillance Program and of food bussiness operators own control programmes. For broilers and turkeys meat, for the matrix which are found in Regulation 2005/2073 a sample consists of 5 pooled sample; and for the matrix which are not found in Regulation 2005/2073, but are mentioned in The National Surveillance Program, a tested unit consists of 1 sample.

*At retail* - According to the provisions of the Regulation 2005/2073/EC, with subsequent

amendments and completions, shall be sampled in the framework of National Surveillance Program and of food bussiness operators own control programmes.

*Diagnostic/analytical methods* used for detection and serotyping Salmonella is microbiological method: EN ISO 6579 - detection and serotyping (Kauffmann White Le Minor scheme).

*Definition of positive finding* – meat from broiler and turkey and products thereof are considered to be positive when Salmonella spp. is isolated by the microbiological method.

**2. Measures in place[(b)]**

A positive laboratory finding of Salmonella spp. it is followed by a notification to RASFF to all levels (central, regional and local). Then all the food chain it is controlled in order to identify the source of contamination. The contaminated batches of broiler/turkey meat are traced back and detent under restrictions, until the results of Salmonella serotyping is communicated and depending on trhe seotype of Salmonella the different measures are applied. If the sample of turkey meat is found positive for Salmonella Enteritidis and/or Salmonella Typhimurium the whole batch of turkey meat is declared unfitted for human consumption and are denaturated. If the sample of of broiler/turkey meat is found positive for Salmonella spp., other than Salmonella Enteritidis and Salmonella Typhimurium, the broiler /turkey meat will admitted for human consumption only if it is undergone to an adequate heat treatment, under veterinary surveillance and if the results of the microbiological analysis of the heat treated broiler/turkey meat is found negative for Salmonella spp. If the sample of broiler/turkey meat products is found positive for Salmonella spp. the whole batch of broiler/turkey meat products are declared unfitted for human consumption and are denaturated.

**3. Notification system in place to the national competent authority[(c)]**

The laboratory that identifies the positive sample has the obligation to notify the positive result through the RASFF to the regional and central authority, and the regional authority will notify the food business operator.

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

In 2013, were isolated 219 strains of Salmonella in meat from poultry and products thereof (all the strains were from the broiler and none of them from the turkey). In 2014, were isolated 92 strains of Salmonella in meat from poultry and products thereof (all the strains were from the broiler and none of them from the turkey). In 2015, were isolated 141 strains of Salmonella in meat from poultry and products thereof (all the strains were from the broiler and none of them from the turkey). In 2016, were isolated 81 strains of Salmonella in meat from poultry and products thereof (all the strains were from the broiler and none of them from the turkey). In 2017, were isolated 109 strains of Salmonella in meat from poultry and products thereof (from which 101 meat from broiler and 8 meat from turkey).

In 2018, were isolated 185 strains of Salmonella in meat from poultry and products thereof (from

which 170 meat from broiler and 15 meat from turkey).

Meat from poultry can be considered to be an important source of infection at human cases in Romania.

**5. Additional information**

Comparison of the Salmonella serotypes found in animals, feedingstuffs, foodstuffs and human helps to suggest possible sources of infection in the food chain.

*** For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

**(a**): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

## 23.Description of Monitoring/Surveillance/Control programmes system*: Salmonella spp. in Egg and egg products - food sample

**1. Monitoring/Surveillance/Control programmes system[(a)]**

The Romanian Surveillance Program is a national program, published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority (N.S.V.F.S.A.), yearly updated and the susceptibility testing of Salmonella from egg and egg products is a part of the program.The *control program* for Salmonella spp. according to the provisions of the Romanian National Surveillance Program, includes sampling and analysis of eggs and egg products, as follows:

- at egg packing center (EPC) - samples of eggs - once a quarter (trimester);
- at the establishments producing liquid egg - samples of eggs and finish products - once a quarter (trimester),
- at the egg processing units - samples of eggs and finish products - once a quarter (trimester);

From *retail* the samples of eggs and finish products for monitoring and testing of Salmonella are taken by the official vets annually and also in any case of: consumer complaints, suspicions or

food borne outbreaks.any situation.

The *type of specimen* taken according to the stage of sampling is:

- at egg packing center (EPC) - Surface of egg shells and egg content
- at the establishments producing liquid egg - Egg white, egg yolk and mixture of white and yolk;
- at the egg processing units - Raw material for egg products (egg white, egg yolk and mixture of white and yolk);
- at retail – egg and egg products.

*Diagnostic/analytical methods* used for detection and serotyping Salmonella is microbiological method: EN ISO 6579 - detection and serotyping (Kauffmann White Le Minor scheme).

*Definition of positive finding* – Eggs and egg products are considered to be positive when Salmonella spp. is isolated by the microbiological method.

**2. Measures in place[(b)]**

A positive laboratory finding of Salmonella spp. it is followed by a notification to RASFF to all levels (central, regional and local). Then all the food chain it is controlled in order to identify the source of contamination. The contaminated batches of eggs and egg products are traced back and detent under restrictions, until the results of Salmonella serotyping is communicated and depending on trhe seotype of Salmonella the different measures are applied. If the sample of eggs and egg products is found positive for Salmonella Enteritidis and/or Salmonella Typhimurium the whole batch of eggs and egg products is declared unfitted for human consumption and are denaturated. If the sample of eggs and egg products is found positive for Salmonella spp., other than Salmonella Enteritidis and Salmonella Typhimurium, the eggs and egg products will admitted for human consumption only if it is undergone to an adequate heat treatment, under veterinary surveillance and if the results of the microbiological analysis of the heat treated eggs and egg products is found negative for Salmonella spp.

**3. Notification system in place to the national competent authority[(c)]**

The laboratory that identifies the positive sample has the obligation to notify the positive result through the RASFF to the regional and central authority, and the regional authority will notify the food business operator.

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

In 2013, 1 strain of Salmonella was isolated in egg. In 2014, were isolated 3 strains of Salmonella in egg. In 2015, were isolated 5 strains of Salmonella in egg. In 2016, were isolated 21 strains of Salmonella in egg. In 2017, were isolated 8 strains of Salmonella in egg and egg products**.**

In 2018, were isolated 2 strains of Salmonella in egg and 31 strains in egg products.

Egg and egg products is not considered to be an important source of infection at human cases in Romania.

**5. Additional information**

Comparison of the Salmonella serotypes found in animals, feedingstuffs, foodstuffs and human helps to suggest possible sources of infection in the food chain.

*** For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

## 24.Description of Monitoring/Surveillance/Control programmes system*: Salmonella spp. in feedingstuffs - feed sample

**1. Monitoring/Surveillance/Control programmes system[(a)]**

The Romanian Surveillance Program is a national program, published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority (N.S.V.F.S.A.), yearly updated and the susceptibility testing of Salmonella in feed is a part of the program. The samples for the official control of feed are taken by the official vets acting, units producing compound feed, at farms, at suppliers of raw materials, at warehouses and at the vehicles registered for the transport of feed. In The National Plan for the official control of animal feedstuffsin the scope of the supervision of Veterinary Inspection, which is approved every year, samples are going to be randomly taken from the feed business operators and tested for Salmonella. The types of specimen taken of sampling are: raw materials, compound feed, fats, premixes, roughage.

According to the provisions of the Regulation 2005/183/EC, with subsequent amendments and completions, the feed business operators of establishments producing raw materials and

compound feed shall take samples for microbiological analysis.

*Diagnostic/analytical methods* used for detection and serotyping Salmonella is microbiological method: EN ISO 6579 - detection and serotyping (Kauffmann White Le Minor scheme).

*Definition of positive finding* – the feedingstuffs are considered to be positive when Salmonella spp. is isolated by the microbiological method.

**2. Measures in place[(b)]**

The feeding stuffs for poultry and other animals must be free from Salmonella. Veterinary Inspection conducts random, regular inspection in feeding stuffs production plants, in particular of microbiological standards, types of internal controls used by the owners of these plants to guarantee the appropriate quality of final product.

A positive laboratory finding of Salmonella spp. it is followed by a notification to RASFF to all levels (central, regional and local). The contaminated batches of feedingstuffs are traced back and detent under restrictions, and the different measures are applied. Operators duties in case of detection of inappropriate microbiological quality of product: notifying the veterinary inspector regarding of the batch of products from which the sample testing were taken; secondary processing of contaminated batch, according to an indicated method, under supervision of veterinary inspection and increase the frequency of sampling.

**3. Notification system in place to the national competent authority[(c)]**

The laboratory that identifies the positive sample has the obligation to notify the positive result through the RASFF to the regional and central authority, and the regional authority will notify the food business operator.

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

In 2013, 27 strains of Salmonella spp. were isolated, from which: 13 feed material of land animal origin, 10 compound feedingstuffs for poultry - laying hens, 6 compound feedingstuffs for pig.

In 2014, 22 strains of Salmonella spp. were isolated, from which: 14 feed material of land animal origin, 6 compound feedingstuffs for poultry - laying hens, 2 feed material of cereal grain origin.

In 2015, 8 strains of Salmonella spp. were isolated, from which: 4 feed material of land animal origin, 1 compound feedingstuffs for pig and 3 feed material of cereal grain origin.

In 2016, 17 strains of Salmonella spp. were isolated in feed, from which: 6 feed material of cereal grain origin, 3 feed material of land animal origin, 5 compound feedingstuffs for poultry and 3 strains in compound feedingstuffs for pig and.

In 2017, 18 strains of Salmonella spp. were isolated in feed, from which: 11 feed material of land animal origin, 3 compound feedingstuffs for poultry, 2 feed material of cereal grain origin

and 2 strains pet food - dog snacks. In 2018, 9 strains of Salmonella spp. were isolated in feed, from which: 1 compound feedingstuffs for pig, 7 compound feedingstuffs for poultry and 1 strain in other feed category.

**5. Additional information**

Comparison of the Salmonella serotypes found in animals, feedingstuffs, foodstuffs and human helps to suggest possible sources of infection in the food chain.

*** For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

## 25. Description of Monitoring/Surveillance/Control programmes system*: Campylobacter spp. in broiler carcases - food sample - neck skin

**1. Monitoring/Surveillance/Control programmes system[(a)]**

The Romanian Surveillance Program is a national program, published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority (N.S.V.F.S.A.), yearly updated and the susceptibility testing of Campylobacter spp. from broilers is a part of the program. The samples for monitoring and testing of Campylobacter spp. are compulsory taken by the official vets acting at slaughterhouses. The *control program* for Campylobacter spp., included in Romanian National Surveillance Program, approved by Order of the N.S.V.F.S.A. no 35/2016 (with subsequent amendments and completions) and in accordance with European Union regulations is carried out according to Reg CE2073/2005.

Sampling of broilers' carcasses were taken from slaughterhouses after chilling stage, as hygiene criteria of the technological process. Sampling rules for poultry carcasses were according to Reg CE 2073/2005.

Methods of sampling (sampling techniques):

*At slaughterhouse* - According to the provisions of Regulation 2005/2073/EC, with subsequent amendments and completions, for the Campylobacter spp. analyses, a minimum of 20 carcass were sampled at random during each sampling session and after chilling. A piece of approximately 10 g from neck skin was obtained from each 4 carcases obtained final 5 samples x 10 g to be tested for Campylobacter.

*Diagnostic/analytical methods* used for enumeration of Campylobacter is microbiological method: EN ISO 10272-2 – Horizontal method for detection and enumeration Campylobacter spp. Part 2. Colony count technique. Species identification was performed by molecular techniques.

**2. Measures in place[(b)]**

Samples exceeded 1000 ufc/g are considered positive and are notificated by RASFF to all levels (central, regional and local).

