Investigation into experimental toxicological properties of plant protection products having a potential link to Parkinson’s disease and childhood leukaemia

EFSA Panel on Plant Protection Products and their Residues (PPR)

European Food Safety Authority (EFSA)

Abstract

In 2013 EFSA published a literature review on epidemiological studies linking exposure to pesticides and human health outcome. As a follow up, the PPR Panel was requested to investigate the plausible involvement of pesticide exposure as a risk factor for Parkinson’s disease (PD) and childhood leukaemia (CHL). A systematic literature review on Parkinson’s disease and Childhood Leukaemia and mode of actions for pesticides was published by EFSA in 2016 and used as background documentation. The Panel used the Adverse Outcome Pathway (AOP) conceptual framework to define the biological plausibility in relation to epidemiological studies by means of identification of specific symptoms of the diseases as AO. The AOP is combining multiple information and provides knowledge of biological pathways, highlight species differences or similarities, identifies research needs and support regulatory decisions. In this context, the AOP approach could help in organizing the available experimental knowledge to assess biological plausibility by describing the link between a molecular initiating event (MIE) and the AO through a series of biologically plausible and essential key events (KEs). As the AOP is chemically agnostic, tool chemical compounds were selected to empirically support the response and temporal concordance of the key event relationships (KERs). Three qualitative and one putative AOP were developed by the Panel. Based on the results obtained, the Panel supports the use of the AOP framework to scientifically and transparently explore the biological plausibility of the association between pesticide exposure and human health outcomes, identify data gaps, define a tailored testing strategy and suggest an AOP’s informed Integrated Approach for Testing and Assessment (IATA).

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Summary

The European Food Safety Authority (EFSA) asked the Panel on Plant Protection Products and their Residues (PPR Panel) to develop a Scientific Opinion investigating experimental toxicological properties of plant protection products having a potential link to Parkinson’s disease and childhood leukaemia.

Following a significant association between pesticide exposure, Parkinson’s disease (PD) and childhood leukemia (CHL) as reported in an external scientific report of EFSA (Ntanzi et al. 2013), the PPR Panel analysed the plausible involvement of pesticides exposure as a risk factor in the pathogenesis of these two diseases. This task is required due to the intrinsic weakness of epidemiological studies that do not allow firm conclusions on causal relationships, but still raise a concern and open a question on suitability of regulatory studies to inform on specific and complex human health outcomes.

In addition to epidemiological studies, experimental data have also provided evidence for neurotoxic effects and biologically plausible mechanisms linking pesticides to PD. Quite contrary, scarce experimental and mechanistic evidence support the association between pesticide exposure and paediatric leukaemia.

The definition of biological plausibility in relation to epidemiological studies, taking into account experimental studies when provided, was achieved by organizing and analysing systematic literature review and the available toxicological data of pesticides active substances in the Adverse Outcome Pathways (AOP) conceptual framework according to OECD criteria (2013, 2014).

An AOP describes the chain of events leading from the first interaction of any chemical with a target (molecular initiating event = MIE) to an adverse outcome (AO), an apical endpoint in accepted regulatory toxicity testing. As such, AOPs are not chemical specific and will not be used to specifically address the issue of linking exposure to a pesticide found to be associated to PD or paediatric leukaemia in epidemiological studies. Rather, the AOP framework will assess the plausibility –if any- that pesticides pose a hazard in the pathogenesis of PD or paediatric leukaemia (i.e. CHL and infant leukaemia –IFL).

According to the OECD guidelines, MIE and AO are sequentially linked by a series of biologically plausible and essential key events (KEs) and their relationship (KERs, key event relationships) should be concordant on dose response, temporality and incidence. The availability and robustness of quantitative experimental data classifies the strength, in a codified assembly of weight of evidence, of the developed AOP. Putative AOPs are based on a hypothesized sequence of KE and KERs supported by biological plausibility and/or statistical inference; qualitative AOPs include assembly and evaluation of the supporting weight of evidence; quantitative AOPs are supported by quantitative relationships and/or computational models that allow quantitative translation of key event measurements into predicted probability or severity of AO.

The Panel adopted a tiered approach to design representative AOPs to evaluate biological plausibility. The starting point was the identification of a sequence of events able to (i) capture complex diseases like PD and CHL in a form of an AOP and (ii) describe the hazard of toxicants. Most relevant requisite was to identify a defined symptom for each disease equivalent to an AO for toxicants, reproducible in animal models, and possibly associable to a defined and measurable regulatory apical endpoint also triggered by chemicals in the regulatory or investigative studies. For PD, the application of the above rationale led to the identification of parkinsonian motor symptoms, i.e. the typical motor deficit observed in humans and in experimental conditions, as an AO representative. As a consequent step, pathological processes relevant to PD progression during adult life, for which there was evidence that they were triggered by chemicals in experimental models, were selected. The choice was based both on a systematic literature review commissioned by EFSA (EN-955, 2016) and on expert knowledge. Chemicals selected from the literature as prototypes to build AOPs relevant for PD were:

1. MPTP, supported by human poisoning data as well as experimental animal data.
2. Rotenone, supported by experimental animal data and a well characterized molecular target; the mitochondrial complex I for which human evidence of the involvement in PD exists.
3. Parathion, being the only pesticide individually associated to PD in epidemiological studies and for which experimental animal data exist.

In line with the selected AO and chemicals and based on the literature retrieved, two qualitative AOPs relevant for PD were built. Two MIEs, binding to mitochondrial complex I and initiation by a chemical of a redox cycling process, were defined. Those MIEs lead to parkinsonian’s motor deficit converging in a sequence of consequent KEs (summarized as mitochondrial dysfunction, impaired proteostasis, degeneration of dopaminergic neurons of the nigrostriatal pathway). Through a detailed analysis of the KERs the strength of association was judged by a weight of evidence approach based on modified Bradford-Hill criteria (i.e. based on biological plausibility, essentiality and empirical support of linkage, quantitative understanding of the linkage, evidence supporting taxonomic applicability and evaluation of uncertainties and inconsistencies). The overall weight of evidence indicates a strong link between the identified MIEs and the AO in the AOPs relevant for PD.

For CHL, the Panel adopted the same rationale as used for PD, supported both by a systematic literature review commissioned by EFSA (EN-955, 2016) and expert knowledge. In this case it became apparent that the term ‘childhood leukaemia’ used in epidemiological studies is general and does not distinguish between infant and childhood leukaemia (IFL) or other forms of pediatric leukaemia. Although both diseases share in utero exposure to relevant environmental risk factors for the development of the disease, they display distinct pathological pathways. Furthermore, while for CHL the Panel was not able to identify tool chemicals able to induce the disease in the experimental models, for IFL enough evidence supported the applicability of the anticancer drug etoposide as a tool. Symptoms and signs of overt paediatric leukaemia were chosen as AO, although the disease as such is not an apical endpoint in the regulatory toxicity studies. Taking into account the above limitations, it has been considered scientifically acceptable to develop a qualitative AOP relevant for IFL and to design only a putative AOP for CHL. The development of these two different AOPs, also in comparison to AOPs relevant for PD, allowed evaluating the flexibility of such an approach. In line with the selected AO and the prototype chemical etoposide for IFL, a MIE ‘in utero topoisomerase II poisoning’ was defined. It was linked to the selected AO through a single KE summarized as ‘in utero MLL chromosomal rearrangement’. The overall weight of evidence suggests that the link between the MIE and the AO is strong and that the proposed events can be used to explore the IFL-triggering hazard of chemicals. As stated, the AOP developed for CHL is based on weaker biological plausibility. However, a hypothetical biological plausibility could exist but cannot be convincingly formulated with the currently available circumstantial information: although epidemiological observations suggest that the association of the disease to in utero exposure to pesticides, complexities in defining a definite MIE and involvement of modulating factors as well as limitations in the standard design of regulatory studies for the exploration of tumour-related endpoints following in utero exposure prevent building a convincing qualitative AOP. In addition, the Panel recognises that an animal model recapitulating the disease is not available and this is also weakening the assessment.

Based on the results obtained, the Panel supports the use of the AOP framework to scientifically and transparently explore the biological plausibility of the epidemiological association between pesticide exposures and human health outcomes. Moreover, pesticides affecting the proposed AOPs should be considered as potentially hazardous with respect to the development of analysed diseases, considering the power of the AOP framework to provide at its best quantitative knowledge of biological pathways leading to an AO on a weight of evidence base.

Although the AOP developed in the present Scientific Opinion only explain a small fraction of the supposed interactions of pesticides, PD and paediatric leukaemia risk, the Panel considered the outcome of this approach promising. Thus, a multitude of AOPs might be developed to investigate the potential link of various pesticides to the different symptoms of the considered diseases. Beside this very relevant point, the AOP framework also represents a suitable scaffold to help identifying data gaps by analysing the weight of evidence for each KER within the defined AOPs. In addition, by suggesting and providing quantitative and measurable markers for critical biological events leading to the development of an AO, the AOP framework may help in the revision of regulatory studies underlining any limitation in the appropriate identification of effects and mode of actions relevant to complex human diseases, PD and paediatric leukaemia in the specific investigated case.
Summarising, the application of an AOP represents a transparent and weighted approach to define and map the causal linkages between key biological processes (MIE and KEs) to an AO that represents an apical endpoint in accepted regulatory toxicity testing. The design of an AOP, according to the OECD guidelines, identifies data gaps and provides information on the best approach to be adopted to investigate a defined toxicity pathway (representative of a relevant pathway of complex human diseases) This helps in identifying data gaps and in tailoring a tiered testing strategies for hazard identification and characterization. When quantitative, an AOP would define a threshold able to trigger the sequence of KEs from the MIE to the AO. Because the AOP process as such is ‘chemically agnostic’, it provides indication of the biological plausibility of a hazard.

Based on these considerations, the contribution of the AOP concept has been evaluated by designing a strategy based on the two AOPs relevant for PD due to their strong weight of evidence and the richness of experimental data. In assessment of risk, the AOP framework cannot be used as a stand-alone procedure but should inform an Integrated Approach for Testing and Assessment (IATA) scheme, integrating the chemical specific toxicokinetic properties. This will enhance confidence that the threshold of activation linking the MIE to the AO indeed triggers the cascade and by this way supports the regulatory process.
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KE1: In utero MLL chromosomal translocation..............................................................................

Adverse Outcome (AO) Infant leukaemia....................................................................................

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1. Introduction

1.1. Background and Terms of Reference as provided by EFSA

According to Regulation (EC) No 1107/2009 on placing of plant protection products on the market, applicants submitting dossiers for approval of active substances shall provide "scientific peer-reviewed open literature [...] on the active substance and its relevant metabolites dealing with side-effects on health [...] and published within the last ten years before the date of submission of the dossier". This should include epidemiological studies, as explicitly listed in Commission Regulation 283/2013 setting out the data requirements for active substances.

In 2013 EFSA published an external scientific report carried out by the University of Ioannina Medical School in Greece on a literature review linking exposure to pesticides and human health effects based on a systematic review of epidemiological studies published between 2006 and 2012 (Ntanzi et al. 2013, EFSA 2013:EN-497). This report summarises the association between pesticide exposure (assessed by different methods) and 23 major categories of human health outcomes. A statistically significant association was observed through fixed and random effect meta-analyses between pesticide exposure and the following health outcomes: liver cancer, breast cancer, stomach cancer, amyotrophic lateral sclerosis, asthma, type II diabetes, childhood leukaemia and Parkinson’s disease.

The results from the meta-analysis of the two latter health outcomes were supported by similar findings in previously and subsequent published studies (additional information 1.4.2 and 1.4.4).

Despite the large volume of available research data and the large number (>6,000) of analyses, firm conclusions could not be drawn for the majority of the outcomes studied. This observation is in line with previous studies on environmental epidemiology and in particular on pesticides which all acknowledge that such epidemiological studies generally suffer from many methodological limitations and large heterogeneities in their conduct. Also, due to the generic terms used for the pesticides assessed in the epidemiological studies, no information could be retrieved on specific pesticides.

In addition, the involvement of pesticide exposure in relation to the etiology of most of the health outcomes reported by Ntanzi et al. (2013) is unknown, and is likely to be influenced by environmental, lifestyle and genetic factors, which may add to the complexity of the interpretation of both epidemiological and experimental data. Consequently, the use of epidemiological studies and their integration in regulatory risk assessment is representing a major challenge for scientists, risk assessors and risk managers and the impact of these studies in regulatory risk assessment is still limited.

Nevertheless, the findings observed in the Ntanzi et al. report raise the question on whether the available experimental data and information on mechanisms of toxicity of pesticides can support these observations and if the regulatory risk assessment carried out to authorise the placing of plant protection products on the market covers the hazard assessment of pesticides with regard to these diseases.

The evaluation of the methodological limitations identified in epidemiological studies included in the Ntanzi et al. report is outside the scope of the mandate and will be addressed in a follow-up mandate.

1.2. Terms of Reference

The PPR Panel is requested to prepare a Scientific Opinion investigating experimental toxicological properties of plant protection products having a potential link to Parkinson’s disease and childhood leukaemia based on the findings in the Ntanzi et al. report (2013). This opinion will:

- Review the available data in the open literature and in regulatory toxicological data of pesticide active substances for which a potential link with a Mode of Action (MOA) relevant for the Parkinson’s disease and childhood leukaemia is known to exist to:
  - Develop a prototype to assess the risk factor by using the principles established for adverse outcome pathways (OECD, 2013).
  - Analyse the plausible involvement of pesticide exposure as a risk factor for the development of Parkinson’s disease and childhood leukaemia.
- Evaluate if, how and to what extent the experimental toxicity studies on mechanisms of toxicity cover effects and modes of action that are relevant to Parkinson’s disease and childhood leukaemia and are in line with the adverse outcome pathways.

- Make recommendations to address eventual data gaps for assessing the link between pesticide exposure and Parkinson’s disease and childhood leukaemia, and potential weaknesses in the current regulatory dossiers in supporting the hazard assessment of pesticides with regard to these diseases.

### 1.3. Interpretation of the Terms of Reference

In the Terms of Reference EFSA has requested a Scientific Opinion on investigating experimental toxicological properties of plant protection products having a potential link to Parkinson’s disease and childhood leukaemia. The terms of reference further elaborates that the PPR Panel should develop Adverse Outcome Pathways (AOP)’s for these two diseases on the basis of a systematic literature review and the available toxicological data of pesticide active substances.

The Adverse Outcome Pathways framework facilitates functional understanding of complex biological systems and the pathways of toxicity that results in adverse outcomes (AO). The AOP has a broader scope than the WHO IPCS mode of action (MoA) concept, which illustrates how to organize and apply mechanistic information on chemical’s MoA to understand human relevance of animal data (Meek et al. 2014). In this perspective, the MoA is chemical specific while the AOP is not.

The methodology provides a framework to collect and evaluate relevant chemical, biological and toxicological information in such a way that it is useful for risk assessment (OECD, 2013). The OECD has incorporated the IPCS framework on MoA in its guidance document on developing and assessing AOPs (Handbook series no. 184) in order to evaluate the biological plausibility of the relationships between the identified key events. These key events must be experimentally measurable and causally linked to the AO, which is usually associated with the findings of an in vivo OECD test guideline. The AOP identified must not contradict any steps of normal biological processes since they need to be biologically plausible.

The human relevance of the MoA framework has been applied in a number of specific case studies on compounds with a focus on quantitative time- and dose-response relationships. Modified Bradford Hill’s criteria for a causal relationship in epidemiological studies are also applied to the AOP concept as a critical foundation for overall weight of evidence evaluation. Therefore, if data are available, the causative link between the identified molecular initiating event (MIE), intermediate key events and final adverse outcome should be described in a quantitative manner, thus increasing the confidence for use in the regulatory context.

For the scope of this scientific opinion, any AOP (e.g., putative, qualitative and/or quantitative) will be useful for hazard identification or priority setting for further testing and development. The Panel understands that the ToR does not encompass full risk assessment (i.e. exposure assessment) of pesticides potentially involved in the diseases. Thus, the opinion will neither address specific exposures to pesticides found to be associated to Parkinson’s disease and childhood leukemia in epidemiological studies, nor consider exposure scenarios of specific active substance and their uses as specified in dossiers submitted for EU approval and the subsequent evaluation.

The ToR, instead, addresses the potential uses of the AOP concept in the regulatory risk assessment including the definition of biological plausibility in relation to the epidemiological studies. The mandate is intended to support the future hazard assessment of pesticides; thus, the AOPs will describe the biological plausibility and essentiality for the identified MIE and its relationship with intermediate key events leading to a defined AO. For the empirical support the panel will use data obtained from experimental studies of tool chemicals to establish concordance on dose response, temporality and incidence within the AOP scheme. The mandate will also analyse to what extent the available experimental toxicity studies cover the identified pathways of toxicity that are relevant for the development of the two diseases. Furthermore, the potential gaps of knowledge and uncertainties in the current pesticide data requirements and dossiers will be identified.

By making its evaluation the Panel realized that the health outcomes from the epidemiological studies were not distinguishing between parkinsonan disorders and Parkinson’s disease; and between...
childhood leukaemia and infant leukaemia. Conversely, the Panel addressed more specific health
outcome i.e. parkinsonian motor deficit, childhood leukaemia and infant leukaemia.

In the context of this Scientific Opinion, the Panel made use of exposure data only to quantitatively
understand the concentration at the target site able to trigger the sequence of events up to the AO.

In conclusion, according to the ToR the opinion will:

1. Analyse the plausible involvement of pesticide exposure as a risk factor for the development of
   Parkinson’s disease, childhood and infant leukaemia on the basis of adverse outcome pathways for
   these diseases.

2. Use AOP as guidance to evaluate, if the experimental toxicity studies on mechanisms of toxicity
   cover effects and modes of action relevant to Parkinson’s disease and childhood/infant leukemia.

3. Develop a prototype approach to assess pesticides as risk factors for complex diseases by using
   the principles (OECD, 2013) established for adverse outcome pathways.

1.4. Additional information

This chapter is intended to inform the reader on:

- data requirements for pesticide approval in regard to the hazards associated with neurotoxicity,
  carcinogenicity and haematology as they are expected to include apical endpoints relevant for the
  diseases considered in this opinion

- a summary of the epidemiological information linking exposure to pesticides and the diseases
  considered in this opinion

- an introduction to the adverse outcome pathway (AOP) conceptual framework

1.4.1 Data requirements for pesticide approval in regard to neurotoxicity

Previous data requirements under Directive 91/414/EEC concerning the placing of plant protection
products on the market:

Under Directive 91/414/EEC, in order to apply for the inclusion of an active substance in Annex I, a
dossier satisfying the requirements of Annex II has to be submitted.

The toxicological and metabolism requirements, listed in point 5 of the annex II of the directive should
permit to make a decision as to whether, or not, the active substance could be included in Annex I, to
specify appropriate conditions or restrictions of use, to classify the active substance as to hazard, to
establish relevant reference values as regard human health to perform risk assessment for man, to
identify relevant first aid measures.

- The need of such supplementary studies on the active substance (as indicated in point 5.8.2
  of annex II) must be made on a case by case basis, taking into account the results of the
  available toxicological and metabolism studies and the most important exposure routes.

- A specific data requirement is dedicated to delayed neurotoxicity (point 5.7). The test
  submitted should permit to evaluate if the active substance induces delayed neurotoxicity
  after acute exposure. Such test has to be performed for substances of similar or related
  structures to organophosphates.

Current data requirements under REGULATION (EC) No 1107/2009 concerning the placing of plant
protection products on the market and repealing Directives 79/117/EEC and 91/414/EEC:

Under REGULATION (EC) No 1107/2009 an active substance is approved at EU level, following assessment against a set of agreed criteria. Those criteria cover both the risks arising from the use of plant protection products which contain it as was already the case under Directive 91/4141/EEC but also the intrinsic properties of the active substance (i.e. an assessment of its hazard).

Indeed different categories of active substances are defined in REGULATION (EC) No 1107/2009 (active substances candidate for substitution, low risk active substances, basic substances) based on their hazard which impact the conditions of their approval.

Neurotoxicity among other criteria is taken into account to categorise active substances. In this way, an active substance:

- shall not be considered of low risk or as basic substance where it has neurotoxic effects (article 22 and 23)
- shall be approved as a candidate for substitution, if there are reasons for concern linked to developmental neurotoxic effects (article 24).


As was already the case under Directive 91/414/EEC, potential neurotoxic effects shall be carefully addressed and reported in routine required toxicological studies (acute toxicity studies point 5.2, short-term toxicity studies point 5.3, long term toxicity and carcinogenicity studies point 5.5 and reproductive toxicity studies point 5.6).

Compared to Directive 91/414/EEC, neurotoxicity requirements have been given more importance, the main differences are:

- in point 5.6.2 dedicated to developmental toxicity requirements, it is mentioned that information on developmental neurotoxicity may be required when such effects are indicated by observation in other studies or suspected based on the mode of action of the active substance.
- Point 5.7 is not restricted to delayed neurotoxicity requirements but includes both neurotoxicity in rodents (point 7.1) and delayed polyneuropathy studies (point 5.7.2).
  Regarding neurotoxicity in rodents, inclusion of neurotoxicity investigations in routine toxicity studies shall also be considered.

1.4.1.1 Triggers for neurotoxicity testing

The circumstances in which neurotoxicity studies should be performed are listed in Regulation (EU) No 283/2013.

- Specific neurotoxicity studies in rodents (point 7.1) shall be performed in case of one those following conditions:
  - there is indication of neurotoxicity in routine toxicity studies carried out with the active substance;
  - the active substance is a structurally related to known neurotoxic compound;
  - the active substance has a neurotoxic mode of pesticidal action.
- Delayed neurotoxicity studies shall be performed for active substances with similar or related structures to compounds capable of inducing delayed polyneuropathy such as organophosphates.
- Developmental neurotoxicity study may be performed when indication of such effects have been triggered in previous toxicity studies.

As a result, specific neurotoxicity studies are not routinely required for all pesticide active substances.
Triggers to perform those tests are well defined for acetylcholine esterase inhibitors for which delayed neurotoxicity studies are systematically carried out and pesticides with neurotoxic mode of pesticidal action for which at least acute neurotoxicity study in rodent has to be performed.

In other cases, specific neurotoxicity testing becomes obligatory only if neurotoxicity has been observed during organ toxicity testing or in case of structural analogy with a known neurotoxic compound. However, clear and consistent criteria to trigger submission of such data are still lacking and "routine" required in vivo toxicity studies may be not sensitive enough to alert on potential neurotoxicity.

The development of a neurotoxicity testing strategy including robust and reliable in vitro assays along with other alternative methods could be of value, as also raised as one of the main conclusions and recommendations in the EFSA opinion on Acetamiprid and Imidacloprid (EFSA 2013). Furthermore, understanding of toxicity mechanisms is given an increasing importance in risk assessment and therefore alternative methods including in vitro assays could also provide useful information on toxicity mechanisms involved.

### 1.4.1.2. Test guidelines – what do they cover

In the EU pesticides neurotoxicity testing for regulatory purposes is based on in vivo animal test methods. Commission Communication provides the list of test methods and guidance documents relevant to the implementation of Regulation (EU) No 283/2013.

In the table below, the test guidelines for neurotoxicity testing but also test guidelines of organ toxicity testing that can highlight neurotoxic effects are summarized (including principle of the assay, the clinical effects, the functional tests and the pathology examinations performed in regard to neurotoxicity).
<table>
<thead>
<tr>
<th>Neurotoxicity Study in Rodents</th>
<th>Test procedure</th>
<th>Detailed clinical observations</th>
<th>Functional tests</th>
<th>Pathology</th>
<th>Remarks</th>
</tr>
</thead>
<tbody>
<tr>
<td>OECD 424 (1997)</td>
<td>Animal: Rat young adults 20 (10M&amp;10F)/group</td>
<td>In the home cage and open field including: autonomic activity, body position, activity level, gait posture, reactivity to handling, placing or other environmental stimuli, presence of clonic or tonic movements, convulsions or tremors, stereotypes, behaviour, aggression, secretions, excretions</td>
<td>Sensory reactivity to different stimuli [auditory, visual, proprioceptive stimuli...]</td>
<td>At least 5M and 5F/group, perfused in situ and used for detailed neurohistopathology.</td>
<td>OECD GUIDANCE DOCUMENT FOR NEUROTOXICITY TESTING: In case of stand-alone study, the remainder of the animals may be used for specific neurobehavioural, neuropathological, neurochemical, electrophysiological procedures.</td>
</tr>
<tr>
<td></td>
<td>Exposure: Acute or 28 days, 90 days or chronic (1 year or longer)</td>
<td>As a standalone study or combined with repeated dose toxicity studies</td>
<td>Limb grip strength</td>
<td>Histopathology of representative sections of: - Brain forebrain, cerebrum, hippocampus, midbrain, cerebellum, pons, medulla oblongata, eye with optic nerve &amp; retina, - Spinal cord at the cervical and lumbar swellings, dorsal root ganglia, dorsal and ventral root fibres, - Peripheral nerve proximal sciatic nerve, the proximal tibial nerve and the tibial nerve calf muscle branches</td>
<td>If other data available on potential neurotoxicity (e.g. structure-activity, epidemiological data...) inclusion of more specialized tests of sensory and motor function or learning and memory to be considered.</td>
</tr>
<tr>
<td></td>
<td>Frequency depending on the duration of the study: - prior to first exposure - several times - at the end of the study</td>
<td>Motor activity measured with an automated device capable of detecting both decreases and increases in activity</td>
<td>Frequency depending on the duration of the study: - prior to first exposure - several times - At the end of the study</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Developmental Neurotoxicity Study</td>
<td>Animal: pregnant rats (at least 20 litters/group)</td>
<td>In the home cage and open field (see OECD)</td>
<td>Behavioural ontogeny Frequency: at least 2</td>
<td>Brain weights (PND 11-22 &amp; PND70)</td>
<td>Alternatively OECD 443 Extended One-Generation Reproductive Toxicity</td>
</tr>
</tbody>
</table>

Table 1: Neurotoxicity test guidelines
### OECD 426 (2007)

<table>
<thead>
<tr>
<th>Exposure: from GD6 to PND21. Study termination at PND 70</th>
</tr>
</thead>
<tbody>
<tr>
<td>At least 3 dose levels + control</td>
</tr>
<tr>
<td>20/sex (1/sex/litter)</td>
</tr>
<tr>
<td>Frequency depending on the duration of the study:</td>
</tr>
<tr>
<td>Pre-weaning: weekly</td>
</tr>
<tr>
<td>Adolescence: at least every 2 weeks</td>
</tr>
<tr>
<td>Young adults: at least every 2 weeks</td>
</tr>
<tr>
<td>Motor activity measures pre-weaning)</td>
</tr>
<tr>
<td>Frequency: 1-3 times (pre-weaning) once (young adults)</td>
</tr>
<tr>
<td>Motor and sensory function</td>
</tr>
<tr>
<td>Frequency: adolescence once (adolescence) once (young adults)</td>
</tr>
<tr>
<td>Learning and memory tests</td>
</tr>
<tr>
<td>Frequency: adolescence once (adolescence) once (young adults)</td>
</tr>
<tr>
<td>Neuropathological examination</td>
</tr>
<tr>
<td>(at PND 11-22 immersion or perfusion fixation and PND 70 perfusion fixation)</td>
</tr>
<tr>
<td>Morphometric evaluation</td>
</tr>
<tr>
<td>Representative sections of Brain: olfactory bulbs, cerebral cortex, hippocampus, basal ganglia, thalamus, hypothalamus, midbrain (tectum, tegmentum, and cerebral peduncles), pons, medulla oblongata, cerebellum).</td>
</tr>
<tr>
<td>In adults at study termination,</td>
</tr>
<tr>
<td>eye with optic nerve and retina</td>
</tr>
<tr>
<td>Spinal cord at the cervical and lumbar swellings, the dorsal and ventral root fibers, the proximal</td>
</tr>
<tr>
<td>Sciatic nerve, the proximal tibial nerve (at the knee), and the tibial nerve calf</td>
</tr>
</tbody>
</table>

Study could be carried out. In this guideline cohort is assigned to developmental neurotoxicity testing.
### Delayed Neurotoxicity of Organophosphorus Substances

**-Following Acute Exposure OECD 418 (1995)**

- **-28-day Repeated Dose Study OECD 419 (1995)**

<table>
<thead>
<tr>
<th>Animal: hen young adults</th>
<th>Behavioural abnormalities, Ataxia</th>
<th>Forced motor activity, such as ladder climbing</th>
<th>Biochemistry 24 &amp; 48 h after dosing 6 hens</th>
</tr>
</thead>
<tbody>
<tr>
<td>Acute exposure. 1 dose group &amp; vehicle control group &amp; positive control (TOCP) group</td>
<td>Frequency: immediately after treatment daily</td>
<td>Frequency: at least twice a week</td>
<td>Brain and lumbar spinal cord prepared and assayed for NTE activity</td>
</tr>
<tr>
<td>Exposure: 28 days. 3 dose levels + control</td>
<td></td>
<td></td>
<td>Histopathology 21D post-treatment (OECD 418) 14D post-treatment (OECD 419) 6 hens</td>
</tr>
</tbody>
</table>

**Repetitive dose 28-day oral toxicity study in rodents OECD 407 (2008)**

<table>
<thead>
<tr>
<th>Animal: Rat young adults 10 (5M&amp;5F)/group</th>
<th>In the home cage and open field (see OECD 424)</th>
<th>Sensory reactivity</th>
<th>Brain weight</th>
</tr>
</thead>
<tbody>
<tr>
<td>3 doses tested + 1 control group</td>
<td>Limb grip strength Motor activity</td>
<td>Frequency: once</td>
<td>Histopathology of representative sections of:</td>
</tr>
<tr>
<td>Exposure: 28 days</td>
<td>Frequency:</td>
<td>May be omitted when the study is conducted as a preliminary study to a subsequent subchronic</td>
<td>Brain (cerebrum, cerebellum and medulla/pons), Spinal cord</td>
</tr>
</tbody>
</table>

**Dedicated to organophosphorus compounds.**

NTE = neuropathy target esterase

OCP = tri-o-cresylphosphate
<table>
<thead>
<tr>
<th>Study Description</th>
<th>Animals</th>
<th>Dose and Grouping</th>
<th>Exposure</th>
<th>Frequency</th>
<th>Outcome Measured</th>
<th>Notes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Repeated dose 90-day oral toxicity study in rodents</td>
<td>Rat young adults</td>
<td>20 (10M&amp;10F)/group</td>
<td>90 days</td>
<td>weekly</td>
<td>no</td>
<td>Brain weight, Histopathology of representative sections of: Brain (cerebrum, cerebellum and medulla/pons), Spinal cord (at three levels: cervical, mid-thoracic and lumbar), Peripheral nerve (sciatic or tibial)</td>
</tr>
<tr>
<td>OECD 408 (1998)</td>
<td>3 doses tested + 1 control group</td>
<td>In the home cage and open field (see OECD424)</td>
<td>- prior to first exposure - weekly</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>Exposure: 90 days</td>
<td>(90-day) study</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Repeated dose 90-day oral toxicity study in non-rodents</td>
<td>generally Dog</td>
<td>8 (4M&amp;4F)/group</td>
<td>90 days</td>
<td>weekly</td>
<td>Sensory reactivity, Limb grip strength, Motor activity</td>
<td>Brain weight, Histopathology of representative sections of: Brain (cerebrum, cerebellum and medulla/pons), Spinal cord (at three levels: cervical, mid-thoracic and lumbar), Peripheral nerve (sciatic or tibial)</td>
</tr>
<tr>
<td>OECD 408 (1998)</td>
<td>3 doses tested + 1 control group</td>
<td>In the home cage and open field (see OECD424)</td>
<td>- prior to first exposure - weekly</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>Exposure: 90 days</td>
<td>(90-day) study</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
### Chronic Toxicity Studies

**OECD 452 (2009)**

<table>
<thead>
<tr>
<th>Animal: Rodent young adults</th>
<th><strong>In the home cage and open field</strong> (see OECD424)</th>
</tr>
</thead>
<tbody>
<tr>
<td>40 (20M&amp;20F)/group</td>
<td>Frequency:</td>
</tr>
<tr>
<td>Non rodent young adults 8 (4M&amp;4F)/group</td>
<td>- prior to first exposure</td>
</tr>
<tr>
<td>3 doses tested + 1 control group</td>
<td>- end of the first week</td>
</tr>
<tr>
<td>Exposure: 52 weeks</td>
<td>- then <strong>monthly</strong></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>functional deficits.</th>
<th>optionally for chemicals where previous repeated dose 28-day and/or 90-day toxicity tests indicated the potential to cause neurotoxic effects.</th>
</tr>
</thead>
</table>

<table>
<thead>
<tr>
<th>Brain weight</th>
<th>Histopathology of representative sections of:</th>
</tr>
</thead>
<tbody>
<tr>
<td>Brain (cerebrum, cerebellum and medulla/pons), Spinal cord (at three levels: cervical, mid-thoracic and lumbar), Peripheral nerve (sciatic or tibial).</td>
<td></td>
</tr>
</tbody>
</table>

| Alternatively OECD 453 Combined Chronic Toxicity/Carcinogenicity Studies. |
| Combined Chronic Toxicity/Carcinogenicity Studies could be carried out. |
1.4.2 Epidemiological studies linking pesticide exposure with Parkinson’s disease and Parkinsonism

The association between pesticide exposure and Parkinson’s disease (PD) has been investigated in numerous epidemiological studies. Priyadarshi et al. (2000) conducted the first meta-analysis on 19 studies published between 1989 and 1999 and found a positive and significant association between pesticide exposure and PD (OR 1.94; 95% CI 1.49-2.53), although with significant heterogeneity among studies. Further systematic reviews and meta-analyses conducted since then have lent support to this association (Breckenridge et al., 2016; Hernández et al., 2016a).

The EFSA external scientific report (Ntzani et al., 2013) reviewed thirty-two studies assessing the association between pesticide exposure and PD published between 2006 and 2012. Most of the studies (80%) involved occupational exposures where general pesticide use was assessed retrospectively by means of questionnaires. Only a minor proportion of studies was prospective in design (10%) or assessed exposure by biomonitoring techniques (particularly for the lipophilic organochlorines DDT and HCB, which represent 10% of the studies). The EFSA external scientific report performed meta-analyses for general pesticide use, DDT and paraquat exposures (which included 26, 5 and 9 studies, respectively). A significantly increased risk of PD was observed for exposure to pesticides in general, although with high heterogeneity (OR 1.49; 95% CI 1.28-1.73, random effect model) and for paraquat exposure (OR 1.32; 95% CI 1.09-1.60, fixed effect metaanalysis), which showed moderate heterogeneity. No significant association was observed for DDT. These results are in accordance with the largest studies carried out on the association between pesticide exposure and PD published from 2000 to 2013. The observed association between pesticides and PD holds true even though the latest meta-analyses were published considerably later, and contain a large number of additional data, relative to the earlier meta-analyses. This indicates consistency of results over time. Moreover, different methodologies used to synthesize the available evidence resulted in the same overall result.

Tanner et al (2011) performed a different kind of analysis in which pesticides were classified by presumed mechanism of toxic action rather than by functional categories or chemical class. Significant associations were found between PD and the use of pesticides grouped as ‘inhibitors of mitochondrial complex I’ or as ‘inducers of oxidative stress’, thus providing support in humans to findings from experimental studies. Use of rotenone, or any of the group of complex I inhibitors, was associated with PD (OR 2.5 and 1.7, respectively). An interesting sub-analysis, intended to provide evidence for temporal concordance, included only studies in which exposure to rotenone was documented up to 15 years before PD diagnosis, and an association of similar magnitude was still observed. Similarly, use of paraquat, or any of the group of oxidative stressors, was associated with PD (OR 2.5 and 2.0, respectively).

A further meta-analysis on 12 cohort studies published between 1985 and 2011 reported a combined OR of 1.28 (95% CI 1.03–1.59, random effects model), although with high heterogeneity and inconsistency among studies (van Maele-Fabri et al., 2012). The 28% increased risk did not vary substantially when omitting studies with extreme weight values, and the highest increased risks were observed for studies with a better design.

The last meta-analysis conducted so far (Breckenridge et al., 2016) found that most of the studies (88%) of pesticide exposure relied on self-reported pesticide use obtained either through personal interviews (49%) or by other methods. Despite an extensive effort to correct potential statistical artefacts (correcting for publication bias, stratifying by study characteristics, fixed and random effect models, etc.), the association between pesticide use and PD was statistically significant for this meta-analysis (OR 1.22; 95% CI 1.18–1.27 for fixed effects model and OR 1.56; 95% CI 1.37–1.77 for the random effects model). Use of herbicides or insecticides was associated with statistically significantly increased PD risk using the fixed effects model (OR 1.20 and 1.32, respectively). Similar results were obtained with the random effects model. High herbicide and high insecticide use were independently and significantly associated with an increased risk of PD; conversely, use of fungicides failed to be significantly associated with PD. Regarding paraquat use, a statistically significant association was found for PD (OR 1.69 and 1.47 using the fixed or random effects model, respectively). Moreover, a high paraquat use showed a significantly greater risk of PD as compared to non-use (OR 1.75; 95% CI 1.19–2.57, fixed effects meta-analysis). ORs for paraquat use, calculated using the fixed effects
model, were statistically significant regardless of interview type (in-person or other), method of paraquat use ascertainment (self-reported or other) and confounder adjustment (Breckenridge et al., 2016).

A relevant factor to take into account is that the use of personal protection measures and hygiene practices are important modifiers of the association between occupational pesticide exposure and PD, as these practices appear to reduce the risk of PD associated with the use of paraquat, permethrin and trifluralin (Furlong et al., 2015).

The vast majority of studies on PD are case-control in design, with the number of prospective cohort studies being much smaller. While almost all studies found a positive association between exposure to pesticides and PD, the association was not always statistically significant. A small number of studies found a negative association; however none of them reached statistical significance (Breckenridge et al., 2016). The consistency of the size of the effect (OR/RR) between meta-analyses combining case-control studies and cohort studies (particularly prospective cohorts) strengthens the hypothesis that exposure to pesticides may be an etiological factor of PD (van Maaë-Fabri et al., 2012). The overall appraisal of meta-analyses available so far suggests that there is sufficient evidence to conclude an association between pesticide exposure (broad definition) and PD, but not enough to support a causal relationship with specific pesticide classes or compounds.

The above observational studies on the relationship of PD and pesticides have intrinsic weaknesses, and their design does not allow conclusions on causal relationships. Limitations include the lack of an accurate exposure estimate (from both a qualitative and quantitative standpoint), the scarcity of information on dose-response relationships (which is difficult to achieve because of the long latency period of PD) and a lack of temporal concordance (most studies are case-control in design). A particular weakness is that exposure is not assessed for defined chemical entities, but rather for broad categories like "pesticide" or functional "classes of pesticides". Even when pesticide subgroups were used, they often provided no useful information and the subgroups herbicides and insecticides cannot be evaluated independently because most of the herbicide-exposed subjects were also exposed to insecticides. This fact is illustrated by the statistically significant correlations observed between ORs derived from the same studies, e.g. pesticide use and insecticide use (r = 0.82), pesticide use and paraquat use (r = 0.84), herbicide use and insecticide use (r = 0.66) and insecticide use and fungicide use (r = 0.90) (Breckenridge et al., 2016). Another general limitation is that subjects seldomly recall the specific class of pesticides used, and when doing so, such statements cannot be validated. The studies found in general that the risk of PD increases with longer exposure durations, but no other indications of a dose-response relation were found. It needs to be noted that environmental, lifestyle and genetic risk factors may exist that have not been corrected for in the epidemiological studies. For instance, allelic variants and single nucleotide polymorphisms (SNP) in certain genes (e.g. ABCB1 transporter (Narayan 2015), nitric oxide synthase (Paull 2016a)) can strongly affect the association of pesticide exposure and PD. Thus, effects of environmental chemicals may only get manifest on certain genetic backgrounds (Hernández et al., 2016b; Logroscino et al., 2005); in addition, different exposed populations might have unknown differences in the frequency of vulnerable genotypes... This adds a layer of uncertainty for the interpretation of the study data, in addition to the general limitations of study size (power). Concerning the latter, it has been argued that the inconsistency of findings in human populations regarding paraquat exposure and PD might be accounted for by the statistical variation of results in relatively small studies (Tanner et al., 2011).

More studies are needed to identify individual pesticides that might be associated with PD, in particular with prospective cohort design and with a better characterisation of exposure at the level of individual pesticides. While the available epidemiological studies support an association between pesticides and PD, complementary experimental research is needed to overcome the limitations inherent to those studies. The ultimate goal is that experimental and mechanistic data lend support and biological plausibility to the human epidemiological data.

### 1.4.3 Data requirements in the pesticide regulations for the exploration of carcinogenicity and haematological endpoints

Data requirements under REGULATION (EC) No 1107/2009 concerning the placing of plant protection products on the market.
Under REGULATION (EC) No 1107/2009 an active substance is approved at EU level, following assessment against a set of agreed criteria.

The required toxicological data should permit to identify the hazard of an active substance, to propose a classification according to CLP Regulation, to set relevant reference values as regard human health in order to perform risk assessment and to finally draw a conclusion as to whether, or not, the active substance could be approved with potential appropriate conditions or restrictions of use.

In routine required toxicological studies, all potentially adverse effects observed should be investigated and reported including genotoxicity, carcinogenicity and haematological endpoints.

Furthermore, in REGULATION (EC) No 1107/2009 active substances are categorized according to their intrinsic hazard, which impact the conditions of their approval.

Genotoxicity and carcinogenicity among other criteria are taken into account to categorise active substances. In this way, an active substance:

- shall not be approved if it is or has to be classified as mutagen category 1A or 1B or as carcinogen category 1A or 1B, in accordance with the CLP criteria (article 4 and annex II points 3.6.2 & 3.6.3)
- shall be approved as a candidate for substitution, if it is or has to be classified as carcinogen category 1A or 1B and has not be excluded (article 24 and annex II point 4)
- shall not be considered of low risk or as basic substance if it is or has to be classified as mutagen or as carcinogen (article 22 and annex II point 5).

For approval of pesticides under REGULATION (EC) No 1107/2009, the data requirements are set out in Regulation (EU) No 283/2013.

As regard to genotoxicity (point 5.4) and carcinogenicity (point 5.5) specific dedicated studies are routinely performed for all pesticide active substances.

As regard to haematological endpoints, they are investigated in the different repeated dose studies required (i.e. short term studies point 5.3, long term studies point 5.5).

Commission Communication provides the list of test methods and guidance documents relevant to the implementation of Regulation (EU) No 283/2013.

1.4.3.1 Genotoxicity testing:

The genotoxicity tests should address the three genotoxic endpoints, namely gene mutations, structural and numerical chromosome aberrations. The aims of the tests battery to be performed are to:

- predict genotoxic potential of active substances,
- identify genotoxic carcinogens at an early stage,
- elucidate the mechanism of action of some carcinogens.

In order to address the genotoxicity profile of pesticide substances a step-wise approach is followed with in vitro testing preceding in vivo testing.

First step: In vitro tests

The basic in vitro tests battery comprises two gene mutation tests (one in bacterial cells and one in mammalian cells) and a test investigating structural and numerical chromosomal alterations.

Studies to investigate gene (point) mutation:

- Bacterial Reverse Mutation Test (OECD TG 471)

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5 http://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:52013XC0403(02)&from=EN
In vitro Mammalian Cell Gene Mutation Tests Using the Hprt or xprt genes (OECD TG 476)
In vitro Mammalian Cell Gene Mutation Tests Using the Thymidine Kinase Gene (OECD TG 490)

Studies to investigate chromosome aberrations:
- In vitro Mammalian Chromosomal Aberration Test (OECD TG 473)
- In vitro Mammalian Cell Micronucleus Test (OECD TG 487)

For active substances harbouring structural alerts not detected by the standard test battery, specific tests investigating properly those alerts may be required.

Second step: In vivo tests
If all the results of the in vitro studies are clearly negative, at least one in vivo study is performed. The appropriate test to be conducted is an in vivo micronucleus assay.
If an equivocal or a positive test result is obtained in any in vitro test, the additional testing needed is considered on a case-by-case basis taking into account all relevant information.
In vivo tests performed should cover the genotoxic endpoint(s) identified as positive or equivocal in vitro and investigate appropriate target organs.

Studies to investigate gene mutations:
- Transgenic Rodent Somatic and Germ Cell Gene Mutation Assays (OECD TG 488)

Studies to investigate chromosome damage:
- Mammalian Erythrocyte Micronucleus Test (OECD TG 474)
- Mammalian Bone Marrow Chromosome Aberration Test (OECD TG 475)

Studies to investigate primary DNA damage:
- In vivo Alkaline Mammalian Comet assay (OECD TG 489)
- Unscheduled DNA Synthesis (UDS) Test with Mammalian Liver Cells in vivo (OECD TG 486)

In the table below, the test guidelines for the exploration of genotoxicity under Regulation (EU) No 283/2013 are summarized.
Table 2: Genotoxicity test guidelines

<table>
<thead>
<tr>
<th>Test guideline</th>
<th>Test system</th>
<th>Endpoints</th>
<th>Remarks</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Bacterial Reverse Mutation Test</strong></td>
<td>Strains of S.typhimurium TA1535; TA1537 or TA97a or TA97; TA98 TA100 and E.coli WP2 strains or S. typhimurium TA102</td>
<td>Detection of gene mutations substitution, addition or deletion, frame-shift and base-pair substitutions</td>
<td>First screening test Easy to use Very large data base of results available</td>
</tr>
<tr>
<td><strong>OECD 471 (1997)</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>In vitro Mammalian Cell Gene Mutation Tests</strong></td>
<td>HPRT: CHO, CHL and V79 lines of Chinese hamster cells, L5178Y mouse lymphoma cells, and TK6 human lymphoblastoid cells XPRT: CHO-derived AS52 cells</td>
<td>Detection of gene mutations including base pair substitutions, frame-shift, small deletions and insertions</td>
<td>XPRT (contrary to HPRT) may allow the detection of large deletions and possibly mitotic recombination due to its location on X-chromosome.</td>
</tr>
<tr>
<td><strong>HPRT or XPRT genes OECD 476 (2015)</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>In vitro Mammalian Cell Gene Mutation Tests</strong></td>
<td>L5178Y mouse lymphoma cells and TK6 human lymphoblastoid cells</td>
<td>Detection of gene mutations including point mutations, frame-shift mutations, small deletions.</td>
<td>Preference to the Mouse lymphoma assay (MLA) most commonly performed. Allows also detection chromosomal events (large deletions, chromosome rearrangements and mitotic recombination)</td>
</tr>
<tr>
<td><strong>TK gene OECD 490 (2015)</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>In vitro Mammalian Chromosomal Aberration Test</strong></td>
<td>Cell lines including Chinese Hamster Ovary (CHO), Chinese Hamster Lung V79, Chinese Hamster Lung (CHL)/IU, TK6 or primary cell cultures, including human or other mammalian peripheral blood lymphocytes.</td>
<td>Detection of chromosomes aberrations Chromatid- and chromosome-type aberrations should be recorded separately and classified by subtypes (breaks, exchanges)</td>
<td>Resource intensive, time consuming and good expertise required. Not appropriate to detect aneugens.</td>
</tr>
<tr>
<td><strong>OECD 473 (2014)</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>In vitro Mammalian Cell Micronucleus Test</strong></td>
<td>Various Human or rodent cell lines or primary cell cultures</td>
<td>Detection of both structural and numerical chromosome aberrations Can be combined with special techniques to additional mechanistic information e.g.: fluorescence in situ hybridisation (FISH)</td>
<td>Rapid and easy to conduct The only in vitro test that can efficiently detect both clastogens and aneugens.</td>
</tr>
<tr>
<td><strong>OECD 487 (2014)</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Transgenic Rodent Somatic and Germ Cell Gene Mutation Assays</strong></td>
<td>Transgenic rodents: Muta™Mouse</td>
<td>Detection of gene mutations base pair substitutions, frameshift</td>
<td>Allows detection of mutations in both somatic tissues and germ lines</td>
</tr>
<tr>
<td>Test Description</td>
<td>Species</td>
<td>Test Purpose</td>
<td>Additional Information</td>
</tr>
<tr>
<td>------------------</td>
<td>---------</td>
<td>--------------</td>
<td>------------------------</td>
</tr>
<tr>
<td>Mammalian Erythrocyte Micronucleus Test OECD 474 (2014)</td>
<td>Rodents (usually)</td>
<td>Detection of both structural and numerical chromosome aberrations</td>
<td>Can be combined with special techniques to additional mechanistic information e.g., fluorescence in situ hybridisation (FISH)</td>
</tr>
<tr>
<td>Mammalian Bone Marrow Chromosome Aberration Test OECD TG 475 (2014)</td>
<td>Rodents (usually)</td>
<td>Detection of structural chromosomal aberrations</td>
<td>Not designed for detection of aneuploidy</td>
</tr>
<tr>
<td><em>In vivo</em> Alkaline Mammalian Comet assay OECD 489 (2014)</td>
<td>Rodents (usually)</td>
<td>Detection of primary DNA damages</td>
<td>DNA single and double strand breaks</td>
</tr>
<tr>
<td>Unscheduled DNA Synthesis (UDS) Test with Mammalian Liver Cells <em>in vivo</em> OECD 486 (1997)</td>
<td>Rat (commonly used)</td>
<td>Detection of DNA repair</td>
<td>Sensitivity has been questioned.</td>
</tr>
</tbody>
</table>
1.4.3.2. Long-term toxicity and carcinogenicity testing

The aims of the long term toxicity testing are to:

- identify adverse effects resulting from long-term exposure to the active substance,
- identify target organs, where relevant,
- establish the dose-response relationship,
- establish the NOAEL and, if necessary, other appropriate reference points.

As for carcinogenicity testing, it shall permit to:

- identify carcinogenic effects resulting from long-term exposure to the active substance,
- establish the species, sex, and organ specificity of tumours induced,
- establish the dose-response relationship,
- identify the maximum dose eliciting no carcinogenic effect where possible,
- determine the mode of action and human relevance of any identified carcinogenic response where possible.

A long-term oral toxicity study and a long-term carcinogenicity study (two years) in rat are to be conducted; where possible these studies shall be combined. A second carcinogenicity study in mouse is to be conducted, unless it can be scientifically justified that this is not necessary. In that case, a scientifically validated alternative carcinogenicity model may be used instead of a second carcinogenicity study.

The relevant regulatory test guidelines are as follows:

- Carcinogenicity Studies (OECD TG 451)
- Chronic Toxicity Studies (OECD TG 452)
- Combined Chronic Toxicity/Carcinogenicity Studies (OECD TG 453)

1.4.3.3. Haematological endpoints:

No specifically dedicated study is required. However, haematological endpoints among other toxicological endpoints are systematically addressed in routine required repeated dose studies (short-term toxicity studies, long term toxicity and carcinogenicity studies point).

Haematological parameters are also to be investigated in the extended one-generation reproductive toxicity study while they are not part of the investigated endpoints of the two-generation reproductive toxicity study. Moreover, when warranted by available information, the extended one-generation study protocol can include a cohort dedicated to detailed investigation of developmental immunotoxicity.

The haematological parameters monitored in repeated dose studies are:

- Red blood cells parameters (haematocrit, haemoglobin concentration, erythrocyte count)
- Total and differential leucocyte count
- Platelet count
- Blood clotting time/potential

The relevant regulatory test guidelines are as follows:

- Short term studies:
  - Repeated dose 28-day oral toxicity study in rodents (OECD TG 407)
  - Repeated dose 90-day oral toxicity study in rodents (OECD TG 408)
  - Repeated dose 90-day oral toxicity study in non-rodents (OECD TG 409)
Long term/carcinogenicity studies:
  - Carcinogenicity Studies (OECD TG 451)
  - Chronic Toxicity Studies (OECD TG 452)
  - Combined Chronic Toxicity/Carcinogenicity Studies (OECD TG 453)

Reproductive toxicity study:
  - Extended One-Generation Reproductive Toxicity Study (OECD TG 443)

In the table below, the test guidelines for the exploration of carcinogenicity and haematological endpoints under Regulation (EU) No 283/2013 are summarized (including the test procedure, the haematological parameters investigated and the organs going through to histopathological examination relevant to pick up hematopoietic disorders).
### Table 3: Cancerogenicity test guidelines and hematological endpoints in the regulatory toxicological studies

<table>
<thead>
<tr>
<th>Test guideline</th>
<th>Test procedure</th>
<th>Haematology</th>
<th>Histopathology Organs of interest for leukaemia</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Repeated dose 28-day oral toxicity study in rodents</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>OECD 407 (2008)</td>
<td>Animals: Rat young adults 5 M &amp; 5F/group 3 doses tested + 1 control group Exposure: 28 days</td>
<td>Parameters: RBC parameters Total and differential leucocyte count, Platelet count Blood clotting time/potential Frequency: once at the end of the test period</td>
<td>Bone Marrow, Thymus, Spleen, LN, liver</td>
</tr>
<tr>
<td><strong>Repeated dose 90-day oral toxicity study in rodents</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>OECD 408 (1998)</td>
<td>Animals: Rat young adults 10 M &amp; 10F/group 3 doses tested + 1 control group Exposure: 90 days</td>
<td>Parameters: RBC parameters Total and differential leucocyte count, Platelet count Blood clotting time/potential Frequency: once at the end of the test period</td>
<td>Bone Marrow, Thymus, Spleen, LN, liver</td>
</tr>
<tr>
<td><strong>Repeated dose 90-day oral toxicity study in non-rodents</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Animals: Dog young adults 4M &amp; 4F/group</td>
<td>Parameters: RBC parameters</td>
<td>Bone Marrow, Thymus, Spleen, LN, liver</td>
</tr>
</tbody>
</table>
### OECD 409 (1998)

- **3 doses tested + 1 control group**
- **Exposure:** 90 days

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Total and differential leucocyte count, Platelet count, Blood clotting time/potential</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Frequency:</strong></td>
<td>- Prior to first exposure&lt;br&gt;- Monthly or midway&lt;br&gt;- At the end of test period</td>
</tr>
</tbody>
</table>

### Chronic Toxicity Studies

#### OECD 452 (2009)

- **Animals:** Rodent young adults<br>20M & 20F/group
- **Non rodent young adults:** 4M & 4F/group
- **3 doses tested + 1 control group**
- **Exposure:** 52 weeks

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Bone Marrow, Thymus, Spleen, LN, liver</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Total and differential leucocyte count,</strong> Platelet count, Blood clotting time/potential</td>
<td></td>
</tr>
<tr>
<td><strong>Frequency:</strong></td>
<td>- At 3, 6, and 12 months and at the end of test period</td>
</tr>
</tbody>
</table>

If the chemical has an effect on the haematopoietic system, reticulocyte counts and bone marrow cytology may also be performed although not routinely conducted.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>RBC parameters</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Frequency:</strong></td>
<td></td>
</tr>
</tbody>
</table>

**Note:** 24% of the dissatisfaction with the results is attributed to the lack of data.
### Carcinogenicity Studies

**OECD 451 (2009)**

- **Animals:** Rodent young adults
  - 50 M & 50F/group
- **Exposure:**
  - 104 weeks rat
  - 78 weeks mouse
- **3 doses tested + 1 control group**
- **Blood smears may also be prepared for examination, particularly if bone marrow is the target organ**
- **Frequency:**
  - At the end of test period
  - At the discretion of the study director
- **Bone Marrow, Thymus, Spleen, LN, liver**
- **Non neoplastic histopathological findings**
- **Neoplastic histopathological findings**

### Combined Chronic Toxicity/Carcinogenicity Studies

**OECD 453 (2009)**

- **Animals:** Rodent young adults
  - 50 M & 50F/group (carcinogenicity phase)
  - 10 M & 10F/group (chronic phase)
- **Exposure:**
  - 52 weeks rat (chronic phase)
- **3 doses tested + 1 control group**
- **Parameters:**
  - RBC parameters
  - **Total and differential leucocyte count,**
    - Platelet count
  - Blood clotting time/potential
  - **Min 10 M & 10 F/group**
- **If the chemical has an effect on the haematopoietic system, reticulocyte counts and bone marrow cytology may also be performed although not routinely conducted**
- **Frequency:**
  - Bone Marrow, Thymus, Spleen, LN, liver
  - Non neoplastic histopathological findings
  - Neoplastic histopathological findings
### Extended One-Generation Reproductive Toxicity Study

**OECD TG 443 (2012)**

<table>
<thead>
<tr>
<th>Parameters</th>
<th>At 3, 6, and 12 months and at the end of test period</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Total and differential leucocyte count</strong></td>
<td></td>
</tr>
<tr>
<td>RBC parameters</td>
<td></td>
</tr>
<tr>
<td>Platelet count</td>
<td></td>
</tr>
<tr>
<td>Blood clotting time/potential</td>
<td></td>
</tr>
<tr>
<td>Parents: all</td>
<td></td>
</tr>
<tr>
<td>Cohort F1A: <strong>10 M &amp; 10 F/group</strong></td>
<td></td>
</tr>
</tbody>
</table>

**Exposure:**
- **P:** 10 weeks (2w pre-mating, 2w mating 6 w post-mating)
- **F:** 6 weeks (in utero +pre-weaning) + 0 to 22 weeks according to cohorts.
- **F1A:** 6 weeks (in utero +pre-weaning) + 10 weeks

**Frequency:**
- Once at the end of the test period

**Cohort 1A:** Bone marrow + lymph nodes e of 10 M and 10 F: group

- Splenic lymphocyte subpopulation analysis (CD4+ and CD8+ T lymphocytes, B lymphocytes, and natural killer cells)
- Splenic lymphocyte subpopulation analysis (CD4+ and CD8+ T lymphocytes, B lymphocytes, and natural killer cells) ➔ to evaluate if exposure impacts immunological steady state distribution

**P and F1A:** spleen, liver and thymus all animals

**LN:** Lymph Nodes
1.4.3.4. Previous data requirements under Directive 91/414/EEC concerning the placing of plant protection products on the market

As detailed above, in the data requirements of the previous regulation 91/414/EEC genotoxicity, carcinogenicity and haematological endpoints where similarly mandatory to address.

In regards to genotoxicity testing, *Salmonella Typhimurium reverse* mutation test, *in vitro* mammalian cytogenetic test and *in vitro* mammalian cell gene mutation test were the only acceptable tests. Even when all *in vitro* tests were negative, one *in vivo* test was to be carried out being the micronucleus test (OECD 474). If indicated from the *in vitro*, further *in vivo* testing could be triggered being chromosomal aberration (475) or unscheduled DNA synthesis (486). Thus, the former data requirements were less comprehensive, in particular in regards to *in vivo* mutagenicity testing and for most of the *in vivo* genotoxicity tests, i.e. the *in vivo* bone marrow micronucleus test, proof of actual bone marrow exposure was often not shown but was only assumed. This is currently being critically assessed in each case during the re-assessment of the active substances.

In relation to carcinogenicity and haematological testing, the former data requirements were as the current except that extended one generation study (OECD 443) was not available and not required.

1.4.4. Epidemiological studies linking pesticide exposure with childhood leukemia

There is an increasing concern about chronic low-level pesticide exposure during pregnancy or childhood and its influence on childhood cancers. Epidemiological studies have suggested that maternal exposure to certain household pesticides during pregnancy may increase the risk of childhood leukaemia; however, these studies are limited because no specific pesticides were directly associated with the risk of leukaemia, but rather the broad term pesticide exposure (Lu et al., 2015).

The EFSA external scientific report (Ntzani et al., 2013) updated the meta-analysis conducted by Turner (2010) on residential pesticide exposure during pregnancy and found an increased risk of childhood leukaemia associated with exposure to unspecified pesticides (OR: 1.30; 95% CI: 1.06–1.56). When exposure was restricted to insecticides, a somewhat stronger association was observed (OR: 1.69; 95% CI: 1.35–2.11). In contrast, meta-analyses on studies examining preconception exposure failed to show statistically significant results. Ntzani et al. (2013) also updated the meta-analysis of Turner et al. (2010) on pesticide exposure during childhood and found a significant increased risk of childhood leukaemia (OR: 1.36; 95% CI: 1.19–1.55). In spite of these positive associations, the evidence must be carefully interpreted because most studies were of small size, exposure was assessed through non-validated self-reported questionnaires (that are prone to misclassification) and concern was raised on publication bias. Also, only few studies included data on leukaemia subtypes.

More recently, meta-analyses have been carried out on occupational and residential exposure to pesticides and risk of childhood leukaemia. Maternal occupational pesticide exposure during pregnancy and/or paternal occupational pesticide exposure around conception have indicated an increased risk of leukaemia in the offspring. Bailey et al. (2014) pooled data from 13 case-control studies participating in the Childhood Leukaemia International Consortium (CLIC) and found a significant increased risk of acute myeloid leukaemia (AML) in children born from mothers exposed to pesticides during pregnancy (OR: 1.94; 95% CI: 1.19–3.18), which is consistent with previous meta-analyses; however, no significant risk was found for paternal exposure around conception (OR: 0.91; 95% CI: 0.66–1.24). In relation to acute lymphocytic leukaemia (ALL), Bailey et al. (2014) observed a 20% increased risk with paternal exposure around conception (OR: 1.20; 95% CI: 1.06–1.38), which appeared to be more evident for children with T-cell ALL; however, no association was found between maternal exposure during pregnancy and risk of ALL (OR: 1.01; 95% CI: 0.78–1.30).

In a separate study investigating residential pesticide exposure, Bailey et al. (2015) pooled data from 12 case-control studies in the CLIC and found an increased risk of ALL associated with exposure to any pesticide shortly before conception, during pregnancy and after birth. The three exposure windows had essentially the same OR: 1.39 (95% CI: 1.25–1.55), 1.43 (95% CI: 1.32–1.54) and 1.36 (95% CI: 1.23–1.51), respectively. Little variation was found by time period, type of pesticide or among other subgroups. Regarding AML, an increased risk was found for exposure to any pesticide...
in the few months prior to conception (OR: 1.49; 95% CI: 1.02–2.16), and during pregnancy (OR: 1.55; 95% CI: 1.21–1.99); however, exposure after birth did not show a significantly increased risk (OR: 1.08, 95% CI: 0.76–1.53). The relative similarity in ORs between leukaemia types, time periods and pesticide types may suggest similar exposure patterns and effects across the time periods in ALL and AML, exposure to multiple pesticides or recall bias.

The meta-analysis conducted by Chen et al. (2015) found that children exposed to indoor but not outdoor residential insecticides had an increased risk of childhood leukaemia (OR: 1.47; 95% CI: 1.26–1.72). A significant association was also found for herbicide exposure during childhood (OR: 1.26; 95% CI: 1.10–1.44).

Almost all the available studies addressing pediatric leukaemia included both infant leukaemia and childhood leukaemia in the same diagnosis. Very few studies examined the risk of pesticide exposure with infant leukaemia (< 1 year) as a separate entity. The Brazilian Collaborative Study Group of Infant Acute Leukaemia found an increased risk of infant leukaemia in mothers exposed to domestic insecticides during pregnancy (OR: 2.18, 95% CI: 1.53–2.13) with a rather small samples size of 91 cases (Pombo de Oliveira et al., 2006). A further study also conducted in Brazil (Ferreira et al., 2013) found that ever use of pesticides during pregnancy was associated with ALL (OR: 2.10; 95% CI: 1.14–3.86) and AML (OR: 5.01; 95% CI: 1.97–12.7) in children <1 year of age. In particular, maternal exposure to permethrin was associated with a significantly higher risk of leukaemia in children <1 year of age (OR: 2.47; 95% CI: 1.17–5.25 for ALL; and OR: 7.28; 95% CI: 2.60–20.38 for AML).

Observational studies on pesticide exposure and paediatric leukaemia have important weaknesses to establish causal relationships. The consistency of findings across studies may be due to the considerable overlap in the studies included in the different meta-analyses carried out. Limitations include the lack of an accurate exposure estimate (from both a qualitative and quantitative standpoint), lack of temporal concordance (most studies were case-control in design) and little information on dose-response relationship. In addition, the sound epidemiological evidence available may be challenged by endogenous or exogenous factors, such as genetic polymorphisms, diet, lifestyle and co-exposure to other environmental agents. Hence, accounting for simultaneous exposure to multiple agents would help to delineate true associations, but this has not been possible for most of the available evidence because of difficulties in properly assessing multiple exposures. The question arises on whether, and to what extent, experimental and mechanistic data can lend support to the human data.

In evaluating the etiological role of environmental factors in the pathogenesis of childhood leukaemia, there is a need to know the evidence for an association between exposure to certain environmental factors and the incidence of the disease assessed by epidemiological studies. Furthermore, evidence from experimental research is also required to know the possible mechanisms that would explain an observed or hypothesised association between the exposure to certain environmental factors and the incidence of childhood leukaemia.

In observational studies the quality of exposure assessment is crucial, especially in deriving dose-response relations. Moreover, the reduction of bias and the adjustment for confounding factors are important in assessing the evidence for causality of associations. Because of the controversy regarding to the role of pesticide exposure in childhood leukaemia, a weight of evidence analysis based on Bradford Hill criteria was performed to evaluate the available scientific evidence linking pesticide exposure with childhood leukaemia (Health Council of the Netherlands. Childhood leukaemia and environmental factors. The Hague: Health Council of the Netherlands, 2012; publication no. 2012/33).

**Strength.** The observed associations between pesticide exposure through parental occupational exposure or residential exposure and childhood leukaemia are rather weak (OR/RR < 2–3) and not always statistically significant. However, the risk of misclassification of exposure, common to many epidemiological studies, leads to underestimation of the real risk, thus decreasing the strength of the association.

**Consistency.** Despite exposure is often not identical in most situations, almost all meta-analyses published so far showed a trend toward increased risk with minor differences. Overall, pesticide
exposure during pregnancy tends to support a causal relationship; however, many individual studies included in the different meta-analyses are largely the same ones.

Specificity. The aetiology of childhood leukaemia is multifactorial, resulting from the interplay of genetic or environmental factors. It is not possible to associate specific pesticide exposures with childhood leukaemia because of the low prevalence of this disease and the imprecise exposure assessment. On the other hand, pesticide exposure is associated with many other diseases. While most of the epidemiological studies evaluated are focussed on childhood leukaemia and other diseases/outcomes are usually not considered, this does not mean that other outcomes do not occur, simply studies were not designed to address them.

Temporal. When risk factors for childhood leukaemia are investigated in case-control studies, exposure is usually measured retrospectively, so temporality cannot be properly addressed like in prospective cohort studies. Besides, responder and recall bias, might influence the accurate timing of exposure. Many epidemiological studies have assessed exposure during pregnancy or even before (prior to conception) such that the risk factor precedes the development of the disease. Nonetheless, the time window at which pesticides might exert its causative action (prior to conception, during early, mid or late pregnancy or during childhood) is not clear. However, exposures during childhood appear to be less consistently associated with childhood leukaemia than exposures during pregnancy.

Biological gradients. Exposure-response relationships can only be assessed when exposure is measured adequately and with sufficient precision. However, exposure is often assessed using questionnaires, or at best with biomonitoring techniques on spot-samples. Accordingly, exposure assessment (and even accumulated exposure to individual chemicals) is difficult to perform and often poorly characterised. In the case of childhood leukaemia, an additional limitation is that exposure can occur at different stages of the development (early pregnancy, late pregnancy or postnatally) and effects of chemicals at each stage may be different. Additionally, children and their parents are exposed to mixtures of different agents, and chemical interactions are not usually studied as well as the potential combined effect to the same agent(s) between prenatal and postnatal stages.

Many of the epidemiological studies did not assess the risk of childhood leukaemia in response to the frequency or intensity of pesticide exposures. The only weak support for a positive exposure-response relationship found that the risk of leukaemia increased with the frequency of pesticide use (Van Maele-Fabry et al., 2011).

Biological plausibility. The growing experimental studies and animal models on the biology of childhood leukaemia show increased evidence for effects for chemicals, thus strengthening the biological plausibility of an association. However, there are no experimental models on specific pesticides (and hence no dose-response relationship) and the animals used failed to recapitulate all the features of the human disease. Besides, for pesticides and childhood leukaemia, the qualitative and quantitative evidence on the biological mechanisms underlying the first initiating events at molecular levels is lacking. Pesticides are biologically active molecules that may play some role in cancer aetiology. Consequently, in the European Union the use of pesticides showing some evidence of carcinogenicity or genotoxicity has been restricted or banned. Nevertheless, potential gaps in the regulatory studies, interspecies variability in target cells, and the use of co-formulants or potential epigenetic factors cannot be ruled out.

Regarding coherence, the cause and effect interpretation should not seriously conflict with the generally known facts of the natural history and biology of the childhood leukaemia. However, the natural history of this disease is far from being adequately understood, thus coherence cannot be properly assessed.

Another Bradford Hill criterion is analogy. This means that if it is known that the effect of one type of exposure can lead to childhood leukaemia, a similar effect from another type of exposure might also. However, the different variety of types of exposures associated to the disease (ionizing radiation, electromagnetic fields, chemicals other than pesticides) are of little help and prevents analogy from being a useful consideration in practice.

In addition to the Bradford Hill considerations, alternative explanations for epidemiological associations other than causality should be considered: chance, bias (specifically exposure misclassification) and confounding. If these are unlikely, a causal relation is more likely.
1.4.5 The Adverse Outcome Pathway (AOP) framework as a conceptual tool to support the biological plausibility of epidemiology studies

Regulatory studies, traditionally based on animal experimentation, are intended to explore for any potential hazard but they are not specifically designed to inform on specific and complex human health outcomes. New data type and methods can be more effective in hazard identification, but there is a need to define which data could be used and/or be more valuable for compound specific risk assessment and which could be informative on data gaps in the standard regulatory assessment or add an insight for their interpretation.

The inclusion of epidemiology findings into risk assessment is an attempt to integrate human data with toxicological data and approaches elucidating mechanisms or pathways of toxicity, rather than rely only on the standard regulatory requirements. Furthermore, human data are compelling and trigger important considerations on the risk perception that are frequently reported in the media. Many epidemiological studies include pesticides and their integration (why and how) or exclusion in the risk assessment should be legitimate. In this top-down context, epidemiology findings can be used for validation purposes; however, in the context of risk assessment, they can trigger alternative approaches to investigate the biological plausibility, overcoming their own limitations or help when human data are not corroborated by the regulatory toxicological studies (Li et al., 2012). Thus, the complex scientific process for the identification of human risk has to involve both epidemiological and experimental data. Furthermore, when epidemiological data are lacking, experimental data are relevant to inform on the biological plausibility as part of the overall weight of evidence.

The AOP is an organizational framework, it combines information from multiple fields of inquiry and provides knowledge of biological pathways, highlight species differences or similarities, identifies research needs and support regulatory decisions (Villeneuve D. et al. 2014 a and b). In this context, the AOP approach could help in organizing the available experimental knowledge to assess biological plausibility and to implement the overall risk assessment. The PPR Panel is therefore recognizing the value of using all the available information on a pesticide active substance when conducting the risk assessment and is considering the AOP framework as a systematic and transparent tool for organizing, reviewing and interpreting complex information from different sources. The AOP, being a conceptual framework to mechanistically understand apical hazards, the human health outcome should be included as part of the hazard assessment and the AOP will serve as tool for hazard identification.

In this perspective, the AOP framework is considered conceptually similar to the International Programme of Chemical Safety (IPCS) Mode of Action/Human Relevance framework (Meek et al., 2003; Seed et al. 2005) with major differences being the dominant applications to which it is applied and the inclusion of the formal incorporation of Bradford Hill’s considerations. AOPs are not intended to be chemical specific in the sense that they are not developed to describe what a single chemical does, but rather to describe how any chemical triggering the molecular initiating event (MIE) in a sufficient dose and temporal relationship might perturb adversely a physiological pathway using chemical tools when necessary. Consequently, describing an AOP does not require chemical-specific information but the application of the pathway in a predictive context relevant for risk assessment. Nevertheless, it requires understanding of the chemical tool-specific properties like potency or ADME properties as these data will be informative for dictating the magnitude and duration of the perturbation at the MIE.

In 2012 the OECD launched the AOP development programme followed by the publication in 2013 of the OECD Guidance Document on Developing and Assessing AOPs, addressing conventions and terminology, information content of an AOP description, weight of evidence evaluation and standardization and rigor for developing AOPs. Conventionally, an AOP consists of a single sequence of key events connecting the MIE to an AO; the idea is to have a tool that pragmatically simplifies complex biological events (OECD 2013 and 2014).

The MIE is defined as a specialized type of KE that represents the initial point of chemical interaction on the molecular level, within an organism, that results in a perturbation that starts the AOP. The AO is defined as a specialized type of KE that is generally accepted as being of regulatory significance on the basis of correspondence to an established protection goal or equivalence to an apical endpoint in an accepted regulatory guideline toxicity test.
The MIE and the AO are linked by a series of KEs defining a direct relationship among them (KER, Key Event Relationship) where the KE should provide some ability to predict or infer the state of the downstream KEs and their relationships have to be supported by biological plausibility and scientific evidence, with a quantitative understanding in a codified assembly of weight of evidence. The availability and robustness of experimental data will classify the AOP developed into a given category, but the AOP will be considered as a living document that can change of category on the basis of new available data. In moving down from a putative AOP to a quantitative AOP it is expected to see an increase in: strength of evidence, understanding, transparency, defensibility, quantitative precision, cost, data needs and time.

Table 4: AOP categories

<table>
<thead>
<tr>
<th>Stages of AOP Development</th>
<th>Characteristics</th>
</tr>
</thead>
<tbody>
<tr>
<td>Putative AOPs:</td>
<td>Hypothesized set of KEs and KERs primarily supported by biological plausibility and/or statistical inference</td>
</tr>
<tr>
<td>Qualitative AOPs:</td>
<td>Include assembly and evaluation of the supporting weight of evidence – developed in AOP knowledgebase in accordance with internationally-harmonized OECD guidance</td>
</tr>
<tr>
<td>Quantitative AOPs:</td>
<td>Supported by quantitative relationships and/or computational models that allow quantitative translation of key event measurements into predicted probability or severity of adverse outcome</td>
</tr>
</tbody>
</table>

In this context it is clear that an objective and complete AOP doesn’t exist as methods and/or new experiments can change the existing one. It is also clear that any stage of AOP development has a potential utility as the level of development desired/required depends on its potential application.

The PPR Panel was then intended to apply the AOP approach to investigate and possibly provide, in an objective and transparent way, the biological plausibility supporting the potential link between exposure to pesticides and Parkinson’s disease/parkinsonian disorders and childhood/infant leukaemia. These human health outcomes were selected because they are consistently observed in different meta-analyses and represent relevant disease models for the application of the approach. While the link between environmental factors and Parkinson’s disease/parkinsonian disorders is relatively data rich, data supporting the link to childhood/infant leukaemia is more scarce. This would allow to evaluate the flexibility of the approach and to make a comparative evaluation on data similarity and/or data gaps between the standard regulatory requirements and alternative studies designed to investigate toxicological endpoints specific for the diseases.

