

# From transnational research collaboration to regional Standards: the case of the EPPO Diagnostic Protocol on *Xylella fastidiosa*

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#### **EPPO IN A FEW WORDS**



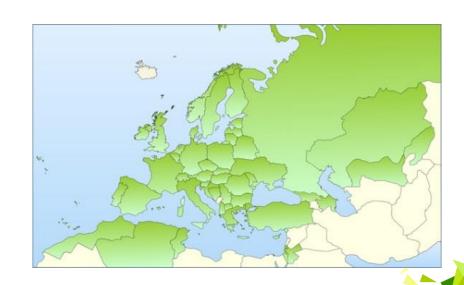
# ONE OF 9 REGIONAL PLANT PROTECTION ORGANIZATIONS RECOGNISED UNDER THE INTERNATIONAL PLANT PROTECTION CONVENTION CREATED IN 1951 NOW 51 MEMBER COUNTRIES

Remit set out in the EPPO Convention – in practice supporting member countries in particular in the areas of:

- Plant quarantine
- Efficacy of plant protection products
- Invasive alien plants
- Biological control agents

#### by:

- Developing and adopting regional technical Standards
- Disseminating information (information services)
- Facilitating networking in the region through Panel meetings, conferences, workshops.



#### STANDARD SETTING IN EPPO

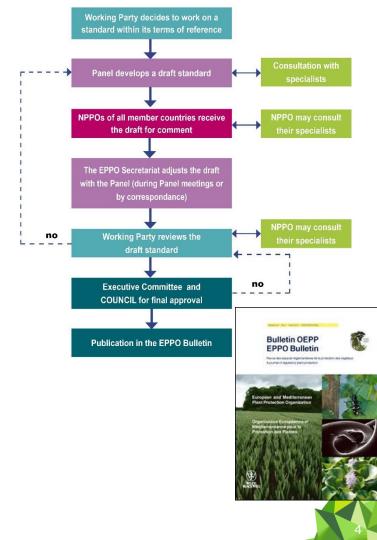
- Long standing and active program for Standard setting in several areas, including diagnostics.
- Objectives: to achieve a harmonized approach to detection and identification for regulated pests.
- The work started in 1998 is conducted by the Panels on Diagnostics.
- Panels are composed of specialists from EPPO member countries.







- Standards are written according to a "common format and content".
- First drafts of Standards prepared by an assigned expert author(s) or by a drafting team and reviewed by different EPPO groups of experts.
- Standards are approved following an approval procedure which involves a formal written consultation of all EPPO Member countries.
- Standards are published in the EPPO Bulletin and are freely available. Also available from the EPPO Global Database https://gd.eppo.int/



# PM 7/24 DIAGNOSTIC PROTOCOL FOR XYLELLA FASTIDIOSA: HISTORY OF THE STANDARD



First version approved in 2003 (published 2004)



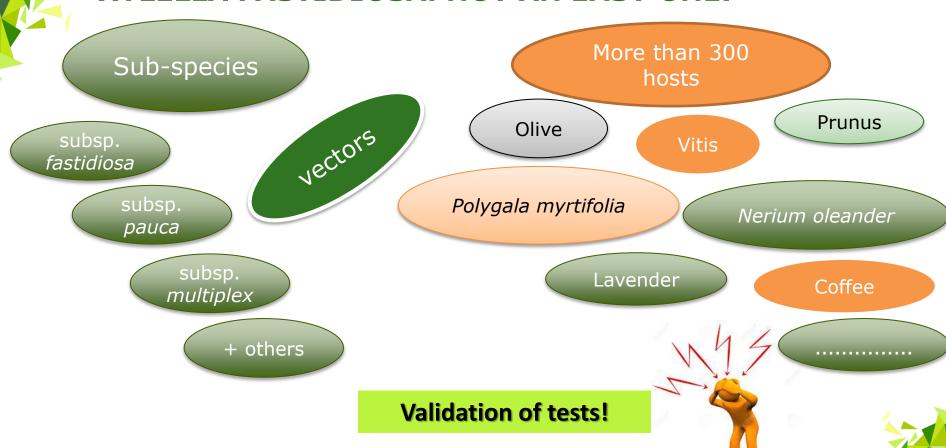
#### **Focused on Vitis and Citrus**



- Revision initiated in 2015 at the Panel on Diagnostics in Bacteriology
- Expert Working Group formed with experts on XF diagnostic from Austria, France, Italy, the Netherlands, Slovenia and Spain.
- Contribution received from experts from the US and Brazil