**3. Notification system in place to the national competent authority[(c)]**

The laboratory that identifies the positive sample has the obligation to notify the positive result through the RASFF to the regional and central authority, and the regional authority will notify the food business operator.

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

In 2015 were taken a total number of 43 samples of meat from broilers, in own check, in order to detect Campylobacter spp., from which 5 were positive. In 2015, voluntary samples usually taken for diagnostic purposes (HACCP and own checks). During the years 2016-2017, no samples were analised in order to detect Campylobacter spp (it did not run a national program for monitoring).

In 2018, according provisions of Regulation 2005/2073/EC, were taken a total number of 1830 samples of meat from broilers (neck skin) in order to detect Campylobacter spp., from which, 81 of these had quantify loads of Campylobacter > 1000 cfu/g. In the 81 strains tested for the identification of the Campylobacter species have been identified 54 species of C. jejuni and 30 species of C. coli. In 3 of them both species were identified**.**

**5. Additional information**

**There are no data to provide**

*** For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing,

serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

**27. Description of Monitoring/Surveillance/Control programmes system*:**

**26. Description of Monitoring/Surveillance/Control programmes system*: Listeriosis organ/ tissues, abortion material, milk,**

**1. Monitoring/Surveillance/Control programmes system[(a)]**

The surveillance is made according with the Order of the President of the National Sanitary Veterinary and Food Safety Authority no.35/2016. Investigations are initiated by the owners of the animals. Testing is performed on owner request and on clinical suspicion.Passive surveillance is performed in case of abortions, stillbirth and other reproductive symptoms.

**2. Measures in place[(b)]**

**3. Notification system in place to the national competent authority[(c)]**

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

**5. Additional information**

* **For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

**27. Description of Monitoring/Surveillance/Control programmes system*:**

**Listeria monocytogenes in food sample - all foodstuffs**

**1. Monitoring/Surveillance/Control programmes system[(a)]**

The Romanian Surveillance Program is a national program, published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority (N.S.V.F.S.A.), yearly updated and the susceptibility testing of Listeria monocytogenes from foodstuffs is a part of the program. The samples for monitoring and testing of Listeria monocytogenes are taken by the the official veterinarians from County Sanitary Veterinary and Food Safety Directorates (C.S.V.F.S.D.) from production plant and from retail, as a *food safety criteria*.

The *type of specimen* taken according to the stage of sampling is:

- At the production plant - Ready-to-eat food before placed on the market.
- At the reatil - Ready-to-eat food placed on the market during their shelf-life.

From *retail* the samples of food for monitoring and testing of Listeria monocytogenes are taken by the official vets also in any case of: consumer complaints, suspicions or food borne outbreaks.

The sampling designs were according to the provisions of the Romanian National Surveillance, which is according with the provisions of Regulation 2005/2073/EC, with subsequent amendments and completions.

*Analytical methods* used from testing of Listeria monocytogenes are microbiological methods: Microbiological method: EN ISO 11290-1 –for detection or Microbiological method: EN ISO 11290-2 - for enumeration.

*Definition of positive finding* - the food sample are considered to be positive when Listeria monocytogenes is isolated by the microbiological method.

**2. Measures in place[(b)]**

A positive laboratory finding of Listeria monocytogenes is followed by a notification by RASFF to all levels (central, regional and local). Then all the food chain is controlled in order to identify the origin of the contamination, if it is possible. The contaminated products are traced back and are withdrawn from human consumption.

**3. Notification system in place to the national competent authority[(c)]**

The laboratory that identifies the positive sample has the obligation to notify the positive result through the RASFF (Rapid Alert System for Food and Feed) to the regional and central authority, and the regional authority will notify the food business operator.

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and**

**sources of infection[(e)]**

In 2013, 47 strains of Listeria monocytogenes were isolated, of which 1 strains were isolated from milk and dairy products (cheeses) and 45 strains were isolated from other foods (fresh meat, meat products, meat preparation, fish and fishery products, other processed food products and prepared dishes).

In 2014, 41 strains of Listeria monocytogenes were isolated, of which 4 strains were isolated from milk and dairy products (cheeses and milk) and 37 strains were isolated from other foods (fresh meat, meat products, meat preparation, fish and fishery products, other processed food products and prepared dishes).

In 2015, 27 strains of Listeria monocytogenes were isolated, of which 2 strains were isolated from milk and dairy products (cheeses and milk) and 25 strains were isolated from other foods (fresh meat, meat products, meat preparation, fish and fishery products, other processed food).

In 2016, 16 strains of Listeria monocytogenes were isolated, of which 2 strains were isolated from dairy products and 14 strains were isolated from other foods (meat preparation, meat products, prepared dishes, bakery products and snails).

In 2017, 33 strains of Listeria monocytogenes were isolated, of which *1 strain* was isolated from milk and dairy products (cheeses), *31 strains* were isolated from other foods (fresh meat, minced meat, meat preparation, meat products, prepared dishes, bakery products and snails) and *1 strain* of them was isolated in feedingstuffs (silo forage). In 2017, it can be observed an increase trend of Listeria monocytogenes positive cases in Romania compared with the period 2015-2016.

In 2018, 193 strains of Listeria monocytogenes were isolated, of which 12 *strain* was isolated from milk and dairy products (cheeses), *113 strains* were isolated from other foods (fresh meat, minced meat, meat products, prepared dishes, processed fishery products and snails) and *68 strains* of them was isolated in frozen vegetables Samples of frozen vegetables have been sampled as a result of Listeria monocytogenes outbreaks in several countries.

**5. Additional information**

There are no data to provide

*** For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

**(**e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

## 28. Description of Monitoring/Surveillance/Control programmes system*:

## Verotoxigenic E. coli (VTEC) in food sample

### 1. Monitoring/Surveillance/Control programmes system[(a)]

The Romanian Surveillance Program is a national program, published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority (N.S.V.F.S.A.), yearly updated and the susceptibility testing of *Escherichia coli*, including Verotoxigenic *E. coli (VTEC)*, from foodstuffs is a part of the program.

The samples for monitoring and testing of *Verotoxigenic E. coli* are taken by the the official veterinarians from County Sanitary Veterinary and Food Safety Directorates (C.S.V.F.S.D.) from production plant and from retail, as a *food safety criteria*.

The sampling designs were according to the provisions of the Romanian National Surveillance, which is according with the provisions of Regulation 2005/2073/EC, with subsequent amendments and completions.

*Analytical method used is:* ISO/TS 13136:2012 - Microbiology of food and animal feed -Real-time polymerase chain reaction (PCR) - based method for the detection of food-borne pathogens - Horizontal method for the detection of Shiga toxin-producing Escherichia coli (STEC) and the determination of O157, O111, O26, O103, O104 and O145 serogroups (taking into account the most recent adaptation by the European Union reference laboratory for *Escherichia coli*, including the detection of STEC O104:H4).

*Definition of positive finding* - the food samples are considered to be positive when STEC has been isolated using the method specified above.

### 2. Measures in place[(b)]

A positive laboratory finding of Verotoxigenic *E. coli* (VTEC) is followed by a notification to RASFF to all levels (central, regional and local). Then all the food chain is controlled in order to identify the origin of the contamination, if it is possible. The contaminated products are traced back and are withdrawn from human consumption.

### 3. Notification system in place to the national competent authority[(c)]

The laboratory that identifies the positive sample has the obligation to notify the positive result through the RASFF (Rapid Alert System for Food and Feed) to the regional and central

authority, and the regional authority will notify the food business operator**.**

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

In 2012, under a national program for monitoring, were tested 446 samples, which from: 203 was carcase swabs, 121 bovine minced meat, 85 mixet meat- meat preparation - from bovine and sheep , 37 mixet meat- minced meat - from bovine and sheep. There were no positive samples for Escherichia coli STEC.

In the period 2013-2015 no samples analised for monitoring *Escherichia coli* VTEC (it did not run a national program for monitoring).

In 2016, were tested 1793 which from 74 samples were positive for *Escherichia coli* STEC (STEC strain isolated) and 287 of them had a "presumptive presence STEC" according to ISO / TS 13136:2012. From all the analyzed samples, 1479 were tested in the frame of the national program for monitoring STEC issued by N.S.V.F.S.A.; 155 samples in the haemolytic uraemic syndrome Romanian outbreak, 23 sprouts samples in national surveillance program and 136 were HACCP and own check samples.

In 2017, were tested 154 samples of which 5 were pozitive and 2 were presumptive according ISO/TS 13136:2012. In 2018, were tested 64 samples of which 2 were pozitive according ISO/TS 13136:2012.

**5. Additional information**

In 2016, according to the provisions of the Romanian National Surveillance Program approved by Order of the President of the National Sanitary Veterinary and Food Safety Authority and the Regulation 2005/2073/EC, with subsequent amendments and completions, sprouts were included to surveillence for Shiga toxin producing E. coli (STEC) O157, O26, O111, O103, O145 and O104:H4. Subsequently to the O26 STEC haemolytic uraemic syndrome Romanian outbreak, the Romanian National Sanitary Veterinary and Food Safety Authority (N.S.V.F.S.A.) issued a national monitoring program (decision no. 6241/2016) for STEC detection from meat and meat products and milk and milk products.

*** For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

**29. Description of Monitoring/Surveillance/Control programmes system*:**

**Histamine in Fishery products - food sample**

**1. Monitoring/Surveillance/Control programmes system[(a)]**

The Romanian Surveillance Program is a national program, published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority (N.S.V.F.S.A.), yearly updated and detection of Histamine from foodstuffs is a part of the program.

The samples for monitoring and testing of Histamine are taken by the the official veterinarians from County Sanitary Veterinary and Food Safety Directorates (C.S.V.F.S.D.) from retail, as a *food safety criterion* and also in case of consumer complaints, suspicions or food borne outbreaks.

The sampling designs were according to the provisions of the Romanian National Surveillance, which is according with the provisions of Regulation 2005/2073/EC, with subsequent amendments and completions. Taken of sampling is at the following fish species: Scombridae, Clupeidae, Engraulidae, Coryfenidae, Pomatomidae, Scombresosidae from fishery products which are not enzyme matured in brine and fishery products which have undergone enzyme maturation treatment in brine.

*Analytical method used is:* High-performance liquid chromatography (HPLC).

*Definition of positive finding* - the food sample are considered to be positive when that contains histamine at a concentration with more than 100 mg/kg (category 1), more than 200 mg/kg (category 2) or more than 400 mg/kg (category 3).

**2. Measures in place[(b)]**

A positive laboratory finding of Histamine is followed by a notification by RASFF to all levels (central, regional and local). Then all the food chain is controlled in order to identify the origin of the contamination, if it is possible. The contaminated products are traced back and are withdrawn from human consumption.

**3. Notification system in place to the national competent authority[(c)]**

The laboratory that identifies the positive sample has the obligation to notify the positive result through the RASFF (Rapid Alert System for Food and Feed) to the regional and central

authority, and the regional authority will notify the food business operator.

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

In 2013, there were analyzed 170 samples from fish species and all samples had values less than 100 mg/kg (no positive samples were detected).

In 2014, there were analyzed 124 samples from fish species and no positive samples were detected.

In 2015, there were analyzed 116 samples from fish species and no positive samples were detected.
In 2016, there were analyzed 102 samples from fish species and no positive samples were detected.

In 2017, there were analyzed 59 samples from fish species and no positive samples were detected.
In 2018, there were analyzed 1532 samples from fish species according with the provisions of Regulation 2005/2073/EC, of which 8 samples were with noncompliance results (3 from category 1, more than 200 mg/kg and 5 from category, more than 400 mg/kg).