2. Introduction to Parkinson’s disease, parkinsonian disorders and application of the AOP conceptual framework

Parkinson’s disease (PD) is a chronic progressive neurodegenerative disorder with a higher prevalence in the aged male population (Cereda et al. 2016). It is a chronic disease as the mean duration is 15 years from the recognition of the disease until death (Schulman et al. 2011) and is progressive as the clinical signs and their severity are linked to the spread and progression of the pathology. Although the clinical symptoms include slowness of movement, resting tremor, rigidity and disturbances in balance, it is now recognized that additional non-motor symptoms can occur as a result of the progression of the disease. Some or all of these clinical signs can however be observed in different disorders and the resulting syndrome is defined as “parkinsonism”. When parkinsonism is the prominent part of the disorder, these are referred as “parkinsonian disorders” and include PD (Dickinson, 2012). The primary pathology is however common to all parkinsonian disorders and is represented by a selective degeneration of dopaminergic neurons in the substantia nigra pars compacta (SNpc), which project mainly to the striatum, in association with the development of cytoplasmatic, protein-rich inclusions, called Lewy body (LB). One of the main components of LB is the aberrant oligomeric α-synuclein (a pre-synaptic neuronal protein) and a parallelism exist between the presence of motor and non-motor symptoms and the finding of α-synuclein pathology beyond the
SNpc. This is the basis of the Braak paradigm (Braak et al. 2003 and 2008), which proposes a staging system to describe the spread and progression of the pathology resulting from multiple detailed post-mortem analysis in PD patients. The sequential occurrence of alterations and the involvement of different structures of the nervous system, including the peripheral one, is a key aspect of the disease that is relevant to understand the contribution of environmental factors and their role in the initiation of the disease (Pan-Montejo et al. 2012).

Complex molecular landscape of PD

Indeed, although the molecular aetiology of the disease is unknown, it is most likely caused by a complex interplay of genetic and environmental factors with multiple interacting pathways including synaptic and mitochondrial dysfunction, impaired protein degradation, α-synuclein pathobiology and neuroinflammation (Fujita et al. 2014). Some cases may have a clear genetic cause while others can be caused by effects of toxins (e.g. MPTP) and/or a gene-environment interaction; however, although these degenerative disorders can be inherited or idiopathic they all have as a common denominator the loss of dopaminergic neurons projecting from the substantia nigra to the putamen (Dickinson, 2012). In this context, the role of pesticides as potential environmental risk factors for PD has long been suspected and recurrent through multiple epidemiological meta-analyses, though the specific causative agents and the mechanisms underlying the disease are not fully understood (Baltazar et al. 2014, Franco et al. 2010, Shulman et al. 2011, Pryadarsnii et al. 2000, Ntanzi et al. 2013). For this reason PD is of high interest for the pesticides risk assessment and several experimental models have been proposed (Baltazar et al. 2014, Cicchetti et al. 2009, Drechsel et al 2008, Moretto et al. 2011).

However, the linkage of a complex and unique human disease with experimental toxicological studies is still representing an important challenge for risk assessment. This is because, regulatory toxicology studies, as good as they are for exploring any potential hazards, are not designed to understand relevant mechanisms of toxicity and, particularly, they can be of limited sensitivity when hazards are likely consequent to long-term, low-dose exposure to toxicants, or when multiple toxicants are interacting on the same AO through different MIE or when the genetic background is influencing the adverse outcome. In this context, the AOP could represent a scientifically valid, transparent and pragmatic tool for hazard identification and could be used to support the biological plausibility of the observed event by means of introducing the human health outcome in the pathway. Due to the complexity of the disease, multiple MIEs and AOPs can be developed for PD. For this reason, the PPR Panel considered as initial step in the construction of AOPs of interest for PD the general scientific consensus that mitochondrial and protein dysfunctions, aggregation of toxic oligomers of α-synuclein, oxidative stress and neuroinflammation are involved in the degeneration of dopaminergic neurons in the SNpc, and that loss of these neurons is leading to symptoms of PD. Based on the existing knowledge supporting such a consensus, the PPR Panel built up a number of initial schemes from which two AOPs were selected for further development. Tool chemicals were selected based on data availability and their use as a prototype chemicals in experimental models of PD.

Tool chemicals for the AOP building

In this context, 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP), rotenone and paraquat, are likely to be the most widely used chemical substances to induce loss of dopaminergic neurons. In particular, MPTP is of high interest as it was able to produce Parkinson-like motor disorders in human after few days following exposure to an illicit substance of abuse containing this compound as a contaminant. MPTP is able to cross the blood-brain barrier and is selectively taken up by dopaminergic neurons after metabolic activation by MAO-B of astrocytes to MPP+. Rotenone is a highly lipophilic insecticide/piscicide which, unlike MPP+, lacks specificity for dopaminergic neurons but this chemical is able to reproduce features of PD when chronically administered to rodents at low doses as has been reported in detail in a seminal paper (Betarbet et al. 2000). The susceptibility of dopaminergic neurons is likely due to their sensitivity to the toxicity induced by rotenone rather than toxicokinetic (i.e, metabolic) characteristics. For both substances, the neurotoxic effect is considered consequent to inhibition of complex I in the mitochondrial respiratory transport chain leading to mitochondrial dysfunction. However, both MPP+ and rotenone can produce neuronal loss by a large number of processes and this was considered an important limitation in the construction of the AOPs (Aguilar et al. 2015). It is also worth to note that these substances were mainly used as tools to reproduce in-vivo and/or in-vitro models of PD or to study mechanisms relevant for PD rather than for hazard identification.
Paraquat is an herbicide belonging to the chemical class of bipyridyl quaternary ammonium. Although the general toxicity of paraquat and its target organs is well characterized, its neurotoxic effect has been mainly explored in the last decade after several reports of brain damage in individuals exposed to lethal doses of paraquat (Baltazar et al. 2014). The mechanism underlying paraquat neurotoxicity is not fully elucidated, although several pathways have been proposed: the toxicity is essentially linked to its redox potential. Paraquat has a complex toxicokinetic and this also includes interaction with microglia (Baltazar et al. 2014). As toxicokinetic and metabolism considerations are not relevant for the construction of the AOPs, in this context, paraquat will be used as a tool chemical to define an AOP dealing with oxidative stress, mitochondrial dysfunction and neuroinflammation (Baltazar et al. 2014).

3. Plausibility of the involvement of pesticide exposure as a risk factor for Parkinson’s disease; and contribution of the AOP concept to support plausibility

3.1. Biological plausibility in support of pesticide-associated Parkinson’s disease

In addition to the above mentioned epidemiological studies, laboratory experiments have provided evidence for neurotoxic effects and biologically plausible mechanisms linking pesticides to PD. Biologically plausible mechanisms for PD causation have been postulated for specific pesticides, including inhibition of mitochondrial complex I by rotenone, induction of oxidative stress by paraquat, and inhibition of aldehyde dehydrogenase by the dithiocarbamate fungicides maneb, ferbam or mancozeb (reviewed in Breckenridge et al., 2016).

Studies with rotenone are consistent with the assumed role of respiratory chain complex I and mitochondrial dysfunction in PD pathogenesis; data on paraquat are in line with the assumed role of oxidative stress in the disease; the toxicity of maneb in experimental animals also involved mitochondrial dysfunction. These experimental toxicants selected from the group of pesticides have triggered many of the features known from PD in animal models. They have in particular been shown to trigger dopaminergic neuronal cell death in the S. nigra, similar to the pathology observed in PD (Drechsel and Patel, 2008; Hatcher et al., 2008).

Despite the large body of epidemiological and experimental evidence linking pesticide exposure to PD, the exact etiological factors remain elusive, and pathogenic mechanism(s) triggering neuronal loss and PD progression are not completely known. Advances concerning the plausibility of the association have been made in the following areas:

a) Repeated and multiple chemical exposures.

Pesticides currently used do not strongly bioaccumulate in the human body, whereas in the past this was not the case. Prolonged effects may therefore arise from long exposure periods or previous exposure to more bioaccumulating compounds. Alternatively, single exposure may cause minute, clinically undetectable neurotoxic effects that, if accumulated over the course of decades, might lead to triggering of disease or to the enhancement of ongoing endogenous disease progression. In this context it is important that PD symptoms become clinically apparent only after considerable dopaminergic cell death has been ongoing. Most likely, it takes years of only few individual neurons dying per day or month, until the threshold for clinical symptoms is reached.

The majority of work identifying potential dopaminergic toxicants associated with PD comes from studies examining mechanisms and risks arising from a single chemical. However, human environmental exposures are much more dynamic and they likely involve numerous risk modifiers including multiple chemicals or chemical mixtures. Pesticides consist of a wide range of chemical structures with diverse mechanisms of toxicity and not necessarily all of them contribute to the development of PD. The effect of pesticide mixtures has to be considered for risk assessment. The multi-hit hypothesis supporting neurodegeneration and PD, suggests that the brain may be capable of withstandng the effects of an individual chemical targeting dopaminergic neurons. However, when multiple chemicals target numerous sites within the dopaminergic system, defense mechanisms may be compromised resulting in cumulative damage and neuronal death (Hatcher et al., 2008).

Furthermore, exposure to different pesticides may initiate a number of neurotoxic mechanisms that
may converge later in a chain of linked events eventually leading to nigrostriatal dopaminergic cell death and impaired motor function. This might explain why pesticides dissimilar in their chemical structure and unlikely affecting the same cellular structure, trigger similar downstream events (e.g., mitochondrial dysfunction and oxidative stress).

Paraquat is a herbicide that has long been considered a potential risk factor for PD because of its structural similarity to MPP⁺, the active metabolite of MPTP. While much of the focus has been put on paraquat, other classes of pesticides are also known to impair dopaminergic neurons. Exposure to maneb, a dithiocarbamate fungicide, has been linked to neurological impairments in agricultural workers, and there are epidemiological data showing that neurodegeneration occurs more frequently in environments where workers are co-exposed to paraquat and maneb (Thrash et al., 2007).

Paraquat and maneb administered individually to mice caused no neurological damage, but when administered as a mixture, produced traits characteristic of PD (Thiruchelvam et al., 2000). A further study on mice demonstrated enhanced sensitivity of the ageing nigrostriatal dopaminergic pathway to the combination of paraquat and maneb, resulting in irreversible and progressive neurotoxicity (Thiruchelvam et al., 2003). These results were partially supported by a case control study on 362 incident PD cases recruited between 2001 and 2007, where ambient exposures to the pesticides paraquat, maneb and ziram were estimated. The combined exposure to these pesticides at workplaces increased three-fold the risk of PD, whereas the combined exposure to only ziram and paraquat, excluding maneb exposure, was still associated with a 80% increase in risk (Wang et al., 2011).

A further type of chemical interaction may occur, if one chemical, given at an early time in life, sensitizes to another chemical, given at a later time point. This was observed in an experiment where (perinatal exposure during gestation and lactation exposure (i.e. developmental) to low dieldrin levels altered dopaminergic neurochemistry in offspring and exacerbated MPTP toxicity later in life (Richardson et al., 2006).

Potentially interaction of chemicals may also occur through the links of epigenetic changes (Balmer et al., 2014) or of neuroinflammation, even in cases in which exposure periods are far apart. For instance, the short exposure to MPTP in humans resulted in an injury that initiated self-perpetuating pathological processes, and neuroinflammation persisted for many years (Langston et al., 1999), making the respective brain regions vulnerable to potential second-hits.

b) Genetic factors and gene x environment interactions.

There is growing evidence suggesting that genetics may affect susceptibility to PD among the subgroup of people exposed to pesticides. Exposure to pesticides (or to specific pesticides) over the course of decades could initiate or accelerate the underlying neurodegenerative process; however, without concurrent genetic or metabolic risk factors pesticides may not necessarily lead to the disease. While a minority of PD cases may be primarily due to a specific genetic or environmental risk factor, most cases are likely due to gene-environment interactions (Fujita et al. 2014). This kind of interactions may explain why despite the large number of people regularly exposed to pesticides not everyone develops the disease; it may only affect those carrying a genetic vulnerability. Highly penetrant mutations in some genes (SNCA, Parkin, DJ-1, PINK 1, LRRK2 and VPS35) produce rare, monogenic forms of PD, while unique variants within LRRK2 and GBA show incomplete penetrance and are strong risk factors for the disease (Hernández et al., 2016b). On the other hand, polymorphisms of genes encoding enzymes involved in the metabolism of pesticides or in cell damage mechanisms, in particular PON1, PON2, NQO1, NAT2, NOS and ALDH-2 may point towards an inherent population-specific genetic predisposition (Fong et al., 2005; Furlong et al., 2016; Manthripragada et al., 2010; Punia et al., 2011; Wan et al., 2011, Paull et al., 2016a). However, most studies addressing gene-environment interactions are limited by the small sample size and recall bias inherent to case-control studies.

c) Oxidative stress.

The role of oxidative stress in the etiopathology of PD is well established (Surmeier et al., 2011; Zhou et al., 2008, Schuldicheft et al, 2013). The metabolism of DA can lead to the generation of hydrogen peroxide (H₂O₂) and other reactive oxygen species (ROS). This exposes dopaminergic neurons of the SNpc to a higher level of oxidative stress than other brain regions. There is evidence that some pesticides would enhance these oxidative stress events. Structurally diverse pesticides can do this,
based on several different mechanisms that eventually converge on a shift of the redox balance of the dopaminergic cell. For instance, paraquat toxicity is related to its ability to redox cycle, accepting an electron from an appropriate donor with subsequent reduction of oxygen to produce superoxide while also regenerating the parent compound. Moreover, paraquat may enhance oxidative stress by activating the NADPH oxidase of microglia cells (Drechsel and Patel, 2008). A third way to increase oxidative stress would be to activate glial cells (neuroinflammation), which may directly mediated neurotoxicity or exacerbate the toxic outcomes already initiated within neurons following exposure to toxic chemicals (Ramsey and Tansey, 2014). An increased risk for PD has also been associated with diquat, a bipyreryl herbicide structurally related to paraquat. Exposure to diquat was reported to cause Parkinsonism in a farmer acutely exposed to a concentrated solution of the herbicide (Sechi et al., 1992). Moreover, part of the toxic mechanism of dithiocarbamate fungicides (e.g., maneb) has been associated with the dopamine oxidation and chelation of metals, leading to alterations in cellular redox status. Permanent parkinsonism has been reported following chronic occupational exposure to maneb (Merco et al., 1994), which supports the potential role of this fungicide in the etiology of PD. Although dopaminergic areas of the brain (striatum, substantia nigra and nucleus accumbens) have the highest levels of the antioxidant enzyme paraoxonase (PON2), levels in males are 2- to 3-fold lower than in females. These lower PON2 levels may provide a weaker defenses against oxidative stress in male dopaminergic neurons and may support the higher incidence of PD in males (Furlong et al., 2016).

d) Aldehyde dehydrogenase (ALDH) inhibition

ALDH enzymes are responsible for detoxification of exogenous and endogenous aldehydes by oxidizing aldehydes to carboxylic acids. Aldehyde metabolites have been suggested to be involved in the pathogenesis of PD; for instance, 4-hydroxy-nonenal (4-HNE), a common aldehyde product of lipid peroxidation, promotes the formation of α-synuclein oligomers (Zhang et al., 2015). ALDH also continuously detoxifies 3,4-dihydroxyphenylacetaldehyde (DOPAL). This degradation product of dopamine is generated in neurons by monoamine oxidase (MAO), and has been involved in the loss of dopaminergic neurons in PD as a result of generating hydroxyl radicals. ALDH activity can be inhibited by pesticides such as the metal-complexed dithiocarbamates (e.g., maneb, ziram), imidazoles (benomyl, triflumizole), phthalimides (captan, folpet) and organochlorines (dieldrin) (Fitzmaurice et al., 2013; Fitzmaurice et al., 2014).

e) Mitochondrial dysfunction

Inhibition of complex I of the mitochondrial electron transport chain is a biologically plausible mechanism for the development of PD that has gained growing relevance. Damaged mitochondrial DNA, as a footprint of mitochondrial oxidative stress is e.g. found in PD brains (Sanders et al., 2014). Both intoxication with MPTP and that with rotenone directly result in inhibition of complex I and in mitochondrial dysfunction (reviewed in Breckenridge et al., 2016). Dopaminergic neurons of the SNpc have been shown to be uniquely sensitive because of their higher production of mitochondrial H2O2 in response to complex I inhibition as compared to cortical neurons (Sanders et al., 2014). Inhibition of complex I activity can lead to the generation of ROS, which then target and inhibit the respiratory chain leading to subsequent ROS production and further mitochondrial damage. The consequent failure in energy production may disrupt the vesicular storage of dopamine, leading to increased free cytosolic concentrations of this auto-oxidizable neurotransmitter (Drechsel and Patel, 2008). Two other pesticides, maneb and dieldrin, have been suggested to also inhibit the respiratory chain. For instance, exposure to maneb has been found to result in inhibition of mitochondrial complex III. This contributes to ROS production and mitochondrial dysfunction (Drechsel and Patel, 2008; Zhang et al., 2003). Organochloride pesticides related to dieldrin have been suggested to impair sequestration of dopamine into neurotransmitter vesicles, and the resultant increase in cytosolic dopamine may increase the risk of oxidative stress (Miller et al., 1999; Vergo et al., 2007).

f) Congruence of clinical features

Parkinsonism is a complex syndrome with a heterogeneous set of clinical features. For instance, parkinsonism observed in humans due to high-dose exposure to manganese or carbon monoxide, has clinical features that differ from those that I are normally –related to idiopathic PD. For instance, there is a poor response to dopaminergic therapy. This situation is different for cases of PD associated to pesticides (Tanner et al., 2011). Similar clinical features were found in PD cases that did or did not have exposure to rotenone, paraquat, or groups of pesticides with similar mechanisms. This
observation suggest that PD associated with these agents is clinically typical, and this provides further plausibility for a role of pesticide exposure in the etiology of typical PD.

3.2. To what extent do experimental toxicity studies on mechanisms of toxicity cover mechanisms relevant for PD, and what is the contribution of the AOP in supporting biological plausibility

3.2.1. Rationale of the working approach

At present, different and separate sets of information exist concerning the following five domains: pesticide exposure, toxicant MoA, experimental studies, disease pathogenesis and the occurrence of PD. The combination of information from these domains may shed light on the questions (i) whether the statistical correlation of pesticide exposure and occurrence of PD is mechanistically plausible, (ii) whether there are causal links, and (iii) if such links can be confirmed or refuted by experimental testing.

As a starting point, data are available from regulatory toxicity studies that link pesticides to traditional endpoints (e.g. histopathology). In addition, for some toxicants a mechanism of action is known. The first question relevant to the working group’s mission was to investigate, whether a mechanism of pathogenesis could be assigned to PD in form of an AOP. The second open issue addressed was to investigate whether experimental studies would yield information concerning the mechanism of action of toxicants. The third step was then to investigate whether mechanisms of action of toxicants overlapped with mechanisms of disease pathogenesis (AOPs) relevant for PD. Finally, the answer to these questions was used to establish plausible links between the exposure to pesticides and the risk of developing PD.

3.2.2. Capturing of a complex disease (PD) by AOP

The AOP concept has been developed by toxicologists to describe the hazard of toxicants. The concept has not been envisaged to cover complex human disease. This has several reasons: defined MIE may not exist for diseases; diseases may follow a multi-hit principle instead of linear chains of events; diseases have multiple symptoms instead of one final unhealthy outcome; pathogenesis of chronic degenerative diseases is likely to be based on cyclic events; KEs of chronic disease are difficult to capture or to be modified experimentally; data on diseases and disease pathogenesis are different in type and in the way they can be obtained than data of poisonings with toxicants; experimental data on disease are either difficult to obtain and to reproduce or they cannot be obtained at all.

Considering the above arguments, the development of “AOP for diseases” will only be possible in some favorable situations. “AOP relevant for a certain disease” is a more exact definition than the more superficial but easy to remember term “disease AOP”. The process of AOP development is greatly facilitated, if the disease has a variant that is known to be induced by a defined toxicant; if defined molecular interventions are known to block the pathogenesis of the disease; if complete sets of data are known on defined stages of the disease; and if biomarkers or measures obtainable by non-invasive methods describe the progression of the disease.

The most important restriction is that the AOP should not be defined for the disease as such, but for a sharply defined symptom of the disease (as equivalent to an adverse outcome for toxicants). A second important condition is that this endpoint can be reproduced in animal models, and that chemicals exist that trigger the same endpoint in the animal models; this implies that example (tool) chemicals are available that are likely to trigger the envisaged “disease AOP”. As such conditions were fulfilled here, it was scientifically acceptable to work on model AOPs relevant to PD (here for simplicity sometimes called “PD AOP”).

3.2.3. Selection of the AO

Parkinson’s disease is a human-specific clinical syndrome, usually not observed in animals. The key clinical signs are bradykinesia, rigidity, resting tremor and postural instability. In addition, the disease may be associated with vegetative symptoms (intestinal disturbances, disturbed sleep pattern), cognitive decline and affective symptoms (most frequently depression). Many different AOPs may thus be associated to the disease. This is because first, the disease has several adverse outcomes, and
second, several AOP may converge onto each of these AO, according to the OECD definition of AOP. For proof of concept, ‘parkinsonian motor symptoms’, i.e. what is described in patients mainly as bradykinesia and rigor, were chosen as AO. Parkinsonian motor symptoms were defined here as the typical motor deficit observed in human disease and in experimental conditions, as a result of the loss of dopaminergic neurons of the nigrostriatal pathway. Other AO could have been chosen. For instance, cognitive function (Paull et al., 2016b) and tremor would have been candidate endpoints.

The choice of the Panel was driven by the relative specificity of the endpoint for PD, by the possibility to associate the AO to known and defined pathologic changes, and by the transferability to animal models. Parkinsonian motor symptoms were considered to be relatively specific, to be found universally in all cases of PD, have a well-defined underlying pathology, and to be measurable and modifiable (by drugs) in experimental animals. Notably, parkinsonian motor symptoms are not 100% specific for PD, but this is not a necessary condition for an AO.

3.2.4. Choice of example AOP relevant both for parkinsonian motor symptoms and for pesticides as risk factors

Having chosen the AO, the next question was which types of assumed pathological sequences were to be reflected by a proof-of-concept AOP. The decision was taken to consider only pathological processes occurring during adult life. It has been hypothesized that PD may also have developmental origins (Landrigan 2005), and pesticides may have effects on early brain development, but this potential etiology was deliberately not considered here. For practical reasons, pathological processes were preferred for which there was sound and ample evidence that they were triggered by chemicals in experimental animals, and preferably also in humans. Having decided on these criteria, and on the AO, the literature was screened for chemicals that triggered parkinsonian motor symptoms. For this, on the one hand a systematic literature review commissioned by EFSA (EN-955, 2016) was consulted, on the other hand, expert knowledge on the state of experimental parkinsonism research was used. On this basis, the Panel decided to develop two relevant AOP up to a quality level sufficient for submission to the OECD. These two AOP (described in detail in Appendix A) are mainly based on data for three chemicals (MPTP, rotenone, paraquat) that had particularly abundant documentation and that could be used to define the corresponding AOP.

This decision process has some important implications for the interpretation of this opinion. The most important one is that the AOP developed here may only explain a small fraction of the supposed interaction of pesticides and PD risk. As the initial molecular structures and biochemical pathways disturbed by a toxicant are highly compound-specific, there is no such thing as a ‘pesticide AOP’ or a mode of action that makes the connection of pesticide exposure and PD risk plausible. This also applies to smaller subclasses, such as herbicides, fungicides or insecticides (Breckenridge et al., 2016). The aim of the Panel was to test whether the hazard posed by individual pesticides could be linked to the pathogenesis of PD via AOP. If the outcome of this approach is considered promising, then a multitude of AOPs would need to be developed to allow linking of many different pesticides to various symptoms of PD. Some pesticides may fail to fit any of these AOPs, which could be an interesting finding as such. On the other hand, several of these AOP may share common key events, such as oxidative stress, and this would considerably reduce the development work to the definition of partial AOP and their connection to common KE.

3.2.5. Use of tool chemicals to determine whether their mechanism of action overlaps with AOP for PD

The three most data rich chemicals were selected from the literature to build AOP that would describe their hazard. MPTP was chosen, as there are well-documented human poisoning data, large sets of primate data and very extensive sets of rodent data, documented by several hundred publications per year (Daneshian 2015). Rotenone was chosen because of the numerous data from rodent models, and because its molecular target, the mitochondrial complex I is particularly well-characterized. Notably, MPTP is assumed to have the same target, and also for human disease pathology there is good evidence that this target plays a role (Schildknecht et al., 2013 + 2015). Paraquat was chosen, first as there is good evidence for its toxicity in animal models, and second as this has been an individual compound (as opposed to the group ‘pesticides’) that was associated to PD in epidemiological studies. In line with the example chemicals chosen, two MIE were defined: binding to
mitochondrial complex I and initiation of a redox cycling process. These were linked to the AO via two AOP.

This process was fundamentally different from biomedical and systems biology initiatives to define disease pathogenesis. For instance, a universal PD map has been developed (Fujita et al 2014) that incorporates the biomedical knowledge on disease processes relevant to PD. This map takes into account multiple genetic susceptibility factors and modulating events, and its organisation is non-linear. Nevertheless, the two AOP chosen by the panel can be identified also on this complex map as relevant pathways (amongst others) and thus are consistent with current medical knowledge on the disease process of PD. The prototype AOPs are fully reported in the Appendix.

3.2.6. Evaluation of the AOP concerning consistency and strength of evidence

A large part of the effort to develop AOPs was used for their evaluation, and the documentation of this process. The strength of association was judged by a weight of evidence approach based on modified Bradford-Hill criteria. This is fully described in the Appendix A.

Based on the overall weight of evidence, the Panel concluded that the link between the MIEs and the AOs as proposed in the developed AOP is strong and that the proposed KEs (including the MIEs and the AO) can be used as a tool for exploring the hazard of a chemical to trigger parkinsonian motor deficits.

One key conclusion from this is that, if a chemical triggers the MIE or an intermediate KE of such an AOP to a sufficiently large extent, it is likely that it will also trigger the downstream KE, including the AO. This would be a large conceptual advance in predicting chemical hazard in terms of increasing the risk for chronic human disease. Another important feature also resulted from the evaluation: it is highly important to obtain as quantitative data as possible on the KE relationships in order to practically apply hazard predictions based on AOP.

3.2.7. Support of hazard plausibility by AOP

Based on above considerations, the Panel is supporting the use of the AOP framework to explore the biological plausibility of the epidemiological association between pesticide exposure and Parkinson's disease. The recommendation is that pesticides affecting the AOPs developed here should be considered as potentially hazardous (with respect to the development of PD). The same would apply to other AOPs linked to PD, and that would need to be developed in the future.

To avoid misunderstandings, it needs to be stressed that the Panel pursued the development of AOP and the recommendations of their use specifically for the identification of hazard, and not for the assessment of risk. This is fully in line with the standard backbone of risk assessment, i.e. to evaluate whether there is any hazard at all, and if so, to proceed with more complex evaluation of the risk.

This exclusive focus on hazard is logical and necessary, as the AOP framework does not consider (external or internal) exposure data or any toxicokinetic and metabolic processes. To fully rationalize this, it needs to be recalled that an AOP is a 'pathway', i.e. a series of biochemical reactions and pathological events. From this, it becomes evident that the pathway as such cannot have pharmacokinetic parameters. These latter ones are associated with individual compounds that trigger the pathway, and they are evidently unique for each chemical, i.e. cannot be associated to the AOP as such. For practical risk assessment, this means that potential triggering of an AOP by a chemical corresponds to the step of hazard evaluation. The next step within the mode of action framework of risk assessment would then be the consideration of exposure and specific ADME properties of a given compound to come to an overall conclusion on the likelihood of a pesticide to trigger PD.

3.2.8. Conclusions from AOP on suitability of current testing methods

The Panel is interpreting the AOP as a practical, transparent and pragmatic tool to integrate knowledge on mechanisms of toxicity with the measurement of apical endpoints of toxicity. In the case of 'AOP relevant for human disease', as developed here, the integration of different levels of information goes one step further. The AOP integrates mechanistic knowledge on disease pathogenesis, apical endpoints, as measured in experimental toxicity studies and clinical symptoms of the disease. This situation allows solutions to the question, in how far the apical endpoints measured
in animal studies adequately reflect endpoints of disease. Already during the process of development
of the small number of AOPs of this Panel assignment, it became obvious that there are limitations of
the standard regulatory studies when dealing with hazards linked to human complex multi-hit diseases
like Parkinson’s disease and parkinsonian disorders in general. The AO (parkinsonian motor deficits)
and the KE linked to degeneration of DA neurons of the nigrostriatal pathway (which are common to
both AOPs developed by the Panel) are typical apical endpoints that would in theory be identifiable in
the regulatory toxicity studies. However, a review of the standard technology and approach used for
such studies, showed that changes in these endpoints would most likely be missed, even if large
adverse effects were present (e.g. loss of 30% of all nigral dopaminergic neurons). The identification
of neuropathology would require specific sectioning of the respective area (which is not done in
standard OECD 90 or day guideline studies), and it would require immunohistochemical approaches
instead of standard H/E staining. The motor deficit would also not be identifiable if neuronal loss in the
nigrostriatal pathway was below the threshold activating motor deficits (i.e. below 50-70% loss).

The lessons learned from the AOP suggest that even if histological sectioning of the S. nigra and
staining for dopaminergic markers were included in a guideline study, severe adverse effects of test
chemicals may still be missed. Both AOPs indicate that the perturbation of the key events shows not
only a dose concordance, but also that triggering of some downstream KE requires disturbance of the
upstream KE for a prolonged period of time. This has major implications for the study design. For
instance, dosing should be tailored in a way to continuously trigger the MIE for a long time. This may
not be the case, if toxicants are dosed only once or twice a week, and only 3-4 times altogether. With
an inappropriate dosing schedule, changes in the downstream KE or AO (i.e. the apical endpoints of
regulatory studies) may be very low, or even absent. In view of these considerations, it is suggested
to use AOP, and the mechanistic information derived form there, to optimise the design of hazard
identification studies according to the expected mechanisms of toxicity. Moreover, AOP can be used to
indicate data gaps in cases of inconsistent experimental studies, and to provide guidance for improved
study design to address data gaps, inconsistencies and uncertainties. This also comprises suggestions
on additional endpoints to be assessed, either as direct indicators of hazard, or as mechanistic support
to improve data interpretation and species extrapolation.

3.3. AOP as informative source for appropriate identification of data
gaps and testing strategy

3.3.1. AOP as a scaffold to help identifying data gaps

Due to the nature of the AOP that is building KER and thus showing causality of events with a WoE
approach, the AOP concept is very well suited for identifying data gaps. Based on the epidemiological
data linking pesticide exposure to PD and the definition of the AO being ‘parkinsonian motor deficits’
several modes of action were identified linking an initiating event to the KE essential for the AO. This
essential KE is the death of dopaminergic neurons of the nigrostriatal pathway with drop of DA, which
is essential for motor control. Thus, for the AOPs developed by the Panel, the causality of substance
binding to and subsequent inhibition of complex I or mitochondrial ROS formation by redox cycling
both leading to mitochondrial dysfunction, impaired proteostasis, death of DA neurons of the
nigrostriatal pathway and parkinsonian motor deficits is biologically plausible and essential.

Assessment of data gaps within an AOP is feasible by analysing the weight of evidence (WoE) for each
KER within an AOP. In the case of KER ‘Binding of inhibitor to NADH-ubiquinone oxidoreductase
(complex I) leads to its inhibition, the WoE is strong. Despite this high level certainty, there are
several open questions within this KER: (1) low doses of complex I inhibitors with only partial
inhibitory function do not compromise cellular ATP levels suggesting an alternative mechanism
contributing to long-term, low-dose nigrostriatal toxicity; (2) few data on complex I inhibitor
concentration-response using human brain cells/mitochondria thus lacking sufficient quantitative
human data. Also the KER ‘A Redox Cycling compound leads to mitochondrial ROS formation and
dysfunction’ has a high WoE. This is especially true for substances with an electron reduction potential
more negative than O$_2$, which effectively produce superoxide. Generation of superoxide and
subsequent mitochondrial dysfunction has been well described in different taxae. The second level of
KER ‘Inhibition of Complex I leads to mitochondrial dysfunction’ also has a strong WoE as complex I
inhibition causes loss in mitochondrial membrane potential with decrease in ATP production, elevated
levels of ROS, followed by reduced activities of enzymes of the mitochondrial respiratory chain causing
ultimate mitochondrial dysfunction: a process, which is also very well studied. Although there is the notion that other mechanisms than complex I inhibition might be responsible for dopaminergic cell death by complex I inhibiting substances, the over all data supporting this KER is outweighing. The KER 'Mitochondrial dysfunction results in an impaired proteostasis' has a strong WoE because there is a high biological plausibility that proteasome activity is dependent on mitochondrial function and that increased ROS formation interferes with proteasomal function. However, there is data gap on the sequence of events triggering proteasomal dysfunction. This is the case as there is a vicious cycle concerning α-synuclein aggregation and proteasomal dysfunction and it is not clear which one is occurring in a first instance. Some studies suggest that induced oxidative stress leads to α-synuclein aggregation that triggers proteasomal dysfunction. Other studies report that initial proteasomal dysfunction induced by ROS causes α-synuclein aggregation. Moreover, the role of alterations in the cytoskeleton contributing to proteasomal dysfunction is not clear. E.g. tubulin co-localizes with α-synuclein in Lewy bodies and tubulin function is ATP-dependent. WoE for the KER 'Impaired proteostasis leads to degeneration of DA neurons of the nigrostriatal pathway' is strong. Yet the essentiality for impaired proteostasis for nigrostriatal cell death is moderate e.g. acute MPTP exposure leads to specific DA cell death without the formation of Lewy bodies. This might be due to the acute exposure scheme followed in the assay. Effects of long-term and low-dose exposure on proteostasis would be of interest and is representing a data gap in that it is not know how long this KE needs to be perturbated to trigger DA neuronal death. In addition, different features of imbalanced proteostasis can trigger one another (e.g. disturbed protein degradation, pathological protein aggregation, microtubule dysfunction); and each of them can lead to cell death. Therefore, the "single" event triggering axonal degeneration or neuronal death is not known. For instance, for α-synuclein aggregation, it is not clear whether this causes death because some vital function of neurons is lost, or whether some protein increases e.g. because of inhibited chaperone-mediated autophagy. The involvement of the KER 'neuroinflammation leading to nigrostriatal cell death and vice-versa' by interaction of a chemical with microglia/astrocyte cells as a MIE is discussed controversially. Some compounds like paraquat might directly activate microglia/astrocyte cells by ROS production through redox cycling by interaction with inflammatory cells NADPH oxidase. Moreover, neuroinflammation is debated as a modulatory KE possibly enhancing nigrostriatal toxicity of chemicals. In the two AOP related to Parkinson's disease, neuroinflammation was placed as a late event, paralleling degeneration of dopaminergic cells of the nigrostriatal pathway. More exactly, the placement of neuroinflammation in the AOP assumes that degeneration is an important trigger of neuroinflammation, and that neuroinflammation contributes to degeneration. This cyclic nature of events is common to many chronic disease processes. In the case of neuroinflammation even further cycles may be involved that have not been considered here: (i) Possibly some features of neuroinflammation are already triggered by earlier KE, and (ii) neuroinflammation may further enhance early KE of the AOP. This complex relationship of neuroinflammation to other KE makes it difficult to define thresholds for its activation. Furthermore, there is a data gap in the precise understanding on how activation of neuroinflammatory cells might contribute to DA toxicity and how to quantify it. There is strong WoE for the KER 'Degeneration of DA neurons of nigrostriatal pathway leads to parkinsonian motor symptoms'. Impaired motor symptoms are expected to be clinically visible when striatal dopamine levels drop by approximately 80%, corresponding to a DA neuronal loss of approximately 60%. However, in vivo experimental studies gave inconsistent results upon compound treatment. Yet the precise reasons for inconsistencies in results in well-performed in vivo studies are not known, indicating a data gap.

3.3.2. Present data gaps in regulatory studies

In humans, the main neurological symptoms of Parkinson’s disease (PD) are tremor, rigidity, bradykinesia, and postural instability, which can be accompanied by non-motor symptoms such as olfactory deficits/anosmia, sleep impairments, depression, cognitive impairment, constipation, incontinence and autonomic dysfunctions.

Pathologically, PD is characterized by the loss of dopaminergic neurons in the substantia nigra pars compacta (SNpc) and the presence of cytoplasmic protein aggregates, Lewy bodies (LB), in remaining dopaminergic cells and a loss of dopamine (DA) in the striatum. Although PD animal models developed for better understanding of the disease and development of new therapeutics do not exactly reproduce the human disease, they exhibit some of the hallmarks of PD (both motor dysfunction and pathological outcomes). With regard to neurotoxicity requirements for pesticides

regulatory assessment, the question is if the guidelines followed may identify these specific motor
dysfunction and pathological outcomes.

**Motor dysfunction**

Detailed clinical observations including: autonomic activity, body position, activity level gait posture,
reactivity to handling, placing or other environmental stimuli, presence of clonic or tonic movements
have to be performed in all OECD toxicity guidelines.

Motor activity should be measured once in short-term repeated dose toxicity studies (OECD 407, 408
and 422) and several times in specific neurotoxicity studies (OECD 424, OECD 426 and cohort 2 of
OECD 443). The same test (measures horizontal and/or vertical movements in a test chamber) is
implemented in both routine studies and neurotoxicity studies.

However, this is not a requirement in chronic toxicity studies unless neurotoxic effects have been
reported in the shorter studies.

In PD animal models, co-ordination and balance are evaluated by rotation, rota
dor or pole tests, and
gait abnormalities by forepaw stride length test (Le, 2014). Those tests are not required by any
repeated dose toxicity OECD guidelines and they can be optionally incorporated in the design of
neurotoxicity studies OECD 424 and OECD 426.

**Pathology outcomes**

Brains should be weighed and histopathological examination performed on brain, spinal cord and
peripheral nerves in all OECD guidelines.

Perfusion fixation of brains for neuropathology evaluation is only required in both OECD 424 and 426,
while morphometric evaluation should be performed in OECD 426 but is only optional in OECD 424.

In order to detect damage on substantia nigra, appropriate samples of the brain should be obtained
(i.e. rostral midbrain section through the anterior colliculus).

The standard three brain sections performed in repeated dose toxicity studies do not contain the
substantia nigra while in OECD 424 and 426 adequate samples from all major brain regions should be
taken (e.g., olfactory bulbs, cerebral cortex, hippocampus, basal ganglia, thalamus, hypothalamus,
midbrain (tectum, tegmentum, and cerebral peduncles), pons, medulla oblongata, cerebellum) to
ensure a thorough examination.

Furthermore, in order to capture the hallmarks of PD, specific procedures could be necessary as:

- Immunostaining to detect α-synuclein (AS) aggregates.
- Detection of TH, the enzyme responsible for catalyzing the conversion of the amino acid L-
tyrosine to L-3,4-dihydroxyphenylalanine (L-DOPA) by immunocytochemistry followed by
  stereological protocol for cell counting with an optical dissector system to capture the loss of
dopaminergic neurons (Tieu et al. 2003).
- Immunocytochemistry of specific markers: DA transporters (DAT) and vesicular monoamine
  transporter type 2 (VMAT2) to measure striatal dopamine decrease.

All such procedures are not routinely carried out in a standard toxicological data package submitted
for pesticide approval.

In regard to the regulatory requirements, identifying hallmarks of PD may be challenging for active
substance for which no previous data indicating potential neurotoxic effect is available. Indeed, only
motor activity measurements performed in short-term organ toxicity studies could give rise to a
presumption. If there a no signals of neurotoxic effect in those studies then specific neurotoxicity
studies will not be required and motor activity will not been assessed in chronic toxicity study
(although longer exposure may lead to different results). In the same way, histopathological measures
carried out in routine studies may be not specific enough to stress PD outcomes.

In case of suspected neurotoxicity (like pesticidal mode of action or structural similarity to known
neurotoxicants , neurotoxicity study (OECD 424) is required and inclusion of more specialized tests of
sensory, motor function or learning and memory, specific pathological procedures should be
considered in order to examine these possible effects (in this case PD) in greater detail.
Neuroinflammation

The identification of the several different features of neuroinflammation during the AOP construction process showed an important shortcoming of regulatory experimental test procedure: the lack of specific methods to assess neuroinflammation. The standard neurotoxicity testing does not require measurements of any marker of neuroinflammation, except for fuel additives, where testing for a potential increase in glial fibrillary acidic protein (GFAP), as marker of astrocyte reactivity, is mandatory according to US EPA (40 CFR 79 67). This is a deficiency for two reasons: (i) neuroinflammation is not easily identified by standard histopathological methods (e.g. neutrophil infiltration as in many peripheral tissues is rarely observed in the brain); (ii) neuroinflammation is obviously a good indicator of a multitude of different damage processes., i.e. it indicates a toxic action of a compound even if other damage parameters are only slightly affected (and thus remain undetected by standard methods).

The latter point is related to the relatively low specificity of neuroinflammation. Indeed this process is not exclusively observed in Parkinson’s disease, but in most neurodegenerative diseases (Whitton, 2007; Tansey and Goldberg, 2009; Niranjan, 2014; Verthaupt, 2014). Neuroinflammation can also be triggered by several classes of toxicants (Monnet-Tschudi et al., 2007). This relative non-specificity (i.e. the capture of many different AOP with one apical endpoint) makes the testing of neuroinflammation an interesting additional endpoint in regulatory toxicology to provide an alert of ongoing damage that may otherwise have been missed. Nevertheless, neuroinflammation testing is still a challenging issue since it requires multiple endpoints and careful consideration of the test data.

This is because neuroinflammation is a complex event (not a single biochemical reaction), involving different cell types (mainly microglial cell and astrocytes), responding to diverse (sometimes yet unknown) inflammogens or signals from injured neurons (Graeber and Streit, 1990; Monnet-Tschudi et al., 2007; Kraft and Harry, 2011; Claycomb et al., 2013). Activated glial cells release a large panel of mediators, which can (i) have positive or negative consequences on the adjacent neurons; (ii) change composition during the long duration of the neuroinflammatory process; (iii) or lead to a self-sustained vicious circle (Carson et al., 2006; Glass et al., 2010; Aguzzi et al., 2013). Thus, neuroinflammation depends strongly on the pathogenic context. The problem is that the negative/neurodegenerative consequences of neuroinflammation do not only depend on the intensity of the glial reaction (quantity), but rather on the type of the neuroinflammatory process (quality). For instance, activated microglia can be in the M1 (pro-degenerative) or the M2 (protective) state (Maresz et al., 2008; Perego et al., 2011; Ponomarev et al., 2007; Kigerl et al., 2009). Both phenotypes can be observed concomitantly (von Tobel et al., 2014) and the features of neuroinflammation can change over time, e.g. with a neurodegenerative phenotype appearing late, after cessation of exposure, as observed after repeated treatments with the herbicide paraquat (Sandström et al., 2014). Therefore, it is not possible to define a threshold that should be reached to trigger the next key event, but the phenotype, the production and the composition of the inflammatory mediators, such as pro-inflammatory cytokines, reactive oxygen (ROS) or nitrogen species (RNS) (Dong and Benveniste, 2001; Brown and Bal-Price, 2003) should rather be considered in order to predict the consequences of the neuroinflammatory process. In addition, as inhibition of one or two features of neuroinflammation leads only to partial protection of dopaminergic neurons and terminals following rotenone, MPTP, or paraquat exposure (for references, see table of quantitative relationships in KER neuroinflammation to neurodegeneration of nigrostrial pathway), it is a combination of several factors and not a single one, that trigger the neurodegenerative process. Therefore, neuroinflammation cannot be sufficiently characterized by measurement of a single parameter. All these considerations makes it for the time being a challenge to include neuroinflammation into the standard regulatory studies. However, the future mechanistically-driven hazard identification approaches implies also the development of in vitro testing and several test systems for neuroinflammation have been developed, based on cocultures of neurons and glial cells in 2D and 3D, using human or rodent cells as starting point (Monnet-Tschudi et al., 2007; Sandström et al., 2014, Aleppey et al. 2014, Efremova 2015 and 2016).

4. Introduction to Childhood Leukaemia

Pediatric leukaemia is a common childhood cancer (representing 30% of all cancers in children under the age of 15) with an incidence peak between three and five years of age. The disease is phenotypically and genetically heterogeneous, targeting B-cell, T cell or myeloid progenitors and can be additionally stratified according to the differentiation stage at which the hematopoietic stem and
progenitor cells (HSPC) are blocked. The HSPC being the target cell, fetal hematopoiesis and \textit{in utero} exposure are key elements that have to be considered for the assessment of the relationship between pesticide exposure and the disease. Fetal hematopoiesis starts in the aorta gonad-mesonephrons region and colonises the fetal liver and eventually, just before birth, the bone marrow (Wang et al. 2011). The fetal liver hematopoiesis is therefore representing the sensitive target as it is entailing a massive active proliferation of progenitor cells, rendering the HSPC susceptible to oncogenic transformation following DNA damage during pregnancy (Emerenciano et al. 2007). Although the etiology of the acute leukemia is not defined, \textit{in utero} exposure to environmental factors represents a relevant etiological suspect; nevertheless, the paucity of mechanistic data is still representing a major obstacle to understand which toxicological pathways are involved. This is also corroborated by the likely multifactorial origin of the disease with the risk derived from environmental exposure and influenced by genetic susceptibility (Hernandez and Hernandez 2016). In addition, recent mechanistic data has supported previous epidemiological data on the role of late infections in clonal evolution of ALL. Whether environmental (i.e. pesticides) cues are affecting infection development during childhood remains unknown (Greaves 2006).

Of note, almost all the available epidemiological evidences are not making a distinction between infant and childhood leukaemia which are two distinct etiological and pathological entities and this is complicating the interpretation of the epidemiological outcome where the terms paediatric or childhood leukaemia is frequently generalised. Although chromosomal translocation is likely representing the common initiating oncogenic event for both disease, the infant leukemia (IFL) shows a unique biological feature which is the common association with the rearrangements of the MLL gene, a master gene that regulates the normal progression of the human hematopoietic development and differentiation (Hernandez and Menendez 2016). It has to be recognized, however that there ALLs with normal karyotypes i.e. not translocation detected. It’s obvious that the MIE in these cases is more obscure. Although the MLL (and analogous gene) rearrangement is representing (one of) the key event for the initiation of the disease in the HSPC (or an earlier mesenchymal cell), it is likely that it is sufficient only for the development of the overt IFL and additional factors would contribute to the aberrant proliferation of the initiated cells in childhood leukemia. These might depend on alternative (epi)-genetic cooperating lesions at a critical developmental window. In addition, and relevant for this Scientific Opinion, epidemiological and genetic studies suggest that MLL rearrangement may result from \textit{in utero} exposure to DNA topoisomerase-II poisons, including but not limited to the chemotherapeutic agent etoposide (Hernandez and Menendez 2016). A chain of pathogenetic events linking the \textit{in utero} exposure to Topo-II poisons to IFL is fully reported in Appendix 3 and is representing the attempt made by the Panel to build up a qualitative AOP to mechanistically support the biological plausibility that exposure to pesticides could be linked to the development of IFL.

\textit{In utero} exposure to environmental risk factors is also relevant for the development of childhood leukaemia (CHL). However, for the CHL, although the initiation event is still involving a structural or numerical chromosomal alteration, the development of the leukaemia requires the activation of cell proliferation. The longer latency period for the CHL (when compared to the IFL) clearly indicates that the initiating event is not enough for the conversion of a preleukaemic clone into cancer, strongly suggesting that a second, very likely post-natal, hit is necessary. Dysfunction of the immune system and delayed infections have been frequently linked to CHL leukaemia by means of mechanistic considerations like a dysregulated immune response consequent to a low repertoire of infections during the early development of the immune system and an aberrant congenital response to infections (Hernandez and Menendez 2016). In its attempt to build an AOP specific for the CHL, the Panel found no sufficient evidence to identify a mechanistically plausible MIE and no chemicals were identified to empirically support the toxicity pathway. Nevertheless, considering the relevance of the debate linking pesticide exposure and potential development of CHL, the Panel developed a hypothetical AOP which is fully reported in Appendix.
5. Plausibility of the involvement of pesticide exposure as a risk factor for Infant and Childhood Leukaemia; and contribution of the AOP concept to support plausibility.

5.1 Biological plausibility in support of pesticide-associated IFL and CHL

In contrast to the epidemiological studies mentioned above, there is scarce experimental and mechanistic evidence supporting the association between exposure to pesticides (or any other chemical except such as benzene) during different developmental stages and paediatric leukaemia. While for childhood leukaemia there is no tool chemical capable of inducing the disease under experimental conditions, for infant leukaemia there is enough evidence for the anticancer drug etoposide. Despite the distinct natural history and pathogenesis of infant and childhood leukaemia, both entities share a chromosomal translocation as the major initiating oncogenic event.

Most of the studies available in the open literature pertaining to pesticides do not directly link pesticide exposure to development of childhood leukaemia. Nevertheless, they do provide some evidence of the genotoxic or cancer-promoting capacities of some pesticides based on cellular studies, suggesting the potential of these compounds to trigger leukaemogenesis. An in vitro study showed that a human leukaemic (K562) cell line exposed to 1 µg/mL isofenphos for 72 hours exhibited an enhanced proliferation and poor cellular differentiation (Rought et al., 2001). In addition, human peripheral lymphocytes exposed to 0.1–10 µg/mL isofenphos for 1 h exhibited dose-dependent damage to chromosomal DNA (using the comet assay) as well as disruption of the cholinergic nuclear signalling pathway, which collectively could lead to genomic instability and leukaemogenesis (Williams et al., 2004). On the other hand, human K562 cells exposed to 0.1 µM of diazinon resulted in hypermethylation of several genes involved in cell cycle arrest such as cyclin-dependent kinase inhibitors (CDKN1A and CDKN1C) as well as tumour suppressor genes such as p53 and PTEN (Zhang et al., 2012). Furthermore, human mammary carcinoma MCF-7 cells exposed to low concentrations of diazinon http://www.who.int/foodsafety/areas_work/chemical-risks/jmpr/en/) or fenitrothion (0.001-10 nM) for 24 hours exhibited a higher degree of micronucleus formation (Ukpebor et al., 2011).

Lu et al (2015) found that human foetal liver HSPCs exposed to chlorpyrifos for 24 h resulted in MLL rearrangements and double-strand DNA breaks in a dose- and time-dependent manner. This study suggested that chlorpyrifos might act as a TOP2 poison similarly to benzoquinone (a benzene metabolite) and bioflavonoids, thus supporting the relevance of TOP2 poisons in the pathogenesis of infant leukaemia.

Pesticides other than OPs have been in some way associated to leukaemogenesis. For instance, human neuroblastoma SH-SY5Y cells and human T-cell leukaemia Jurkat cells exposed to methylpyrazole insecticides (tebufenpyrad, bixafen, fenpyroximate or tolfenpyrad) for 1 h showed increased induction of γ-H2AX (a marker of double strand DNA breaks) attributed to the generation of oxidative stress as an effect of impairment of the mitochondrial electron transport chain (Grailiot et al., 2012). Furthermore, exposure of the CEM x 174 cell line, a hybrid of human T and B cells, to 50 µM heptachlor, chlordane or toxaphene for 24-36 h showed decreased protein levels of the tumour suppressors p53 and Rb (Rought et al., 1998; Rought et al., 1999). Low concentrations of heptachlor (5-10 µM) suppressed doxorubicin-induced caspase-3 activity and subsequent activation of apoptosis in this cell line (Rought et al., 2000). Human peripheral lymphocytes exposed to 20 µg/mL of a commercial formulation of the fungicide dinocap for 24 h exhibited increased chromosomal aberrations, formation of sister chromatid exchanges and decreased mitotic index (Celik et al., 2005). In vitro studies with the chloroalkylthiocarbamoxide fungicides captan and captanol at a concentration of 1 µM have shown to decrease the activity of topoisomerase II by 50 and 20%, respectively (Rahden-Staroń et al., 2002). Similarly, thiram (a dithiocarbamate fungicide) inhibits topoisomerase II at 10 µM (Rahden-Staroń et al., 1993). However, the in vivo genotoxic potential of these fungicides (i.e., genetic deletions and/or mutations) occurred only at very high doses in Drosophila (10-100 mM) (Rahden-Staroń, 2002).

In assessing the above studies coming from the open literature, findings from regulatory studies should also be taken into account. Tebufenpyrad, bixafen, fenpyroximate, captan, chlorpyrifos and thiram are approved in EU, and none of them are classified for genotoxicity for the time being, thus
the mandatory regulatory studies did not show genotoxic potential. Captan is classified for
carcinogenicity. For the rest of the pesticides not approved in the EU none of them are currently
classified as being genotoxic, while two are classified being carcinogenic; namely chlordane and
captafol. Thus, although a thorough assessment of the genotoxic potential of the mentioned
pesticides have not been undertaken, the panel finds that the few in vitro studies available from the
open literature so far to support the epidemiological evidence for the association between childhood
leukaemia and exposure (in utero and/or after birth) to some classes of pesticides is limited. Also,
there is limited evidence from in vivo studies. However, it remains uncertain whether this association
arises from a causal or non-causal relationship and biological studies to provide evidence for a
potential mechanism have been inconclusive. Almost all in vitro studies used immortalised cell lines or
primary human lymphocytes from adults and 3-week-old mice, which are not appropriate cell models
for studying childhood leukaemia. The only one study using foetal liver HSPCs can be considered as
the best cell model for this purpose.

This clearly indicate how complex is to define and weight biological plausibility when both regulatory
studies and experimental studies from the open literature are contradicting.

The mechanisms underlying the association between pesticides and childhood leukaemia are currently
poorly understood and more studies are needed to better understand this association. There is
agreement in the scientific community that a well defined key event involved in paediatric
leukaemogenesis is the induction of chromosomal rearrangements. The mechanistic linkage between
pesticide exposure and this genetic damage may be accounted for by topoisomerase II (TOP2)
poisons (in particular for infant leukaemia) or generation of oxidative stress leading directly or
indirectly to DNA damage.

a) TOPO2 poisoning (inhibition)

Topoisomerase II has critical functions in both DNA replication and transcription processes. Under
physiological circumstances, the active site tyrosine in TOP2 serves as a nucleophile to initiate the first
transesterification reaction to form a covalent adduct with the backbone phosphate in DNA, thus
generating a transient break. The second transesterification reseals the DNA break and regenerates
the free tyrosine (Chen et al, 2013). In contrast, exposure to TOP2 poisons can lead to the
stabilization of the transient DNA/Top2 cleavage complex resulting in an increased frequency of DNA
double-strand breaks and error-prone non-homologous end-joining (NHEJ) repair. For this reason,
these chemicals are called top2 poisons to distinguish them from catalytic inhibitors of the enzyme.
Cells harboring accumulated breaks in DNA are not able to enter into the mitotic phase of the cell
cycle, thus undergoing cell death.

Some anticancer drugs (i.e., etoposide, doxorubicin); environmental chemicals (i.e., benzene, some
pesticides) and natural substances (i.e., bioflavonoids) are TOP2 poisons with DNA cleavage activity
(Pendleton et al., 2014). Amongst the TOP2-poisons chemicals only etoposide has strong evidence for
causing acute leukaemia in human via the general process of the AOP described herein. For the other
Top2 poisons, including bioflavonoids, the evidence is weaker.

Etoposide is a semisynthetic derivative of podophyllotoxin that exhibits cytotoxicity by inhibiting DNA
synthesis as described above. However, if cells manage to bypass cell death, the accumulation of DNA
DSBs can lead to chromosomal translocations and further generation of fusion gene products
(particularly MLL rearrangement). Evidence supporting the causal relationship between etoposide-
induced TOP2 inhibition and the MLL rearrangement is strong regarding treatment-related acute
leukaemia (Cowell and Austin 2012; Pendleton et al 2014). Between 2 and 12% of patients that
receive epipodophyllotoxin develop secondary AML, with the mean latency period from drug
administration to the onset of secondary leukaemia being about 2 years. The risk of secondary AML
appears to be dependent on both treatment schedule and dose. Typically, epipodophyllotoxin-induced
AML occurs after multiple doses administered in brief intravenous infusions with cumulative doses
ranging from 5,200 mg/m2 to 19,200 mg/m2 (Ezoe, 2012). Dose-response relationships between
etoposide and treatment-related leukaemia are difficult to unravel, but risk of leukaemia seems to
increase with larger total exposure to etoposide. There is no doubt that the fusion genes are caused
by etoposide treatment because MLL rearrangements have not been detected in bone marrow
samples banked before the start of the treatment of the first malignancy (Pendleton et al., 2014).
Chemical-induced DNA breakpoints are associated with predicted Top2 cleavage sites (ie MLL), supporting an essential role for TOP2-mediated breakage. The high frequency of Top2 recognition sites in specific DNA regions and the high expression of this enzyme in human CD34+ HSPCs represent favorable conditions for breakage following exposure to Topo2 poisons. Because CD34+ HSPCs appear to be more sensitive to DNA damage than committed progenitor cells, exposure to low levels of different chemicals may induce DNA breakage at certain sites in HSPCs, increasing the risk of chromosomal rearrangements (Bueno et al. 2009; Montecucco et al., 2015; Thys et al., 2015; Hernández and Menéndez, 2016).

Studies on identical twins and neonatal blood samples strongly implicate an in utero occurrence of the key events (Sanjuan-Pla et al 2015). Furthermore, a study in pregnant mice demonstrated that in utero exposure of the foetus to etoposide causes the MLL chromosomal translocation analogously as in humans but with different gene fusion partners (Nanya et al., 2015). Indirect evidence from human prehaematopoietic/mesenchymal stem cells and foetal liver HSPCs strengthens the biological plausibility. Experimental evidence in these cell lines has demonstrated that etoposide causes DSBs in MLL and partner genes, which leads to the formation of fusion genes and their products (Sanjuan-Pla et al 2015).

Nanya et al. (2015) has shown that in utero exposure to etoposide induces MLL translocations in ATM-knockout mice, which are defective in the DNA damage response, but not in wild-type mice. Moreover, foetal liver HSPCs were more susceptible to etoposide than maternal bone marrow mononuclear cells, pointing out the life stage-related susceptibility in regards to Top2 inhibition also in the mouse. However, in utero exposure to etoposide failed to induce leukaemogenesis (Nanya et al 2015). Whereas etoposide can induce a large number of MLL rearrangements, most of them occur within non-coding regions, without eliciting direct oncogenic consequences. The appropriate oncogenic event needs to occur in a target cell within a relatively small and spatially restricted cell population during the appropriate and epigenetically plastic developmental window. Thus, it is a very rare event and difficult to support empirically.

Li et al (2014) developed a cell model based on the hypothesis that cells are capable of clearing low-level DNA damage with existing repair capacity. When the number of DSBs exceeds a certain value, ATM and p53 become fully activated through reversible mechanism, leading to elevated repair capacity. The dose-response relationships for activation of p53 and the formation of micronuclei in the target cell model indicate that critical concentrations of etoposide are in the range of 0.01 to 0.1 µM (Li et al. 2014). This range is in agreement with the increased levels of DSBs observed in human foetal liver CD34+ cells at a concentration of 0.14 µM of etoposide; however, MLL translocations were detectable at higher concentrations (Moneypenny et al 2006; Bueno et al. 2009).

Despite the limited number of studies investigating the role of chemicals in the pathogenesis of paediatric leukaemia, the consistent observation of the inhibition of Top2 activity suggests that this might be a key mechanism induced by chemicals with leukaemogenic potential. Aside from this, there are also other common mechanisms observed among the studies involving chemicals, such as oxidative stress.

b) Oxidative stress

Oxidative stress has been implicated in haematotoxicity induced by benzene and pesticides (Choi et al., 2016).

Under some circumstances, oxidative lesions can lead to DNA DSB formation in HSPCs. Environmental exposures to numerous chemicals, including many pesticides, have been shown in vivo and in vitro to generate reactive oxygen species (ROS) that can ultimately induce DNA oxidative damage, leading to single-strand breaks (SSBs) and DSB formation in the DNA (Sedelnikova et al., 2010). For example, OP insecticides (chlorpyrifos, methyl-parathion, malathion), methyl-carbamates (methomyl) and the herbicide paraquat all cause oxidative DNA damage followed by DNA SSBs and DSBs (Esperanza et al., 2015; Guanggang et al., 2013; Muniz et al., 2008; Ojha and Srivastava, 2014). There is also evidence of pesticide-induced oxidative stress and DNA damage in agricultural workers (Muniz et al., 2008). Additionally, oxidative species may interact with biological molecules to disrupt normal DNA synthesis and repair, and so inhibition/inactivation of antioxidant proteins or DNA repair enzymes may also be an underlying molecular mechanism (Kryston et al., 2011).
ROS are not known to directly cause DSBs, however DSBs could be generated if two SSBs oppose each other on complementary strands or could occur as secondary lesions at the replication fork or during an intermediate step in a repair process (Li et al., 2013). DNA DSBs are the most harmful initial event in molecular and cell carcinogenesis. Unrepaired DSBs may result in structural chromosomal abnormalities, whole or partial chromosome loss and genetic recombination, but can also lead to the breakdown of DNA replication, causing apoptosis to prevent a possible mutation being passed or during replication (ap Rhys and Bohr, 1997).

Specific oncogene activation in different tumour models has been linked to DNA DSBs and the activation of DNA-damage checkpoints. Efficient DSB repair is crucial for the maintenance of genomic integrity. In response to DNA damage, phosphatidylinositol-3 kinase-related kinases ATM (ataxia telangiectasia mutated) and ATR (ATM- and Rad3-related) are initially activated and subsequently phosphorylate a variety of proteins that regulate the DNA-damage response. DNA DSBs observed in some studies with chlorpyrifos and atrazine may be due to the ability of these compounds to generate highly reactive molecules/radicals (Huang et al., 2015; Lu et al., 2015). Aldicarb has caused a dose-dependent DNA damage, with single-strand breaks being produced by low concentrations during short time, which could be repaired, whereas high concentration led to DSBs which were difficult to repair (Li et al., 2003). Tetrachlorohydroquinone, the major toxic metabolite of pentachlorophenol, can induce DNA lesions as this metabolite contributes to the release of free radicals which have been linked to tumour promotion (Chen et al., 2015).

When the ROS are elevated beyond physiological levels, oxidative stress can cause HSPC dysfunction. These cells are extremely sensitive to oxidative stressors, such as anti-cancer agents, radiation, and the extensive accumulation of ROS. NADPH oxidase has been proposed for ionizing radiation-induced persistent and prolonged intracellular ROS generation in human CD34+ HSPCs, such that the resulting oxidative stress is associated with inhibition of the clonogenic potential of these cells (Yamaguchi and Kashiwakura, 2013).

ROS-induced DSBs in human foetal liver CD34+ HSPCs following maternal exposure to chemicals trigger recombination/repair pathways by non-homologous end-joining (NHEJ). The majority of damaged HSPCs may either successfully repair the DNA DSBs or fail to do so and undergo apoptotic cell death. If the DNA DSBs within particular breakpoint cluster regions (bcr) are not correctly repaired, chromosomal translocations or deletions may occur, although this possibility is very unlikely and would only happen in a small fraction of cells. For fusion genes to be leukaemogenic, DSBs must occur simultaneously in two chromosomes and must also involve the coding region of the genes to generate an exon-exon in-frame functional chimeric gene product. Importantly, this has to occur in a HSPC that has managed to bypass cell death and displays a sustainable lifespan and clonal potential to propagate the chimeric gene product (Hernández and Menéndez, 2016).

In the case of infant leukaemia, and based on the very short latency of the disease, it is not completely clear whether the fusion gene generated from chromosomal translocations requires additional cooperating oncogenic hits for leukaemogenesis. Recurrent activating mutations of genes associated with cellular proliferation, such as components of the RAS signalling pathway, are important for tumour maintenance rather than initiation in human HSPCs (Prieto et al., 2016). The transformation mediated by the aberrant proteins encoded by fusion genes might depend on alternative (epi)-genetic cooperating lesions at a critical developmentally-earlier window of stem cell vulnerability to develop overt leukaemia (Sanjuan-Pla, et al., 2015).

For childhood leukaemia, chromosomal translocations resulting in aberrant chimeric proteins alter the normal transcriptional program of HSPCs and block normal B-cell and/or myeloid differentiation. In contrast to MLL-associated infant leukaemia, chromosomal translocations linked to childhood leukaemia (i.e., TEL-AML1) are not sufficient to cause the disease by themselves. As TEL-AML1 fusion gene is observed in cord blood from about 1% of normal newborns, a significant proportion of the population carries self-limiting preleukaemic clones, the majority of which do not result in disease. The longer latency and that only a fraction of children carrying the translocation develop the disease unequivocally indicates that the initiating chromosomal translocation per se is unlikely to convert a preleukaemic clone into an overt disease, consequently secondary cooperating (epi)-genetic events are needed. In this regard, developmental dysfunction of the immune system followed by an aberrant immune response upon delayed infections has been linked to the development of childhood leukaemia (Greaves, 2002; Pui et al., 2008; Teitell and Pandolfi, 2009, Wiemels, 2012).
5.2 To what extent do experimental toxicity studies on mechanisms of toxicity cover mechanisms relevant for IFL and CHL, and what is the contribution of the AOP in supporting biological plausibility?

5.2.1 Rationale of the working approach

At present, the scientific evidence available sheds light on the following topics: pesticide exposure, toxicant MoA (e.g. etoposide), experimental studies, disease biology and the occurrence of paediatric leukaemia (including both infant and childhood leukaemia). The combined information allows understanding better whether the statistical association of pesticide exposure with the development of paediatric leukaemia is mechanistically plausible; whether causal links can be raised; and if such links can be confirmed or refuted by experimental testing.

As a starting point, there is sound epidemiological evidence in support of the association between pesticide exposure (either occupational or environmental/residential, either pre- or postnatal) and pediatric leukaemia. However positive data from regulatory toxicity studies linking pesticides to traditional endpoints (e.g. genotoxicity) are lacking. In addition, for some chemicals, a mechanism of action is known (e.g. etoposide), but this is not the case for pesticides for which the evidence is weak.

The first question relevant to be addressed was to evaluate whether a pathogenic mechanism could be assigned to paediatric leukaemia in form of an AOP, which could be done only for infant leukaemia but not for childhood leukaemia because of the lack of a clearly delineated molecular initiating events.

The second issue raised was to ascertain whether experimental studies provide information concerning the mechanism of toxic action of chemicals. The next step was then to investigate whether toxicity pathways of chemicals overlapped with the mechanisms supporting the disease pathogenesis (AOPs) relevant for infant and childhood leukaemia. Finally, the answer to these questions was used (see chapter 8.4.) to establish plausible links between exposure to pesticides and the risk of developing paediatric leukaemia.

5.2.2 Capturing complex diseases (IFL and CHL) by AOP

A major obstacle in developing AOP of events leading to adverse outcome in paediatric leukaemia stems from the complex nature of this disease. The molecular initiating event and the adverse outcome highly depend on the subtypes of the disease (e.g. infant or childhood leukaemia, lymphoblastic or myeloid leukaemia).

Furthermore, the most important restriction is that the AOP should not be defined for the disease as such, but for a sharply defined feature of the disease (as equivalent to an endpoint or adverse outcome for toxicants). A second important condition is that this endpoint can be reproduced in animal models, and that there exist chemicals that trigger the endpoint in the animal models. This implies that example chemicals are available that are likely to trigger the envisaged “disease AOP”. As such conditions were partially fulfilled herein (only for IFL), it was scientifically acceptable to work on model AOPs relevant to IFL but only an hypothetical AOP could have been developed for CHL.

Although the apical outcome is the same for those clinical conditions, the natural history of the disease is different for IFL and CHL and the MIE has been identified only for IFL. This can be considered as a developmental disease with all the relevant pathogenic steps occurring in utero and with an exceptionally short latency. In contrast, childhood leukaemia fits better to the two-hit model confirmed in the natural history of several cancer s and hypothesized for leukaemogenesis. The initiating hit would occur in utero and the promoting event would occur postnatally. Besides, whereas a single big hit is enough for IFL (as exemplified by MLL rearrangement), a similar hit is not enough to trigger childhood leukaemia.

5.2.3 Selection of the AO

Leukaemia is a group of cancers that presents with the proliferation of immature, clonal, myeloid or lymphoid precursors leading to progressive marrow failure and ultimately death. Clinical symptoms and signs of paediatric leukaemia usually reflect bone marrow infiltration by leukaemic blasts and/or extramedullary disease. The major clinical signs consist of neutropenia, thrombocytopenia and anaemia, with these signs being responsible of symptoms such as increased susceptibility to infections (with fever), bruising, bleeding, pallor and fatigue. Accordingly, many different AOPs might be...
associated with the disease because of the several adverse outcomes of the disease, as well as pathway may converge into the same AO or share the same KE. Despite leukaemia has been identified in several animals and animals models have been developed to study the disease, the disease as such is not an apical endpoint in the regulatory toxicity studies and a mechanistic understanding of apical hazard suggestive of the AO is needed.

For proof of concept, symptoms and signs of overt paediatric leukaemia were chosen as AO. However, human leukaemia features have not been fully recapitulated in experimental animal models and the major candidate endpoints consist of chromosomal translocations that show similarities with those found in humans (sometimes paediatric patient-derived leukaemia xenografts with MLL or TEL/AML1 translocations). The wide range of genetic and epigenetic changes needed for the expansion of preleukaemic clones prevents from delineating a sharp definition of paediatric leukaemia, either IFL or CHL.

5.2.4 Choice of example AOP relevant both for IFL/CHL and for pesticides as risk factors (see Appendix B)

Once the AO had been defined, the next question was to describe the sequence of pathogenic events that could be incorporated into a proof-of-concept AOP. For practical reasons, etoposide (a non-pesticide chemical; a chemotherapeutic drug currently used for the treatment of cancer at various sites) was chosen as a tool example for IFL leukaemia because of the sound and consistent evidence in humans and experimental animals pointing out the pathophysiological processes triggered by this chemical. However, no chemical has been identified so far with the capability of triggering the toxicological pathway leading to childhood leukaemia. Notwithstanding this limitation for building an AOP, the systematic literature review commissioned by EFSA (Ntzani et al., 2013) concluded that there was sound epidemiological evidence linking pesticide exposure at diverse developmental stages and paediatric leukaemia. Moreover, expert knowledge on the state of experimental paediatric leukaemia research was used. On this basis, the Panel decided to develop two relevant AOPs one of them based on data for etoposide and the second AOP was a putative one because of the lack of empirical data at clinical, cellular, mechanistic or regulatory level that support any particular chemical with the onset of childhood leukaemia. However, there is abundant data that could be used to define the rest of the corresponding AOP.

This decision process has some important implications for the interpretation of this scientific opinion. The most important one is that the AOPs developed herein fail to support strongly how the different pesticides can lead to any of the different types of paediatric leukaemia. Besides, as the initial molecular targets and biochemical pathways disturbed by a toxicant are highly chemical-specific, a pesticide AOP cannot be defined. Likewise, there is no a plausible mode of action that relates exposure to any individual pesticide (or pesticide classes) with paediatric leukaemia. Therefore, the PPR Panel decided to test whether the hazard posed by pesticides could be linked to the pathogenesis of paediatric leukaemia via AOPs. However, a single AOP may not capture all events that contribute to the relevant adverse outcome, instead sets of AOPs sharing at least one common event may capture more realistically potential toxic effects. If this approach is considered useful, then a multitude of AOPs could be developed for the many different pesticides currently used for improving our knowledge on their mechanism of toxic action. It can be anticipated that not all pesticides will fit any of these AOPs and also that several of these AOPs may share common key events, or may converge into common intermediate key events, which would allow the definition of partial AOPs and their connection to common KE.

5.2.5 Use of tool chemicals to check whether their mechanism of action overlaps with AOP for IFL and CHL

Etoposide was selected from the biomedical literature as the most promising tool chemical to build AOP that would describe its hazard. The rationale is that well-documented human and experimental data, both in vivo and in vitro, supporting the involvement of etoposide in the development of leukaemia. Its molecular target, topoisomerase II, is particularly well-characterized. Also, for infant leukaemia biology there is good evidence that this target plays a key role (Pendleton et al., 2014). In contrast, no tool chemical has been identified in the open literature for childhood leukaemia. Since data are lacking to delineate the molecular initiating event for this disease, efforts were made to fill a
conceptual gap by presenting a hypothetical framework that provides sufficient biological plausibility
based on an analogy approach derived from toxic mechanism following exposure to ionizing radiation.
Accordingly, the example chemical chosen for IFL allowed to define a MIE (topoisomerase II
inhibition/poisoning) whereas the lack of MIE for CHL restricted the exercise to develop a putative
AOP linked to the adverse outcome. The approach followed takes into account multiple genetic
susceptibility factors and modulating events such that it falls within “system toxicology”. Analogously
to systems biology, this approach is intended to decode the toxicological blueprint of an active
substance that interact with biological targets that function as a network in cells, tissues or organisms
(Sturla et al., 2014).
The two AOPs chosen by the PPR Panel can be fitted on this complex scenario as relevant pathways
and are consistent with current medical knowledge on the disease biology of paediatric leukaemia.
The prototype AOPs are fully reported in the Appendix B.

5.2.6 Evaluation of the AOP concerning consistency and strength of evidence

Data generated from experimental models collectively contribute to the weight of evidence supporting
the proposed AOP. A large part of the effort to develop AOPs was used for their evaluation, and the
documentation of this process. The strength of association was judged by a weight of evidence
approach based on modified Bradford-Hill criteria. This is fully described in the Appendix B.
Based on the overall weight of evidence, the PPR Panel concluded that the link between the MIE and
the AO as proposed in the developed AOPs for IFL is strong and that the proposed key events
(together with the MIE and the AO) can be used as a tool for exploring the IFL-triggering hazard of a
chemical. Once an AOP has been established, the MIE can be used to develop screening assays for
compounds that might affect the AOP and networks of interacting AOPs (Knudsen et al., 2015).
However, this framework is not valid for CHL because of the lack of a clearly defined MIE as
aforementioned. This limitation is partially overcome by a set of hypothetical mechanisms that can
provide a plausible biological basis for the epidemiological evidence gathered on the association
between pesticides exposure and CHL.
One key conclusion that can be drawn from this approach is that any chemical triggering the MIE, or
an intermediate key event of the proposed AOP, to a sufficiently quantitative extent, is also expected
to trigger the downstream key events and eventually the AO. This represents an important conceptual
advance in predicting chemical hazards in terms of increasing the risk for human disease. Another
important conclusion is the need of obtaining and using quantitative data as much as possible on the
key event relationships for a practical application of hazard predictions based on AOPs.

