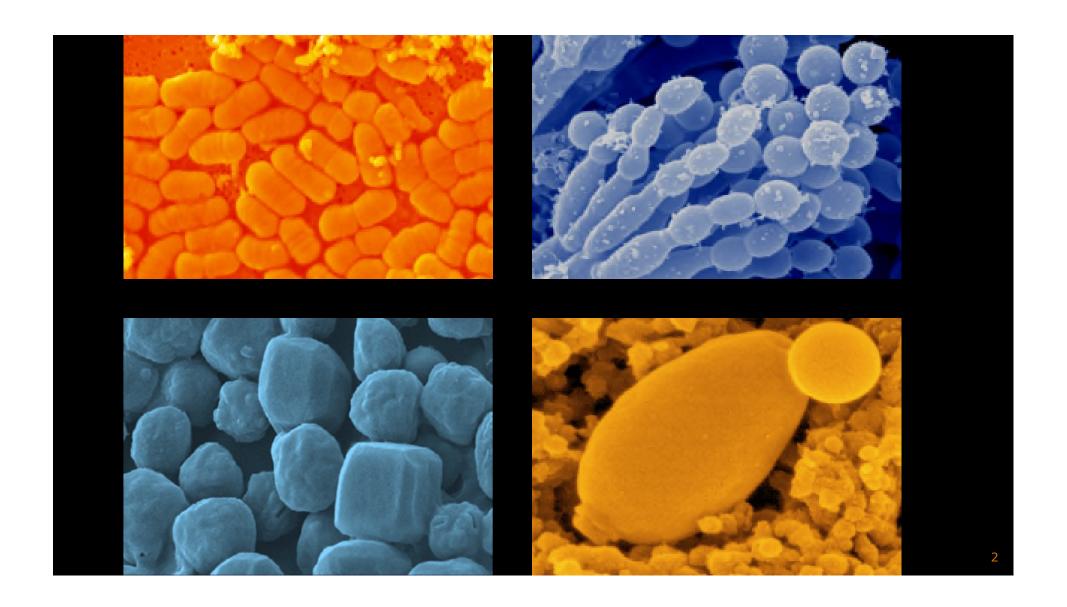


Pier Sandro Cocconcelli- EFSA FEEDAP Panel







### MICROORGANISMS INTENTIONALLY USED IN THE FOOD CHAIN

- More than 100 bacterial species
  Arthrobacter globiformis ...... Streptococcus thermophilus
- Filamentous fungi

  Fusarium solani ..... Trichothecium domesticum
- ■Yeasts

  Candida famata ......Williopsis mrakii
- ■Virus

  Bacteriophages Baculovirus -Potovirus





### OUTLINE

Characterisation of production strains of additives obtained by microbial fermentation:

- options for change
- future challenges
- Opportunities
- General principles
  - Regulatory Frameworks
  - Focus on risks
  - Most updated Risk Assessment approaches





### **Taxonomical identification**

- Key aspect of the safety assessment
  - QPS or non-QPS
  - Virulence/Toxigenic potential
- Accurate and update identification
  - Molecular Taxonomy (e.g. 16S rRNA gene ITS)
  - Species dependent approach
  - Whole Genome Sequence (phylogenomics)
- Re-classifications, synonymous and new names





### Strain Identification

- Unique identification of microbial products
  - Viable strains
  - Additive producers
- Essential for the safety assessment
- Critical for a safe lineage assessment (e.g. *E. coli* K12 derivatives)
- DNA based methods
  - PFGE PCR based (e.g. RAPD, REP, ERIC, AFLP)
  - Sequenced based (e.g. MLST)
  - Whole Genome Sequence (e.g. SNPs)





## **Toxigenic potential/Virulence**

- QPS:
  - waive for tox studies for consumer safety
  - no tolerance studies with target animal species
- Qualifications
  - Absence of toxigenic potential
  - Lack of acquired AMR genes
  - End use (viable vs production purposes)
- QPS + genetic modification:
  - Genetic modification = no safety concern
  - Alignment with food enzymes





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### Non-QPS:

- History of safe use:
  - Limitations
  - Scientific substantiation
- Criteria
  - Known toxigenic potential
  - Literature searches
  - Whole Genome Sequence
    - Annotation
    - Analysis

Metabolite	Organism
Aspergillomarasmine	A. oryzae
Cyclopiazonic acid	A. oryzae
Kojic acid	A. oryzae
Malformins	A. niger
Malformin A	
Malformin C	
Maltoryzine	A. oryzae
Naphtho-r-pyrones	A. niger
Aurasperone D	
Nigerazine B	A. niger
Nigragillin	A. niger
3-Nitropropionic acid	A. oryzae
Ochratoxin A	A. $niger$
Oxalic acid	A. niger
Trichodermin	T. reesei
Violacetin	A. oryzae

Blumenthal, 2004. Reg. Toxicol. Pharmacol. 39: 214-228





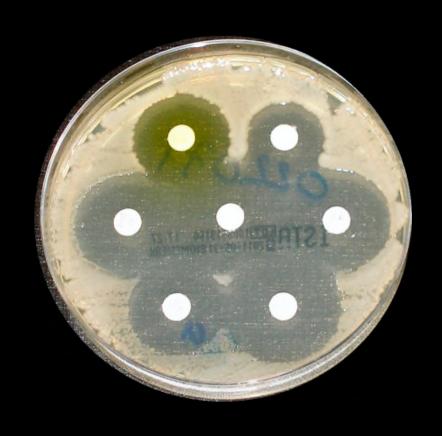
## Safe Strain Lineage

- On-going procurement on safe strain lineages
  - 36 Bacterial species
  - 42 fungal species
- Extended Literature Review
- Genomic Data