**5. Additional information**

**There are no data to provide**

*** For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

**30.Description of Monitoring/Surveillance/Control programmes system*:**

**Staphylococcal enterotoxins - food sample**

1. Monitoring/Surveillance/Control programmes system[(a)]

The Romanian Surveillance Program is a national program, published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority (N.S.V.F.S.A.), yearly updated and detection of Staphylococcal enterotoxins from foodstuffs is a part of this program. The samples for monitoring and testing of Staphylococcal enterotoxins are taken by the the official veterinarians from County Sanitary Veterinary and Food Safety Directorates (C.S.V.F.S.D.) from retail, as a *food safety criterion* and also in case of consumer complaints, suspicions or food borne outbreaks.

The sampling designs were according to the provisions of the Romanian National Surveillance, which is according with the provisions of Regulation 2005/2073/EC, with subsequent amendments and completions..

*Analytical method used* is European method from EURL-CPS and detection of staphylococcal enterotoxin encoding genes performed by Multiplex PCR.

*Definition of positive finding* - the food sample are considered to be positive when staphylococcal enterotoxins have been detected.

2. Measures in place[(b)]

A positive laboratory finding of Staphylococcal enterotoxins is followed by a notification by RASFF to all levels (central, regional and local). Then all the food chain is controlled in order to identify the origin of the contamination, if it is possible. The contaminated products are traced back and are withdrawn from human consumption.

3. Notification system in place to the national competent authority[(c)]

The laboratory that identifies the positive sample has the obligation to notify the positive result through the RASFF (Rapid Alert System for Food and Feed) to the regional and central authority, and the regional authority will notify the food business operator.

4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]

In 2013 were analyzed 411 samples from which 1 sample was positive.

In 2014 were analyzed 215 samples, from which 2 were positive (Staphylococcal enterotoxins D).

In 2015 were analyzed 79 samples and neither of them were found positive. In 2016 were analyzed 389 samples and 1 sample of them were found positive.

In 2017 were analyzed 247 samples and and neither of them were found positive.It can be observed declining of samples analyzed in the 2017 year in Romania compared with the year 2016.

In 2018 were analyzed 1206 samples, from which 4 were positive. Detection of staphylococcal enterotoxin encoding genes was performed by Multiplex PCR (positive for types C, H and I).

**5. Additional information**

There are no data to provide

*** For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

**31. Description of Monitoring/Surveillance/Control programmes system*:**

**Norovirus and Hepatitis A virus in food sample**

**1. Monitoring/Surveillance/Control programmes system[(a)]**

Since 2016, according to the provisions of the Romanian National Surveillance Program approved by Order of the President of the National Sanitary Veterinary and Food Safety Authority, fruits were included to surveillence for Norovirus and Hepatitis A virus. The samples for monitoring and testing of Norovirus and Hepatitis A virus are taken by the the official veterinarians from County Sanitary Veterinary and Food Safety Directorates (C.S.V.F.S.D.) from production plant and from retail, as a *food safety criteria*.

*Analytical method used is:* The laboratory procedures detects simultaneusly both norovirus and hepatitis A virus from a sample, according ISO/TS 15216-2:2013.

*Definition of positive finding* - the food sample are considered to be positive when Norovirus and Hepatitis A virus has been isolated using a method specified above.

**2. Measures in place[(b)]**

A positive laboratory finding of Norovirus and Hepatitis A virus is followed by a notification by RASFF to all levels (central, regional and local). Then all the food chain is controlled in order to identify the origin of the contamination, if it is possible. The contaminated products are traced back and are withdrawn from human consumption.

**3. Notification system in place to the national competent authority[(c)]**

The laboratory that identifies the positive sample has the obligation to notify the positive result through the RASFF (Rapid Alert System for Food and Feed) to the regional and central authority, and the regional authority will notify the food business operator.

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

In 2016, under a national program for Surveillance, 16 fruits samples were tested from Norovirus

and Hepatitis A virus and none of them were positive.

In 2017, under a national program for Surveillance, 23 fruits samples were tested from Norovirus

and Hepatitis A virus and none of them were positive.

In 2018, under a national program for Surveillance, 20 fruits samples were tested from Norovirus

and Hepatitis A virus and none of them were positive.

**5. Additional information**

**There are no data to provide**

*** For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

**32. Description of Monitoring/Surveillance/Control programmes system*:**

**Cronobacter in food sample**

**1. Monitoring/Surveillance/Control programmes system[(a)]**

The Romanian Surveillance Program is a national program, published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority (N.S.V.F.S.A.), yearly updated and detection of *Cronobacter* from infant formula is a part of this program.The samples for monitoring and testing of Cronobacter are taken by the the official veterinarians from County Sanitary Veterinary and Food Safety Directorates (C.S.V.F.S.D.) from retail, as a *food safety criteria*.

Methods of sampling (sampling techniques):

*At retail* - According to the provisions of Regulation 2005/2073/EC, with subsequent amendments and completions, for the Cronobacter spp. Analyses.

*Diagnostic/analytical methods* used for detection of Cronobacter spp. is microbiological method: ISO 22964 – Horizontal method for detection of Cronobacter spp.

*Definition of positive finding* - the food sample are considered to be positive when Cronobacter spp. has been detected using microbiological method.

**2. Measures in place[(b)]**

A positive laboratory finding of Cronobacter is followed by a notification by RASFF to all levels (central, regional and local). Then all the food chain is controlled in order to identify the origin of the contamination, if it is possible. The contaminated products are traced back and are withdrawn from human consumption.

**3. Notification system in place to the national competent authority[(c)]**

The laboratory that identifies the positive sample has the obligation to notify the positive result through the RASFF (Rapid Alert System for Food and Feed) to the regional and central authority, and the regional authority will notify the food business operator.

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

In the years 2012-2015 no samples were analysed for Cronobacter.

In 2018, 18 infant formula samples were tested from Cronobacter, and none of them were positive.

**5. Additional information**

There are no data to provide

*** For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

## 33. Description of Monitoring/Surveillance/Control programmes system*: Trichinella spp. in pigs (organ/tissue) - food sample

**1. Monitoring/Surveillance/Control programmes system[(a)]**

The sampling designs were according to the provisions of the Romanian National Surveillance Programme published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority, yearly updated which is according with the provisions of Regulation 2005/2075/EC (repealed by Regulation 2015/1375/EC), sampling is performed for all pigs slaughtered, intended to human consumption, in order to detect Trichinella spp.

The *type of specimen* taken is diaphragm pillars and In the absence of diaphragm pillars, the follwing specimens are taken: the rib part or the breastbone part of the diaphragm, the jaw muscles, tongue or abdominal muscles.

*Diagnostic/analytical methods* used for detection Trichinella is artificial digestion methods on individual samples and/or on pooled samples.

| |
|---|
| *Definition of positive finding* – animal in which Trichinella spp. larvae have been detected. |
| **2. Measures in place[(b)]** |
| Sampling is compulsory for all pigs slaughtered in order to detect Trichinella spp. and to avoid human trichinelosis**.** A positive laboratory finding of Trichinella spp. it is followed by a notification to RASFF to all levels (central, regional and local). Pig meat infested with Trichinella spp. is withdrawn from human consumption and sent to the rendering establishments, in order to be denatured |
| **3. Notification system in place to the national competent authority[(c)]** |
| The laboratory that identifies the positive sample has the obligation to notify the positive result through the RASFF (Rapid Alert System for Food and Feed) to the regional and central authority, and the regional authority will notify the food business operator. |
| **4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]** |
| During the year 2011, in Romania were detected a total number of 264 positive cases of Trichinella spp. in pigs. During the year 2012, in Romania were detected a total number of 171 positive cases of Trichinella spp. in pigs. In 2012 for pigs raised in backyards was observed a decrease of percent of positive cases, with 33,97%, compared with 2011.<br><br>In 2013, 193 positive cases in fattening pigs from backyards were detected and was observed an increase of percent of positive cases, with 12,90 % compared with 2013.<br><br>In 2014, 141 positive cases in fattening pigs from backyards were detected.<br><br>In the year 2015, 87 positive cases in fattening pigs from backyards were detected . In the period 2014-2015, it can be observed declining trend of positive cases in Romania compared with 2013.<br><br>In 2016, the 151 cases of trichinella detected are related to 120 positive cases registered in fattening pigs raised in backyards (meat from pig not raised under controlled housing conditions) and 31 positive cases to fattening pigs raised in farms (meat from pig not raised under recognised controlled housing conditions).The cases of Trichinella detected are related to the positive cases registered in meat and products thereof<br><br>In 2017, the 120 cases of trichinella detected are related to fattening pigs raised in backyards (meat from pig not raised under controlled housing conditions). In 2017, in one of the samples two different species were identified (coinfection with T. Pseudospiralis and T. Britovi).<br><br>In 2018, the 134 cases of trichinella detected were registered to fattening pigs raised in backyards (meat from pig not raised under controlled housing conditions). |

**5. Additional information**

All positive samples (larvae detected in meat from pigs), were sent for identify the species of Trichinella, some of them are identified by the N.R.L. for Trichinella, which is in Institute of Hygiene and Veterinary Public Health and same of the larvae were sent to EURL Parasites Roma.

Comparison of the Trichinella species found in pigs, meat from pig and human caseshelps to suggest possible sources of infection in the food chain.

* **For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant. If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

**34.Description of Monitoring/Surveillance/Control programmes system*: TUBERCULOSIS, MYCOBACTERIAL DISEASES - Cattle (bovine animals) - Farm - animal sample - Surveillance – Official sampling – Census**

**1. Monitoring/Surveillance/Control programmes system[(a)]**

Status as officially free of bovine tuberculosis during the reporting year
The entire country free
Romania is not official free for Bovine tuberculosis.
***Monitoring system***
*Sampling strategy*
The program is applied in all teritory of Romania, in non-profesional and commercial farms to all bovines and buffaloes aged over 6 weeks
*Frequency of the sampling*
Annualy
*Type of specimen taken*
From all reagent animals, slaughtered for clarification diagnosis of tuberculosis, samples must be taken individual for laboratory examinations, as follows:
a) lymph nodes of the head - retrofaringian left and right, left and straight mandibular,left and right parotidian,
b) left and right tracheobronchic lymph nodes, anterior and anterior mediastinal lymph nodes back; c) internal and external iliac lymph nodes, upper and lower retromammery lymph nodes inferior and popliteal;
d) portions of tissues and organs: pleura, pulmonus, liver, spleen, kidneys, genital organs, mammary gland. From positive animals to tuberculin or immunological tests which at post-mortem inspection does not show any lesions, the following groups of lymphocytes will be harvested:

a) submaxillar, retrofaringian, bronchial, mediastinal;
b) eventually the mesenteric ones, if they are enlarged in volume, portals and retromammary.
*Diagnostic/analytical methods used*
The diagnostic method used is the skin test, more precisely intradermal compartive test (TCS) as described in Annex B of Council Directive 64/432/EEC. Laboratory diagnosis is confirmed by morphopathology + direct microscopic examination + biological test on guinea pigs.
Additionally: cultural examination + phenotypic typing + genetic typing

**2. Measures in place[(b)]**

-

**3. Notification system in place to the national competent authority[(c)]**
Tuberculosis is a compulsory notifiable disease on the entire territory of Romania. The notification is carried out according to the national Order 77/2005 for the approval of the Sanitary Veterinary Norm regarding the notification of animal diseases, with all subsequent amendments, published in the Official Journal of Romania, no. 964/31 October 2005; this order represents the official transposition of Council Directive 82/894/EEC on the notification of animal diseases within the European Community and Order no. 79 of 18 September 2008 for approval of sanitary veterinary internal and officially declared the notification of infectious animal diseases as amended;.