5.2.7 Support of hazard plausibility by AOP

Based on above considerations, the Panel is supporting the use of the AOP framework to explore the
biological plausibility of the epidemiological association between pesticide exposure and paediatric
leukaemia. The recommendation is that pesticides affecting the AOPs proposed in this opinion should
be considered as potentially hazardous with respect to the development of paediatric leukaemia. The
same would apply to other AOPs linked to this disease that could be developed in the future.
It is stressed again that the PPR Panel recommended the use of AOP specifically for hazard
identification, and not for the assessment of risk. According to the risk assessment process, once any
hazard has been identified, there is a need to proceed with a more complex evaluation of the risk. The
AOP framework is exclusively focused on hazard and does consider neither exposure data nor
toxicokinetic (including metabolism) processes. The series of biochemical reactions and pathological
events of an AOP by definition cannot have pharmacokinetic parameters. Dose of chemicals are taken
into account for defining a threshold above which individual compounds may trigger the pathway, and
they are evidently unique for each chemical but are not associated with the AOP as such.
Nevertheless, certain features of paediatric leukaemia should be considered for the hazard plausibility.
The scientific evidence undoubtedly indicates that exposure to chemicals takes place in utero and
even prior to conception (prenatal exposures). This assumption implies the need to account for
toxicokinetic factors as chemicals will need to go across the placenta to reach foetal targets. In
addition, many compounds are not active by themselves but need to be bioactivated by maternal (and
to a lower extent foetal) biotransformation processes as occurs with etoposide for infant leukaemia or
with benzene for adult myeloid leukaemia. The active chemical, either the parent compound or a metabolite, needs to reach the proper target in the embryo/fetus at a proper time window of development and at a concentration high enough to trigger the initiating events defined in the AOP. However, these considerations are not directly related to the chain of pathogenic events involved in the AOP as these are meaningful only for hazard plausibility. Another feature of paediatric leukaemia is related to toxicodynamic factors since differences in susceptibility regarding ontogeny processes may be relevant. For instance, IFL is considered a 'developmental disease' showing different features and pathogenesis than childhood leukaemia, as more immature haematopoietic precursors are involved. The physiological role and susceptibility of these precursors to chemicals may vary depending on the embryonic/foetal stage of development.

5.3 Data gaps and suggestion for testing strategy. Also include the AOP as informative source for appropriate identification of data gaps and testing strategy

5.3.1 AOP as a scaffold to help identifying data gaps

Since the AOP concept shows causality of events under a weight of evidence approach, it is very well suited for identifying data gaps. Based on the epidemiological data linking pesticide exposure to paediatric leukaemia, and the AO being defined as 'overt leukaemia', several modes of action were identified linking an initiating event to the key event essential for the AO. This essential KE is the chromosomal translocations within HSPCs, which are cells essential for haematopoiesis. Thus, for the AOPs developed, the causality of substance binding to and subsequent inhibition of Top2, non-repaired DNA DSBs or leading to chromosomal translocations, differentiation block of HSPCs, clonal expansion of preleukaemic clones and overt paediatric leukaemia is biologically plausible and essential. However, the main challenge of developing AOPs for leukaemia is the complex nature of the disease. For example, a tumour suppressor gene could be mutated or transcriptionally inactivated while in another instance an oncogene could be activated to trigger leukaemogenesis. Different genetic aberrations are associated with different subtypes of leukaemia. In addition, although leukemia is a cancer with a low mutation rate, paediatric (childhood and infant) leukemia are the second cancer with the least somatic mutations of all cancers sequenced so far (Bardini et al. 2010, 2011; Dobbins et al 2013; Andersson et al 2015). This stable genome makes difficult to unravel the etiology and pathogenesis of paediatric cancer and there are no many genetic tags to be traced back for associating exposure to specific compounds and then validate the pathway.

Assessment of data gaps within an AOP is feasible by analysing the weight of evidence for each KER within an AOP. For IFL, the KER ‘In utero exposure to DNA Top2 poisons leading to MLL chromosomal translocation’ has a strong WoE, although there are still some open questions. For instance, the appropriate target cell model that recapitulates the production of DSB as a result of Top2 ‘poisoning’ has not been identified so far. Approximately 80% of IFL cases have the MLL rearrangement, but the remaining 20% carry other chromosomal aberrations leading to different fusion genes that eventually result in the same leukaemia phenotype. In utero etoposide-treatment in a murine model, failed to induce leukaemogenesis because the appropriate chromosomal rearrangement is a rare event that needs to occur in a target cell within a relatively small and spatially restricted cell population during the appropriate, epigenetically plastic, developmental window. Moreover, although the risk of IFL seems to increase with larger total exposure to etoposide, dose-response relationships between etoposide and treatment-related leukaemia are difficult to unravel. In contrast, for CHL there is no evidence at the molecular level as to to how some chemicals interact with biological targets to elicit DNA damage. This is not a straightforward question as genetic damage in HSPCs may be properly repaired in most cases, but if not cells undergo apoptosis. The exact nature of how and when this damage is not repaired has not yet been clarified as many factors are involved, thus contributing to a stochastic process with the final occurrence of the disease being very unlikely. The genetic damage (i.e. chromosomal translocations) has to occur in a particularly vulnerable genetic locus, within the proper cell, and in a specific time window; however, details of this entire process and how it happens are not clear.

The second level of KER for IFL ‘In utero MLL chromosomal translocation leading to infant leukaemia’ also has a high weight of evidence as the potential of both differentiation blockage and clonal
expansion are inherent properties of the MLL-rearranged fusion product. Thus, weight of evidence, indicates that IFL originates from one ‘big-hit’ occurring during a critical developmental window of stem cell vulnerability. However, although the MLL-rearrangement is essential to develop leukaemia, it alone may not be sufficient and further (epi)genetic factors would contribute to convey a proliferative advantage to preleukaemic clones to develop overt leukaemia. On the other hand, the MLL-AF4 knock-in mice developed leukaemia only after a prolonged latency, thus not recapitulating an important feature of IFL.

For CHL, the KE ‘**In utero** chromosomal translocations leading to differentiation arrest of HSPCs’ has a high weight of evidence as this process has been very well studied, although the identity of leukaemia-initiating mutations that result in preleukaemic clones is still an important open question. The block of differentiation of HSPCs confers self-renewal properties to these cells and provides proliferative advantage to lymphoid progenitors. However, chromosomal translocations are insufficient by themselves to cause overt disease. Additional postnatal events are needed for the development of full-blown disease, but they are not yet sufficiently well understood. Experimental models should be developed in cell lines and in mice to accurately recapitulate human leukaemogenesis. Additionally, oncogenes and chromosomal translocations should be studied in the appropriate cellular context, which consist of primary human haematopoietic cells. If the initiating oncogenic alteration is not occurring in the right cell, mouse models would unlikely recapitulate the human disease and would constitute an inaccurate model of human leukaemia.

The KE ‘**Differentiation arrest of HSPCs leading to clonal expansion of leukaemogenic cells**’ has a sound weight of evidence as murine models with human precursor cells harbouring the TEL-AML1 fusion gene need the acquisition of additional genetic abnormalities to result in overt leukaemia. However, the reproducibility and accuracy of these models have yet to be validated for humans, providing a data gap. Functional studies are needed to unveil the key mechanisms driving the evolution of these progenitors/stem cells into the appropriate type of leukaemia. Besides, individual patients harbour multiple genetic subclones of leukaemia-initiating cells, with a complex clonal architecture which limits to build a consistent AOP. Owing to the technical challenge of distinguishing and isolating distinct cancer subclones, many aspects of clonal evolution are poorly understood. For instance, it remains to be demonstrated to what extent epigenetic diversity contributes to subclonal heterogeneity in acute leukaemia.

There is scarce scientific evidence for the KER ‘clonal expansion of leukaemogenic cells leading to overt childhood leukaemia’ since there are data gaps in the precise understanding on how leukaemic clones grow and expand. However, the biological plausibility of this KER is large as the pathobiology of the disease together with its evolutionary genetic landscape clearly indicates a causal linkage between the expansion of leukaemic clones within either the myeloid or lymphoid lineage and the onset of clinical phenotype of the disease.

### 5.3.2 Conclusions from AOP on suitability of current testing methods and present data gaps in regulatory studies

According to Regulation 1107/2009 on placing of plant protection products on the market, carcinogenicity and haematological endpoints must be evaluated for hazard identification and characterization of active substances in order to decide on their approval. In addition, Regulation 283/2013 setting out the data requirements for active substances indicates that genotoxicity and carcinogenicity studies are always required and that haematological endpoints will be addressed and reported in general toxicity studies (repeated doses - short-term and long term - and reproductive toxicity studies). Haematological endpoints addressed in these studies include red blood cell parameters, total and differential leukocyte count, platelet count and blood clotting time among others. In addition, haematopoietic organs are investigated in repeated dose and carcinogenicity studies, so that substances inducing leukaemia in rodents are expected to be identified in the basic data set.

The adequate evaluation of the genotoxic potential of a chemical is consistently addressed in regulatory dossiers by the assessment of different endpoints, i.e. induction of gene mutations, structural and numerical chromosomal alterations. These endpoints can only be covered by the use of diverse test system as no individual test can simultaneously provide information on all of them. The bacterial reverse mutation assay covers gene mutations and the *in vitro* micronucleus test allows
the identification of both structural and numerical chromosome aberrations. If all these endpoints are clearly negative the substance can be reasonably regarded as devoid of any genotoxic potential. Conversely, in the case of inconclusive, contradictory or equivocal results from this basic battery of tests, further in vitro testing must resolve the situation. In the case of positive results, further tests in vitro are appropriate either to optimise any subsequent in vivo testing or to provide additional useful mechanistic information. In vivo tests (mammalian erythrocyte micronucleus or transgenic rodent gene mutation assays) should relate to the genotoxic endpoint(s) identified as positive in vitro and to appropriate target organs or tissues (EFSA, 2011). According to this testing strategy, a substance inducing leukaemia by a genotoxic mode of action is supposed to be captured by the genotoxicity tests battery.

However, the cell lines or primary cell cultures routinely used for regulatory in vitro genotoxicity testing may not be representative of HSPCs. The cell systems used in the different tests consist of adult cells and it is a widely recognised assumption that exposure of these cell systems at high dosages (until cytotoxicity occurs) would stress the cells to the extent that genotoxic properties would be found if the chemical was actually genotoxic. However, very early HSPCs have been considered particularly sensitive to genotoxicity because of the unfolded nature of their DNA, immature repair systems, high division rate, high Top2 expression and activity level, etc.

Concerning assessment of carcinogenicity and haematological end-points, the design of regulatory studies does not include a prenatal exposure. While a specific window of exposure is explored in the two-generation reproduction toxicity studies, haematological measurements and histopathology of haematopoietic organs are not performed.

The only regulatory test guideline where haematology and histopathology of haematopoietic organs are investigated in animals exposed in utero to the testing chemicals is the extended one-generation reproroduction toxicity study. However, only 10 male and 10 female rats per group are examined for those parameters in order to assess the potential impact of the substance on the immune system. Nonetheless, this regulatory study is not intended to explore any carcinogenic event, which would require 50 animals/sex/group. Since this guideline has been adopted recently (2011), this kind of study has been submitted in very few cases compared to the two-generation reproduction toxicity study.

For adult leukaemogenesis induced by external factors, the following four main patterns were outlined by the US-EPA (Eastmond, 1997): a) the primary type of lymphohaematopoietic cancer induced by chemicals and ionizing radiation in humans is myeloid leukaemia; b) potent human leukaemia-inducing agents produce significant myelotoxicity and structural chromosomal aberrations in humans and animal models; c) administration of human leukaemia-inducing agents to mice results in more lymphohaematopoietic tumours that, unlike to happen in humans, are primarily lymphoid in origin; d) the rat is considerably less responsive than the mouse to the induction of lymphohaematopoietic neoplasia following administration of human leukaemogenic agents.

Chemical- and radiation-induced lymphohaematopoietic tumours are complex processes involving multiple genes, chromosomal alterations and altered differentiation. In addition, other factors such as metabolic capabilities, DNA repair and genetic susceptibilities may influence cancer incidence. Given the complexity and multiplicity of steps, animal models are unlikely to reproduce precisely all the critical stages involved in development of chemical-induced leukaemias or lymphomas in humans (Eastmond, 1997).

Other limitations of rodent models for assessing the risk of leukaemias are represented by the different classification schemes of haematopoietic neoplasms used for rodents and humans. In mouse, histopathological distinction between malignant and non-malignant myeloproliferations is also hard to establish. Likewise, distinction between lymphoma and leukaemias is often difficult, particularly in mouse, which can lead to misclassifications. Rats are relatively resistant to chemically induced leukaemogenesis; however, Fischer 344 rats, which are commonly used in carcinogenesis studies, exhibit a high incidence of spontaneous large granular lymphocyte leukaemia (LGLL). In contrast, spontaneous leukaemias are rare in other rat strains. The background incidence of LGLL in F344 rats has increased over time reaching more than 50% in males (Irons, 1994). Therefore, the usefulness of this strain for haematopoietic neoplasms exploration is questionable. Despite significant interspecies differences, rodents are valuable models for immunotoxic and myelotoxic effects, including
leukaemogenesis. Furthermore, chronic animal bioassays using mouse models have been shown to be effective in identifying human (adult) leukaemia-inducing agents (Eastmond, 1997).

Paediatric leukaemias represent a diverse group of diseases with distinct biological features compared with adult leukaemias. B-cell acute lymphoblastic leukaemia, the most frequent leukaemia found in children, is characterized by an uncontrolled expansion of immature B-cell (pre B phenotype). However, in the particular case of infant B-cell acute lymphoblastic leukaemia, a very early haematopoietic precursor (pro-B phenotype) is involved. These progenitor cells are initiated in utero, usually as a result of structural or numerical chromosomal aberrations and/or gene mutations. A wide range of acquired chromosomal translocations have been associated with early stages of acute leukaemias pathogenesis, with MLL gene being frequently involved. Special techniques not routinely performed, such as fluorescence in situ hybridisation (FISH), can be combined to classic genotoxicity protocols in order to identify specific translocations with dedicated probes. This would allow obtaining additional mechanistic information.

In summary, assuming that the critical events of paediatric leukaemia consist of in utero induction of chromosome aberrations followed in cases of childhood leukaemia by an aberrant post-natal immune response to common infections, it is evident that the current animal tests/models do not cover these critical events.

### 5.3.3 Consideration on testing strategy

The above considerations point out that the current testing paradigm is not able to detect the potential leukaemogenic hazard posed by exposure to chemicals during early life stages (prenatal and early postnatal). There appear to be different sensitivities between cells for in vitro genotoxicity testing (notably HSPCs are considered more sensitive to genotoxic damage than other cells) and some in vivo tests (ie., the chromosomal aberration test and the micronucleus test) have shown a poor sensitivity, likely because of the low exposure of haematopoietic cells in vivo. Besides, the carcinogenicity study design does not cover the relevant window of exposure and the model does not include a second hit that has been capture in experimental models (ie., aberrant immune response to delayed infections). The only test guideline that covers this developmental period of susceptibility is the extended one generation test, in which haematology parameters and histopathology of haematopoietic organs are assessed on animals exposed in utero and during the juvenile period. However, this guideline has a low power to detect the leukaemogenic potential of a chemical because of the low number of animals examined. Besides, since this is a recent testing protocol there is scarce data on chemical substances already on the market.

The EFSA Scientific Committee recommended a step-wise approach for the generation and evaluation of data on genotoxic potential. This approach consist of: a) a basic battery of in vitro tests; b) consideration of whether specific features of the test substance might require substitution of some of the recommended in vitro tests by other in vitro or in vivo tests in the basic battery; c) in the event of positive results from the basic battery, review of all the available relevant data on the test substance; and d) where necessary, conduct of an appropriate in vivo study (or studies) to assess whether the genotoxic potential observed in vitro is expressed in vivo (EFSA, 2011).

For a practical testing approach to leukaemogenesis potential of chemicals, in vitro genotoxicity should be tested in the relevant cells, particularly HSPCs, and technologies should be applied to detect structural or numerical chromosomal abnormalities (ie, FISH). Besides, evidence of target cell exposure is necessary for in vivo tests. In relation to whether or not the carcinogenicity studies (or the combined chronic toxicity/carcinogenicity studies) are appropriate to capture the carcinogenic potential of chemical exposures during developmental phases, consensus among the scientific (and regulatory) community is needed to reach sound and feasible recommendations as to how to proceed. It seems reasonable that improved models for in vitro testing should be used as a screening tool and that optimized carcinogenicity studies should be triggered when sufficient positive evidence has been obtained from lower tier tests. AOP can be used to design a testing strategy.
6. Application of the AOP concept to support the regulatory process; using parkinsonian motor deficits as an example.

AOPs are not chemical specific and do not include ADME data of the chemical. Therefore, the Panel decided to evaluate the effective application of the AOP concept to the regulatory process by designing an exemplary strategy based on the two AOPs relevant for PD. The choice of the Panel was mainly motivated by the completeness of these two AOPs. In this process, the Panel is proposing to use the AOPs to build up an IATA (Integrated Approaches to Testing and Assessment) strategy to evaluate whether the exposure to a pesticide (e.g. triggering mitochondrial dysfunction) causes dopaminergic neurodegeneration and ultimately parkinsonian motor deficits.

The AOP-based KE testing cannot be used as a stand-alone approach but needs integration of ADME data and embedding into an IATA framework. This will give the confidence that the threshold of KEs activation will indeed trigger the full cascade of events and that a dose, temporal and response concordance is maintained.

In designing a test battery for parkinsonian motor deficits in the context of IATA the following considerations should be taken into account:

- Duration of exposure e.g. how long proteostasis needs to be impaired for inducing DA neuronal death
- In vitro concentrations relevant to hazard assessment
- Metabolic capacity of the test system
- Assays permitting evaluation of KERs and modulatory factors or measuring recognized biomarkers reflecting KERs activation
- Predictive capacity of the assays e.g. taking into account a role of glial cells, species differences etc.

6.1. Which types of data are needed to predict risk of parkinsonian motor deficits applying an AOP-informed IATA.

Elaboration on the IATA proposal is not part of this mandate; however, for understanding the value of an AOP-based testing strategy and placing the AOP in a larger context for any potential application in risk assessment, the Panel considered an introduction to the AOP-based IATA as necessary. For the proposed AOP-informed IATA ‘Assessment of nigrostriatal toxicity’ the Panel adapted the general IATA concept from the AOP-informed IATAs for skin irritation and corrosion (NV/JM/MONO(2014)19). Ten modules were identified for assessing if a compound poses a hazard for inducing Parkinsonian motor deficits in humans (Table 5).

Table 5: Parts and Modules of the proposed IATA for assessment of nigrostriatal toxicity (adapted from NV/JM/MONO(2014)19).

<table>
<thead>
<tr>
<th>Part 1 (Existing information, physico-chemical properties and non-testing methods)</th>
<th>Module</th>
<th>Data</th>
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<tr>
<td>1</td>
<td>Existing information - Existing human epidemiological data</td>
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<td>2</td>
<td>Existing guideline studies including nervous system evaluation (OECD TG 424, 452,453)</td>
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<tr>
<td>3</td>
<td>In vitro neurotoxicity data - Other in vivo and in vitro data with the focus on dopaminergic neurons</td>
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<tr>
<td>4</td>
<td>ADME data - Physicochemical properties</td>
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The first step consists in collecting all available data (including regulatory toxicology studies) and human epidemiological information (Modules 1 & 2) followed by integration of toxicokinetic information (information on ADME, Module 3; Table 1, Figure 1). Moreover, information on physicochemical properties, e.g. compound’s redox potential, are gathered (Module 5). Here, the focus lies on the compounds’ abilities to (a) be taken up by dopamine transporter or other transporters, (b) generate ROS, or (c) interfere with complexes of the mitochondrial respiratory chain. Apply (Q)SAR and read across where possible (module 6). All these data should be evaluated by a WoE approach (Module 7). The WoE approach should be structured and possibly quantitative, should inform on data gaps for decision making. At this stage, decision should be taken if new data are needed. However, it is beyond the scope of this opinion to detail on WoE analysis in general, but guidance on transparent WoE analysis is being developed by EFSA. Part 3; if new data are required, an AOP-informed IATA will be used to set-up the most appropriate testing approach (module 8 to 10 and Figure 1). After data generation, the WoE has to be re-evaluated and decision taken on hazard assessment.
The Panel recommends that the following points are considered for an AOP informed IATA for ‘Parkinsonian motor deficits’.

- Should be based on human cells when necessary
- Testing for neurotoxicity performed only on neuronal monoculture might not be sufficient because glia cells could be modifiers of the toxicity; either by promoting toxicity through e.g. pro-inflammatory stimuli or by buffering toxicity through e.g. activation of the cellular antioxidative defense (Henn et al. 2009, Efremova et al, 2015 and 2016).
- Dopaminergic neurons should be the preferred neuronal subtype for testing.
- In cases where cell-cell contacts affect cellular responses a 3D format may be considered (Alepee et al. ALTEX 2014; Yamada & Cuckierman Cell 2007).
- Cells used for testing should be without ethical concern and of high availability. hiPSC (human induced pluripotent stem cells) are envisioned to solve the ethical issues and limitation of material when working with cells of human origin. They can be differentiated into (dopaminergic) neurons and astroglia functioning also in vivo when transplanted into rodent brains (Wernig et al. 2008; Palm et al. 2015).
- The result of the tests must be in line with the temporal and concentration concordance as described in the AOP.
- Apply quality control and good cell culture practice (GCCP) principles to ensure reproducibility of the results.
Initially, the testing strategy should consider all KEs followed by selection of the most predictive assays.

According to the proposed AOPs, the following endpoints could be considered for testing: 1) Oxidative stress, 2) Mitochondrial dysfunction, 3) Proteasomal dysfunction, 4) α-synuclein accumulation and 5) Dopaminergic cell death (Fig. 2). Moreover, the testing strategy should include considerations on the temporality of events as described in the AOPs, by selecting the appropriate timing for endpoint measurements which has to reflect the the time concordance. E.g. oxidative stress as KE1 should occur before disturbance of proteasomal function as KE3.

The Panel also recommends that in the context of these AOPs, length of treatment/exposure should allow for triggering the KE down (Betarbet et al. 2000, Fornai et al. 2003). The Panel does not expect this battery consisting of, for example, 5 assays (Fig. 2) measuring the endpoints mentioned above, to be the final testing strategy, but recommends a statistical analysis of data with a test set of compounds for selecting a minimum amount of tests that would predict the adverse outcome with the highest predictivity value. The Panel is also recommending that a tiered approach should be considered.

**Fig. 2:** Proposed KERs-based testing strategy established on the AOPs for ‘Parkinsonian Motor Deficits’. Assay 1: measurement of oxidative stress; Assay 2: assessment of mitochondrial function; Assay 3: determination of proteasomal function as a measure of proteostasis; Assay 4: identification of α-synuclein accumulation; Assay 5: assessment of dopaminergic cell death.

The structure provided by the ten modules described above (Table 1) allow for composing an IATA. Ideally, this IATA should make the maximum use of existing data, being resource efficient and minimising or eliminating the requirement for animal experiments.
7. Uncertainties

A number of uncertainties were identified by the Panel for each AOP developed; such uncertainties are reported in detail in the Appendices. The Panel has however identified general uncertainties concerning human epidemiological studies, experimental evidences and AOP development methodologies.

- Epidemiological studies

Human data from epidemiological studies and meta-analyses have been used in this Scientific Opinion to define the adverse outcome (AO) and this is considered an uncertainty per se due to the known intrinsic limitations of the epidemiological studies.

Definition of the human health outcome, when this is a complex, multifactorial human disease was considered by the Panel as an uncertainty. This issue is exemplified by epidemiological studies showing an association with “childhood leukemia” (i.e., the diagnosis of leukemia in infants and children), whereas the AOP development led to the identification of two distinct diseases i.e. Infant leukemia and Childhood leukemia. The current epidemiological data do not allow on concluding whether the association with pesticide exposure concerns infant and/or childhood leukemia. This problem leads to significant data heterogeneity in the epidemiological studies.

Exposure estimates was considered by the Panel as a major limitation and uncertainty of the epidemiological studies. Indeed, this uncertainty includes two components: a) the generic definition of the substances of concern which in most cases refers to large usage groups of diverse pesticides (e.g., “insecticides”) and only in some cases refers to pesticide structural groups (“organophosphates”, “chboro-S-triaines”). This generic definition of exposure cannot identify the individual substances contributing to the risk; b) lack of detailed quantitative information concerning internal exposure. For chronic diseases, such as PD, it is difficult to integrate biomonitoring (biomarkers of exposure) and epidemiology (health outcomes) information, leading the investigators to use methodologies with low accuracy, e.g. pesticide usage. Furthermore, in a realistic field scenario, humans are exposed to several substances and coformulants contained in actual pesticide products, representing an additional source of uncertainties.

The design of epidemiological studies and the heterogeneity of the target populations may introduce a considerable heterogeneity among studies that apparently investigate the same outcomes weakening the reliability of meta-analyses and this was considered by the Panel as an uncertainty.

An uncertainty directly relevant to AOP development is the limitation of knowledge about the natural history of multifactorial human diseases, involving the presence of different phenotypes, genetic factors and environmental factors, other than pesticides. This limitation of knowledge bears a direct effect about the characterization of KEs as well as of modulating factors involved in the KERs.

- Animal studies

Regulatory toxicity studies have shown intrinsic limitations leading to uncertainties (Papparelle et al. 2013). These are also relevant in relation to this scientific opinion as regulatory toxicity studies may not include endpoints that model or predict the relevant adverse outcomes deriving from the mechanistic understanding of human health outcomes. The Panel also recognises lack of (human) disease-specific animal models as an uncertainty. This lack is due to the problem that the adverse outcome is complex and multifactorial and only limited knowledge is available. In particular, the Panel notes the deficiency in predictive in vivo models for both IFL and CHL.

- AOP development

Some uncertainties in the process of AOP development were identified by the Panel.

Lack of criteria for data selection (assembling evidence) might account for differences in the strength of an AOP. In addition there is uncertainty about the consistent way to include the role of modulating factors into AOP development, particularly when dealing with AO derived from complex human diseases. Finally, there is uncertainty whether the developed AOP are the only ones linking PD, IFL and CL to pesticide toxicological MoAs.
8. Discussion and Conclusions

Human health risk assessment for pesticides is mostly based on experimental toxicity studies performed in laboratory animals. These studies are conducted at relatively high doses with the highest dose expected to be the maximum tolerated dose for the experimental animal species and strain to be tested and for the pre-defined study design. The outcome of these pivotal studies is extrapolated to humans exposed to relatively low environmental doses. With some exceptions, human data are only available through the epidemiological studies which are called to be incorporated into the risk assessment according to Regulation N. 1107/2009 when available, and as indicated in Regulation 283/2013 setting out data requirements for active substances. It is then essential for the evaluation of epidemiological data to weigh, integrate and make use of all the available information coming from multiple experimental studies. This complex task is getting even more difficult when epidemiological data are dealing with multi-factorial, multi-hit, chronic diseases, for which toxicological models or disease-specific animal models are limited. This Scientific Opinion is exploring methods and principles to guide and investigate the use of experimental data and available knowledge; it aims at developing a mechanistically-driven approach that aims to evaluate the evidence of cause-effect relationship i.e. biological plausibility and coherence. As described in the mandate, the Panel selected Parkinson's disease and childhood leukaemia as human health outcomes based on the associations observed for these diseases and exposure to pesticides which are consistently reported in multiple meta-analyses.

In this attempt to integrate epidemiological studies on pesticide exposure and human diseases by developing AOPs to assess the biological plausibility of the associations, the Panel recognised a number of limitations. The premise of the mandate, i.e. that pesticide exposure is associated with PD and CHL, in itself has some uncertainty/limitations in the nature of the epidemiological data. These arise especially from the lack of detailed quantitative information regarding exposure and data heterogeneity. In regard to the link between pesticide exposure and PD, since the Ntzani report (NTZANI et al. 2012) the association has been further consolidated in a systematic review by Hernández et al. (2016). In this work the authors could confirm that PD is significantly associated with pesticide exposure; however, they could not conclusively identify a single pesticide, or a specific pesticide group, associated with a significant risk of PD besides paraquat. Regarding the epidemiological evidence of pesticide exposure and the risk of CHL since the Ntani report (Ntzani et al. 2012), the association has been investigated by additional meta-analyses. Again the association between prenatal occupational exposure was observed both for ALL and AML, but recognising the considerable uncertainty in regard to the assessment of pesticide exposure. Again, besides an association with "generic insecticides" the data did not allow to conclude about a single pesticide, or a specific pesticide group, associated with a significant risk of CHL.

To summarise, assuming that the observed outcomes are not due to other confounders, to date they cannot be linked to a specific pesticide active substance, but only to the exposure to pesticides. However, exposure is rarely to a pesticide active substance alone, but to pesticide formulations and/or multiple active substances. In reality a plethora of co-formulants that are largely uncharacterised in regard to toxicological effects (other than acute toxicity by different routes, irritation and sensitization) should be considered. The recent assessment and discussion of the epidemiological findings between exposure to glyphosate and different cancer outcomes (RAR glyphosate 2015, Addendum 2015) also highlighted the major uncertainties in associating exposure to a certain pesticide active substance to an adverse outcome, concerning not only the lack of quantitative exposure estimates, but the simultaneous co-exposure to other co-formulants. Another critical issue, that was also relevant for the conclusion of the EU assessment in regard to the carcinogenic potential of glyphosate, is the strength of the biological plausibility of the epidemiological observations.

Why an AOP?

The terms of reference of this mandate included exploring the use of the AOP framework for supporting both a mechanistic-driven hazard identification and biological plausibility of epidemiological associations, in order to incorporate the human health adverse outcome as part of the hazard identification process. The AOP framework was thus selected as a flexible and transparent tool for the review, organization and interpretation of complex information coming from different sources. In this perspective, the AOP was intended to overcome one of the major limitations of many epidemiological studies i.e. lack of understanding of biological plausibility and this aspect has been investigated in this opinion.
The Panel, in proposing the AOP framework as an integral part of this mandate was also considering several aspects which were considered to be relevant namely: exploring how to possibly improve toxicity testing in regard to effects involved in complex and multifactorial diseases like PD and CHL; integration of in vitro predictive tools into testing in order to shift towards a more “toxicity pathways” –based framework; highlight species differences or similarities; identify data gaps, research needs and requirements for development of toxicological assays and IATA. All these aspects have to be dealt with in order to support regulatory decision in a scientifically robust framework.

To support his mandate, the Panel committed a systematic review specifically tailored to serve as a basis for defining and mapping the causal linkages between an MIE and a final AO and possibly identify all relevant publications related to the mechanisms and chemicals involved in the pathogenesis of PD and CHL. In case of PD this turned out to be a considerable number, making it a challenge to select the relevant publications but at the same time avoid “cherry-picking”. The Panel noted that the use of the systematic literature review framework also showed some limitations. Using the outcome of the systematic literature review in building an AOP, it was evident that relevant information was sometimes not captured despite the very large number of publications retrieved by the search. The most likely reason was explored and identified as the use of a structured search protocol with strict inclusion and exclusion criteria. Although it was recognised that an extensive literature search would have been more appropriate, the relevant scientific expertise present in the working group in the field of neurodegenerative diseases and childhood leukaemia with knowledge of the relevant available literature, overcame this potential limitation. As part of the process the working group of the Panel also met with authors of relevant publications to gain methodological details which were considered important in the context of the AOP framework.

Indeed, an AOP need not necessarily be based on a systematic literature review. The quality assessment of the literature is filtered and guaranteed by the strength of the structured weight of evidence analysis proposed in the AOP framework; this includes the evaluation of the empirical support to assess the reproducibility, dose, time and response concordance for the selected chemical tools.

It was noted that the regulatory acceptance of an AOP where the majority of the components e.g. the empirical data supporting a KER, is not supported by reproduced data would be very limited. Such an AOP could however, serve the purpose of identifying where more data should be generated.

An additional consideration for the regulatory use of AOPs is variability. Variability (due to intrinsic factors such as genetic polymorphisms, species, age, gender, as well as “environmental” factors such as diet, lifestyles etc.) is likely to be a considerable contributing factor for some KEs. To some extent this is already considered and discussed in building an AOP, but it is recognized that modulating factors should be given more space in the AOP development. Whereas the role of modulating factors is a general issue in toxicology (e.g. the use of uncertainty factors), to understand how modulating factors can impact on a threshold is a specific challenge for AOP development. Indeed, by properly accounting for modulating factors, AOP might provide a scientific background to build up specific uncertainty factors.

The Panel also observed that the studies included in the empirical support can be quite heterogeneous in terms of design and route of exposure, and this can be interpreted as a source of inconsistencies. However, some elements of the study design are important for hazard characterisation and ultimately for risk assessment. For example the intra-peritoneal route of administration is usually not considered a relevant exposure route for pesticide risk assessment since this route would be most unlikely for humans. The current toxicological studies conducted in laboratory animals are intended to serve first for hazard identification and then for hazard characterisation, in order to determine a suitable point of departure (NOAEL or Benchmark Dose). Accordingly, in the context and scope of this Scientific Opinion, the Panel intended the development of AOPs for the purpose of solely hazard identification to support biological plausibility based on mechanistic knowledge. If hazard identification was based on a route of administration not relevant for risk assessment, the Panel still considered this to be acceptable and in line with the principle for developing AOPs. However, for a quantitative AOP with the purpose of being used for hazard characterisation this would need different considerations, again depending on problem formulation. If, for example, the scope is to define a threshold able to trigger the sequence of KEs from the MIE to the AO, the route of administration of the used tool compound will still not be relevant; instead, the concentration(s) at the target(s) able to activate the MIE and
KE(s) will be relevant, independently of the route of administration. Toxicokinetic and metabolism information will be indeed very important when dealing with compound specific hazard characterisation by applying the MOA and/or IATA framework; this information will tell the risk assessor if the concentration of the specific compound at the target MIE will be relevant or not for its activation. In any case, the doses used in the empirical support with the tool chemicals should always be assessed to define the strength of the dose response concordance. In addition, as well as for the animal studies, effects detected at excessive doses – close to the maximal tolerated dose/cytotoxicity – would always require a careful assessment of the biological relevance of the observed finding.

As detailed in this opinion the core studies of the regulatory dossiers do not necessarily capture the potential hazard of pesticides in regard to PD and CHL. This is not unexpected when considering the complexity of these diseases and the fact that regulatory toxicology studies are intended and designed to explore multiple hazards and should be considered as standalone experiment i.e. one species, one strain, one NOAEL for the endpoints explored in the context of the study design. For the purpose of analysing the biological plausibility linking human health outcomes to pesticide exposure, AOPs can serve as an important tool, particularly when the regulatory animal toxicological studies are negative but the evaluation of the apical endpoint (or relevant biomarkers) was considered inadequate based on the AOP.

The scientific and regulatory relevance of AOP at the different levels of maturity (whether putative, qualitative or quantitative), would depend on the fitness-for-purpose in a given context; the problem formulation will therefore drive the building of an AOP, with the expectation that the AOP reflects the current knowledge and the WoE evaluation is transparent and complete.

A putative AOP is intended as a set of hypothesized key events and key event relationships primarily supported by biological plausibility. The Panel considered that for the problem formulation, as expressed in the terms of reference, a putative AOP can be useful in order to give indications on the strength of the relationship between the AO (intended as a human health outcome) and pesticides affecting the pathway. In addition, by detecting and/or identifying data gaps and/or research needs, putative AOPs could serve to inform IATA or give guidance for further works.

The Panel considered that qualitative AOPs (intended as an AOP including the assembly and evaluation of the supporting weight of evidence following the OECD guidance for AOP development) should be the starting and standard approach in the process of integration of the epidemiological studies into risk assessment by supporting (or identifying the lack of support for) the biological plausibility of the link between the exposure to pesticides affecting the pathway and the AO, intended as the human health outcome. This should be based on the agreement of the current understanding of the AO and the strength of the weight of evidence will define the boundaries of its scientific validity. In developing qualitative AOPs the Panel realized that these can be also used as screening tools.

Quantitative AOPs (intended as supported by quantitative relationship that allow quantitative translation of key event measurement into predicted probability or severity of adverse outcome) can cover any need, including a complete hazard assessment of a chemical, by identifying regulatorily relevant point of departures for reference values; the quantitative AOP can also support the inclusion of chemical specific factors like internal exposure and metabolism. Fully quantitative AOP’s are very data-demanding; thus it is, envisaged that they would be a second step in a regulatory prioritisation process. However, the Panel recognized that moving from qualitative to quantitative AOPs, would potentially represent an important step forward to a more effective use of pathway-based data to support risk assessment and build up a predictive network.

The Panel considered that the use of properly developed AOPs is important to guide on future tailored and tiered testing strategies for hazard identification and characterization and consequently a proposal for an AOP-based IATA framework for identifying the risk of PD was made. The most sensitive, robust, reliable test for a KE can be proposed to be further validated and, if needed, ultimately becoming a part of the OECD testing program for chemicals. This is considered a very important element of integrating non-animal data for regulatory use (NRC 2007). The Panel further concluded that the AOP framework is a powerful tool to support the most appropriate design for in vitro and in vivo studies and increase the sensitivity of methods and experimental designs for capturing and possibly characterising a given hazard. As an AOP is expected to reflect the current knowledge, this would imply that a large number of studies and methods will be included in the description and empirical
support for KE and KE relationship (KER). In this perspective, the quality of the studies and scientific validation of the methods are relevant for regulatory purposes; in the meanwhile, the Panel considered that decision on how to make the best use of the available information should depend on the problem formulation. For the scope of this Scientific Opinion, i.e., to explore the AOP framework as a tool for supporting biological plausibility in relation to epidemiological studies, it is essential that the description of the WoE would be complete and transparent; merging in the WoE of the biological plausibility, essentiality and strength of empirical support in a ranking order of relevance for each KE and KER will foster the decision made. If AOPs are intended to inform IATA or define the optimal study design for hazard identification/characterization, elements of scientific validation and study quality should be taken into consideration.

The Panel identifies AOPs as a critical element to facilitate the move towards a mechanism-based risk assessment instead of the current testing paradigm relying heavily on apical effects observed in animal studies (as recommended by EFSA, as well as by ECHA and OECD). Shifting the risk assessment paradigm and mechanistic understanding would reduce limitations of the animal data in predicting human health effects for the single pesticide, and also support the current efforts on carrying out cumulative risk assessment of pesticide exposure. Regarding the grouping of pesticides for cumulative risk assessment and in particular the refining of groups, the Panel concluded in 2013 that the current read-out from animal studies (apical endpoints) were not tailored for this purpose and recommended that a better mechanistic understanding of toxicity should be achieved, and in particular recommended development of AOPs (EFSA 2013 – SO on dissimilar MoA). In this perspective, AOP networks represent the functional unit of prediction of the AO as AOPs are not triggered in isolation but they rather interact. Indeed, key events and KERs are shared by multiple AOPs (as exemplified in this Opinion by neuroinflammation and neurodegenerative diseases). Rather than looking at hazards in isolation, developing AOPs in a modular approach gradually describes the complexity of potential interactions at cell, tissue, organ, system and organism levels, thus meeting the concept of systems biology. The Panel recommends that AOPs could serve the purpose of building more mechanistically driven cumulative assessment groups; considering this specific mandate, this opinion underlines how biological plausibility can support the link between pesticide exposure and complex human health outcomes (e.g., parkinsonian disorders) by evidencing how multiple MIEs can lead to the same AO. The Panel also appreciated that interactions between AOPs can be easily made visible if AOPs are downloaded in the AOP-Wiki.

In regard to the future process of AOP development, the collaborative AOP-Wiki of the AOP knowledge base (https://aopkb.org) serves as a useful platform and the upload of AOPs into the AOP-Wiki is encouraged by the Panel. The use of the AOP-Wiki would provide multiple benefits in terms of easy approach to AOPs network, transparency, and accurate peer-review will be ensured by the formal review process of the proposed AOPs. This effort is still under development and from the learnings of this mandate future work and improvements could be suggested. Common KEs should be shared only if approved following the peer-review process. Overall, to increase the future regulatory acceptance of AOPs and the key events based testing, more emphasis should be put on the transparent data selection for building AOPs; accordingly, weight of evidence analysis should be conducted and built into the AOP wiki. Collier et al. (2016) have recently suggested a framework which also includes quantitative considerations, especially on the strength of linkages between key events: such considerations would include criteria for soundness, applicability and utility, clarity and completeness, uncertainty and variability and last evaluation and review.

The Panel concluded that:

- The AOP framework is a useful tool for risk assessment to explore if an AO is biologically plausible or not; by means of mechanistically describing apical endpoints, the AOP contributes to the hazard identification and characterization steps in risk assessment. As the AOP framework is chemically agnostic, it will consolidate the chemical specific risk assessment with the aid of, and within, the MOA and/or IATA framework.

- The prototype AOPs developed by the Panel support that pesticides affecting the proposed MIEs and the pathways are risk factors for the development of the diseases i.e. Parkinson’s disease and infant leukaemia, considering risk factor as hazard identification. This conclusion is based on a weight of evidence assessment appropriately characterized by defining questions on biological plausibility and empirical support for the key events relationship and...
on the essentiality of the key events. In addition, inconsistencies and uncertainties were identified and reported for each of the developed AOP.

- The Panel recognised that the systematic literature review and meta-analysis indicated that the plausible involvement of pesticide exposure as a risk factor for the development of Parkinson’s disease and infant leukaemia could by linked to further AOPs development in addition to the ones developed by the Panel. This limitation in the current Scientific Opinion does not weaken the overall conclusion about the plausible involvement of pesticides in the pathogenesis of PD and IFL.

- The Panel concluded that the AOP developed for childhood leukaemia is not bringing a definitive evidence of biological plausibility. However, circumstantial evidences indicate that a hypothetical biological plausibility could exist but can not be formulated with the current available information. These circumstantial evidences are mainly derived by the epidemiological observation that the disease is prevalent following in utero exposure to pesticides and that exploration of tumor related endpoints following in utero exposure has limitations in the standard design of regulatory studies. In addition, the Panel recognise that an animal model recapitulating the disease is not available and this is also weakening the assessment.

- The AOP framework was considered by the Panel as an appropriate tool to understand if chemical hazards related to the relevant human diseases (Parkinson’s disease, infant and childhood leukaemia) can be explored and detected in the standard regulatory studies. Although apical endpoints in the regulatory studies i.e. histological evaluation of the nigrostriatal pathway and neurological examination (for Parkinson diseases), blood analysis, genotoxicity testing, immunological parameters in reproductive studies and cancerogenesis assays (for infant and childhood leukemias), can potentially inform on some KEs or the AOs, the mechanistic understanding of the apical endpoints indicate that the regulatory toxicological studies have limitations because of the study design or because of the sensitivity of the test system. Tailored studies or more sensitive tools should be considered to prove that a chemical has negligible hazard if it is affecting the pathway.

9. **Recommendations**

- Overall, the Panel recommends the AOP conceptual framework to assess the biological plausibility; or lack of biological plausibility, of the association between pesticide exposure and human health effects reported in epidemiological studies by means of including the observed effect in the AO and consequently in the hazard identification process.

- The AOPs should stimulate regulators to ask for the application of additional testing based on the mechanistic understanding of the KEs. Therefore, the Panel recommends that the AOP should be used as a mechanistic tool to support biological plausibility and mechanistic understanding of apical hazards when toxicity studies are considered insufficient, inconclusive or inadequate, but the substance is known to affect the pathway.

- The AOPs should stimulate additional research work in order to provide a more robust quantitative evaluation of the threshold effects for the different KEs, using the same tool compounds used for their development. Quantitative evaluation should foster the regulatory use of the AOP and should include, where possible, a concentration response analysis by means of identification of a non-effect threshold and a minimum threshold effect able to trigger the pathway. A biologically relevant battery of assays preferably based on human cells, able to recapitulate the key events of the AOP and predictive of the concentration of the chemical leading to the AO should be developed.

- The systematic literature review indicated that the plausible involvement of pesticide exposure as a risk factor for the development of Parkinson’s disease and infant/childhood leukaemia could by linked to additional AOPs other than the ones developed by the Panel. The Panel recognizes this limitation of the current Scientific Opinion and recommends that additional AOPs should be developed with the intent of using the AOP to support the biological plausibility of additional MIEs and pathways but also to develop an AOP network to be used as a functional unit for the prediction of the diseases.
The Panel recommends that the AOP network should be considered as a tool for the refinement of the Cumulative Assessment Groups to be used in cumulative risk assessment of pesticides.

The Panel recommends that for compounds affecting the AOPs developed for parkinsonian motor symptoms, the histological evaluation of the nigrostriatal pathway should be performed by means of application of proper stereology protocols and inclusion of special stains in addition to H&E. The Panel also recommends that biomarkers e.g. α-synuclein could be considered to help in the study design i.e. dose selection and length of the treatment, when compounds are known to affect the pathway but the regulatory endpoints are negative.

In addition the Panel recommends using mixed neurons/glia cocultures for in vitro testing.

The Panel proposes neuroinflammation as a key event for the two AOPs developed for the AO “parkinsonian motor symptoms” The Panel does not, however, recommend the use of neuroinflammatory biomarkers for the time being. Although the Panel is supporting their utility and validity from the scientific point of view, they are still too challenging and complex for regulatory uses. However, this does not include the use of immuno-markers for cell phenotyping.

The Panel recognizes the limitations of the standard regulatory studies as evidenced by the AOP developed in this Scientific Opinion and recommends that an AOP-informed IATA should be developed to support the testing strategy. The Panel also recommends that an AOP-informed IATA framework should be developed for the IFL and the CHL and that the testing strategy should be based on non-animal testing as a first approach for new data generation.

The Panel recommends that the standard OECD guidance on histological evaluation of the brain in the 90 days toxicity study (OECD TG 408) and in general in the toxicity studies performed in vivo, should be revised in order to include a more in depth evaluation of brain structures involved in Parkinson disease i.e. the nigrostriatal pathway.

The Panel recommends that genotoxicity assays should consider the sensitive detection of TopoII poisons; in particular, this could be achieved by considering the inclusion of liver HSPCs as target cells in genotoxicity testing. Therefore, sensitive cell systems representing the early-embryo phase for exploring genotoxicity endpoints should be considered.

The Panel recognises that the use of a non-validated cell system needs more scientific work in order to provide robust data on specificity and sensitivity for an appropriate use in risk assessment.

The Panel recommends that the epidemiological studies and meta-analysis should make a distinction between infant and childhood leukaemia which are etiologically and pathologically different entities.

The Panel recommends that tailored and tiered testing strategies should be developed and the assays should be anchored to the KEs identified in the AOPs. Accordingly, the test system should be selected to model the human biology of KEs.

In order to facilitate the regulatory relevance, the AOPs developed for this mandate should be submitted to the AOP-Wiki and undergo the rigorous peer-review by the OECD.

Based on the experience gained in developing AOPs, the Panel recommends that the transparency and weight of evidence in building AOPs should be strengthened in the future. An agreed approach during the process of AOP preparation (assembling evidence) for the evaluation of data quality of individual studies and for aggregating lines of evidence, possibly in a more quantitative and structured way, is recommended. The framework suggested by Collier et al (2016) could serve this purpose. Also, the Panel notes that for the future development of AOPs and AOP-network of the AOP-Wiki, that careful updates of KER of common KE must be implemented.

The Panel recommends to apply in vitro methods as a first approach for gaining mechanistic information to support the biological plausibility linking exposure to pesticide to human health outcomes.
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Appendix A – AOP developed for parkinsonian motor deficit

AOP1: Inhibition of the mitochondrial complex I of nigra-striatal neurons leads to parkinsonian motor deficits

Abstract

This AOP describes the linkage between inhibition of complex I (CI) of the mitochondrial respiratory chain and motor deficit as in parkinsonian disorders. Binding of an inhibitor to complex I has been defined as the molecular initiating event (MIE) that triggers mitochondrial dysfunction, impaired proteostasis, which then cause degeneration of dopaminergic (DA) neurons of the nigro-striatal pathway. Neuroinflammation is triggered early in the neurodegenerative process and exacerbates it significantly. These causatively linked cellular key events result in motor deficit symptoms, typical for parkinsonian disorders, including Parkinson’s disease (PD), described in this AOP as an Adverse Outcome (AO). Since the release of dopamine in the striatum by DA neurons of the Substantia Nigra pars compacta (SNpc) is essential for motor control, the key events refer to these two brain structures. The weight-of-evidence supporting the relationship between the described key events is based mainly on effects observed after an exposure to the chemicals rotenone and 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP), i.e. two well-known inhibitors of complex I.

Data from experiments with these two chemicals reveal a significant response-response concordance between the MIE and AO and within KEs. Also essentiality of the described KEs for this AOP is strong since there is evidence from knock out animal models, engineered cells or replacement therapies that blocking, preventing or attenuating an upstream KE is mitigating the AO. Similarly, there is proved experimental support for the KERs as multiple studies performed with modulating factors that attenuate (particularly with antioxidants) or augment (e.g. overexpression of viral-mutated α-synuclein) a KE up show that such interference leads to an increase or attenuation/prevention of KE down or the AO.

Information from in vitro and in vivo experiments is complemented by human studies in brain tissues from individuals with sporadic Parkinson’s disease (Keeney et al., 2006) to support the pathways of toxicity proposed in this AOP.

This AOP is reported in line with the OECD Guidance Document for developing and assessing AOPs [ENV/JM/MONO(2013)6] and with the supplemented user’s handbook.
1. Molecular Initiating Event (MIE): Binding of an inhibitor to NADH ubiquinone oxidoreductase (complex I)

1.1 How this Key Event works:
Electron transport through the mitochondrial respiratory chain (oxidative phosphorylation) is mediated by five multimeric complexes (I–V) that are embedded in the mitochondrial inner membrane (Fig. 1). NADH-ubiquinone oxidoreductase is the Complex I (CI) of the electron transport chain (ETC). It is a large assembly of proteins that spans the inner mitochondrial membrane. In mammals, it is composed of about 45-47 protein subunits (45 in humans) of which 7 are encoded by the mitochondrial genome (i.e., ND1, ND2, ND3, ND4, ND4L, ND5, and ND6) and the remaining ones by the nuclear genome (Greenamyre, 2001). CI oxidizes NADH elevating the NAD+/NADH ratio by transferring electrons via a flavin mononucleotide (FMN) cofactor and several iron-sulfur centers to ubiquinone (Friedrich et al., 1994) (Fig. 2).
Fig. 2. The electron transport chain in the mitochondrion.

CI (NADH-coenzyme Q reductase or NADH dehydrogenase) accepts electrons from NADH and serves as the link between glycolysis, the citric acid cycle, fatty acid oxidation and the electron transport chain. Complex II also known as succinate-coenzyme Q reductase or succinate dehydrogenase, includes succinate dehydrogenase and serves as a direct link between the citric acid cycle and the electron transport chain. The coenzyme Q reductase or Complex III transfers the electrons from CoQH2 to reduce cytochrome c, which is the substrate for Complex IV (cytochrome c reductase). Complex IV transfers the electrons from cytochrome c to reduce molecular oxygen into water. Finally, this gradient is used by the ATP synthase complex (Complex V) to make ATP via oxidative phosphorylation. mtDNA: mitochondrial DNA; nDNA: nuclear DNA (Friedrich et al 1994).

Binding of an inhibitor to CI inhibits the NADH–ubiquinone oxido-reductase activity, i.e. blocks the electron transfer. Recent studies suggest that a wide variety of CI inhibitors share a common binding domain at or close to the ubiquinone reduction site (Ito et al., 2003). Furthermore, the structural factors required for inhibitory actions have been characterized on the basis of structure-activity relationships (Miyoshi, 1998, Hideto, 1998).

Based on molecular docking simulations, in silico models mimicking the binding of chemicals to the pocket of NADH ubiquinone oxidoreductase have been created according to the crystal structure of mitochondrial CI. To investigate the ability of chemicals to bind to the active pocket, around 100 individual docking simulations have been performed. These confirmed the possible site of interaction between the chemical and the pocket of CI. In particular, Miao YJ and coworkers recently investigated the IC50 values of 24 chemicals (annoonaceous acetogenins) for inhibition of mitochondrial CI (Miao et al., 2014).

Based on their binding sites, CI inhibitors are classified as follows (Degli Esposti, 1998) (Fig. 3):

(i) type A inhibitors are antagonists of fully oxidized ubiquinone binding;
(ii) type B inhibitors displace the partially reduced ubisemiquinone intermediate;
(iii) type C inhibitors are antagonists of the fully reduced ubiquinol product.

The affinity of the different types of CI inhibitors to their diverse CI binding sites is described in the paragraph Evidence for Chemical Initiation of this Molecular Initiating Event (see below) in the context of a specific type of inhibitor.
Fig. 3. Schematic representation of CI and proposed inhibition binding sites by inhibitors of class A, B and C. Nicotinamide adenine dinucleotide (NADH, reduced and NAD, oxidized), flavin mononucleotide (FMN) and Ubiquinone (Q) (Haefeli et al., 2012, Fig. 46).

1.2 How it is measured or detected

Two different types of approaches have been used. The first is to measure binding as such, and the corresponding assays are described below; the second is to infer binding indirectly from assays that quantify e.g. CI activity and to assume that the activity can only be altered upon binding.

The second type of approach is dealt with in the chapter entitled KE1: Inhibition of NADH ubiquinone oxidoreductase (complex I). However, it has to be noted here that indirect assays can lead to wrong conclusions. For instance, some compounds may trigger oxidative stress without actually binding to CI. Such compounds, by triggering the generation of reactive oxygen species (ROS), may damage CI protein components, thus causing a reduction of CI activity.

1.3 Measurement of binding by quantitative autoradiography

To assess binding of an inhibitor at the rotenone binding site of CI in tissues (e.g. in the substantia nigra or in the striatum), the standard approach is to quantify the displacement of a radioactively labelled ligand of this binding site by the toxicant under evaluation. Most commonly, binding of \([^3H]\)-labeled dihydrorotenone (DHR) is measured and compared in control tissue and treated tissue.

Binding of this rotenone-derivative is detected by autoradiography. Unselective binding is determined by measurement of \([^3H]\)-DHR binding in the presence of an excess of unlabeled rotenone. Since a rotenone-derivative is used for the assay, only CI inhibitors that bind to the rotenone-binding site in CI are detected. This was observed for e.g., meperdine, amobarbital, or MPP⁺. This method allows a spatial resolution of CI expression and the mapping of the binding of a competitive inhibitor on CI.

The method can be used for (a) in vitro measurements and for (b) ex vivo measurements:

a) In vitro measurements. Tissues are embedded in a matrix for cutting by a cryostat. The tissue slices are then mounted onto slides. For the binding experiment, they are incubated with the test compound in the presence of labeled \([^3H]\)-DHR. Then the tissue slices are washed and prepared for autoradiographic detection (Greenamyre et al. 1992; Higgins and Greenamyre, 1996).

b) Ex vivo measurements. As rotenone can pass the blood brain barrier, the in vitro method was further extended for in vivo labeling of CI in the brains of living animals, and detection of binding after preparation of the tissue from such animals. Animals are exposed to test compounds and \([^3H]\)-DHR is applied intraventricularly for 2-6 h before the brain is dissected and arranged for the preparation of tissue slices (Talpade et al. 2000). In untreated animals, this method allows a precise spatial...
resolution of the expression pattern of CI. In animals with impaired CI activity, either as a result of CI deficiencies, or upon treatment with CI inhibitors, the assay allows an assessment of the degree of CI inhibition.

1.4 Complex I Enzyme Activity (Colorimetric)

The analysis of mitochondrial OXPHOS CI enzyme activity can be performed using human, rat, mouse and bovine cell and tissue extracts (abcam: http://www.abcam.com/complex-i-enzyme-activity-microplate-assay-kit-colorimetric-ab109721). Capture antibodies specific for CI subunits are pre-coated in the microplate wells. Samples are added to the microplate wells which have been pre-coated with a specific capture antibody. After the target has been immobilized in the well, CI activity is determined by following the oxidation of NADH to NAD+ and the simultaneous reduction of a dye which leads to increased absorbance at OD=450 nm. By analyzing the enzyme's activity in an isolated context, outside of the cell and free from any other variables, an accurate measurement of the enzyme's functional state can be evaluated.

1.5 Evidence supporting taxonomic applicability (tissue type, taxa, life stage, sex)

CI has a highly conserved subunit composition across species, from lower organisms to mammals (Cardol, 2011). Fourteen subunits are considered to be the minimal structural requirement for physiological functionality of the enzyme. These units are well conserved among bacterial (E. coli), human (H. sapiens), and Bovine (B. taurus) (Vogel et al., 2007b; Ferguson, 1994). However, the complete structure of CI is reported to contain between 40 to 46 subunits and the number of subunits differs, depending on the species (Gabaldon 2005; Choi et al., 2008). In vertebrates CI consists of at least 46 subunits (Hassinen, 2007), particularly, in humans 45 subunits have been described (Vogel et al., 2007b). Moreover, enzymatic and immunochemical evidence indicate a high degree of similarity between mammalian and fungal counterparts (Lummen, 1998). Mammalian CI structure and activity have been characterized in detail (Vogel et al., 2007a; Vogel et al., 2007b), referring to different human organs including the brain. There is also a substantial amount of studies describing CI in human muscles, brain, liver, as well as bovine heart (Janssen et al., 2006; Mimaki et al. 2012) (Okun et al., 1999).

1.6 Evidence for Chemical Initiation of this Molecular Initiating Event (MIE)

The most studied examples of chemicals that inhibit CI are: rotenone and 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) (Desplats et al., 2012; Lin et al., 2012; Sava et al., 2007). Both, rotenone (pesticide) and MPP+ (the active metabolite of MPTP) are well known to reproduce the anatomical, neurochemical, behavioural and neuropathological features of PD-like syndrome (Betarbet et al., 2000; Greenamyre et al., 2001). Indeed, overwhelming evidence has accumulated in the existing literature suggesting such a link, and therefore these two inhibitors of CI will be discussed in the context of all the KEs identified in this AOP.

1.6.1. Rotenone affinity to complex I binding sites

Rotenone, a colorless, odorless, crystalline ketonic chemical compound (a flavonoid) naturally occurring in the seeds and stems of several plants, such as the jicama vine plant, and the roots of several members of Fabaceae, is a classical, high affinity and irreversible inhibitor of CI and is typically used to define the specific activity of this enzymatic complex. Rotenone is extremely lipophilic, it crosses biological membrane easily and can get into the brain very rapidly. Dose-dependent relative affinities of rotenone to the inhibitory site of CI is shown in Fig. 4B (for more details Grivennikova et al., 1997).

Most of the studies suggest that hydrophobic inhibitors like rotenone or Piericidin A most likely disrupt the electron transfer between the terminal Fe-S cluster N2 and ubiquinone (Fig. 4A).
**Rotenone structure**

![Rotenone structure](image)

**Fig. Rotenone structure (left); schematic diagram of CI**

**Fig. 4A.** NADH ubiquinone oxidoreductase, illustrating molecular mode of action and binding site of Rotenone (and Rotenone-like compounds) IMS: inter-membrane space (based on Lummen, 1998).

**Fig. 4B.** Dose-dependent relative affinities of rotenone to the inhibitory site of CI (for more detail see Grivennikova et al., 1997).
1.6.2. MPTP affinity to complex I binding sites

MPTP is not directly binding to CI and it is therefore non-toxic to DA neurons. MPTP exerts its toxicity after it is metabolized by mono-amino-oxidase, type B (MAO B) in astrocytes to 1-methyl-4-phenylpyridinium (MPP+). This metabolite binds to CI, and is toxic. MPP+ is a good substrate for dopamine transporters (DAT), expressed selectively by DA neurons (Greenamyre et al., 2001). Due to both a positive charge and an amphoteric character, MPP+ specifically accumulates in mitochondria, where despite a lower affinity to the binding site of CI than rotenone, it reaches high enough intra-mitochondrial concentrations to inhibit CI activity (Ramsay et al., 1991). The binding affinity of MPP+ is low (mM range), and it can be totally reversed by washing out. However, prolonged treatment results in a severe, progressive and irreversible inhibition of CI, most likely by indirect mechanisms involving oxidative damage (Cleeter et al., 1992). Competitive binding experiments with rotenone and MPP+ suggest that the two compounds bind to the same site of the CI (Ramsay et al., 1991).

1.6.3. General characteristics of other complex I inhibitors

Besides rotenone, there is a variety of CI inhibitors, both naturally occurring, such as Piericidin A (from Streptomyces mobaraensis), acetogenins (from various Annonaceae species), as well as their derivatives, and synthetically manufactured compounds, like pyridaben and various piperazin derivatives (Ichimaru et al., 2008). They have been used to probe the catalytic activity of CI especially in order to clarify its ubiquinone binding site and indeed, most of these compounds inhibit the electron transfer step from the Fe-S clusters to ubiquinone (Friedrich et al., 1994).

Therefore, classification of CI inhibitors is based on their types of action. Type A inhibitors, like piericidin A, 2-decyl-4-quinazolinyl amine (DQA), annonin VI and roliniastatin-1 and -2, are considered to be antagonists of the ubiquinone substrate. For piericidin A, it has been shown that it inhibits NADH:Q2 activity in a partially competitive manner. Contrary to type A, type B inhibitors, like the commonly used rotenone, have hydrogen-bonding acceptors only in the cyclic head of the molecule and are non-competitive towards ubiquinone, but are believed to displace the semiquinone intermediate during the catalysis (Fig. 2). Finally, inhibitors classified as type C, like stigmatellin and capsaicin, form a third group of hydrophobic CI inhibitors that are believed to act as antagonists of reduced ubiquinone (Degli Esposti 1998, Friedrich et al. 1994, Haefeli 2012) (Fig. 2).

Competition studies with representatives of all three different types of inhibitors revealed that type A and B, and type B and C, but not type A and C, compete with each other for binding. This led to a suggestion that all CI inhibitors acting at the ubiquinone binding pocket share a common binding domain with partially overlapping sites (Okun et al. 1999).

Some inhibitors bind to the outside of the ubiquinone reduction site and do not fit the preceding classification. Examples of such compounds are ADP-ribose, which competes for substrate binding at the NADH site (Zharova and Vinogradov, 1997), and diphenyleneiodonium (DPI) that covalently binds to reduced flavin mononucleotide (FMN) in the hydrophilic part of the enzyme, blocking the electron transfer to the Fe-S clusters (Majander et al., 1994).

There are also new, commercially available insecticides/acaricides with potential to inhibit mitochondrial respiration, such as benzimidazole, bullatacin, 6-chlorobenzothiadiazole, cyhalothrin, Fenazaquin Fenpyroximate, Hoe 110779, Pyridaben, Pyrimidifen, Sandoz 547A, Tebufenpyrad and Thiangazole (Greenamyre et al., 2001). It is clear that they are capable of inhibiting the mammalian CI by binding to and blocking ubiquinone-dependent NADH oxidation with high efficacy (Lummen, 1998).
References


2. KE1: Inhibition of NADH ubiquinone oxidoreductase (complex I)

2.1 How this Key Event works

Under physiological conditions complex I (CI) couples the oxidation of NADH to NAD\(^+\) by reducing flavin mononucleotide (FMN) to FMNH\(_2\). FMNH\(_2\) is then oxidized through a semiquinone intermediate. Each electron moves from the FMNH\(_2\) to Fe-S clusters, and from the Fe-S clusters to ubiquinone (Q).

Transfer of the first electron results in the formation of the free-radical (semiquinone) form of Q, and transfer of the second electron reduces the semiquinone form to the ubiquinol form (CoQH\(_2\)). Altogether, four protons are translocated from the mitochondrial matrix to the inter-membrane space for each molecule of NADH oxidized at CI. This leads to the establishment of the electrochemical potential difference (proton-motive force) that may be used to produce ATP (Garrett and Grisham, 2010).

Binding of an inhibitor attenuates or completely blocks the activity of CI, i.e. the oxidation of NADH is impaired and protons are not moved. This causes two major consequences: first, electrons are channelled toward oxygen instead Q. This impairs normal oxygen reduction into water at complex IV and leads to the formation of the ROS superoxide at other sites of the respiratory chain. Superoxide may cause damage of proteins, lipid and DNA of the cell, or damage components of the mitochondria after transformation into e.g. hydrogen peroxide. These processes result in mitochondrial dysfunction (Voet and Voet, 2008). The second consequence is the increase of the NADH/NAD\(^+\) ratio in mitochondria. This affects the function of key dehydrogenase enzymes in the citric acid cycle and can lead to its block, resulting in an inhibition of mitochondrial ATP production and mitochondrial respiration.

The functional consequences of CI inhibition have been titrated in a time- and dose-dependent manner (Barrientos and Moraes, 1999), with mitochondrial dysfunction measured by a range of different assays (Barrientos and Moraes, 1999; Greenamyre et al., 2001). These included quantification of ROS derived from mitochondria, and of cellular respiration (see KE2: Mitochondrial dysfunction).

2.2 How it is measured or detected

As CI has an enzymatic function as such, but also contributes to the overall function of oxidative phosphorylation, there are two fundamental approaches to assess CI inhibition. The first approach measures the enzymatic activity of the complex itself; the second one assesses the overall activity of oxidative phosphorylation of entire mitochondria, and indirectly infers from this a potential dysfunction of CI.

2.3 Direct detection of complex I activity

This type of assay is always performed in homogenates of cells or tissues, and requires at least a partial purification of mitochondria or respiratory chain components. In order to focus on CI activity, the activities of Complexes III (e.g. antimycin A) and complex IV (e.g. cyanide) need to be blocked by pharmacological inhibitors in these setups.

2.3.1 Forward Electron Transfer

Submitochondrial particles or intact isolated mitochondria are incubated with NADH as electron donor and with an electron acceptor to measure the flow of electrons from NADH, through CI to the acceptor. As readout, either the consumption of NADH or the reduction of the electron acceptor is followed photometrically or fluorometrically (Lenaz et al. 2004; Spinazzi et al. 2012; Long et al. 2009; Kirby et al. 2007). The physiological electron acceptor of CI is Coenzyme Q10 (CoQ10). Due to its hydrophobicity, it is not suitable for use in an experimental in vitro setup. Short-chain analogs of CoQ10, such as CoQ1 or decylubiquinone (DB) with a 10 carbon-atom linear saturated side chain are hence applied as alternatives. With these non-physiological electron acceptors, it is important to consider that the activity of CI can easily be underestimated. As water-soluble electron acceptors, either ferricyanide or 2,6-dichlorophenoldiphosphophenol (DCIP) are used. However the reduction of such compounds is not strictly coupled to the transduction of energy. To identify the portion of rotenone-inhibitable CI activity, all samples investigated are assayed in parallel following treatment with...
rotenone. In contrast to the autoradiography assays, direct CI activity detection allows the identification also of CI inhibitors that bind to sites of CI different from the rotenone binding site.

### 2.3.2 Reverse Electron Transfer

An alternative setup for the direct measurement of CI activity with minimal interference by the activities of complex III and complex IV make use of the observation of a general reversibility of oxidative phosphorylation and electron flow across the mitochondrial respiratory chain (Ernster et al. 1967). With this method, electrons enter the respiratory chain via complex II. Based on the reverse flux, this method allows the complete circumvention of complexes III and IV. As electron donor, succinate is applied, together with NAD$^+$ as electron acceptor. Formation of NADH from NAD$^+$ can be determined photometrically. The succinate-linked NAD$^+$ reduction can be performed either with intact isolated mitochondria or with submitochondrial particles. For the direct assessment of CI activity, submitochondrial particles are used. For assays with intact mitochondria, the succinate-linked reduction of NAD$^+$ is performed in the presence of ATP as energy source. Potassium cyanide (KCN) is added for inhibition of forward electron transport towards complex IV.

### 2.3.3 Complex I activity dipstick assay

To assess CI activity and its inhibition in cell or tissue homogenates without interference by other components of the respiratory chain, CI-selective antibodies attached to a matrix (e.g. multiwell plates) are used (Willis et al., 2009). Homogenized tissue can directly be added for capturing of CI, the unbound supernatant is washed away and leaves a complex of the antibody and mitochondrial CI.

For activity determination, NADH as electron donor and nitroblue tetrazolium (NBT) as acceptor are added. Reduced NBT forms a colored precipitate, its signal intensity is proportional to the amount of CI bound to the antibody. CI inhibitors can directly be added for an assessment of their inhibitory potential. This method, when applied in e.g. 96-well or 384-well plates, allows screening of large sets of potential CI inhibitors without any interference by other elements of the mitochondrial respiratory chain.

### 2.4 Indirect measurements of complex I activity

Such assays mostly require / allow the use of live cells.

### 2.4.1 Oxygen consumption

Electrons, fed into the mitochondrial respiratory chain either by CI or complex II, ultimately reduce molecular oxygen to water at complex IV. In a closed system, this consumption of oxygen leads to a drop of the overall O$_2$ concentration, and this can serve as parameter for mitochondrial respiratory activity. Measurements are traditionally done with a Clark electrode, or with more sophisticated optical methods. At the cathode of a Clark electrode, oxygen is electrolytically reduced, which initiates a current in the electrode, causing a potential difference that is ultimately recorded. Clark electrodes however have the disadvantage that oxygen is consumed. Furthermore, interferences with nitrogen oxides, ozone, or chlorine are observed (Stetter et al., 2008). To circumvent these limitations, optical sensors have been developed that have the advantage that no oxygen is consumed, combined with a high accuracy and reversibility. Optical oxygen sensors work according to the principle of dynamic fluorescence quenching. The response of the respective fluorescence dye is proportional to the amount of oxygen in the sample investigated (Wang and Wolfbeis, 2014). In a model of isolated mitochondria in the absence of complex II substrates, oxygen consumption can serve as surrogate readout for the assessment of the degree of CI inhibition. It is however essential to realize that also complex III and complex IV activities are involved and their inhibition also results in a decline in O$_2$ consumption. In addition to that, CI inhibitors can lead to a one-electron reduction of molecular oxygen at the site of CI to yield superoxide. The amount of superoxide formed hence contributes to the consumption of oxygen, but this must not be interpreted as oxygen consumption as a result of controlled and coupled electron flux through the complexes of the mitochondrial respiratory chain. A modern convenient method to measure oxygen consumption is provided by the Seahorse technology of extracellular flux (XF) analysis, in which cells are kept in a very small volume, so that changes of oxygen levels can be detected very sensitively by an oxygen sensor. To allow manipulation of the mitochondria in cells, the cell membrane can be permeabilized with saponin (SAP), digitonin (DIG) or recombinant perfringolysin O (rPFO) (XF-plasma membrane permeabilizer (PMP) reagent), to allow
addition of specific substrates to measure activity of different respiratory chain complexes, including CI. (Salabei et al., 2014).

2.4.2 Intracellular ATP levels

Intracellular ATP levels originate both from mitochondria and from glycolysis. If glycolytic ATP production is impaired or inhibited, the cellular production of ATP is a measure of mitochondrial function. If it is assumed that the ATP consumption remains constant, then the steady state ATP levels can serve as indirect readout for mitochondrial activity, and the latter depends on the functioning of CI. Inhibitors of CI reduce cellular ATP levels, but it has to be remembered that intracellular ATP levels are also affected by inhibitors of other parts of the respiratory chain, of the citric acid cycle or of the transport of energy substrates. For a proper interpretation of assay results, it has to be ascertained in each particular test system, that ATP production from other sources is excluded and that the cellular ATP consumption remains constant. ATP levels can be easily measured from lysates of in vitro cell cultures or from tissues by a luminometric luciferase/luciferin assay. The amount of light emitted is proportional to the amount of ATP in the sample (Nguyen et al. 1988) (Leist, 1997).

2.4.3 Other approaches

As mitochondrial activity is coupled to many cellular functions, there is a multitude of other indirect assays that are sensitive to inhibitors of CI. Some of these tests may indeed be very sensitive, while they have a low specificity. Thus, their application requires usually a good control of the experimental system and care with the interpretation of the data. One exemplary approach is the measurement of NADH/NAD\(^+\) ratios in mitochondria by imaging methods. This provides resolution on the level of individual mitochondria within a living cell (van Vliet et al., 2014).

2.5 Evidence Supporting Taxonomic Applicability

The CI is well-conserved across species from lower organisms to mammals. The central subunits of CI harboring the bioenergetic core functions are conserved from bacteria to humans. CI from bacteria and from mitochondria of \textit{Yarrowia lipolytica}, a yeast genetic model for the study of eukaryotic CI (Kerscher et al., 2002) was analyzed by x-ray crystallography (Zickermann et al., 2015, Hofhaus et al., 1991; Baradaran et al., 2013). The CI of the mitochondria of eukaryotes and in the plasma membranes of purple photosynthetic bacteria are closely related to respiratory bacteria and the close homology of sequences, function, and prosthetic groups shows a common ancestry (Friedrich et al., 1995).
References


3. KE2: Mitochondrial dysfunction (ENV/JM/WRPR(2016)34; 2016)

3.1 How this Key Event works

Mitochondria play a pivotal role in cell survival and cell death because they are regulators of both energy metabolism and apoptotic/necrotic pathways (Fiskum, 2000; Wieloch, 2001; Friberg and Wieloch, 2002). The production of ATP via oxidative phosphorylation is a vital mitochondrial function (Kann and Kovács, 2007; Nunnari and Suomalainen, 2012). The ATP is continuously required for signalling processes (e.g., Ca$^{2+}$ signalling), maintenance of ionic gradients across membranes, and biosynthetic processes (e.g., protein synthesis, heme synthesis or lipid and phospholipid metabolism) (Kang and Pervaiz, 2012; Green, 1998; Hajnóczky et al., 2006; McBride et al., 2006). Inhibition of mitochondrial respiration contributes to various cellular stress responses, such as deregulation of cellular Ca$^{2+}$ homeostasis (Graier et al., 2007) and ROS production (Nunnari and Suomalainen, 2012; reviewed Mei et al., 2013).

It is well established in the existing literature that mitochondrial dysfunction may result in: (a) an increased ROS production and a decreased ATP level, (b) the loss of mitochondrial protein import and protein biosynthesis, (c) the reduced activities of enzymes of the mitochondrial respiratory chain and the Krebs cycle, (d) the loss of the mitochondrial membrane potential, (e) the loss of mitochondrial motility, causing a failure to re-localize to the sites with increased energy demands, (f) the destruction of the mitochondrial network, (g) increased mitochondrial Ca$^{2+}$ uptake, causing Ca$^{2+}$ overload (reviewed in Lin and Beal, 2006; Graier et al., 2007), (h) the rupture of the mitochondrial inner and outer membranes, leading to the release of mitochondrial pro-death factors, including cytochrome c (Cyt. c), apoptosis-inducing factor, or endonuclease G (Braun, 2012; Martin, 2011; Correia et al., 2012; Cozzolino et al., 2013), which eventually leads to apoptotic, necrotic or autophagic cell death (Wang and Qin, 2010). Due to their structural and functional complexity, mitochondria present multiple targets for various compounds.