Work from December 2015 to March 2016

#### XYLELLA FASTIDIOSA: NOT AN EASY ONE!



#### MAIN CHANGES: MORE PICTURES OF POSSIBLE SYMPTOMS



Fig. 3 Leaf scorch symptoms on almond. Courtesy D. Boscia, CNR-Institute for Sustainable Plant Protection (IT).



Fig. 4 Scorch symptoms with distinct leaf burn surrounded by a dark line of demarcation between green and dead tissue. Courtesy P.M. Brennan University of Georgia (US).



Fig. 11 Symptoms of quick olive decline syndrome. Courtesy D. Boscia, CNR-Institute for Sustainable Plant Protection (IT).



Fig. 12 Symptoms of quick olive decline syndrome. Courtesy D. Boscia, CNR-Institute for Sustainable Plant Protection (IF).



Fig. 9 Leaf scorch symptoms on Coffea sp. Courtesy M. Bergsma-Vlami, NPPO (NL).



Fig. 10 'Crespera' symptoms on Coffea sp. including curling of leaf margins, chlorosis and deformation (asymmetry). Courtesy M. Bergsma-Vlami, NPPO (NL).



Fig. 19 Marginal leaf scorch symptoms caused by *Xylella fastidiosa* subsp. *pauca* on oleander. Courtesy D. Boscia, CNR-Institute for Sustainable Plant Protection (IT).



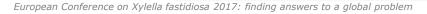
Fig. 20 Symptoms on Polygala myrtifolia. Courtesy B. Legendre, Anses, Plant Health Laboratory (FR).



#### MAIN CHANGES: MORE DETAILS ON SAMPLE PREPARATION

Type of sample	Host plants/ type of tissue	Minimum number of leaves per laboratory sample	Approximate weight of the laboratory sample
	Petioles and/or midribs or leaves of large size such as Coffea sp.; Ficus sp., Vitis sp., Nerium oleander.	5	0.5-1g
Samples from individual plants with leaves	Petioles and/or midribs of leaves of small size such as Polygala myrtifolia and Olea sp.	25	0.5-1 g
	Plant species without petioles or with small petiole and midrib	25	0.5-1 g
Composite sample from several plants from a single lot with leaves	Samples of asymptomatic plants collected from e.g. imported consignments or nursery monitoring	100-200	10 to 50 g
Dormant plants or cuttings	Xylem tissue	Not applicable	0.5-1 g

# Asymptomatic plants tricky as often!



# MAIN CHANGES: NEW TESTS FOR DETECTION AND IDENTIFICATION

Serological tests (plant material only)

- ELISA
- Immunofluorescence
- DTBIA (Direct Tissue Blot Immunoassay)

Molecular tests (plants and vectors)

- Conventional PCR (Minsavage)
- Real-time PCR (Francis; Harper)
- LAMP (Loop mediated isothermal amplification, Harper)

Different tests included to take into account the different situations in the EPPO region

#### **Sub species assignation:**

- MLST Yuan
- Conventional PCR Pooler & Hartung; Hernandez-Martinez (simplex & multiplex)

Unlike other protocols for bacteria isolation is not recommended as screening test.