**4. Results of investigations and national evaluation of the situation, the trends [(d)] and sources of infection[(e)]**

In 2013, was detected 181 positive bovines.
In the year 2014, 45 positive cases in bovine from backyards were detected.
In 2015, the 241 cases of bovine tuberculosis were detected.
Following tests in 2016, positive cattle were diagnosed with M. bovis (18 bovines) and M. caprae (190 bovines).
Following tests in 2017, positive cattle were diagnosed with M. bovis (64 bovines) and M. caprae (317 bovines).
Following tests in 2018, positive cattle were diagnosed with M. bovis (9 bovines) and M. caprae (253 bovines).

**5. Additional information**

All positive samples (larvae detected in meat from pigs), were sent for identify the species of Trichinella, some of them are identified by the N.R.L. for Trichinella, which is in Institute of Hygiene and Veterinary Public Health and same of the larvae were sent to EURL Parasites Roma.
Comparison of the Trichinella species found in pigs, meat from pig and human caseshelps to suggest possible sources of infection in the food chain.

* **For all combinations of zoonotic agents and matrix (Food, Feed and Animals) for 'Prevalence' and 'Disease Status': one text form reported per each combination of matrix/zoonoses or zoonotic agent**

(a): Sampling scheme (sampling strategy, frequency of the sampling, type of specimen taken, methods of sampling (description of sampling techniques) + testing scheme (case definition, diagnostic/analytical methods used, diagnostic flow (parallel testing, serial testing) to assign and define cases. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(b): The control program/strategies in place, including vaccination if relevant.If applicable a description of how eradication measures are/were implemented, measures in case of the positive findings or single cases; any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation, if applicable. If programme approved by the EC, please provide link to the specific programme in the Commission`s website.

(c): Mandatory: Yes/No.

(d): Minimum five years.

(e): Relevance of the findings in animals to findings in foodstuffs and for human cases (as a source of infection).

## 35. Food-borne Outbreaks

### 1. System in place for identification, epidemiological investigations and reporting of food-borne outbreaks

The Romanian Surveillance Program is a national program, published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority (N.S.V.F.S.A.), and surveillance of food borne outbreaks is a part of the program.

The samples are taken by the the official veterinarians from County Sanitary Veterinary and Food Safety Directorates (C.S.V.F.S.D.) from retail in case of consumer complaints, suspicions or food borne outbreaks**.** The municipal public health authorities are responsible for detecting, preventing diseases related to food and water and for notifying to the other authorities involved. Ill perssons and the overall epidemiological investigation are the responsibilities of the regional authorities (public health and veterinary public health authorities).

The Institute for Hygiene and Veterinary Public Health (I.H.V.P.H.), which is a public institution with legal personality, designated as national reference authority in the field of food safety, under the responsibility of N.S.V.F.S.A. collects from County Sanitary Veterinary and Food Safety Directorates (C.S.V.F.S.D.) and reports to the N.S.V.F.S.A. all food borne outbreaks data (in the field of food and feed safety).

### 2. Description of the types of outbreaks covered by the reporting

During 2017 there were 12 outbreaks , 2 episodes was weak-evidence and 10 episodes were with strong evidence, within which 425 people ill and 211 people hospitalized.

### 3. National evaluation of the reported outbreaks in the country[(a)]

In 2014 it recorded a total of 27 food borne outbreaks were reported, 6 episodes was weak-evidence and 21 episodes were with strong evidence, 379 people ill and 199 people hospitalized.
In 2015 it recorded a total of 21 outbreaks,18 episodes were with strong evidence and 3 episodes were weak-evidence, 397 people ill and 270 people hospitalized.
In 2016 it recorded a total of 19 outbreaks ,13 episodes were with strong evidence and 6 episodes was weak-evidence, 312 people ill and 220 people hospitalized.

In 2016 it recorded a total of 19 outbreaks ,13 episodes were with strong evidence and 6 episodes was weak-evidence, 312 people ill and 220 people hospitalized.

In 2017 there were recorded 12 outbreaks, 2 episodes was weak-evidence and 10 episodes were with strong evidence, within which 425 people ill and 211 people hospitalized.

In 2017, Staphylococus was the most frequently identified agent in food borne disease outbreaks (6 episodes with 167 human cases and 71 people hospitalised); followed by

Salmonella as the agent identified (4 episodes with 147 human cases and 51 people hospitalised) and Trichinella (2 episodes with 111 human cases and 89 people hospitalised). From all 12 outbreaks recorded in 2017, 4 episodes were mixed outbreaks and had agents as well Staphylococus and Escherichia coli. Also 1 episode was with unknown vehicle and the case was classified based on clinical and epidemiological data according to surveillance methodology. Most of the outbreaks, were reported to be linked to the public consumption, 11 general FBOType, and and only 1 by huosehold FBO Type (private type). The types of foods involved in food borne disease outbreaks reported were : buffet meals, mixed food (prepared dishes), meat and products thereof, cheese, sweets and other food. The causative agents, in the incriminated foodstuff, were confirmed in laboratory and also based on epidemiological investigation or epidemiological suspected.The most important factors contributing to food borne disease outbreaks reported were unsatisfactory hygiene conditions and carriers, cross-contamination and infected food handler.

In 2018 there were recorded 29 outbreaks, 5 episodes was weak-evidence and 24 episodes were with strong evidence, within which 738 people ill and 324 people hospitalized. *In 2018*, Salmonella (6 episodes with 494 human cases and 181 people hospitalised) and Trichinella (6 episodes with 64 human cases and people hospitalised) were the most frequently identified agent in food borne disease outbreaks; followed by Staphylococus as the agent identified (4 episodes with 44 human cases and 39 people hospitalised) and E.coli (3 episodes).

2 outbreaks of special interest were recorded of these, with *Clostridium botulinum* as etiological agent with 4 human cases and 3 people hospitalised, where *the death of one of them* was recorded.

5 outbreaks were *unknown* etiological agent (unidentified in food or human cases), with 83 human cases and 18 people hospitalised and the cases were classified based on clinical and epidemiological data according to surveillance methodology.

From all 29 outbreaks recorded in 2018, 4 episodes were mixed outbreaks. Of these, 1 outbreak was of interest, with 4 infectious agents (Listeria monocytogenes, Salmonella infantis, Staphylococcus aureus and Escherichia coli).

Most of the outbreaks, were reported to be linked to the public consumption, 24 general FBOType, and and 5 by huosehold FBO Type (private type). The types of foods involved in food borne disease outbreaks reported were: mixed food (prepared dishes, meat and products thereof, cheese) buffet meals, other food and water. The causative agents, in the incriminated foodstuff, were confirmed in laboratory and also based on epidemiological investigation or epidemiological suspected.The most important factors contributing to food borne disease outbreaks reported were unsatisfactory hygiene conditions and carriers, infected food handler**,** Inadequate heat treatment and cross-contamination. *Compared to 2017, in 2018 was observed a 142% increase in the number of outbreaks reported*

**4. Descriptions of single outbreaks of special interest**

No unique outbreaks of special interest were recorded.

**5. Control measures or other actions taken to improve the situation**

According with the provisions Romanian National Programme for Surveillance of Zoonoses, Rapid Alert System for Food and Feed and the National Sanitary Veterinary and Food Safety Authority Order no. 34/2006, which transposed Directive 2003/99/EC.

In addition there is a collaboration protocol between the Ministry of Health and the National Sanitary Veterinary and Food Safety Authority for the control work on the risks presented by food for public health and consumer protection.

*In 2018 the data collection method was changed, the outbreaks being collected in the form of an excel file, immediately after the end of the epidemiological investigation unlike in 2017 when the collection was in an on-line application, the frequency being quarterly. We cannot suggest that this increase is related to the change in the reporting system used in 2018 compared to 2017. We must monitor comparative developments at least and the following year in order to provide evidence on how to improve the reporting system.*

**6. Any specific action decided in the Member State or suggested for the European Union as a whole on the basis of the recent/current situation**

The Institute for Hygiene and Veterinary Public Health do not have any data provided

**7. Additional information**

There are no data to provide

(a): Trends in numbers of outbreaks and numbers of human cases involved, relevance of the different causative agents, food categories and the agent/food category combinations, relevance of the different type of places of food production and preparation in outbreaks, evaluation of the severity of the human cases.

**36. Institutions and laboratories involved in antimicrobial resistance monitoring and reporting**

Institute for Diagnosis and Animal Health – central animal health diagnostic institute, NRL for Antimicrobial resistance - for monitoring and reporting
Sanitary Veterinary and Food Safety Directorate Counties: Alba, Brăila, Buzău, Dâmbovița, Giurgiu, Ialomița, Iași, Mureș, Prahova and Satu Mare – for monitoring

The monitoring of antimicrobial resistance is made according with the Order of the President of

the National Sanitary Veterinary and Food Safety Authority (N.S.V.F.S.A.) yearly updated, which is according to the provisions of Commission Decision 2013/652/EU on the monitoring and reporting of antimicrobial resistance in zoonotic and commensal bacteria.The samples for monitoring of antimicrobial resistance (according to allocation under the National Sampling Plan) are taken by the official veterinarians from County Sanitary Veterinary and Food Safety Directorates (C.S.V.F.S.D.) and all the strains isolated in foodstuffs derived from products of animal origin are tested for the antimicrobial resistance at the National Reference Laboratory (N.R.L. - AR). The NRL - AR is organized within the Institute for Hygiene and Veterinary Public Health (I.H.V.P.H.), which is a public institution with legal personality, designated as national reference authority in the field of food safety, under the responsibility of N.S.V.F.S.A.

The I.H.V.P.H. collects from regional laboratories (Sanitary Veterinary and for Food Safety Laboratories) and reports to the N.S.V.F.S.A. all antimicrobial resistance data.

Short description of the institutions and laboratories involved in data collection and reporting

## 36. General Antimicrobial Resistance Evaluation

**1. Situation and epidemiological evolution (trends and sources) regarding AMR to critically important antimicrobials[(a)] (CIAs) over time until recent situation**

Starting with 2015 in Romania runs the program for the monitoring of antimicrobial resistance for for each combination of bacterial species and food categories, every two years, according to the provisions of Commission Decision 2013/652/EU. The antimicrobials included in monitoring are: Ampicillin, Azithromycin, Cefotaxime, Ceftazidime, Chloramphenicol, Ciprofloxacin, Colistin, Gentamicin, Meropenem, Nalidixic acid, Sulfamethoxazole, Tetracycline, Tigecycline, Trimethoprim (first panel) and Cefepime, Cefoxitin, Ceftazidime, Ceftazidime + clavulanic acid, Cefotaxime, Cefotaxime + clavulanic acid, Ertapenem, Imipenem, Meropenem, Temocillln (Second panel), and are tested by the micro-dilution method according to the method described by the EUCAST and CLSI, accepted as ISO 20776-1:2006.

The cut-off values used in testing are those listed in Decision 2013/652/UE, and yearly updated and provided by EURL-AR and EFSA in the Manual for reporting on antimicrobial resistance (listed in Panel of antimicrobial substances to be included in AMR monitoring, interpretative thresholds for interpreting resistance and concentration ranges).

In 2015, were tested for identification *Salmonella spp*., 399 samples meat from pig-carcasse (carcasses swabs) from which a number of 23 were positive. Non one of them, was not resistant for cephalosporins. In 2015, were tested for identification *Escherichia coli,* 399 sample fresh meat from pig from which 63 were positive and 244 sample fresh meat from bovine from which a number of 28 were positive.

In 2016, were tested for identification *Salmonella spp*.,1871 samples neck skin of broilers

(Gallus gallus - carcase), from which a number of 82 were positive. Non one of them was not resistant for cephalosporins. In 2016, were tested for identification *Escherichia coli,* 315 samples fresh meat from broilers (Gallus gallus)**,** from which a number of 190 were positive. *1 one of them was resistant for carbapenems*.