3.2 How it is measured or detected

Mitochondrial dysfunction can be detected using isolated mitochondria, intact cells or cells in culture as well as in vivo studies. Such assessments can be performed with a large range of methods (revised by Brand and Nicholls, 2011) for which some important examples are given. All approaches to assess mitochondrial dysfunction fall into two main categories: the first approach assesses the consequences of a loss-of-function, i.e., impaired functioning of the respiratory chain and processes linked to it. Some assays to assess this have been described for KE1, with the limitation that they are not specific for CI. In the context of overall mitochondrial dysfunction, the same assays provide useful information, when performed under slightly different assay conditions (e.g., without addition of complex III and IV inhibitors). The second approach assesses a ‘non-desirable gain-of-function’, i.e. processes that are usually only present to a very small degree in healthy cells, and that are triggered in a cell upon mitochondria failure.

3.2.1 Mitochondrial dysfunction assays assessing a loss-of function

3.2.1.1. Cellular oxygen consumption

See KE1 for details regarding oxygen consumption assays. The oxygen consumption parameter can be combined with other endpoints to derive more specific information on the efficacy of mitochondrial function. One approach measures the ADP-to-O ratio (the number of ADP molecules phosphorylated per oxygen atom reduced (Hinkle, 1995 and Hafner et al., 1990). The related Phosphate/Oxygen (P/O) ratio is calculated from the amount of ADP added, divided by the amount of O consumed while phosphorylating the added ADP (Ciapaite et al., 2005).

3.2.1.2. Mitochondrial membrane potential ($\Delta \psi_m$)

The mitochondrial membrane potential ($\Delta \psi_m$) is the electric potential difference across the inner mitochondrial membrane. It requires a functioning respiratory chain in the absence of mechanisms that dissipate the proton gradient without coupling it to ATP production. The classical, and still most quantitative method uses a tetraphenylphosphonium ion (TPP$^+$)-sensitive electrode on suspensions of isolated mitochondria.
The Δψm can also be measured in live cells by fluorimetric methods. These are based on dyes which accumulate in mitochondria because of Δψm. Frequently used are tetramethylrhodamineethylester (TMRE), tetramethylrhodamine, methyl ester (TMRM) (Petronilli et al., 1999) or 5,5′,6,6′-tetrachloro-1,1′,3,3′-tetraethylbenzimidazole carbocyanide iodide (JC-1). In particular, mitochondria with intact membrane potential concentrate JC-1, so that it forms red fluorescent aggregates, whereas de-energized mitochondria cannot concentrate JC-1 and the dilute dye fluoresces green (Barrientos et al., 1999). Assays using TMRE or TMRM measure only at one wavelength (red fluorescence), and depending on the assay setup, de-energized mitochondria become either less fluorescent (loss of the dye) or more fluorescent (attenuated dye quenching).

**3.2.1.3 Enzymatic activity of the electron transport system (ETS)**

Determination of ETS activity can be determined following Owens and King's assay (1975). The technique is based on a cell-free homogenate that is incubated with NADH to saturate the mitochondrial ETS and an artificial electron acceptor [1 - (4 -iodophenyl) -3 - (4 -nitrophenyl) -5- phenylte trazolium chloride (INT)] to register the electron transmission rate. The oxygen consumption rate is calculated from the molar production rate of INT-formazan which is determined spectrophotometrically (Cammen et al., 1990).

**3.2.1.4. ATP content**

For the evaluation of ATP levels, various commercially-available ATP assay kits are offered (e.g. Sigma, http://www.abcam.com/atp-assay-kit-colorimetricfluorometric-ab83355.html), based on luciferin and luciferase activity. For isolated mitochondria various methods are available to continuously measure ATP with electrodes (Llaudet et al., 2005), with lumimometric methods, or for obtaining more information on different nucleotide phosphate pools (e.g. Ciapaite et al., 2005).

**3.2.2 Mitochondrial dysfunction assays assessing a gain-of function**

**3.2.2.1 Mitochondrial permeability transition pore (PTP) opening**

The opening of the PTP leads to the permeabilization of mitochondrial membranes (Lemasters et al., 2009; Fiskum, 2000), so that different compounds and cellular constituents can change intracellular localization. This can be measured by assessment of the translocation of cytochrome c, adenylate kinase or the apoptosis-inducing factor (AIF) from mitochondria to the cytosol or nucleus. The translocation can be assessed biochemically in cell fractions, by imaging approaches in fixed cells or tissues, or by live-cell imaging of GFP fusion proteins (Single et al., 1998; Modjtahedi et al., 2006). An alternative approach is to measure the accessibility of cobalt to the mitochondrial matrix in a calcein fluorescence quenching assay in live permeabilized cells (Petronilli et al., 1999).

**3.2.2.2 mtDNA damage as a biomarker of mitochondrial dysfunction**

Various quantitative polymerase chain reaction (QPCR)-based assays have been developed to detect changes of DNA structure and sequence in the mitochondrial genome (mtDNA). mtDNA damage can be detected in blood after low-level rotenone exposure, and the damage persists even after CI activity has returned to normal. With a more sustained rotenone exposure, mtDNA damage can be also detected in skeletal muscle. These data support the idea that mtDNA damage in peripheral tissues in the rotenone model may provide a biomarker of past or ongoing mitochondrial toxin exposure (Sanders et al., 2014).

**3.2.3. Generation of ROS and resultant oxidative stress**

**3.2.3.1 General approach**

Electrons from the mitochondrial ETS may be transferred ‘erroneously’ to molecular oxygen to form superoxide anions. This type of side reaction can be strongly enhanced upon mitochondrial damage. As superoxide may form hydrogen peroxide, hydroxyl radicals or other ROS, a large number of direct ROS assays and assays assessing the effects of ROS (i.e., indirect ROS assays) are available. Direct assays are based on the chemical modification of fluorescent or luminescent reporters by ROS species. Indirect assays assess cellular metabolites, the concentration of which is changed in the presence of...
ROS (e.g., glutathione, malonaldehyde, isoprostanes, etc.). In living animals, the effects of oxidative stress can be detected by analysis of specific biomarkers in the blood or urine.

### 3.2.3.2. Measurement of the cellular glutathione (GSH) status

GSH is regenerated from its oxidized form (GSSG) by the action of a NADPH-dependent reductase (GSH + NADPH + H+ → 2 GSH + NADP+). The ratio of GSH/GSSG is therefore a good indicator for the cellular NADP+/NADPH ratio (i.e. the redox potential). GSH and GSSH levels can be determined by HPLC, capillary electrophoresis, biochemically with DTNB (Ellman’s reagent, 5,5’-dithio-bis-[2-nitrobenzoic acid]) or by means of luminescence-based assays (for example, GSH-Glo™ Glutathione Assay). As excess GSSG is rapidly exported from most cells to maintain a constant GSH/GSSG ratio, a reduction of total glutathione levels is often considered a good surrogate measure for oxidative stress.

### 3.2.3.3. Quantification of lipid peroxidation

Measurement of lipid peroxidation has historically relied on the detection of thiobarbituric acid (TBA)-reactive compounds, such as malondialdehyde generated from the decomposition of cellular membrane lipid under oxidative stress (Pryor et al., 1976). This method is quite sensitive, but not highly specific. A number of commercial assay kits are available for this assay using absorbance or fluorescence detection technologies. The formation of F2-like prostanoid derivatives of arachidonic acid, termed F2-isoprostanes (IsoPs) has been shown to be more specific for lipid peroxidation. A number of commercial ELISA kits have been developed for IsoPs, but interfering agents in samples require partial purification before analysis. Alternatively, gas chromatography–mass spectrometry (GC-MS) may be used as a robust, specific and sensitive method.

### 3.2.3.4. Detection of superoxide (O$_2^-$) production

Generation of superoxide by inhibition of CI and the methods for its detection are described by Grivennikova and Vinogradov (2006). A range of different methods is also described by BioTek (http://www.biotek.com/resources/articles/reactive-oxygen-species.html). The reduction of ferricytochrome c to ferrocytochrome c may be used to assess the rate of superoxide formation (McCord and Fidovich, 1968). Like in other superoxide assays, specificity can only be obtained by measurements in the absence and presence of superoxide dismutase. Chemiluminescent reactions have been used for their increased sensitivity. The most widely used chemiluminescent substrate is lucigenin. Coelenterazine has also been used as a chemiluminescent substrate. Hydroxyamine dyes are fluorogenic sensors for superoxide and hydroxyl radical, and they become membrane impermeable after oxidation (trapping at the site of formation). The best characterized of these probes are Hydro-Cy3 and Hydro-Cy5. Generation of superoxide in mitochondria can be visualized using fluorescence microscopy with MitoSOX™ Red reagent (Life Technologies). MitoSOX™ Red reagent is a cationic derivative of dihydroethidium that permeates live cells and accumulates in mitochondria.

### 3.2.3.5. Detection of hydrogen peroxide (H$_2$O$_2$) production

There are a number of fluorogenic substrates, which serve as hydrogen donors that have been used in conjunction with horseradish peroxidase (HRP) enzyme to produce intensely fluorescent products in the presence of hydrogen peroxide (Zhou et al., 1997; Ruch et al., 1983). The more commonly used substrates include diacetyl dichloro-fluorescein, homovanillic acid, and Ampliplex® Red (https://www.thermofisher.com/order/catalog/product/A22188). In these assays, increasing amounts of H$_2$O$_2$ leads to increasing amounts of fluorescent product (Tarpley et al., 2004).

### 3.3 Evidence Supporting Taxonomic Applicability

Mitochondrial dysfunction is a universal event occurring in cells of any species (Farooqui and Farooqui, 2012). Many invertebrate species (e.g., *D. melanogaster* and *C. elegans*) are considered as potential models to study mitochondrial functionality. New data on marine invertebrates, such as molluscs and crustaceans and non-Drosophila species, are emerging (Martinez-Cruz et al., 2012). Mitochondrial dysfunction can be measured in animal models used for toxicity testing (Winklhofer and Haass, 2010; Waerzeggers et al. 2010) as well as in humans (Winklhofer and Haass, 2010).
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Promega GSH-Glo Glutathione Assay Technical Bulletin, TB369, Promega Corporation, Madison, WI.


4. KE3: Impaired proteostasis

4.1 How this key Event works

The concept of proteostasis refers to the homeostasis of proteins in space and time, i.e. the correct balance between protein synthesis, modification, transport and degradation. Disturbance of proteostasis results in pathological changes either by loss of function events (lack of a pivotal protein/protein function) or by a gain of undesired functions (aggregation of a protein leading to the formation of inclusions and new structures in cells and disturbing turnover of many unrelated proteins).

Proteostasis regulation is the main defence mechanism against toxic proteins, whose accumulation could greatly compromise normal cellular function and viability. Therefore, the chaperone and degradation systems assuring the removal of misfolded and aggregated proteins, as well as damaged, dysfunctional cellular organelles (e.g., defective mitochondria) play a key role in cellular homeostasis (Lee et al., 2012).

The two major degradation systems are the ubiquitin–proteasome system (UPS) and the autophagy–lysosome pathway (ALP) (Korolchuk et al., 2010; Kroemer et al., 2010; Ravikumar et al., 2010). The UPS works through the attachment of multiple ubiquitin molecules to a protein substrate, followed by the subsequent degradation of the tagged polyubiquitinated protein by the proteasome (Ciechanover, 1998; Ciechanover and Brundin, 2003). A compromised function of the UPS leads to the accumulation of ubiquitylated proteins, such as α-synuclein, (Ii et al. 1997; Spillantini et al. 1997; Sulzer and Zecca 2000). The accumulation of polyubiquitinated proteins, as a consequence of a dysfunctional proteasome activity, is observed in some pathologies, and experimental inhibition of the proteasome has been shown to trigger parkinsonian neurodegeneration (McNaught and Jenner 2001; Hardy et al., 2001).

ALP involves the engulfment of cytoplasmic materials into autophagosomes, which are degraded by lysosomal enzymes after fusion of autophagosomes with lysosomes (Kuma et al., 2004) or direct import of proteins into lysosomes (Cuervo, 2004; Mizushima et al., 2008). Autophagy also plays an essential role for the removal of damaged organelles, such as mitochondria. Both, excessive autophagy or reduced autophagic flux can compromise cell survival (Rothermel and Hill, 2007), and several genetic forms of PD are linked to the autophagy-related genes Pink1, Parkin or Uchl1.

Autophagy enables cell survival during mitochondrial stress by clearing the damaged organelles (Lee et al., 2012).

One of the main aggregated proteins found to accumulate in nigrostriatal cells during Parkinson's disease is α-synuclein. Aggregation of α-synuclein can obstruct normal cellular transport, leading to impaired intracellular trafficking and/or trapping of cellular organelles in inappropriate locations, this resulting in synaptic and cell dysfunctions (Bartels et al., 2011) (Bellucci A., et al., 2012; Cookson MR., 2005; Games D., et al., 2013; Hunn BH., et al., 2015).

Importantly, accumulation of α-synuclein affects mitochondrial trafficking. The polarity and correct function of different types of cells depend on an efficient transport of mitochondria to areas of high energy consumption (Sheng, 2014). Therefore, the correct distribution of mitochondria to various parts of a cell is essential to preserve cell function (Schwarz, 2013; Zhu et al., 2012).

4.2 How it is measured or detected

4.2.1 Evaluation of UPS function

4.2.1.1 General turnover assays

Quantitative evaluation can be based on the detection of increased ubiquitin or ubiquinated proteins, as well as proteasomal subunits, either by immunocyto/histochemistry or by western blotting (Rideout et al., 2001; Ortega and Lucas, 2014). UPS activity can be continuously monitored by quantitating (by mean of flow cytometry or microscopy) the level of e.g. EGFP-degron fusion proteins that are selectively degraded by the proteasome (Bence et al., 2001).
4.2.1.2 Proteasome activity assay

Various fluorogenic substrates (e.g., Suc-Leu-Leu-Val-Tyr-AMC for the chymotrypsin-like activity) can be used for the determination of proteasomal activity in in vivo or in vitro applications. These substrates may be applied to tissue or cell homogenates, but specific measurements require partial purification of the proteasome (Kisselev and Goldberg, 2005).

4.2.1.3 Detection of α-synuclein (AS) aggregates

The most common methods to detect AS aggregates use immunostaining for AS (in cells or in tissues). In cell culture, AS may also be epitope-tagged or coupled to GFP to allow an indirect detection. The detection of small, not microscopically-visible AS aggregates is indicative of protease-resistance. Tissue slices may be exposed to proteases before immunostaining for AS. Alternatively, small or large aggregates may be biochemically enriched by differential centrifugation and proteolytic treatment, and then analyzed, e.g., by western blot, mass spectrometry or ELISA-like immunoquantification.

4.2.2. Evaluation of ALP function

4.2.2.1 Quantification of lysosomes or autophagosomes

Disturbances of ALP often result in counter-regulations that can be visualized by staining of lysosomes or parts of the autophagy system. Several weakly basic dyes can be used to stain acidic organelles (lysosomes) in live cells. For example, the dye LysoTracker Red stains lysosomes and can be used to monitor autophagy (Klionsky et al., 2007; Klionsky et al., 2008). The autofluorescent drug monodansylcadaverine (MDC) has also been used as autophago-lysosome marker (Munafó and Colombo, 2002). A convenient way to stain lysosomes in tissue or fixed cells is the use of antibodies against the Lysosomal- Associated Membrane Protein 1 (LAMP-1) (Rajapakse et al., 2015) or against cathepsins (Foghgaard et al., 2001).

For qualitative or semiquantitative estimates of lysosomes and related organelles, transmission electron microscopy has been frequently used (Barth et al., 2010).

4.2.2.2 Monitoring of autophagy-related molecules

The amount and the localization of autophagy-related proteins can change during disturbance of the ALP. Especially in cell culture, but also in transgenic mice, various techniques have been used to monitor autophagy by mean of fluorescence-tags or other substrates, e.g., ATG, autophagy-related protein or autophagy substrates, to monitor their fate in cells and thus provide information on disturbed ALP, or the over-expression of GFP-LC3, in which GFP (green fluorescent protein) is expressed as a fusion protein at the amino terminus of LC3 (microtubule-associated protein 1A/1B-light chain 3), which is the a mammalian homologue of *S. cerevisiae* ATG8 (Kadowaki and Karim, 2009).

4.2.2.3 Monitoring autophagic flux

The lysosomal degradation of the autophagic cargo constitutes the autophagic flux, which can be measured by assessing the rate of turnover of long-lived proteins that are normally turned over by autophagy (Bauvy et al., 2009) This is performed by labelling intracellular proteins with either $[^{14}C]$-leucine or $[^{14}C]$-valine, followed by a long culture period in standard medium. The release of radioactive leucin or valin into the culture medium corresponds to the protein degradation rate in cells, and it may be measured by liquid scintillation counting.

4.2.2.4 Monitoring the conversion of LC3-I to LC3-II

The progression of autophagy (autophagic flux) can be studied by the conversion of LC3-I into LC3-II (i.e. a post-translational modification specific for autophagy) by mean of Western blot analysis. The amount of LC3-II correlates with the number of autophagosomes. Conversion of LC3 can be used to examine autophagic activity in the presence or absence of lysosomal activity (Klionsky et al., 2007; Klionsky et al., 2008). The technology can also be used in vivo, e.g. by the use of transgenic mice that overexpress GFP–LC3 (Kuma et al., 2004).
4.2.2.5. Evaluation of intracellular transport of mitochondria and other organelles

A range of technologies has been used to visualize mitochondrial dynamics in live cells (Jakobs, 2006; Grafstein and Forman, 1980). They usually employ a combination of mitochondrial labelling with fluorescent dyes (e.g. DiOC₆ (3, 3′-Dihexyloxacarbocyanine iodide), JC-1 (5,5′,6,6′-Tetrachloro-1,1′,3,3′ tetraethylbenzimidazolylcarbo-cyanine iodide), MitoTracker, MitoFluor probes, etc.), followed by video- or confocal microscopy for live cell imaging (Schwarz, 2013; Pool et al., 2006). Most frequently, mitochondrial mobility is observed along neurites, and measurable endpoints may be mitochondrial speed and direction with regard to the cell soma (Schildknecht et al. 2013). Additionally, also mitochondrial fusion and fission have been monitored by such methods (Exner et al., 2012). The transport of other organelles along neurites may be monitored using similar methods, and the microtubule structures that serve as transport scaffold may be co-stained.

4.3 Evidence supporting taxonomic applicability

The ubiquitin proteasome system is highly conserved in eukaryotes, from yeast to human. Ubiquitin is a small (8.5 kDa) regulatory protein that has been found in almost all tissues of eukaryotic organisms. For instance, drosophila has been used as PD model to study the role of ubiquitin in α-synuclein induced-toxicity (Lee et al., 2009). Human and yeast ubiquitin share 96% sequence identity. Neither ubiquitin nor the ubiquitination machinery is known to exist in prokaryotes.

Autophagy is ubiquitous in eukaryotic cells and is the major mechanism involved in the clearance of oxidatively or otherwise damaged/worn-out macromolecules and organelles (Esteves et al., 2011). Due to the high degree of conservation, most of the knowledge on autophagy proteins in vertebrates is derived from studies in yeast (Klionsky et al., 2007). Autophagy is seen in all eukaryotic systems, including fungi, plants, slime mold, nematodes, fruit flies and insects, rodents (i.e., laboratory mice and rats), and humans. It is a fundamental and phylogenetically conserved self-degradation process that is characterized by the formation of double-layered vesicles (autophagosomes) around intracellular cargo for delivery to lysosomes and proteolytic degradation.
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In neuronal processes, called Lewy neurites, as well as by neurons showing a more clinical signs, 60% of SNpc neurons are lost, a prerequisite for the maintenance of the physiological function (Fujita et al. 2014). The nigrostriatal system is anatomically located in the basal ganglia circuit which comprises the motor system structures caudate nucleus, putamen, globus pallidum and substantia nigra. The caudate nucleus and the putamen are collectively called striatum (David Robinson in: Neurobiology, Springer edition, 1997). The system plays a unique integrative role in the control of movement as part of a system called the “basal ganglia motor loop”. This anatomical loop includes structures in the thalamus, motor and somatosensory cortex and wide regions of surrounding cortex. Neurons of the SN produce dopamine (DA) and project to the striatum. They give dopaminergic excitatory (D1 receptors) and inhibitory (D2 receptors) inputs to striatal interneurons (GABAergic). These control thalamic output to the motor cortex. Degeneration within the SNpc leads to a decreased thalamic activation of the motor cortex. (Shulman et al, 2011).

The biological function of the nigrostriatal pathway depends on the intactness of its anatomical structure. Preservation of the striatum terminals and of neuronal cell bodies of DA neurons in the SNpc is a prerequisite for the maintenance of the physiological function (Fujita et al. 2014). The nigrostriatal system is anatomically located in the basal ganglia circuit which comprises the motor system structures caudate nucleus, putamen, globus pallidum and substantia nigra. The caudate nucleus and the putamen are collectively called striatum (David Robinson in: Neurobiology, Springer edition, 1997). The system plays a unique integrative role in the control of movement as part of a system called the “basal ganglia motor loop”. This anatomical loop includes structures in the thalamus, motor and somatosensory cortex and wide regions of surrounding cortex. Neurons of the SN produce dopamine (DA) and project to the striatum. They give dopaminergic excitatory (D1 receptors) and inhibitory (D2 receptors) inputs to striatal interneurons (GABAergic). These control thalamic output to the motor cortex. Degeneration within the SNpc leads to a decreased thalamic activation of the motor cortex. (Shulman et al, 2011).

The dopaminergic cells localized in the SNpc synthesize the transmitter substance dopamine (DA) and make extensive contacts within the caudate and putamen (the striatum). These DA neurons have a
complex morphology and high energy demand. They are provided with very long and dense
arborisations projecting into the striatum where DA is released. This unique morphological
characteristics demand a high level of energy to maintain the activity at the synaptic level, to
compensate for the risk of depolarization of the unmyelinated fibres and to support a long distance
axonal transport. This puts a tremendous burden on mitochondrial functions (Pissadaki et al. 2013).
SNpc neurons are provided with specific calcium channels, the L-type Cav 1.3 which are intended to
regulate the autonomous firing as "pacemaker". The high demand of calcium buffering arising from
this is handled by the endoplasmic reticulum (ER) and by the mitochondria. This is a function specific
for SNpc DA neurons, as the dopaminergic neurons belonging to the ventral tegmental area (VTA) are
using Na⁺ channels as a pacemaker. Additional peculiarities of the neurons of the nigrostriatal
pathway are the high number of synapses and the higher probability of these neurons to accumulate
misfolded proteins, including α-synuclein. Furthermore, the nigrostriatal metabolic pathway of DA is
known to induce oxidative and nitrative stress (Fujita et al. 2014, Asanuma et al. 2003, Cantuti-
Castelvetri et al. 2003, Pissadaki et al. 2013) making DA neurons particularly sensitive to oxidative
stress (Lotharius and Brundin, 2002). DA neurons in SNpc also have a relatively low mitochondria
mass which may contribute to the vulnerability of these neurons (Liang et al. 2007). In addition,
increased levels of iron have been observed in SN of PD patients (Gotz et al. 2004) and the high
content of iron in dopamine neurons has been reported to trigger oxidative/nitrative stress and
subsequent neurodegeneration (Ayton and Lei 2014; Benshachar et al. 1991). As a consequence,
these neurons are particularly sensitive to various stressors that can contribute to their preferential
loss (Fujita et al. 2014).

5.2 How it is measured or detected
The presence of DA cells in the SNpc and DA terminals in the striatum can be visualized using
different phenotypic histological markers. Changes can be captured by measurement of markers
specific for dopaminergic neurons such as tyrosine hydroxylase dopamine transporter (DAT) and
vesicular monoamine transporter type 2 (VMAT2). Degenerating and/or degenerated neurons can be
detected by the silver stains and the Fluoro Jade stains.

The silver degeneration stain is considered as the gold standard method to trace degeneration of
axons. By this matter, products from disintegrated cells are visualized (Switzer R., 2000; Betarbet et
al. 2000).

- Fluoro Jade stain is a fluorochrome derived from fluorescein used in neuroscience disciplines
to label degenerating neurons. It is an alternative technique to traditional methods for
labeling degenerating neurons such as silver degeneration staining. Fluoro-Jade may be
preferred to other degeneration stains due to the simplicity of staining procedures, which are
a common drawbacks of conventional stains. However, the mechanism by which fluoro-jade
labels degenerating neurons is unknown (Betarbet et al. 2000, Schmued et al. 1997).

- Detection of TH, the enzyme responsible for catalyzing the conversion of the amino acid L-
tyrosine to L-3,4-dihydroxyphenylalanine (L-DOPA), a precursor for dopamine. Detection of
TH can be done either by immunocytochemistry (at the protein level) followed by cell
counting (quantitative evaluation) or by western blot followed by densitometry analysis

- Counting of cells, immunostained for TH, or counting of nuclei by e.g. with Nissel’s, DAPI
(Kapuscinski, 1995) or Hoechst stain (Latt et al. 1976) should be done following standard
morphometric protocols. However, inclusion of stereological cell counts to assess
neurodegeneration is representing the most sensitive method to confirm quantitatively this

- Quantification of dopaminergic neurons in SNpc: the average number of DA neurons in adult
mouse SN is approximately 8.000 to 14.000, depending on strain (Zaborszky and Vadasz
2001). Their distribution is not homogeneous with difference in density between the caudal
and rostral part of the SN. The gold standard for counting neurons is then to use an unbiased
stereological protocol for cell counting with an optical dissector system (Tieu et al. 2003). This
requires a computerized stereology software. The count should include TH⁺ neurons as well
the total count of neurons using a non-specific cell stain (e.g. Nisell’s, Fox3).
Quantification of dopaminergic terminals in the striatum: the density of dopaminergic terminals is not homogeneous in the striatum, increasing from the rostral to the caudal part and representative regions of the striatum should be assessed. This can be done by digitalization of the fibres and quantification by optical density or quantification of the fiber density identified by TH+ immunoreactivity (Tieu et al. 2003; Fernagut et al. 2007). Alternatively, striatal tissue can be isolated for immunobloting of TH or DAT.

DA transporters (DAT) and vesicular monoamine transporter type 2 (VMAT2) can be visualized and quantified using immunocytochemistry (single cell levels) or western blot followed by densitometry analysis, to quantify the changes in their expression. (Hirata et al. 2007; Fornai et al. 2003; Tong et al. 2011; Ciliax et al. 1995).

DA, DOPAC (DA metabolite) and HVA (homovanillic acid, formed from dopamine that escapes conversion to norepinephrine in noradrenergic neurons throughout the body as well as from dopamine synthesized in dopaminergic neurons that are mainly in brain( Kopin et al. 1988)) content in the striatum can be quantified through several methodologies such as capillary electrophoresis, spectrofluorimetry and high performance liquid chromatography (HPLC). The commonly used detectors for chromatography include MS, UV, optical fiber detector, electrochemical detector and fluorescence detector (Zhao et al. 2011, Fornai et al. 2005, Magnusson et al. 1980).

Indentification of LB in standard histological sections stained with haematoxylin and eosin, they are characterized by the presence of pale eosinophilic vacuoles (Betarbet 2000 and 2006, Pappolla 1988, Dale 1992).


Imaging techniques: 18-fluoro-dopa positron emission tomography (PET) quantification of various dopamine presynaptic markers (e.g. dopamine transporter DAT, vesicular monoamine transporter type 2 VAT2) identified by single photon emission tomography (SPECT). They permit to visualize the loss of nigrostriatal DA neurons in patients (Shapira 2013).

5.3 Evidence Supporting Taxonomic Applicability

Parkinson’s disease (PD) is a progressive age-related human neurodegenerative disease with a multifactorial pathogenesis implicating various genetic and environmental factors and is more prevalent in males (Fujita et al. 2014). However, the anatomy and function of the nigrostriatal pathway is conserved across mammalian species (Barron et al. 2010). Pathological changes, similar to the one described in PD, have been reproduced with chemicals such as rotenone and MPTP. These chemicals have been tested successfully in primates and mice. The mouse C57BL/6 strain is the most frequently used strain in the reported experiments. A difference in vulnerability was observed, particularly for rats, depending on the strain and route of administration. The Lewis strain gives more consistency in terms of sensitivity when compared to the Sprague Dawley. A genetic-based susceptibility has been also described for mice following parquat treatment, underlining the relevance of the genetic component in Parkinsonism syndroms with the C57BL/6J strain resulting the more susceptible(Jiao et al. 2014; Yin et al. 2011). In addition to rodents, the pesticide rotenone has been also studied in Caenorhabditis elegans (C.elegans), Drosophila, zebrafish and Lymnaea Stagnalis (L.stagnalis) (Johnson et al., 2015).
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6.1 How this KE works

Neuroinflammation or brain inflammation differs from peripheral inflammation in that the vascular response and the role of peripheral bone marrow-derived cells are less conspicuous. The most-easily detectable feature of neuroinflammation is the activation of microglial cells and astrocytes. It is evidenced by changes in shape, increased expression of certain antigens, and accumulation and proliferation of these glial cells in affected regions (Aschner, 1998; Graeber & Streit, 1990; Monnet-Tschudi et al, 2007; Streit et al, 1999; Kraft and Harry, 2011; Claycomb et al., 2013). Upon stimulation by cytokines or inflammogens (e.g. from pathogens or from damaged neurons), both glial cell types activate inflammatory signalling pathways, which result in increased expression and/or release of inflammatory mediators such as cytokines, eicosanoids, and metalloproteinases (Dong & Benveniste, 2001), as well as in the production of reactive oxygen (ROS) and nitrogen species (RNS) (Brown & Bal-Price, 2003). Different types of activation states are possible for microglia and astrocytes, resulting in different responses concerning pro-inflammatory/anti-inflammatory signalling and other cellular functions (such as phagocytosis) (Streit et al., 1999; Nakajima and Kohsaka, 2004).

Therefore, neuroinflammation can have both neuroprotective/neuroreparative and neurodegenerative consequences (Carson et al., 2006; Monnet-Tschudi et al, 2007; Aguzzi et al., 2013; Glass et al., 2010). Under normal physiological conditions, microglial cells scan the nervous system for neural integrity (Nimmerjahn et al., 2005) and for invading pathogens (Aloisi, 2001; Kreutzberg, 1995; Kreutzberg, 1996; Rivest, 2009). They are the first type of cell activated (first line of defence), and can subsequently lead to astrocyte activation (Falsig, 2008). Two distinct states of microglial activation have been described (Gordon, 2003; Kigerl et al, 2009; Maresz et al, 2008; Mosser & Edwards, 2008; Perego et al; Ponomarev et al, 2005): The M1 state is classically triggered by interferon-gamma and/or other pro-inflammatory cytokines, and this state is characterized by increased expression of integrin alpha M (Itgam) and CD86, as well as the release of pro-inflammatory cytokines (TNF-alpha, IL-1beta, IL-6), and it is mostly associated with neurodegeneration. The M2 state is triggered by IL-4 and IL-13 (Maresz et al, 2008; Perego et al, 2011; Ponomarev et al, 2007) and induces the expression of mannose receptor 1 (MRC1), arginase1 (Arg 1) and Ym1/2; it is involved in repair processes. The activation of astrocytes by microglia-derived cytokines or TLR agonists resembles the microglial M1 state (Falsig 2006).

### 6.2 How it is measured or detected

Neuroinflammation, i.e. the activation of glial cells can be measured by quantification of cellular markers (most commonly), or of released mediators (less common). As multiple activation states exist for the two main cell types involved, it is necessary to measure several markers of neuroinflammation:

- Microglial activation can be detected based on the increased numbers of labeled microglia per volume element of brain tissue (due to increase of binding sites, proliferation, and immigration of cells). A specific microglial marker, used across different species, is CD11b. Alternatively, various specific carbohydrate structures can be stained by lectins (e.g. IB4). Beyond that, various well-established antibodies are available to detect microglia in mouse tissue (F4/80), phagocytic microglia in rat tissue (ED1) or more generally microglia across species (Iba1). Transgenic mice are available with fluorescent proteins under the control of the CD11b promoter to easily quantify microglia without need for specific stains.

- The most frequently used astrocyte marker is GFAP (99% of all studies) (Eng et al., 2000). This protein is highly specific for astrocytes in the brain, and good clinically-validated antibodies are available for immunocytochemical detection. In neuroinflammatory brain regions, the stain becomes more prominent, due to an upregulation of the protein, a shape change/proliferation of the cells, or better accessibility of the antibody. Various histological quantification approaches can be used. Occasionally, alternative astrocytic markers, such as vimentin of the S100beta protein have been used for staining of astrocytes (Struzynska et al., 2007).

- All immunocytochemical methods can also be applied to cell culture models.

- In patients, microglial accumulation can be monitored by PET imaging, using [11C]-PK 11195 as microglial marker (Banati et al., 2002).

- Activation of glial cells can be assessed in tissue or cell culture models also by quantification of sets of activation markers. This can for instance be done by PCR quantification of inflammatory factors, of by measurement of the respective mediators, e.g. by ELISA-related immuno-quantification. Such markers include:

  - Pro- and anti-inflammatory cytokine expression (IL-1β; TNF-α, IL-6, IL-4); or expression of immunostimulatory proteins (e.g. MHC-II)

  - Itgam, CD86 expression as markers of M1 microglial phenotype

  - Arg1, MRC1, as markers of M2 microglial phenotype

(for description of techniques, see Falsig 2004; Lund 2006; Kuegler 2010; Monnet-Tschudi et al., 2011; Sandström et al., 2014; von Tobel et al., 2014)
6.3 Evidence supporting taxonomic applicability

Neuroinflammation is observed in human, monkey, rat, mouse, and zebrafish, in association with neurodegeneration or following toxicant exposure. Some references (non-exhaustive list) below for illustration:

In human: Vennetti et al., 2006 in monkey (Macaca fascicularis): Charleston et al., 1994, 1996 in rat: Little et al., 2012; Zurich et al., 2002; Eskes et al., 2002 in mouse: Liu et al., 2012 in zebrafish: Xu et al., 2014.

6.4 Regulatory examples using the KE

Measurement of glial fibrillary acidic protein (GFAP), whose increase is a marker of astrocyte reactivity, is required by the US EPA for fuel additives (40 CFR 79.67), but is optional for other toxicant evaluation.
References


ENV/JM/WRPR(2016)34. 2016. Joint Meeting of the Chemicals Committee and the Working Party on Chemicals, Pesticides and Biotechnology. Adverse Outcome pathway on ionotropic glutamatergic receptors and cognition


7. **Adverse Outcome: parkinsonian motor deficits**

7.1 How this key events works

A large number of neurological disorders are characterized by a clinical syndrome with motor symptoms of bradykinesia, tremor, rigidity and postural instability. As these clinical features are common to multiple disorders, the clinical syndrome is referred as "parkinsonism" and when parkinsonism is representing the prevalent part of the syndrome, these are referred as "parkinsonian..."
disorders”. Parkinson’s Disease (PD) is one of parkinsonian disorders and can have an idiopathic, genetic or toxic (i.e. MPTP induced parkinsonism) cause (Dickson 2012). The pyramidal motor system comprises bundles of neurons originating in the motor centers of the cerebral cortex to terminate in the brainstem or in the spinal cord where they are responsible for voluntary control of motor functions (Brooks 1971). The extrapyramidal system, which is in the center of AO, is the part of the motor system primarily involved in the control and regulation of involuntary motor control, and in fine tuning (Barnes 1983). Especially the initiation and maintenance of complex movement patterns or of neuronal regulatory pathways involved in postural control of the body are regulated by the nigrostriatal system that is affected in parkinsonian states. The CNS input is modulated by extrapyramidal circuits before the execution of complex motor movements. The modulated information from the basal ganglia is looped back through the thalamus to the cortex, from where final motor signals are sent via the pyramidal system; i.e. the basal ganglia system is not involved in the control of motor neurons and striatal muscles, but it modulates the signals from the cortex to these systems. Thus, an impaired input of dopamine into the striatum leads to an impairment of this modulation loop, and a disturbance of basal ganglia feedback to the thalamic and cortex. This ultimately manifests in key parkinsonian symptoms such as tremor, rigidity, or bradykinesia (Bernheimer 1973). These conditions can be generated experimentally by dopamine depletion with reserpine (Carlsson), by inhibition of dopamine receptors, by mechanical or chemical ablation of nigrostriatal dopamine neurons (cut of the median forebrain bundle or injection of the toxicant 6-OH-dopamine) or the application of toxicants that leading to a relatively selective death of dopaminergic neurons in the substantia nigra (e.g. MPTP) and therefore a reduction of dopamine in the striatum (Kolata 1983).

The basal ganglia loop include the ventral striatum, the neostriatum composed of the putamen and the caudate nucleus, the globus pallidus pars externa (GPe), the globus pallidus pars interna (GPi), the subthalamic nucleus (STN), the substantia nigra pars reticulata (SNpr) and the substantia nigra pars compacta (SNpc) (Obeso 2008). The main input sites into basal ganglia are the striatum and the STN where cortical (glutamatergic) innervations terminate in a topographically organized manner that largely reflects the organization in the cortex (Fallon 1978, Takada 1998). Both the Gpi and the SNpr represent the main output nuclei projecting into the thalamus (Parent 1999, Alexander 1990). The connection between input and output nuclei is functionally organized into a “direct” and an “indirect” pathway (Silverdale 2003). These two pathways in parallel regulate the activity of the basal ganglia output neurons of the GPi and STN and are modulated by dopamine in the striatum. The dopaminergic terminals in the striatum originate from dopaminergic projections from the SNpc. Striatal dopamine modulates the activity of inhibitory GABAergic medium spiny neurons that make up 90% of all neurons in the striatum (Smith 1994). Medium spiny neurons that preferentially express the D1 dopamine receptor are involved in the direct pathway and directly project into the two main output nuclei (Gpi and SNpr). Activation of the D1 medium spiny neuronal direct pathway results in a reduction of the inhibitory basal ganglia output (Gpi and SNpr) leading to a dis-inhibition of thalamic target neurons (Bolam 2000). These events ultimately lead to an elevated activity in the respective cortical neurons, i.e. D1 signalling in the striatum leads to an increase in motor activity.

Medium spiny neurons predominantly expressing the D2 dopamine receptor mostly project to the GPe (Gerfen 1990). Activation of D2 expressing neurons leads to an inhibition of their activity. D2 neurons of the indirect pathway connect the striatum with Gpi/SNpr via synaptic connections in the GPe and the STN. Activating neurons originating in the STN project into the Gpi/SNpr are glutamatergic. From the STN, activating glutamatergic neuronal projections into the Gpi/SNpr lead to a basal, low activation. Activation of the indirect pathway by striatal dopamine from the substantia nigra hence leads to a low basal inhibitory GABAergic output into thalamic structures, and thus allows a strong motor cortex activation of the thalamic.
Fig. 5 Functional anatomy of basal ganglia.

A) Normal conditions. Striatal (STR) dopamine mainly originates from projections originating in the substantia nigra pars compacta (SNc). The STR is mainly composed of inhibitory GABAergic medium spiny neurons (MSN). MSN involved in the direct pathway directly project to the globus pallidus pars interna (GPI) and the substantia nigra pars reticulata (SNpr) leading to a basal inhibition of these output nuclei. MSN involved in the indirect pathway send inhibitory projections to the globus pallidus pars externa (GPe). Their activity is dampened by dopamine binding to D2 receptor expressing MSN in the striatum.

B) Lack of striatal dopamine. Under conditions of a lack of striatal dopamine, inhibitory GABAergic neurons, originating in the striatum, receive less activation, resulting in a declined inhibition of GPI and SNpr inhibitory output. In the indirect pathway, the lack of dopamine causes a lack of its inhibitory influence on inhibitory GABAergic projections into the GPe. This accelerated inhibition of the GPe results in a decline in its inhibitory output into the STN. The decline in STN inhibition allows its overactivation, resulting in an excessive activation of stimulatory glutamatergic projections into the GPi and SNpr. (according to Silverdale 2003)

Parkinson’s Disease is characterized by a decline in striatal dopamine input from the substantia nigra pars compacta (Smith 1994). Under normal conditions, ganglial output via GPI/SNpr nuclei causes a moderate inhibitory influence on cortical and brainstem motor neurons. A reduction in striatal dopamine leads to an underactivation of D1 receptor-expressing medium spiny neurons of the direct pathway. This insufficient activation of the inhibitory GABAergic medium spiny neurons results in a reduction of its normal inhibitory influence on GPI and SNpr output nuclei. As a consequence, dopamine depletion leads to the overactivation of the inhibitory GABAergic GPI/SNpr output via the direct pathway (Mitchell 1989).

In the indirect pathway, the reduced activation of D2 receptors expressing neurons leads to an overactivation of inhibitory output nuclei projecting into the GPe. The resulting inhibitory output of the GPe is hence reduced, thus leading to a declined inhibition of the STN. Overactivation of the stimulatory glutamatergic projections originating in the STN leads to the hyperactivation of the output GPI/SNpr nuclei. As a consequence of striatal dopamine depletion, the direct pathway becomes underactivated and the indirect pathway becomes overactivated. This leads to an overactivation of the basal ganglia output nuclei. Due to their inhibitory influence on thalamocortical motor centers, the resulting reduced cortical activation leads to the prominent impairment of motor functions in parkinsonian states (Silverdale 2003).

The model of direct and indirect pathways linking striatal dopamine content with the basal ganglia output nuclei has been criticized in recent years as it ignores the influence of extrastriatal dopamine (Smith 2000), or the fact that some medium spiny neurons express dopamine receptors of both the D1...
and of the D₂ type (Surmeier 1996). Principal validity of the model and the central role of striatal dopamine was e.g. demonstrated by L-DOPA-mediated supplementation of striatal dopamine content in unprimed PD patients that causes a partial reduction in the overactivation of GPi/SNpr output (Yuan 2010, Heimer 2006). As an alternative way for symptomatic treatment of parkinsonian conditions, deep brain stimulation of either the STN or the Gpi was shown to relieve from parkinsonian motor features (Mazzone 2003, Odekerken 2013).

7.2 How is it measured or detected

For the analysis of striatal dopamine content and its correlation with motor control, both biochemical analysis methods on the cellular and tissue level as well as behavioral tests are required. Available test models are mice and rats on the one hand and non-human primates and humans on the other. Motor impairment features associated with parkinsonian states in man serve as reference standard. Monkey models have the advantage to largely reflect complex motor impairment patterns observed in humans which are rather difficult to assess in rodents. Rodent models in contrast are cost-efficient and allow both biochemical analysis that require major invasive methods as well as basic behavioral tests. Due to the limitations in the assessment of moderate motor impairment in rodents and the well-established correlation between striatal dopamine content and impaired motor output, analysis of striatal dopamine is often applied as surrogate readout for the assessment of motor deficits.

7.2.1 Detection of striatal dopamine (total or extracellular)

The standard method used in the majority of experimental work is the determination of total contents of dopamine and its two degradation metabolites HVA and DOPAC. For this purpose, the striatum is quickly removed from experimental animals, homogenized in a suitable acidic buffer, and the dopamine (metabolites) determined by HPLC with electrochemical detector or by HPLC-MS.

For live in vivo detection of extracellular dopamine levels, a microdialysis probe is inserted into the striatum. Microdialysis can be performed in anesthetized animals or freely moving animals; basal dopamine levels or stimulated levels (amphetamine, KCl) can be recorded. Dopamine and its metabolites are detected in the dialysate either by HPLC or by HPLC-mass spectrometry analysis (Saraswat 1981, Cui 2009, Gonzalez 2011).

7.2.2 Detection of dopamine neuron terminals in the striatum

As alternative to the detection of striatal dopamine that is to a large extent limited to live detection setups due to its instability in tissues, the number of remaining dopamine neurons in the substantia nigra pars compacta was suggested as alternative readout (Burns 1983). It allows the analysis of ex vivo samples without the limitations associated with the instability and reactivity of extravesicular dopamine. Although the number of surviving dopamine neurons in the SNpc in PD or in complex-I inhibitor challenged test animals is a valuable parameter on its own, it was discovered that the number of DA neurons in the SNpc not necessarily correlates with the amount of dopamine released in the striatum. Tyrosine hydroxylase (TH) was regularly stained as marker for DA neurons, however it was observed that TH expression was very variable following MPTP intoxication in the absence of cell death and therefore has only limited suitability for the assessment of DA neuronal numbers (Aznavour 2012). Second, many DA neurites and terminals displayed damage or degradation in the absence of death of the corresponding neuronal cell (Ling 2015). Hence, even in the presence of viable DA neurons in the SNpc, their corresponding terminals could no longer be able to release dopamine into the striatum. Staining of DA neuronal terminals in the striatum is therefore used as a more reliable indirect marker for striatal dopamine content. For the analysis of nigrostriatal terminals, the dopamine transporter (DAT) is visualized either by antibody-mediated staining in tissue slices or by the application of radioactively labeled DAT ligands that allow their application both in vivo and in ex vivo samples (Morris 1996).

7.2.3 Behavioral tests: Rodent models

Rotation: the rotation model of Ungerstedt et al (Ungerstedt 1970) is based on the unilateral lesion of the nigrostriatal dopamine neuron system either in rodents or in non-human primates. The lesion can be produced either surgically, or by stereotaxic infusion of e.g. 6-OHDA into the nigrostriatal system of one hemisphere, or by infusion of MPTP through one carotid (single sided). After the lesion,
animals are left to recover, then the dopamine system is stimulated by injection of amphetamine. The asymmetry of remaining dopamine neurons (only on one side) triggers spontaneous asymmetric motor behaviour, i.e. rotations of the animals. Each full turn of an animal is recorded, the respective numbers of left- and right turns are plotted versus time, respectively. In the standard rotation model, monkeys become hypokinetic in the limbs on the contralateral side of the brain hemisphere treated. Rats preferentially rotate towards the side of the lesion upon treatment with drugs that trigger activation of the remaining dopamine neurons.

Rotarod: assessment of motor coordination. The animals are placed on a rotating rod that is subjected to linear acceleration. The latency to fall from the rod is detected (Jones 1968).

Hang test: Detection of neuromuscular strength. Mice are placed on a horizontal grid. When the animals grabbed the grid with their fore- and hindpaws, the grid is inverted with the animal hanging upside down. In a typical setup, mice are required to remain on the grid for at least 30 s (Tillerson 2002).

Forepaw Stride length during walking. Ink is applied to the forepaws and the mice walk across a blank sheet of paper. Training of the animals to walk across the white paper in a straight line without stopping is performed before the respective treatment. The distance between single steps on each side are measured (Klapdor 1997).

Grid test: Mice hang upside down for 30 s on the grid that is also used for the Hang test and are recorded on video for closer analysis. With this method, the average forepaw distance is measured by assessing the distance covered, divided by the number of successful forepaw steps. In the course of the analysis, the number of unsuccessful forepaw steps are detected and displayed as percentage of the total number of steps performed (Crawley 1999).

Akinesia: the animal is placed on a flat surface and the latency until it has moved all of its four limbs is assessed.

Open field test: Infrared beams detect the animals activity for the determination of parameters such as the time spent locomoting, the distance travelled, or the number of rearings.

Pole test: the animal is placed on a gauze-taped pole with the head upwards below the top of the pole. Two parameters are detected: 1) time until animals turn by 180°; 2) time until the animals reach the floor.

7.2.4 Non-invasive imaging of DA neuron terminals

Positron emission tomography (PET): Based on its appropriate half life time of ca. 2 h for clinical investigations, fluorine-18 labeled L-[18F]-fluorodopa is routinely used in trace amounts for intravenous administration. Striatal uptake of L-[18F]-fluorodopa is followed by applying positron emission tomography (PET) (Leenders 1986).

Single photon emission computed tomography (SPECT): monitoring of dopamine transporter (DAT). Iodine-123-β-CIT is used as a sensitive ligand for dopamine and serotonin transporters and was applied in monkeys and humans (Winogrodzka 2003).

7.2.5 Human neurological tests

A recent systematic review and evaluation of currently used rating scales for the assessment of motor impairment and disability in PD patients identified the 1) Columbia University rating scale, 2) the Northwestern University Disability Scale, and 3) the Unified Parkinson's Disease rating scale as the most evaluated and reliable scales available (Ramaker 2002). All scales evaluate several parameters, some of which are not motor related. Thus, only subscales are useful for readout of motor symptoms (e.g. 13 of the 42 UPDRS parameters). Of these, not all are equally dependent on nigrostriatal dopamine. Examination needs to be done by a trained neurologist.

7.2.6 Regulatory examples using this Adverse Outcome

Neurotoxic effects shall be carefully addressed and reported in routine required regulatory toxicological studies (acute toxicity studies, short-term toxicity studies, long term toxicity and carcinogenicity studies and reproductive toxicity studies). Regarding neurotoxicity in rodents, inclusion
of neurotoxicity investigations in routine toxicology studies shall also be considered. For pesticide active substances the circumstances in which neurotoxicity studies should be performed are listed in Regulation (EU) No 283/2013:

Specific neurotoxicity studies in rodents shall be performed in case of one those following conditions:

- there is indication of neurotoxicity in routine toxicity studies carried out with the active substance;
- the active substance is a structurally related to known neurotoxic compound;
- the active substance has a neurotoxic mode of pesticidal action.

As a result, specific neurotoxicity studies are not routinely required for all pesticide active substances. Specific neurotoxicity testing becomes obligatory only if neurotoxicity has been observed during histopathological evaluation or in case of structural analogy with a known neurotoxic compound. Motor activity should be measured once in short-term repeated dose toxicity studies (OECD 407, 408 and 422) and several times in specific neurotoxicity studies (OECD 424, OECD 426 and cohort 2 of OECD 443). However, this is not a requirement in chronic toxicity studies unless neurotoxic effects have been reported in the shorter studies. The same test (measures horizontal and/or vertical movements in a test chamber) is implemented in both routine studies and neurotoxicity studies. Coordination and balance are evaluated by rotation or rotarod or pole tests, and gait abnormalities by forepaw stride length test. Those tests are not required by any repeated dose toxicity OECD guidelines and they can be optionally incorporated in the design of neurotoxicity studies OECD 424 and OECD 426.

Although motor deficits is the AO in this AOP, degeneration of DA neurons, is also considered an adverse effect in the regulatory framework, even in the absence of clear clinical symptoms or motor deficits. Morphological assessment of brain structures is a standard requirement in the regulatory toxicological studies supporting the risk assessment of chemical substances and it is a regulatory expectation that the anatomical structures belonging to the nigrostriatal pathway would be included and evaluated as part of the standard evaluation of the brain. Treatment related neuronal degeneration, when occurring as a consequence of the treatment, is generally dose-dependent in incidence and severity. However, if not accompanied by clinical signs or behavioral changes indicative of central nervous system pathology, minimal loss of DA neurons would likely remain undetected in the standard histological evaluation, due to the presence of non DA neurons or as a consequence of the subjectivity of non-quantifiable analysis, unless specific markers are used. As multiple forms of perturbation can affect the neurons, some changes are potentially still reversible (e.g. loss of TH or DA) and irreversibility should be confirmed as part of the assessment. It is then important to apply a sensitive and appropriate method (Switzer 2000) and evaluation of the phenotypic markers in the striatum and in the SNpc should be always performed as a minimum standard (Minnema et al 2014) when investigating perturbation of the nigrostriatal pathway. It should additionally considered that rat is likely to be a poor model to capture this kind of hazard, as demonstrated by the poor sensitivity of rat to MPTP or related compounds and this should be taken into account for the design and interpretation of the studies.

Dissimilarities of chemical induced animal models to human disease are also important and should be carefully weighted when considering the duration and schedule of the study/treatment. Differently from the human disease, with the MPTP animal model, the damage occurs rapidly, is hardly progressive, is little age-dependent and formation of Lewy bodies is sometime not occurring (Efremova et al. 2015). Therefore, for different animals models, the standard 90 days toxicity study could not match with the chronic and progressive characteristics of the human disease and compensatory changes influencing DA metabolism and turnover and protein catabolis can occur during the treatment period with an impact on the time of onset of the lesion (Ossowska et al. 2005).
**References**


KEY EVENTS RELATIONSHIPS (KERS)

1st KER: Binding of inhibitor to NADH-ubiquinone oxidoreductase (complex I) leads to its inhibition

1.1 How does this Key Event Relationship work

It is well documented that binding of an inhibitor to CI inhibits its activity (see MIE). Naturally occurring and synthetic CI inhibitors have been shown to inhibit the catalytic activity of CI, leading to partial or total inhibition of its activity in a dose response manner (Degli Esposti and Ghelli, 1994; Ichimaru et al. 2008; Barrientos and Moraes, 1999; Betarbet et al., 2000). Indeed, binding of inhibitors stops the electron flow from CI to ubiquinone. Therefore, the Fe-S clusters of CI become highly reduced and no further electrons can be transferred from NADH to CI. This leads to the inhibition of the NADH oxidoreductase function, i.e. CI inhibition.

1.2 Weight of Evidence for the KER

The weight of evidence supporting the relationship between binding of an inhibitor to NADH-ubiquinone oxidoreductase and its inhibition is strong.

1.2.1 Biological Plausibility

There is an extensive understanding of the functional relationship between binding of an inhibitor to NADH-ubiquinone oxidoreductase (CI) and its inhibition. As the first entry complex of mitochondrial respiratory chain, CI oxidizes NADH and transfers electrons via a flavin mononucleotide cofactor and several Fe-S complexes to ubiquinone. The electron flow is coupled to the translocation of protons from the matrix to the intermembrane space. This helps to establish the electrochemical gradient that is used to fuel ATP synthesis (Greenamyre et al., 2001). If an inhibitor binds to CI, the electron transfer is blocked. This compromises ATP synthesis and maintenance of Δψm, leading to mitochondrial dysfunction. As CI exerts a higher control over oxidative phosphorylation in synaptic mitochondria than in non-synaptic mitochondria in the brain (Davey and Clark, 1996), specific functional defects observed in PD may be explained.

It is well documented that CI inhibition is one of the main sites at which electron leakage to oxygen occurs. This results in a production of ROS, such as superoxide (Efremov and Sazanow, 2011) and hydrogen peroxide, which are main contributors to oxidative stress (Greenamyre et al., 2001).

1.2.2 Empirical support for linkage

A variety of studies show a significant correlation between binding of an inhibitor to CI and its inhibition, usually measured by the decreased mitochondrial respiration. Different classes of CI inhibitors, such as rotenone, MPP+, piericidin A, acetogenins, pyridaben, and various piperazin derivatives (Ichimaru et al. 2008) have been shown to bind to the ubiquitin site of CI, leading to a partial or total inhibition of oxidoreductase activity in a dose response manner (Grivennikova et al., 1997; Barrientos and Moraes, 1999; Betarbet et al., 2000).

The reduction of CI activity is well documented in a variety of studies using isolated mitochondria or cells, as well as in in vivo experiments and in human post mortem PD brains. Usually it is measured by assays described in 2nd Key Event Relationship (KER): Inhibition of complex I leads to mitochondrial dysfunction.

It has been shown that binding of rotenone to CI (e.g. Betarbet et al., 2000, Greenamyre et al., 2001) or MPP+ (e.g. Krug et al., 2014; Langston, 1996) can reproduce the anatomical, neurochemical, behavioural and neuropathological features of PD. Therefore, the empirical support for this KER will be mainly based on the experiments performed after exposure to rotenone or MPP+.

- The binding of rotenone to CI resulted in time- and dose-dependent inhibition of CI activity measured in sub-mitochondrial particles. The kinetics of the active CI inhibition was determined after exposure to rotenone at 20, 30 and 40 nM at different times of exposure (30 sec, 1 min or 2 min) (Grivennikova et al., 1997). This study suggests that two rotenone...
binding sites exist in CI: one affecting NADH oxidation by ubiquinone and the other one operating in ubiquinol-NAD+ reductase action.

- Partial inhibition of CI produces a mild, late-onset mitochondrial damage. The threshold effect seen in brain mitochondria (25–50% decrease in activity) may not directly impact ATP levels or Δψm but could have long-term deleterious effects triggered by oxidative stress, as it has been shown that an electron leak upstream of the rotenone binding site in CI leads to ROS production (Greenamyre et al., 2001).

- Exposure of rats to rotenone (2 days, 2 mg/kg) produced free brain rotenone concentration of 20–30 nM and resulted in 73% inhibition of specific binding to CI of [3H] dihydrotrotenone (Betarbet et al., 2000). However, oximetry analysis indicated that in brain mitochondria (but not liver mitochondria) this rotenone concentration (30 nM maximum) was insufficient to inhibit glutamate (CI substrate)-supported respiration (Betarbet et al., 2000) suggesting that this rotenone concentration did not alter mitochondrial oxygen consumption in isolated brain mitochondria.

- Rotenone has been reported to be a specific and potent mitochondrial CI inhibitor with IC50 values from 0.1 nM to 100 nM depending on the system and methods used (Lambert and Brand, 2004; Ichimaru et al., 2008; Chinopoulos and Vizi, 2001; Beretta et al., 2006).

- Mesencephalic cultures prepared from C57/BL6 mice and treated with 5, or 10 nM rotenone for 24 h inhibited CI activity by 11% or 33%, respectively (Choi et al., 2008).

- The inhibition of CI was studied in the human osteosarcoma-derived cell line (143B) after the exposure to rotenone or using a genetic model (40% loss of CI activity in human xenomitochondrial cybrids (HXC) lines). Different degrees of CI inhibition were quantitatively correlated with levels of decreased cellular respiration (Barrientos and Moraes, 1999). Only when CI was inhibited by 35-40% (< 5 nM rotenone), cell respiration decreased linearly until 30% of the normal rate. Increasing concentrations of rotenone produced further but slower decrease in CI activity and cell respiration (Fig. 1). Cells with the complete rotenone-induced CI inhibition still maintain a cell respiration rate of approximately 20% because of an electron flow through complex II. At high concentrations (5–6-fold higher than the concentration necessary for 100% CI inhibition), rotenone showed a secondary, toxic effect at the level of microtubule assembly (Barrientos and Moraes 1999).

- Bovine sub-mitochondrial particles were used to test rotenone affinity binding at 20 nM. This concentration of rotenone reduced the NADH oxidation rate by approximately 50% (Okun et al., 1999)

- MPP+ (an active metabolite of MPTP) is an inhibitor of CI (Nicklas et al., 1987; Mizuno et al, 1989; Sayre et al., 1986). Inhibition of the mitochondrial CI by MPP+ supresses aerobic glycolysis and ATP production (Book chapter in Cheville 1994).

- MPP+ binds loosely to CI and causes reversible inhibition of its activity: approximately 40% inhibition was observed at 10 mM concentration within 15 min of incubation. However, prolonged incubation (> 15min) produces up to 78% of irreversible inhibition of CI (Clee ter et al., 1992).

**Human studies**

- There are many studies that show impaired catalytic activity of CI in multiple PD post-mortem brain tissues. For example (Parker and Swerdlow, 1998), five PD brains were used to measure activities of complexes I, III, IV, and of complexes I/III together (NADH: cytochrome c reductase). These measurements were performed in purified frontal cortex mitochondria and revealed a significant loss of CI activity in these PD samples as compared to controls.

- Human data indicate that impairment of CI activity may contribute to the pathogenic processes of PD (for example, Greenamyre et al., 2001; Schapira et al., 1989; Shults, 2004).
1.3 Uncertainties or inconsistencies

- It is not clear the number of subunits constituting CI in mammals, as according to the existing literature different numbers are cited (between 41-46) (Vogel et al., 2007a; Hassinen, 2007).
- The majority of data claims that mammalian CI is composed of 46 (Greenamyre et al., 2001; Hassinen, 2007) or 45 subunits (Vogel et al., 2007a). It is not sure whether there may exist tissue-specific subunits of CI isoforms (Fearnley et al., 2001). It is unclear, which subunit(s) bind rotenone or other inhibitors of CI.
- Additionally, it is not clear whether CI has other uncharacterized functions, taking into consideration its size and complexity (43-46 subunits vs. 11 subunits of complex III or 13 subunits of complex IV) (Greenamyre et al., 2001).
- There is no strict linear relationship between inhibitor binding and reduced mitochondrial function. Low doses of rotenone that inhibit CI activity partially do not alter mitochondrial oxygen consumption. Therefore, bioenergetic defects can not account alone for rotenone-induced neurodegeneration. Instead, under such conditions, rotenone neurotoxicity may result from oxidative stress (Betarbet et al., 2000). Few studies used human brain cells/human brain mitochondria. Therefore, full quantitative data for humans are not available.

1.4 Quantitative understanding

The kinetics of binding and CI inhibition by rotenone has been quantitatively evaluated in a dose-dependent manner using the sub-mitochondrial particles (Grivennikova et al., 1997). The consequences of CI inhibition were quantitatively measured by a variety of assays that are used to study mitochondrial dysfunction (see Key Event Relationship (KER): Inhibition of Complex I leads to mitochondrial dysfunction). There are also many in vitro and in vivo studies combining the quantification of CI inhibition and DA cell death (e.g. Choi et al., 2008, Betarbet et al., 2000, see KER Mitochondrial dysfunction induces degeneration of nigrostriatal pathway).

The binding of different classes of inhibitors (e.g., pesticides, drugs and other toxins) to CI has been determined quantitatively and $I_{50}$ and $K_I$ values are available. Potency relative to that of rotenone has been determined under the same conditions in beef mitochondria or submitochondrial particles using the ratio of the $K_I$ values, when they were available (Degli Esposti, 1998; Okun et al., 1999). Rotenone $I_{50}$ value is defined as 20 nM (Okun et al., 1999).

Example of a quantitative evaluation of concentration-dependent CI inhibition by rotenone (from Barrientos and Moraes, 1999, Fig. 6).
**Fig. 6.** Effect of CI (NADH decylubiquinone reductase) inhibition on endogenous cell respiration. Cells were treated with different concentrations of rotenone for 4 h before measuring cell respiration in whole cells and CI activity in isolated mitochondria. Complete CI inhibition was achieved with 100 nM rotenone. The cell respiration was inhibited also in a dose-dependent manner but showed different inhibition kinetics and a saturation threshold. For comparison, the genetically-altered cell line HXC had an approximately 40% CI reduced activity and an approximately 80% residual cell respiration. HXC, human xenomitochondrial cybrids.

Time- and concentration-relationship of NADH oxidase inhibition by rotenone (Fig. 7. from Grivennikova et al., 1997).

**Table 1:** Quantitative evaluation of the 1st KER: Binding of inhibitor to NADH-ubiquinone oxidoreductase (MIE; KE upstream) leads to its inhibition (KE downstream)

<table>
<thead>
<tr>
<th>MIE (KE upstream) Binding of inhibitor to NADH-ubiquinone oxidoreductase (nM)</th>
<th>KE (downstream) Inhibition of CI (%), approximately</th>
<th>Comments (in vivo, in vitro or human studies)</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Administration of rotenone at 2 mg/kg per day for 2 days resulted in free rotenone concentration of 20–30 nM in the brain.</td>
<td>75%</td>
<td>DA neuronal cell death determined after rotenone administration at 1 to 12 mg/kg per day, Sprague Dawley and Lewis rats infused continuously by jugular vein, 7 days up to 5 weeks</td>
<td>Betarbet et al., 2000</td>
</tr>
</tbody>
</table>
### 20 nM rotenone

| Direct binding studies using bovine and Musca domestica sub-mitochondrial particles | 50% | Binding studies that defined the $I_{50}$ and $K_d$ values for three classes of CI inhibitors (12 chemicals) including rotenone. | Okun et al., 1999 |

### Human skin fibroblasts exposed to 100 nM Rotenone for 72 hr

| 20% | In the same experiment mitochondria morphology, motility was also evaluated. | Koopman et al., 2007 |

### 0-2.5 nM Rotenone

| 5/10 nM Rotenone | No effect | Treatments with 5 or 10 nM rotenone killed 50% or 75% DA neurons respectively. | Choi et al., 2008 |

### 5/10 nM Rotenone

| Mesencephalic neurons were cultured from E14 C57/BL6 mouse embryos for 6 days and then treated with rotenone for 24 hr | 11% and 33%, respectively |

### 1-2.5-5-7.5-10-20 nM

<table>
<thead>
<tr>
<th>10-20: 35-50- 65-80 %</th>
<th>In this study time course of the active and deactivated enzymes inhibition by rotenone and Piericidin A is study in a dose-dependent manner.</th>
</tr>
</thead>
<tbody>
<tr>
<td>5- 75 %</td>
<td>Binding studies in sub-mitochondrial particles prepared from bovine heart after 20 min of exposure to rotenone.</td>
</tr>
</tbody>
</table>

### 1-10-20-80 nM

| 55-78 % | In the same study similar experiments were performed using HXC cell line (see Fig. 1 above). | Barrientos and Moraes 1999 |

### 1-20 nM

| 80% | 87% | 100% |

### 5-10 nM

| 143B Cells (human osteosarcoma), exposed for 4 hrs to rotenone | 80% | 87% | 100% |

### 50-100 nM

| 80% | 87% | 100% |

### 1-20-80 nM

| 55-78 % | In the same study similar experiments were performed using HXC cell line (see Fig. 1 above). | Barrientos and Moraes 1999 |

### 1.5 Evidence Supporting Taxonomic Applicability

The CI is well-conserved across species from lower organism to mammals. The central subunits of CI harboring the bioenergetic core functions are conserved from bacteria to humans. CI from bacteria and from mitochondria of *Yarrowia lipolytica*, a yeast genetic model for the study of eukaryotic CI (Kerscher et al., 2002) was analyzed by x-ray crystallography (Zickermann et al., 2015).

However, the affinity of various chemicals to cause partial or total inhibition of CI activity across species is not well studied (except for rotenone).
References

Barrientos A, and Moraes CT. 1999. Titrating the Effects of Mitochondrial Complex I Impairment in the Cell Physiology. 274(23)16188–16197 http://www.jbc.org/content/274/23/16188.full.pdf Fig 1 p.16190. © the American Society for Biochemistry and Molecular Biology.


2nd KER: Inhibition of Complex I lead to mitochondrial dysfunction.

2.1 How does this Key Event Relationship work

Inhibited CI is unable to pass off its electron to ubiquinone and it cannot translocate protons across the mitochondrial inner membrane. This creates a back-up of NADH within the mitochondrial matrix (Brown and Borutaite, 2004). This leads to an arrest of the citric acid cycle and a failure to build a proton gradient (mitochondrial membrane potential, Δψm) across the inner membrane. This results in impaired ATP production. In addition, the direct transfer of electrons from CI to oxygen is increased. This leads to oxidative stress as ROS (e.g. superoxide, hydrogen peroxide) are produced, which can damage DNA, proteins, lipids and other cell components and function (Sanders et al., 2014).

2.2 Weight of Evidence

The weight of evidence supporting the relationship between inhibition of CI and mitochondrial dysfunction is strong. The mechanisms behind this KER are partially understood and well documented based on in vitro as well as in vivo experiments (e.g., Sanders et al., 2014), complemented by data from human post-mortem PD brain evaluations (Parker et al., 1989; Greenamyre et al., 2001; Sherer et al., 2003; Schapira et al., 1989).

2.2.1 Biological Plausibility

The biological plausibility that inhibition of CI activity triggers mitochondrial dysfunction is strong. It is well understood, how the inhibition of CI can lead to mitochondrial dysfunction as measured by: a) decreased oxygen consumption, b) decrease or loss of ATP production, c) decrease of Δψm, d) the loss of mitochondrial protein import and protein biosynthesis, e) reduced activities of enzymes of the mitochondrial respiratory chain and the Krebs cycle, f) elevated levels of ROS, g) the loss of mitochondrial motility, causing a failure of mitochondria to re-localize to sites of increased energy demands (such as synapses), h) destruction of the mitochondrial network, i) increased mitochondrial uptake of Ca²⁺ causing mitochondrial Ca²⁺ overload (Graier et al., 2007) and opening of mitochondrial PTP, j) rupture of the mitochondrial inner and outer membranes, leading to release of mitochondrial pro-death factors, including cytochrome c, AIF and endonuclease G (Braun, 2012; Martin, 2011; Correia et al., 2012; Cozzolino et al., 2013). These pathological mechanisms are extremely well studied.

2.2.2 Empirical support for linkage

Many studies show that the pathophysiological consequences of a partial or total CI inhibition are linked to mitochondrial dysfunction. In many of these experiments the cellular damage caused by mitochondrial dysfunction is reduced (or entirely prevented) by treatment with antioxidants.

Different degrees of CI inhibition by rotenone have been studied in the human osteosarcoma-derived cell line (143B). A quantitative correlation between increasing inhibition of CI and mitochondrial dysfunction (as shown by inhibition of mitochondrial respiration, reduced ATP production, increased ROS release and lipid peroxidation, as well as decreased Δψm) was established (Fig. 1 and Table 1 based on Barrientos and Moraes, 1999).

Based on the existing literature it is suggested that rotenone exerts toxicity via oxidative stress, rather than via decrease of ATP synthesis (bioenergetics effects).

A few examples illustrating mitochondrial damage and oxidative stress in animal model of PD and human cells induced by:

Rotenone

- Rotenone administered subcutaneously for 5 weeks (2.5 mg/kg/d) caused a selective increase (by ~2 folds) in oxidative damage in the striatum, as compared to the hippocampus and cortex, accompanied by massive degeneration of DA neurons (~80% decrease) in the substantia nigra. Rotenone reduced intracellular ATP levels in the striatum (by >40%), increases malondialdehyde (MDA, indicative of lipid peroxidation, by ~60%), reduced GSH levels (by ~20%), thioredoxin (by ~70%), and manganese superoxide dismutase (SOD, by...
Rotenone was administered 2.5 mg/kg body weight to male Wistar rats for 4 weeks in the presence or absence of ferulic acid (FA, at the dose of 50 mg/kg) that has antioxidant and anti-inflammatory properties. Rotenone administration caused DA neuronal cell death (~50%), significant reduction in endogenous antioxidants, such as superoxide dismutase (~75%), catalase (~40%), and glutathione (~50%), and induced lipid peroxidation evidenced by increased MDA formation (~2 folds). Treatment with FA rescued DA neurons in substantia nigra pars compacta area and nerve terminals in the striatum, as well as restored antioxidant enzymes, prevented depletion of glutathione, and inhibited lipid peroxidation induced by rotenone (Ojha et al., 2015).

Many studies have shown that mitochondrial aldehyde dehydrogenase 2 (ALDH2) functions as a cellular protector against oxidative stress by detoxification of cytotoxic aldehydes. Dopamine is metabolized by monoamine oxidase to yield 3,4-dihydroxyphenylacetalddehyde (DOPAL) then converts to a less toxic acid product by ALDH. The highly toxic and reactive DOPAL has been hypothesized to contribute to the selective neurodegeneration of DA neurons. In this study, rotenone (100 nM, 24 hr) in both SH-SYSY cells and primary cultured substantia nigra (SN) DA neurons, was shown to reduce DA cell viability (~40%), decrease Δψm (~40%, as shown by TMRM), induce mitochondrial ROS production (~30%, as shown by increase of MitoSox Red), and increased cytosolic protein levels of proteins related to the mitochondrial apoptotic pathway (i.e. Bax, cytochrome c, active caspase-9 and active caspase-3) (~2 folds for all proteins).

The neuroprotective mechanism of ALDH2 was observed as overexpression of wild-type ALDH2 gene (but not the enzymatically deficient mutant ALDH2*2 (E504K)) reduced rotenone-induced DA neuronal cell death, prevented rotenone-induced reduction in TMRM signal (95.7±1.6% v.s. 67±3.5%), and prevented rotenone-induced increase in MitoSox Red intensity (103±1% v.s. 134±0.6%). Additionally, pre-treatment of cells with Alda-1 (activator of ALDH2) (1–10 μM, for 24 hr) prevented rotenone-induced loss of Δψm and ROS production in a dose-dependent manner. These results were confirmed by in vivo studies. Rotenone (50 mg/kg/day, oral administration for 14 days) or MPTP (40 mg/kg/day, i.p. for 14 days) both administered to C57BL/6 mice caused significant SN TH+ DA neuronal cell apoptosis (~50%). Alda-1 attenuated rotenone-induced apoptosis by decreasing ROS accumulation, reversing Δψm depolarization, and inhibiting the activation of proteins related to mitochondrial apoptotic pathway. The present study demonstrates that rotenone or MPP+ induces DA neurotoxicity through oxidative stress. Moreover, Alda-1 is effective in ameliorating mitochondrial dysfunction by inhibiting rotenone or MPP+ induced mitochondria-mediated oxidative stress that leads to apoptosis (Chiu et al., 2015).

Rotenone-induced mitochondrial dysfunction was observed in human neuroblastoma cells exposed to 5 nM rotenone for 1-4 weeks. After 3-4 weeks of treatment, rotenone-treated cells showed evidence of oxidative stress, including loss of GSH (by 5%) and increased oxidative DNA (qualitative, measured by using antibodies to 8-oxo-dG) and protein damage (223 ± 29% of control, as shown by the large increase in protein carbonyls in the insoluble fraction) (Sherer et al. 2002). This chronic rotenone treatment markedly sensitized cells to further oxidative challenge since in response to H2O2 cytochrome c release from mitochondria and caspase-3 activation occurred earlier and to a greater extent in rotenone-treated cells vs Ctr (1.44 ± 0.02% vs 0.38 ± 0.07% apoptosis/hr). This study indicates that chronic, low-level CI inhibition by rotenone induces progressive oxidative damage, and caspase-dependent neuronal cell death (Sherer et al., 2002).

By using anti-oxidant, kaempferol (6 μM, 1 hr prior addition of rotenone) and rotenone (50 nM, max up to 24 hr) on SH-SYSY cells, kaempferol was found to counteract rotenone-induced ROS production (especially superoxide: with kaempferol, ethidium fluorescence decreased below the control (Ctr) levels), rotenone-induced mitochondrial oxidative dysfunction (protein carbonyls values: 2.5 in Ctr, 6.2 with rotenone, 2.7 with kaempferol +
In preclinical studies protective effects in PD animal models. Rats treated with rotenone and PLL derived alkaloids showed decreased ROS, stabilized \( \Delta \psi_m \), and the opening of the mitochondrial PTP - which is triggered by ROS production - was inhibited. In addition, rotenone-induced apoptosis was abrogated in the presence of these alkaloids (Wang H. et al., 2014).

**Antioxidant (Piperaceae; PLL)** with some anti-inflammatory activities demonstrated in preclinical studies protective effects in PD animal models. Rats treated with rotenone and PLL derived alkaloids showed decreased ROS, stabilized \( \Delta \psi_m \), and the opening of the mitochondrial PTP - which is triggered by ROS production - was inhibited. In addition, rotenone-induced apoptosis was abrogated in the presence of these alkaloids (Wang H. et al., 2015).