283318-2015: Italy-Parma: OC/EFSA/FEED/2015/01 — Database on the taxonomical identification and potential toxigenic capacities of non-QPS production strains of industrially produced food and feed additives

Publication date:	12-08-2015	Deadline:	15-10-2015
Document:	Contract notice		
Authority name:	European Food Safety Authority (EFSA)		

# **Antimicrobial susceptibility**









### **Antimicrobial resistance**



EFSA Journal 2012;10(6):2740

#### SCIENTIFIC OPINION1

Guidance on the assessment of bacterial susceptibility to antimicrobials of human and veterinary importance<sup>2</sup>

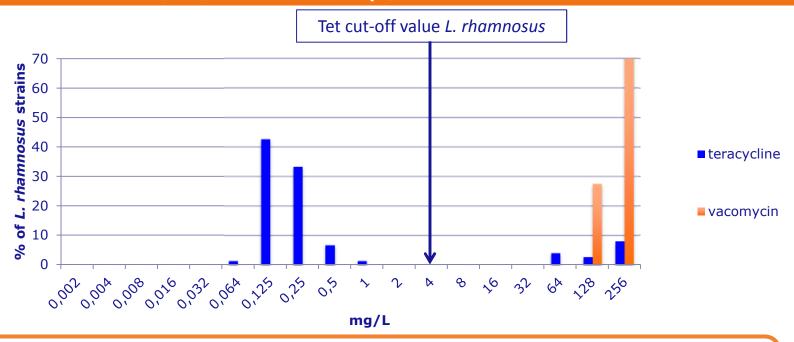
EFSA Panel on Additives and Products or Substances used in Animal Feed (FEEDAP)<sup>3,4</sup>

European Food Safety Authority (EFSA), Parma, Italy





# **INTRINSIC** vs ACQUIRED



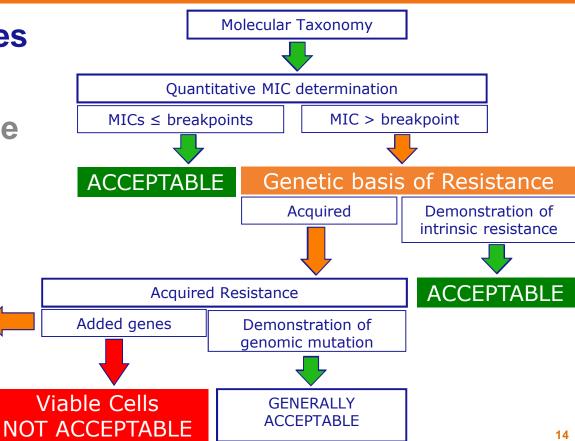
- Where all strains within a given taxonomic group show phenotypic resistance to an antimicrobial, such resistance can be considered intrinsic to the taxonomic group
- Molecular basis of intrinsic resistance are generally well described.





- Microbial Additives
  - Viable cells
  - Absence of viable cells

Production strain: Absence of DNA in the products









Minutes of the 102<sup>nd</sup> Plenary Meeting Held on 4-6 March 2014, Parma

#### Annex II

Update on the requirements for the assessment of additives produced by fermentation

- If atypically antibiotic resistance is detected, it should be demonstrated that the DNA from the production strain is not present in the final product.
- This can be done by PCR targeting known DNA sequences coding for the resistance or other sequences specific to the production strain.
- The specificity of the target sequence should be demonstrated







Minutes of the 102<sup>nd</sup> Plenary Meeting Held on 4-6 March 2014, Parma

#### Annex II

Update on the requirements for the assessment of additives produced by fermentation

### Demonstration of the absence of DNA by PCR

- Sequence to target (specificity, length)
- Methods (PCR qPCR)
- Controls (for PCR, for DNA extraction)
- Limits of detection / thresholds





# Susceptibility testing

Phenotypic analysis

Inclusion of new species, including production species (e.g. C. glutamicum)

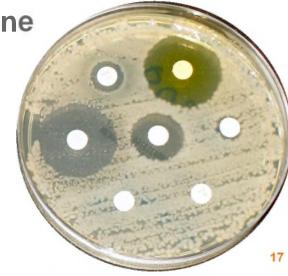
Revisit lists of antibiotics (e.g. colistine

and ciprofloxacin for *E. coli*)

Updated cut-off values

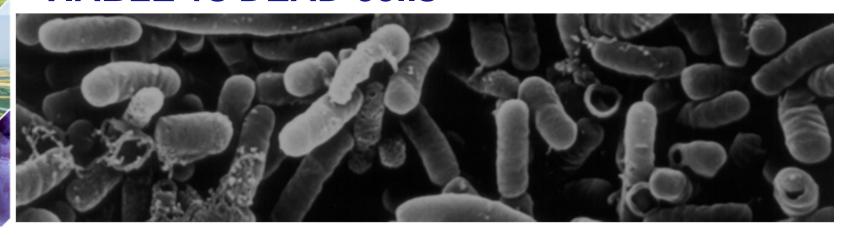
Whole Genomic Sequence data for resistance mechanisms

Presence/absence of DNA









- Critical point for the Risk assessment of GMM and non-GMM
- Definition the amount of product to be tested
- Resuscitation approaches
- Background/contaminant microbiota





### **Genetic modifications**

- Most common cause of inconclusive opinions
- New options and methodologies not well covered in the current Guidance Document
- Reiterated questions on aspects not well clear
- New assessment criteria, based on experience





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### **Current GMM Guidance Document:**

- From the GMO Panel
- GM food and feed
- GM fermentation products | Food additives

Food enzymes
Food additives
Feed additives

### New Guidance Document for GM fermentation products:

- From the FEEDAP Panel
- Based on experience
- Harmonised among Panels (FEEDAP, CEF)