In 2017, were tested for identification *Salmonella spp*., 300 samples meat from pig-carcasse (carcasses swabs), from which a number of 4 were positive. Non one of them, was not resistant for cephalosporins. In 2017, were tested for identification *Escherichia coli,* 298 sample fresh meat from pig*,* from which 44 were positive and 146 sample fresh meat from bovine from which a number of 5 were positive.

In 2018, were tested for identification *Salmonella spp*., 3138 samples neck skin of broilers (Gallus gallus - carcase) from which 35 were positive and 125 sample neck skin of turkey (carcase) from which a number of 13 were positive. Non one of them, was not resistant for cephalosporins. In 2018, were tested for identification *Escherichia coli***,** 297 samples fresh meat from broilers (Gallus gallus) from which a number of 99 were positive.

**2. Public health relevance of the findings on food-borne AMR in animals and foodstuffs**

The Institute for Hygiene and Veterinary Public Health do not have any data provided

**3. Recent actions taken to control AMR in food producing animals and food**

Starting with 2016, the National Sanitary Veterinary and Food Safety Authority (N.S.V.F.S.A.) has developed a national strategy to combat antimicrobial resistance in veterinary medicine (National Guide), antibiotic resistance being a public health security issue. The NATIONAL GUIDE refers to the prudent use of antimicrobial substances in animals and, in particular, to limiting the development of antimicrobial resistance and its purpose is to provide veterinarians, farmers, authorities the veterinary industry, the drug industry, associations and academia, practical guidance on the prudent use of antimicrobials, in particular antibiotics, in veterinary medicine.

**4. Any specific action decided in the Member State or suggestions to the European Union for actions to be taken against food-borne AMR threat**

The Institute for Hygiene and Veterinary Public Health do not have any data provided

**5. Additional information**

There are no data to provide

(a): The CIAs depends on the bacterial species considered and the harmonised set of substances tested within the framework of the harmonised monitoring:

- For *Campylobacter* spp., macrolides (erythromycin) and fluoroquinolones (ciprofloxacin);

- For *Salmonella* and *E. coli*, 3rd and 4th generation cephalosporins (cefotaxime) and fluoroquinolones (ciprofloxacin) and colistin (polymyxin);

**37.General Description of Antimicrobial Resistance Monitoring*;**

**Escherichia coli, non-pathogenic in meat from broilers - fresh**

**1. General description of sampling design and strategy[(a)]**

In 2018, the sampling designs were according to the Order of the Romanian National Sanitary Veterinary and Food Safety Authority President (N.S.V.F.S.A.) service note no 7145/2018 and the provisions of Commission Decision 2013/652/EU on the monitoring and reporting of antimicrobial resistance in zoonotic and commensal bacteria (Grant Decision SI2.778865/2018). For isolation of ESBL-, AmpC- and carbapenemase, **meat from broilers (Gallus gallus) - fresh** were taken from retail (according to allocation under the National Sampling Plan).The samples for monitoring of antimicrobial resistance (according to allocation under the National Sampling Plan) are taken by the official veterinarians from County Sanitary Veterinary and Food Safety Directorates (C.S.V.F.S.D.) and all the samples were tested for the antimicrobial resistance only at the National Reference Laboratory (N.R.L. - AR).

The Hygiene and Veterinary Public Health (I.H.V.P.H.) collects all the strains isolated from **meat from broilers (Gallus gallus) - fresh** taken from retail.

**2. Stratification procedure per animal population and food category**

For detection (isolation and serotyping) E. coli in **meat from broilers (Gallus gallus) - fresh**, the samples were collected from 30 counties, respectively from 60 cutting plants and 30 supermarkets. Samples were collected from regional county (County Sanitary Veterinary and Food Safety Directorate – C.S.V.F.S.D.) and analised in the Institute for Hygiene and Veterinary Public Health. Each sample had a unic number recorded in a standard form sampling**.** The isolates were serotyped in the NRL - E.coli and the antimicrobial resistance testing was performed in the NRL-AR (Institute for Hygiene and Veterinary Public Health).

**3. Randomisation procedure per animal population and food category**

Samples were collected through a random selection according to the provisions of N.S.V.F.S.A. President Order 7145/2018 on the monitoring and reporting of antimicrobial resistance in zoonotic and commensal bacteria. The distribution of planned samples from the national plan, was carried out by the official veterinarians within the county (C.S.V.F.S.D.) based on the principle of representativeness, randomized on days / weeks / months / batch / epidemiological unit / the specificity of the sampling method / the specific matrix / type of unit / activity and other criteria set out in the monitoring plan of antimicrobial resistance. Each sample had a unic

number recorded in a standard form sampling.

**4. Analytical method used for detection and confirmation[(b)]**

The method used for detection of the antimicrobial resistance is broth microdilution (ISO 20776) testing and quality control were performed according to CLSI (Clinical and Laboratory Standards Institute) documents and standards.

**5. Laboratory methodology used for detection of antimicrobial resistance[(C)]**

Laboratory protocol for isolation of ESBL-, AmpC- and carbapenemase producing E. coli from fresh meat DTU Food, biochemical confirmation provided by EURL- AR. All the antimicrobials (panel 1 and panel 2) included in monitoring, according to the Decision 2013/652/EU, were tested and the cut-off values used in testing are those provided by EURL- AR and by EFSA in the Manual for reporting on antimicrobial resistance. The antimicrobials included in monitoring were: Ampicillin, Azithromycin, Cefotaxime, Ceftazidime, Chloramphenicol, Ciprofloxacin, Colistin, Gentamicin, Meropenem, Nalidixic acid, Sulfamethoxazole, Tetracycline, Tigecycline, Trimethoprim (first panel) and Cefepime, Cefoxitin, Ceftazidime, Ceftazidime + clavulanic acid, Cefotaxime, Cefotaxime + clavulanic acid, Ertapenem, Imipenem, Meropenem, Temocillln (Second panel).

**6. Results of investigation**

In 2018, were tested for identification *Escherichia coli,* 297 samples ***fresh meat from broilers*** (Gallus gallus) from which a number of 99 were positive.

**7. Additional information**

According to the provisions of the Order of President of National Sanitary Veterinary and Food Safety Authority no.34/2006, transposing into Romanian legislation the Directive 2003/99/EC, the strains isolated in foodstuffs were tested for the antimicrobial resistance. Isolation and serotyping of strains of the E. coli from samples, was performed by the NRL and all the antimicrobial resistance data is collected only in Institute of Hygiene and Veterinary Public Health.

*** to be filled in per combination of bacterial species/matrix**

(a): Method of sampling (description of sampling technique: stage of sampling, type of sample, sampler), Frequency of sampling, Procedure of selection of isolates for susceptibility testing, Method used for collecting data.

(b): Analytical method used for detection and confirmation: according to the legislation, the protocols developed by the EURL-AR should be used and reported here. In the case of the voluntary specific monitoring on Carbapenemase-producers, the selective media used (commercial plates, 'in house' media) should be also reported here. In general, any variation with regard to the EURL-AR protocols should be stated here, number of isolates isolated per sample, in particular for *Campylobacter* spp..

(c): Antimicrobials included, Cut-off values

## 38. General Description of Antimicrobial Resistance Monitoring*;

## Salmonella spp., in broiler carcases (neck skin) - food sample

### 1. General description of sampling design and strategy[(a)]

In 2018, the sampling designs were according to the Romanian National Sanitary Veterinary and Food Safety Authority President (N.S.V.F.S.A.) service note no 7145/2018 and the provisions of Commission Decision 2013/652/EU on the monitoring and reporting of antimicrobial resistance in zoonotic and commensal bacteria (Grant Decision SI2.778865/2018). For detection and serotyping of Salmonella, **meat from broilers (Gallus gallus) – carcase (neck skin)** and **meat from turkey – carcase (neck skin)** from slaughterhouses, according Regulation 2073/2005/EC. The samples for monitoring of antimicrobial resistance (according to allocation under the National Sampling Plan) are taken by the official veterinarians from County Sanitary Veterinary and Food Safety Directorates (C.S.V.F.S.D.) and all the samples were tested for the antimicrobial resistance only at the National Reference Laboratory (N.R.L. - AR).

The Hygiene and Veterinary Public Health (I.H.V.P.H.) collects all the strains isolated from **meat from poultry** - **carcase (neck skin)** taken from slaughterhouses.

### 2. Stratification procedure per animal population and food category

For detection (isolation and serotyping) Salmonella in meat from broilers (Gallus gallus) – carcase (neck skin) and meat from turkey – carcase (neck skin), samples were collected from regional county (County Sanitary Veterinary and Food Safety Directorate – C.S.V.F.S.D.) within the Romanian National Surveillance Program, and analised in (Laboratory Sanitary Veterinary and Food Safety Directorate – L.S.V.F.S.D.) or Institute for Hygiene and Veterinary Public Health (I.H.V.P.H.).

All the isolates were serotyped in the NRL - Salmonella and the antimicrobial resistance testing was performed in the NRL-AR (I.H.V.P.H.).

### 3. Randomisation procedure per animal population and food category

Samples were collected according to the provisions of to Regulation CE 2073/2005, by the official veterinarians within the county (C.S.V.F.S.D.) based on the principle of representativeness, randomized on days / weeks / months / batch / epidemiological unit / the specificity of the sampling method / the specific matrix / type of unit / activity and other criteria set out in the monitoring plan of antimicrobial resistance.

### 4. Analytical method used for detection and confirmation[(b)]

The method used for detection of the antimicrobial resistance is broth microdilution (ISO 20776) testing and quality control were performed according to CLSI (Clinical and Laboratory

Standards Institute) documents and standards.

**5. Laboratory methodology used for detection of antimicrobial resistance(C)**

Laboratory protocol for isolation of Salmonella is DTU Food, biochemical confirmation provided by EURL- AR. All the antimicrobials (panel 1 and panel 2) included in monitoring, according to the Decision 2013/652/EU, were tested and the cut-off values used in testing are those provided by EURL- AR and by EFSA in the Manual for reporting on antimicrobial resistance. The antimicrobials included in monitoring were: Ampicillin, Azithromycin, Cefotaxime, Ceftazidime, Chloramphenicol, Ciprofloxacin, Colistin, Gentamicin, Meropenem, Nalidixic acid, Sulfamethoxazole, Tetracycline, Tigecycline, Trimethoprim (first panel) and Cefepime, Cefoxitin, Ceftazidime, Ceftazidime + clavulanic acid, Cefotaxime, Cefotaxime + clavulanic acid, Ertapenem, Imipenem, Meropenem, Temocillln (Second panel).

**6. Results of investigation**

In 2018, were tested for identification *Salmonella spp*., 3138 samples ***neck skin of broilers*** (Gallus gallus - carcase) from which 35 were positive and 125 sample neck skin of turkey (carcase) from which a number of 13 were positive. Non one of them, was not resistant for cephalosporins.

**7. Additional information**

According to the provisions of the Order of President of National Sanitary Veterinary and Food Safety Authority no.34/2006, transposing into Romanian legislation the Directive 2003/99/EC, the strains isolated in foodstuffs were tested for the antimicrobial resistance. Serotyping of strains of the **Salmonella** and the antimicrobial resistance, was performed by the NRL and all the antimicrobial resistance data is collected only in Institute of Hygiene and Veterinary Public Health.

*** to be filled in per combination of bacterial species/matrix**

(a): Method of sampling (description of sampling technique: stage of sampling, type of sample, sampler), Frequency of sampling, Procedure of selection of isolates for susceptibility testing, Method used for collecting data.

