

ALLERGENICITY ASSESSMENT

Applications for Authorisation of Genetically Modified Food and Feed Products submitted under Regulation (EC) No 1829/2003



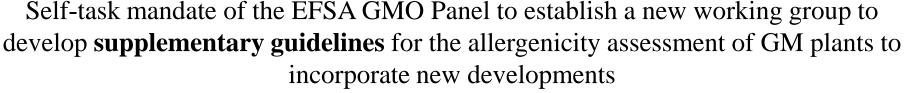


Outline

- 1. Background information
- 2. Pepsin resistance
- 3. Non-IgE mediated immune adverse reactions to foods
- 4. Endogenous allergens
- 5. Concluding remarks
- 6. Implementation



1. Background information



(published: July 2014; deadline: July 2016)

Background

Assessment of the two 2013 publications commissioned and published by EFSA

- 1. "Literature review: 'in vitro' digestibility tests for allergenicity assessment" by E.N. Clare Mills et al., 2013a
- 2. "Literature review: non-IgE-mediated immune adverse reactions to foods" by E.N. Clare Mills *et al.*, 2013b

Inclusion of certain allergens (as defined in OECD Consensus documents) in the compositional analysis and consequently, the quantitative measurement of individual allergens.

Proposal: Develop supplementary guidelines for the allergenicity assessment of GM plants to incorporate new developments



2. Pepsin resistance: historical perspective

- **Codex Guidelines** recommend testing for resistance to pepsin digestion since "a correlation exists between resistance to digestion by pepsin and allergenic potential"
- Implementing Regulation (EU) No 503/2013 as well as 2011 EFSA Guidance recommendations are focused on *physiological digestion* instead of *pepsin resistance*
- In vitro digestibility tests for allergenicity assessment (Mills et al., 2013a)
 recommends standardisation of methodology and replication of the dynamic
 digestive process; the HESI sponsored efforts to standardise the pepsin
 resistance assay not discussed

EFSA guidance documents and Implementing Regulation (EU) No 503/2013 deviate from internationally agreed guidelines



2. Pepsin resistance: current status

- Most dietary proteins are digested and provide nutritional value
- The ability to resist pepsin digestion is an intrinsic property of the protein
- Current pepsin resistance assay provides information to predict the potential dietary exposure to a protein
- Pepsin's enzymatic activity is optimal between pH 1.5-2.0. Not clear how assays outside of this optimal range will further inform on protein safety
- Codex guidelines do not require pepsin resistance assays to strictly mimic physiological conditions of human digestion
 - No validated in vitro method exists that can mimic human digestion conditions
 - Human digestion is highly variable and depends on many factors including age, diet, genetic make-up



3. Non-IgE mediated immune adverse reactions to foods: *historical perspective*

- Non-IgE-mediated immune adverse reactions to foods (Mills et al., 2013b)
 reviews the science of identifying gluten and related food proteins that cause Tcell mediated Celiac Disease
- Testing scheme proposal presented for in vitro or in vivo immunoreactivity when/if a trigger is identified. However,
 - No testing platforms are identified and/or validated
 - No predictive tests for novel proteins and the likelihood of T-cell stimulation are available.
 - No serum test for celiac disease is available
- Mills et al., 2013b paper is consistent with Martel et al., 2010 paper which looked at examining enzymes. Also consistent with 2012 International Symposium in Prague (PATC/HESI) that examined the state of the science in sensitisation

Not clear how the risk assessment process and hypothesis-driven analyses would be used effectively for GMO protein risk assessment



3. Non-IgE mediated immune adverse reactions to foods: *current status*

- University of Nebraska food allergen database FARRP (<u>www.allergenonline.org</u>) includes gluten and related proteins (since inception)
- Risk management of significant "in silico" bioinformatic
 alignments to gluten proteins is consistent with alignments to
 any allergen proteins; would require a closer look into risk or
 avoidance of novel protein
 - Current industry practices would **not** take any novel proteins with alignments to allergens through development
 - Current weight-of-evidence approach encompass celiac disease risks for novel proteins