- Adult male Sprague–Dawley rats were intranigraly infused with rotenone (6 \( \mu \)g in 1 \( \mu \)l) alone or in the presence of L-deprenyl (0.1, 1, 5 and 10 mg/kg; i.p.) at 12 h intervals for 4 days. Rotenone alone (100 \( \mu \)M, 30 min) increased the levels of hydroxyl radials in the mitochondrial P2 fraction 2,3-DHBA (122.90 \( \pm \) 5.4 pmol/mg protein) and 2,5-DHBA (146.21 \( \pm \) 6.3 pmol/mg protein). L-deprenyl (100 nM–1 mM) dose-dependently attenuated rotenone-induced \( \cdot \)OH generation in the mitochondrial P2 fraction. L-deprenyl-induced attenuation in the rotenone-mediated 2,3-DHBA generation was from 17 \( \pm \) 1.1% to 67 \( \pm \) 4.3%, respectively, for 100 nM–1 mM of the MAO-B inhibitor. Also, rotenone caused about 51 \( \pm \) 3.3% reduction in GSH levels in the cell body region, SN and 34 \( \pm \) 1.1% decrease in the nerve terminal region, NCP (nucleus caudatus putamen). L-deprenyl alone did not cause any significant difference in the GSH content in either region. L-deprenyl treatment dose-dependently attenuated the rotenone-induced GSH depletion in SN from 51 \( \pm \) 3.1% to 44 \( \pm \) 2.1%, 32 \( \pm \) 1.7% and 9 \( \pm \) 1.0%, respectively, for doses of 1, 5 and 10 mg/kg. Additionally, SOD activity was assayed in rotenone-lesioned animals, which were treated with L-deprenyl at different doses (1–10 mg/kg). SN exhibited 2- and 3-fold activity of Cu/Zn-SOD (i.e. cytosolic SOD fraction) and Mn-SOD (i.e. particulate SOD fraction), respectively, compared to the nerve terminal region, NCP.

- MtDNA damage is detectable prior to any signs of neuronal degeneration and is produced selectively in midbrain neurons. The selective vulnerability of midbrain neurons to mtDNA damage was not due to differential effects of rotenone on CI since rotenone suppressed respiration equally (\(-60\%\)) in midbrain and cortical neurons compared to vehicle. However, in response to CI inhibition, midbrain neurons produced more mitochondrial \( \text{H}_2\text{O}_2 \) (5 min of rotenone increased Mitopy1 fluorescence of \(-10\%\) in midbrain mitochondria vs vehicle, and progressively for the duration of measurement), than cortical neurons. The selective mtDNA damage in midbrain could serve as a molecular marker of vulnerable nigral neurons in PD. Oxidative damage to cell macromolecules in human PD and the rotenone model have been recently reviewed (Sanders et al., 2014).

- To model the systemic mitochondrial impairment, rats were exposed to rotenone. A single rotenone dose (10 nM, for 24 hr) induced mtDNA damage in midbrain neurons (>0.4 lesions/10kb vs 0 lesions/10kb in vehicle), but not in cortical neurons; similar results were obtained in vitro in cultured neurons. Importantly, these results indicate that mtDNA damage is detectable prior to any signs of neuronal degeneration and is produced selectively in midbrain neurons. The selective vulnerability of midbrain neurons to mtDNA damage was not due to differential effects of rotenone on CI since rotenone suppressed respiration equally (\(-60\%\)) in midbrain and cortical neurons compared to vehicle. However, in response to CI inhibition, midbrain neurons produced more mitochondrial \( \text{H}_2\text{O}_2 \) (5 min of rotenone increased Mitopy1 fluorescence of \(-10\%\) in midbrain mitochondria vs vehicle, and progressively for the duration of measurement), than cortical neurons. The selective mtDNA damage in midbrain could serve as a molecular marker of vulnerable nigral neurons in PD. Oxidative damage to cell macromolecules in human PD and the rotenone model have been recently reviewed (Sanders et al., 2014).

- In SK-N-MC human neuroblastoma cells, rotenone (10 nM - 1 \( \mu \)M, 48 hr) caused dose-dependent ATP depletion (~35% reduction by 100 nM rotenone vs Ctr), oxidative damage (100% increase of carbonyls levels upon 100 nM rotenone), and death (100 nM rotenone after 48 hr caused 1.1 AU (arbitrary units) increase of cell death vs untreated – 0.00 AU -). α-
Tocopherol pre-treatment (62.5 or 125 μM 24 hr before rotenone (10 nm)) attenuated rotenone toxicity (Sherer et al., 2003).

MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine) or MPP+ (1-methyl-4-phenyl-pyridinium ion)

- MPTP converted into MPP+ inhibits mitochondrial CI activity, resulting in excessive intracellular ROS production followed by further mitochondrial dysfunction leading to mitochondrial-dependent apoptosis. Lutein, a carotenoid of xanthophyll family (antioxidant) reversed MPTP-induced mitochondrial dysfunction, oxidative stress, apoptotic cell death and motor abnormalities. These results revealed that antioxidant protected DA neurons and diminished mitochondrial dysfunction and apoptotic death (Nataraj et al., 2015).

- Antioxidant (salidroside; Sal) pre-treatment protected DA neurons against MPTP/MPP+ induced toxicity in a dose-dependent manner by: (1) reducing the production of ROS, (2) regulating the ratio of Bcl-2/Bax, (3) decreasing cytochrome-c and Smac release, and inhibiting caspase-3, caspase-6, and caspase-9 activation, which are known to trigger apoptosis following mitochondrial dysfunction. Sal acted as an effective neuroprotective agent through modulation of the ROS-induced mitochondrial dysfunction in vitro and in vivo (Wang S. et al., 2015).

- In an in vitro study, MPP+ (1 mM, 24 hr) was found to elicit production of ROS (by 2 fold vs Ctr) and reduce by 50% SOD (by about 50%) and catalase (by about 65%) activity in SH-SY5Y human neuroblastoma cells. Pre-treatment with the antioxidant astaxanthin (AST; 50 μM, 24 hr) inhibited MPP+-induced production of ROS and attenuated both SOD and catalase activity decrease. Furthermore, MPP+ (1 mM, 48 hr) increased caspase-3 activity to 243% of the Ctr and also increased cleaved caspase-3 in the cells (qualitative). Addition of 50 μM AST attenuated MPP+-induced caspase-3 activation (57% suppression). MPP induced also a 70% reduction of Δψm and cytochrome c release (qualitative), while AST prevented both these effects. The protective effects of AST on MPP+ induced mitochondrial dysfunction was due to its anti-oxidative properties and anti-apoptotic activity via induction of expression of SOD and catalase (as shown above) and regulating the expression of Bcl-2 and Bax (Bax/Bcl-2 ratio increased to 1.6-fold vs Ctr upon treatment with MPP+, while AST prevented the MPP+-induced increase of the Bax/Bcl-2 ratio). These results were confirmed by in vivo studies (Lee et al., 2011).

- DA neurons in primary mesencephalic cultures treated with MPP+ (100 μM, for 48 hr) underwent reduction of cell viability (~55% MTT reduction), LDH release (~90%), about 60% reduction of TH+ cells, disruption of Δψm (~45% decline) and ROS production (~60% increase), upregulation of Nox2 (~45%) and Nox4 (~60%), while promoting a decrease of both SOD (~45%) and GSH activity (~85%). Additionally, MPP induced apoptosis via mitochondrial dysfunction, as shown by induction of cytochrome c (~55%), cleaved-caspase-3 (~75%), upregulation of Bax expression (~55%), and downregulation of Bcl2 (~60%). Liiuwei dihuang (LWDH), a widely used traditional Chinese medicine (TCM), has antioxidant characteristics. LWDH- WH, derivative of LWDH (0.01-10 μg/ml, added 1 hr prior to MPP+ addition) reduced oxidative damage via increasing antioxidant defence (SOD, GSH), decreasing ROS production, and down-regulating NADPH oxidases (Nox2 and Nox4). LWDH-WH also inhibited neuronal apoptosis by increasing anti-apoptotic protein Bcl-2 expression, and down-regulating apoptotic signalling (Bax, cytochrome c, cleaved-caspase-3) in MPP+-treated neurons. All these protective effects were induced in a dose-dependent manner (Tseng et al., 2014).

- PC12 cells treated with MPP+ (500 μM, for 24 hr) underwent reduction of viability (~55% MTT reduction), oxidative stress (~160% increase in ROS production) and downregulation of heme oxygenase-1 expression (~2 folds). Pre-treatment with edaravone, a novel free radical scavenger, (25, 50, 75, 100 μM, for 1 h prior MPP+ treatment) protected PC12 cells against MPP+-cytotoxicity via inhibiting oxidative stress and up-regulating heme oxygenase-1 expression in a dose-dependent manner (Cheng et al., 2014).
The protective effects of antioxidant, apigenin (AP), naturally occurring plant flavonoids were observed on the MPP+-induced cytotoxicity in cultured rat adrenal pheochromocytoma cells (PC12 cells). The PC12 cells were pre-treated with various concentrations of the test compound for 4 h, followed by the challenge with 1,000 µM MPP+ for 48 h. Pre-treatment with AP (3 - 6 - 12 µM) before MPP+ significantly reduced the level of intracellular ROS and elevated Δψm in the MPP+-treated PC12 cells. In addition, AP markedly suppressed the increased rate of apoptosis and the reduced Bcl-2/Bax ratio induced by MPP+ in the PC12 cells. The findings demonstrated that AP exerts neuroprotective effects against MPP+-induced neurotoxicity in PC12 cells, at least in part, through the inhibition of oxidative damage and the suppression of apoptosis through the mitochondrial pathway (Liu et al., 2015).

Brain mitochondria isolated from ventral midbrain of mitochondrial matrix protein cyclophilin D (CYPD) knockout mice were significantly less sensitive to acute MPP+ (20 µM) -induced effects. CYPD ablation attenuated in vitro Ca2+-induced mitochondrial dysfunction and ROS generation upon Ca2+ loading, both in the absence and in the presence of MPP+, compared to wild-type mice. CYPD ablation conferred a protection to mitochondrial functions upon in vivo treatment with MPTP.

Ventral midbrain mitochondria (that constitutes ≤ 5% of SNpc DA neurons) isolated from brains of wild type (wt) mice acutely treated with MPTP (single MPTP 20 mg/kg injection, analysis done after 4 hr), as compared with saline-treated mice, showed a reduction of CI (by 53%), a reduced rate of phosphorylating respiration (by 38%), a reduced respiratory control index (by 37%), and a decreased ADP/O ratio (by 18%).

Ventral midbrain mitochondria isolated from brains of CYPD knockout mice acutely treated with MPTP, as compared with MPTP-treated wt mice, exhibited higher activity of CI (~80%, vs 53% wt), higher rate of phosphorylating respiration (~82%, vs 62% wt), a better respiratory control index (~79%, vs 63% wt), and a higher ADP/O ratio (~90% vs 82% wt) (Thomas et al., 2012).

CYP plays as a regulatory component of a calcium-dependent permeability transition pores (PTP), and the data suggest that PTP is involved in MPP+-induced mitochondrial damage. Under oxidative stress, the prolonged opening of the PTP results in calcium overload and with time mitochondrial dysfunction as they get de-energized, depolarized, triggering apoptotic or necrotic cell death (Bernardi, 1999).

There are many other studies showing that MPP+ induces NADH-dependent SOD formation and enhances NADH-dependent lipid peroxidation in submitochondrial particles, confirming that oxidative stress is induced by MPP+ (e.g. Takeshige, 1994; Ramsay and Singer, 1992).

Based on the human post mortem studies of PD brains it is well established that oxidative stress and mitochondrial dysfunction accompany the pathophysiology of PD (e.g. Dias et al., 2013; Zhu and Chu, 2010; Hartman et al., 2004; Fujita et al., 2014).

Examples of human data confirming the presence oxidative stress and mitochondrial dysfunction in PD post mortem brains:

- A significant decrease in CI activity has been identified in a large study of post-mortem PD brains, specifically in substantia nigra compared with age matched controls. In idiopathic PD all 10 patients studied had significant reductions of CI activity (Parker et al., 1989). It is hypothesize that the CI dysfunction may have an etiological role in the pathogenesis of PD (Greenamyre et al., 2001; Sherer et al., 2003, Schapira et al., 1989).

- The structure and function of mitochondrial respiratory-chain enzyme proteins were studied post-mortem in the substantia nigra of nine patients with PD and nine matched controls. Total protein and mitochondrial mass were similar in the two groups. CI and NADH cytochrome c reductase activities were significantly reduced, whereas succinate cytochrome c reductase activity was normal. These results indicated a specific defect of CI activity in the substantia nigra of patients with PD (Schapira et al., 1990).

- Post mortem human studies show that CI deficiency in PD is anatomically specific for the substantia nigra, and they are not present in another neurodegenerative disorder involving...
the substantia nigra. These results suggest that CI deficiency may be the underlying cause of DA cell death in PD (Schapira et al., 1990; Schapira, 1994).

- The mitochondrial respiratory chain function was studied in various brain regions as well as in skeletal muscle and in blood platelets from patients with idiopathic PD and from matched controls. The evidence suggests that the CI deficiency in PD is limited to the brain and that this defect is specific for the substantia nigra (Mann et al., 1992).

- Immunoblotting studies on mitochondria prepared from the striata of patients who died of PD were performed using specific antisera against Complexes I, III and IV. In 4 out of 5 patients with PD, the 30-, 25- and 24-kDa subunits of CI were moderately to markedly decreased. No clear difference was noted in immunoblotting studies on subunits of Complexes III and IV between the control and PD. The authors claim that deficiencies in CI subunits seem to be one of the most important clues to elucidate pathogenesis of PD (Mizuno et al., 1989).

- Redox markers have been found unchanged in PD patient-derived vs Ctr-derived fibroblasts at baseline. Basal mitochondrial respiration and glycolytic capacity resulted similar at baseline between PD and Ctr fibroblasts, while rotenone-sensitive respiration (analysed by using 0.5 μM rotenone) resulted lower in PD fibroblasts vs Ctr (174.74 ± 48.71 vs 264.68 ± 114.84) (Ambrosi et al., 2014).

- Augmented oxidative metabolism has been detected in PD brains by magnetic resonance studies, in conjunction with energy unbalance. Decreased glucose consumption (22% mean reduction), likely reflecting a decrease in neuronal activity, has been reported in the nigrostriatal system of PD patients (Pielt et al., 1996). These symptoms were hypothesized to be indicative of mitochondrial dysfunction as early markers, present in the brain of patients with PD even in the absence of overt clinical manifestations (Rango et al., 2006). In particular, by using high temporal and spatial resolution 31P magnetic resonance spectroscopy (31P MRS) technique authors studied mitochondrial function by observing high-energy phosphates (HEPs) and intracellular pH in the visual cortex of 20 PD patients and 20 normal subjects at rest, during, and after visual activation. In normal subjects, HEPs remained unchanged during activation, but rose significantly (by 16%) during recovery, and pH increased during visual activation with a slow return to rest values. In PD patients, HEPs were within the normal range at rest and did not change during activation, but fell significantly (by 36%) in the recovery period; pH did not reveal a homogeneous pattern with a wide spread of values. Energy unbalance under increased oxidative metabolism requirements, that is, the post-activation phase, discloses a mitochondrial dysfunction that is present in the brain of patients with PD even in the absence of overt clinical manifestations (Rango et al., 2006).

There are many other studies providing evidence that oxidative stress and mitochondrial dysfunction play an important role in PD pathophysiology (see indirect KER Mitochondrial dysfunction induced DA neuronal cell death of nigrostriatal pathway).

2.3 Uncertainties or inconsistencies

- Some studies suggest that rotenone may have effects other than CI inhibition, and it has been claimed that rotenone induces microtubule disruption, rather than ETC CI inhibition (Feng, 2006; Ren et al., 2005).

- Some studies suggested that there was no evidence for significant change in mitochondrial CI function in PD patients' brains (Jenner et al., 1992).

- It is still unclear whether the site of superoxide production in CI inhibited mitochondria is CI itself or not (Singer and Ramsay, 1994).

2.4 Quantitative Understanding of the Linkage

Based on the available data, the threshold effect seen in brain mitochondria indicates that modest CI inhibition (∼25-50% decrease in activity) may not directly impact ATP levels or Δψm. Indeed, low levels of CI inhibition produces an oxidative stress without any significant changes in mitochondrial respiration (Betarbet et al., 2000; Greenamyre et al., 2001) or causes not significant changes in ATP levels (Sherer et al., 2003).
In particular, in rotenone-infused animals (2.0 mg/kg per day for 2 days), \( [3^H] \) dihydrorotenone binding to CI in brain was reduced by about 73%. Based on this degree of binding inhibition, the rotenone concentration in brain was estimated to be between 20–30 nM. Complexes II and IV were unchanged by rotenone infusion (Betarbet et al., 2000).

However, such defects have long-term deleterious effects. It is well documented that there is a site of electron leak upstream of the rotenone binding site in CI (i.e., on the ‘NADH side’ of the complex) (Hensley et al., 1998) leading to the superoxide (O\(_2^−\)) and followed up by H\(_2\)O\(_2\) production by CI (Greenamyre et al., 2001). The relative role of each ETC complex in forming superoxide differs by tissue; however CI is a major source of O\(_2^−\) in the brain (Halliwell, 2007).

Thus, a low inhibition of CI activity that is insufficient to affect cell respiration may lead to mitochondrial damage and chronic up-regulation of ROS production. Therefore, it is suggested that rotenone that binds to CI with an affinity of 10−20 nM induces toxicity not by bioenergetics effects but rather via accumulative oxidative stress. Sustained oxidative stress leads to decrease levels of reduced glutathione; activation of superoxide dismutase (SOD) (scavenger of O\(_2^−\)), catalase and indeed, treatments with antioxidants reduce the oxidative stress-induced damage. Such data are abounded in the existing literature based both on in vivo and in vitro studies and a few examples are described in the

**2.5. Empirical support for linkage**

The selective CI defects (other complexes were unaffected) (Schapira et al., 1990a) and induced mitochondrial damage followed by oxidative stress is also described in PD patients brains as documented by: (a) reduced glutathione levels (Jenner et al., 1992); (b) increased content of 8-oxodeoxyguanine, a marker of oxidatively damaged nucleic acids (Alam et al., 1997; Mecocci et al., 1993); (c) increased level of malondialdehyde (marker of lipid peroxidation) (Navarro et al., 2009); (d) increased cholesterol lipid hydroperoxide (Dexter et al., 1994); (e) increased protein oxidation measured e.g. by elevated levels of methionine sulfoxide formation or protein carbonyl content (Alam et al., 1997). These studies in human brain present a semiquantitative evaluation of the oxidative stress, as there is no data showing KER between the various degrees of CI inhibition and mitochondrial damage (ROS production) and the parameters described above. However, these studies clearly confirmed that oxidative stress in PD patient brain is increased as shown by the measured biomarkers (Sanders and Greenamyre, 2013).

In *in vitro* and *in vivo* animal studies there are some data showing the quantitative relationship between the oxidative stress produced by inhibition of CI and mitochondrial damage measured by the same assays, as described in human studies, and a few examples of such experiments are discussed below.

The quantitative evaluation of the causative relationship between the CI inhibition (KE up) induced by rotenone (4 hr exposure) and mitochondrial dysfunction (KE down) measured in human-chimpanzee isolated mitochondria (xenomitochondrial cybrids; HXC) by a decreased cell respiration and Δψm, increased ROS production and lipid peroxidation showed linear, time- and concentration-dependent effects (below Fig.8 from Barrientos and Moraes, 1999).
Fig. 8. A dose- and time-dependent effect of CI inhibition by rotenone on (A) reactive oxygen species production (ROS), (B) Lipid peroxidation and (C) mitochondrial membrane potential (Δψm) studied in the human osteosarcoma-derived cell line (143B) or using a genetic model (40% CI inhibited in HXC lines) (for further information see Barrientos and Moraes, 1999, Fig. 5).

The endogenous respiration was inhibited in a dose-dependent manner but showed different inhibition kinetics. Only when CI was inhibited by 35-40% (< 5 nM rotenone), cell respiration started decreasing (a threshold for inhibition for cell respiration triggered by rotenone). Between 40 and 60% of CI inhibition (5-10 nM), cell respiration decreased linearly until 30% of the normal rate. Increasing concentrations of rotenone produced further but slower decrease in CI activity and cell respiration. 100% CI inhibition was achieved with 100 nM rotenone but the cells still maintained a cell respiration rate (through complex II), approximately 20% and the rate of ROS production increased by a maximum of 20-25% (4 hr treatment). ROS production was saturated at 100 nM rotenone but an initial effect was observed already at 1-5 nM (Barrientos and Moraes, 1999). Inhibition of CI activity...
triggered decrease of cell respiration by different concentrations of rotenone and resulted in mitochondrial damage measured not only by ROS production, but also by lipid peroxidation and decreased Δψm. Inhibition of CI by 25, 50, 75 and 100% decreased cell respiration by 5, 20, 53, 81%, increased ROS production by 48, 81, 157, 216%, increased lipid peroxidation by 8, 27, 45, 55% and decreased Δψm by 6, 13, 20, and 37% respectively (approximately).

Similar studies were also performed using different types of neuronal cells.

Hoglinger and colleagues, by using DA neurones derived from the rat (embryonic day 15.5) ventral mesencephalon, showed that CI inhibition by rotenone at 30 nM, (or MPP+ 3 µM) for 24 hr decreased ATP levels (by > 80%) within the first 6 hr, and neuronal cell death within 24 hr. When residual ATP levels remained above 20%, there was no or little neuronal loss, suggesting that 20% of normal ATP level was the minimum compatible with neuronal survival. Rotenone (and MPP+) increased ROS (≥40% over control levels) already at low concentrations that were subtoxic or only moderately toxic (i.e., 10-30 nM for rotenone, 10-30 µM for MPP+) (Fig. 9) (Hoglinger et al., 2003).

Shamoto-Nagai and colleagues showed that 25 or 50 nM rotenone decreased ATP levels over time. In particular, the intracellular ATP level was reduced to 18.0% and 19.6% of control after 44 hr of treatment with 25 and 50 nM of rotenone, respectively, and thereafter the decreased level was sustained (Fig. 10, left) (Shamoto-Nagai et al., 2003). Also, The production of ROS-RNS increased 6 hr after the rotenone treatment, and the increase was about 1.5-fold of the basal value. With treatment with the higher (50 nM) concentration of rotenone, DCF production level was restored to the basal level after 48 hr, whereas, at the lower concentration (25 nM), DCF production increased again at 48 hr and then declined to the basal value after 90 hr (Fig. 10, right) (Shamoto-Nagai et al., 2003).
Fig. 10. Effect of rotenone on ATP level (left) and on ROS and RNS production (right) in SH-SY5Y cells. SH-SY5Y cells were treated with 25 nM (circles) or 50 nM (triangles) of rotenone. * indicates significant difference from control (P < .05) (from Shamoto-Nagai et al., 2003, Figs. 2, 3).

Human neuroblastoma cell line (SK-N-MC) exposed to 5 nM rotenone chronically, for 4 weeks caused reduction in GSH by 44%, GSSG by 40%. These effects were not observed after two weeks of exposure. Total cellular GSH levels were reduced after 4 weeks of exposure by 50% (Sherer et al., 2002). Similarly, in the same study, 1-2 weeks of treatment did not alter protein carbonyl levels (oxidative protein damage) but exposure for 3-4 weeks caused a large increase in carbonyls in the insoluble fraction by approximately 223% of control. Systemic in vivo rotenone infusion (up to 5 weeks, 3.0 mg/kg/day) modestly elevated soluble protein carbonyls in the rat cortex by approximately 19%, in the striatum by 27% and the largest elevation occurred in the DA neurons of midbrain, around 41% (no effect in cerebellum or hippocampus) (Sherer et al., 2003).

The prolonged treatment with rotenone (3-4 weeks, not 1-2 weeks) caused also a marked increase in 8-oxo-dG immune-reactivity (i.e., oxidative DNA damage) and redistribution of cytochrome c (Sherer et al., 2002).

The same group showed that exposure of SK-N-MC cells for 6-8 hr to low concentrations of rotenone (100 pM, 1 nM, 10 nM and 100 nM) produced a concentration-dependent decrease in ATP levels by 0, 2.5, 10, and 32.2 % respectively (Sherer et al., 2003).

The oxidative stress (mitochondrial damage) induced by rotenone exposure was confirmed in ex-vivo studies using brain sections at the level of the substantia nigra that were treated with 50 nM rotenone over 1 week. A significant increase of protein carbonyls (indicative of oxidative damage to proteins; biomarkers of oxidative stress) was observed (~ 25%) when compared to the untreated slices. Exposure to 100 μM tocopherol, antioxidant (vitamin E) significantly protected the neurons from the oxidative damage induced by 50 nM rotenone over 1 week (~ 25%), as shown by lower protein carbonyl levels (~ 3%), with very similar effects observed with 20 nM rotenone over 2 weeks (Testa et al., 2005).

The same assays for mitochondrial dysfunction evaluation after exposure to rotenone, MPTP or other chemicals were used through a range of different studies (Sherer et al., 2003, Betarbet et al., 2000) and the role of CI inhibition in PD is discussed in many published reviews (Sanders and Greenamyre, 2013, Greenamyre et al., 2001, Schapira et al., 1990a and 1990b).

Conclusions: It is well documented in human PD brain studies as well as in vivo and in vitro existing data that CI inhibition induces mitochondrial dysfunction as shown by measuring the decreased cellular respiration and induced oxidative damage to protein, lipids and nucleic acids, as well as compromised function of anitoxidant defense mechanisms (e.g. decreased levels of reduced glutathione). As discussed above, oxidative damage is largely reversed by antioxidants treatments. These data are largely semi quantitative only, as the full dose- and time response curves are available. They indicate that low levels of CI inhibition for long periods of time (4-5 weeks) mostly increase ROS production, having negative effects on DA neurons in SNpc, which seem to be affected more than other neuronal cell types in other brain structures (reviews e.g. by Sanders and Greenamyre, 2013; Greenamyre et al., 2001, Schapira et al., 1990a and 1990b etc.).

2.5 Evidence Supporting Taxonomic Applicability

Mitochondrial CI in eukaryotes has highly conserved subunit composition based on protein databases (Cardol, 2011).

The characterization of induced mitochondrial dysfunction phenotypes in zebrafish was studied in the presence of CI and CII inhibitors (Pinho et al., 2013).

Exposure of Caenorhabditis elegans (C. elegans) to rotenone, reduced bioluminescence (an assay for mitochondrial dysfunction) after both relatively short (2 hr) and longer exposures (24 hr) to a range of concentrations. A sharp decline in bioluminescence (maximal inhibition) relative to controls occurred at the lowest rotenone concentration of 2.5 μM. This decline in bioluminescence was consistent with reduced cellular ATP (Lagido et al., 2015).
The results obtained from *C. elegans* exposed to rotenone suggested that chronic exposure to low concentration (2 or 4 μM) caused mitochondrial damage through persistent suppression of mitochondrial biogenesis and mitochondrial gene expression leading to mitochondrial dysfunction that contributed to DA neuron degeneration (Zhou et al., 2013).

*Drosophila melanogaster* has been proven suitable to study signaling pathways implicated in the regulation of mitochondrial function and integrity, such as the PINK1/parkin pathway (controlling mitochondrial integrity and maintenance), DJ-1 and Omi/HtrA2 genes (associated with the regulation of mitochondrial functionality). Notably, PINK1, PARKIN, and DJ-1 genes are associated with recessive forms of PD (Guo, 2012). *Drosophila* flies lacking DJ-1 result to be viable, but show an increased sensitivity to oxidative stress induced upon rotenone or Paraquat (an herbicide inducer of CI-dependent ROS) feeding (Menzies et al. 2005; Meulener et al. 2005; Meulener et al. 2006). Moreover, it has been reported in *Drosophila* that inhibition of CI by mean of sublethal chronic exposure to rotenone (<750 μM) via the feeding medium caused a selective loss of DA neurons in all of the brain regions and locomotor impairments, while L-dopa (3,4-dihydroxy-L-phenylalanine) rescued the behavioral deficits (but not neuronal death) (Coulom and Birman, 2004).

MPTP causes Parkinsonism in primates including humans. However, rodents (rats) are much less susceptible to MPTP+ but are fully susceptible to MPP+ (due to the differences in toxicokinetics). In all species, CI inhibition leads to mitochondrial dysfunction. Mitochondrial dysfunction is an universal event occurring in cells of any species (Farooqui and Farooqui, 2012).
References


PPPs, Parkinson’s disease and childhood leukaemia


Third KER: Mitochondrial dysfunction results in an impaired proteostasis

3.1 How this Key Event Relationship work

In any cell type, including neurons, the protein homeostasis (proteostasis) plays a key role in cellular functions. There are two major systems involved in the removal of damaged cellular structures (e.g. defective mitochondria) and misfolded or damaged proteins, the ubiquitin-proteasome system (UPS) and the autophagy–lysosome pathway (ALP). These processes are highly energy demanding and highly susceptible to oxidative stress. Upon mitochondrial dysfunction UPS and ALP functions are compromised resulting in increased protein aggregation and impaired intracellular protein/organelles transport (e.g. Zaltieri et al., 2015; Song and Cortopassi, 2015; Fujita et al., 2014; Esteves et al., 2011; Sherer et al., 2002).

3.2 Weight of Evidence

The weight of evidence supporting the relationship between mitochondrial dysfunction and impaired proteostasis, including the impaired function of UPS and ALP that results in decreased protein degradation and increase protein aggregation is strong.

3.2.1 Biological Plausibility

The biological relationship between Mitochondrial dysfunction and Impaired proteostasis (unbalanced protein homeostasis) that involves dysregulation of proteins degradation (misfolded or damaged) as well as removal of cell organelles is partly understood. Under physiological conditions, mechanisms by which proteostasis is ensured include regulated protein translation, chaperone assisted protein folding and functional protein degradation pathways. Under oxidative stress, the proteostasis function becomes burdened with proteins modified by ROS (Powers et al., 2009; Zaltieri et al., 2015). These changed proteins can lead to further misfolding and aggregation of proteins (especially in non-dividing cells, like neurons). Particularly in DA cells, oxidative stress from dopamine metabolism and dopamine auto-oxidation may selectively increase their vulnerability to CI inhibitors (such as rotenone) and cause additional deregulation of protein degradation (Lotharius and Brundin, 2002; Esteves et al., 2011). As most oxidized proteins get degraded by UPS and ALP (McNaught and Jenner, 2001), mitochondrial dysfunction and subsequent deregulation of proteostasis play a pivotal role in the pathogenesis of PD (Dagda et al., 2013; Pan et al., 2008; Fornai et al., 2005; Sherer et al., 2002).

It is also well documented that increased oxidative stress changes the protein degradation machinery and leads to a reduction of proteasome activity (Lin and Beal, 2006; Schapira, 2006).

3.2.2 Empirical support for linkage

Based on the existing in vitro and in vivo data it is suggested that mitochondrial dysfunction impairs protein homeostasis through oxidative and nitrosative stress resulting in protein aggregation, disruption of microtubule assembly and damaged intracellular transport of proteins and cell organelles.

Mitochondrial dysfunction by rotenone or MPP+ reduces UPS activity

- Mitochondrial dysfunction induced by systemic and chronic CI inhibition by rotenone, results in a selective inhibition of proteasomal function in the midbrain (not in cortical or striatal homogenates) of rats that had lost the TH-positive terminals in the striatum. Initially, proteasomal activity showed an acute increase prior to a decrease by 16-31 %, during chronic rotenone exposure (3.0 mg/kg/day, through osmotic pump during 5 weeks). In the same animals a significant and selective increase in ubiquitinated proteins (~ 25%) was observed in the ventral midbrain of lesioned rats, indicating an increase in the proteins levels that have been marked for degradation by UPS. These results were confirmed immunocyto-chemically, pointing out that ubiquitin levels were elevated selectively in DA neurons present in SNpc (Betarbet et al., 2006).

- Nigral neurons in chronically rotenone-treated rats (up to 5 weeks, infusion of rotenone at 2.5 mg/kg/day) accumulate fibrillar cytoplasmic inclusions that contain ubiquitin and α-synuclein
(the main protein of Lewy bodies observed in PD) (qualitative data, obtained by immunoelectron microscopy) (Betarbet et al., 2000).

- Inhibition of proteasomal function was also observed in in vitro systems using SK-N-MC human neuroblastoma. Exposure to 5 nM rotenone, for up to 4 weeks caused 60% increase in the levels of ubiquinated proteins, suggesting that chronic exposure to rotenone increased the level of misfolded or oxidized proteins targeted for degradation by UPS (Betarbet et al., 2006).

- To determine whether rotenone-induced proteasomal inhibition was due to CI inhibition or direct effects of rotenone on the UPS, proteasomal activity was determined in SKN-MC cells expressing the rotenone-insensitive single-subunit NADH dehydrogenase of *Saccharomyces cerevisiae* (ND11), which acts as a "replacement" for the entire CI in mammalian cells (Bai et al., 2001; Seo et al., 2000, 2002). The obtained results confirmed that rotenone-induced proteasomal dysfunction is due to CI inhibition and not to direct effects of rotenone on proteasomal function (Betarbet et al., 2006). In the same study the decreased proteasomal activity and an accumulation of ubiquitinated proteins was completely prevented by continuous treatment with α-tocopherol (62.5 μM added 1 week prior to and continuously thereafter along with 5 nM rotenone) (qualitative data), confirming that oxidative damage played a major role in rotenone-induced proteasomal dysfunction rather than bioenergetic defects. Indeed, chronic, low levels of rotenone exposure did not changed significantly ATP levels (111.5 ± 1.5% of control), but produced ROS (not shown in this study). Similar results were published by Shamoto-Nagai's group (Shamoto-Nagai et al. 2003).

- Rotenone significantly lowered UPS activity in a concentration dependent manner in HEK (human embryonic kidney cells) and SK-N-MC human neuroblastoma cells even after 24 h exposure to doses as low as 10 nM. It caused a reduction in the 20S proteasome activity (by 5-25%) and of the 20S proteasome subunit (by 20-60%) (as shown by increase of GFP-U fluorescence) (Chou et al., 2010). Similar results were obtained using other pesticides that inhibit CI, including pyridaben and fenazaquin (Wang et al., 2006). This effect was mediated by oxidative stress as anti-oxidants, such as butylated-hydroxy toluene (BHT), and catalase attenuated rotenone-induced UPS inhibition. Additionally, nitric oxide (NO) and peroxinitrite contributed to this effect as well, since neuronal nitric oxide synthase (nNOS) inhibitor (LNMMA) attenuated rotenone-induced proteasome inhibition by 20% (Chou et al., 2010) indicating that both oxidative and nitrative stress can directly inhibit the proteasome activity through increased degradation of proteasome subunits. The same mechanisms of proteasome inhibition were suggested by many other studies (e.g. Szweda et al, 2002; Osna et al., 2004; Shamoto-Nagai et al., 2003).

- CI inhibition-induced proteasomal dysfunction has been reported in ventral mesencephalic cultures following acute rotenone or MPP+ exposure (Hoglinger et al., 2003). In DA neurones derived from rat (embryonic day 15.5) ventral mesencephalon, it has been shown that proteasome inhibition (by 100 nM epoxomicin) exacerbated the neurotoxicity of CI inhibitors (by mean of rotenone 30 nM, or MPP+ 3 μM, for 24 hr). All three proteasomal peptidase activities (i.e., chymotrypsin (CT)-like, trypsin (T)-like, and peptidylglutamyl-peptide hydrolase (PGPH) activity) significantly decreased in cultures upon 6 hr treatment with 30 nM rotenone (by 50+-60%) or 30 μM MPP+ (by 25-30%) (Hoglinger et al., 2003).

- CI inhibition-induced proteasomal dysfunction has been reported in human SH-SY5Y neuroblastoma cells following acute rotenone exposure (Shamoto-Nagai et al., 2003). After 96 hr of incubation with 25 or 50 nM rotenone, the activity was reduced respectively to 28.7% and 21.9% of control, and adding ATP did not increase the activity. After 120 hr, the activity was virtually undetectable (with or without added ATP). On the contrary, the levels of the proteins composing proteasome did not change with rotenone treatment (Shamoto-Nagai et al., 2003).

- The ability of rotenone to cause proteasome inhibition via disruption of microtubules (MT) assembly has been also documented. In human embryonic kidney (HEK) and neuroblastoma SK-N-MC cells rotenone (10-100-100 nM, 24 hr) was found to inhibit 26S UPS activity (by 25%, at 10 nM) (Chou et al., 2010). Rotenone was found to interfere with MT assembly at concentrations as low as 10 nM, providing evidence that there could be additional...
mechanisms implicated in the rotenone induced UPS inhibition, possibly mediated by nitric oxide (NO). In the same study, nocodazole, a MT disrupter (positive control), strongly inhibited the UPS activity (e.g., 10 μM nocodazole caused ~80% decrease of 26S UPS activity) (Chou et al., 2010).

- Oxidative stress triggered by the MPP+ inhibited CI (1 mM, for 2-6-24 hr) led to a decrease in proteolytic activity, as shown in NT2 human teratocarcinoma cells containing mitochondrial DNA (p+) and NT2 cells depleted of mtDNA (p0) (Domingues et al., 2008). In particular, MPP+ (1 mM, 2 hr) elevated ubiquitylated protein content (by ~3 fold compared to untreated Ctr), and after 24 hr induced a significant decrease of chymotrypsin-like activity (by ~30%) and peptidyl-glutamy peptide hydrolytic-like activity (by ~75%) compared to untreated cells (Domingues et al., 2008).

- Mice following continuous MPTP infusion (1-5-30 mg/kg daily) exhibited inhibition of the UPS (respectively by 40-50-60%) and increased inclusions of ubiquitin and α-synuclein in the neurons in the substantia nigra (Fornai et al., 2005).

- A mouse model of mitochondrial CI deficiency (Ndufs4-/- mice) showed an impaired 20S proteasomal activity (by ~50%), leading to increased ubiquitin protein levels (by ~40%) in the substantia nigra (not in cortex and hippocampus), increased of ubiquitin+/TH+ neurons (by ~2 fold, compared to WT mice), and increased ubiquitinated neurofilaments in the midbrain (values of 1.2 - 2.8 vs 1.0 in WT) (Song and Cortopassi, 2015).

**Human studies**

- PD patients appear to have an impaired UPS. The presence of aggregated, poly-ubiquitinated proteins in Lewy Bodies indicates that proteolytic dysfunction and proteo-toxicity are critical steps in the pathogenic cascade of PD (Betarbet et al., 2005). In this regard, impairment of proteasomal activity and reduced expression of proteasomal subunits have been reported selectively in substantia nigra of sporadic PD post-mortem brains (McNaught et al., 2003; McNaught and Jenner, 2001). In particular, in PD, there was a 40.2% reduction in the amount of α-subunits in the SNc. On the opposite α-subunits levels were increased by 9.2% in the cerebral cortex and by 29.1% in the striatum in PD compared to Ctr (McNaught et al., 2003).

- Chymotrypsin-like, trypsin-like, and peptidyl glutamyl-peptide hydrolytic (PGPH) 20/26S proteasomal activities were significantly decreased in the substantia nigra (by 43.9%, 45.9%, and 44.6% respectively) (not in the cortex or striatum) in PD patients. At the same time, in PD there was a marked increase in the levels of PA700 subunits (the 19S regulatory complex of the 26S proteasome) in the frontal cortex and/or the striatum compared to controls, while in the SNpc PA700 subunits resulted decreased up 33%, whereas levels of nigral PA28 were almost undetectable in both normal and PD subjects (McNaught et al., 2003).

- Steady-state levels of soluble AF-6 (modulates parkin ubiquitin-ligase activity) have been found significantly lower in the caudate/putamen (~66% lower) as well as in the SN of PD patients (~66% lower). AF-6 was also detected in ~25% of mature Lewy bodies and in occasional Lewy neurites in the substantia nigra of the four PD brains analysed, and may contribute to the disruption of mitochondrial homeostasis (Haskin et al. 2013).

- HDAC6 has recently been identified by immunocytochemistry as a constituent in Lewy bodies of PD and dementia with LBs (DLB), as well as in glial cytoplasmic inclusions in multiple system atrophy (MSA) (Kawaguchi et al. 2003; Miki et al. 2011; Chiba et al. 2012). HDAC6 is considered a sensor of proteasomal inhibition and a cellular stress surveillance factor. Upon proteasomal inhibition, HDAC6 is relocated and recruited to polyubiquitin-positive aggresomes. HDAC6 inhibition elicits tubulin acetylation and restores microtubule (MT)-dependent transport mechanisms in neurons (Richter-Landsberg and Leyk, 2013).

- Basal activity of 20S proteasome was significantly reduced (by ~33%) in PD as compared to control fibroblasts. Higher accumulation of ubiquitinated proteins (by ~2 fold), representative of impaired 26S proteasome function, were found in PD as compared to Ctr cells at baseline. In the presence of rotenone (20 and 500 μM, 6 hr) PD-derived fibroblasts showed a higher induction of 20S proteasome activity (~15% higher) as compared to Ctr fibroblasts, with no
Mitochondrial dysfunction by rotenone or MPP+ deregulates ALP activity

- Exposure to rotenone (10 μM, 24 hr) induced neurotoxicity in human neuronal SH-SY5Y cells (number of dead cells was 8 folds higher than Ctr group) and pre-treatment with rapamycin (3 μM, 48 hrs) (strong inducers of autophagy) robustly protected against rotenone-mediated toxicity (number of dead cells was 3 folds higher than Ctr group) and this was due to the induction of autophagy. Indeed, suppression of autophagy (by silencing of Atg5) blocked the neuroprotection of rapamycin (Pan et al., 2009).

- Similar results were produced using kaempferol (6 μM, 1 hr prior addition of rotenone) and rotenone (50 nM, max up to 24 hr) on SH-SY5Y cells. Kaempferol was found to counteract rotenone-induced effects (see KERZ) and these protective effects were related to induction of autophagy (6 hr kaempferol induced LC3-II formation, as shown by Western blot) (Filomeni et al., 2012).

- Treatment of SH-SY5Y cells with high doses of rotenone (500 nM, 48 hr) induced Atg5–Atg12 dependent autophagy, which leads to lysosomal dysfunction, increased p62 levels, and an aberrant accumulation of α-synuclein (Pan et al., 2009; Dadakhuaev et al., 2010). In particular, in α-synuclein expressing SH-SY5Y cells Atg5–Atg12 were increased by addition of rotenone and rapamycin (100 nM, 48 hr). Co-treatment with rotenone and autophagy inhibitors (e.g., 3-MA, bafilomycin or wortmannin) similarly diminished the level of Atg5–Atg12 in α-synuclein expressing cells (western blot analyses) (Dadakhuaev et al., 2010).

- A few studies have suggested that rotenone can act as an inducer of autophagic flux. For instance, treating human embryonic kidney cells (HEK 293) and U87 glioma cells with rotenone (50 μM, for 0-72 hr) caused cell death (in HEK 293 cells, rotenone induced 30% cell death, after 72 hr; in U87 cells, 40%) by upregulating autophagy and mitophagy (as shown by increase of cells with AVOs (indicative of autophagosomes and autolysosomes, analysed by flow cytometry): by ~14% in HEK 293 cells, and by ~20% in U87 cells, as compared to untreated cells, 0%), a process that is supposed to be triggered by mitochondrial superoxide (Chen et al., 2007).

- Increased autophagic flux has been observed in SH-SY5Y cells and primary cortical neurons treated respectively with 1 μM and 250 nM of rotenone. Rotenone elicited increases in autophagy (~ 2 folds vs Ctr) and mitophagy (i.e., as shown by the percentage of GFP-LC3 puncta colocalizing with mitochondria (~ 4 folds vs Ctr), indicating a preferential increase in “mitophagosomes” relative to total autophagosomes. Additionally, rotenone induced a decrease in p62 (SQSTM1), levels (~40% decrease with 250 nM), consistent with increased autophagic flux. This effect was reversed by co-treating cells with bafilomycin A2, a specific inhibitor of vacuolar-type H(+)-ATPase, or by RNAi (knockdown of ATG7 and ATG8/LC3). The mechanism by which LC3 recognizes damaged mitochondria in rotenone-treated neurons involves, among others, the externalization of cardiolipin and recruitment of LC3 at the mitochondria initiating rotenone-induced-mitophagy and lysosomal-mediated degradation of mitochondria (Chu et al., 2013).

- In the study by Wu et al., (2015) chronically rotenone-treated rats (male Lewis rats received rotenone 1mg/kg subcutaneously twice a day for 8 weeks) had a robust loss of TH+ neurons in striatum (~50%) and in SNpc (~30%). However, in the remaining DA neurons of SNpc, cytoplasmic inclusions containing α-synuclein were observed (~7% of α-synuclein+/TH+ cells vs ~2% in Ctr), probably due to rotenone-induced decreased degradation of the autophagosomes (upregulation of LC3-II by ~30%, Beclin 1 by ~10%, and p62 by ~150%, after 24 hr rotenone) indicating decreased ALP function. Compared with the control group, the nigral DA neurons of the rotenone-treated group exhibited an increased diffuse distribution of LAMP2 (~15% vs ~25% Ctr) and cathepsin D (~22% vs ~60% Ctr) instead of punctuate pattern, indicating impaired lysosome integrity and a redistribution of cathepsin D from lysosomes to the cytosol. In parallel in vitro studies by the same group showed that PC12 cells exposed to rotenone (500 nM for 24 hr) underwent increased protein levels (but not mRNA levels) of α-synuclein (~4.5 folds vs Ctr), indicating an impairment of protein
degradation. In TEM pictures, the majority of neurons displayed mitochondrial swelling, crista fragmentation, and accumulation of double membrane structures containing damaged mitochondria, which were stalled autophagosomes (Wu et al., 2015).

- Similar results, showing impaired autophagic flux resulting in α-synuclein accumulation and the rupture of lysosomes in neuronal cell lines exposed to rotenone have been described in many other studies (e.g. Mader et al., 2012; Sarkar et al., 2014).

- Rotenone produced bidirectional effects on macroautophagy (decrease or increase). This may be attributed to differences in the dosage, the duration, and cell type which can produce variable levels of ROS and mitochondrial damage (Pan et al., 2009; Dadakhujaev et al., 2010; Chen et al., 2007; Filomeni et al., 2012; Mader et al., 2012).

- MPP+ (2.5 mM, 24 - 48 hr) increased autophagy (~14 folds increase vs Ctr, of LC3-II) and mitochondrial loss in SH-SY5Y cells (a DA neuronal cell line widely used as a cell culture model of PD) by increased MAP kinase signalling (MEK inhibition by UO126 reversed by both autophagy and mitochondrial loss elicited by MPP+) (Zhu et al., 2007).

- Another study from the same group showed that longer MPP+ treatment (250 μM, 2 weeks) induced formation of enlarged, coarse GFP-LC3 puncta, in a time- and dose-dependent manner (~1.8% of cells presenting coarse GFP-LC3 puncta, vs ~0.2% in Ctr, at 14 days with 250 μM rotenone) (Zhu et al., 2012).

- An in vitro study on MN9D cells (a fusion of embryonic ventral mesencephalic and neuroblastoma cells, used as a model of DA neurons) showed that MPP+ (50 μM, for 24 hr) blocked autophagic flux, as evidenced by increased steady-state levels of p62 (qualitative data, Western blot), increased of autophagic vacuoles numbers (~3 folds vs Ctr) along with lysosomal depletion and dysfunction presumably due to leakage of lysosomes, impaired lysosomal biogenesis, and increased proteasomal-mediated degradation of proteins (as shown by time-dependent increase of ubiquitinated proteins, by IC) (Lim et al., 2011).

- In another study human neuroblastoma BE-M17 cells were treated with MPP+ (0.25-2.5 mM, 24 hr); Lamp1 protein levels were decreased in a dose-dependent manner in MPP+-treated cells (by ~40% at 2.5 mM), without concomitant decreases in mRNA expression levels. Also, LC3-II increased in a dose-dependent manner with MPP+ treatment (~3000% increase at 2.5 mM vs Ctr), indicating lysosome depletion and autophagosome accumulation upon MPP+ treatment. These data were confirmed in vivo: lysosomal depletion and accumulation of autophagosomes (as shown by ~600% increase of LC3-II, and ~40% decrease of Lamp1, after 1 day of MPTP injection compared to saline) occurred also in MPTP-intoxicated mice (30 mg/kg/day, for 5 consecutive days) (Dehay et al., 2010).

- Other in vitro data support a negative role of MPTP on autophagic flux. Mice were i.p. injected with 2 mg/ml MPTP (30 mg/kg) for 7 days. Suppression of autophagic flux induced by MPTP (~20% reduction vs Ctr) was detrimental to neuronal survival (as shown by ~60% decrease of TH+ neurons). Treating mice with the autophagy inducer rapamycin after seven days of MPTP treatment (daily i.p. injections of 2 mg/ml MPTP (30 mg/kg) for 7 days, followed by 0.1 ml of 20 μg/ml rapamycin by i.v. for an additional 7 days), significantly increased the number of surviving dopamine neurons (~60% TH+ neurons vs ~30% with MPTP alone, as compared to Ctr 100%) and the levels of TH protein (~75% vs ~60% with MPTP alone, as compared to Ctr 100%) and decreased the levels of α-synuclein aggregates (~210% of α-synuclein protein level, vs ~300% with MPTP alone, as compared to Ctr 100%) (Liu et al., 2013).

- Treating mice with the autophagy inducer rapamycin after seven days of MPTP treatment (daily i.p. injections of 2 mg/ml MPTP (30 mg/kg) for 7 days, followed by 0.1 ml of 20 μg/ml rapamycin by i.v. for an additional 7 days), significantly increased the number of surviving dopamine neurons (~75% of TH protein level vs ~60% with MPTP alone) and decreases the levels of α-synuclein aggregates (~210% of α-synuclein protein level, vs ~300% with MPTP alone) (Liu et al., 2013).

MPP+ induced dysregulation of macroautophagy in neurons is discussed in recently published reviews (e.g. Cherra et al., 2010; Jiang et al., 2010).
The potential other mechanisms by which rotenone or MPTP induce mitochondrial dysfunction are further discussed in recent publications (e.g. Dagda et al., 2013; Esteves et al., 2011).

Human studies

- In PD patient postmortem cortical tissues, levels of oligomeric α-synuclein in SNpc (~1000% vs Ctr samples) and expression of LC3-II levels (~130% vs Ctr samples) were up-regulated (Yu et al., 2009) (for further info, see the review from Vekrellis et al., 2011).
- The pathological observations in PD autopsy brains showed that LC3-II levels were elevated in the SNpc and amygdala of PD brain samples, suggesting an increase in macroautophagy (but they did not reach statistical significance). LC3 colocalized with α-synuclein in most LBs and Lewy neurites in PD SNpc as well as in small punctate α-synuclein immunoreactive inclusions (IC images) (Alvarez-Erviti et al., 2010).
- Analogously, another study reported that brain homogenates derived from the temporal cortex of dementia with LB (DLB) patients vs non-demented controls were characterized by higher levels of both mTor (~130% vs Ctr) and p-mTor (~10 folds higher than Ctr), and levels of Atg7 (molecular initiator of autophagy) were moderately reduced in DLB cases compared to Ctr (~40% lower than Ctr). Consistent with the studies in human brains, levels of both mTor and p-mTor were increased in the membrane fractions from brains of α-synuclein tg mice compared to non tg controls (respectively, by ~250% and ~200% vs Ctr), and levels of Atg7 were reduced in α-synuclein tg brains compared to non tg controls (~75% less than Ctr) (Crews et al., 2010).
- Another study showed that post-mortem brain samples derived from PD patients, compared to age-matched controls, presented significant reductions of LAMP1, CatD, HSP73, and 20S proteasome (calculated by optic density (OD) measures) (Chu et al., 2009).

<table>
<thead>
<tr>
<th>Group</th>
<th>LAMP1 OD</th>
<th>CatD OD</th>
<th>HSP73 OD</th>
<th>20S proteasome OD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age-matched control</td>
<td>2069.10 ± 329.52</td>
<td>1809.35 ± 533.47</td>
<td>2604.92 ± 494.56</td>
<td>1660.84 ± 229.87</td>
</tr>
<tr>
<td>PD</td>
<td>1261.54 ± 107.77</td>
<td>1094.64 ± 378.10</td>
<td>1799.27 ± 376.19</td>
<td>1172.65 ± 273.28</td>
</tr>
</tbody>
</table>

These data globally indicate that the functions of both the UPS and ALP systems seem compromised in PD patients.

- HDAC6, which plays a central role in autophagy by controlling the fusion process of autophagosomes with lysosomes, has recently been identified as a constituent in Lewy bodies of PD and glial cytoplasmic inclusions of multiple system atrophy (Richter-Landsberg and Leyk, 2013).

Impaired UPS and ALP function leads to α-synuclein aggregation

α-synuclein is one of the most abundant neuronal proteins (Vekrellis et al., 2011). Several PD-related mutations and environmental toxicants cause autophagy dysfunction and lead to the accumulation of misfolded proteins in DA neurons, including α-synuclein. Both monomeric and aggregated forms of α-synuclein can be degraded by macroautophagy, whereas only wild-type α-synuclein (not Ala30Pro, Ala53Thr and Glu46Lys mutant forms) is degraded by the process of chaperone-mediated autophagy (CMA) (Vekrellis et al., 2011).

- Rotenone-induced α-synuclein aggregation has the ability to inhibit proteasome activity due to its propensity to assemble into filaments (as reviewed in Zaltieri et al., 2015). In particular, expression of α-synuclein was found to inhibit proteasome activity in SH-SYSY cells. Increased levels of GFP-CL1 band were observed in cells coexpressing GFP-CL1 and α-synuclein (~9000
By using stable PC12 cell lines expressing wild-type (WT) or A53T mutant human α-synuclein it has been shown that cells expressing mutant α-synuclein showed: (1) disruption of the ubiquitin-dependent proteolytic system, manifested by small cytoplasmic ubiquitinated aggregates and by an increase in polyubiquitinated proteins (qualitative data); (2) marked accumulation of autophagic-vesicular structures (qualitative data); (3) reduction of lysosomal hydrolysis and chymotrypsin-like proteasomal function (by ~30%, compared to WT) (Stefanis et al., 2001).

Rotenone- (or MPP+)-induced inhibition of CI results in calcium (Ca2+) release from mitochondria. Calcium rise and oxidative stress cooperatively can promote α-synuclein aggregation (Follett et al., 2013; Goodwin et al., 2013; Nath et al., 2011).

For instance, to investigate the influence of raised Ca2+ in response to plasma membrane depolarization on the aggregation of α-synuclein, HEK293T and SH-SYSY neuroblastoma cells have been used and depolarized by addition of KCl to the cell culture medium. After KCl treatment (50 mM) increase of cellular Ca2+ was observed (~90% increase 20 min after KCl treatment), leading to the formation of frequent perinuclear α-synuclein focal aggregates at 26–74 hr post-treatment (qualitative IC images). By adding TMO (a selective T-type Ca2+ channel blocker) no α-synuclein aggregates were detected (Follett et al., 2013).

Similarly, increased intracellular free Ca2+ (obtained by treating cells with either calcium ionophore or thapsigargin) induced the formation of α-synuclein aggregates in α-synuclein-GFP-transfected 1321N1 glioma cells (~65% increase compared to Ctr-untreated cells) (Nath et al., 2011).

On the other hand, α-synuclein can control mitochondrial calcium homeostasis by enhancing endoplasmic reticulum-mitochondria interactions. Silencing of endogenous α-synuclein (siRNA-α-syn) in HeLa cells was found to impair mitochondrial Ca2+ transients (~35% decrease compared to Ctr-scrambled siRNA) and morphology (Cali et al., 2012). Also, α-synuclein oligomerization exacerbates calcium dysregulation by increasing mitochondria permeability transition (Danzer et al., 2007). Therefore, it is possible that mitochondrial dysfunction-induced calcium rise precede the onset of α-synuclein accumulation leading to UPS inhibition (Chou et al., 2010).

It has been demonstrated that rotenone increased the intracellular calcium levels, triggering aggregation and phosphorylation of α-synuclein in a calcium-dependent manner. The aggregation of α-synuclein in PC12 cells following rotenone exposure was observed in a dose and time-dependent manner (1, 10 and 100 mM for 48 hrs, 3 days, 1 and 3 weeks) (~4-fold increase of α-syn with 100 mM rotenone for 48 hr, vs Ctr; and also, ~2.5 fold increase of α-syn with 1 mM rotenone for 1 week, vs Ctr) as evaluated via a variety of methods, including western blotting, immunofluorescence and electron microscopy. The observed attenuation of autophagy and α-synuclein aggregation was reversed by scavenging calcium (by using the calcium chelator BAPTA at 10 μM). Aggregated α-synuclein is typically degraded by autophagy, but rotenone impaired this process (Yuan et al., 2015).

Under physiological conditions, α-synuclein is degraded by both the proteasome and autophagy. Mutant α-synuclein inhibits ALP functioning by tightly binding to the receptor on the lysosomal membrane for autophagy pathway control (e.g. Pan et al., 2009; Betarbet et al., 2000).

The strongest evidence supporting that mitochondrial dysfunction precedes the onset of α-synuclein pathology derives from studies on rotenone and MPTP in which repetitive exposure of rodents and monkeys to these chemicals via oral, intraperitoneal, intragastric, or nasal administration resulted in the pathological accumulation of α-synuclein in central as well as peripheral neurons (Cannon et al., 2009; Drolet et al., 2009; Mandel et al., 2004; Pan-Montejo et al., 2012 and 2010; Tristão et al., 2014). For example, male Lewis rats were injected with rotenone (2.0 mg/kg, i.p.) and sacrificed at 0, 4, 8, 16, or 32 h after injection and showed α-
Mice were implanted with somal dysfunction (Fornai et al., 2005). Continuous MPTP administration (respectively, 1-5-30 mg/kg MPTP daily) for the indicated time periods (Fig. 6) (Fornai et al., 2005). Continuous MPTP infusions caused also a long-lasting activation of glucose uptake. Additionally, in mice lacking α-synuclein, the MPTP-induced inhibition of the UPS system and the production of inclusion bodies were reduced (e.g., Ctr mice showed ~40% inhibition of postglutamyl peptidase (PGPH) activity, vs ~13% inhibition observed in α-synuclein KO mice) (Fig. 9), suggesting that α-synuclein could play an important role in UPS inhibition induced by MPP+ (Fornai et al., 2005). These data suggest that continuous, low-level exposure of mice to MPTP causes a Parkinson-like syndrome in α-synuclein-dependent manner (Fornai et al., 2005).

These results are supported by other studies showing that α-synuclein heterozygous mice are resistant to MPTP toxicity (Dauer et al., 2002; Drolet et al., 2004). MPTP exposure (0.5, 5, 50 µM, 48 hr) increases in a dose-dependent manner the α-synuclein protein level in mesencephalic neurons in culture (e.g., ~70% increase at 5 µM vs Ctr) (Duka et al., 2006).

Increased expression of α-synuclein predisposes DA neuronal cells to proteasomal dysfunction (~50% decrease compared to Ctr-vector cells) (Sun et al., 2005).

Accumulation/overexpression of α-synuclein, both wild type and mutant, potentiates inhibition of proteasomal activity. Cells expressing mutant α-synuclein showed a reduction of lysosomal hydrolysis and chymotrypsin-like UPS function (by ~30%, compared to WT) (Stefanis et al., 2001).

Proteasomal inhibition (by mean of lactacystin, a proteasome inhibitor, used at different concentrations for 24 hr) contributes to the accumulation of α-synuclein as it has been described by immunostaining in PC12 cells (Rideout et al., 2001) and in primary mesencephalic neurons (McNaught et al., 2002).

α-Synuclein levels were selectively increased in the ventral midbrain (VMB) region of rotenone-infused rats with or without lesion (~110% increase vs Ctr) (Fig. 3) (Betarbet et al., 2006). Rotenone was administered up to 5 weeks, at 2.5 mg/kg/day. Additionally, 4 weeks of...
in vitro rotenone exposure (5 nM, on SK-N-MC human neuroblastoma cells) increased α-
synuclein levels by 24%, while lactacystin (9 µM, overnight) did not induce any detectable
changes in α-synuclein levels. α-Tocopherol attenuated the rotenone-induced increase in α-
synuclein (comparable to Ctr) (Fig. 10). Furthermore, levels of ubiquitinated proteins detected
in solubilized protein fractions from SK-N-MC cells resulted increased (by 60%) with rotenone
treatment (5 nM), and even more (by 484%) with rotenone combined with lactacystin (Fig.
11) (Betarbet et al., 2006).

- CI inhibition-induced proteasomal dysfunction has been reported in human SH-SY5Y
neuroblastoma cells following acute rotenone exposure (Shamoto-Nagai et al., 2003). The
proteasome activity decreased in the cells treated with rotenone (25 or 50 nM) in a time- and
dose-dependent way. ATP addition restored the reduction of proteasome activity in the cells
treated with 25 nM rotenone for 72 hr. However, after 96 hr of incubation with 25 or 50 nM
rotenone, the activity was reduced respectively to 28.7% and 21.9% of control, and adding
ATP did not increase the activity. After 120 hr, the activity was virtually undetectable (with or
without added ATP) (Fig. 6). On the contrary, the levels of the proteins composing
proteasome did not change with rotenone treatment (Shamoto-Nagai et al., 2003).

**Cytoskeletal damage further enhances disturbed proteostasis**

- α-synuclein can trigger hyperphosphorylation of Tau. Treatment of primary mesencephalic
neurons acutely (48 hr) or subchronic treatment of wild-type (WT) mice with MPP+/MPTP
results in selective dose-dependent hyperphosphorylation of Tau at Ser396/404 (p-Tau). The
presence of α-synuclein was absolutely mandatory to observe MPP+/MPTP-induced increases
in p-Tau levels, since no alterations in p-Tau were seen in transfected cells not expressing α-
synuclein or in α-synuclein−/− mice. MPP+/MPTP also induced a significant accumulation of α-
synuclein in both mesencephalic neurons and in WT mice striatum. Sub-chronic MPTP
exposure increased phosphorylated-Tau in striatum of WT (but not α-Syn−/− mice) causing
microtubule (MT) cytoskeleton instability that affects cellular microtubule transport (including
axonal transport) (Qureshi and Paudel, 2009; Duka et al., 2006). For instance, MPTP was
found to elicit an increase of phosphorylated Tau at Ser262 by 2.8-, 4.5-, 4.6-, and 4.0-fold
higher in 1, 5, 25, and 50 µM MPTP-treated cells than the basal level observed in Ctr/vehicle-
treated cells, respectively. Additionally, MPTP caused a dose-dependent increase in the
intracellular α-synuclein level in M17 human neuroblastoma cells (~3.5 fold increase in cells
treated with 25 µM MPTP vs Ctr) (Qureshi and Paudel, 2009). These results were confirmed
by other studies (e.g. Dauer et al., 2002; Drolet et al., 2004 etc.).

- α-synuclein accumulation followed by MT depolymerisation induces disruption in axonal
transport, which leads to an accumulation of damaged organelles, aggregated/misfolded
proteins and impaired vesicular release. Dopamine is leaking from the vesicles to the cytosol
promoting an increase in oxidative stress, potentiated by dopamine oxidation (Feng, 2006;
Kim et al., 2007). When microtubule network is disrupted, the amount of free tubulin
increases, triggering α-synuclein fibrillation (Payton et al., 2001).

- Axonal transport might be impaired by misfolded α-synuclein through perturbation of
microtubule assembly (Esposito et al., 2007; Lee et al., 2002; Chen et al., 2007 ), especially
together with MAPT protein (Qureshi and Paudel, 2011; Giasson et al., 2003). It induces not
only microtubule disruption but also impairs microtubule-dependent trafficking (Lee at al.,
2006). MT-dependent transport is important for maintaining the Golgi structure, and thus,
depolymerization of the MT leads to a specific pattern of Golgi fragmentation (Cole et al.,
1996). When the MT network was disrupted by nocodazole treatment (5 µg/mL) or α-
synuclein was overexpressed, this normally compact organelle was fragmented and dispersed
(IC images) as shown in COS-7 cells (Lee at al., 2006). Similarly, overexpression of α-
synuclein in differentiated SH-SY5Y cells caused Golgi fragmentation (e.g., ~190% increased
fragmented Golgi) at 12 m.o.i. (multiplicity of infection) of α-synuclein vs Ctr) (Lee at al.,
2006).

- It was found that α-synuclein mutants associated with PD exhibit reduced transport in
neurons, as shown in rat primary neuronal cortical cultures transfected with wild-type (WT),
A53T or A30P α-synuclein. For instance, the rate of transport (expressed in µm/hr) was
Damaged cytoskeletal proteins disrupt also mitochondrial trafficking. Mitochondria use cytoskeletal system as tracks for their directional movement (Nogales, 2000). The function, therefore, damage to microtubules perturbs transport of mitochondria through axons, increasing their retrograde movement. These changes in mitochondrial dynamics lead to a decrease of mitochondria numbers in axons and mitochondria accumulation in cells (De Vos et al., 2007; Miller and Sheetz, 2004). Depletion of mitochondria quantity and function in axons occurs in neurodegenerative disorders (Brownlees et al., 2002; Stamer et al., 2002).

Since mitochondria are ATP suppliers and microtubules need ATP to accomplish their function, mitochondrial dysfunction has a profound effect on axonal transport and function (De Vos et al., 2008).

Mitochondrial dysfunction may damage mitochondrial trafficking through calcium dysregulation. Cytosolic Ca\(^{2+}\) is one of the best-studied regulators of mitochondrial movement. Elevation of cytosolic Ca\(^{2+}\) stops both the anterograde and retrograde trafficking of mitochondria in neurons and in many cell lines (Chang et al. 2006; Szabadkai et al. 2006). In H9c2 cells simultaneous measurements of free Ca\(^{2+}\) levels and mitochondrial dynamics showed that 50% reductions in mitochondrial movement occurred at concentrations of approximately 400 nM Ca\(^{2+}\), and a complete arrest in the low micromolar range (Yi et al. 2004; Saotome et al., 2008). These are indirect proofs suggesting that inhibition of CI, followed by mitochondrial dysfunction, could damage mitochondrial trafficking. Also, chronic exposure to rotenone (50 nM at different times of exposure) was reported to reduce mitochondrial movement in differentiated SH-SY5Y cells (e.g., ~30% reduction of mitochondrial movement (\(\mu\)m/sec) after 8 days of rotenone treatment vs Ctr) (Borland et al., 2008).

### 3.3 Uncertainties or inconsistencies

The exact molecular link from mitochondrial dysfunction to disturbed proteostasis is not known. It is not clear which is the oxidative modification that drives the process.

The sequence of events taking place after inhibition of CI is not entirely clear (Zaltieri et al., 2015). Some studies suggest that induced oxidative stress leads to \(\alpha\)-synuclein aggregation that triggers proteosomal dysfunction (Betarbet et al., 2006). Such order of events is suggested to take place in vivo (McNaught and Jenner, 2001). However, in other studies opposite sequence of events is proposed suggesting that first proteosomal dysfunction take place that leads to \(\alpha\)-synuclein aggregation.

A vicious circle is observed here as \(\alpha\)-synuclein aggregation potentiates proteosomal dysfunction and v/v. In this vicious cycle it is difficult to establish exact quantitative relationship of these two events.

Whether \(\alpha\)-synuclein is a substrate for proteasome remains controversial since both positive and negative data have been reported (Paxinou et al., 2001). Furthermore, polyubiquitination of \(\alpha\)-synuclein, a prerequisite for 26S proteasomal degradation has yet to be reported (Stefanis et al., 2001). It is also not clear whether polyubiquitination of \(\alpha\)-synuclein is necessary for its degradation. However, \(\alpha\)-synuclein gets targeted by the UPS in the SH-SY5Y neuroblastoma cell line. Phosphorylated \(\alpha\)-synuclein gets targeted to mono- or di-ubiquitination in synucleinopathy brains (Hasegawa et al., 2002), but it is not clear if this modification can play any role in proteasomal degradation since monoubiquitination of proteins serves mainly as a signal for endocytosis or membrane trafficking.

On the contrary to the increased \(\alpha\)-synuclein levels observed in the midbrain, decreased \(\alpha\)-synuclein levels were found in the cerebellums of PD patients when compared to controls, suggesting an imbalance of \(\alpha\)-synuclein levels in different parts of the brain (Westerlund et al., 2004).
• Although mitochondrial alterations have been reported in PD patients (Ikawa et al., 2011) and disease models, it is not clear whether they represent a primary pathogenic mechanism. In particular, the critical interplay between mitochondrial dysfunction and oxidative stress, which has been widely reported in PD (Dias et al., 2013) and could constitute either a cause or a consequence of mitochondrial damage, hampers an effective comprehension of the above-mentioned studies. Oxidative stress can constitute a bridge connecting mitochondrial dysfunction to the induction of $\alpha$-synuclein misfolding, aggregation, and accumulation, but otherwise it may be also triggered by these latter events that in turn could induce mitochondrial alterations (Zhu and Chu, 2010; Dias et al., 2013).

• It is still unclear whether the involvement of $\alpha$-synuclein in chronic MPTP toxicity reflects a physiological function for $\alpha$-synuclein that has been activated in the wrong context, or whether $\alpha$-synuclein produces an accidental pathogenicity that contributes to MPTP toxicity but is unrelated to the normal function of $\alpha$-synuclein (Fornai et al., 2005).

• The inconsistent effects of MPP+ on autophagy (up or down regulation) are reported. It may be attributed to differences observed between immortalized cell lines and primary neurons, different timing or dose. While dysregulation of autophagy is always described, the direction is not clear. Further studies are required to clarify this issue.

• MPTP administration does not induce Lewy body formation (in contrast to rotenone) characteristic of PD, even after repeated injections (Drolet et al., 2004; Dauer et al., 2002).

• There is also controversy over whether the increase in autophagic markers is protective or, on the contrary, causative of neuronal death.

• MPP+ may have effects apart from CI inhibition, e.g., on microtubules but it is still unclear whether this is a primary effect. Indeed, MPP+ binds to microtubules in PC12 cells and inhibits their polymerization and stability (Cappelletti et al., 1999; Cappelletti et al., 2001).

• It is not clear whether microtubules disruption may be associated with $\alpha$-synuclein aggregation since tubulin was shown to co-localize with $\alpha$-synuclein in Lewy bodies. Furthermore, tubulin folding is dependent on ATP and GTP hydrolysis, and mitochondrial dysfunction with subsequent energy failure could trigger microtubules disruption. Cytoskeletal microtubule (MT) injury is likely to be responsible for altered rearrangement and movement of cell organelles, being a common feature of several neurodegenerative diseases including PD (Wade, 2009; Mattson et al., 1999).

• It is not clear whether rotenone could cause microtubules depolymerization in vivo and in vitro (Brinkley et al., 1974) by binding to the colchicine site on tubulin heterodimers (Marshall et al., 1978). Ren and Feng (2007) found that microtubule depolymerization induced by rotenone caused vesicle accumulation in the soma and kills neurons.

### 3.4 Quantitative evaluation of KERs

As described in the studies above (Empirical support for linkage) a quantitative or semi-quantitative relationship has been established between rotenone-induced mitochondrial dysfunction and the impairment of UPS/ALP function. Below some representative studies are reported as examples for how such quantitative evaluations can be performed.

• Human neuroblastoma SK-N-MC or human embryonic kidney (HEK) cells were exposed to rotenone at 100 nM for 24 or 48 hrs (for further details see Chou et al., 2010).

Examples of quantitative evaluation of this KER
Fig. 11. Dose and time dependent striatal proteasome activity after MPTP continuously infused up to 28 days measured by relative chymotrypsin-like, trypsin-like, and peptidyl-glutamyl-peptide hydrolysing (PGPH) proteasome activities in mice. Delayed and prolonged inhibition of proteasome activity after continuous MPTP administration (1, 5, or 30 mg/kg MPTP daily) for the indicated time periods. Asterisks indicate statistically significant differences (P < 0.05) from baseline proteasome activity (single asterisk) or from both baseline proteasome activity and activity after lower MPTP doses (1 and 5 mg/kg, daily, double asterisk; n = 5 mice) (Fornai et al., 2005, Fig. 2B).
**Fig. 12.** Effect of a α-synuclein deletion on MPTP toxicity. Proteasome activity in control and α-synuclein KO mice continuously infused for 28 days with MPTP (30 mg/kg of body weight daily, striatum concentration approximately 13 µM). Proteasome activities in the substantia nigra are depicted as percent of control (means +/- SEMs) as a function of time after beginning of the infusions (five mice per group). Asterisks indicate statistically significantly different values (P < 0.05) from controls (Fornai et al., 2005, Fig. 5c)).

![Distribution of alpha-synuclein in rotenone-infused rats](image)

**Fig. 13.** α-Synuclein levels were selectively increased in the ventral midbrain (VMB) region of rotenone-infused rats with or without lesion. α-Synuclein levels, as determined from Western blot analysis, from rotenone-treated rats were expressed as a percentage of values from control vehicle-infused rats. Results are mean ± SEM (n = 3 control, 6 rotenone with lesion, 3 rotenone with no lesion) *P < 0.05 vs. vehicle-infused rats (from Betarbet et al., 2006, Fig. 3A).

![α-synuclein levels following chronic rotenone exposure and proteasome inhibition](image)

**Fig. 14.** Bar graph showing the effects of rotenone and lactacystin on α-synuclein levels after 4 weeks of rotenone exposure (5 nM) in vitro, on SK-N-MC human neuroblastoma cells. Rotenone alone increased α-synuclein levels, but lactacystin alone did not. α-Tocopherol attenuated the rotenone-induced increase in α-synuclein. Results are mean ± SEM (n = 4). *P < 0.05 vs. solvent-treated cells. CC, control cells; RC, rotenone-treated cells; C-Lac or CL, lactacystin treated cells; R-lac or RL, rotenone and lactacystin treated cells; R-AT, rotenone and α-tocopherol treated cells (from Betarbet et al., 2006, Fig. 5B).
Fig. 15. Levels of ubiquitinated proteins were estimated in solubilized protein fractions from SK-N-MC cells collected at the end of each week of rotenone treatment (5 nM), using gel electrophoresis and immunoblotting. Quantitative analysis demonstrated significant increases in ubiquitinated protein levels 4 weeks after rotenone treatment and after proteasomal inhibition with lactacystin. Band intensities were expressed as % of control. Results represent mean ± SEM. *P < 0.05 compared to control (from Betarbet et al., 2006, Fig. 8C).

(1) (A) 25 nM (B) 50 nM

Proteasome activity (% of control)

Fig. 16. Effects of rotenone on the activity of proteasome. Proteasome activity in the cytoplasmic fraction of cells treated with 25 nM (A) or 50 nM (B) rotenone was measured fluorometrically in the absence (open triangles and circles) or presence (solid triangles and circles) of exogenously added ATP (2 mM) (from Shamoto-Nagai et al., 2003, Fig. 6).