(b): Analytical method used for detection and confirmation: according to the legislation, the protocols developed by the EURL-AR should be used and reported here. In the case of the voluntary specific monitoring on Carbapenemase-producers, the selective media used (commercial plates, 'in house' media) should be also reported here. In general, any variation with regard to the EURL-AR protocols should be stated here, number of isolates isolated per sample, in particular for *Campylobacter* spp..

(c): Antimicrobials included, Cut-off values

**39.General Description of Antimicrobial Resistance Monitoring*;**

**Campylobacter spp. in broiler carcases (neck skin) - food sample**

**1. General description of sampling design and strategy[(a)]**

The Romanian Surveillance Program is a national program, published in Romanian Official Journal as Order of the President of the National Sanitary Veterinary and Food Safety Authority (N.S.V.F.S.A.), yearly updated and the susceptibility testing of Campylobacter spp. from broilers is a part of the program. After strain confirmation and species identification, the strains are tested

for the antimicrobial resistance only at the National Reference Laboratory (N.R.L. - AR).

The Hygiene and Veterinary Public Health (I.H.V.P.H.) collects all the strains isolated from broilers in official laboratories.

**2. Stratification procedure per animal population and food category**

The NRL-AR (I.H.V.P.H.) tested all Campylobacter strains isolated in official laboratories and NRL Campylobacter for antimicrobial resistance.

**3. Randomisation procedure per animal population and food category**

Samples were collected according to the provisions of to Regulation CE 2073/2005, by the official veterinarians within the county (C.S.V.F.S.D.) based on the principle of representativeness, randomized on days / weeks / months / batch / epidemiological unit / the specificity of the sampling method / the specific matrix / type of unit / activity and other criteria set out in the monitoring plan of antimicrobial resistance.

**4. Analytical method used for detection and confirmation[(b)]**

Analytical methods used for enumeration of Campylobacter is microbiological method: EN ISO 10272-2 – Horizontal method for detection and enumeration Campylobacter spp. Part 2. Colony count technique. Species identification was performed by molecular techniques.

**5. Laboratory methodology used for detection of antimicrobial resistance[(C)]**

Laboratory protocol for isolation of Campylobacter is DTU Food, biochemical confirmation provided by EURL- AR. The method used for detection of the antimicrobial resistance is broth microdilution (ISO 20776) testing and quality control were performed according to CLSI (Clinical and Laboratory Standards Institute) documents and standards. The cut-off values used in testing are those provided by EURL- AR and by EFSA in the Manual for reporting on antimicrobial resistance. The antimicrobials included in monitoring were: Erytromycin, Ciprofloxacin, Tetracycline, Gentamicin Nalidixic acid, Streptomicyn.

**6. Results of investigation**

In 2018, were tested for identification Campylobacter spp.,1830 samples neck skin of broilers (Gallus gallus - carcase) and 81 samples meat fresh of broilers (Gallus gallus).

In 2018, 42 strains were tested for detection of the antimicrobial resistance of which 33 were isolated from samples ***neck skin of broilers*** and 9 were isolated from samples meat fresh of broilers.

**7. Additional information**

According to the provisions of the Order of President of National Sanitary Veterinary and Food Safety Authority no.34/2006, transposing into Romanian legislation the Directive 2003/99/EC, the strains isolated in foodstuffs were tested for the antimicrobial resistance. Species identification of strains of the **Campylobacter** and the antimicrobial resistance, was performed by the NRL and all the antimicrobial resistance data is collected only in Institute of Hygiene and Veterinary Public Health.

*** to be filled in per combination of bacterial species/matrix**

(a): Method of sampling (description of sampling technique: stage of sampling, type of sample, sampler), Frequency of sampling, Procedure of selection of isolates for susceptibility testing, Method used for collecting data.

(b): Analytical method used for detection and confirmation: according to the legislation, the protocols developed by the EURL-AR should be used and reported here. In the case of the voluntary specific monitoring on Carbapenemase-producers, the selective media used (commercial plates, 'in house' media) should be also reported here. In general, any variation with regard to the EURL-AR protocols should be stated here, number of isolates isolated per sample, in particular for *Campylobacter* spp..

(c): Antimicrobials included, Cut-off values

## 40. Institutions and laboratories involved in antimicrobial resistance monitoring and reporting

Institute for Diagnosis and Animal Health – central animal health diagnostic institute, NRL for Antimicrobial resistance - for monitoring and reporting
Sanitary Veterinary and Food Safety Directorate Counties: Alba, Brăila, Buzău, Dâmbovița, Giurgiu, Mureș, Prahova and Satu Mare – for monitoring

Short description of the institutions and laboratories involved in data collection and reporting

## General Antimicrobial Resistance Evaluation

**1. Situation and epidemiological evolution (trends and sources) regarding AMR to critically important antimicrobials[(a)] (CIAs) over time until recent situation**

Write text here please

**2. Public health relevance of the findings on food-borne AMR in animals and foodstuffs**

Write text here please

**3. Recent actions taken to control AMR in food producing animals and food**

Write text here please

**4. Any specific action decided in the Member State or suggestions to the European Union for actions to be taken against food-borne AMR threat**

Write text here please

**5. Additional information**

Write text here please

(a): The CIAs depends on the bacterial species considered and the harmonised set of substances tested within the framework of the harmonised monitoring:

- For *Campylobacter* spp., macrolides (erythromycin) and fluoroquinolones (ciprofloxacin);
- For *Salmonella* and *E. coli*, 3rd and 4th generation cephalosporins (cefotaxime) and fluoroquinolones (ciprofloxacin) and colistin (polymyxin);

## General Description of Antimicrobial Resistance Monitoring*; Broilers/E.coli, non-pathogenic

**1. General description of sampling design and strategy[(a)]**

According to Commission Implementing Decision No 652/2013 *Escherichia coli* strains isolated from broilers caecal samples which are tested for antimicrobial susceptibility were obtained from monitoring programmes, based on randomised sampling design. The commensal E. coli, ESBL/AmpC/carbapenemase producing E. coli isolates are originate from randomly selected farms and randomly selected within the slaughterhouses.
Type of specimen taken: 838 caecal samples from slaughtered broilers.
Frequency of the sampling: the collected samples at slaughter were evenly distributed over each month of the year to enable the different seasons to be covered, respectively from May to December 2018. They were sampled between 1 and 6 slaughter batches per week, respectively between 2 and 151 slaughter batches per year from different slaughterhouses. Only one representative sample of caecal content (10 caeca) per flock, derived from a different number of carcasses were gathered to account for clustering.
Methods of sampling (description of sampling techniques): Within slaughterhouses, after the mass gastrointestinal examination, the official vet wills perform caeca sampling on special designated location, that to avoid carcasses contamination with the intestinal contents. To avoid cross contamination, the cecum has to be sampled with caution by careful manual traction at the junction with the intestine;
- for a slaughtered animals lot, it shall be sampled 10 caeca, from 10 birds, which have to be randomly chosen on cutting line (avoiding the first part of the batch to be slaughtered, collecting samples from non-consecutive birds). The traceability has to be assured for each batch sample;
- caeca must be intact and full;

- caeca sample will be collected in a single sterile bag/pack for a transport.It is labeled with a unique number which is identical with the analysis request number, and sealed
- samples should not be exposed to extreme temperatures and as soon as possible have to be transported to the laboratory for testing them.
Procedures for the selection of isolates for antimicrobial testing: there were isolated from broilers 838 commensal E. coli strains and 574 presumptive ESBL/AmpC producing E. coli strains.
8 commensal E. coli strains were resistant to 3rd generation cephalosporinases.
They were tested for antimicrobial resistance 170 commensal E. coli strains and 574 ESBL/AmpC producing E. coli strains. The selection of the commensal E. coli strains for antimicrobial testing were based on geographical origin of the samples/farm and date of sampling.
Methods used for collecting data: in accordance with SN of NSVFSA no 7145/26279/09.05.2018, respectively Annex XI - Report of the results of AMR monitoring. The document contains the information requested in Part B of Decision No 652/2013. The data were collected by NRL-AR and transmitted to NSVFSA.

**2. Stratification procedure per animal population and food category**

They were sampled and tested 838 caecal samples from slaughtered broilers originate from randomly selected farms and randomly selected slaughterhouses, respectively 26 slaughterhouses from 19 different counties situated in different country regions.

**3. Randomisation procedure per animal population and food category**

The random sampling plan was stratified per slaughterhouse by allocating the number of samples collected per slaughterhouse proportionally to the annual throughput of the slaughterhouse. Sampling was performed on a random selection regarding sampling days, during each month; cecum samples were chosen at random, regardless of the origin of the slaughtered animals (farms/flocks in Romania).

**4. Analytical method used for detection and confirmation[(b)]**

The isolation of indicator commensal Escherichia coli was based on an 'in house' method and for the specific monitoring of ESBL-/AmpC-/Carbapenemase-producers were used the protocols developed by the EURL-AR. The specific monitoring on Carbapenemase-producers was voluntary and the selective media used were commercial plates.

**5. Laboratory methodology used for detection of antimicrobial resistance[(C)]**

Micro-dilution method performed according to the method described by EUCAST and CLSI, accepted as ISO 20776-1:2006. Antimicrobials included in monitoring are: Ampicillin, Azithromycin, Cefotaxime, Ceftazidime, Chloramphenicol, Ciprofloxacin, Colistin, Gentamicin, Meropenem, Nalidixic acid, Sulfamethoxazole, Tetracycline, Tigecycline, Trimethoprim (first panel) and Cefepime, Cefoxitin, Ceftazidime, Ceftazidime + clavulanic acid, Cefotaxime, Cefotaxime + clavulanic acid, Ertapenem, Imipenem, Meropenem, Temocillin (second panel), according to the Decision 2013/652/EU.
Cut-off values used in testing are in conformity with Decision 2013/652/EU (tabel 1 and tabel 4).

**6. Results of investigation**

During 2018 there were tested 838 caecal samples from slaughtered broilers for detection of commensal E. coli and ESBL/AmpC/carbapenemase producing E. coli. There were isolated 838 commensal E. coli strains and 574 ESBL/AmpC producing E. coli strains. 8 commensal E. coli strains were resistant to 3rd generation cephalosporinases. They were tested for antimicrobial resistance 170 commensal E. coli isolates and 574 ESBL/AmpC producing E. coli isolates. .