Validation data in http://dc.eppo.int/validationlist.php



# FLOW DIAGRAMS TAKING INTO ACCOUNT SPECIFIC SITUATIONS (E.G. AREA FREE OR NOT)

Symptomatic/asymptomatic plants

Screening test(s)
Serological tests (IF, DTIAB, ELISA)

Conventional PCR/ real-time PCR test/LAMP

When two tests are performed they should be based on different biological principles or targeting different parts of the genome

Test(s)
negative

X. fastidiosa
not detected

Inconsistent tests results



Retesting and/or resampling recommended

At least two tests positive

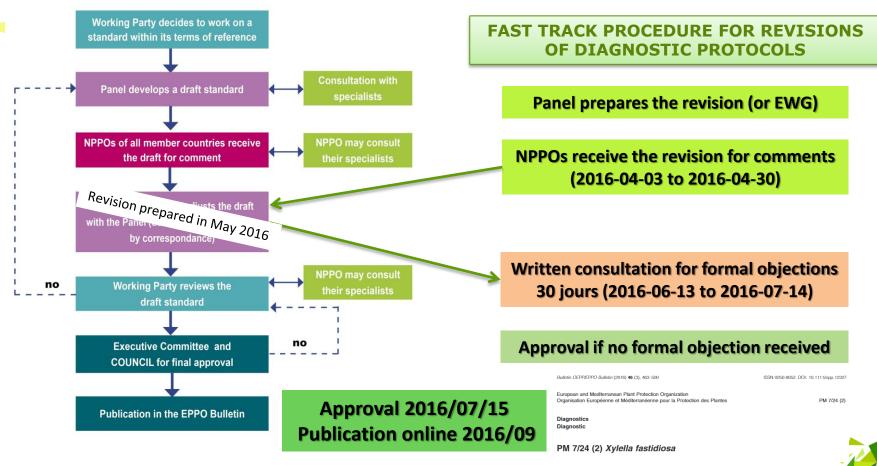


X. fastidiosa detected

For testing of
(a)symptomatic plants in
an outbreak area or in
buffer zone a single test
including serological tests
may be performed

It is advised to include molecular tests for detection on asymptomatic plants material in pest free area

#### **APPROVAL IN JULY 2016**





#### A LOT HAS HAPPENED SINCE MAY 2016

Several research projects on *Xylella fastidiosa* launched at national or regional level and outcomes are becoming available.

# NEED TO INCORPORATE RESULTS FROM RESEARCH IN A REVISION TO ENSURE THAT THE DP IS UP TODATE

#### 9 Feedback on this Diagnostic Protocol

If you have any feedback concerning this Diagnostic Protocol, or any of the tests included, or if you can provide additional validation data for tests included in this Protocol that you wish to share please contact diagnostics@eppo.int.

#### **NATIONAL RESEARCH PROJECTS**







# TRANSNATIONAL RESEARCH PROJECTS



2015-F-146: Harmonized protocol for monitoring and detection of *Xylella fastidiosa* in its host plants and its vectors (**PROMODE**)

23 research organisations (Europe, North Africa, North America)

- Develop sampling methods in symptomatic and asymptomatic plant materials and insect vectors
- Development of procedures for the highly sensitive detection in host plants and vectors (LAMP, digital PCR, NGS MLST)
- Review of available diagnostic methods and development of a common diagnostic method through interlaboratory validation
  - One proficiency test organized in 2017 on plant material
  - A proficiency test in progress on insect vectors





# TRANSNATIONAL RESEARCH PROJECTS



2016-F-221: *Xylella fastidiosa* and its insect vectors Cicadella 10 research organisations (Europe, North Africa)

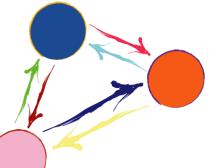
- Evaluation of sampling methods for vector species in the environment.
- Development of real-time tests for known and suspected vector species (including North American species) to provide taxonomic support (especially nymphs) and rapid sample screening.
- Improvement of the real time LAMP detection method of X. fastidiosa in potential vectors.
- Validation of the non-destructive DNA extraction methods already developed in potential vectors.
- Improvement of the non-destructive extraction methods to other insect.



#### **EU H2020 FUNDED PROJECTS**



WP4 Harmonization and validation of Xf diagnostic procedures



Euphresco projects
&
national projects



WP 4 Implementation and validation of diagnostics for early and rapid detection of target pathogens in host plants and vectors..