Table 2: Quantitative evaluation of the KER

<table>
<thead>
<tr>
<th>KE (upstream) Mitochondrial dysfunction (rotenone, nM)</th>
<th>KE3 (downstream) Impaired proteostasis UPS inhibition (% approx.) measured by:</th>
<th>Comments</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>26S UPS activity + catalase (anti-oxidant)</td>
<td>HEK cells exposed for 2 4hr</td>
<td>Chou et al., 2010</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Activity</td>
<td>Study Details</td>
<td></td>
</tr>
<tr>
<td>------------------------------</td>
<td>--------------------------------------------------</td>
<td>--------------------------------------------------------------------------------</td>
<td></td>
</tr>
<tr>
<td></td>
<td>20S proteasome activity</td>
<td>SK-N-MC human neuronal cell line (exposed for 24 hr)</td>
<td></td>
</tr>
<tr>
<td>1 mg/kg daily</td>
<td>20</td>
<td>Mice continuously infused with MPTP for 28 days</td>
<td></td>
</tr>
<tr>
<td>5 mg/kg daily</td>
<td>30</td>
<td></td>
<td></td>
</tr>
<tr>
<td>30 mg/kg daily</td>
<td>40</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mice continuously infused with MPTP for 28 days</td>
<td>0</td>
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<td></td>
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</tbody>
</table>
The studies presented in the above Table showed that rotenone caused a reduction in UPS activity (measured by 26S and 20S proteasome activity) in a dose-dependent manner. Further studies showed that rotenone increases proteasome subunit degradation, but does not alter synthesis (Western blot and RT-PCR studies, reviewed in Chou et al., 2010). Dose- and time-dependent striatal proteasome activity is also shown after MPTP continuously infused up to 28 days measured by relative chymotrypsin-like, trypsin-like, and peptidyl-glutamyl-peptide hydrolysing (PGPH) proteasome activities in mice (Fornai et al. 2005).

- PD patient-derived fibroblasts (vs Ctr fibroblasts) showed reduction of UPS function (by ~33%) and higher accumulation of ubiquitinated proteins (by ~2 fold) in PD as compared to control fibroblasts at baseline. Treatment with rotenone (20, 500 μM, 6hr) caused a higher induction of 20S proteasome activity in PD fibroblasts vs Ctr. An increase of LC3-II accumulation (indicative of autophagic vesicle accumulation) in both groups (PD and Ctr) after exposure to 500 μM rotenone was observed (Ambrosi et al. 2014).

- Human neuroblastoma cells (SK-N-MC) after short treatment with rotenone (1 week) elevated soluble α-synuclein protein (41 ± 16% increase) levels without changing mRNA levels, suggesting impairment of α-synuclein degradation via UPS. Chronic rotenone exposure (4 weeks) increased levels of insoluble α-synuclein (29 ± 9% increase) and ubiquitin (87 ± 14% increase) (Sherer et al., 2012).

- SHSY-5Y cells treated with rotenone (500 nM, 24 h) showed a ~2 fold increase in DCF fluorescence compared to untreated cells (indicative of intracellular ROS). Additionally, rotenone elevated cytosolic calcium (about 35-40% increase vs Ctr), ER-stress (about 45% increase vs Ctr), impaired UPS function (~3 fold increase of insoluble protein aggregate vs Ctr). Inhibition of Rac1 (Rho-like GTPase) mitigated the oxidative/nitrosative stress, prevented calcium-dependent ER-stress, and partially rescued UPS function (Pal et al. 2014).

- Human neuronal SH-SY5Y cells treated with rotenone (10 μM, for 24 hr showed accumulation of high molecular weight ubiquitinylated bands (by immunoblotting – qualitative - assay), and increase of both mitochondrial- (~5 fold increase vs Ctr) and cytosolic- cytochrome c fractions (~1.2 fold increase vs Ctr). Rapamycin pre-treatment (3 μM, for 48 hr prior addition of rotenone) diminished rotenone-induced effects, as shown by enhanced degradation of ubiquitinated proteins, and reduced levels of cytosolic cytochrome c. Also, rapamycin promoted mitophagy (as shown by lysosome and mitochondria co-localization within the cells) (Pan et al. 2009).

References


Borland MK, Trimmer PA, Rubinstein JD, Keeney PM, Mohanakumar KP, Liu L and Bennett JP. 2008. Chronic, low-dose rotenone reproduces Lewy neurites found in early stages of Parkinson’s disease,


Mitochondrial Dysfunction: The...
PPP5s, Parkinson’s disease and childhood leukaemia


4th KER: Impaired proteostasis leads to degeneration of DA neurons of the nigrostriatal pathway.

4.1 How this key event relationship works

One of the critical functions in the long-lived cells such as neurons is the clearing system for the removal of the unfolded proteins. This function is provided by two major systems, the Ubiquitin Proteosome System (UPS) and the Autophagy-Lysosome Pathway (ALP) (Tai H-C, 2008, Korolchuck VI et al. 2010 and Ravikumar B et al. 2010). Impaired proteostasis with formation of misfolded α-synuclein aggregates deregulates microtubule assembly and stability with reduction in axonal transport and impairment of mitocondrial trafficking and energy supply (Esposito A. et al. 2007, Chen L. et al. 2007; Borland et al. 2008; O’Malley 2010; Fujita et al. 2014; Weihofen et al. 2009). Pathological consequences of these deregulated process include interference with the function of synapses, formation of toxic aggregates of proteins, impaired energy metabolism and turnover of mitochondria and chronic endoplasmic reticulum stress; all eventually leading to degeneration of DA neurons in the nigrostriatal pathway (Fujita et al. 2010, Shulman et al. 2011, Dauer et al. 2003, Orimo et al.2008, Raff et al. 2005; Schwarz 2015).

4.2 Weight of evidence

The weight of evidence for the relationship between impaired proteostasis and degeneration of dopaminergic neurons of the nigrostriatal pathway is strong. The biological plausibility is based on the knowledge of the physiological cellular process governing the clearing processes of degraded proteins and organells and on the observations done in genetic and idiopathic forms of Parkinson's disease. Dose and time concordance support a strong response-respose relationships which is also supported by the very well known chronic and progressive behaviour of the Parkinson's disease. Although essentiality has been demonstrated in multiple models and lines of evidence, including knockout animals, a single molecular chain of events cannot be established; therefore essentiality for this KES relationship was considered moderate.

4.2.1 Biological plausibility

The fact that impaired proteostasis can induce degeneration of DA neurons of the nigrostriatal pathway is well known and based on the understanding of the physiological cellular processes involved in removing degraded/misfolded proteins as they are critical for normal mitochondria and axonal transport. Accumulation of misfolded and/or aggregated α-synuclein and the presence of abnormal mitochondria is a consequence of deregulation of this clearing process, and the Lewy bodies, a pathological hallmark of sporadic PD, stain specifically for proteins associated with UPS (Fornai et al., 2003; Gai et al., 2000; McNaught et al., 2002). Impaired proteostasis has been described in humans affected by sporadic PD (McNaught et al.; 2001, 2003), and changes induced by excess cellular levels of degraded proteins in nigral dopaminergic neurons cause a progressive decline in lysosome function, ie ALP system, contributing to neurodegeneration (Decressac 2013). In this context, the ALP system is likely working in a complementary way, with the UPS being the major cleaning system in the soma and the ALP playing a role at pre-synaptic sites (Friedman et al., 2012).

Pathological observations from patients affected by PD and from animal models show an increased number of autphagic vacuoles or autophagic markers (Alvarez-Erviti L. et al., 2010; Crews L. et al. 2010). Additional observations support the role of impaired proteostasis in nigrostriatal toxicity such as: several genetic variants of sporadic PD are due to susceptible genes able to participate in or modify proteostasis (Shulman et al. 201, Fornai et al. 2003, Shimura et al. 2000, Leroy et al.1998) and striatal microinfusion of proteasome inhibitors induce selective nigrostriatal toxicity with loss of DA and DA metabolites (DA, DOPAC and HVA) in the striatum, retrograde loss of nigral DA cell and intracytoplasmic inclusions positive for protein of the UPS (Fornai et al. 2003).

Transgenic overexpression of mutant or wild-type forms of α-synuclein in mice causes neuropathological changes including dystrophic neurites and α-synuclein positive LB-inclusion (Dauer et al. 2003, Masiliah et al. 2000). However, they fail to reproduce specific cell death in the nigrostriatal pathway. In contrast, injection of human α-synuclein expressing viral vectors into the SN of adult rats causes a selective death of dopaminergic neurons and formation of LB inclusions (Dauer et al. 2003, Kirik et al. 2002, Lo Bianco et al. 2002). These effects were observed with adeno-associated virus –

Impaired proteostasis and formation of proteins aggregates also affect the axonal transport and mitochondrial trafficking. α-synuclein mutants accumulate in the neuronal soma when overexpressed, reducing the axonal transport (Kim-Han et al., 2011; Saha et al., 2004); in addition, overexpressed vesicle-associated α-synuclein binds to the microtubules with a detrimental role on axonal transport (Kim-Han et al., 2011; Yang et al. 2010). Postmortem studies on PD patients are indicative of axonal damage. It appears that axonal changes precede neuronal loss, supporting the idea that axonal impairments are early events in neurodegenerative disorders (Orimo 2005 and 2008, Raff 2002, Braak et al. 2004).

These changes, and observation from animals models using the chemical stressor MPTP (Meissner et al. 2003, Serra et al 2002, Hasbani et al. 2006) are supporting the notion that DA neurons of the nigrostriatal pathway degenerate through a “dying back” axonopathy (Raff et al. 2002). It was demonstrated that axonal degeneration follows an active process distinct from cell body loss in a Wallerian degeneration slow (WldS) mutant mouse transgenic model. In this model, axonal degeneration in a variety of disorders is inhibited. In WldS mice, acute treatment with MPTP (20 mg/kg ip for 7 days) resulted in attenuated nigrostriatal axon degeneration and attenuated DA loss, but cell bodies were not rescued (Hasbani et al. 2006). Indeed, multiple evidences from genetic and experimental models (particularly using MPTP as a stressor) support an early and critical role of axonal impairment with early occurrence of Lewy neurites preceding Lewy bodies formation and cell death (O’Malley 2010).

In addition, a strong link between mitochondrial dysfunction and PD came from the discovery that mutations in PINK1, α-synuclein, LRRK2, parkin and DJ-1, all linked with genetic causes of PD, can affect mitochondrial function (Rappold et al.2014, O’Malley 2010). Deregulation of mitochondrial dynamics (fission, fusion and movement of mitochondria) can affect neuronal activity and viability and imbalance of mitochondrial dynamics have been reported in experimental models of PD with mutated α-synuclein (Tieu, 2014) or chronic model of primary neuronal cells treated with low concentrations (0.1 nM) of rotenone (Arnold et al. 2011). Progression of neuronal changes with formation of Lewy neurites and reduction of mitochondrial movement leading to cell death has been also observed in vitro in a chronic cell-based model (SH-SY5Y neuroblastoma cell line) treated with low concentration of Rotenone (50nM for 21 days). In this assay, reduction in mitochondrial movement was associated with a progressive damage, first including formation of Lewy neurites, followed by cell death (Borland et al.2008).

### 4.2.2 Empirical support for linkage


Neurotoxic external doses of both rotenone and MPTP are well characterized and reported; however, the corresponding brain concentration is much less frequently quoted. In order to understand the brain concentration for both compounds, data were retrieved from Betarbet et al. 2000 and 2006 for rotenone and from Fornai et al. 2005 and Thomas et al. 2012 for MPTP. In all cases, the compounds were administered by infusion and, at least for MPTP, the brain concentrations were taken after chronic infusion and are expected to be at the steady state. For MPTP only, brain concentration was expressed as ng/mg protein (Fornai et al. 2005) or as ng/mg weight tissue (Thomas et al. 2012). To do the final estimate we assumed a density for protein as 1.4 (Quillin and Matthews 2000) and a protein content in the brain of about 10% (Schwartz et al. 2012). Density for brain tissue was assumed to be 1. The final concentration was 12 µM (Fornai et al. 2005) and 47 µM (Thomas et al. 2012).
It should be noted that the upstream key event includes multiple pathological events, eventually leading to the downstream key event. As it is difficult to assess real time changes for a series of complex and dynamic events in a single experiment, most of the empirical supporting evidences are performed by exploring single factors (e.g. impairment of ALP or UPS or axonal transports) and their role in the degeneration of DA neurons.

**MPTP/MPP⁺**

- Inhibition of the UPS was observed following continuous infusion of MPTP at 1, 5 and 30mg/kg/day for 28 days in mice. A dose related decrease in the enzyme activity of the UPS was observed and was associated with a dose-related decrease of TH positive terminals (densitometry analysis) in the dorsal and ventral striatum. This effect was accompanied by a dose-related cell loss in the SN (counting of TH positive cells) at 5 and 30 mg/kg/day. At 30 mg/kg/day the authors reported cytoplasmic inclusions positively staining for ubiquitin and α-synuclein in neurons of the SN (and locus coeruleus). In the same experiment, acute administration of MPTP (single injection of 30 mg/kg/ or 4 separate injections of 20 mg/kg) induced a transient inhibition of the UPS activity, neuronal loss but no intracytoplasmatic inclusions, indicating that a continuous infusion is necessary to induce permanent inhibition and pathological changes similar to the one observed in PD (Fornai et al. 2005).

- In mice lacking α-synuclein, continuous infusion of up to 30 mg/kg/day for 28 days of MPTP neuronal cell death and behavioral symptoms were almost alleviated (Fornai et al. 2005, Dauer et al. 2002).

- Administration of MPTP to mice (30 mg/kg/day ip for 5 days) produced autophagosome (AP) accumulation (increase in LC3II) and dopaminergic cell death which was preceded by a decrease in the amount of lysosomes in DA neurons. MPTP also induced mitochondrial-derived ROS and permeabilization of the lysosomal membrane. This resulted in a decrease in Lamp 1 lysosome structural protein and accumulation of undegraded AP and release of lysosomal enzymes into the cytosol. The effect observed in-vivo was quantitatively confirmed in-vitro (human neuroblastoma cell line BEM17(M17EV)). MPP⁺ was tested in-vitro at the concentrations of 0.25 to 2.5 μM and induced a concentration-related decrease in Lamp1, increase in LC3II, increase in cell death and decrease in lysotracker. In the same in-vitro system, MPP⁺ also induced lysosome membrane permeabilization. In the same experiment, induction of lysosome biogenesis by the autophagy-enhancer compound rapamycin attenuated the dopaminergic neurodegeneration, both in vitro and in vivo, by restoring lysosomal levels (Dehay et al. 2010).

- In an in-vitro microchamber that allowed specific exposure of neuritis of murine mesencephalic neurons, treatment with 1 to 5 μM of MPP⁺ induced impairment of mitochondrial transport, neurite degeneration (degeneration of proximal dendrites) and autophagy, before cell death (Kim-Han et al. 2011). The number of TH positive cell bodies and neurites was reduced at 1 μM, and axonal fragmentation and LC3 dots increased while tubulin density decreased (Kim-Han et al. 2011).

- Mice treated with MPTP at 20mg/kg/day ip for 5 days showed loss of DA neurons in SN which was attenuated by the pharmacological block of mitochondrial fission protein Drp1. Drp 1 blockade also promoted mitochondrial fusion and enhanced the release of DA from the striatal terminals in a PINK1 knockout model showing a defective DA release (Rappold et al. 2014; Tieu et al. 2014).

- In differentiated (d6) LUHMENS cell system stably expressing eGFP/mito-tRFP, treatment with MPP⁺ (5μM) for 24 hours revealed a reaction in the total number of mitochondria in neuritis and a significant reduction in velocity. Partial protection from MPP⁺ dependent mitochondrial immobilization in neuritis as well as drop in mitochondria numbers in neuritis was detects following co-treatment with the anti-oxidant Vitamin C (Schildknecht et al. 2013)
Proteasome inhibitors

- Intracerebral microinfusion of proteasome inhibitors (lactacystein or epoxomycin at 1000 µM) induced loss of TH and DAT immunostaining and decrease in DA and DOPAC in DA terminals in the striatum and loss of nigral cells in SN (counting of TH positive cells).
- Formation of cell inclusions (positively immunostained for α-synuclein and ubiquitin) and apoptosis were observed after treatment with proteasome inhibitors (0.1 to 50 µM) in an in-vitro system (PC 12 cells). The concentration response curve for apoptosis was shifted to the right compared to the concentration response curve for cellular inclusions indicating that inclusions occurred earlier and independently of cell death. A maximum effect was reached between 1 and 10 µM (Fornai et al.2003).

Rotenone

- Administration of rotenone, via osmotic mini pumps implanted to rats (3 mg/kg/day for 7 days) induced decrease of TH in substantia nigra and striatum and decrease in α-synuclein, in its native form, in substantia nigra and striatum, while monoubiquitinated alpha-synuclein increased in the same regions. Valproic acid (VPA) treatment (effective inhibitor of histone deacetylases) significantly counteracted the death of nigral neurons and the 50% drop of striatal dopamine levels caused by rotenone administration. VPA treatment also counteracted both type of α-synuclein alterations. Furthermore, monoubiquitinated alpha-synuclein increased its localization in nuclei isolated from substantia nigra of rotenone-treated rats, an effect also prevented by VPA treatment. Nuclear localization of alpha-synuclein has been recently described in some models of PD and its neurodegenerative effect has been ascribed to histone acetylation inhibition (Monti et al. 2010).
- Chronic oral administration of rotenone at 30mg/kg/day in mice produced neuronal loss and degeneration of TH positive terminals in the striatum accompanied by an increase in α-synuclein, ubiquininated proteins and decrease in proteasomai activity. Concomitant treatment with 4-PBA (a chemical chaperone able to reverse the mislocalization and/or aggregation of proteins) inhibited rotenone-induced neuronal death and decreased protein level of α-synuclein (Inden et al. 2007).
- Treatment of Lewis rat with 2 mg/kg/day of rotenone, administered sc for 8 weeks impaired autophagic flux, induced lysosomal dysfunction and degeneration of DA neurons (decrease in number of TH positive cells and decrease in density of TH positive fibers) in SNpc. The effect of rotenone was paralleled by an increase in LC3 immunopositive dots and upregulation of the LC3II in DA neurons. A concomitant effect was observed and characterized by a decrease in LAMP2 and cathepsin immunodots with a diffuse morphological pattern, possibly indicative of decreased lysosomal membrane integrity and leaking to cytosol. In-vitro (PC12 cells) at 500 nM, rotenone also induced increases in α-synuclein, microtubule associated protein 1, light chain 3-II, Beclin 1, p62, increased lysosome permeability and induced cell death. In PC12 cell, the concomitant treatment with trehalose (autophagic inducer) attenuated the rotenone-induced cell death while in-vivo trehalose treatment decreased the rotenone-induced dopaminergic neurons loss (Wu et al. 2015).
- Rotenone LD50 of 10 nM in differentiated SH-SY5Y cells decreased autophagic flux at both 2 and 24h. Up-regulation of autophagy by rapamycin protected against cell death while inhibition of autophagy by 3-methyladenine exacerbated cell death (Giordano et al. 2014).
- Treatment of embryonic midbrain neuronal cells with 0.1 to 10 µM rotenone for 30 minutes induces a decrease in polymerized tubulin and increased the number of apoptotic TH+ cells. Similar effects were observed with colchicine treatment, a well-known microtubule-depolyrizing agent and prevented by taxol, a well-known microtubule –stabilizing agent. The effect was considered specific to DA neurons as the effect on apoptosis and cell death was much less evident in GABAergic and glutamatergic neurons (Ren et al. 2005).
Human evidences

- Inclusion bodies in DA neurons (ie Lewy bodies), a pathological hallmark for sporadic PD, stains specifically for proteins associated with the UPS (Fornai et al. 2003, Gai et al. 2000, Mcnaught et al. 2002), including α-synuclein, parkin and ubiquitin; possibly indicating that failure of the UP system represents a common step in the pathogenesis of PD and impairment of the proteasome system was found in humans affected by sporadic PD (McNaught et al. 2001, 2003).

- Lysosomal breakdown and autophagosome (AP) accumulation with co-localization of lysosomal markers in Lewy Bodies is reported to occur in PD brain samples where Lewy bodies were strongly immunoreactive for the autophagosome markers (LC3II). (Dehay et al. 2010).

- Postmortem studies on PD patients show axonal pathology that is likely to precede the loss of neuronal bodies In this investigation, TH immunoreactive fibers had almost entirely disappeared with preservation of neuronal bodies (Orimo et al. 2005 and 2008).

4.3 Uncertainties or inconsistencies

- MPTP can induce damage to nigrostriatal neurons without formation of Lewy bodies (hallmark of PD). Acutely intoxicated humans and primates with MPTP lack LB-like formation (Dauer 2003; Forno 1986, 1993). Similarly, discontinuous administration of rotenone, even at high doses, damages the basal ganglia but produce no inclusions (Heikkila et al. 1985; Ferrante et al. 1997, Lapontine 2004). To reproduce the formation of neuronal inclusions, continuous infusion of MPTP or rotenone is necessary.

- Acute intoxication with rotenone seems to spare dopaminergic neurons (Dauer et al 2003, Ferrante 1997). In addition, in rats chronically infused with rotenone showed a reduction in striatal DARPP-32-positive, cholinergic and NADPH diaphorase-positive neurons (Hoglinger 2003) or in other brain regions. These results would suggest that Rotenone can induce a more widespread neurotoxicity (Aguilar 2015).

- The vulnerability of the dopaminergic pathway still remains circumstantial. The selectivity of MPP+ for dopaminergic neurons is due to its selective uptake via dopamine transporter (DAT), which terminates the synaptic actions of dopamine (Javitch 1985, Pifl, 1993, Gainetdinov 1997, Hirata 2008). Selectivity of Rotenone for dopaminergic neurons is not fully understood (Hirata 2008).

- Transgenic overexpression of α-synuclein induces neurotoxicity (ie neuronal atrophy, distrophic neuritis, astrocytosis and LB-like formation). However they fail to cause death of dopaminergic neurons. Nevertheless, injection of the human protein or mutated form expressing viral vectors into the SN, are able to induce all the pathological changes characteristic of PD. This discrepancy could be due to the higher expression of α-synuclein in the viral vector model or because in these models, α-synuclein overexpression would occur suddenly in adult animals (Dauer et al. 2003). In addition, transgenic expression of C-terminal truncated α-synuclein also leads to motor symptoms but neuronal degeneration is not reported (Halls et al. 2015).

- There is conflicting literature on whether increased autophagy would be protective or enhances damage. Similarly, a conflicting literature exists on extent of inhibition or activation of different protein degradation system in PD and a clear threshold of onset is unknown (Fornai et al. 2005).

- Several mechanisms may affect the axonal transport in neurons showing swelling of neurites positive for α-synuclein. These include e.g. ROS production, lysosome and mitochondria membranes depolarization, increased permeability and microtubule depolymerization (Kim-Ham 2011, Borland 2008, Choi 2008). As both MPTP and Rotenone could directly trigger these effects, a clear mechanistic understanding leading to cell death is difficult to identify (Aguilar et al. 2015).
Different features of imbalanced proteostasis can trigger one another (e.g. disturbed protein degradation, pathological protein aggregation, microtubule dysfunction); and each of them can lead to cell death. Therefore, the “single” triggering event triggering axonal degeneration or neuronal death is not known. For instance, for α-synuclein aggregation, it is not clear whether this causes death because some vital function of neurons is lost, or whether some protein increases e.g. because of inhibited chaperone-mediate autophagy (Kaushik et al. 2008, Cuervo et al. 2014).

Real-time changes in DA axons are difficult to assess, accounting for the limitation of testing models of structural or trafficking impairment in-vivo.

### 4.4 Quantitative evaluation of KER

As described in the empirical support, a quantitative relationship has been established between chemical stressors inducing impaired proteostasis and loss of DA neurons of nigrostriatal pathway. The response-response relationship was evident in most of the studies and, where possible a relationship in dose-response could be also observed. A chronic dose regimen for the chemical stressor was necessary in most of the studies and this is confirming that a long lasting perturbation of the key event up is necessary to affect neuronal loss consistent with the presence of intracytoplasmatic inclusions. However, some inconsistency in the measurement of the endpoints relevant for impaired proteostasis were observed, probably because they also act as compensatory factors (Betarbet et al. 2006). The acute administration of MPTP (single injection of 30 mg/kg/ or 4 separate injections of 20 mg/kg) induced a transient inhibition of the UPS activity and neuronal loss but no intracytoplasmatic inclusions i.e Lewy body were observed, supporting the temporal relationship among the two events (Fornai et al. 2005).

### Table 3: Quantitative evaluation of the KER

<table>
<thead>
<tr>
<th>Measured endpoint relevant for the KEup (KE3)</th>
<th>Measured endpoint relevant for the KEdown (KE4)</th>
<th>Model</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Approx. 40% inhibition of UPS</td>
<td>Approx. 38% decrease in TH density in dorsal striatum</td>
<td>MPTP 1mg/kg/day IV infusion for 28 days in mice</td>
<td>Fornai et al. 2005</td>
</tr>
<tr>
<td>Approx. 50% inhibition of UPS</td>
<td>Approx. 40% decrease in number of TH positive cells/mm² in SN and approx. 25% decrease in TH in dorsal striatum</td>
<td>MPTP 5mg/kg/day IV infusion for 28 days in mice</td>
<td></td>
</tr>
<tr>
<td>Approx. 60% inhibition of UPS</td>
<td>Approx. 86% decrease in number of TH positive cells/mm² in SN and approx. 50% decrease in TH in dorsal striatum and approx. 50% in ventral striatum</td>
<td>MPTP 30mg/kg/day IV infusion for 28 days in mice</td>
<td></td>
</tr>
<tr>
<td>Approx. 40% proteasome inhibition</td>
<td>Approx. 70% decrease in DA and 50% decrease in DOPAC in striatum and 30% cell loss in SN</td>
<td>ic infusion of lactacystin (proteasome inhibitors) in rats 100 µM</td>
<td>Fornai et al. 2003</td>
</tr>
<tr>
<td>Approx. 50% increase in mRNA expression for α-synuclein</td>
<td>Decrease in TH immunoreactivity (approx. 50%), in TH-positive nerve terminals in the striatum</td>
<td>Transgenic model overexpressing α-synuclein</td>
<td>Kirk et al. 2002</td>
</tr>
<tr>
<td>Approx. 16-13% reduction in proteosomal activity</td>
<td>Degeneration of nigrostriatal dopaminergic neurons in 50% of animals</td>
<td>Chronic iv treatment (up to 5 weeks) of Lewis rat with rotenone at 2-3 mg/kg day (free brain Rotenone 20-30 nM)</td>
<td>Betarbet et al. 2000 and 2006</td>
</tr>
<tr>
<td>Approx. 50% increase in α-synuclein</td>
<td>Approx. 57% reduction in TH immunoreactivity in SNpc neurons at 30</td>
<td>Oral chronic administration (28 days) of rotenone (0.25, 1, 2.5,</td>
<td>Inden et al. 2007</td>
</tr>
<tr>
<td>Treatment</td>
<td>Effect</td>
<td>Reference</td>
<td></td>
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<tr>
<td>-----------</td>
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</tr>
<tr>
<td>30 mg/kg/day</td>
<td>Decrease in TH and DAT in the striatum (approx. 30% and 70% respectively) and ventral midbrain area (approx. 60%) at 30 mg/kg/day</td>
<td>5, 10 or 30 mg/kg/day to mice</td>
<td></td>
</tr>
<tr>
<td>Increase in LC3 positive dots in nigral DA neurons (approx. 380%), upregulation of LC3II (approx. 40%), Beclin 1 (approx. 33%) and P62 (approx. 50%) autophagic substrate</td>
<td>Approx. 40% decrease in the number of TH neurons (SNpc) and density of TH positive fibers (approx. 50%) (striatum).</td>
<td>2 mg/kg/day for 8 wks sc of Rotenone in Levis rats</td>
<td></td>
</tr>
<tr>
<td>Approx. 8 fold increase in the number of TH+ neurons with granular LC3</td>
<td>Approx. 40% decrease in the number of TH immunoreactive neurons.</td>
<td>Primary dopaminergic neurons following treatment with MPP+ (LD50 of 5µM/L)</td>
<td></td>
</tr>
<tr>
<td>Decrease in mitochondrial speed (approx. 100% decrease in anterograde speed and approx. 28% increase in retrograde speed)</td>
<td>Approx. 70% decrease in positive TH neuronal bodies at 48 hours</td>
<td>Treatment with up to 5 µM (1 to 5 µM) of MPP+ in TH positive mesencephalic neurons in an in-vitro microchamber segregating system</td>
<td></td>
</tr>
<tr>
<td>Reduction in mitochondrial movement was statistically significant from day 8 and was greatest on day 16 at 50 nM (approx. day 3 19%, day 6 7%, day 8 62%, day 14 37%, day 16 200%)</td>
<td>Approx. 60% of cell loss by day 21</td>
<td>In vitro SH-SY5Y neural cells treated with 50 nM rotenone for 21 days</td>
<td></td>
</tr>
<tr>
<td>30% increase over control in static mitochondria and 50 decrease over control in number of mitochondria</td>
<td>Significant decline of intracellular ATP at 24 hours</td>
<td>Differentiated (d6) LUHMENS stably expressing eGFP/mito-tRFP, treated with MPP+ (5µM) for 24 hours</td>
<td></td>
</tr>
</tbody>
</table>

**Note:** The table above summarizes various effects observed in different experimental conditions, including changes in TH and DAT levels, LC3 positive dots, mitochondrial speed, and ATP levels. The references cited provide further details on the methods and results of these experiments.
Fig. 17: Neurotoxicity induced by continuous MPTP administration. (a) Representative tyrosine hydroxylase (TH)-stained sections of the substantia nigra from mice that were continuously treated for 28 days with control pump infusions or with infusions of 1, 5, or 30 mg MPTP/kg daily. (Scale bar, 600 μm.) (b and c) TH-positive cell counts in the substantia nigra (b) and semiquantitative densitometric measurements of the TH signal in striatum (c) (n = 10 mice per group). (d) Striatal monoamine levels in MPTP-treated mice (n = 10 mice per group). Asterisks indicate statistically significant differences (P < 0.05) of a sample compared to control (single asterisks) or to both the control and the lower MPTP dose (double asterisks).

**Fig. 18.** Effect of an α-synuclein deletion on MPTP toxicity. (b) Uptake of $[^{14}\text{C}]2$-DG in littermate wild-type and α-synuclein KO mice that were continuously infused for 7 days with control or MPTP (30 mg/kg daily) solution. Pictures display false-color autoradiograms. (c) Proteasome activity in control and α-synuclein KO mice continuously infused with MPTP (30 mg per kg of body weight daily). Proteasome activities in the substantia nigra are depicted as percent of control (means ± SEMs) as a function of time after beginning of the infusions (five mice per group). In a and c, asterisks indicate statistically significantly different values ($P < 0.05$) from controls.


**Evidence Supporting Taxonomic Applicability**

Multiple animal models have been used to mimic PD (Johnson et al. 2015). There are no sex restriction; however, susceptibility to MPTP increases with age in both non-human primates and mice (Rose et al. 1993, Irwin et al. 1993, Ovadia et al. 1995).
References


5th KER: Neuroinflammation leads to degeneration of the dopaminergic neurons of nigrostriatal pathway

5.1 How this KER works

Cells of the innate (microglia and astrocytes) and adaptive (infiltrating monocytes and lymphocytes) immune system of the brain have, like other immune cells (in peripheral tissues), various ways to kill neighboring cells. This is in part due to evolutionary-conserved mechanisms evolved to kill virus-infected cells or tumor cells; in part it is a bystander phenomenon due to the release of mediators that should activate other cells and contribute to the killing of invading microorganisms. An exaggerated or unbalanced activation of immune cells can thus lead to parenchymal (neuronal) cell death (Gehrmann et al., 1995). Mediators known to have such effects, and that are also known to be produced during inflammation in the brain comprise components of the complement system and cytokines/death receptor ligands triggering programmed cell death (Dong and Benveniste, 2001). Besides these specific signals, various secreted proteases (e.g. matrix metalloproteinases), lipid mediators (e.g. ceramide or gangliosides) or reactive oxygen species can contribute to bystander death of neurons (Chao et al., 1995; Nakajima et al., 2002; Brown and Bal-Price, 2003; Kraft and Harry, 2011; Taetzsch and Block, 2013). Especially the equimolar production of superoxide and NO from glial cells can lead to high steady state levels of peroxynitrite, which is a very potent cytotoxicant (Yuste et al., 2015). Already damaged neurons, with an impaired anti-oxidant defence system, are more sensitive to such mediators.

An important role of microglia in the brain is the removal of cell debris (Xu et al., 2015). Healthy cells continuously display anti-“eat me” signals, while damaged and stressed neurons/neurites display “eat me” signals that may be recognized by microglia as signal to start phagocytosis (Neher et al., 2012), thus accelerating the loss of DA neurites in the striatum.

Activated microglia surrounding DAergic neurons in PD express the M1 neurodegenerative phenotype (Hunot et al., 1999), which promote proliferation and function of CD4+ T cells (for review Appel et al., 2010), which in turn induce DA neuron toxicity, as assessed by experiments with immunodeficient mice (Brochard et al., 2009). Possible infiltration of other myeloid cells, such as monocytes or macrophages through a compromised blood-brain barrier, may also be involved in phagocytosis and neurodegeneration (Déboyer et al., 2012 ; Pey et al., 2014).

5.2 Weight of evidence

5.2.1 Biological plausibility

Histopathological studies have shown that glial activation is a hallmark of every neurodegenerative disease, including Parkinson’s disease (Whitton, 2007 ; Tansey and Goldberg, 2009 ; Niranjan, 2014 ; Verkhratiky et al., 2014). PET studies in PD patients have revealed that microglial activation in the substantia nigra is an early event in the disease process (Iannaccone et al., 2012), and that it is extremely persistent. The role of astrocytes is less clear than the one of microglia, but reactive astrocytes are able to release neurotoxic molecules (Mena and Garcia de Ybenes, 2008; Niranjan, 2014). However, astrocytes may also be protective due to their capacity to quench free radicals and secrete neurotrophic factors. The activation of astrocytes reduces neurotrophic support to neurons, and the proportion of astrocytes surrounding dopaminergic neurons in the substantia nigra is the lowest for any brain area suggesting that dopaminergic neurons are more vulnerable in terms of glial support (for review, Mena and Garcia de Ybenes, 2008).

In vitro co-culture experiments have demonstrated that reactive glial cells (microglia and astrocytes) can kill neurons (Chao et al., 1995 ; Brown and Bal-Price, 2003 ; Kraft and Harry, 2011 ; Taetzsch and Block, 2013), and that interventions with e.g. i-NOS inhibition can rescue the neurons (Yadav et al., 2012; Brzozowski et al., 2015). Direct activation of glial cells with the inflammomogen LPS has also resulted in vivo in the death of DA neurons (Sharma and Nehru, 2015; Zhou et al., 2012; Li et al., 2009).

Circulating monocytes and lymphocytes.

Neuroinflammation can disrupt blood-brain barrier integrity (Zhao et al., 2007), facilitating infiltration of circulating monocytes and lymphocytes (Machado et al., 2011; Quian et al., 2010). T cell infiltration
has been found in CNS tissue of PD patients (Miklossy et al., 2006; Qian et al., 2010), and in animal models, in which depletion or inactivation of lymphocytes has been found to protect striatal DA terminals (for review, Appel et al., 2010).

5.2.2 Empirical support for linkage

LPS injections

Lipopolysaccharide (LPS, a known activator of microglia) injected into the substantia nigra successfully replicated the pathogenic features of Parkinson’s disease in rats. An increase in the mRNA expression of pro-inflammatory cytokines (TNF-alpha, IL-1 beta) was observed 7 days post-injection; alterations in oxidative stress markers (ROS, lipid peroxidation, NO formation, NADPH oxidase activity, GSH system, SOD and catalase) became significant 14 days post-injection, and this was followed by a significant decline in tyrosine hydroxylase (TH), as marker of dopaminergic neurons (Sharma and Nehru, 2015). LPS-induced downregulation of TH expression seemed to depend on the pro-inflammatory cytokine IL-1 beta, since it was not observed in LPS-injected IL-1 knockout mice (Tanaka et al., 2013).

Progressive hypokinesia, selective loss of dopaminergic neurons in substantia nigra and reduction of striatal dopamine content, as well as alpha-synuclein aggregation in substantia nigra was also achieved by unilateral intranasal instillation of LPS every other day for 5 months, mimicking a progressive inflammation-mediated chronic pathogenesis of Parkinson’s disease (He et al., 2013). It is important to note that LPS administrated either directly in the brain, intraperitoneally or in utero results in a delayed and progressive loss of nigral DA neurons that persists well after the initial inflammatory stimulus (for review, Taetzsch and Block, 2013).

Rotenone

Chronic systemic rotenone exposure reproduces features of Parkinsons’ disease with loss of DA neurons and putative Lewis bodies in substantia nigra, accompanied by neuroinflammation and oxidative stress, and reduction of TH immunoreactivity in striatum together with an increase in reactive astrocytes (Betarbet et al., 2000; Ferris et al., 2013). In this chronic rotenone model (2-3 mg/kg per day up to 4 weeks), microglia activation precedes neuronal death (Sherer et al., 2003). Several interventions aiming at blocking several features of microglial activation (NADPH oxidase, myeloperoxidase, phagocytosis, opening of K<sub>ATP</sub> channels,...) protected DA neurons from death (Wang et al., 2014; Emmrich et al., 2013; Chang et al., 2013; Salama et al., 2013; Zhou et al., 2007; Gao et al., 2003). An enhanced sensitivity of dopaminergic neurons to rotenone-induced toxicity was observed with aging, in parallel with the increase of glial cell activation in older rats (Phiney et al., 2006).

In vitro, little neurotoxicity was detected in primary DA neuron cultures (low glia-content) exposed to rotenone, whereas significant and selective dopaminergic neurodegeneration was observed in neuron/glia cultures (Gao et al., 2002).

MPTP/MPP+

Following MPTP treatment, microglial cells are activated by a mechanism secondary to dopaminergic neuron injury (Zhou et al., 2005). However, elevation of interferon-gamma and TNFalpha in substantia nigra was detected before the death of DAergic neurons (Barcia et al., 2011); and serum levels of IFN-gamma and TNFalpha remain elevated for years in monkeys exposed to MPTP (Barcia et al., 2011). The role of microglia in the progression of DA neurodegeneration is suggested by in vivo and in vitro experiments in which feature of microglial reactivity (TNF-alpha, i-NOS, NADPH-oxidase, ROS generation) were blocked (Brzozowski et al., 2015; Wang et al., 2006; Liu et al., 2015; Wang et al., 2014; Chung et al., 2011; Borrajo et al., 2014; Bodea et al., 2014; Sriram et al., 2002; Feng et al., 2002; Dehmer et al., 2000; Ferger et al., 2004). Some evidence from above studies also extends to astrocytes (Sathe et al., 2012; Khan et al., 2014). For instance, systemic administration of nicotine (stimulating the anti-inflammatory role of alpha 7 nicotinic acetylcholine receptors on astrocytes and microglia) reduced MPTP-induced motor symptoms, and protected against neurodegeneration in the substantia nigra by (Liu et al., 2012; 2015).

Entrance into the brain of bone marrow-derived cells expressing i-NOS may also play a deleterious role in neurodegeneration (Kokovay and Cunningham, 2005). Indeed, pharmacological inhibition or
deletion of CD95 in peripheral myeloid cells hampered brain infiltration and was protective for MPTP-induced DA loss in striatum (Gao et al., 2015; Chung et al., 2015). Similarly, therapies aiming at suppressing immune reactivity, such as administration of Treg cells (CD4+CD25+ regulatory T cells) lead in MPTP treated mice, to a robust nigrostriatal protection associated to an inhibition of microglial reactivity (Reynolds et al., 2010).

Paraquat

Paraquat alone (10mg/kg, 2x/week, for 4 weeks) or in combination with manebo (30 mg/kg) induces a loss of DAergic neurons in the substantia nigra paralleled by an increase in microglial reactivity (Cicchetti et al., 2005; Mitra et al., 2011). In a paraquat rat model, microglial reactivity was observed 4 weeks post-injection, whereas degeneration of DAergic neurons was only observed 2 weeks later (Sant-Pierre et al., 2006).

Direct treatment of primary microglial cells with paraquat (5-15 microM) showed no morphological change and no upregulation of IL-10, IL-1beta, IL-2, IL-4, TNF-alpha, GM-CSF or INF-gamma, suggesting that paraquat cannot activate directly microglial cells (Klintworth et al., 2009), despite contrasting observations in the microglial cell lines BV2 (Miller et al., 2007) or N9 (Bonneh-Barkay et al., 2005). But « priming » of microglial cells by a first exposure to paraquat (10 mg/kg) (Purisai et al., 2007), by LPS (2-4 mg/kg) (Purisai et al., 2007), or by a viral mimic (Bobyn et al., 2012) increased the vulnerability of DA neurons to further paraquat treatments. Interestingly, if minocycline (45 mg/kg), an antibiotic known to decrease microglial reactivity, was applied together and after the first priming paraquat treatment, subsequent exposure to paraquat failed to cause DA neurodegeneration (Purisai et al., 2007). If paraquat treatments were made in mice lacking functional NADPH oxidase, no DA neurodegeneration was detected (Purisai et al., 2007), identifying again NADPH-oxidase as a key factor (Wu et al., 2005).

In particular, the NADPH oxidase isofrom NOX2 located on microglia plasma membranes transfers electrons to paraquat inducing the formation of the paraquat radical cation (Rappold et al. 2011). Radical paraquat may then (i) react with oxygen efficiently producing superoxide and regenerating paraquat, and/or (ii) enter DA neurons being a substrate for the dopamine transporter (DAT) (Rappold et al., 2011). This second possibility is supported by the observation that cells expressing DAT efficiently uptake paraquat only in the presence of microglia, but not when NOX2 activity is specifically abolished (Rappold et al. 2011). Neurodegeneration may be then triggered (i) by the amplification of the extracellular redox signalling (Purisai et al., 2007, Bonneh-Barkay et al., 2005) and/or (ii) establishing a new round of redox cycling once paraquat is taken up into DA neurons. Accordingly, expression of DAT sensitise HEK293 cells to paraquat (50 microM) induced intracellular ROS production and cell death, as well as mutant mice with hypomorphic DAT are resistant to paraquat neurotoxicity (Rappold et al. 2011).

Besides NADPH-oxidase, other inflammatory factors are involved in DA neurodegeneration: for example, iNOS, NF-kappaB or p38 MAPK, since their blockade reverted the 50% decrease of TH immunoreactivity, as well as IL1-beta and NO increased expression in striatum observed following paraquat or paraquat and manebo treatments (Yadav et al., 2012). Similarly, IFN-gamma silencing prevented the paraquat-induced morphological signs of microglial activation, the NADPH-oxidase expression, as well as the time-dependent changes in the pro-inflammatory enzymes i-NOS and COX-2, of cytokines (IL-1beta, TNF alpha), and of signaling molecules (JNK and p38 MAPK), and protected against paraquat-induced DA neurodegeneration (Mangano et al., 2012).

Protection against paraquat-induced DA neurodegeneration can also be achieved by providing trophic support (intranigral or peripheral injection of GDNF or GM-CSF, respectively), which is reduced upon paraquat treatment (Mangano et al., 2011).

5.3 Uncertainties or inconsistencies

- Mice deficient in microglia (depletion by a ganciclovir-thymidine kinase system under the CD11b promoter) were still susceptible to MPTP toxicity, while mixed cell cultures prepared from these deficient mice showed partial protection (Kinugawa et al., 2013).
- Although some publications show strong protection by COX-2 inhibition/deletion, others showed that mice deficient for COX-2 were partly protected against MPTP-induced decrease of
DAergic neurons in substantia nigra, but not against DA terminal loss in striatum (Feng et al., 2000).

- Mice deficient in IL6 (IL6-/−) showed an increased vulnerability of the nigrostriatal pathway following MPTP treatment associated to a normal astrogliosis but a transient microgliosis, suggesting that transient microgliosis and IL6 may have also protective effects (Cardenas and Bolin, 2003).

- MMTV integration site 1 (Wnt 1) is a key transcript involved in DAergic neurodevelopment, and is dynamically regulated during MPTP-induced DA degeneration and glial activation. MPTP-activated astrocytes of the ventral midbrain were identified as candidate source of Wnt 1 by in situ hybridization and RT-PCR in vitro, suggesting that reactive astrocytes may be rather involved in neuroprotective/neurorescue pathways, as further demonstrated by deletion of Wnt 1 or pharmacological activation of Wnt/β-catenin signaling pathway (L’Episcopo et al., 2011).

- The role of microglia, NADPH-oxidase and oxidative stress in paraquat-induced neurodegeneration is well established. Nevertheless, the mechanism connecting these three elements remain poorly understood since direct evidence for extracellular and/or intracellular formation of radical paraquat and superoxide is controversial.

- Rotenone (1-3 nM) applied directly on BV2 microglial cells increased their phagocytosis and the release of pro-inflammatory cytokines (TNF-alpha, IL-1 beta), suggesting that microglial cell can also be a primary target of rotenone (Zhang et al., 2014). However, these results in a transformed microglial cell line contrast with the experiments performed on isolated primary microglial cells, where rotenone (10-50 nM) was not able to trigger a direct activation (Klintworth et al., 2009).

- The regulation of inducible nitric oxide synthase (for production of peroxynitrite) differs strongly between rodents and human, and thus, the role of NO in human remains unclear (Ganster et al., 2001).

- While in human long-term use of anti-inflammatory drugs (NSAIDs, aspirin, iboprufen) for preventing PD onset or for slowing the progression is still controversial, a new strategy is emerging aiming at targeting microglial cells by modulating their activity, rather than simply trying to counteract their inflammatory neurotoxicity. The advantage of this therapeutic approach could be to reduce neuroinflammation and neurotoxicity, while at the same time strengthening intrinsic neuroprotective properties (Pena-Altamira et al., 2015).

### 5.4 Quantitative relationship

As it is rather the features and the duration of the inflammatory response that determine the extent of the nigrostriatal pathway neurodegeneration, the best way to propose a quantitative or semi-quantitative evaluation of the links between KE<sub>up</sub> and KE<sub>down</sub> is to use studies where any feature of neuroinflammation is inhibited and to quantify the protection of the Daergic neurons and terminals. Thus it will give an evaluation of how much neurodegeneration depends on the neuroinflammatory process. Below are some examples for illustration.

#### Table 4: Quantitative evaluation of the KER

<table>
<thead>
<tr>
<th>KE upstream Neuroinflammation</th>
<th>KE downstream Neurodegeneration of dopaminergic nigrostriatal pathway</th>
<th>Reference</th>
<th>Type of study</th>
<th>Comment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Inhibition of any feature of neuroinflammation (microglia/astrocyte)</td>
<td>How much nigrostriatal pathway degeneration depends on KE&lt;sub&gt;up&lt;/sub&gt; as assessed by protection when any KE&lt;sub&gt;up&lt;/sub&gt; feature is inhibited</td>
<td></td>
<td></td>
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<tr>
<td>K&lt;sub&gt;A TP&lt;/sub&gt; channel opener</td>
<td>TH immunoreactivity : Total recovery</td>
<td>Zhou et al., 2007</td>
<td>In vivo Rotenone 2.5 mg/kg/d + in vitro</td>
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<tr>
<td>(iptakalim) induced decrease of TNF-alpha and COX2 mRNA expression and TNF-alpha content, as well as microglial reactivity (OX42, ED1)</td>
<td>DA uptake TH immunoreactivity About 50% more neuronal death in presence of glia (80 % of protection with apocynin)</td>
<td>Gao et al., 2002</td>
<td>In vitro Rotenone 5-20 nM</td>
<td></td>
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<tr>
<td>NADPH oxidase</td>
<td>DA uptake TH immuno : 20% protection</td>
<td>Gao et al., 2003</td>
<td>In vitro Rotenone 5-10 nM</td>
<td></td>
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<tr>
<td>Neuron enriched cultures Neuron-Glia co-cultures +apocynin</td>
<td>Protection of DA terminals in striatum, but no effect in substantia nigra</td>
<td>Salama et al., 2012</td>
<td>In vivo Rotenone 3mg/kg/d</td>
<td></td>
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<tr>
<td>NADPH oxidase</td>
<td>Protection of neuron : 40% cell viability 50-60% TH immuno + number of dendrites</td>
<td>Chang et al., 2013</td>
<td>In vitro Rotenone 30 nM MPP+ 0.1 microM</td>
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<tr>
<td>Mice knockout for NADPH ox gp91&lt;sup&gt;77&lt;/sup&gt;</td>
<td>DA uptake and TH immuno : 30-40 % of protection</td>
<td>Wang et al., 2014</td>
<td>In vitro LPS 20 ng/ml MPP+ 0.15 microM</td>
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<tr>
<td>Myeloperoxidase</td>
<td>MPP+ caused 40% decrease of TH+ neurons Nicotine induced a 30% recovery</td>
<td>Liu et al., 2012, 2015</td>
<td>In vivo MPTP 20mg/kg Nicotine 5mg/kg In vitro on isolated microglia and astrocytes</td>
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<tr>
<td>(HOCI from H2O2)</td>
<td>TNF-alpha of microglial origin</td>
<td>Borrajo et al., 2013</td>
<td>In vitro + in vivo MPP+ 0.25 microM</td>
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<tr>
<td>Resveratrol decreased NO, ROS, phagocytosis in microglia and astrocytes</td>
<td>20 % of recovery of TH immunoreactivity</td>
<td>Liu et al., 2015</td>
<td>In vitro Co-cultures MPP+ 5 microM indirect</td>
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<td>Phagocytic signaling between neuron and microglia i.e. block of vitronectin and P2Y6 on microglia or annexin or phophatidylserine on neuron (eat-me signal)</td>
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<td>Decrease in the number of activated microglia by L-thyroxin</td>
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<td>in substantia nigra, not in striatum</td>
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<tr>
<td>NADPH oxidase : NOX2</td>
<td>20 % of recovery of TH immunoreactivity</td>
<td>Borrajo et al., 2013</td>
<td>In vitro + in vivo MPP+ 0.25 microM</td>
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<tr>
<td>Diphenyleneiodonium : long acting NOX2 inhibitor</td>
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<td>Control of microglial and astrocyte reactivity by Alpha 7 nicotinic Ach receptor</td>
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<td>present on microglia and astrocyte Its activation decreased microglial and astrocyte reactivity</td>
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<td>TNF-alpha of microglial origin</td>
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<tr>
<td>By blocking angiotensin-1 receptors, NADPH-oxide, Rho-kinase and NF.kB</td>
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<tr>
<td>Infusion of the anti-inflammatory cytokine TGF beta</td>
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<td>protects from MPP+ induced cell loss by decreasing CD11b, i-NOS, TNFalpha, IL+ beta, and increasing IGF-1. Silencing of TGFbR1 gene abolished the protective effect</td>
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</table>

**K<sub>A TP</sub> channel opener** (iptakalim) induced decrease of TNF-alpha and COX2 mRNA expression and TNF-alpha content, as well as microglial reactivity (OX42, ED1)

**NADPH oxidase**
- Neuron enriched cultures
- Neuron-Glia co-cultures +apocynin

**NADPH oxidase**
- Mice knockout for NADPH ox gp91<sup>77</sup>
- Co-culture neuron-glia

**Phagocytic signaling** between neuron and microglia i.e. block of vitronectin and P2Y6 on microglia or annexin or phosphatidylserine on neuron (eat-me signal)

**Decrease in the number of activated microglia by L-thyroxin**
- In substantia nigra, not in striatum

**Myeloperoxidase**
- (HOCI from H2O2)
- Resveratrol decreased NO, ROS, phagocytosis in microglia and astrocytes

**NADPH oxidase : NOX2**
- Diphenyleneiodonium : long acting NOX2 inhibitor

**Control of microglial and astrocyte reactivity by Alpha 7 nicotinic Ach receptor**
- present on microglia and astrocyte
- Its activation decreased microglial and astrocyte reactivity

**TNF-alpha of microglial origin**
- By blocking angiotensin-1 receptors, NADPH-oxide, Rho-kinase and NF.kB

**Infusion of the anti-inflammatory cytokine TGF beta**
- protects from MPP+ induced cell loss by decreasing CD11b, i-NOS, TNFalpha, IL+ beta, and increasing IGF-1. Silencing of TGFbR1 gene abolished the protective effect
| **i-NOS inhibition**  
caused a decrease of astrocyte and microglial reactivity as assessed by GFAP and OX6, respectively  
(n-NOS inhibition had no effect) | TH immunoreactivity  
Dose-dependent recovery with 1400W (0.1-100 microM) | Brzozowski et al., 2015 | In vitro  
MPP+  
43 microM |
| **Inhibition of laminin receptor**  
on microglia  
i.e. regulating cell-ECM interactions induced a decrease of microglia phagocytosis and of O$_2^-$ production | Dose-dependent partial recovery (about 35% of TH immunoreactivity) | Wang et al., 2006 | In vitro  
MPP+  
0.1-0.5 microM |
| **Inhibition of glial activation-mediated oxidative stress**  
by Fluoxetine, anti-depressant) | 30% of recovery of TH immunoreactivity in Substantia nigra and total recovery of DA terminals in striatum | Chung et al., 2011 | In vivo  
MPTP 20 mg/ kg ip |
| **Mice lacking both TNFR**  
Induced a decrease of GFAP in striatum  
Double KO, if only KO for TNFR1 or TNFR2, no protection | TH staining in striatum, DA content and GFAP staining, all returned to control level | Siriram et al., 2014 | In vivo  
MPTP 12.5 mg/kg sc |
| **Mice-deficient for COX2**  
Microglial cells are the major cells expressing COX2 | MPTP caused in substantia nigra  
40% loss in wild type  
45% loss in COX1/-  
20% loss in COX2/-  
in striatum  
70% loss of DA in all 3 types of mice | Feng et al., 2002 | In vivo  
MPTP 20 mg/kg sc |
| **S100B+/- in astrocytes**  
caused decreased microgliosis, TNF-alpha and RAGE | 12% of protection for TH+ neuron  
30% of protection for Nissl-labelled neurons | Sathe et al., 2012 | In vivo  
MPTP 30 mg/kg ip |
| **Glia Maturation Factor (GMF)**  
overexpression  
or  
GMF-/- showed decreased TNF-alpha, IL-1beta, ROS and NFkappaB downregulation | Overexpression of GMF exacerbate DA neuron degeneration  
GMF-/ - induced a protection of 40% of TH+ neurons | Khan et al., 2014 | In vitro  
Mesencephalic neuron/glia cultures  
MPP+ 5,10,20 microM |
| **Pharmacological inhibition or deletion of CD95 in peripheral myeloid cells**  
(monocytes, macrophages, microglia, leucocytes) hampered infiltration in the brain of peripheral myeloid cells | Total preservation of DA level in striatum  
Total protection of TH+ neurons in Snigra (25% affected in wild type mice) | Gao et al., 2015 | In vivo  
MPTP 30 mg/kg ip |
| **Glucocorticoid receptor (GR)**  
deletion in microglia  
increased their reactivity and induced a persistent activation | 2X aggravation of TH+ neuronal loss in Snigra | Ros-Bernal et al., 2011 | In vivo  
MPTP 20 mg/kg ip |
| **TNF -/- mice**  
No protection in substantia nigra TH density in striatum: return to control level | | Ferger et al., 2004 | In vivo  
MPTP 20 mg/kg ip |
| **Intra-venous transplantation of mesenchymal stem cells**  
Cell migration in substantianigra and release of TGFbeta (anti-inflammatory)  
Reparation of BBB, decreased | About 15% protection of TH+ neurons in Snigra | Chao et al., 2009 | In vivo  
MPTP 20 mg/kg ip |
<table>
<thead>
<tr>
<th>Evidence Supporting Taxonomic Applicability</th>
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<tbody>
<tr>
<td>Rodent models have been mainly used to study the impact of neuroinflammation on DAergic nigrostriatal pathway degeneration, without any sex restriction. Neuroinflammation preceding neuronal death was detected in monkeys exposed to MPTP (Barcia et al., 2011); and in human, neuroinflammation is considered as an early event in the disease process (Innaccone et al., 2012).</td>
</tr>
</tbody>
</table>
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otenone and paraquat do not directly activate microglia or lth 8(7): 

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resistant to 1-methyl-4-phenyl1, 2, 3, 6-tetrahydropyridine-induced damage of dopaminergic 


(TNF-alpha) and pharmacological inhibition of TNF-synthesis attenuates MPTP toxicity in mouse 


Ferris CF, Marella M, Smerkers B, Barch 


expressed in Parkinson's disease and induces, in vitro, production of nitric oxide and tumor 


activation in very early dementia with Lewy bodies, comparison with Parkinson's disease. 


prevents 1-methyl-4-phenylpyridinium (MPP(+))-induced loss of mesencephalic dopaminergic 

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CD11b contribute to the microglial-induced death of dopaminergic neurons in vitro but not in vivo 


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exposure and neurotoxicity. International journal of environmental research and public health 8(7): 

2980-3018.


6th KER: Degeneration of dopaminergic neurons of the nigrostriatal pathway directly leads to neuroinflammation

6.1 How does this KER work?

Several chemokines and chemokine receptors (fraktalkine, CD200) control the neuron-microglia interactions and a loss of this control on the side of neurons can trigger microglial reactivity without any further positive signal required (Blank and Prinz, 2013; Chapman et al., 2000; Streit et al., 2001). Upon neuronal injury, signals termed "Damage-Associated Molecular Patterns (DAMPs)" are released by damaged neurons to promote microglial reactivity (Marin-Teva et al., 2011; Katsumoto et al., 2014). These are for instance detected by Toll-like receptors (TLRs) (for review, see Hayward and Lee, 2014). TLR-2 functions as a master sensing receptor to detect neuronal death and tissue damage in many different neurological conditions including nerve transection injury, traumatic brain injury and hippocampal excitotoxicity (Hayward and Lee, 2014). Astrocytes, the other cellular actor of neuroinflammation besides microglia (Ranshoff and Brown, 2012) are also able to sense tissue injury via e.g. TLR-3 (Farina et al., 2007; Rossi, 2015), and neuronal injury can result in astrocytic activation (Efremova, 2015).

6.2 Weight of evidence

6.2.1 Biological Plausability

Kreutzberg and coworkers (1995, 1996) showed that neuronal injury generally leads to activation of microglia and astrocytes. This is a general phenomenon: for instance it is always observed in ischemic damage (stroke; often in the form of glial activation following neuronal injury (Villa 2007)) as well as in stab or freeze injuries (Allahyari and Garcia, 2015). It is also observed regularly when neurons are killed by highly specific neurotoxicants that do not affect glia directly, such as injection of quinolinic acid or of 6-hydroxydopamine into the striatum (Hernandez-Baltazar et al., 2013; Arlicot et al., 2014).

The vicious circle of neuronal injury triggering glial activation and glial activation triggering/enhancing neurodegeneration is often assumed to be a key element in the pathogenesis of neurodegenerative diseases, not just PD, but also (Alzheimer's disease, prion disease and many others) (Hirsch and Hunot, 2009; Tansey and Goldberg, 2009; Griffin et al., 1998; McGeer and Mc Geer, 1998; Blasko et al., 2004; Cacquevel et al., 2004; Rubio-Perez and Morillas-Ruiz, 2012; Thundyil and Lim, 2014; Barbeito et al., 2010).

6.2.2 Empirical support for linkage

MPP+

The chemokine fractalkine (regulating neuron-glia interactions) was found to be released by neurons after unilateral injection of MPP⁺ in substantia nigra. It induced microglial activation by binding on the microglial receptor CXCR1 (Shan et al., 2011). Similarly, in chronically MPTP-injected macaques, metalloproteinases-9 (MMP-9) released by injured neurons favor glial activation (Annese et al., 2015). Advanced glycation endproducts (AGEs), which are endproducts of reactions involving ROS, colocalized with DAergic neurons 2 days post last MPTP injection, suggesting neuronal injury (Teismann et al., 2012). In contrast, the receptors for AGEs (RAGEs) were found on microglial cells and astrocytes (Teismann et al., 2012). RAGE can activate NF-kappaB, the transcription factor involved in the inflammatory response (Abdelsalam and Safar, 2015). Ablation of RAGE proved to be protective against MPTP-induced decreases of TH⁺ neurons, by decreasing NF-kappaB p65 nuclear translocation and by mitigating microglia and astrocyte reactivities (Teismann et al., 2012).

Rotenone

Rotenone-induced neurotoxicity was less pronounced in neuron-enriched cultures, than in neuron-glia co-cultures (Gao et al., 2002), suggesting that neuron-glia interactions are critical for rotenone-induced neurodegeneration. Indeed, CD200-CD200R signaling regulates neuron-glia interactions and holds microglia in a quiescent state (Biber et al., 2007). Therefore, inhibition of CD200R by blocking antibodies increased rotenone-induced DA neurotoxicity in neuron-glia mesencephalic co-cultures (Wang et al., 2011). Aging is associated with a decrease of CD200 expression (Wang et al., 2011) and...
deficits in neuronal CD200 production is also observed in several animal models of Parkinson’s disease (Sung et al., 2012; Wang et al., 2011; Zhang et al., 2011). Inhibition of RAGE, which is upregulated in the striatum following rotenone exposure and in response to neuroinflammation, decreases rotenone-induced apoptosis by decreasing mitochondrial cytochrome c release and caspase-3 activation and suppresses NF-kappaB activation, as well as the downstream inflammatory markers TNF-alpha, i-NOS and myeloperoxidase (Abdelsalam and Safar, 2015), showing again intermingled links between neuronal injury/death and neuroinflammation.

**Paraquat**

Non-lethal neuronal damage is sufficient to trigger neuroinflammation: in 3D rat brain cell cultures, repeated treatment with concentrations of paraquat that did not kill the neurons, microglia and astrocytes were activated (Sandström et al., 2014). Paraquat alone (10mg/kg, 2x/week, for 4 weeks) or in combination with maneb (30 mg/kg) induces a loss of DAergic neurons in the substantia nigra paralleled by microglial activation (Cicchetti et al., 2005; Mitra et al., 2011). Neuronal injury is facilitated by uptake of paraquat via DA transporters (Rappold et al., 2011). In this model, paraquat-induced neuronal perturbations are sufficient to induce neuroinflammation, but then neuroinflammation exacerbates the neurodegenerative process (Purisai et al., 2007).

### 6.3 Inconsistencies and uncertainties

- Triggering of glia by injured neurons may not necessarily be due to the damage of neurons, but it may also be due to released synuclein (Sanchez-Guajardo, 2010).

- In a AAV alpha-synucleinopathy model, it was shown that cytoskeletal perturbation and accumulation of alpha-synuclein were sufficient to induce microglial reactivity, suggesting that neuroinflammation appears early in the disease process and is not a result triggered by cell death (Chung et al., 2009).

- Direct effects of toxicants on glia cannot be completely excluded. They have been reported for most toxicants in one or the other publication (rotenone, paraquat, MPP⁺) (Zhang et al., 2014; Rappold et al., 2011; Brooks et al., 1989). The overwhelming evidence speaks against such effects for rotenone and MPP⁺ (Klintworth et al., 2009), but for paraquat there is evidence of direct interaction with microglial membrane NADPH oxidase (Rappold et al., 2011).

- As paraquat has several MIE (Czerniczyniec et al., 2015; Rappold et al., 2011), these may involve both neurons and microglia.

### 6.4 Quantitative relationship

Some examples of quantitative relationships between KE_{up} and KE_{down} are given below. For KE_{down} Neuroinflammation, only the features measured are cited, as neuroinflammation is a complex KE involving several cell types and measured by changes in the expression /release of several markers.

**Table 4:** Quantitative evaluation of the KER

<table>
<thead>
<tr>
<th>KE upstream</th>
<th>KE downstream</th>
<th>Compound</th>
<th>Reference</th>
<th>Comment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Degeneration of DAergic nigrostriatal pathway</td>
<td>Neuro-inflammation</td>
<td>MPTP 20mg/kg i.p. 4 injections at 2h intervals</td>
<td>Annese et al., 2013</td>
<td>MMP-9 released by neurons as trigger of neuroinflammation</td>
</tr>
<tr>
<td>about 25 % decrease of TH⁺ neurons 24h-72h post-injection</td>
<td>Microglial and astroglial reactivities in substantia nigra and striatum</td>
<td>MPTP 20mg/kg i.p. 4 injections at 2h intervals</td>
<td>Chung et al., 2013</td>
<td>MMP-3-induced disruption of BBB</td>
</tr>
<tr>
<td>about 60 % decrease of TH⁺ neurons in subnigra and of DA terminals in striatum 7days</td>
<td>increase in ED1⁺ cells (macrophagic microglia or invading monocytes)</td>
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Evidence Supporting Taxonomic Applicability

Beside the rodent models, the concept of vicious circle with neuronal injury leading to neuroinflammation and neuroinflammation triggering or enhancing neurodegeneration is described in several neurodegenerative diseases in human, without any sex restriction (Hirsch and Hunot, 2009; Tansey and Goldberg, 2009; Griffin et al., 1998; McGee and Mc Geer, 1998; Blasko et al., 2004; Catquevel et al., 2004; Rubio-Perez and Morillas-Ruiz, 2012; Thundyil and Lim, 2014; Barbeito et al., 2010). Aging is an aggravating factor and increases the risk for developing a neurodegenerative disease (Kawas et al., 2000; Blasko et al., 2004).
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Thundyil J, Lim KL. 2014. DAMPs and Neurodegeneration. Ageing research reviews.


7th KER: Mitochondrial dysfunction leads to the degeneration of dopaminergic neurons of the nigrostriatal pathway

7.1 How this key event relationship works

Neurons are characterized by the presence of neurites, the formation of action potentials, and the release and re-uptake of neurotransmitters into the synaptic cleft. The presence of long extensions implies a significant enlargement of total cell surface. In combination with the transmission of action potentials that require a continuous maintenance of active transport processes across the membrane, the steady state energy demand of these neurons is significantly higher compared with non-neuronal cells. Dopaminergic (DA) neurons located in the substantia nigra pars compacta (SNpc) that project into the striatum are unique with respect of the total length of their neurites and the number of synapses that are significantly higher compared with other neuronal cell types (Bolam et al., 2012). Besides this complex morphology DA neurons have a distinctive physiological phenotype that could contribute to their vulnerability (Surmeier et al., 2010). Other features such as high energy demand, high calcium flux, dopamine autoxidation process as well as high content of iron and high content of microglia makes these DA neurons at vulnerable population of cells to oxidative stress produced by mitochondrial dysfunction. These architectural features of SNpc DA neurons render this cell type as particularly vulnerable to impairments in energy supply. Mitochondrial dysfunction, either evoked by environmental toxins such as the complex I inhibitor rotenone or MPTP, by oxidative modifications of components of the mitochondrial respiratory chain, or by genetic impairments of mitochondrial ATP generation hence have direct influence on the function and integrity of SNpc DA neurons.

7.2 Weight of evidence for the KER

7.2.1 Biological plausibility

Mitochondria are organelles essentials for multiple cellular processes, including production of ATP, maintenance of calcium homeostasis, management of ROS production and apoptosis. Mitochondrial dynamics are also critical for the maintenance of cellular homeostasis, which involve multiple factors controlling mitophagy (Youle et al. 2012). Deregulation of mitochondrial functions may impact any...
neuronal population; however, SNpc DA neurons are indeed the most vulnerable population in PD.

Multiple factors are related to their vulnerability: These include autonomous activity, broad action potentials, low intrinsic calcium buffering capacity, poorly myelinated long branched axons and terminal fields, and use of a catecholamine neurotransmitter, often with the catecholamine-derived neuromelanin pigment (Sulzer et al. 2013; Surmeier et al.2010).

The above mentioned factors imply a significantly higher total cell surface and a high energy requirement in order to maintain the re-distribution of ions across the membrane following an action potential. In addition, SNpc DA neurons are characterized by significantly higher numbers of synapses compared with other neuronal types or with DA neurons of different anatomical localizations (Anden et al., 1966; Kawaguchi et al., 1990; Kita et al., 1994; Bevan et al., 1998; Wu et al., 2000; Tepper et al., 2004). In humans, ca. 10 times higher numbers of synapses compared with rats are expected, making human DA neurons particularly vulnerable (Bolam et al., 2012; Matsuda et al., 2009). These extreme bioenergetics demands pose SNpc DA neurons energetically "on the edge". Any stressor that might perturb energy production would hence lead to conditions under which the energy demand would exceed energy supply, resulting in cell damage and ultimately to cell death.

The mechanistic link between mitochondrial dysfunction and loss of SNpc DA neurons also comes from evidence of mutated proteins related to mitochondrial function in familial PD, resulting in reduced calcium capacity, increased ROS production, increase in mitochondrial membrane permeabilization and increase in cell vulnerability (Koopman et al. 2012; Gandhi et al. 2009). In addition, excessive ROS production can damage mitochondrial DNA and activate the intrinsic pathway of apoptosis (Tait et al. 2010). Additional sources of oxidative stress come from the autodestruction of dopamine and the active generation of ROS by activated glia cells; however, the mitochondrial respiratory chain itself represents a source of constant superoxide formation, even under normal conditions (Moosmann et al., 2002).

Furthermore, imbalance of mitochondrial dynamics have been reported in a wide range of experimental models of PD and inhibition of the mitochondrial fission proteins (i.e. Drp1) promote mitochondrial fusion and fission and enhanced the release of dopamine from the nigrostriatal terminals (Tieu et al. 2014).

Additional link between mitochondrial dysfunction and the degeneration of DA neurons of the nigrostriatal pathway comes from studies indicating a reduced activity of mitochondrial complex I in human idiopathic PD cases in the substantia nigra (Keeney et al., 2006; Parker et al., 1989, 2008; Swerdlow et al., 1996). The impairment in complex I activity was directly correlated with an elevated sensitivity of SNpc DA neurons and their demise. Transfer of mitochondria from human platelets collected from idiopathic PD subjects into fibroblasts or neuronal cells resulted in elevated levels of basal oxidative stress, a declined supply with ATP, and an elevated vulnerability towards exogenous stressors such as the complex I inhibitors rotenone or the redox cycler paraquat (Swerdlow et al., 1996; Gu et al., 1998). Systemic application of complex I inhibitors such as rotenone or MPTP lead to a preferential loss of nigrostriatal DA neurons, while other brain areas or peripheral cells are not affected to the same degree (Langston et al., 1983).

### 7.2.2 Empirical support for linkage

The experimental support linking mitochondrial dysfunction with the degeneration of DA neurons of the nigrostriatal system is based on the analysis of mitochondria from PD patients, from genetic mouse models, from in vitro knockdown and overexpression systems, and from in vitro and in vivo toxin models.

- **In vitro/rotenone:** Prevention of ROS formation protects from cell death. The concept of mitochondrial dysfunction as a consequence of defects in complex I has been fueled by observations of impaired complex I activity in the SNpc, muscle, and in platelets of PD patients. Human neuroblastoma SK-N-MC cells, exposed to rotenone, displayed a time- and concentration-dependent decline in viability. Transfection of rotenone-insensitive single subunit NADH dehydrogenase (NDI 1) allowed a replacement of endogenous complex I activity. NDI 1 transfected cells showed no oxidative damage, no declined mitochondrial activity, or cell death. A significant amount of endogeneously formed ROS at complex I was identified in SK-N-MC cells and in a chronic midbrain slice culture exposed to rotenone.
Antioxidants such as α-tocopherol prevented cell death evoked by rotenone, but not the rotenone-induced drop in ATP (Sherer et al. 2003).

- In vitro/rotenone/MPP⁺: Antioxidants protect from rotenone/MPP⁺ cell death. Analysis of post-mortem nigrostriatal material from PD patients regularly revealed the presence of elevated levels of oxidative modified proteins, lipids, and DNA. These observations indicate an elevated formation of ROS in the cells affected by the disease and triggered the concept of antioxidants as a potential intervention strategy to slow down the progression of PD. In MES23.5 cells, a reduction in viability, DA content, NADH levels, as well as an increase in ROS formation and elevated nuclear condensation was observed upon treatment with MPP⁺. Rosmarinic acid is well known for its radical scavenging activities and displayed a complete protection from MPP⁺-mediated cell death and rescued NADH levels. In addition, it lead to a partial protection from the loss of DA and resulted in a rate of nuclear condensation that was about half of that observed with MPP⁺ alone (Du et al. 2010).

- The flavonoid rutin has been demonstrated to protect from oxidative stress in 6-OHDA induced motor deficits in rats as well as to inhibit the formation of nitric oxide and proinflammatory cytokines (Khan et al., 2012). In a model of SH-SY5Y cells, exposure to rotenone lead to a reduction in viability by ca. 50% that was almost completely protected in the presence of rutin. Rotenone-dependent increase of ROS formation and an elevation of intracellular Ca²⁺ was significantly dampened by the presence of rutin, similar to its rescue from rotenone-dependent decrease in mitochondrial membrane potential (Park et al., 2014).

- Comparable observations were made with the quinone triterpene celastrol that protected SH-SY5Y cells exposed to rotenone almost completely from cell death, from a rotenone-dependent elevation in ROS levels, and from a rotenone-dependent loss of the mitochondrial membrane potential (Choi et al., 2014).

- In vitro/different complex I inhibitors: Inhibition of complex I triggers oxidant formation and cell death. The majority of experimental PD studies were either conducted using rotenone or MPP⁺. In order to demonstrate that the concept of complex I inhibition and its ROS-mediated triggering of mitochondrial dysfunction and cell demise can be regarded as a general principle, alternative complex I inhibitors were applied to substantiate previous observations made with rotenone. In human SK-N-MC neuroblastoma cells, rotenone as well as the pesticides fenazaquin, fenpyroximate, pyridaben, tebufenpyrad, pyridaben were tested. In all cases, a time- and concentration-dependent decrease in intracellular ATP and cell viability was observed. Expression of the rotenone-insensitive NADH dehydrogenase from Saccharomyces cerevisiae (NDI 1) prevented from the toxicity of the different complex I inhibitors completely.

- In vitro/rotenone: Mitochondrial dysfunction-dependent cell death is prevented by antioxidants. In a human neuroblastoma SH-SY5Y model, exposed either to the complex I inhibitors MPP⁺ or rotenone, the imine antioxidants iminostilbene, phenothiazine, phenoxazine in the low nanomolar concentration range partially protected from MPP⁺ or rotenone toxicity. A reduction in the membrane potential evoked by MPP⁺ and rotenone was completely prevented by these antioxidants (Hajteva et al., 2009).

- In vitro/rotenone: Circumvention of dysfunctional mitochondria protects from cell death. Assuming a direct causal relationship between complex I inhibition, mitochondrial dysfunction, and the demise of DA neurons, the circumvention of endogenous complex I by expression of the NADH dehydrogenase of Saccharomyces cerevisiae (NDI 1) provided initial evidence for the essential role of complex I inhibition in this sequence of events. As an alternative electron carrier, capable of transferring electrons from NADH to cytochrome c, methylene blue was identified. In hippocampal HT-22 cells, a rotenone-mediated reduction in the oxygen consumption rate was completely reversed by the addition of methylene blue. A rotenone-mediated decline in cell viability by 70% was almost completely prevented by 0.1 µg/ml methylene blue. In rats, rotenone-mediated decline in striatal DA was entirely prevented by methylene blue, the observed elevation of ROS formation evoked by rotenone was reduced to control levels, and rotarod performance impairments evoked by rotenone were completely
avoided by administration of methylene blue. These observations illustrate a causal relationship between dysfunctional mitochondria, the degeneration of nigrostriatal DA neurons, and impaired motor performance (Wen et al. 2011).