**7. Additional information**

-

*** to be filled in per combination of bacterial species/matrix**
(a): Method of sampling (description of sampling technique: stage of sampling, type of sample, sampler), Frequency of sampling,

Procedure of selection of isolates for susceptibility testing, Method used for collecting data.
(b): Analytical method used for detection and confirmation: according to the legislation, the protocols developed by the EURL-AR should be used and reported here. In the case of the voluntary specific monitoring on Carbapenemase-producers, the selective media used (commercial plates, 'in house' media) should be also reported here. In general, any variation with regard to the EURL-AR protocols should be stated here, number of isolates isolated per sample, in particular for *Campylobacter* spp..
(c): Antimicrobials included, Cut-off values

## General Description of Antimicrobial Resistance Monitoring*; Broilers/Campylobacter jejuni

**1. General description of sampling design and strategy[(a)]**

According to Commission Implementing Decision No 652/2013 *Campylobacter jejuni* strains isolated from broilers caecal samples which are tested for antimicrobial susceptibility were obtained from monitoring programmes, based on randomised sampling design. *Campylobacter jejuni* isolates are originate from randomly selected farms and randomly selected within the slaughterhouses.
Type of specimen taken: 838 caecal samples from slaughtered broilers.
Frequency of the sampling: the collected samples at slaughter were evenly distributed over each month of the year to enable the different seasons to be covered, respectively from May to December 2018. They were sampled between 1 and 6 slaughter batches per week, respectively between 2 and 151 slaughter batches per year from different slaughterhouses. Only one representative sample of caecal content (10 caeca) per flock, derived from a different number of carcasses were gathered to account for clustering.
Methods of sampling (description of sampling techniques): Within slaughterhouses, after the mass gastrointestinal examination, the official vet wills perform caeca sampling on special designated location, that to avoid carcasses contamination with the intestinal contents. To avoid cross contamination, the cecum has to be sampled with caution by careful manual traction at the junction with the intestine;
- for a slaughtered animals lot, it shall be sampled 10 caeca, from 10 birds, which have to be randomly chosen on cutting line (avoiding the first part of the batch to be slaughtered, collecting samples from non-consecutive birds). The traceability has to be assured for each batch sample;
- caeca must be intact and full;
- caeca sample will be collected in a single sterile bag/pack for a transport.It is labeled with a unique number which is identical with the analysis request number, and sealed
- samples should not be exposed to extreme temperatures and as soon as possible have to be transported to the laboratory for testing them.
Procedures for the selection of isolates for antimicrobial testing: there were isolated from broilers 754 Campylobacter strains: 338 Campylobacter jejuni and 416 Campylobacter coli.
They were tested for antimicrobial resistance 338 Campylobacter jejuni strains.
Methods used for collecting data: in accordance with SN of NSVFSA no 7145/26279/09.05.2018, respectively Annex XI - Report of the results of AMR monitoring. The document contains the information requested in Part B of Decision No 652/2013. The data were collected by NRL-AR and transmitted to NSVFSA.

**2. Stratification procedure per animal population and food category**

They were sampled and tested 838 caecal samples from slaughtered broilers originate from randomly selected farms and randomly selected slaughterhouses, respectively 26 slaughterhouses from 19 different counties situated in different country regions.

**3. Randomisation procedure per animal population and food category**

The random sampling plan was stratified per slaughterhouse by allocating the number of samples collected per slaughterhouse proportionally to the annual throughput of the slaughterhouse. Sampling was performed on a random selection regarding sampling days, during each month; cecum samples were chosen at random, regardless of the origin of the slaughtered animals (farms/flocks in Romania).

**4. Analytical method used for detection and confirmation[(b)]**

The isolation of Campylobacter jejuni was performed according to SR EN ISO 10272-1:2017 and OIE Manual.

**5. Laboratory methodology used for detection of antimicrobial resistance(C)**

Micro-dilution method performed according to the method described by the EUCAST and CLSI, accepted as ISO 20776-1:2006. Antimicrobials included in monitoring are: Ciprofloxacin, Erythromycin, Gentamicin, Nalidixic acid, Streptomycin and Tetracycline, according to the Decision 2013/652/EU. Cut-off values used in testing are in conformity with Decision 2013/652/EU (tabel 2).

**6. Results of investigation**

During 2018 there were tested 838 caecal samples from slaughtered broilers for detection of Campylobacter jejuni. There were isolated from broilers 754 Campylobacter strains: 338 Campylobacter jejuni and 416 Campylobacter coli. They were tested for antimicrobial resistance 338 Campylobacter jejuni strains.

**7. Additional information**

-

*** to be filled in per combination of bacterial species/matrix**

(a): Method of sampling (description of sampling technique: stage of sampling, type of sample, sampler), Frequency of sampling, Procedure of selection of isolates for susceptibility testing, Method used for collecting data.

(b): Analytical method used for detection and confirmation: according to the legislation, the protocols developed by the EURL-AR should be used and reported here. In the case of the voluntary specific monitoring on Carbapenemase-producers, the selective media used (commercial plates, 'in house' media) should be also reported here. In general, any variation with regard to the EURL-AR protocols should be stated here, number of isolates isolated per sample, in particular for *Campylobacter* spp..

(c): Antimicrobials included, Cut-off values

## General Description of Antimicrobial Resistance Monitoring*; Broilers/Salmonella spp.

**1. General description of sampling design and strategy(a)**

"According to Commission Implementing Decision No 652/2013 Salmonella spp. strains isolated from broilers boot swabs samples which are tested for antimicrobial susceptibility, were obtained in the framework of the National Salmonella control Programme in broilers of Gallus gallus, established according to Article 5(1) of Regulation (EC) No 2160/2003.
Methods used for collecting data: in accordance with SN of NSVFSA no 7145/26279/09.05.2018, respectively Annex XI - Report of the results of AMR monitoring. The document contains the information requested in Part B of Decision No 652/2013. The data were collected by NRL-AR and transmitted to NSVFSA.

**2. Stratification procedure per animal population and food category**

The selection of the Salmonella spp. strains for antimicrobial testing were based on serotype, geographical origin of the samples/farm and date of sampling. They were randomly selected 170 isolates from a total of 707 Salmonella strains, taking in account one isolate per epidemiological unit (flock) and year, geographical representativeness and an even distribution of the date of sampling over the year. One strain was resistant to 3rd generation cephalosporinases.

**3. Randomisation procedure per animal population and food category**

**4. Analytical method used for detection and confirmation(b)**

The isolation Salmonella spp. was performed according to ISO 6579:2002/A1:2007 and SR EN ISO 6579-1:2017 and the serotyping of isolated strains according to ISO/TR 6579-3:2014.

**5. Laboratory methodology used for detection of antimicrobial resistance(C)**

Micro-dilution method performed according to the method described by EUCAST and CLSI, accepted as ISO 20776-1:2006. Antimicrobials included in monitoring are: Ampicillin, Azithromycin, Cefotaxime,

Ceftazidime, Chloramphenicol, Ciprofloxacin, Colistin, Gentamicin, Meropenem, Nalidixic acid, Sulfamethoxazole, Tetracycline, Tigecycline, Trimethoprim (first panel) and Cefepime, Cefoxitin, Ceftazidime, Ceftazidime + clavulanic acid, Cefotaxime, Cefotaxime + clavulanic acid, Ertapenem, Imipenem, Meropenem, Temocillin (second panel), according to the Decision 2013/652/EU.
Cut-off values used in testing are in conformity with Decision 2013/652/EU (tabel 1 and tabel 4).

**6. Results of investigation**

During 2018 there were isolated and identified 707 Salmonella spp. strains from broilers There were selected and tested for antimicrobial resistance 170 Salmonella spp.

**7. Additional information**

-

*** to be filled in per combination of bacterial species/matrix**

(a): Method of sampling (description of sampling technique: stage of sampling, type of sample, sampler), Frequency of sampling, Procedure of selection of isolates for susceptibility testing, Method used for collecting data.

(b): Analytical method used for detection and confirmation: according to the legislation, the protocols developed by the EURL-AR should be used and reported here. In the case of the voluntary specific monitoring on Carbapenemase-producers, the selective media used (commercial plates, 'in house' media) should be also reported here. In general, any variation with regard to the EURL-AR protocols should be stated here, number of isolates isolated per sample, in particular for *Campylobacter* spp..

(c): Antimicrobials included, Cut-off values

## General Description of Antimicrobial Resistance Monitoring*; Fattening turkeys /E.coli, non-pathogenic

**1. General description of sampling design and strategy[(a)]**

According to Commission Implementing Decision No 652/2013 *Escherichia coli* strains isolated from fattening turkeys caecal samples which are tested for antimicrobial susceptibility were obtained from monitoring programmes, based on randomised sampling design. The commensal E. coli, ESBL/AmpC/carbapenemase producing E. coli isolates are originate from randomly selected farms and randomly selected within the slaughterhouses.
Type of specimen taken: 18 caecal samples from slaughtered fattening turkeys.
Frequency of the sampling: the collected samples at slaughter were evenly distributed over each month of the year to enable the different seasons to be covered, respectively from May to November 2018. They were sampled between 1 to 3 slaughter batches per month, from one slaughterhouse. Only one representative sample of caecal content (10 caeca) per flock, derived from a different number of carcasses were gathered to account for clustering.
Methods of sampling (description of sampling techniques): Within slaughterhouses, after the mass gastrointestinal examination, the official vet wills perform caeca sampling on special designated location, that to avoid carcasses contamination with the intestinal contents. To avoid cross contamination, the cecum has to be sampled with caution by careful manual traction at the junction with the intestine;
- for a slaughtered animals lot, it shall be sampled 10 caeca, from 10 birds, which have to be randomly chosen on cutting line (avoiding the first part of the batch to be slaughtered, collecting samples from non-consecutive birds). The traceability has to be assured for each batch sample;
- caeca must be intact and full;
- caeca sample will be collected in a single sterile bag/pack for a transport.It is labeled with a unique number which is identical with the analysis request number, and sealed
- samples should not be exposed to extreme temperatures and as soon as possible have to be transported to the laboratory for testing them.
Procedures for the selection of isolates for antimicrobial testing: there were isolated from broilers 18 commensal E. coli strains and 13 presumptive ESBL/AmpC producing E. coli strains.

None of the commensal E. coli strain were resistant to 3rd generation cephalosporinases.
They were tested for antimicrobial resistance 18 commensal E. coli strains and 13 ESBL/AmpC producing E. coli strains.
From fattening turkeys were isolated from broilers 18 commensal E. coli strains and 13 presumptive ESBL/AmpC producing E. coli strains. They were tested for antimicrobial resistance 18 commensal E. coli strains and 13 ESBL/AmpC producing E. coli strains.
Methods used for collecting data: in accordance with SN of NSVFSA no 7145/26279/09.05.2018, respectively Annex XI - Report of the results of AMR monitoring. The document contains the information requested in Part B of Decision No 652/2013. The data were collected by NRL-AR and transmitted to NSVFSA.

**2. Stratification procedure per animal population and food category**

They were sampled and tested 18 caecal samples from slaughtered fattening turkeys originate from 9 different farms from 2 different counties and one slaughterhouse.

**3. Randomisation procedure per animal population and food category**

The random sampling plan was stratified per slaughterhouse by allocating the number of samples collected per slaughterhouse proportionally to the annual throughput of the slaughterhouse. Sampling was performed on a random selection regarding sampling days, during each month; cecum samples were chosen at random, regardless of the origin of the slaughtered animals (farms/flocks in Romania).

**4. Analytical method used for detection and confirmation(b)**

The isolation of indicator commensal Escherichia coli was based on an 'in house' method and for the specific monitoring of ESBL-/AmpC-/Carbapenemase-producers were used the protocols developed by the EURL-AR. The specific monitoring on Carbapenemase-producers was voluntary and the selective media used were commercial plates.

**5. Laboratory methodology used for detection of antimicrobial resistance(C)**

Micro-dilution method performed according to the method described by EUCAST and CLSI, accepted as ISO 20776-1:2006. Antimicrobials included in monitoring are: Ampicillin, Azithromycin, Cefotaxime, Ceftazidime, Chloramphenicol, Ciprofloxacin, Colistin, Gentamicin, Meropenem, Nalidixic acid, Sulfamethoxazole, Tetracycline, Tigecycline, Trimethoprim (first panel) and Cefepime, Cefoxitin, Ceftazidime, Ceftazidime + clavulanic acid, Cefotaxime, Cefotaxime + clavulanic acid, Ertapenem, Imipenem, Meropenem, Temocillin (second panel), according to the Decision 2013/652/EU.
Cut-off values used in testing are in conformity with Decision 2013/652/EU (tabel 1 and tabel 4).

**6. Results of investigation**

During 2018 there were tested 18 caecal samples from slaughtered fattening turkeys for detection of commensal E. coli and ESBL/AmpC/carbapenemase producing E. coli. There were isolated 18 commensal E. coli strains and 13 ESBL/AmpC producing E. coli strains. None of the commensal E. coli strains was resistant to 3rd generation cephalosporinases. They were tested for antimicrobial resistance 18 commensal E. coli isolates and 13 ESBL/AmpC producing E. coli isolates. .