#### **NEEDS FOR IMPROVEMENT OF THE EPPO DP**

Revision of the DP discussed at the last meeting of the Panel on Diagnostics in Bacteriology (2017-05, Bari IT)

TPS results of projects presented

### **CONCLUSIONS OF THE REVIEW**



The protocol broadly serves the needs of the laboratories but most critical points are:

Difficulty to isolate the bacterium

Determination of subspecies

The EPPO DP EWG should be reactivated and should include new experts from the XF-ACTORS and PONTE projects to ensure that research results are included.





# First meeting in September 2017 (12/13, Paris)

Experts from France, Germany, Italy and Spain

# Second meeting November 2017 (12, Mallorca)

- Experts from Belgium, France, Italy, the Netherlands, Spain.
- Experts from US.

Proposals for revision made based on the data gathered by the experts in the framework of the different projects





## SEPTEMBER MEETING MAIN ISSUES DISCUSSED (1)

#### Isolation of the bacterium

 Addition of a sonication step before plating



DNA extraction: more instructions for QuickPick in manual use

#### Real-time PCR as screening test in plant material:

Analytical sensitivity of Harper *et al.* (2010) higher than Francis *et al* (2006). The Taqman version of Francis does not detect some American strains of Xf

DP now recommends to perform the real-time Harper et al. (2010) first.

Real-time Li et al., 2013 under evaluation in XF-ACTORS, PONTE and Euphresco

## SEPTEMBER MEETING MAIN ISSUES DISCUSSED (2)

Assignment of isolates to subspecies (important in the European context)

Difficulties to assign subspecies with MLST on plant extracts some gene targets not amplified resulting in lack of amplicons to be sequenced; different host plants producing incomplete allelic profiles.

Proposal made: sequences determined for at least two housekeeping genes can also be used for assignment of the subspecies

Difficulty of making links between ST in MLST database and subspecies

Table correspondence between ST and subspecies added

## SEPTEMBER MEETING MAIN ISSUES DISCUSSED (3)

# Difficulties with MLST on different hosts or in case of mixed infections

Laboratories adapt the MLST protocol to optimise it.

**Proposal made:** recommendation in the DP that if erratic amplification occurs, PCR parameters can be adjusted:

- dilution of the DNA extract (to limit inhibition) or
- increase of DNA input,
- use of a different Tag polymerase/Mastermix,
- decrease of annealing temperature from 65° C to 60° C or 58° C or
- increase of primer concentration from 0.3 to 0.5 μM.

## Multiplex PCR from Hernandez-Martinez et al. (2006)

Should not be used on plant extracts





## **NOVEMBER MEETING MAIN ISSUES DISCUSSED (1)**

# Changes proposed by the EWG accepted

# Discussion during the Euphresco meeting:

- Real-time PCR Harper in rare cases late Ct values observed. Comment to include in the protocol to warn the users with some recommendations.
- New triplex real-time PCR developed to consider for a further revision.
- Nested PCR developed



# **NEXT STEPS**

- Country Consultation in early 2018 (ideally after the results of the TPS on insects and TPS on Li et al., 2013)
- Adoption of a revision before summer 2018
- Planning of the next revision to start before summer 2018
  What could be improved:



Sampling of asymptomatic material Testing of vectors
Additional validation data
New kit (AGDIA kit currently being

New kit (AGDIA kit currently being evaluated INRA & UNIBA)

**Inclusion of NGS, nested PCR...** 

Need for guidance on identification of vectors (separate protocol).



othing is Written

In Stone



#### CONCLUSIONS

Research projects are essential for Standard setting activities in particular through:

- the development and validation of new tests,
- the optimization of tests or the production of additional validation data.

#### But also to

 ensure that laboratories implement the tests as expected (with e.g. organization of proficiency tests or training of laboratories)







EPPO's achievements is only possible thanks to the collaboration of experts from our region but also from other parts of the world

Thanks to all!