- In vivo/rotenone: Circumvention of dysfunctional mitochondria prevents from nigrostriatal cell degeneration. Circumvention of a dysfunctional complex I by the rotenone-insensitive NADH dehydrogenase NDI 1 in vivo and its influence on nigrostriatal DA neuron integrity was demonstrated in a rat model with an unilateral injection of a recombinant adeno-associated virus, carrying the NDI 1 gene into close special vicinity to the SNpc. The animals were treated with rotenone after the unilateral expression of NDI 1. NDI 1 almost completely prevented from the rotenone-mediated loss of TH staining in the SNpc and the striatum. Striatal DA levels that were reduced by ca. 50 % by rotenone, in the presence of NDI 1, DA levels were also almost identical to the values of untreated controls. These observations highlight a causal relationship between the inhibition of complex I and the degeneration of nigrostriatal DA neurons (Marella et al. 2008).

- In vitro/DA: Exogenously added oxidants lead to mitochondrial dysfunction and cell death. Next to an elevated formation of reactive oxygen species evoked by endogenous defects in complex I or in response to pharmacological inhibitors of complex I, nigrostriatal DA neurons are characterized by the neurotransmitter dopamine and its tendency to undergo autoxidation when exposed to physiological pH and oxygen tension conditions. To assess the role of DA-mediated oxidative stress as a cause of mitochondrial dysfunction and its influence on cell viability, PC12 cells were exposed to DA. The observed increase in intracellular ROS was completely reversed by the presence of the antioxidant N-acetyl-cysteine (NAC). The amount of oxidative modified protein increased by DA treatment, its rise was completely prevented by the presence of NAC, and partially prevented by the presence of exogenously added GSH. DA-dependent PC12 cell death, decline in the transmembrane potential and in intracellular ATP, and decline in complex II/III activities were observed and were all completely prevented by the presence of NAC (Jana et al., 2011).

- In vitro/ GSH depletion: Oxidative stress causes mitochondrial dysfunction and neurodegeneration. Several reports indicated a declined activity of complex I in the brain, but also in muscle and platelets of PD patients. In order to investigate the mutual interaction between pro-oxidative conditions and complex I activity, a PC12 subclone was generated, allowing the inducible downregulation of γ-glutamyl-cystein synthetase involved in the synthesis of glutathione (GSH). This system allows a controlled decrease of intracellular GSH by ca. 50 % and a decrease in mitochondrial GSH by ca. 40 %. Under these conditions, intracellular and intramitochondrial ROS increased by ca. one third, mitochondrial complex I activity and ATP levels were reduced by ca. two thirds. The observed inhibition of complex I was completely reversed by DTT. These observations indicate that an impairment of complex I activity as a key event in the initiation of mitochondrial dysfunction and ultimately cell death, can be evoked by elevated levels of oxidants, respectively by a declined cellular antioxidant capacity (Jha et al., 2000).

- In vitro/ GSH depletion: Oxidative stress causes mitochondrial dysfunction and neurodegeneration. PD is characterized by the depletion of glutathione (GSH) in the SNpc. Declined cellular levels of GSH were reported to be associated with morphological changes of mitochondria (Perry et al., 1982; Jain et al., 1991). To investigate the influence of declined GSH levels, N27 cells were exposed to buthionine-S-sulfoximine (BSO), an inhibitor of glutamate cysteine ligase and hence of de novo GSH synthesis. The BSO concentration chosen allowed a reduction in intracellular GSH levels by 50 % in the absence of cell death. Chronic GSH depletion resulted in the S-nitrosation of complex I and its inhibition. Both effects were completely reversed by the addition of DTT (Chinta et al., 2006).

- Isolated mitochondria: Exogenous oxidants cause mitochondrial dysfunction. In order to further address the aspect on how DA autoxidation contributes to mitochondrial dysfunction and DA neurodegeneration, isolated rat brain mitochondria were exposed to DA, resulting in an inhibition of complex I by ca. 30 % and in an inhibition of complex IV by ca. 50 %. Both activities of complex I and complex IV were completely protected from DA-dependent inactivation by the presence of GSH. These observations point to a direct inhibitory action of...
endogenous DA and its autoxidation derivatives on the activity of the mitochondrial respiratory chain. (Khan et al., 2005)

- In vitro/cybrid cells: Sensitization of neuronal cells for degeneration by transfer of dysfunctional mitochondria. In a subclone of human neuroblastoma cells (SH-SY5Y), devoid of mitochondrial DNA, mitochondria from platelets of PD patients were transplanted. Analysis after 5-6 weeks in culture after transplantation of mitochondria indicated a 20 % reduction in complex I activity, a 2-fold increase in the basal formation of reactive oxygen species, and a ca. 2-fold higher sensitivity towards the mitochondrial PD toxin MPP+ (Swerdlow et al., 1996)

- In vitro/cybrid cells: Sensitization of neuronal cells for degeneration by transfer of dysfunctional mitochondria. In a subclone of the human A549 cell line, devoid of mitochondrial DNA, mitochondria of platelets from PD patients were transplanted. Complex I activity in platelets of PD patients displayed a reduction of 25 % compared with age-matched controls. After transplantation into the A549 cells, complex I activity was reduced by 25% in its activity (Gu et al., 1998)

- In vivo: Induction of mitochondrial dysfunction by Drp1 deletion leads to neuronal cell loss. Maintenance of functional mitochondria in a cell is regulated by fission/fusion processes that allow the elimination of damaged mitochondria and the spreading of intact mitochondria. Deletion of the central fission protein dynamin related protein 1 (Drp1) leads to an elimination in DA neuron terminals in the caudate putamen and to a loss of DA neuron cell bodies in the midbrain. In Drp1 deficient mice, mitochondrial mass decreases, particularly in axons (Berthet et al., 2014)

- In vivo: Induction of mitochondrial dysfunction by Tfam knockdown leads to neuronal cell loss. Mitochondrial transcription factor A (Tfam) is a key regulator of mitochondrial biogenesis. Conditional knockout mice with a selective disruption of the gene for mitochondrial Tfam in DA neurons indicated a reduction in mtDNA levels and deficiencies in the respiratory chain in midbrain DA neurons that progressed to DA cell death. The demise of DA neurons in the SNpc was associated with the onset of PD symptoms such as a reduction in locomotor activity of these mice by ca. 30 %. The decrease in locomotor activity was reversed by L-DOPA treatment (Ekstrand et al., 2007)

- In vivo: MPTP dependent mitochondrial dysfunction and cell death is protected by PGC-1α overexpression. Peroxisome proliferator-activated receptor gamma coactivator 1α (PGC-1α) is a key regulator of mitochondrial biogenesis and metabolism. Transgenic mice overexpressing PGC-1α show protection against MPTP intoxication (50 %). The SNpc in these mice is characterized by elevated levels of SOD2, Trx2. Resveratrol is a known activator of SIRT1, leading to enhanced PGC-1α gene transcription. In MPTP mice, resveratrol protected TH-positive neurons by 80% from cell loss (Mudo et al., 2012)

- In vivo: Prevention of mitochondrial dysfunction protects from nigrostriatal cell loss. In order to demonstrate the causative connection between complex 1-dependent mitochondrial dysfunction and the degeneration of DA neurons, a series of in vivo experiments were conducted that indicated partial restoration by antioxidants or by compounds supporting a dysfunctional mitochondrial ATP generation. In MPTP challenged mice that additionally received Q10 treatment, a 37 % higher striatal DA level compared with the MPTP group was detected. TH positive staining in the striatum dropped by ca. 65 % after MPTP. In the MPTP + Q10 group, the loss in striatal TH staining was reduced to ca. 40 % compared with the untreated controls. (Beal et al., 1998).

- In MPTP challenged marmosets, TH positive cell body numbers were reduced by ca. 60 %, co-administration with ebselen resulted in a reduction of TH staining of only ca. 25 % (Moussaoui et al., 2000).

- In MPTP challenged mice, a reduction of striatal DA by ca. 70 % was detected. Co-treatment with creatine resulted in a reduction of DA levels of only 42 %. In the same setup, TH positive neuron number in the SNpc was reduced by 70 % in response to MPTP, in the presence of creatine, a drop of only 4 % was observed (Matthews et al., 1999).
In vivo/rotenone: Antioxidants prevent from rotenone-dependent nigrostriatal cell death. Rotenone administered subcutaneously for 5 weeks (2.5 mg/kg/d) caused a selective increase in oxidative damage in the striatum as compared to the hippocampus and cortex, accompanied by massive degeneration of dopaminergic neurons in the substantia nigra. Antioxidant polydatin (Piceid) treatment significantly prevented the rotenone-induced changes in the levels of glutathione, thioredoxin, ATP, malondialdehyde and the manganese superoxide dismutase (SOD) in the striatum, confirming that rotenone-induced mitochondrial dysfunction resulted in oxidative stress (Chen et al., 2015).

In vivo/rotenone: Degeneration of DA neurons depends on oxidative stress evoked by mitochondrial dysfunction. Many studies have shown that mitochondrial aldehyde dehydrogenase 2 (ALDH2) functions as a cellular protector against oxidative stress by detoxification of cytotoxic aldehydes. Dopamine is metabolized by monoamine oxidase to yield 3,4-dihydroxyphenylacetalddehyde (DOPAL) then converts to a less toxic acid product by ALDH. The highly toxic and reactive DOPAL has been hypothesized to contribute to the selective neurodegeneration of dopamine (DA) neurons. In this study, the neuroprotective mechanism of ALDH2 was observed as overexpression of wild-type ALDH2 gene, but not the enzymatically deficient mutant ALDH2*2 (E504K), reduced rotenone-induced DA neuronal cell death. Application of a potent activator of ALDH2, Alda-1, was effective in protecting against rotenone-induced (100 nM, 24 hr exposure) apoptotic cell death in both SH-SY5Y cells and primary cultured substantia nigra (SN) DA neurons. These results were confirmed by in vivo studies. Intraperitoneal administration of Alda-1 to C57BL/6 mice treated with rotenone (50 mg/kg/day, oral administration for 14 days) or MPTP (40 mg/kg/day, i.p. for 14 days) significantly reduced death of SN tyrosine hydroxylase-positive dopaminergic neurons. The attenuation of rotenone-induced apoptosis by Alda-1 resulted from decreasing ROS accumulation, reversal of mitochondrial membrane potential depolarization, and inhibition of activation of proteins related to mitochondrial apoptotic pathway. The present study demonstrates that rotenone or MPP+ induces DA neurotoxicity through oxidative stress. Moreover, Alda-1 is effective in ameliorating mitochondrial dysfunction by inhibiting rotenone or MPP+ induced mitochondria-mediated oxidative stress that leads to apoptosis (Chiu et al., 2015).

7.3 Uncertainties or inconsistencies

Several in vitro studies applying rotenone to evoke mitochondrial dysfunction came to the conclusion that rotenone-dependent ROS formation, and not the rotenone-evoked drop in ATP is the primary cause for cell degeneration. These observations are largely based on experimental systems employing the rotenone insensitive NADH dehydrogenase NDI 1. Expression of NDI 1 protected rotenone exposed cells from degeneration. The presence of NDI 1 however results in a substitution of ATP. Endogenously expressed complex I is still present in these models and it can be assumed that rotenone exposure would still lead to a complex I-dependent formation of ROS that precludes the modeling of a precise cause-consequence relationship between either ATP depletion or elevated ROS levels with the demise of DA neurons.

Several studies indicate a dominant role of ROS in the degeneration of DA neurons, based on models in which rotenone/MPP+ mediated mitochondrial dysfunction and cell degeneration was protected by the presence of exogenously added antioxidants. Maintenance of the endogenous redox potential however is a highly ATP-dependent process. Clear-cut separations between the respective contribution of ROS or the role of an inhibited mitochondrial ATP synthesis on the degeneration of DA neurons is hence difficult to postulate.

Studies with chronic partial GSH depletions indicated that an experimental reduction of GSH/GSSG by ca. 50 % has no influence on cell viability. Reports involving rotenone and MPP+ however regularly observe degeneration of DA neurons under conditions of GSH depletion around 50 %. These observations indicate a more prominent role of the intracellular drop of ATP evoked by the complex I inhibitors in the process of cell degeneration.

Studies in which oxidative stress is generated e.g. by the application of DA or 6-OHDA not only observed a challenge of the cellular redox potential, but also reversible and irreversible
inhibitory mechanisms of mitochondrial respiratory chain complexes (nitration, S-nitrosation) that are accompanied by an inhibition of the respiratory chain in the absence of pharmacological complex I inhibitors. These observations illustrate the close mutual interaction between oxidative stress and the inhibition of mitochondrial respiration and point to a profound role of direct mitochondrial inhibition also under oxidative stress conditions.

- Mitochondrial dysfunction is generally associated with conditions of oxidative stress. Dysfunctional mitochondria can act as potent source of superoxide. Oxidative stress associated with PD however not only originates from mitochondrial ROS, but also from DA autoxidation and the Fenton reaction, as well as from inflammatory activated adjacent glia. Interpretations on the role of oxidative stress in DA neurons and its role in DA neurodegeneration is hence hampered by the fact that the respective origin of the reactive oxygen species formed (mitochondria, DA autoxidation, inflammation of glia cells) is rather difficult to identify and often shows overlappings (Murphy et al., 2009; Starkov et al., 2008, Cebrian et al., 2015).

- In PD patients, a reduction in complex I activity in the SNpc, but also in peripheral tissue and cells such as platelets, was reported. Studies with isolated mitochondria indicated that for efficient inhibition of mitochondrial ATP formation, an inhibition of complex I by ca. 70 % is necessary (Davey et al., 1996). Reports on the reduction of complex I activity in PD patients however repeatedly indicated an inhibition of only 25-30 % (Schapira et al., 1989; Schapira et al., 1990; Janetzky et al., 1994).

- Data available on the respective inhibition of the components of the respiratory chain are highly dependent on the experimental setup used. Analysis of mitochondrial respiratory chain complex activities in mitochondrial homogenates provide results different from data obtained with intact, isolated mitochondria. These aspects need to be considered in the interpretation of such data (Mann et al., 1992; Parker et al., 2008; Mizuno et al., 1989; Schapira et al., 1990; Cardellach et al., 1993).

### 7.4 Quantitative understanding

Quantitative understanding for this KE relationship mainly comes from in-vitro and engineered systems, using rotenone and MPTP as main chemical stressors. A clear response- response effect is evident as well as temporality was mainly supported by evidence that modulation of the KE up was attenuating or preventing the KE down. Evidence of dose relationship was limited, as most of the time a single, generally high, concentration was used.

**Table 5:** Quantitative evaluation of the KER.

<table>
<thead>
<tr>
<th>KE 2 upstream</th>
<th>KE 4 downstream</th>
<th>Comments</th>
<th>Reference</th>
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<tr>
<td>Mitochondrial membrane potential reduced by 50 % upon rotenone treatment. Back to 80 % compared to controls in the presence of the flavonoid rutin. Intracellular Ca(^{2+}) elevated by a factor of 3 by rotenone, reduction to an increase of 1.5 in the presence of rutin. ROS increased by a factor of 6.5; increase of ROS by a factor of 2 in the presence of rutin.</td>
<td>Rotenone (10 µM) resulted in a reduction of cell viability by 50 %. In the presence of rutin, cell viability was only reduced by 10 % upon rotenone treatment</td>
<td>SH-SY5Y cells exposed to rotenone (10 µM) for 24 h. When applied alone, rutin displayed no toxic effects, up to 100 µM. Rutin was added to the cells 30 min prior rotenone at concentrations from 0-10 µM</td>
<td>Park et al., 2014</td>
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Mitochondrial membrane potential reduced by ca. 66% upon rotenone treatment; in the presence of celastrol, reduction by ca. 55%.

ROS formation increased by a factor of 2 in the presence of rotenone; ROS increase by a factor of 1.5 in the presence of celastrol.

Cell viability was reduced by 50% by rotenone; in the presence of the triterpene celastrol, cell viability was only reduced by ca. 10%.

SH-SY5Y + rotenone (10 µM). Celastrol (2.5 nM) was applied 90 min prior to rotenone. Cells were incubated with the two compounds for a period of 24 h.

5 month old male Sprague-Dawley rats (ca. 500 g) received intracerebral injection of recombinant adenovirus with the NADH dehydrogenase NDI 1 gene.

TH staining in the SNpc in arbitrary units:
Control (25)
Rotenone (14)
Rotenone + NDI 1 (22)

TH staining in the striatum
Control (70)
Rotenone (40)
Rotenone + NDI 1 (65)

DA levels in the striatum:
Control (2.5)
Rotenone (1.3)
Rotenone + NDI 1 (2.2)

Marella et al., 2008

MPP⁺ experiments

Decline in mitochondrial transmembrane potential by MPP⁺; 50% prevention from this decline by rosmarinic acid.

NADH levels were reduced by ca. 50% in the presence of MPP⁺; loss of NADH was completely prevented by the presence of rosmarinic acid.

ROS levels increased by 50% in the presence of MPP⁺. Rosmarinic acid lead to a reduced increase of ROS by only 20% compared with the untreated control.

Cell viability reduced by MPP⁺ by 30%, complete protection by the presence of the antioxidant rosmarinic acid.

Striatal DA content reduced by 40% by MPP⁺ treatment, partially protected by rosmarinic acid back to a value of 25% reduction compared with the untreated control.

MES23.5 cells exposed to MPP⁺ (200 µM) for 24 h.

Rosmarinic acid (1 nM) was applied 30 min prior to MPP⁺ treatment.

Du et al. 2010
Reduction in mitochondrial membrane potential by 60% (MPP⁺), by 50% (rotenone), complete recovery by the co-incubation with ISB, PHT, PHO

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Effect on Cell Viability</th>
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<tbody>
<tr>
<td>SH-SY5Y + MPP⁺</td>
<td>Cell viability reduced by 66%; ISB, PHT, PHO partially protected from cell death with a reduction in cell viability by ca. 20%</td>
</tr>
<tr>
<td>SH-SY5Y + rotenone</td>
<td>Reduction in cell viability by 60%</td>
</tr>
<tr>
<td>Partial protection by ISB, PHT, PHO to a reduction in cell viability by 25-50%</td>
<td></td>
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<tr>
<td>SH-SY5Y + BSO</td>
<td>Reduction in cell viability by 80%</td>
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<tr>
<td>ISB, PHT, PHO partially protected with a residual decline in cell viability by ca. 20%</td>
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SH-SY5Y + MPP⁺ (200 µM) or rotenone (150 nM) or BSO (150 µM) for 60 h and 72 h.

Antioxidants tested:
- Iminostilbene (ISB)
- Phenothiazine (PHT)
- Phenoxazine (PHO)

The antioxidants were applied 2 h prior to rotenone, MPP⁺, or BSO treatment

**Circumvention of endogenous complex I**

Wt cells exposed to rotenone: increase in carbonyl content as marker of oxidative stress by 100%; completely prevented in NDI 1 expressing cells.

In midbrain slice cultures exposed to rotenone: increase in carbonyl content by 20%.

Rats exposed to rotenone: increase in carbonyl content: 27% in the striatum, increase by 41% in the midbrain.

SK-N-MC cells: rotenone evoked cell death protected by ca. 90% in NDI 1 expressing cells.

Rotenone induced cell death prevented by 80% by α-tocopherol (62.5 µM and 125 µM).

SK-N-MC human neuroblastoma cells transfected with the rotenone insensitive NADH dehydrogenase NDI 1;

Cells were treated with rotenone (100 nM) for 48 h or with BSO (10 µM) for 24 h.

When both compound were used in a combined experiment, cells were first treated with BSO (10 µM) for 24 h, then rotenone (10 nM) was added for additional 36 h.

Application of the complex I inhibitors:
- Rotenone
- Fenazaquin
- Fenpyroximate
- Pyridaben

SHT-SY5Y + MPP⁺ 200 µM or rotenone 150 nM or BSO 150 µM for 60 h and 72 h.

Antioxidants tested:
- Iminostilbene (ISB)
- Phenothiazine (PHT)
- Phenoxazine (PHO)

The antioxidants were applied 2 h prior to rotenone, MPP⁺, or BSO treatment.

**Circumvention of endogenous complex I**

**Antioxidants tested:**
- Iminostilbene (ISB)
- Phenothiazine (PHT)
- Phenoxazine (PHO)

**Application of the complex I inhibitors:**
- Rotenone
- Fenazaquin
- Fenpyroximate
- Pyridaben

**Time and concentration-dependent cell death with rotenone and a series of other complex I inhibitors:**

NDI 1 expressing cells were resistant towards the different complex I inhibitors.

SK-N-MC human neuroblastoma cells expressing the rotenone-insensitive NADH dehydrogenase NDI 1 from saccharomyces cerevisiae.

All complex I inhibitors applied were added at the concentrations: 10 nM, 100 nM, 1 µM.

Pyridaben was applied at 1 µM.
| **Tebufenpyrad** | **Pyridaben** | **µM, 10 µM, 100 µM.**
Viability was assessed after 48 h, ATP was detected after 6 h. Carbonyl content was detected after 24 h. |
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</tr>
</thead>
<tbody>
<tr>
<td>Oxygen consumption rate doubled by MB in the absence of complex I inhibitor.</td>
<td>HT22 cell viability reduced by 70 % by rotenone.</td>
<td>The study included:</td>
</tr>
<tr>
<td>Oxygen consumption reduced by 50 % by rotenone; completely reversed to control levels by the presence of MB.</td>
<td>In the presence of MB, reduction by only 10 % of cell viability was observed.</td>
<td>- Isolated rat heart mitochondria exposed to rotenone (5 µM) (instant treatment)</td>
</tr>
<tr>
<td>Complex I-III activity reduced by 95 % by rotenone. Reversed to control levels by the presence of MB.</td>
<td>In rats treated with rotenone, rotarod retention time was reduced by 50 % by rotenone. Completely reversed to control levels by the co-administration of MB.</td>
<td>- Hippocampal HT-22 cells exposed to rotenone (2-8 µM) for 24 h.</td>
</tr>
<tr>
<td>In rats, rotenone evoked a reduction of striatal DA by 50 %; completely reversed to control levels by MB</td>
<td>Complex I-III activity in the striatum of rats was reduced by 50 %, residual inhibition of 10 % observed in rats that were additionally treated with MB</td>
<td>- Rats receiving rotenone (5 mg/kg/day via osmotic minipumps for 8 days</td>
</tr>
<tr>
<td>Cybrid cells with PD mtDNA display a reduction in complex I activity by 20 %.</td>
<td>Cybrid cells: increase in basal formation of reactive oxygen species by 80%, 2-times higher sensitivity towards MPP⁺⁺ as stressor SH-SY5Y cells devoid of mtDNA; fused with platelets from PD patients for mitochondria transfer: cybrid cells.</td>
<td>Test of methylene blue (MB) (10 and 100 ng/ml in isolated mitochondria; 1 and 10 µg/ml in HT 22 cells) to circumvent the complex I/III blockade</td>
</tr>
<tr>
<td><strong>Oxidative stress causes mitochondrial dysfunction</strong></td>
<td>Treatment with MPP⁺⁺ (40 or 80 µM) for 24 h or 48 h</td>
<td>Swedlow et al., 1996</td>
</tr>
<tr>
<td><strong>Isolated mitochondria:</strong></td>
<td><strong>PC12 cells exposed to DA:</strong></td>
<td><strong>PC12 cells and isolated rat brain mitochondria exposed to dopamine (100-400 µM).</strong></td>
</tr>
<tr>
<td>Exposure to DA: loss of ca. 50 % membrane potential. Completely protected by GSH or N-acetyl-cystein (NAC)</td>
<td>Increase in intracellular ROS by a factor of 2; completely reversed by NAC</td>
<td>N-acetyl cysteine or GSH for protection were added at a concentration of 2.5 mM.</td>
</tr>
<tr>
<td>Decline of mitochondrial respiration capacity by</td>
<td>Quinoprotein formation increased by a factor of</td>
<td>In experiments including isolated mitochondria, NAC</td>
</tr>
</tbody>
</table>

Swedlow et al., 1996

Jana et al., 2011

Wen et al. 2011
90 %.

In the presence of NAC or GSH, only a reduction by 25-30 % was observed.

PC12 cells exposed to DA, then isolation and analysis of mitochondria: inhibition of complex I activity by ca. 50 %, prevented by co-incubation with NAC.

Inhibition of complex II and III; prevented by NAC.

Intact PC12 exposed to DA:

Mitochondrial transmembrane potential reduced by ca. 50 %; prevented by NAC

Intracellular ATP reduced by ca. 50 %; Cell death increased by DA by ca. 30 %, caspase 3 activity increased by a factor of 3; all increases prevented by the presence of NAC.

Reduction of intracellular GSH by 50 % and of intramitochondrial GSH by 60 % leads to:

Mitochondrial ROS increased by 30 %
ATP levels reduced by 66 %
Mitochondrial activity reduced by 66 %
State 3 respiration reduced by 60 %
Complex I activity inhibited by 60 %

No cell toxicity under the applied conditions

N27 cells exposed to BSO (2.5 µM) for 7 days:

Total glutathione was declined by ca. 50 % by this chronic treatment; absence of cell toxicity under these conditions. DTT for restoration of complex I activity was added at 1 mM.

Reduction of GSH levels by ca. 50 % result in:

Complex I inhibition by 40 %; completely reversed by DTT.

Whole cell ROS increased by 30 %

PC12 cells with inducible knockdown of glutamyl cysteine synthetase (inhibition of GSH synthesis) by addition of 25 µg/ml doxycycline.

Treatment for 24 h with doxycycline resulted in a GSH decline by ca. 50 %.

Jha et al., 2000

Chinta et al., 2006
Evidence Supporting Taxonomic Applicability

There are no sex or age restrictions for the applicability of this KE and mitochondrial are essential for most of eukaryotic cells. Rotenone and MPTp have been tested successfully in primates and mice. The mouse C57BL/6 strain is the most frequently used strain in the reported experiments. A difference in vulnerability was observed, particularly for rats, depending on the strain and route of administration. The Lewis strain gives more consistency in terms of sensitivity when compared to the Sprague Dawley. In addition to rodents, the pesticide rotenone has been also studied in Caenorhabditis elegans (C.elegans), Drosophila, zebrafish and Lymnaea Stagnalis (L.stagnalis) (Johnson et al., 2015), indicating that the system is preserved across species.
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8th KER: Degeneration of DA neurons of nigrostriatal pathway leads to parkinsonian motor deficits (bradykinesia, rigor, and tremor)

8.1 How Does This Key Event Relationship Work

Degeneration of dopaminergic (DA) neuron terminals in the striatum and the degeneration of DA neurons in the substantia nigra pars compacts (SNpc) are the defining histopathological events observed in idiopathic, familial, and toxicant-evoked cases of Parkinson's Disease (PD) (Tolwani et al. 1999; Bove et al. 2012). The loss of nigrostriatal DA neurons leads to a decline in the levels of DA in the striatum (Koller et al. 1992). Striatal DA is involved in the modulation of extrapyramidal motor control circuits. A decline in striatal DA leads to an overactivation of the two principal basal ganglia output nuclei (GPI/STN). Therefore, the inhibitory GABAergic neurons that project to thalamo-cortical structures are overactivated and inhibit cortical pyramidal motor output performance. This inhibited output activity is responsible for key clinical symptoms of PD such as bradykinesia and rigor.

8.2 Weight of Evidence

8.2.1 Biological Plausibility

The mechanistic understanding of striatal DA and its regulatory role in the extrapyramidal motor control system is well established (Alexander et al. 1986; Penney et al. 1986; Albin et al. 1989; DeLong et al. 1990; Obeso et al. 2008; Blandini et al. 2000). The selective degeneration of DA neurons in the SNpc (and the subsequent decline in striatal DA levels) have been known to be linked to PD symptoms for more than 50 years (Ehringer et al. 1960). The reduction of DA in the striatum is characteristic for all etiologies of PD (idiopathic, familial, chronic manganese exposure) and related parkinsonian disorders (Bernheimer et al. 1973); and it is not observed in other neurodegenerative diseases, such as Alzheimer's or Huntington's Diseases (Reynolds et al. 1986). In more progressive stages of PD, not only a loss of DA neuronal terminals in the striatum, but also a degeneration of the entire DA neuron cell bodies in the substantia nigra pars compacta (SNpc) was detected (Leenders et al. 1986; Bernheimer et al. 1973). The different forms of PD exhibit variations in the degradation pattern of the SNpc DA neurons. In idiopathic PD, for example, the putamen is more severely affected than the caudate nucleus (Moratalla et al. 1992; Snow et al. 2000). All different PD forms however are characterized by the loss in striatal DA that is paralleled by impaired motor output (Bernheimer et al. 1973). Characteristic clinical symptoms of motor deficit (bradykinesia, tremor, or rigidity) of PD are observed when more than 80% of striatal DA is depleted (Koller et al. 1992). These findings on the correlation of a decline in striatal DA levels as a consequence of SNpc DA neuronal degeneration with the onset of clinical PD symptoms in man provide the rationale for the current standard therapies that aim to supplement striatal DA, either by the application of L-DOPA, or by a pharmacological inhibition of the endogenous DA degradation-enzyme monoamine oxidase B (MAO-B). These treatments result in an elevation of striatal DA that is correlated with an improvement of motor performance (Calne et al 1970). The success of these therapies in man as well as in experimental animal models clearly confirms the causal role of dopamine depletion for PD motor symptoms.

8.2.3 Empirical Support for Linkage

The experimental support linking the degeneration of DA neurons of nigrostriatal pathways with the manifestation of motor symptoms characteristics of parkinsonian disorders comes from human clinical observations as well as from primates, mice and rat in vivo models using DA neuron ablation by toxicants. The levels of striatal DA corrected with the onset of PD symptoms, and dopaminergic
degeneration precede the onset of motor symptoms. The exemplary animal studies selected here are based on the use of MPTP or rotenone. The efficacy of MPTP or rotenone treatment depends on the regimen applied (acute, subacute, chronic administration), the age of the animals, and the strains used. For the interpretation of the studies, it is important that in some animal models the initial depletion of DA is only partially explained by neurite degeneration. The other contributing factors are downregulation of TH, and depletion of DA from synaptic terminals. These effects recover after 1-2 weeks. This makes the time point of measurement important for the correlation of effects. Moreover, the mouse brain has a very high plasticity after damage, so that motor deficits can recover after several weeks although there is pronounced dopaminergic neuro degeneration.

Rat in vivo models

- **Rat/rotenone**: Correlation between striatal DA, SNpc DA neurons, and motor deficits. Lewis rats exposed to systemic rotenone (3 mg/kg/ day i.p.) exhibited a loss of TH positive neurons in the SNpc by 45 %. Motor deficits were assessed by the postural instability test and by the rearing test. While 3 month old animals developed motor symptoms after 12 days of rotenone exposure, 7 month and 12 month old animals developed motor symptoms already after 6 days of exposure. Rotenone treatment elicited a progressive development of motor deficits that was reversible when treated with a DA agonist. Similar to that, the loss of rearing performance evoked by rotenone was reversed by the DA agonist apomorphine. Rotenone elicited terminal loss in the dorsolateral structures. While in the dorsolateral striatum, a significant loss of TH-positive neurites was detected, striatal cell bodies were spared from degeneration. Initial striatal DA levels (75 ng/ mg protein) dropped to 45 % following rotenone treatment (Cannon et al. 2009).

- **Rat/6-OHDA**: Destruction of nigrostriatal DA neurons. Unilateral injection of 6-OHDA into the dopaminergic nigrostriatal pathway leads to a preferential loss of DA neurons that is correlated with the onset of rotational motor deficits (Luthman et al. 1989; Perese et al. 1989; Przedborski et al. 1995).

- **Rats/rotenone**: Correlation between striatal dopamine and motor symptoms; partial reversibility by L-DOPA. Rats were exposed to 2.5 mg/kg rotenone, daily, for 48 days. Dopamine detected in the anterior striatum and posterior striatum was reduced by ca. 50 % after rotenone treatment. Rotenone treatment resulted in a significantly prolonged descent latency compared to control in the bar test and grid test. In the catalepsy test, descent latency dropped from 35 s of the controls to 5 s. In the grid test, a reduction from 30 s (control) down to 4 s (rotenone) was observed. The average distance travelled within 10 min by the animals was reduced from 37 m to 17 m in the rotenone group. Average number of rearings declined from 65 to 30; the time of inactive sitting of 270 s in controls was increased to 400 s in the rotenone group (Alam et al. 2004).

- **Rat/rotenone**: Correlation between striatal dopamine and motor symptoms. Rats were treated with rotenone either at doses of 1.5 mg/kg or 2.5 mg/kg over two months with daily i.p. injections. In the 2.5 mg/kg group, striatal DA levels dropped from 6400 pg/mg in the controls to 3500 pg/mg in the rotenone group. Rotenone treated animals showed an extended descent latency (5 to 50). In a vertical grid test, latency time increased from 9 s to 72 s (Alam et al. 2002).

- **Rats/rotenone**: Correlation between nigrostriatal TH intensity and motor symptoms. Rats were treated with different doses of rotenone for 21 days with daily i.v. or s.c. injections. In the 2.5 mg/kg group, TH intensity in the striatum dropped from 0.2 to 0.12. The average time to initiate a step increased from 5 s in the controls to 11 s in the rotenone group. Spontaneous rearing scores dropped from 80 % of the vehicle treated controls to 20 % in the rotenone group (Fleming et al. 2004).

- **Rat/rotenone**: In middle-aged rats exposed to rotenone (3 mg/kg/day for 6 days), a reduction of striatal DA levels and TH positive neurons by ca. 50 % correlated with impairments rearing performance and postural instability tests (Cannon et al. 2009).

- **Rat/rotenone**: In rats, exposed to rotenone (2.5 mg/kg/day), spontaneous locomotor activity was reduced by ca. 50 % after 1 week of rotenone treatment. This impaired motor
performance was correlated with a loss of striatal DA fibers by 54% and a loss of nigral DA neurons by 28.5% (Höglinger et al. 2003).

Mouse in vivo models

- **Mouse/MPTP:** In mice exposed to MPTP in combination with probenecid, both a chronic treatment scheme (MPTP 25 mg/kg, in 3.5 day intervals for 5 weeks) as well as a subacute treatment scheme (25 mg/kg, 1x per day for 5 days) resulted in a deletion of striatal DA that was directly correlated with impairments in motor symptoms (Petroske et al. 2001).

Mouse/MPTP: In a mouse model exposed to MPTP at 15 day intervals (36 mg/kg), lower rotarod performance was observed after the fourth injection. The decline in motor performance was correlated with the decline in TH-immunoreactivity in the striatum ($r^2 = 0.87$) (Rozas et al. 1998).

- **Mouse/D2 receptor knockout.** Mice deficient in D2 receptors displayed akinesia, bradykinesia and a reduction in spontaneous movement (Baik et al. 1995).

Monkey in vivo models

- **Monkey/MPTP:** Correlation between striatal DA, SNpc DA neuron number and PD symptoms. Macaca exposed to MPTP (i.v) (0.2 mg/kg, daily) display signs of PD at day 15, including motor abnormalities. The transition between the presymptomatic and symptomatic period occurred between day 12 and day 15 of MPTP exposure. At day 15, TH neurons in the SNpc were reduced by 50%, DAT binding autoradiography studies revealed a decline in binding also by 50% at day 15. Compared with control values of 150 pg/µg protein, the DA content of the caudate nucleus dropped to values < 10 pg/µg protein at day 15. In the putamen, DA levels dropped from 175 pg/µg protein to 20 pg/µg protein at day 15 (Bezard et al. 2001).

- **Monkey/MPTP:** The established model of basal ganglia wiring received ample experimental support in recent years. For instance, an increase in the inhibitory output by GPI/STN has been observed in MPTP treated monkeys, similar to the situation in idiopathic PD patients. These findings were corroborated by observations indicating an elevated mitochondrial activity and an elevated firing rate of the inhibitory output nuclei detected on the level of individual neurons (Mitchell et al. 1989; Filion et al. 1991). Lesions in the output ganglia of monkeys lead to a reduction in the output and to an improvement in motor control (Bergman et al. 1990; Aziz et al. 1991). In analogy to these lesion experiments, deep brain stimulation of these regions results in a profound improvement of motor performance in PD patients (Limousin et al. 1999; Ceballos-Baumann et al. 1994).

Human data

- **Human PD:** Association of PD phenotype with impaired striatal DA. In the brains of human PD patients, a significant decrease of striatal DA was observed (Lloyd et al. 1975). In the caudate nucleus, levels of DA dropped from control values of 4 µg/g tissue to levels of 0.2 µg/g. In the putamen, control values were in the range of 5 µg/g and 0.14 µg/g in the PD patient group.
The levels of DA in the striata of DA patients that received L-DOPA treatment was 9-15 times higher compared with non-treated PD cases.

- **Human PD**: Correlation between striatal DA loss and degeneration of DA neurons in the SNpc. Examinations of the brains of PD patients revealed morphological damage in the SNpc, accompanied by the degeneration of DA neurons (Earle et al. 1968).

- **Human PD**: Association of striatal DA levels and motor performance. In order to substitute degenerated DA neurons in the SNpc, human fetal tissue from the ventral mesencephalon was transplanted to the caudate and putamen in idiopathic cases PD as well as in patients that developed PD-related motor deficits as a consequence to MPTP intoxication. Transplanted cells led to a reinnervation of the striatum with DA projections (Widner et al. 1992; Kordower et al. 1995, 1998). In these case studies, patients demonstrated a sustained improvement in motor function (decline in rigidity score by more than 80%).

- **Human PD**: Correlation between nigrostriatal DA neuron content and motor symptoms. Imaging of DAT was performed by the use of 123I-FP-CIT SPECT (single photon emission computed tomography). Clinical PD severity was determined by using the Unified Parkinson Disease Rating Score (UPDRS). In PD patients, DAT binding in the striatum, caudate, and putamen correlated with disease severity and duration of disease (Benamer et al. 2000).

- **Human PD**: correlation between 18F-dopa uptake measured by PET and the onset of motor symptoms detected according the UPDRS. 18F-dopa influx rate constants (Ki/min) were reduced in the midbrain from 0.008 to 0.006, in the right putamen from 0.017 to 0.0036, and in the left putamen from 0.017 to 0.005 (Rakshi et al. 1999).

- **Human PD**: correlation between putamen influx rate (Ki/min). Ki (control): 0.0123; asymptomatic PD (no observable motor deficits): 0.0099; symptomatic PD (clinically evident motor deficits): 0.007. Mean UPDRS value was 15.1 ± 7.5. A correlation coefficient of -0.41 was detected between motor UPDRS and putamen influx (Ki) (Morrish et al. 1995).

- **Human PD**: Correlation of the degree of monoaminergic degeneration in early PD with motor symptoms assest by the UPDRS and the Hoehn and Yahr Stage scale. For PET imaging, 18F-9-fluoropropyl-dihydrotetrabenazine that targets VMAT2 was used. Uptake of the tracer was reduced by 20-36 % in the caudate, by 45-80 % in the putamen, and by 31 % in the substantia nigra. This correlated with a total UPDRS value of 12.1 ± 7.1 in the PD group, respectively with a HY value of 1.0 ± 0.1 in the PD group compared to controls (Lin et al. 2014).

- **Human PD**: Correlation between the decline in 18F-dopa rate constant (Ki) and the onset of motor deficits. The 18F-dopa rate constant Ki was reduced in the caudate nucleus (0.011 down to 0.0043) and inversely correlated with an increase in the UPDRS from 11.9 ± 5.2 to 50 ± 11.6 (Broussolle et al. 1999).

- **Human PD**: Correlation between striatal DAT binding measured by the use of 123I-CIT SPECT and motor deficits. A correlation coefficient between 123I-CIT binding and UPDRS motor scale of -0.56 was detected. A correlation coefficient of -0.64 between 123I-CIT binding and Hoehn and Yahr stage scale was detected. Motor symptoms in the clinically less affected body side show a closer correlation with striatal DAT binding (Pirker et al. 2003).

- **Human PD**: Correlation between the reduction in the putamen uptake of 18F-CFT and the severity of PD motor symptoms. 18F-CFT uptake was reduced to 18 % in the putamen, 28% in the anterior putamen, and to 51 % in the caudate nucleus (Rinne et al. 1999).

- **Human PD**: Reduction in 123I-CIT binding in the putamen by 65 % correlated with a mean UPDRS score of 27.1 (Tissingh et al. 1998).

8.3 Uncertainties or Inconsistencies

- Motor abnormalities observed in PD display large interindividual variations.
- The model of striatal DA loss and its influence on motor output ganglia does not allow to explain specific motor abnormalities observed in PD (e.g. resting tremor vs bradykinesia) (Obeso et al. 2000). Other neurotransmitters (Ach) may play additional roles.
- There are some reports indicating that in subacute rotenone or MPTP models (non-human primates), a significant, sometimes complete, recovery of motor deficits can be observed after termination of toxicant treatment. While the transient loss of striatal DA can be explained by an excessive release of DA under acute toxicant treatment, the reported losses of TH-positive neurons in the SNpc and their corresponding nerve terminals in the striatum are currently not explained (Petroske et al. 2001).
- In MPTP treated baboons, the ventral region of the pars compacta was observed to be more severely degenerated that the dorsal region. This pattern is similar to the degeneration pattern in idiopathic PD in humans. These observations indicate that two subpopulations of nigrostriatal DA neurons with different vulnerabilities might exist (Varastet et al. 1994).
- According to the classical model of basal ganglia organization, DA is assumed to have a dichotomous effect on neurons belonging either to the direct or indirect pathway. More recent evidence however rather indicates that D1 and D2 receptors are expressed on most striatal neurons in parallel (Aizman et al. 2000).

8.4 Quantitative Understanding of the Linkage

An example of quantitative analysis is reported in the table below. The analysis of the empirical data produced with the chemical toxicants supports a strong response-response relationship between the KE up and the KE down which also indicative of the temporal progression and relationship between the degeneration of striatal terminals of DA neurons, loss of DA neurons in the SNpc and the occurrence and severity of the motor deficits. This is also quantitatively supported by studies conducted in human PD patients.

Table 6: Quantitative evaluation of the KER

<table>
<thead>
<tr>
<th>Upstream key event (KE 4)</th>
<th>Downstream key event (AO)</th>
<th>References</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rat models</td>
<td></td>
<td></td>
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<tr>
<td>45 % loss of TH-positive SNpc neurons in 7 month old rats, ca. 40 % loss in 12 month old rats</td>
<td>Bradykinesia, postural instability, rigidity observed in 50 % of cases: 3 month old rats: after 12 days of rotenone 7 + 12 month old rats. After 6 days of rotenone Postural instability test: Distance required for the animal to regain postural stability: 3.5 cm (control) 5 cm (rotenone) Rearing test (rears/ 5 min):</td>
<td>Cannon et al. 2009</td>
<td>Lewis rats + rotenone (3 mg/kg/day, i.p. daily)</td>
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<tr>
<td>Striatal DA reduced from 90 ng/mg (control) down to 45 ng/mg</td>
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<tr>
<td>TH pos. neuron number</td>
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<tr>
<td>18000 (control)</td>
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<td></td>
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<tr>
<td>10000 (rotenone)</td>
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</table>
PPP5s, Parkinson’s disease and childhood leukaemia

Dopamine in the anterior and posterior striatum reduced by ca. 50 %.

Loss of rearing performance evoked by rotenone was reversed by the DA agonist Apomorphine in 3 month old rats.

10 (control)
3 (rotenone)

Catalepsy test: decline from 35 s to 5 s.
Grid test: decline from 30 s to 4 s
Distance travelled in 10 min: reduction from 37 m to 17 m.
Number of rearings: decline from 65 to 30.
Inactivity time increased from 270 s to 400 s.
Partial reversibility by L-DOPA treatment:
L-DOPA: number of rearings increased from 16 to 30.
L-DOPA: inactivity time reduced from 450 s to 360 s.
L-DOPA: increase in the distance travelled from 12 to 16 m.

Alam et al. 2004
Rats + rotenone (2.5 mg/kg) daily over the course of 48 days.

TH staining intensity reduced from 0.2 to 0.12
Rearing scores reduced from 80 % (vehicle controls) to 20 % (rotenone group).
Increase in the average time to initiate a step from 5 s to 11 s.

Fleming et al. 2004
Rats + rotenone 2.5 mg/kg for 21 days i.v. or s.c.

Loss of striatal DA fibers by 54 %
Loss of DA neurons by 28.5 %

Spontaneous locomotor activity after 1 week
100 % (control)
55 % (rotenone)

Höglinger et al. 2003
Rats + rotenone (2.5 mg/kg/day for 28 days

Mouse models

Subacute model:
Striatal DA dropped from 11 ng/mg (control) to 2.5 ng/mg (MPTP) after 3 days.
3H-DA striatal uptake reduced from 2.9 pmol/mg

Petroske et al. 2001
Mouse + MPTP
Subacute model:
<table>
<thead>
<tr>
<th>Test</th>
<th>Control</th>
<th>Early PD</th>
<th>Adv. PD</th>
<th>Right putamen</th>
<th>Control</th>
<th>Early PD</th>
</tr>
</thead>
<tbody>
<tr>
<td>18F-dopa influx rate constants (Ki)</td>
<td>Midbrain: 0.008</td>
<td>Early PD: 0.008</td>
<td>Adv. PD: 0.006</td>
<td>Right putamen:</td>
<td>Control: 0.017</td>
<td>Early PD: 0.006</td>
</tr>
<tr>
<td>Mean duration in the bradykinesia test</td>
<td>19 sec. (day 0)</td>
<td>3 sec. at day 15</td>
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<tr>
<td>Rotarod performance</td>
<td>1250 AUC</td>
<td>200 AUC</td>
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<td>Time on rod at a speed of 20 rpm</td>
<td>125 s in controls</td>
<td>25 s in MPTP animals</td>
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<tr>
<td>Reduction in TH staining intensity of at least 50 %</td>
<td>125 mg/kg MPTP 1x days for 5 days</td>
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<tr>
<td>TH density in the nigrostriatal system correlated with the decline of rotarod performance (r² = 0.87)</td>
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<tr>
<td>TH staining in the nigrostriatal system reduced by ca. 50 % 1 week after initiation of MPTP treatment.</td>
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<tr>
<td>Striatal DA content reduced from 13 ng/ml down to 0.5 ng/ml at 1 week after MPTP treatment.</td>
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<tr>
<td>TH staining in the nigrostriatal system reduced by ca. 50 % 1 week after initiation of MPTP treatment.</td>
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<tr>
<td>Reduction in TH staining intensity of at least 50 % required for detectable influence on motor performance.</td>
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</table>

**PPP, Parkinson’s disease and childhood leukaemia**

- **Chronic model:**
  - Striatal DA content reduced from 13 ng/ml down to 0.5 ng/ml at 1 week after MPTP treatment.
  - TH staining in the nigrostriatal system reduced by ca. 50 % 1 week after initiation of MPTP treatment.
  - Reduction in TH staining intensity of at least 50 % required for detectable influence on motor performance.
  - TH density in the nigrostriatal system correlated with the decline of rotarod performance (r² = 0.87)

- **Monkey models**
  - Approx. 50 % loss of TH positive neurons in the SNpc. DA content in the caudate nucleus reduced to < 10 %; DA content of the putamen ca. 10 % compared with control
  - Mean duration in the bradykinesia test increased from 3 sec. (day 0) to 19 sec. at day 15

- **Human**
  - 18F-dopa influx rate constants (Ki)
  - Midbrain:
    - Control: 0.008
    - Early PD: 0.008
    - Adv. PD: 0.006
    - Right putamen:
      - Control: 0.017
      - Early PD: 0.006

- **Rozas et al. 1998**
  - Mouse + MPTP
  - Rotarod performance reduced from 1800 AUC (control) to 1250 AUC (1 week after initiation of MPTP treatment)
  - Time on rod at a speed of 20 rpm: 125 s in controls, 25 s in MPTP animals

- **Bezard et al. 2001**
  - Macaca + MPTP i.v. 0.2 mg/kg daily for 15 days
  - Mean duration in the bradykinesia test increased from 3 sec. (day 0) to 19 sec. at day 15

- **Rakshi et al. 1999**
  - Human PD patients
  - Early PD:
    - UPDRS: 9 +/- 3
  - Adv. PD:
    - UPDRS: 41 +/- 15
<table>
<thead>
<tr>
<th>Study</th>
<th>Method</th>
<th>Results</th>
</tr>
</thead>
<tbody>
<tr>
<td>Morrish et al. 1995</td>
<td>Human PD</td>
<td>Uptake of 18F-DTBZ (VMAT2 tracer) reduced by: 20-36 % (caudate) 45-80 % (putamen) 31 % (SN)</td>
</tr>
<tr>
<td>Lin et al. 2014</td>
<td>Human PD</td>
<td>Reduction in 18F-CFT uptake in the posterior putamen (by 18 %); in the anterior putamen (by 28 %); in the caudate nucleus (by 51 %)</td>
</tr>
<tr>
<td>Broussolle et al. 1999</td>
<td>Human PD</td>
<td>Correlation between total motor score of the UPDRS and 18F-CFT uptake: Posterior putamen: r = -0.62 Anterior putamen: r = -0.64 Caudate nucleus: r = -0.62</td>
</tr>
<tr>
<td>Rinne et al. 1999</td>
<td>Human PD</td>
<td>Numerical values for 123I-CIT SPECT values in controls and PD cases with a Hoehn and Yahr rating of 2-2.5: Putamen (ipsilateral): Correlation coefficient between striatal 123I-CIT binding and: Str. (ipsilateral) and Bradykinesia:</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Study</th>
<th>Method</th>
<th>Results</th>
</tr>
</thead>
<tbody>
<tr>
<td>Human PD</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control: 6.13</td>
<td>PD: 1.84</td>
<td></td>
</tr>
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<td></td>
</tr>
<tr>
<td>Caudate (ipsilateral):</td>
<td>r = -0.61</td>
<td></td>
</tr>
<tr>
<td>Control: 6.93</td>
<td>PD: 3.66</td>
<td></td>
</tr>
<tr>
<td>Striatum (ipsilateral):</td>
<td>Str. (ipsilateral) and Rigidity:</td>
<td></td>
</tr>
<tr>
<td>Control: 6.28</td>
<td>PD: 2.33</td>
<td></td>
</tr>
<tr>
<td>Binding ratio striatum/cerebellum detected by 123I-CIT / SPECT</td>
<td>Correlation between 123I-CIT binding to DAT and PD motor symptoms rated according to the Hoehn and Yahr scale:</td>
<td></td>
</tr>
<tr>
<td>Control: 8.71 +/- 1.54</td>
<td>r = -0.75</td>
<td></td>
</tr>
<tr>
<td>PD: 4.49 +/- 1.86</td>
<td>Correlation according to the UPDRS:</td>
<td></td>
</tr>
<tr>
<td>Uptake of 123I-CIT in the putamen reduced to 54%; uptake into the caudate nucleus reduced to 65%</td>
<td>Correlation between CIT uptake in the putamen and Hoehn and Yahr stage:</td>
<td></td>
</tr>
<tr>
<td>r = -0.79</td>
<td>Decline in nigrostriatal DAT assessed by 123I-CIT SPECT in PD patients</td>
<td></td>
</tr>
<tr>
<td>Correlation coefficients for 123I-CIT uptake in the striatum and:</td>
<td>Benamer et al. 2000</td>
<td></td>
</tr>
<tr>
<td>UPDRS: r = -0.54</td>
<td>Human PD</td>
<td></td>
</tr>
<tr>
<td>Bradykinesia: r = -0.5</td>
<td>Human PD</td>
<td></td>
</tr>
<tr>
<td>Rigidity: r = -0.27</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tremor: r = -0.3</td>
<td></td>
<td></td>
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<tr>
<td>Correlation coefficients for 123I-CIT uptake in the caudate and:</td>
<td>Rinne et al. 1995</td>
<td></td>
</tr>
<tr>
<td>UPDRS: r = -0.5</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bradykinesia: r = -0.43</td>
<td></td>
<td></td>
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<tr>
<td>Rigidity: r = -0.27</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tremor: r = -0.26</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
8.5 Evidence Supporting Taxonomic Applicability

Parkinsonian disorders are generally recognized as progressive age-related human neurodegenerative diseases more prevalent in males. However, the anatomy and function of the nigrostriatal pathway is conserved across mammalian species (Barron et al. 2010) and no sex and species restrictions were evidenced using the chemical stressors rotenone and MPTP. It should be noted that animal behaviour models can only be considered as surrogates of human motor disorders as occurring in Parkinson’s disease.

References


Bezard E, Dovero S, Prunier C, Ravenscroft P, Chalon S, Guilloteau D, Crossman AR, Bioulac B, Brotchie JM, Gross CE (2001) Relationship between the appearance of symptoms and the level of uptake in the putamen and:

UPDRS: r = -0.57
Bradykinesia: r = -0.53
Rigidity: r = -0.29
Tremor: r = -0.37


**Overall assessment of the AOP**

1. **Concordance of dose-response relationship**

Data from experiments with the stressor compounds rotenone and MPTP (known inhibitors of the mitochondrial Complex I (CI)) reveal a good concordance of the dose-response relationships between the MIE and AO within KEs. Although the different KEs have been measured using different methodologies, comparison of data from multiple in-vitro/in-vivo studies shows a general agreement in dose-response relationship (see table 1 and 2). There is a good consistency when comparing data on KE4 and the AO after exposure to rotenone and MPTP. However, in vivo rodent studies proved that only exposure to low concentrations of rotenone (rat brain concentration between 20-30 nM of rotenone; Betrabet et al., 2000) or MPTP (mice striatum concentration of approximately 12-47 µM MPP+; Fornai et al., 2005; Thomas et al. 2012) after chronic exposure (approximately 5 weeks) reproduced the anatomical, neurochemical and neuropathological features similar to the ones observed in Parkinson's disease (PD). Because of the variability of experimental protocols used, a clear no-effect threshold could not be established; nevertheless, these brain concentrations of rotenone (20-30 nM) and MPP+ (approximately 12-47µM) could serve as probabilistic thresholds for chronic exposure that could reproduce features of PD as both concentrations trigger approximately a 50% inhibition of Complex I (see table 3). Generally, a strong response-response relationship is observed within studies. Some exceptions for this rule are observed between KE3/KE5 and KE4, likely because of the all biological complexity associated with these KEs. In this AOP, neuroinflammation was considered to
have a direct effect on degeneration of DA neurons. However, it was not clear at which conditions it would become a modulatory factor and for practical reasons was not included in table 1, 2 and 3 but considered in the weight of evidence analysis.

### 2. Temporal concordance among the MIE, KEs and AO

There is a strong agreement that loss of DA neurons of the SNpc that project into the putamen is preceded by reduction in DA and degeneration of DA neuronal terminals in the striatum (Bernheimer et al. 1973). The clinical symptoms of a motor deficit are observed when 80% of striatal DA is depleted (Koller et al. 1992) and the sequence of pathological events leading to the adverse outcome has been well-documented (Fujita et al. 2014; O'Malley 2010, Dexter et al. 2013). Temporal concordance (see table 1 and 2) among the KEs can be observed in the experimental models of PD using the chemical stressors rotenone and MPTP (Betarbet 2000 and 2006; Sherer et al. 2003, Fornai et al. 2005). The acute administration of the chemical stressors can trigger a dose-related change from the MIE to impaired proteostasis; however, to trigger KE4 (i.e. degeneration of DA neurons in SNpc with presence of intracytoplasmatic Lewy-like bodies) and motor deficits (AO), proteostasis needs to be disturbed for a minimum period of time (Fornai et al. 2005).

#### Table 7: Response-Response and Temporality concordance table for the tool compound rotenone

|-------------------------------------|---------|-----------------------|---------|-------------------------------|---------|----------------------------|---------|--------------------------------|--------|---------------------------------|

References: Choi et al. 2008 [1]; Betarbet et al. 2006 [2]; Chou et al. 2010 [3]; Barrientos and Moraes 1999 [4]; Okun et al. 1999 [5]; Betarbet et al. 2000 [6]; no data available 1999 [7]; Betarbet et al. 2000 [6]; all animals affected in KE4 showed impaired motor symptoms; a 50% of treated animals showed loss of DA neurons in SNpc.

#### Table 8: Response-Response and Temporality concordance table for the tool compound MPTP/MPP+

<table>
<thead>
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</tr>
</thead>
<tbody>
<tr>
<td>1 mg/kg infusion [1]</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>4 weeks[1]</td>
<td>+aaa</td>
<td>4 weeks [1]</td>
<td>No effect</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
3. Strength, consistency, and specificity of association of AO and MIE

Strength and consistency of the association of the AO with the MIE is strong. There is a large body of evidence from in-vitro and in-vivo studies with chemical stressors, showing association between the MIE that triggers an inhibition of CI and the AO (Sherer et al. 2003; Betarbet et al. 2000 and 2006, Fornai et al. 2005; Thomas et al. 2012). Human data also suggest a link between inhibition of CI and AO (Greenamyre et al. 2001; Schapira et al. 1989; Shults, 2004). Using the two different chemical stressors, rotenone and MPTP, data are consistent and the pattern of activation of the MIE leading of the AO is similar. For rotenone and MPTP, specificity is high; however, there are many inhibitors of the mitochondrial CI without evidence of triggering the AO. When considering these chemicals specificity is low; therefore, kinetic and metabolic considerations should be taken into account to fully demonstrate specificity for these compounds.

4. Weight of Evidence (WoE)

4.1 Biological plausibility, coherence, and consistency of the experimental evidence

The biological plausibility of this AOP is overall considered strong. When using multiple stressors in different studies and assays, the coherence and consistency of the experimental data is well established. Furthermore, in-vivo and in-vitro studies are also in line with the human evidence from PD patients. In addition, although the mechanistic understanding of parkinsonian disorders (and PD in particular) are not fully clear, the KEs and KERs described in this AOP are considered critical for the development of the disease (Fujita et al. 2015, Shulman et al. 2011, Dexter et al. 2013, Dauer et al. 2003).
### Table 9: Biological Plausibility of KERs; WoE analysis

<table>
<thead>
<tr>
<th>Support for Biological Plausibility of KERs</th>
<th>Defining Question</th>
<th>High (Strong)</th>
<th>Moderate</th>
<th>Low (Weak)</th>
</tr>
</thead>
<tbody>
<tr>
<td>MIE=&gt;KE1</td>
<td>Binding of inhibitor to NADH-ubiquinone oxido-reductase leads of complex I</td>
<td><strong>STRONG</strong></td>
<td>Rationale: As describe in this KER there is an extensive understanding of the functional relationship between binding of an inhibitor to NADH-ubiquinone oxido-reductase (CI) and its inhibition. Different complex I ligands, both naturally occurring, like rotenone (from Derris scandens), piericidin A (from Streptomyces mobaraensis), acetogenins (from various Annonaceae species) and their derivatives, and synthetically manufactured like pyridaben and various piperazin derivatives inhibit the catalytic activity of complex I (Degli Esposti, 1994; Ichimaru et al. 2008; Barrientos and Moraes, 1999; Betarbet et al., 2000).</td>
<td></td>
</tr>
<tr>
<td>KE1=&gt;KE2</td>
<td>Inhibition of complex I leads to mitochondrial dysfunction</td>
<td><strong>STRONG</strong></td>
<td>Rationale: There is extensive understanding of the mechanisms explaining how the inhibition of complex I lead to mitochondrial dysfunction (i.e., failure to produce ATP, increase in production of ROS etc). It is well documented that CI inhibition is one of the main sites at which electron leakage to oxygen occurs resulting in oxidative stress (Efremov and Sazanow, 2011; lauren et al. 2010; Greenamyre et al. 2001). These pathological mechanisms are well studied as they are used as readouts for evaluation of mitochondrial dysfunction (Graier et al., 2007; Braun, 2012; Martin, 2011; Correia et al., 2012; Cozzolino et al., 2013)</td>
<td></td>
</tr>
<tr>
<td>KE2=&gt;KE3</td>
<td>Mitochondrial dysfunction results in impaired proteostasis</td>
<td><strong>MODERATE</strong></td>
<td>Rationale: The weight of evidence supporting the biological plausibility behind the relationship between mitochondrial dysfunction and impaired proteostasis, including the impaired function of UPS and ALP that results in decreased protein degradation and increase protein aggregation is well documented but not fully understood. It is well established that the two main mechanisms that normally remove abnormal proteins (UPS and ALP) rely on physiological mitochondrial function. The role of oxidative stress, due to mitochondrial dysfunction, burdens the proteostasis with oxidized proteins and impairs the chaperone and the degradation systems. This leads to a vicious circle of oxidative stress inducing further mitochondrial impairment (Powers et al., 2009; Zaltieri et al., 2015; McNaught and Jenner, 2001). Therefore, the interaction of mitochondrial dysfunction and UPS /ALP deregulation plays a pivotal role in the pathogenesis of PD (Dagda et al., 2013; Pan et al., 2008; Fornai et al., 2005; Sherer et al., 2002).</td>
<td></td>
</tr>
<tr>
<td>KE2=&gt;KE4</td>
<td>Mitochondrial dysfunction leads to the degeneration of dopaminergic neurons of the nigrostriatal pathway</td>
<td><strong>STRONG</strong></td>
<td>Rationale: Mitochondrial dysfunction is essential for ATP production, ROS management, calcium homeostasis and control of apoptosis. Mitochondrial homeostasis by mitophagy is also an essential process for cellular maintenance (Fujita et al. 2014). Because of their anatomical and physiological characteristics, SNpc DA neurons are considered more vulnerable than other neuronal populations (Sulzer et al. 2013; Surmeier et al.2010). Mechanistic evidence of mutated proteins relate the mitochondrial dysfunction in familial PD with reduced calcium capacity, increased ROS production, increase in mitochondrial membrane permeabilization and increase in cell vulnerability (Koopman et al. 2012; Gandhi et al. 2009). Human studies indicate mitochondrial dysfunction in human idiopathic PD cases in the substantia nigra (Keeney et al., 2006; Parker et al., 1989, 2008; Swerdlow et al., 1996). In addition, systemic application of mitochondrial neurotoxins such as rotenone or MPTP leads to a preferential loss of nigrostriatal DA neurons (Langston et al., 1983).</td>
<td></td>
</tr>
</tbody>
</table>
 KE3 => KE4
Impaired proteostasis leads to degeneration of DA neurons of the nigrostriatal pathway

MODERATE

Rationale: It is well known that impaired proteostasis refers to misfolded and aggregated proteins including alpha-synuclein, deregulated axonal transport of mitochondria and impaired trafficking of cellular organelles. Evidences are linked to PD and experimental PD models as well as from genetic studies (McNaught et al. 2001, 2003; Tieu et al. 2014; Arnold 2011; Rappold et al. 2014). Strong evidence for degeneration of the nigrostriatal pathway comes from the experimental manipulations that directly induce the same disturbances of proteostasis as observed in PD patients (e.g. viral mutated alpha-synuclein expression) or in chronic rotenone/MPTP models trigger degeneration of the nigrostriatal pathway (Kirk et al. 2003; Betarbet et al. 2000 and 2006; Fornai et al. 2005). However, a clear mechanistic proof for the understanding of the exact event triggering cell death is lacking. There is only moderate evidence showing that interventions that correct disturbances of proteostasis after exposure to rotenone would prevent neuronal degeneration and that the disturbances of proteostasis correlate quantitatively under many conditions with the extent of nigrostriatal neuronal degeneration.

KE4 => KE5
Neuroinflammation

MODERATE

Rationale: The fact that reactive glial cells (microglia and astrocytes) may kill neurons is well accepted. The mechanisms underlying this effect may include the release of cytotoxic signals (e.g. cytokines) or production of ROS and RNS (Chao et al., 1995; Brown and Bal-Price, 2003; Kraft and Harry, 2011; Taetzsch and Block, 2013). However, the responsible mediators differ from model to model. The evidence that neuroinflammation triggered by neuronal damage can cause neuronal death (vicious circle), is mostly indirect or by analogy (Hirsch and Hunot, 2009; Tansey and Goldberg, 2009; Griffin et al., 1998; McGee and Mc Geer, 1998; Blasko et al., 2004; Cacquevel et al., 2004; Rubio-Perez and Morillas-Ruiz, 2012; Thundyil and Lim, 2014; Barbeito et al., 2010).

KE4 => AO
Degeneration of DA neurons of the nigrostriatal pathway leads to parkinsonian motor symptoms

STRONG

Rationale: The mechanistic understanding of the regulatory role of striatal DA in the extrapyramidal motor control system is well established. The loss of DA in the striatum is characteristic of all aetiologies of PD and is not observed in other neurodegenerative diseases (Bernheimer et al. 1973; Reynolds et al. 1986). Characteristic motor symptoms such as bradykinesia, tremor, or rigidity are manifested when more than 80 % of striatal DA is depleted as a consequence of SNpc DA neuronal degeneration (Koller et al. 1992).

4.2 Essentiality

Essentiality of KEs for this AOP is strong. There is ample evidence from knock out animal models, engineered cells or replacement therapies that blocking, preventing or attenuating an upstream KE is mitigating the AO. In addition, there is experimental support for the KERs as multiple studies performed with modulating factors that attenuate (particularly with antioxidants) or augment (e.g. overexpression of viral-mutated alpha-synuclein) a KE show that such interference leads to an increase of KE down or the AO.

Table 10: Essentiality of KEs; WoE analysis

<table>
<thead>
<tr>
<th>Support for Essentiality of KEs</th>
<th>Defining Question: Are downstream KEs and/or the AO prevented if an upstream KE is</th>
</tr>
</thead>
<tbody>
<tr>
<td>High (Strong)</td>
<td>Direct evidence from specifically designed</td>
</tr>
<tr>
<td>Moderate</td>
<td>Indirect evidence that is sufficient</td>
</tr>
<tr>
<td>Low (Weak)</td>
<td>No or contradictory</td>
</tr>
</tbody>
</table>

www.efsa.europa.eu/efsajournal
| KE | Inhibition of complex I | STRONG | Rationale: Inactivation of the Ndufs 4 gene (knockout mice) that produces CI deficiency causes encephalomyopathy, including ataxia and loss of motor skills (Kruse et al., 2008). NDI1-transduced SK-N-MC cells expressing the rotenone-insensitive single subunit NADH dehydrogenase of yeast (NDI1) that acts as a replacement for the entire CI in mammalian cells were completely resistant to 100 nM rotenone-mediated cell death (at 48 hrs of exposure) indicating that rotenone – induced toxicity requires rotenone binding of CI (Sherer et al., 2003). In all rotenone models, mitochondria CI is inhibited at the dose that cause neurodegeneration (Betarbet et al 2000 and 2006). |
| KE | Mitochondrial dysfunction | STRONG | Rationale: Many studies showing that antioxidants protect the cells against rotenone or MPTP induced oxidative stress are published (Chen et al. 2015; Lu et al., 2015; Saravanan et al., 2006; Chiu et al., 2015, Sherer et al.2003, Nataraj et al.2015, Wu et al. 1994; Tseng et al. 2014; Li et al. 2010; Kim-Han et al. 2011). This provides (indirect) evidence for essentiality of KE2, if production of ROS is assumed as direct consequence/sign of mitochondrial dysfunction. Additional evidence comes from experiments with overexpression or activation of antioxidative enzymes (e.g.SOD or ALDH2) , which also prevent rotenone and MPTP induced neurotoxicity (Mudo et al. 2012; Ciu CC et al. 2015). Furthermore, promotion of mitochondrial fusion or blocking of mitochondrial fission prevents or attenuates rotenone and MPTP induced neurotoxicity (Tieu K. et al. 2014). |
| KE | Impaired proteostasis | MODERATE | Rationale: Indirect evidence for the role of disturbed alpha-synuclein proteostasis: Lacking of alpha-synuclein expression in mice prevented induction of behavioural symptoms, neuronal degeneration in the nigrostriatal pathway and loss of DA neurons after chronic treatment with MPTP (Fornai et al. 2004; Dauer et al. 2002) . Injection of adeno/lenti-associated virus that expresses wild-type or mutant α-syn into rat, mice or non-human primate SN produced loss of dopaminergic neurons, but the effect is not easily reproduced in transgenic mice overexpressing alpha-synuclein (Kirk, 2002; Klein, 2002; Lo Bianco, 2002; Lauwers, 2003; Kirk, 2003). Rationale for the role of autophagy: Early dendritic and axonal dystrophy, reduction of striatal dopamine content, and the formation of somatic and dendritic ubiquitinated inclusions in DA neurons were prevented by ablation of Atg7 (an essential autophagy gene (Friedman et al. 2012)). Rationale for the role of UPS/ALP: Protection from DA neuronal death was also observed in multiple experiments through the pharmacological modulation of the UPS, ALP system; however, there are also contradicting data in the literature. (Inden et al. 2007; Fornai et al. 2003; Dehay et al. 2010; Zhu et al. 2007, Fornai et al. 2005). However, although many lines of evidence exist to support essentiality of impaired proteostasis, a single molecular chain of events cannot be established. |
| KE | Degeneration of DA neurons of nigrostriatal pathway | STRONG | Rationale: Clinical and experimental evidences show that the pharmacological replacement of the DA neurofunction by allografting fetal ventral mesencephalic tissues is successfully replacing degenerated DA neurons resulting in the total reversibility of motor deficit in animal model and partial effect is observed in human patient for PD (Widner et al., 1992; Henderson et al., 1991; Lopez-Lozano et al., 1991; Freed et al., 1990; Peschanski et al., 1994; Spencer et al., 1992). Also, administration of L-DOPA or DA agonists results in an improvement of motor deficits (Calne et al 1970; Fornai et |
The success of these therapies in man as well as in experimental animal models clearly confirms the causal role of dopamine depletion for PD motor symptoms (Connolly et al., 2014; Lang et al., 1998; Silva et al., 1997; Cotzias et al., 1969; Uitti et al., 1996; Ferrari-Tonielli et al., 2008; Kelly et al., 1987; Walter et al., 2004; Narabayashi et al., 1984; Matsumoto et al., 1976; De Bie et al., 1999; Uitti et al., 1997; Scott et al., 1998; Moldovan et al., 2015; Deuschl et al., 2006; Pasano et al., 2010; Castrito et al., 2011; Liu et al., 2014; Widner et al., 1992; Henderson et al., 1991; Lopez-Lozano et al., 1991; Freed et al., 1990; Peschanski et al., 1994; Spencer et al., 1992).

Furthermore, experimental evidence from animal models of PD and from in-vitro systems indicate that prevention of apoptosis through ablation of BCL-2 family genes prevents or attenuates neurodegeneration of DA neurons (Offen et al., 1998; Dietz et al., 2002).

Rationale: Following treatment with Rotenone or MPP+, protection of DA neurons and terminals was observed in vivo and in vitro by inhibiting different feature of neuroinflammation (microglia/astrocyte); however, inhibition was different in different models and considered as an indirect evidence of essentiality (Zhou et al., 2007; Gao et al., 2002 and 2003 and 2015; Emmrich et al., 2013; Salama et al., 2012; Chang et al., 2013; Wang et al., 2014; Liu et al., 2012, 2015; Borrajo et al., 2013; Brzozowski et al., 2015; Wang et al., 2006; Chung et al., 2011; Srinivas et al., 2014; Feng et al., 2002; Sathe et al., 2012; Khan et al., 2014; Ros-Bernal et al., 2011; Ferger et al., 2004; Chao et al., 2009; Rojo et al., 2010; Qian et al., 2011; Dehner et al., 2000; Bodea et al., 2014; Purisai et al., 2007; Manzano et al., 2012; Wu et al., 2005; Yadav et al., 2012). It should be noted that this KE, depending on the situation, can be bypassed.

### Empirical support

Empirical support is strong. Many studies show evidence for the KERs by showing temporal concordance and dose concordance when using different stressors.

#### Table 11: Empirical support for the KERs; WoE analysis

<table>
<thead>
<tr>
<th>Empirical support for KERs</th>
<th>Defining Question</th>
<th>High (Strong)</th>
<th>Moderate</th>
<th>Low(Weak)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Does the empirical evidence support that a change in the KEup leads to an appropriate change in the KE down? Does KEup occur at lower doses and earlier time points than KE down and is the incidence of KEup higher than that for KE down? Are inconsistencies in empirical support cross taxa, species and stressors that don't align with expected pattern of hypothesized</td>
<td>Multiple studies showing dependent change in both exposure to a wide range of specific stressors (extensive evidence for temporal, dose-response and incidence concordance) and no or few critical data gaps or conflicting data.</td>
<td>Demonstrated dependent change in both events following exposure to a small number of specific stressors and some evidence inconsistent with expected pattern that can be explained by factors such as experimental design, technical considerations, differences among laboratories, etc.</td>
<td>Limited or no studies reporting dependent change in both events following exposure to a specific stressor (ie endpoints never measured in the same study or not at all); and/or significant inconsistencies in empirical support across taxa and species that don't align with expected pattern for hypothesized AOP</td>
</tr>
</tbody>
</table>
PPP, Parkinson’s disease and childhood leukaemia

<table>
<thead>
<tr>
<th>AOP?</th>
<th>Rationale:</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>MIE=&gt;KE1</strong></td>
<td>Binding of inhibitor to NADH-ubiquinone oxidoreductase leads to partial or total inhibition of complex I</td>
</tr>
<tr>
<td><strong>KE1=&gt;KE2</strong></td>
<td>Inhibition of complex I leads to mitochondrial dysfunction</td>
</tr>
<tr>
<td><strong>KE2=&gt;KE3</strong></td>
<td>Mitochondrial dysfunction results in impaired proteostasis</td>
</tr>
<tr>
<td><strong>KE2=&gt;KE4</strong></td>
<td>Mitochondrial dysfunction directly leads to degeneration of DA neurons of nigrostriatal pathway</td>
</tr>
<tr>
<td><strong>KE3=&gt;KE4</strong></td>
<td>Impaired proteostasis leads to degeneration of DA neurons of the nigrostriatal pathway</td>
</tr>
<tr>
<td><strong>KE4&lt;=&gt;KE5</strong></td>
<td>Neuroinflammation directly leads to degeneration of DA neurons of the nigrostriatal pathway</td>
</tr>
<tr>
<td><strong>KE4=&gt;AO</strong></td>
<td>Degeneration of DA neurons of nigrostriatal pathway leads to</td>
</tr>
</tbody>
</table>

**Rationale:**
- The inhibition of complex I is well documented in a variety of studies using isolated mitochondria or cells as well as in vivo experiments and in human post mortem PD brains. In many experiments using different inhibitors i.e. rotenone and MPTP, the observed relationship between the two events was temporal, response and dose concordant (Betarbet et al., 2000 and 2006, Okun et al., 1999, Koopman et al., 2007, Choi et al., 2008, Grivennikova et al., 1997, Barrientos and Moraes 1999).
- There is a large amount of studies showing that the inhibition of CI inhibition results in mitochondrial dysfunctions in a response and dose dependent manner (Barrientos and Moraes, 1999).
- Based on the existing in vitro and in vivo data it is suggested that mitochondrial dysfunction impairs protein homeostasis (impairment of the UPS and ALP system) through oxidative and nitrosative stress resulting in accumulation of misfolded proteins (including α-synuclein), disruption of microtubule assembly and damaged intracellular transport of proteins and cell organelles. A number of studies performed with chemical stressors showed evidence of temporal, response and dose concordance (Chou et al. 2010; Betarbet et al. 2000 and 2006; Fornai et al. 2005).
- Multiple in vitro studies indicate dose and response-concordance. Many studies were conducted in vitro, the temporal concordance is difficult to establish; however, can be expected based on the well known temporal sequence of the two KEs. (Park et al., 2014; Choi et al., 2014; Marella et al., 2008; Du et al. 2010; Hajieva et al., 2009; Sherer et al., 2003; Sherer et al., 2007; Wen et al. 2011; Swedlow et al., 1996; Jana et al., 2011; Jha et al., 2000; Chinta et al., 2006).
- The empirical support linking impaired proteostasis with degeneration of DA neurons of the nigrostriatal pathway is strong and comes from in-vivo and in-vitro studies performed with different stressor (i.e. Rotenone, MPTP or proteasome inhibitors) and post-mortem human evidences in PD patients supporting a causative link between the two key events. Temporal, effect and dose concordance was established in a number of experiments (Fornai et al. 2005; Fornai et al. 2003; Betabret et al. 2000 and 2006).
- Multiple in vivo and in vitro experiments support the link between neuroinflammation and degeneration of DA neurons in the nigrostriatal pathway as well as vice versa. The observation of concomitant presence of glial and astrocytic cells and degenerated/degenerating DA neurons is also reported in many studies with a good temporal and response concordance.
- The experimental support linking the degeneration of DA neurons of nigrostriatal pathways with the manifestation of motor symptoms of PD comes from human in vivo observations as well as from monkey, mice and rat in vivo models exposed to an experimental toxin i.e. rotenone and MPTP. Observations in human allow defining correlation between the levels of striatal DA with the onset of motor dysfunction (Lloyd et al. 1975; Hornykiewicz et
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parkinsonian motor symptoms

PPPs, Parkinson's disease and childhood leukaemia

5. Uncertainties and Inconsistencies

- There is no strict linear relationship between inhibitor binding and reduced mitochondrial function. Low doses of rotenone that inhibit CI activity partially do not alter mitochondrial oxygen consumption. Therefore, bioenergetics defect cannot account alone for rotenone-induced neurodegeneration. Instead, under such conditions, rotenone neurotoxicity may result from oxidative stress (Betarbet et al., 2000). Few studies used human brain cells/human brain mitochondria. Therefore, full quantitative data for humans are not available.