**7. Additional information**

-

*** to be filled in per combination of bacterial species/matrix**

(a): Method of sampling (description of sampling technique: stage of sampling, type of sample, sampler), Frequency of sampling, Procedure of selection of isolates for susceptibility testing, Method used for collecting data.

(b): Analytical method used for detection and confirmation: according to the legislation, the protocols developed by the EURL-AR should be used and reported here. In the case of the voluntary specific monitoring on Carbapenemase-producers, the selective media used (commercial plates, 'in house' media) should be also reported here. In general, any variation with regard to the EURL-AR protocols should be stated here, number of isolates isolated per sample, in particular for *Campylobacter* spp..

(c): Antimicrobials included, Cut-off values

## General Description of Antimicrobial Resistance Monitoring*; Fattening turkeys /Campylobacter jejuni

**1. General description of sampling design and strategy[(a)]**

According to Commission Implementing Decision No 652/2013 *Campylobacter jejuni* strains isolated from fattening turkeys caecal samples which are tested for antimicrobial susceptibility were obtained from monitoring programmes, based on randomised sampling design. The Campylobacter jejuni isolates are originate from randomly selected farms and randomly selected within the slaughterhouses.
Type of specimen taken: 18 caecal samples from slaughtered fattening turkeys.
Frequency of the sampling: the collected samples at slaughter were evenly distributed over each month of the year to enable the different seasons to be covered, respectively from May to November 2018. They were sampled between 1 to 3 slaughter batches per month, from one slaughterhouse. Only one representative sample of caecal content (10 caeca) per flock, derived from a different number of carcasses were gathered to account for clustering.
Methods of sampling (description of sampling techniques): Within slaughterhouses, after the mass gastrointestinal examination, the official vet wills perform caeca sampling on special designated location, that to avoid carcasses contamination with the intestinal contents. To avoid cross contamination, the cecum has to be sampled with caution by careful manual traction at the junction with the intestine;
- for a slaughtered animals lot, it shall be sampled 10 caeca, from 10 birds, which have to be randomly chosen on cutting line (avoiding the first part of the batch to be slaughtered, collecting samples from non-consecutive birds). The traceability has to be assured for each batch sample;
- caeca must be intact and full;
- caeca sample will be collected in a single sterile bag/pack for a transport.It is labeled with a unique number which is identical with the analysis request number, and sealed
- samples should not be exposed to extreme temperatures and as soon as possible have to be transported to the laboratory for testing them.
Procedures for the selection of isolates for antimicrobial testing: there were isolated from fattening turkeys 16 Campylobacter strains: 7 Campylobacter jejuni and 9 Campylobacter coli.
They were tested for antimicrobial resistance 7 Campylobacter jejuni strains.
Methods used for collecting data: in accordance with SN of NSVFSA no 7145/26279/09.05.2018, respectively Annex XI - Report of the results of AMR monitoring. The document contains the information requested in Part B of Decision No 652/2013. The data were collected by NRL-AR and transmitted to NSVFSA.

**2. Stratification procedure per animal population and food category**

They were sampled and tested 18 caecal samples from slaughtered fattening turkeys originate from 9 different farms from 2 different counties and one slaughterhouse.

**3. Randomisation procedure per animal population and food category**

The random sampling plan was stratified per slaughterhouse by allocating the number of samples collected per slaughterhouse proportionally to the annual throughput of the slaughterhouse. Sampling was performed on a random selection regarding sampling days, during each month; cecum samples were chosen at random, regardless of the origin of the slaughtered animals(farms/flocks in Romania).

**4. Analytical method used for detection and confirmation[(b)]**

The isolation of Campylobacter jejuni was performed according to SR EN ISO 10272-1:2017 and OIE Manual.

**5. Laboratory methodology used for detection of antimicrobial resistance(C)**

Micro-dilution method performed according to the method described by the EUCAST and CLSI, accepted as ISO 20776-1:2006. Antimicrobials included in monitoring are: Ciprofloxacin, Erythromycin, Gentamicin, Nalidixic acid, Streptomycin and Tetracycline, according to the Decision 2013/652/EU. Cut-off values used in testing are in conformity with Decision 2013/652/EU (tabel 2).

**6. Results of investigation**

During 2018 there were tested 18 caecal samples from slaughtered fattening turkeys for detection of Campylobacter jejuni. There were isolated from broilers 16 Campylobacter strains: 7 Campylobacter jejuni and 9 Campylobacter coli. They were tested for antimicrobial resistance 7 Campylobacter jejuni strains.

**7. Additional information**

-

*** to be filled in per combination of bacterial species/matrix**

(a): Method of sampling (description of sampling technique: stage of sampling, type of sample, sampler), Frequency of sampling, Procedure of selection of isolates for susceptibility testing, Method used for collecting data.

(b): Analytical method used for detection and confirmation: according to the legislation, the protocols developed by the EURL-AR should be used and reported here. In the case of the voluntary specific monitoring on Carbapenemase-producers, the selective media used (commercial plates, 'in house' media) should be also reported here. In general, any variation with regard to the EURL-AR protocols should be stated here, number of isolates isolated per sample, in particular for *Campylobacter* spp..

(c): Antimicrobials included, Cut-off values

## General Description of Antimicrobial Resistance Monitoring*; Fattening turkeys /Salmonella spp.

**1. General description of sampling design and strategy(a)**

According to Commission Implementing Decision No 652/2013 Salmonella spp. strains isolated from fattening turkeys boot swabs samples which are tested for antimicrobial susceptibility, were obtained in the framework of the National Salmonella control Programme in fattening turkeys, established according to Article 5(1) of Regulation (EC) No 2160/2003.
Methods used for collecting data: in accordance with SN of NSVFSA no 7145/26279/09.05.2018, respectively Annex XI - Report of the results of AMR monitoring. The document contains the information requested in Part B of Decision No 652/2013. The data were collected by NRL-AR and transmitted to NSVFSA.

**2. Stratification procedure per animal population and food category**

The selection of the Salmonella spp. strains for antimicrobial testing were based on serotype, geographical origin of the samples/farm and date of sampling.
They were selected 2 isolates from a total of 2 Salmonella strains, taking in account one isolate per epidemiological unit (flock) and year, geographical representativeness and an even distribution of the date of sampling over the year. None of the Salmonella strains was resistant to 3rd generation cephalosporinases.

**3. Randomisation procedure per animal population and food category**

**4. Analytical method used for detection and confirmation(b)**

The isolation Salmonella spp. was performed according to ISO 6579:2002/A1:2007 and SR EN ISO 6579-1:2017 and the serotyping of isolated strains according to ISO/TR 6579-3:2014.

| **5. Laboratory methodology used for detection of antimicrobial resistance[(C)]** |
|---|
| Micro-dilution method performed according to the method described by EUCAST and CLSI, accepted as ISO 20776-1:2006. Antimicrobials included in monitoring are: Ampicillin, Azithromycin, Cefotaxime, Ceftazidime, Chloramphenicol, Ciprofloxacin, Colistin, Gentamicin, Meropenem, Nalidixic acid, Sulfamethoxazole, Tetracycline, Tigecycline, Trimethoprim (first panel) and Cefepime, Cefoxitin, Ceftazidime, Ceftazidime + clavulanic acid, Cefotaxime, Cefotaxime + clavulanic acid, Ertapenem, Imipenem, Meropenem, Temocillin (second panel), according to the Decision 2013/652/EU.<br>Cut-off values used in testing are in conformity with Decision 2013/652/EU (tabel 1 and tabel 4). |
| **6. Results of investigation** |
| During 2018 there were isolated and identified 2 Salmonella spp. strains from fattening turkeys. There were selected and tested for antimicrobial resistance 2 Salmonella spp. strains. |
| **7. Additional information** |
| - |

*** to be filled in per combination of bacterial species/matrix**

(a): Method of sampling (description of sampling technique: stage of sampling, type of sample, sampler), Frequency of sampling, Procedure of selection of isolates for susceptibility testing, Method used for collecting data.

(b): Analytical method used for detection and confirmation: according to the legislation, the protocols developed by the EURL-AR should be used and reported here. In the case of the voluntary specific monitoring on Carbapenemase-producers, the selective media used (commercial plates, 'in house' media) should be also reported here. In general, any variation with regard to the EURL-AR protocols should be stated here, number of isolates isolated per sample, in particular for *Campylobacter* spp..

(c): Antimicrobials included, Cut-off values

## General Description of Antimicrobial Resistance Monitoring*; Laying hens /Salmonella spp.

| **1. General description of sampling design and strategy[(a)]** |
|---|
| According to Commission Implementing Decision No 652/2013 Salmonella spp. strains isolated from laying hens boot swabs and faeces samples which are tested for antimicrobial susceptibility, were obtained in the framework of the National Salmonella control Programme in laying hens, established according to Article 5(1) of Regulation (EC) No 2160/2003.<br>Methods used for collecting data: in accordance with SN of NSVFSA no 7145/26279/09.05.2018, respectively Annex XI - Report of the results of AMR monitoring. The document contains the information requested in Part B of Decision No 652/2013. The data were collected by NRL-AR and transmitted to NSVFSA. |
| **2. Stratification procedure per animal population and food category** |
| The selection of the Salmonella spp. strains for antimicrobial testing were based on serotype, geographical origin of the samples/farm and date of sampling.<br>They were selected 47 isolates from a total of 100 Salmonella strains, taking in account one isolate per epidemiological unit (flock) and year, geographical representativeness and an even distribution of the date of sampling over the year. None of the Salmonella strains was resistant to 3rd generation cephalosporinases. |
| **3. Randomisation procedure per animal population and food category** |
| |
| **4. Analytical method used for detection and confirmation[(b)]** |
| The isolation Salmonella spp. was performed according to ISO 6579:2002/A1:2007 and SR EN ISO 6579-1:2017 and the serotyping of isolated strains according to ISO/TR 6579-3:2014. |
| **5. Laboratory methodology used for detection of antimicrobial resistance[(C)]** |
| Micro-dilution method performed according to the method described by EUCAST and CLSI, accepted as |

ISO 20776-1:2006. Antimicrobials included in monitoring are: Ampicillin, Azithromycin, Cefotaxime, Ceftazidime, Chloramphenicol, Ciprofloxacin, Colistin, Gentamicin, Meropenem, Nalidixic acid, Sulfamethoxazole, Tetracycline, Tigecycline, Trimethoprim (first panel) and Cefepime, Cefoxitin, Ceftazidime, Ceftazidime + clavulanic acid, Cefotaxime, Cefotaxime + clavulanic acid, Ertapenem, Imipenem, Meropenem, Temocillin (second panel), according to the Decision 2013/652/EU.
Cut-off values used in testing are in conformity with Decision 2013/652/EU (tabel 1 and tabel 4).

**6. Results of investigation**

During 2018 there were isolated and identified 100 Salmonella spp. strains from laying hens. There were selected and tested for antimicrobial resistance 47 Salmonella spp. strains.

**7. Additional information**

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*** to be filled in per combination of bacterial species/matrix**

(a): Method of sampling (description of sampling technique: stage of sampling, type of sample, sampler), Frequency of sampling, Procedure of selection of isolates for susceptibility testing, Method used for collecting data.

(b): Analytical method used for detection and confirmation: according to the legislation, the protocols developed by the EURL-AR should be used and reported here. In the case of the voluntary specific monitoring on Carbapenemase-producers, the selective media used (commercial plates, 'in house' media) should be also reported here. In general, any variation with regard to the EURL-AR protocols should be stated here, number of isolates isolated per sample, in particular for *Campylobacter* spp..

(c): Antimicrobials included, Cut-off values