4. Endogenous allergens

- Endogenous allergens are assessed only in commonly allergenic crops such as soybean
- Several soybean allergen lists exists, but only some of the proteins from these lists had sufficient clinical data to suggest they cause soybean allergic disease
- MS, ELISA-, and gel separation-based methods have been developed and can provide comparative measurements of allergen levels
- Methods using human serum are not appropriate for the measurement of allergens in a comparative assessment because of the limited supply of critical reagents
- How to interpret the data?
 - Allergic individuals already avoid the offending foods
 - No allergen threshold information exists for sensitisation
 - No clear link between the amount of allergen exposure and sensitisation
 - Information on the natural variability of allergen expression levels is necessary to establish a context of safety



5. Concluding remarks

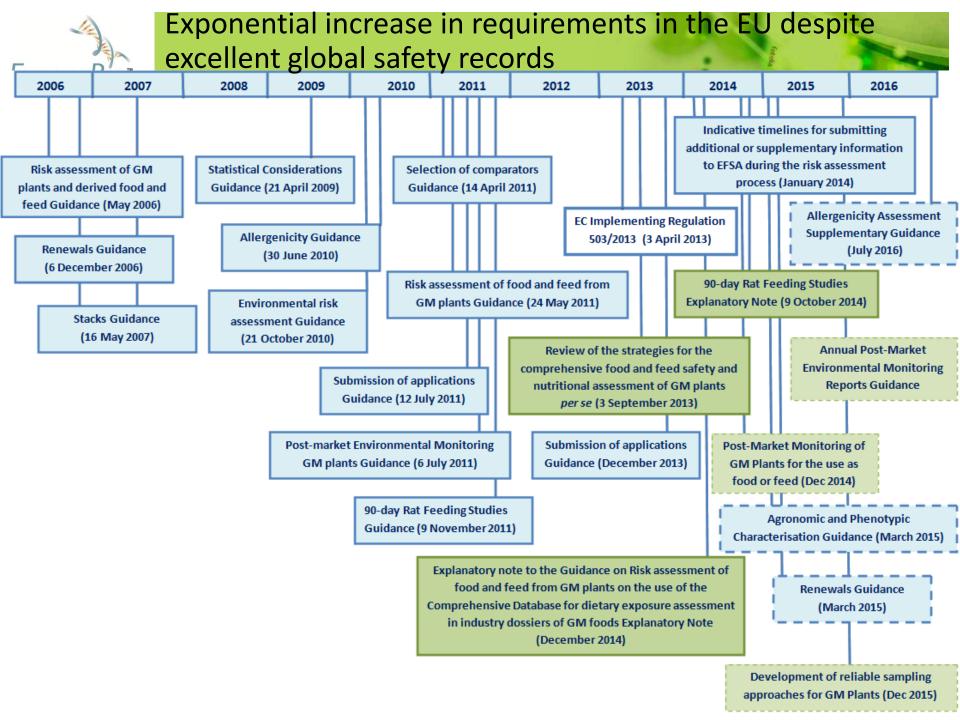
- Codex guidelines recommend testing for resistance to pepsin digestion through a validated method
- ➤ Weight-of-evidence approach can cover the potential celiac disease risks for novel proteins
- No serum testing for celiac disease is available
- OECD consensus document for soybean does not recommend testing of endogenous allergens as part of the comparative assessment
- ➤ No predictive testing for *de novo* sensitisation
- Current allergenicity data package sufficient to conclude on negligible risk



Implementation



- Need for a standard procedure for clear and timely communication of new requirements to applicants
- Need for sufficient and realistic transition periods before attaining mandatory compliance with the requirements set out in new guidance documents
- EFSA guidance should not be applied retroactively
- Need to justify the relevance for risk assessment (hazard x exposure)





Thank you!

28 October 2014 EFSA Technical Meeting 12



Additional Information

28 October 2014 EFSA Technical Meeting 13



1. Background information (continued)

Risk Assessment

- Risk is a function of hazard and exposure; the allergenicity assessment addresses both portions of this equation to identify risk.
- The risk assessment paradigm has to be acknowledged in terms that are consistent with the context of the hazard and exposure characterisations performed in the allergy weight-ofevidence approach.

Hazard defined adverse effects from the newly expressed proteins in GM crops or

from the transformation of the parental crop

Exposure a measure of the likelihood that the hazard comes into significant contact

with humans/animals such that the hazards can have a physiological impact

Characterisations typical of the allergy assessment. These assess whether there are

biophysical similarities between a novel protein and known allergens. There is no single, fully predictive test; therefore, the sum or "weight-of-evidence" from all of the characterisation studies is taken into account.

Study designs should have a hypothesis that can be tested and have endpoints with

interpretable outcomes.



1. Background information (continued)

How do allergy studies fit in with risk assessment?

Key points:

- 1. There is no single, fully predictive test for de novo allergy.
- 2. Because no characterisation studies are perfectly consistent among known allergens, all of the information for newly expressed proteins is assessed for allergenic potential, there is no single test to predict risk.
- 3. Many of the characterisation studies (assessments) feature an evaluation of exposure

Characterize protein <u>hazard</u>

- Not an allergen
- Similarity with known allergens
- Source organism
- Molecular characterisation identifies intended protein



Characterize protein <u>exposure</u>

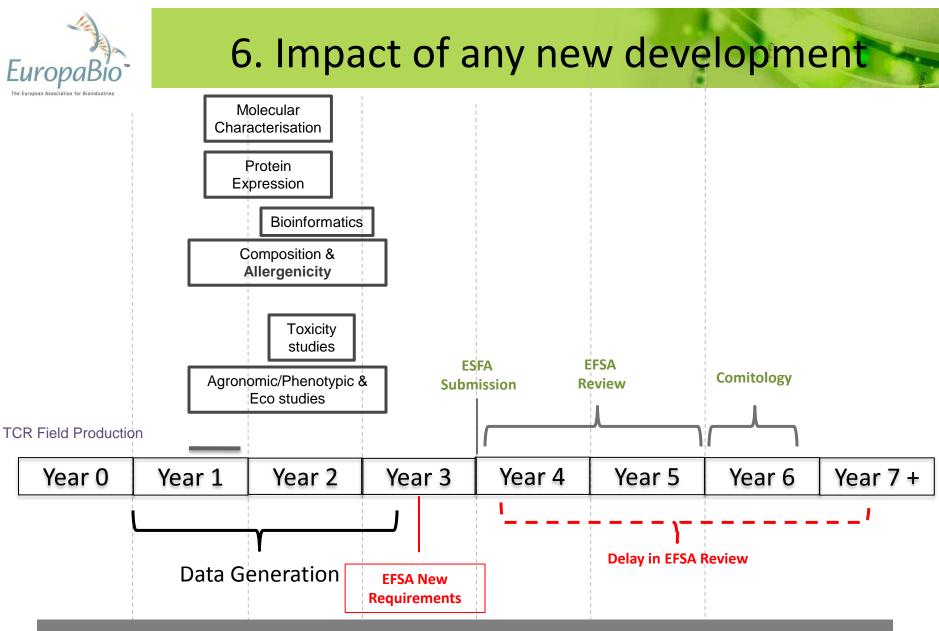
- History of safe exposure oral
- Pepsin (SGF) oral
- Heat Denaturation oral
- Abundance in Food/Feed oral



1. Background information (continued)

Key points in achieving consistent risk assessment

- Should make use of the full background knowledge regarding the novel protein's characterisation and intended use in food/feed.
- Should maintain a *case-by-case* approach
- In general, no specific characterisation should be taken as proof of risk in exclusion of the rest of risk assessment. Specific knowledge that supports safety should be utilized:
 - <u>Examples specific to allergy</u>: bacterial proteins not generally known as allergens; enzymes shown clinically not to act as allergens by oral route
- Achieve consistency in risk assessment
 - Industry provides dietary exposure assessment, but EFSA promotes "worst case" exposure assessment by assuming maximal exposure; this is undefined. It reduces the importance of recognizing the typical, very low concentrations of novel proteins in grain, enzyme clinical studies, etc.



Sufficient and realistic transition period warranted



References

- 1. Astwood JD, Leach JN, Fuchs RL. 1996. Stability of food allergens to digestion in vitro. Nat Biotechnol. 14(10):1269-73.
- 2. EFSA (European Food Safety Authority), 2010. EFSA panel on Genetically Modified Organisms (GMO). Scientific opinion on the assessment of allergenicity of GM plants and microorganisms and derived food and feed. EFSA J. 8, 1–168.
- 3. European Commission Implementing Regulation (EU) No 503/2013 of 3 April 2013 on applications for authorization of genetically modified food and feed in accordance with regulation (EC). No 1829/2003 of the European Parliament and of the council and amending commission regulations (EC) No 641/2004 and (EC) NO 1981/2006. Official J. EU L157, 1-48.
- 4. Fu, T.J., Abbott, U.R., Hatzos, C. 2002. Digestibility of food allergens and nonallergenic proteins in simulated gastric fluid and simulated intestinal fluid-a comparative study. J. Agricultural Food Chemistry. 50:7154–7160.
- 5. Herman, R.A., Woolhiser, M.M., Ladics, G. S., Korjagin, V. A., Schafer, B. W., Storer, N. P., Green, S. B., Kan, L. 2007. Stability of a set of allergens and non-allergens in simulated gastric fluid. Int. J. Food Sci. Nutr. 58: 125–141.
- 6. Ladics GS, Budziszewski GJ, Herman RA, Herouet-Guicheney C, Joshi S, Lipscomb EA, McClain S, Ward JM. 2014. Measurement of endogenous allergens in genetically modified soybeans short communication. Regul Toxicol Pharmacol 70: 75-79.
- 7. L'Hocine, L., Boye, J.I., 2007. Allergenicity of soybean: new developments in identification of allergenic proteins, cross-reactivities and hypoallergenization technologies. Crit. Rev. Food Sci. Nutr. 47, 127–143.
- 8. Martel C, Nielsen GD, Mari A, Licht TR, Poulsen LK. 2010. Bibliographic review on the potential of microorganisms, microbial products and enzymes to induce respiratory sensitization. SCIENTIFIC / TECHNICAL REPORT submitted to EFSA CFP/EFSA/FEEDAP/2009/02.
- 9. Mills, EN, Marsh, JT, Boyle, R, Hoffmann-Sommergruber, K, DuPont, D, Bartra, J, Bakalis, S, McLaughlin, J and Shewry, PR. 2013a. Literature Review: 'in vitro digestibility tests for allergenicity assessment'. EFSA supporting publication 2013: EN-529, 52pp.
- 10. Mills, EN, Marsh, JT, Boyle, R, Hoffmann-Sommergruber, K, DuPont, D, Bartra, J, Bakalis, S, McLaughlin, J and Shewry, PR. 2013b. Literature Review: 'non-lgE-mediated immune adverse reactions to foods'. EFSA supporting publication 2013: EN-527, 40pp.
- 11. OECD, 2012. Revised consensus document on compositional considerations for new varieties of soybean [Glycine max (L.) Merr.]: key food and feed nutrients, antinutrients, toxicants and allergens. Series on the Safety of Novel Foods and Feeds No. 25.
- 12. Piper, D.W., Fenton, F.H. 1965. pH stability and activity curves of pepsin with special reference to their clinical importance. Gut. 6:506–508.
- 13. Thomas, K., Aalbers, M., Bannon, G.A., Bartels, M., Dearman, R.J., Esdaile, D.J., Fu, T.J., Glatt, C.M., Hadfield, N., Hatzos, C., Hefle, S.L., Heylings, J.R., Goodman, R.E., Henry, B., Herouet, C., Holsapple, M., Ladics, G.S., Landry, T.D., MacIntosh, S.C., Rice, E.A., Privalle, L.S., Steiner, H.Y., Teshima, R., Van Ree, R., Woolhiser, M., Zawodny, J., 2004. A multi-laboratory evaluation of a common in vitro pepsin digestion assay protocol used in assessing the safety of novel proteins. Reg. Toxicol. Pharmacol. 39:87–98.
- 14. Guideline for the conduct of food safety assessment of foods derived from recombinant-DNA plants http://www.fao.org/fileadmin/user_upload/gmfp/docs/CAC.GL 45 2003.pdf