- It is molecularly unclear how rotenone binding alters CI function, switching it to ROS production. It is still unclear whether the site of superoxide production in CI inhibited mitochondria is complex I itself or not (Singer and Ramsay, 1994).

- Some studies suggest that rotenone and MPTP may have effects other than CI inhibition, e.g. MPTP and rotenone can induce microtubule disruption (Feng, 2006; Ren et al., 2005; Cappelletti et al., 1999; Cappelletti et al., 2001; Brinkley et al., 1974; Aguilar et al. 2015).

- There are additional feedback possible between KEs, e.g. ROS production from KE2 may damage CI, this leads to enhancement of KE1.

- Some KEs e.g. KE 2, 3, 5 pool molecular processes that may need to be evaluated individually at a later stage.

- The exact molecular link from mitochondrial dysfunction to disturbed proteostasis is still unclear (Malkus et al. 2009; Zaltieri et al. 2015).

- The role of ATP depletion vs. other features of mitochondrial dysfunction is not clear.

- The role of α-synuclein in neuronal degeneration is still unclear as well as the mechanisms leading to its aggregation.

- It is not clear under which conditions KE3 and KE5 become modulatory factors, and when they are essential. MPTP can induce damage to nigrostriatal neurons without formation of Lewy bodies (Dauer 2003; Forno 1986, 1993). Similarly, discontinuous administration of rotenone, even at high doses, damages the basal ganglia but produce no inclusions (Heikkila et al. 1985; Ferrante et al. 1997, Lapontine 2004). To reproduce the formation of neuronal inclusions, continuous infusion of MPTP or rotenone is necessary. Acute intoxication with rotenone seems to spare dopaminergic neurons (Dauer et al. 2003, Ferrante 1997). In addition, in rats chronically infused with rotenone showed a reduction in striatal DARPP-32-positive, cholinergic and NADPH diaphorase-positive neurons (Hoglinger 2003) or in other brain regions. These results would suggest that Rotenone can induce a more widespread neurotoxicity (Aguilar 2015) or the model is not reproducible in all laboratories.

- Inconsistent effects of MPP+ on autophagy (up or down regulation) are reported (Drolet et al., 2004; Dauer et al., 2002). There is conflicting literature on whether increased autophagy would be protective or enhances damage. Similarly, a conflicting literature exists on extent of inhibition or activation of different protein degradation system in PD and a clear threshold of onset is unknown (Malkus et al. 2009; Fornai et al. 2005).

- The selective vulnerability of the SNpc dopaminergic pathway does not have a molecular explanation.

- Priority of the pattern leading to cell death could depend on concentration, time of exposure and species sensitivity; these factors have to be taken into consideration for the interpretation of the study's result and extrapolation of potential low-dose chronic effect as this AOP refers to long-time exposure.

- The model of striatal DA loss and its influence on motor output ganglia does not allow to explain specific motor abnormalities observed in PD (e.g. resting tremor vs bradykinesia) (Obeso et al. 2000). Other neurotransmitters (Ach) may play additional roles. Transfer to animal models of PD symptoms is also representing an uncertainty.

- There are some reports indicating that in subacute rotenone or MPTP models (non-human primates), a significant, sometimes complete, recovery of motor deficits can be observed after termination of toxicant treatment. The role of neuronal plasticity in intoxication recovery and resilience is unclear.

- This AOP is a linear sequence of KEs. However, mitochondrial dysfunction (and oxidative stress) and impaired proteostasis are influencing each other and this is considered an uncertainties (Malkus et al. 2009).
6. Quantitative Considerations

The quantitative understanding of this AOP includes a clear response-response relationship and the identification of a threshold effect. The WoE analysis clearly supports the qualitative AOP as a means to identify and characterize the potential of a chemical to induce DA neuronal loss and the AO. Importantly, both the AO and the KE4 are considered relevant regulatory endpoints for this AOP. The empirical evidence supports existence of a response-response relationship. This response-response is likely triggered by a brain concentrations of approximately 20-30 nM and 17-47 μM of rotenone and MPP+, respectively, and both concentrations trigger approx. a 50% inhibition of mitochondrial complex I and this could be considered as a “threshold”. However, a more detailed dose-response analysis for each KE is lacking as it is not clear which temporal relationship exists for lower CI inhibitory effects. It is clear from the analysis of the AOP that for the identification of these AOs, the design of the in-vivo studies should be tailored as to a MIE which leads to a long-lasting perturbation of the KEs. This provides the most specific and definite context to trigger neuronal death. To observe KEs relevant for this AOP, new endpoints need to be introduced. Although a dose, response and temporal relationship is evident for most KEs, the quantitative relationship between impaired proteostasis and degeneration of DA neurons has yet to be elucidated. Moving from a qualitative AOP to quantitative AOP would need a clear understanding of effect thresholds and this is still representing a major hurdle for several KEs of this AOP.

Table 12: Concordance table for the tool compounds rotenone and MPTP/MPP*

<table>
<thead>
<tr>
<th>Concentration</th>
<th>KE1</th>
<th>KE2</th>
<th>KE3</th>
<th>KE4</th>
<th>AO</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rotenone 20-30 nM rat brain</td>
<td>Approx. 53%</td>
<td>Approx. 20-53%</td>
<td>Approx. 20-60% (decrease</td>
<td>Neuronal loss (50%</td>
<td>Motor impairment</td>
</tr>
<tr>
<td>concentration [1-2]</td>
<td>[4-5]</td>
<td>(decrease in</td>
<td>in UPS (26S activity)</td>
<td>of animal affected) [2]</td>
<td>(100% of animals</td>
</tr>
<tr>
<td></td>
<td></td>
<td>respiration</td>
<td></td>
<td></td>
<td>with neuronal loss</td>
</tr>
<tr>
<td></td>
<td></td>
<td>rate) [1-2]</td>
<td></td>
<td></td>
<td>[2]</td>
</tr>
<tr>
<td>MPP+ 12-47 μM rat brain</td>
<td>Approx. 75%</td>
<td>Approx. 38%</td>
<td>Approx. 60% (decrease</td>
<td>Approx. 50% of</td>
<td>Motor impairment</td>
</tr>
<tr>
<td></td>
<td></td>
<td>phosphorylating</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>respiration) [5]</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>


7. Applicability of the AOP

This proposed AOP is neither sex-dependent nor associated with certain life stage; however, aged animals may be more sensitive. The relevance of this AOP during the developmental period has not been investigated.

In vivo testing has no species restriction. The mouse was the species most commonly used in the experimental models conducted with the chemical stressors; though experimental studies using alternative species have been also performed. (Johnson et al. 2015). However, animal models (rodents in particular) would have limitations as they are poorly representative of the long human life-time as well as of the human long-time exposure to the potential toxicants. Human cell-based models would likely have better predictivity for humans than animal cell models. In this case, toxicokinetics information from in-vivo studies would be essential to test the respective concentrations in-vitro on human cells.

8. Schematic summary of the AOP

Chronic, low level of exposure to environmental chemicals that inhibit complex I could result in mitochondrial dysfunction and oxidative stress, triggering proteasomal dysfunction strongly implicated
in parkinsonian disorders, including aggregation/modifications in α-synuclein protein and organelles trafficking. These cellular key events cause DA terminals degeneration in striatum and progressive cell death of DA neurons in SNpc. Important to notice that at each step, the effects become regionally restricted such that systemic complex I inhibition eventually results in highly selective degeneration of the nigrostriatal pathway.

Fig 18: Schematic summary of the AOP
9. Potential application of the AOP

This AOP has been developed in order to evaluate the biological plausibility that the adverse outcome i.e. parkinsonian motor deficits, is linked to a MIE that can be triggered by chemical substances i.e. pesticides and chemicals in general. The relevance of the AOP has been documented by tools compounds known to trigger the described AOP. By means of using a human health outcome that has been shown in epidemiological studies to be associated with pesticide exposure, the authors intend to draw attention on this AO in the process of hazard identification. This AOP can be used to support the biological plausibility of this association during the process of evaluation and integration of the epidemiological studies into the risk assessment. It is biologically plausible that a substance triggering the pathway, can be associated with the AO and ultimately with the human health outcome, pending the MoA analysis. In addition, this AOP can be used to support identification of data gaps that should be explored when a chemical substance is affecting the pathway. Moreover, the AOP provides a scaffold for recommendations on the most adequate study design to investigate the apical endpoints.

It is important to note that, although the AO is defined in this AOP as parkinsonian motor deficits, degeneration of DA neurons is already per se an adverse outcome even in situations where it is not leading to parkinsonian motor deficits, and this should be taken into consideration for the regulatory applications of this AOP.

The MIE and KEs identified in this AOP could serve as a basis for assays development that could contribute to an AOP informed-IATA construction which can be applied for different purposes such as: screening and prioritization of chemicals for further testing, hazard characterization or even risk assessment when combined with exposure and ADME information.
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AOP 2: Redox-cycling of a chemical initiated by electrons released by the mitochondrial respiratory chain leading to parkinsonian motor deficits

Abstract

This Adverse Outcome Pathway (AOP) describes the linkage between excessive ROS production at the level of the mitochondrial respiratory chain and parkinsonian motor deficits, including Parkinson’s disease (PD). Interaction of a compound with complex I and/or III of the mitochondrial respiratory chain has been defined as the molecular initiating event (MIE) that triggers mitochondrial dysfunction, impaired proteostasis, which then cause degeneration of dopaminergic (DA) neurons of the nigrostriatal pathway. These causatively linked cellular key events result in motor deficit symptoms, typical of parkinsonian disorders including PD, described in this AOP as an Adverse Outcome (AO). This AOP also includes neuroinflammation as a KE and is intending the KER with degeneration of dopaminergic neurons as a causative link but the priority of the temporal sequence is not defined as neurodegeneration can be the cause as well the consequence of the KE neuroinflammation.

Since the role DA neurons of the Substantia Nigra pars compacta (SNpc) projecting into the striatum is essential for motor control, the key events refer to these two brain structures, i.e. SNpc and striatum. The weight-of-evidence supporting the relationship between the described key events is mainly based on effects observed after an exposure to the well-known pesticide paraquat which will be used as a tool chemical to support this AOP.

Schematic representation of the proposed AOP:
**Fig 19:** AOP scheme
MIE: Redox cycling of a chemical initiated by electrons released by the mitochondrial respiratory chain

How this Key Event works:

Redox cycling is a process of alternate reduction and reoxidation steps. It is triggered in the presence of chemicals able to accept an electron from a reductant to form free radicals (Fig. 20). These radicals due to their high reactivity may undergo electron transfer to molecular oxygen generating superoxide anion radical ($O_2^-$) (Kappus, 1986). As a result of electron transfer, the parent compound is regenerated and able to catalyse further $O_2^-$ production. Extent and direction of this reaction depend on both the concentration of the reactants and their reduction potentials relative to the $O_2/ O_2^- (E_0 = -160 mV$ at pH7 for a standard state of $1M O_2$; Sawer and Valentine, 1981). Compounds with more negative electron reduction potential will react faster being thus effective redox cycler. In addition, very negative $E_0$ limit the pool of possible reductants, which have a sufficiently low reduction potential to donate an electron.

Chemicals radicalization appears to be the consequence of one electron reduction often catalysed by a flavoprotein (Cohen 1987). A number of different enzymes are involved, including mitochondrial reductases.

Electron transport through the mitochondrial respiratory chain (oxidative phosphorylation) is mediated by five multimeric complexes (I–V) that are embedded in the mitochondrial inner membrane (Fig 21).

---

Fig. 20: Schematic representation of the mechanism of chemicals redox cycling. (Modified by Cohen and Doherty, 1987).

Fig. 21. The electron transport chain in the mitochondrion. Complex I (NADH-coenzyme Q reductase or NADH dehydrogenase) accepts electrons from NADH and serves as the link between glycolysis, the citric acid cycle, fatty acid oxidation and the electron transport chain. Complex II also known as succinate-coenzyme Q reductase or succinate dehydrogenase, includes succinate dehydrogenase and serves as a direct link between the citric acid cycle and the electron transport chain. The coenzyme Q reductase or Complex III transfers the electrons from
CoQH$_2$ to reduce cytochrome c which is the substrate for Complex IV (cytochrome c reductase). Complex IV transfers the electrons from cytochrome c to reduce molecular oxygen into water. Finally, this gradient is used by the ATP synthase complex (Complex V) to make ATP via oxidative phosphorylation (Friedrich et al. 1994).

Under physiological conditions 1–5% of the oxygen is converted to O$_2^-$ by mitochondria due to electron leakage from the respiratory chain (Wei et al. 2001). Although different respiratory complexes and individual mitochondrial enzymes are sources of O$_2^-$ (Fig 22), leaking electron are primarily produced at two discrete points in the electron-transport chain namely at CI (NADH) and CIII (ubiquinone-cytochrome c reductase) (Selivanov et al. 2011).

**Fig. 22:** Chemical redox cycling in mitochondria. Complex I and Complex III start PQ redox cycle in bovine heart and brain mitochondria respectively, while the involvement of outer mitochondrial membrane NADH-oxidoreductase is controversial. OMM: outer mitochondrial membrane, IMM: inner mitochondrial membrane (Friederich et al., 1994).

NADH-ubiquinone oxidoreductase is the Complex I (CI) of electron transport chain (ETC). It is a large assembly of proteins that spans the inner mitochondrial membrane. In mammals, it is composed of about 45–47 protein subunits (human 45) of which 7 are encoded by the mitochondrial genome (ND1, ND2, ND3, ND4, ND4L, ND5, and ND6) and the remainder by the nuclear genome (Greenamyre, 2001). Complex I oxidizes NADH elevating the NAD+/NADH ratio by transferring electrons via a flavin mononucleotide (FMN) cofactor and several iron-sulfur centers to ubiquinone (Friedrich et al., 1994).

Complex III (CIII) of the ETC is the ubiquinol cytochrome C oxidoreductase, or coenzyme Q reductase. Like CI, CIII is also an assembly of multiple proteins spanning the inner mitochondrial membrane. One of the 11 CIII subunits is encoded by mtDNA, while nuclear DNA codes the remaining 10 proteins. CIII transfers electrons from CoQH$_2$ to reduce cytochrome C, which is the substrate for Complex IV (Fig. 1).

In presence of a redox cycling chemical, the leaking electrons from these complexes are readily accepted and transferred to molecular oxygen starting the redox cycling and boosting O$_2^-$ production.

**How it is measured or detected**

Redox cycling of a chemical can be measured directly or indirectly by different methods.
1. Direct detection of redox cycling by electron paramagnetic resonance (EPR):
A radical with an unpaired electron, like the PQ$^{•+}$ radical, has a distinctive EPR spectrum because of the delocalization of the unpaired electron across the conjugated ring system. Thus, it can be measured by EPR, which is a sensitive, specific method for studying radicals formed in chemical reactions and the reactions themselves (Schweiger & Jeschke 2001 Principles of Pulse Electron Paramagnetic Resonance, Oxford University Press). An example of an EPR spectrum of the PQ$^{•+}$ radical is shown in Fig. 23.

![EPR Spectrum](image)

**Fig. 23:** Detection and quantification of the PQ$^{•+}$ radical by EPR spectroscopy. (A) Typical EPR spectrum of the PQ$^{•+}$ radical (100 mM; trace a) generated in vitro by reduction of PQ$^{2+}$ with a two-fold excess of sodium dithionite. EPR signal of the SO$^{•-}$ radical present in the dithionite solution (10 mM; trace b). Modified after Cocheme and Murphy, 2009 Methods in Enzymology.

2. Direct detection of chemical radical formation by spectrophotometry:
Each chemical radical with a distinct absorbance spectrum than the parent compound can be measured spectrophotometrically in isolated mitochondria. However, due to the fast reaction of the chemical radical with oxygen, these measures have to be performed under anaerobic conditions (Cocheme and Murphy, 2009).

3. Direct detection of chemical radical formation (aromatic cations) by selective electrodes:
Selective electrodes were constructed and used for measuring the concentration of lipophilic cations in real time in mitochondrial incubations (Brand 1995; Murphy and Smith, 2007; Cocheme and Murphy, 2009).

3. Direct detection of superoxide anion formation

The methods for superoxide detection are described by Grivennikova and Vinogradov (2013). A range of different methods is also described by BioTek (http://www.biotek.com/resources/articles/reactive-oxygen-species.html). The reduction of ferricytochrome c to ferrocytochrome c may be used to assess the rate of superoxide formation (McCord, 1968). Like in other superoxide assays, specificity can only be obtained by measurements in the absence and presence of superoxide dismutase. Oxidation of hydroethidine (HE) to 2-OH-E$^{•+}$, together with non specific oxidation to ethidium and dimeric ethidium products to exclude the formation of oxidants other than superoxide, is also used as an indicator of superoxide anion formation (Dranka et al. 2012). Chemiluminescent reactions have been used for their increased sensitivity with lucigenin or coelenterazine as substrates. Hydrocyanine dyes are fluorogenic sensors for superoxide and hydroxyl radical, and they become membrane impermeable after oxidation (trapping at site of formation). The best characterized of these probes are Hydro-Cy3 and Hydro-Cy5. Generation of superoxide in mitochondria can be visualized using fluorescence microscopy with MitoSOX™ Red reagent (Life Technologies). MitoSOX™ Red reagent is a cationic derivative of dihydroethidium that permeates live cells and accumulates in mitochondria.
4. Indirect detection of superoxide anione formation

The enzyme aconitase contains an iron-sulfur cluster at its active site, which is highly sensitive to inactivation by O$_2^-$ (Gardner, 2002). Levels of O$_2^-$ production can, therefore, be inferred from the rate of aconitase inactivation during mitochondrial incubations. Aconitase activity is measured spectrophotometrically by a coupled enzyme assay, linking isocitrate production by aconitase to NADPH formation by isocitrate dehydrogenase (Gardner, 2002; (Cocheme and Murphy, 2009)).

Evidence supporting taxonomic applicability (tissue type, taxa, life stage, sex)


Mitochondria as a major site of mitochondrial superoxide production by PQ are supported in rodents, flies and yeast. Thus, mice heterozygous for MnSOD (the isoform of superoxide dismutase locate in the mitochondrial matrix) (Van Remmen et al. 2004) and flies silenced for MnSOD (Kirby et al., 2002) show greater sensitivity to PQ than the control; flies overexpressing catalase in mitochondria are resistant to PQ, whereas enhancement of cytosolic catalase was not protective (Mockett et al., 2003); human peroxiredoxin 5 in mitochondria protects yeast more efficiently against PQ than expression in the cytosol (Tien Nguten-nhu et al., FEBS let 2003).

Complex I has a highly conserved subunit composition in eukaryotes (Cardol, 2011). Fourteen subunits are considered to be the minimal structural requirement for physiological functionality of the enzyme. These units are well conserved between, Bacterial (E. coli), human (H. sapiens), and Bovine (B. Taurus) (Vogel et al., 2007; Ferguson, 1994). However, the complete structure of Complex I is reported to contain between 40 to 46 subunits and the number of subunits differs, depending on the species (Gabaldon 2005; Choi et al., 2008).

Complex I is well-conserved across species, from lower organism to mammals. In vertebrates it consists of at least 46 subunits (Hassinen, 2007), including human in which 45 subunits were found (Vogel et al. 2007). Moreover, enzymatic and immunochemical evidence indicate a high degree of similarity between mammalian and fungal counterparts (Lummen, 1998). Mammalian complex I structure (Vogel et al., 2007) and activity is characterized in detail, referring to different human organs including brain. There is also substantial amount of studies performed on human muscles, brain, liver as well as bovine heart (Okun et al., 1999).

Yeasts lack Complex I but reduce PQ in dependence on NADPH by intramitochondrial NADPH dehydrogenases (Cocheme et al., 2008).

Evidence for Chemical Initiation of this Molecular Initiating Event (MIE)

The most studied examples of chemicals that accept an electron from the mitochondrial respiratory chain and undergo redox cycling in dopaminergic neurons are the three bipyridyl herbicides paraquat, diquat and benzyl viologen. Substantial evidence has accumulated in the existing literature suggesting a role for these chemical, and paraquat in particular, and this AOP. Therefore, the redox cycler paraquat will be discussed in the context of all KEs identified in this AOP.

1. Paraquat as a mitochondrial electron acceptor.

The cellular toxicity of PQ is essentially due to its redox cycling abilities. Mitochondria are a major source of PQ-induced ROS production in brain (Castello et al. 2007; Fig. 24).
Fig. 24: PQ$^2$-induced H$_2$O$_2$ production in cellular fractions from the brain. Fluorometric (a) and polarographic (b) assays were used to measure H$_2$O$_2$ in the presence of malate and glutamate following the addition of 250 M PQ$^2$ to equal amounts of protein from each rat brain fraction: mitochondria (solid line), cytosol (dotted line), and homogenate (dashed line). (from: Castello et al. 2007, Fig. 2).

The early involvement of mitochondria in PQ-induced oxidative stress has been also demonstrated in whole cells overexpressing reduction-oxidation sensitive fluorescent proteins targeted to mitochondria or the cytosol (Rodriguez-Rocha 2013, Filograna et al., 2016). PQ (0.1-1mM) dose-dependently increases oxidative stress in SK-N-SH cells mitochondrial matrix at 24h with no changes in the cytosol (Fig.25) (Rodriguez-Rocha 2013). Accordingly, PQ 0.5 mM increases mitochondrial ROS production in SH-SY5Y after 6 and 12h with no evidence in the cytosol (Filograna et al., 2016). Significant cytoplasmic oxidative stress is evident only after 48h starting from PQ 0.5 mM, but not for lower concentrations (Fig. 25) (Rodriguez-Rocha 2013). A selective involvement of mitochondria is thus dose and time dependent.
Fig. 25: Alterations of mitochondrial and cytosol redox state following exposure to PQ of cells expressing fluorescent redox probe targeted to mitochondria (Mito-roGFP) or cytosol (roGFP). Cells were co-stained with PI and only viable cells were analyzed. Alteration in the redox state were determined by ratiometric analyses of changes in (Mito-)roGFP fluorescence at 407/488 ex and 530 em normalized with respect to control values. Data represents means ± SE of at least five independent experiments. *p<0.05 vs control values (from Rodriguez-Rocha et al. 2013).

In addition, higher protection against PQ toxicity is reached with mitochondrial, rather than cytosolic, expression of antioxidant enzymes (Mockett et al., 2003; Tien Nguyen-nhu and Knoops, 2003, Rodriguez-Rocha et al., 2013; Filograna et al., 2016). Accordingly, the deficiency of the isoform of mitochondrial superoxide dismutase (MnSOD) or mitochondrial thioredoxin reductase (necessary to maintain the H2O2 detoxifying thioredoxin/peroxiredoxin system) increases sensitivity to PQ (Kirby et al. 2002; Van Remmen et al 2004; Lopert et al., 2012).

Both Complex I (Cocheme and Murphy 2008) and Complex III (Castello et al., 2007; Drechsel and Patel 2009) have been involved in PQ radicalization. In Castello et al 2007, the redox cycle-initiating electrons are accepted from complex III and to a minor part by complex I as inhibition of complex I by rotenone only partially inhibited PQ-induced ROS formation in isolated brain mitochondria or rat midbrain cultures; while PQ-induced ROS formation in these systems was completely blocked after inhibition of complex III by using antimycin A (Drechsel & Patel 2009; Castello et al. 2007). That complex I is not the major source of electrons triggering PQ toxicity is supported by Choi et al. (2008) who demonstrated that silencing a major component of complex I abolishing its activity does not protect against PQ-dependent dopaminergic cell death. On the other hand, Cocheme and Murphy (2008) demonstrated that PQ accumulates into yeast and bovine heart mitochondrial matrix in dependence on mitochondrial membrane potential. In heart mitochondria, PQ is then reduced mainly by Complex I forming the radical which rapidly react with O2 to give O2−. The Authors explain this discrepancy with differences existing between brain and heart mitochondria (Cocheme and Murphy 2008, Drechsel and Patel 2009). The involvement of mitochondrial enzymes other than Complex I and III (VDAC and Cytb5, located at the external mitochondrial membrane) remains controversial (Shimada et al., 2009; Nikiforova et al. 2014) and potentially excluded by the recent observation that the main site of PQ reduction is inside mitochondria (Nikiforova et al. 2014).

2. General characteristics of other mitochondrial redox cyclers

Other mitochondrial redox cyclers include two other bipyridyl herbicides, diquat and benzyl viologen (Fig. 26, A ad B). These share common structural features with paraquat (Fig. 7 C): all compounds are composed of two aromatic rings containing a positively charged nitrogen and are thus good electron acceptors and redox cyclers (Drechsel & Patel 2009; Sandy et al. 1986).
Fig. 26: Molecular structures of: A diquat, B benzyl viologen, C paraquat

Quinones (i.e. menadione, Adriamycin) and nitroaromatic compounds (i.e. nitrofurantoin) also radicalize following one electron reduction by mitochondrial reductases (complex I and III and external mitochondria NADH-oxidoreductase) establishing a redox cycle (Frei et al. 1986; Nikiforova et al., 2014). Intriguingly, free cytosolic dopamine spontaneously oxidizes to produce different quinones like dopamine-\(\cdot\)-quinone and aminochrome. Aminochrome can undergo a one-electron reduction by NAD(P)H flavoproteins generating a leukoaminochrome-\(\cdot\)-semiquinone radical and giving rise to redox cycle with production of superoxide anion (Fig. 27) (Zoccarato et al., 2005; Munoz et al., 2012).

Fig. 27: One electron reduction of aminochrome (adapted from Munoz et al., 2012, Fig. 6).

Aminochrome has been recently suggested to play a role in the death of dopaminergic neurons containing neuromelanin triggering oxidative stress/mitochondrial dysfunction, the formation of \(\alpha\)-synuclein and impaired protein degradation (Munoz et al., 2012).
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KE1: Mitochondrial reactive oxygen species (ROS) formation and 
dysfunction

How this Key Event works:

O2-, generated by redox cycling drives a cascade of active oxygen species (ROS). O2 may:

- spontaneously or in a reaction catalysed by mitochondrial superoxide dismutase (MnSOD) and 
CuZnSOD (primarily cytoplasmic but also present in the peroxisome, lysosome, nucleus and 
mitochondrial intermembrane space) lead to the production of hydrogen peroxide (H2O2), which in 
turn will favour the formation of hydroxyl anion and hydroxyl radical through the Fenton reaction (Fig. 
1; Turrens 2003).

- react with nitric oxide (NO), which can be simultaneously produced in mitochondria by a unique form 
of nitric oxide synthase locted at the mitochondrial matrix ( Turrens 2003), to form peroxynitrite which 
may further convert to peroxynitrous acid and may yield nitrogen dioxide and hydroxyl radical (Pryor 
et al. 1995).

Other possible reactions triggered by free radicals originating from redox-cycle are: hydrogen atom 
abstraction and covalent binding to tissue macromolecules by radical addition to carbon-carbon double 
bonds or by radical combination.
Figure 28. Reactive oxygen species. Two molecules of superoxide can react to generate hydrogen peroxide
\((\text{H}_2\text{O}_2)\) in a reaction known as dismutation, which is accelerated by the enzyme superoxide dismutase (SOD). In the presence of iron, superoxide and \(\text{H}_2\text{O}_2\) react to generate hydroxyl radicals. In addition to superoxide, \(\text{H}_2\text{O}_2\) and hydroxyl radicals, other reactive oxygen species (ROS) occur in biological systems, which can be generated from singlet oxygen by antibody molecules 65,66. The colour coding indicates the reactivity of individual molecules (yellow, limited reactivity; orange, moderate reactivity; red, high reactivity and non-specificity) (Modified by Lambeth 2004).

ROS amount can be counter-balanced by natural antioxidants under physiological conditions. As such, mitochondria are equipped with several antioxidant systems (vitamin E, phospholipid hydroperoxide glutathione peroxide, MnsOD, cytochrome C, catalase, glutathione, glutathione-S-transferase, glutathione-reductase, glutathione peroxidase, peroxideroxins) (Andreyev et al., 2004). Nevertheless, antioxidant response might be overwhelmed by aberrant augmented levels of ROS that react with mitochondrial macromolecules such as lipids, proteins, nucleic acids and carbohydrates (Murphy 2009) leading to mitochondrial dysfunction, cell death and subsequently to organ pathogenesis. Indeed, oxidative stress is considered as a contributor to the pathogenesis of chronic health problems among which neurodegenerative conditions (Halliwell and Gutteridge, 2007).

It is well established in the existing literature that mitochondrial dysfunction may result in: (a) an increased ROS production and a decreased ATP level, (b) the loss of mitochondrial protein import and protein biosynthesis, (c) the reduced activities of enzymes of the mitochondrial respiratory chain and the Krebs cycle, (d) the loss of the mitochondrial membrane potential; (e) the loss of mitochondrial motility, causing a failure to re-localize to the sites with increased energy demands, (f) the destruction of the mitochondrial network, (g) increased mitochondrial Ca\(^{2+}\) uptake, causing Ca\(^{2+}\) overload (reviewed in Lin and Beal, 2006; Graier et al., 2007), (h) the rupture of the mitochondrial inner and outer membranes, leading to the release of mitochondrial pro-death factors, including cytochrome c (Cyt. c), apoptosis-inducing factor, or endonuclease G (Braun, 2012; Martin, 2011; Correia et al., 2012; Cozzolino et al., 2013), which eventually leads to apoptotic, necrotic or autophagic cell death (Wang and Qin, 2010). Due to their structural and functional complexity, mitochondria present multiple targets for various compounds.

How it is measured or detected

I. Reactive oxygen species production

Different ROS including hydrogen peroxide and the hydroxyl radical or the consumption of the ROS detoxifying substance glutathione as well as ROS-dependent cellular damage like lipid peroxidation or oxidation of protein or DNA can be measured by a variety of assays. Direct assays are based on the chemical modification of fluorescent or luminescent reporters by ROS species. Indirect assays assess cellular metabolites, the concentration of which is changed in the presence of ROS (e.g. glutathione, malondialdehyde, 4-hydroxynonenal, isoprostanes, etc.). The assays described below are not comprehensive.

1. Detection of hydrogen peroxide (\(\text{H}_2\text{O}_2\)) production

There are a number of fluorogenic substrates, which serve as hydrogen donors that have been used in conjunction with horseradish peroxidase (HRP) enzyme to produce intensely fluorescent products in the presence of hydrogen peroxide (Zhou et al., 1997; Ruch et al., 1983). The more commonly used substrates include diacetyl dichloro-fluorescein, homovanillic acid, and Amplex® Red (https://www.thermofisher.com/order/catalog/product/A22188). In these assays, increasing amounts of \(\text{H}_2\text{O}_2\) leads to increasing amounts of fluorescent product (Tarpley et al., 2004).

2. Measurement of the cellular glutathione (GSH) status

GSH is regenerated from its oxidized form (GSSH) by the action of a NADPH-dependent reductase (GSSH + NADPH + H\(^+\) → 2 GSH + NADPH\(^+\)). The ratio of GSH/GSSG is therefore a good indicator for the cellular NADPH/NADPH ratio (i.e. the redox potential). GSH and GSSG levels can be determined by HPLC, capillary electrophoresis, biochemically with DTNB (Ellman’s reagent, 5,5’-dithio-bis-[2-nitrobenzoic acid]) or by mean of luminescence-based assays (for example, GSH-Glo™ Glutathione Assay, https://www.promega.co.uk/resources/protocols/technical-bulletins/101/gsh-glo-glutathione-assay-protocol/). As excess GSSG is rapidly exported from most cells to maintain a constant...
3. Measurement of ROS- scavenging enzymes activity

Increased activity of scavenging enzymes like catalase, superoxide dismutase (SOD) and glutathione-S-transferase activity (GST) is indicative of ROS-production. The enzymes are recovered both in cells and tissues homogenates, thus providing a tool to measure the occurrence of ROS ex-vivo (Mitra et al., 2011) as an alternative to the measurement of lipid peroxidation. Measurements are based on the detection of chromogen, sensitive to the ROS specifically produced by the investigated enzyme, by spectrophotometric methods as described by Shina et al. (1972; for catalase); Pabst et al. (1974, for GST) and Kakkar et al. (1984, for SOD).

4. Quantification of lipid peroxidation

Measurement of lipid peroxidation has historically relied on the detection of thiobarbituric acid (TBA)-reactive compounds, such as malondialdehyde generated from the decomposition of cellular membrane lipid under oxidative stress (Pryor et al., 1976). This method is quite sensitive, but not highly specific. A number of commercial assay kits are available for this assay using absorbance or fluorescence detection technologies. The formation of F2-like prostanoid derivatives of arachidonic acid, termed F2-isoprostanes (IsoPs) has been shown to be more specific for lipid peroxidation. A number of commercial ELISA kits have been developed for IsoPs, but interfering agents in samples require partial purification before analysis. Alternatively, gas chromatography–mass spectrometry (GC-MS) may be used as a robust, specific and sensitive method.

5. Detection of peroxynitrite

There are three major approaches for peroxynitrite detection, including electrochemical sensors, detection of nitrotyrosine formation, and fluorescent probes (Chen X, Biomed J. 2014).

II. Mitochondrial dysfunction assays assessing a loss-of function

1. Cellular oxygen consumption

Electrons, fed into the mitochondrial respiratory chain either by complex I or complex II, ultimately reduce molecular oxygen to water at complex IV. In a closed system, this consumption of oxygen leads to a drop of the overall O₂ concentration, and this can serve as parameter for mitochondrial respiratory activity. Measurements are traditionally done with a Clark electrode, or with more sophisticated optical methods. At the cathode of a Clark electrode, oxygen is electrolytically reduced, which initiates a current in the electrode, causing a potential difference that is ultimately recorded. Clark electrodes however have the disadvantage that oxygen is consumed. Furthermore, interferences with nitrogen oxides, ozone, or chlorine is observed (Stetter et al. 2008). To circumvent these limitations, optical sensors have been developed that have the advantage that no oxygen is consumed, combined with a high accuracy and reversibility. Optical oxygen sensors work according to the principle of dynamic fluorescence quenching. The response of the respective fluorescence dye is proportional to the amount of oxygen in the sample investigated (Wang et al. 2014). In a model of isolated mitochondria in the absence of complex II substrates, oxygen consumption can serve as surrogate readout for the assessment of the degree of complex I inhibition. It is however essential to realize that also complex III and complex IV activities are involved and their inhibition also results in a decline in O₂ consumption. In addition to that, CI inhibitors can lead to a one-electron reduction of molecular oxygen at the site of Cl to yield superoxide. The amount of superoxide formed hence contributes to the consumption of oxygen, but must not be interpreted as oxygen consumption as a result of controlled and coupled electron flux through the complexes of the mitochondrial respiratory chain. A modern convenient method to measure oxygen consumption is provided by the Seahorse technology of extracellular flux (XF) analysis, in which cells are kept in a very small volume, so that changes of oxygen levels can be detected very sensitively by an oxygen sensor. To allow manipulation of the mitochondria in cells, the cell membrane can be permeabilized with saponin (SAP), digitonin (DIG) or recombinant perfringolysin O (rPFO) (XF-plasma membrane permeabilizer (PMP) reagent), to allow addition of specific substrates to measure activity of different respiratory chain complexes, including complex I. (Salabei et al., 2014).
The oxygen consumption parameter can be combined with other endpoints to derive more specific information on the efficacy of mitochondrial function. One approach measures the ADP-to-O ratio (the number of ADP molecules phosphorylated per oxygen atom reduced (Hinkle, 1995 and Hafner et al., 1990). The related Phosphate/Oxygen (P/O) ratio is calculated from the amount of ADP added, divided by the amount of O consumed while phosphorylating the added ADP (Ciapaite et al., 2005).

2. Mitochondrial membrane potential (Δψm)

The mitochondrial membrane potential (Δψm) is the electric potential difference across the inner mitochondrial membrane. It requires a functioning respiratory chain in the absence of mechanisms that dissipate the proton gradient without coupling it to ATP production. The classical, and still most quantitative method uses a tetraphenylphosphonium ion (TPP⁺)-sensitive electrode on suspensions of isolated mitochondria.

The Δψm can also be measured in live cells by fluorimetric methods. These are based on dyes which accumulate in mitochondria because of Δψm. Frequently used are tetramethylrhodamine ethyl ester (TMRE), tetramethylrhodamine, methyl ester (TMRM) (Petronilli et al., 1999) or 5,5',6,6'-tetrachloro-1,1',3,3′-tetraethylbenzimidazole carbocyanide iodide (JC-1). In particular, mitochondria with intact membrane potential concentrate JC-1, so that it forms red fluorescent aggregates, whereas de-energized mitochondria cannot concentrate JC-1 and the dilute dye fluoresces green (Barrientos et al., 1999). Assays using TMRE or TMRM measure only at one wavelength (red fluorescence), and depending on the assay setup, de-energized mitochondria become either less fluorescent (loss of the dye) or more fluorescent (attenuated dye quenching).

3. Enzymatic activity of the electron transport system (ETS)

Determination of ETS activity can be determined following Owens and King's assay (1975). The technique is based on a cell-free homogenate that is incubated with NADH to saturate the mitochondrial ETS and an artificial electron acceptor [I - (4 -iodophenyl) -3 - (4 -nitrophenyl) -5-phenylte trazolium chloride (INT)] to register the electron transmission rate. The oxygen consumption rate is calculated from the molar production rate of INT-formazan which is determined spectrophotometrically (Cammen et al., 1990).

4. ATP content

For the evaluation of ATP levels, various commercially-available ATP assay kits are offered (e.g. Sigma, http://www.abcam.com/ATP-assay-kit-colorimetricfluorometric-ab83355.html), based on luciferin and luciferase activity. For isolated mitochondria various methods are available to continuously measure ATP with electrodes (Llaudet et al., 2005), with luminometric methods, or for obtaining more information on different nucleotide phosphate pools (e.g. Ciapaite et al., 2005).

II. Mitochondrial dysfunction assays assessing a gain-of function

1. Mitochondrial permeability transition pore (PTP) opening

The opening of the PTP leads to the permeabilization of mitochondrial membranes (Lemasters et al., 2009; Fiskum, 2000), so that different compounds and cellular constituents can change intracellular localization. This can be measured by assessment of the translocation of cytochrome c, adenosine kinase or the apoptosis-inducing factor (AIF) from mitochondria to the cytosol or nucleus. The translocation can be assessed biochemically in cell fractions, by imaging approaches in fixed cells or tissues, or by life-cell imaging of GFP fusion proteins (Single et al., 1998; Modjtabahi et al., 2006). An alternative approach is to measure the accessibility of cobalt to the mitochondrial matrix in a calcein fluorescence quenching assay in live permeabilized cells (Petronilli et al., 1999).

2. mtDNA damage as a biomarker of mitochondrial dysfunction

Various quantitative polymerase chain reaction (QPCR)-based assays have been developed to detect changes of DNA structure and sequence in the mitochondrial genome (mtDNA). mtDNA damage can be detected in blood after low-level rotenone exposure, and the damage persists even after CI activity...
has returned to normal. With a more sustained rotenone exposure, mtDNA damage can be also
detected in skeletal muscle. These data support the idea that mtDNA damage in peripheral tissues in
the rotenone model may provide a biomarker of past or ongoing mitochondrial toxin exposure
(Sanders et al., 2014).

Evidence Supporting Taxonomic Applicability
Redox cycling is a universal event occurring in any cells of any species as well as in bacteria and yeast
(Cocheme and Murphy, 2008).
Mitochondrial dysfunction is a universal event occurring in cells of any species (Farooqui and Farooqui,
2012). Many invertebrate species (e.g., *D. melanogaster* and *C. elegans*) are considered as potential
models to study mitochondrial functionality. New data on marine invertebrates, such as molluscs and
crustaceans and non-Drosophila species, are emerging (Martínez-Cruz et al., 2012). Mitochondrial
dysfunction can be measured in animal models used for toxicity testing (Winklhofer and Haass, 2010;
Waerzeggers et al 2010) as well as in humans (Winklhofer and Haass, 2010).

However, there seem to be different susceptibilities towards mitochondrial toxins between
mitochondria of different organs. For example, rotenone (complex I inhibitor) severely damage brain
mitochondria, whereas liver mitochondria remained virtually unaffected (Panov et al. 2005). Moreover,
liver mitochondria have much lower Ca2+ capacities, they evidently undergo mitochondrial
permeability transition (mPT), followed by apoptosis much easier than brain mitochondria, which can
withstand much higher Ca2+ concentrations than liver (Panov et al. 2007). Not only do mitochondria
differ between organs, but also between brain regions. Work from Dubinsky’s group found that striatal
mitochondria isolated from rats were more sensitive than cortical mitochondria in their response to
calcium, perhaps due to increased amounts of cyclophilin D, a mitochondrial permeability transition
pore component (Brustovetsky et al., 2003; LaFrance et al., 2005). Independent of the mitochondrial
transition pore, brain region-specific mitochondrial membrane potential and susceptibility towards
dysfunction of mitochondrial oxidative phosphorylation (OXPHOS) was also observed by Pickrell et al.
(2011). Here the striatum was found to be especially sensitive towards disturbance of OXPHOS due to
the high striatal mitochondrial OXPHOS and membrane potential, which is prone to collapse when
OXPHOS activity is reduced. This instance becomes important when studies on compound effects on
isolated mitochondria are not of the correct origin, which would – for studying Parkinsonism – be the
brain, and here the nigrostriatal area. In addition to mitochondrial differences between organs and
intra-organ regions, species-specific mitochondrial activity was also measured. E.g., inhibition of
complex III with Antimycin A causes significantly higher ROS formation in mouse than rat brain
mitochondria suggesting a species-specific susceptibility to compounds interfering with complex III
across species (Panov et al. 2007). If human brain mitochondria are more similar to mouse or rat
mitochondria remains so far enigmatic.
References


Zhou, M., Z.Diwu, Panchuk-Voloshina, N. and R.P. Haughland (1997), A Stable nonfluorescent derivative of resorufin for the fluorometric determination of trace hydrogen peroxide: application in

**KE2. Impaired proteostasis**

See AOP 1 (p. 99) for the description of this KE

**KE3: Neuroinflammation**

See AOP 1 (p. 109) for the description of this KE

**KE4: Degeneration of dopaminergic neurons of nigrostriatal pathway**

See AOP 1 (p. 104) for the description of this KE

**Adverse Outcome: Parkinsonian motor deficits**

See AOP 1 (p. 113) for the description of the AO

**KEY EVENTS RELATIONSHIPS (KERs)**

1st KER: Chemical redox cycling in mitochondria leads to mitochondrial reactive oxygen species (ROS) production and dysfunction

How this Key Event Relationship works

Chemical redox cycling is triggered in the presence of chemicals able to accept an electron from a reductant to form a mono-cation free radical. Compounds with a lower electron reduction potential than O₂ will react fastest and the newly formed free radical, in the presence of oxygen, will re-oxidize generating the superoxide radical O₂⁻ (Kappus, 1986). The radical species may then be reformed from the parent compound reacting with oxygen again and establish a futile redox cycle boosting O₂⁻ production (Cohen and Doherty 1987). Mitochondria may represent the major site of chemical redox cycling, although several membrane and cytosolic enzymes may trigger this reaction. This has been demonstrated for PQ where alterations of mitochondrial redox state occurs earlier in mitochondria than in the cytosol (Castello et al. 2007; Rodriguez-Roche et al. 2013, Filograna et al., 2016) and higher protection from its toxicity is reached with mitochondrial, rather than cytosolic, expression of antioxidant enzymes (Mockett et al., 2003; Tien Nguyen-nhu and Knoops, 2003; Rodriguez-Roche et al. 2013, Filograna et al., 2016). Excessive generation of superoxide within mitochondria, as it occur in the presence of a chemical redox cycler like PQ, will start a cascade of active oxygen species that will overwhelm antioxidant response and damage DNA, proteins, lipids and other mitochondrial components and function (Andreyev et al. 2014, Turrens 2003; Murphy 2009) (Fig. 29).
**Fig. 29:** Schematic representation of the mechanism of paraquat toxicity. A, cellular diaphorases; SOD, superoxide dismutase; CAT, catalase; GPX, glutathione peroxidase; Gred, glutathione reductase; PQ2+, paraquat; PQŸ+, paraquat cation radical; HMP, hexose monophosphate pathway; FR, Fenton reaction; HWR, Haber-Weiss reaction. (Dinis-Oliveira et al., 2006).

**Weight of Evidence**

That PQ-dependent superoxide anion formation causes H₂O₂ in mitochondria is promoted by concentration-response relationships, e.g. in Cocheme et al. 2009 (Fig. 4). In addition, the observation that SOD and GPx overexpressing transgenic mice protect against the PQ-induced Parkinson disease phenotype (Thiruchelvam et al. 2005) supports the evidence as these enzymes are involved in mitochondrial ROS detoxification, which occurs e.g. after paraquat exposure.

**Fig. 30:** Production of H₂O₂ and O₂ by PQ in mammalian mitochondria - effect of respiratory substrate, uncoupler, and respiratory inhibitors. (A to B) Example traces from the Amplex Red assay. Rat heart mitochondria (0.2 mg protein/ml) were incubated at 37°C in KCl buffer supplemented with 0.01% [w/v] BSA. Substrate (5 mM succinate or 5 mM glutamate/malate), PQ (0.1 or 1 mM), the uncoupler FCCP (1 mM), and the complex I inhibitor rotenone (4 mg/ml) were added as indicated. (C) Rates of H₂O₂ efflux determined from the above traces. Data are the means ± SD of three-four determinations. (D) O₂ production in rat heart mitochondria determined by the aconitase inactivation assay. Heart mitochondria (2 mg protein/ml) were incubated for 10 min at 37°C in KCl buffer supplemented with 0.1% [w/v] BSA. Substrate (5 mM succinate or 5 mM glutamate/malate) and PQ (0.1 or 1 mM) were present as indicated. Data are the means ± SD of three determinations (Cocheme et al. 2009).
Biological Plausibility

The biological plausibility evolves from the measured that (i) PQ is reaching the brain (Breckenridge 2013, Prasad 2007, Yin 2011, Liang 2013, Breckenridge 2014), (ii) PQ is taken up into nigrostriatal neurons (Rappold 2011) and mitochondria (Cocheme and Murphy, 2009; Castello et al. 2007) (iii) PQ is a redox cycler inducing $O_2^\cdot•$ production and a cascade of ROS in isolated rat brain mitochondria (Cocheme and Murphy, 2008, Castello et al., 2007) brain homogenates (Castello et al., 2007), yeast (Cocheme and Murphy, 2008) and brain cell cultures mitochondria (Castello et al., 2007, Rodriguez-Rocha et al., 2013, Huang et al., 2012, Dranka et al., 2012, Cantu et al. 2011) (iv) Uncontrolled $O_2^\cdot•$ production and oxidative stress, due to chemical redox-cycling or endogenous $O_2^\cdot•$ over-production, results in mitochondrial dysfunction, namely decreased activity of enzymes of the respiratory chain (Hinerfeld at al. 2014; De Oliveira et al., 2016), diminished ATP production (De Oliveira et al., 2016), decrease in mitochondrial membrane potential (De Oliveira et al., 2016, Huang et al., 2012), mitochondrial DNA damage (Murphy 2009). (v) This ROS formation can be blocked by inhibition of complex III activity (and to a lesser extent by complex I inhibition; Castello et al. 2007; Drechsel & Patel 2009), (vi) overexpression of Gpx rescues parkinsonian phenotype (Thiruchelvam et al. 2005).

Mitochondrial oxidative stress and mitochondrial dysfunction is a contributing factor in the etiology of PD ()

Empirical support for linkage

Existing in vitro and in vivo data shows that compound-induced mitochondrial redox cycling causes mitochondrial ROS formation and dysfunction. PQ dose-dependently increases mitochondrial $O_2^\cdot•$ production both in isolated mitochondria and brain cells. The effect occurs within minutes in isolated mitochondria exposed to PQ and hours in cell cultures. In any biological context the effect is dose dependent and cumulative in time. In vivo evidence supporting PQ-induced oxidative stress exists and is mainly based on the occurrence of lipoperoxidation. It has been demonstrated that PQ induces an early increase in oxidative stress in the mitochondrial matrix due to $O_2^\cdot•$ formation that is followed by subsequent oxidative stress in the cytosol (Rodriguez-Rocha et al. 2013). In both in vivo and in vitro studies mitochondrial dysfunction and cell death is reduced/prevented by overexpression of mitochondrial redox cycling with superoxide anion formation causes ROS formation and mitochondrial dysfunction

In Vitro

- Incubation of rat primary mesencephalic cells or a dopaminergic cell line, N27, with PQ 0.250 - 1 mM for 3 or 4h resulted in a dose-dependent reduction of aconitase activity significant for all the tested doses (Tab. 1) (Cantu et al., 2009; 2011). Aconitase is uniquely sensitive to $O_2^\cdot•$ mediated oxidative inactivation thus being an indirect marker of $O_2^\cdot•$ production. $O_2^\cdot•$ formation was coupled to a dose dependent $H_2O_2$ production after 2-6h exposure of both cell type to PQ. The effect was significant only for PQ 1mM at 2h (Tab.1, 17%), 0.5 and 1 mM at 4h (Tab. 1) and 0.25 – 1mM at 6h (Tab. 1) (Cantu et al., 2009; 2011). Cell death occurred only 18h after PQ exposure (n.e. after 4-6h) (Cantu et al., 2009; 2011). Mitochondrial aconitase has also been shown to be a source of $°OH$, probably Via Fenton chemistry initiated by the co-released Fe$^{2+}$ and $H_2O_2$ (Vasquez-Vivar et al., 2000). 60-70% reduction of mitochondrial aconitase expression in N27 cells resulted in a decreased $H_2O_2$ production, attenuation of respiratory capacity deficiency and death after PQ exposure (Cantu et al. 2011). On the contrary, overexpression of m-aconitase resulted in exacerbation of $H_2O_2$ production and increased primary mesencephalic neuron death (Cantu et al., 2009). Aconitase inhibition by PQ (0.1 and 1mM) has been reported also in yeast and bovine heart mitochondrial within minutes from the exposure (Cocheme and Murphy, 2008). This effect is coupled as well to a dose dependent (PQ 0.1, 0.5 and 1 mM) mitochondrial $H_2O_2$ formation and is a consequence of a mitochondrial membrane potential-dependent uptake of PQ dication (Cocheme and Murphy, 2008).

- In another study perfomed on primary mesencephalic neurons (Cantu et al. 2009) exposure to PQ 0.25 and 0.5 mM reduced aconitase activity of 43% and 58% respectively. A dose and time response response increase in $H_2O_2$
Exposure of human neuroblastoma SK-N-SH cells to PQ dose (0.2 - 1 mM) and time (6-72h) dependently increases the production of $O_2^{-}$, as measured by mitosox and electron paramagnetic resonance. PQ (0.5 mM)-induced $O_2^{-}$ production up to 48 h was due to mitochondria, being prevented by MnSOD (located in the mitochondrial matrix) but not by CuZnSOD (primarily localized in the cytosol). In addition PQ dose-dependently increases oxidative stress in the mitochondrial matrix at 24h and both in mitochondrial matrix and cytosol at 48h. A mitochondrial restricted ROS production after SH-SY5Y cell exposure to PQ 0.5 mM for 6 and 12h was also observed in another study (Filograna et al., 2016). MnSOD pretreatment significantly reduced mitochondrial oxidative stress and neuronal cell death induced by PQ 0.5mM at 48h, while CuZnSOD had no effect (Rodriguez-Rocha et al. 2013). Similar results were obtained by Filograna et al. (2016) in SH-SY5Y after 24h exposure to PQ. All together these data shows that PQ induces an early increase in oxidative stress in the mitochondrial matrix associated with $O_2^{-}$ production, which is followed by subsequent oxidative stress in the cytosol and is a trigger to neural cell death (Rodriguez-Rocha et al. 2013).

Paraquat (250 µM) induced $H_2O_2$ in the mitochondrial, but not in the cytosolic fraction of rat brain homogenates (Castello et al. 2007). These data indicate again that the mitochondrion is the primary place of PQ-induced ROS production in the cell.

Redox cycling of Paraquat (250 µM) involves complex III of the MRC as PQ-dependent $H_2O_2$ production of isolated rat brain mitochondria (2-3 min) or primary midbrain cell cultures (6 hrs) is antagonized by co-treatment with the complex III inhibitor Antimycin A and to a lesser extent by rotenone (inhibitor of complex I; Castello et al. 2007). These data are supported by Drechsel & Patel (2009), who confirmed that complex III of the MRC is the major player in PQ-induced ROS production in Malate and Glutamate-stimulated rat brain mitochondria (100 and 300 µM; measurements over 15 min) and primary midbrain cultures (300 µM, 8 hrs) by co-treatment with Antimycin A, while this group measured involvement of MRC complex I in PQ-induced ROS formation in isolated rat brain mitochondria only after exposure to 1 or 3 mM PQ (15 min measurement).

A neurotoxic concentration of PQ 0.1mM induces production of $O_2^{-}, H_2O_2$ and NO after 24h in SH-SY5Y. Oxidative stress is coupled to impairment of complex I and complex V activity, to a decrease mitochondrial potential and ATP production. All these effects are prevented by a 12h pre-treatment with carnosic acid, a diterpene with antioxidant properties (de Oliveira 2016). ROS production coupled to reduced ATP production and lipid peroxidation were also observed in SH-SY5Y differentiated cells exposed to PQ 10°·M for 48h (McCarthy et al. 2004), indicating that PQ ability to trigger an oxidative damage is function of dose and time of exposure.

In vitro, PQ toxicity both in terms of ROS production, mitochondrial dysfunction and neuronal death is rescued by several antioxidants namely EUK 134 and 189 (synthetic SOD/catalase mimetics) (Peng et al. 2005; Hinerfeld et al., 2014), Coenzyme Q10 (McCarthy et al., 2004), rasagiline and cabergoline through their ability to increase the expression of glutathione (Chau et al., 2009, 2010), carnosic acid through the increased expression of both mitochondrial and total glutathione and several other antioxidant enzymes (de Oliveira 2016). Similar results are obtained by decreasing the expression of mitochondrial enzymes involved in ROS production (i.e. mitochondrial aconitase) prior to PQ exposure (Cantu et al., 2011) or by over-expressing enzymes involved in $O_2^{-}$ dismutation (i.e. mitochondrial superoxide dismutase) (Rodriguez-Rocha et al., 2013; Choi et al., 2006). Accordingly, decreased expression or inhibition of detoxifying enzymes like thioredoxin reductase (involved in the conversion of $H_2O_2$ in $H_2O$) potentiates synergistically increase $H_2O_2$ levels and decreased maximal and reserve respiratory capacity following incubation with PQ oxidative stress and mitochondrial dysfunction in dopaminergic cells (Lopert et al., 2012).

Ex Vivo

Mitochondria isolated from the striatum of Sprague Dawley rats 24h after exposure to PQ 25 mg/kg produce a significant higher amount of $H_2O_2$ compared to controls (+150%) and display decreased complex I and IV activity (-37 and -21%), increased mitochondrial membrane potential, increased lipid peroxidation (+42%) and increased cardiolipin
oxidation/depletion (+12%). No changes were observed in cortical mitochondria from PQ treated animals. (Czerniczyniec et al., 2015). Increased O$_2^-$ production (50% and 20% for cortical and striatal mitochondria respectively), decreased aconitase activity (30% Cx, 50% Str), increased lipid peroxidation (20% Cx, 30% Str) and release of cytochrome c and AIF were also observed in mitochondria isolated from the cortex and the striatum of Sprague Dawley exposed to PQ (10 mg/kg) over 4 weeks (one injection weekly) (Czerniczyniec et al., 2013). These results show that both acute and prolonged in vivo exposure to PQ promotes mitochondrial O$_2^-$and ROS production coupled to mitochondrial dysfunction with the striatum more sensitive than the cortex.

**In Vivo**

- Paraquat (10 mg/kg i.p.) once a week for three weeks causes loss of dopaminergic neurons (TH+) after two weeks in mice in vivo. In parallel, 4-hydroxynonenal (4-HNE, time course) and nitrotyrosine proteins (single time point) (as markers of PQ-induced oxidative stress) were measured in TH+ cells of these animals. Lipid peroxidation at TH+ neurons is already significant after the 1st PQ injection (+200%) and increases up to 600% on the 2nd PQ injection. No nigral dopaminergic cell loss occurs after the 1st PQ injection, while a significant reduction of neurons is triggered by the 2nd injection (30%), suggesting a relationship with lipid peroxidation. That ROS was involved in the dopaminergic cell death was not only shown by these markers of peroxidation, but also shown by transgenic, human ferritin overexpressing mice (characterized by a decreased susceptibility to oxidative stress), which were protected against PQ-induced dopaminergic cell death and 4-HNE generation (McCormack et al. 2005).

- Mice exposed to PQ (5, 10, 20, 40, 80 mg/kg, twice a week, ip) displayed a dose-dependent increase in superoxide, catalase and glutathione s-transferase activity as measured in homogenate obtained from substantia nigra, (SN) frontal cortex and the hippocampus. ROS-scavenging activity dose dependently increased in all the three areas both at sublethal (PQ 5-10 mg/kg) and lethal doses (PQ 20-80 mg/kg) (Tab.1, data referred only to SN and non lethal doses). At PQ 5 and 10 mg/kg, ROS scavenging enzyme activity was specific for the brain since no increase was observed in peripheral organs. Part of the mice exposed to PQ 10/mg kg were also supplemented with α-tocopherol (20 mg/kg, after the last dose of PQ for five consecutive days, ip), which decreased SOD, catalase and GST activity in all three brain areas. All the animals displayed significant DA neuronal death, microglia activation and motor dysfunction at PQ 10 mg/kg (Mitra et al., 2011).

- In vivo administration of synthetic superoxide dismutase/catalase mimetics like EUK-134, 189, (mice, PQ 7mg/kg, ip every 2 days for 10 times; Peng et al. 2005), M40401 (rats, PQ 50·g infused in the SN); PEP-SOD (mice, PQ 10 mg/kg, ip,once; Choi 2006) fusion protein protects against PQ neurotoxicity. On the other hand, depletion of antioxidants systems exacerbates PQ toxicity.

- Subcutaneous administration of PQ (10 mg/kg, twice/week, 3 weeks) to null mice for glutathione (major antioxidant to maintain redox equilibrium in cells) significantly decreased aconitase activity (20%) and complex I activity (20%) in the striatum but not in the cortex. PQ has no effect in wild type mice (Liang et al. 2013).

**Quantitative Understanding of the Linkage**

PQ ability to trigger mitochondrial ROS production (O$_2^-$ and correlated species) by redox cycling has been demonstrated in vitro, both in isolated mitochondria, mitochondrial brain homogenates and cells and ex-vivo from brain mitochondria isolated from PQ-treated rats. In vivo evidence of oxidative stress, as a consequence of PQ exposure, is mainly supported by the occurrence of lipoperoxidation, accumulation of oxidized protein or by mean of sodium salicylate molecular trap.

PQ (0.1-1 mM) induces ROS production within minutes in isolated mitochondria and mitochondrial brain fraction (Cocheme and Murphy, 2008; Castello et al., 2007), while in cells this process is detectable after 2-6h from the exposure in dependence on the dose (Rodriguez-Rocha et al., 2013; Cantu et al., 2011, Huang et al, 2012, Dranka et al. 2012). Based on the work of Cantu et al. (2009), which compare O$_2$ and H$_2$O$_2$ production by PQ (0.25-0.5 mM) along different time points, O$_2$ formation
slightly precedes $\text{H}_2\text{O}_2$ production at the lowest PQ concentrations (Tab.1). In addition, at these time points no death is usually detected in cells exposed to PQ up to 1mM, pointing at ROS production as an early event preceding cell death.

Table 13: Quantitative evaluation of the KER.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>PQ redox cycling with superoxide formation</th>
<th>ROS formation (KE1)</th>
<th>reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rat primary mesencephalic cell culture PQ at 0.25 to 1mM</td>
<td>Inhibition of aconitase after 3: 43% at 0.25 mM 58% at 0.5 mM</td>
<td>Increase in $\text{H}_2\text{O}_2$ At 2 hours 17% at 1mM At 4 hours 28% at 0.5 mM and 64% at 1 mM At 6 hours 31% at 0.25mM, 59% at 0.5mM and 119% at 1mM</td>
<td>Cantu 2009</td>
</tr>
<tr>
<td>N27 cell culture, PQ at 0.3 mM to 1 mM</td>
<td>Inhibition of aconitase, 80% at 0.5 mM, 98% at 1mM at 4h</td>
<td>Increase in $\text{H}_2\text{O}_2$ at 4-6h 25% at 0.3 mM and 33% at 1mM</td>
<td>Cantu 2011</td>
</tr>
<tr>
<td>SK-N-SH human neuroblastoma cells treated with PQ 0.2mM up to 1 mM. 6 to 62 hours sampling</td>
<td>Dose and time related increase of $\text{O}_2$ by electro paramagnetic resonance spectroscopy. 50% at 0.2 mM, 80% at 0.5 mM and 150% at 1mM at 24 hours</td>
<td>Increase in DHE ROS production 800% at 0.5mM at 48 hours</td>
<td>Rodriguez Rocha 2013</td>
</tr>
<tr>
<td>SD rat treated at 25 mg/kg and observed 24 hours later</td>
<td></td>
<td>$\text{H}_2\text{O}_2$ increase of 150% in isolated mitochondria from SN neurons corresponding to 42% mitochondrial lipid peroxidation Decrease in Complex I 33% and Complex IV 21% Increase mitochondrial membrane potential</td>
<td>Czerniczyniec 2015</td>
</tr>
<tr>
<td>SD rat treated at 10 mg/kg weekly for 4 weeks</td>
<td>Increase in $\text{O}_2$ production in isolated mitochondrial of 20% Decrease in aconitase activity in mitochondrial of 50% in striatum</td>
<td>Increase in lipid peroxidation in isolated mitochondria of 30%</td>
<td>Czerniczyniec 2015</td>
</tr>
<tr>
<td>C57BL/6 mice treated with 10 mg/kg PQ i.p. once a week for three weeks</td>
<td></td>
<td>Increased neuronal lipid peroxidation measured 1 day after weekly injection each: 10mg kg ip 200% increase in lipid peroxidation at 2 and 4 days post inj 500 600% in lipid</td>
<td>McCormack et al. 2005</td>
</tr>
</tbody>
</table>
poxidation after 2nd injection 2/4 days after
After third injection limited response due to significant neuronal cell loss

Swiss albino mice ip at 5 and 10 mg/kg twice a week for four weeks
SOD activity ex vivo
At 5 mg/kg increase of 42%
At 10 mg/kg increase of 75%
Glutathione s transferase activity ex vivo
At 5 mg/kg increase of 25%
At 10 mg/kg increase of 75%
Catalase activity ex vivo
At 5 increase of 17%
At 10 increase of 50%

Mitra 2011
Neuronal cell loss of 40% TH positive and Fox 3 positive and motor dysfunction symptoms at 10 mg/kg and 10 % at 5 mg/kg. Motor symptoms only at 5 mg/kg

Uncertainties or inconsistencies

- Besides mitochondria, NADPH-oxidase 1 (NOX1) (Cristovao et al., 2012) and plasma membrane microglia NOX (Rappold et al., 2011) also contribute to PQ-induced ROS production. Furthermore, in vitro data suggest that for time points of exposure longer than 48h oxidative stress occurs both at mitochondria and cytosol in dependence to the dose. Thus it is difficult to discriminate the source of PQ-induced ROS and the early involvement of mitochondria in vivo due to the extensive treatments and to the indirect detection of oxidative stress mainly by mean of lipoperoxidation, protein oxidation. Mitochondrial involvement is suggested by ex-vivo studies (Czerniczyniec et al., 2013; 2015).

- Mitochondrial loss of function (i.e. decrease in mitochondrial membrane) might sometimes be the consequence of cell death rather than directly resulting from oxidative stress. This is due to the estimation of this parameter at time points already characterized by a significant cell death without a double staining, which allow discriminating between alive and dead cells. The observation that loss of mitochondrial membrane potential on PQ exposure is only detected in the population of dead cells when cells are double stained for mitochondrial membrane potential and plasma membrane integrity (Rodriguez-Rocha, 2013) support this uncertainty.
References


Castello PR, Drechsel DA, Patel M. Mitochondria are a major source of paraquat-induced reactive oxygen species production in the brain. J Biol Chem. 2007 May 11;282(19):14186-93


Chau KY, Cooper JM, Schapira AH. Rasagiline protects against alpha-synuclein induced sensitivity to oxidative stress in dopaminergic cells. Neurochem Int. 2010 Nov;57(5):525-9.


de Oliveira MR, Ferreira GC, Schuck PF. Protective effect of cardosic acid against paraquat-induced redox impairment and mitochondrial dysfunction in SH-SY5Y cells: Role for PI3K/Akt/Nrf2 pathway. Toxicol In Vitro. 2016


2nd KER: Mitochondrial dysfunction results in an impaired proteostasis

How this Key Event Relationship work

See AOP 1 (p. 153)

Weight of Evidence

See AOP 1(p. 153)

Biological Plausibility

See AOP 1(p. 153)

Empirical support for linkage

Based on the existing in vitro and in vivo data it is suggested that mitochondrial dysfunction impairs protein homeostasis through oxidative and nitrosative stress resulting in protein aggregation, damaged intracellular transport of proteins and cell organelles.

- Paraquat 0.5 mM decreases mitochondrial complex V activity, ATP production and proteasome activity in SH-SY5Y cells. All these effects increase in time (from 6 to 48h) and are significant at 24 and 48h of treatment. In addition, PQ significantly decreases proteasome 19S subunit but not 20S- only at 48h. However, since this 19S subunit drops later than proteasome activity decrease, it could not have caused proteasome dysfunction. Significant increased levels of a-syn and ubiquitinated proteins are also evident at 24 and 48h following PQ exposure. SH-SY5Y death occurred only at 48h. Cell death is dose dependent (PQ 0.05 – 1 mM) and is significant at 0.5 and 1mM (57 and 75% respectively). PQ induces mitochondrial dysfunction and proteasome impairments leading to neuronal death (Yang and Tiffany-Castiglioni, 2007).

- Reduced mitochondrial membrane potential and proteasome inhibition has been also observed for 0.2 mM PQ as early as 3h after exposure in SH-SY5Y cells. A slight but significant effect also occurs at 0.02 mM PQ at longer time (6h). 0.2 mM PQ-induced effects precede neuronal death (12h; no death observed at 0.02 mM). Transfection of the heat shock protein HDJ-1 (that attenuate protein aggregation without altering ROS production, as measured by DCF) in SH-SY5Y cells attenuates 0.2 mM PQ-induced mitochondrial membrane potential decrease at 6h (from 50% to 80%). This suggests that protein aggregation also contribute to the loss of mitochondrial membrane potential (Ding and Keller, 2001).

- Paraquat (10 mg/kg, once a week for 3 weeks) in combination with DJ-1 deficiency decreases ATP levels, proteasome activities, proteasome subunits levels and increases ubiquitinated proteins in the ventral midbrain including SNpc. None of these effects is observed at the striatum (Yang et al., 2007). DJ-1 has been suggested to contribute to mitochondrial integrity due to its localization in the mitochondrial matrix and inter-membrane space (Zhang et al, 2005 ) and its antioxidant action (Taira et al., 2004 ). Likewise, exposure to PQ and deficiency of DJ-1 might cooperatively induce mitochondrial dysfunction resulting in ATP depletion and contribute to proteasome dysfunction in the brain.

- Paraquat (10 mg/kg i.p.,) induced significant increase in lipid peroxides (LPO) in ventral midbrain (VM), striatum (STR) and frontal cortex (FCtx), maximum in VM after 5 doses (2.4 times the control). An elevated LPO level was still present in VM after 28 days. Moreover, the activity of 20S proteasome in STR was altered (increased 40-50%) after a single dose and slightly reduced after 5 doses (Prasad et al., 2007). The temporal activation of proteasomal activity at 1 and 24h after single dose was explained by the fact that carbonylated proteins moderately undergo degradation by UPS (Poppek and Grune, 2006). Sublethal proteasome inhibition induces neurons to increase proteasome activity and promotes resistance to oxidative injury (Lee et al., 2004).
## Quantitative evaluation of KERs

### Table 14: Quantitative evaluation of the KER.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Mitochondrial dysfunction (KE2)</th>
<th>Impaired protein degradation (KE3a)</th>
<th>reference</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td>SH-SY5Y cells, PQ 0.5 mM, 12, 24 and 48h</td>
<td>decreased activity of complex V (% of control; significant): 12h ne 24h 70% 48h 50% decreased ATP levels (% of control): 12h ne 24h 76% 48h 39%</td>
<td>decreased proteasome activity (% of control): 12h ns 24h 40% 48h 23% Decreased protein level of 19S subunit (% of control): 12h ne 24h ne 48h 32% ne on 20S a and b at any time Increased level of ubiquitinated proteins (% of control): 12h ne 24h 154.5% 48h 167% Increased protein level of α-syn: 12h ns 24h 236% 48h 305%</td>
<td>Yang and Tiffany-Castiglioni 2007</td>
<td>PQ induced significant SHSY5Y cells death only at 48h thus mitochondrial dysfunction and impaired protein degradation occurs before neurons die. Furthermore, the lack of effect on 20S subunits suggests that the observed paraquat effects were not nonspecific cytotoxic events. Levels of 19S dropped at 48 but not 24 h after paraquat treatment, and therefore could not have caused the proteasome dysfunction observed.</td>
</tr>
<tr>
<td>SH-SY5Y cells, PQ 20 and 200 μM, different time points</td>
<td>Reduced mitochondrial membrane potential (% of control): 20 μM- 6h approx. 80%? Reduced of 20% vs control 200μM- 3h approx. 60%? Reduced 40% vs control 6h approx 40% reduced 60% vs control</td>
<td>Reduced proteasome activity (% of control) 20uM- 6h 85% significant reduced of 15% vs control 200uM- 1h approx. 80% reduced of 20% vs control 3h approx 60% reduced of 40% vs control 6h approx. 55% reduced of 65% vs control</td>
<td>Ding and Keller, 2001</td>
<td>Death at 6h not measured, significant death at 24h for 20uM and 12h for 200uM Co-treatment with 20·M PQ + epoxomicyn 1·M (proteasome inhibitor) exacerbate PQ-induced mitochondrial membrane potential decrease (to 75% vs control or 60% vs 20nM PQ treated only) and cell death. The ability of increased</td>
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<td>Logically grouped information</td>
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<tr>
<td>SHSY5Y transfected with HDJ-1 (member of the Hsp40 family, attenuate protein aggregation), PQ 200uM for 6h</td>
<td>Partial significant (20% vs PQ treated only) recovery of mitochondrial membrane potential</td>
<td>Partial significant (25% vs PQ treated only) recovery of proteasome activity levels of HDJ-1 to attenuate proteasome inhibition did not appear to be due to a decrease in ROS levels, or altered levels of proteasome subunits.</td>
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<tr>
<td>Mice WT and DJ-deficient, 10 mg/kg PQ, once a week for three weeks</td>
<td>ATP levels in VMB decreased of 30% in DJ deficient (vs control)</td>
<td>Partial significant (20% vs PQ treated only) recovery of mitochondrial membrane potential</td>
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<tr>
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<td>-Proteasome activity in VMB reduced approx. 30% (vs control)</td>
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<td>-Ubiquitinated proteins increased levels in VMB 1.5 times the control</td>
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<td>- Proteasomal subunits (18S and 20S) levels decreased in VMB of approx. 30% (vs control)</td>
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<td></td>
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<td>Yang et al. 2007</td>
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<td>Effects evident only in VMB (include SNpC) and not in striatum and only in DJ-deficient mice. DJ-deficient as WT for all the parameters.</td>
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<td>Additional measurements:</td>
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<td>- Motor symptoms decreased of 40% (vs control) in DJ-defic only;</td>
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<td>- Dopamine levels decreased 30% (vs control)in DJ-defic only (BUT dopamine level in DJ mice not treated is higher than in WT control)</td>
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<td>- TH+ neurons stereol count: NO effects</td>
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<td>Thus concordance motor symptoms and decreased dopamine, but not effect on neurons: authors suggested that behavioural and neurochemical consequences manifest before dopamine neuron degeneration -</td>
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<tr>
<td>PQ 10 mg/kg i.p. (administered 3 times/week for a total of 1, 3 or 5 doses) in C57BL/6J mice</td>
<td>Increased tissue level of lipid peroxides (LPO) after a single (and persistent up to 28 days) and repeated doses, maximum in VM after 5 doses (2.4 times the control, lower in STR (80%) and least (66%) in FrCtx</td>
<td>INCREASED activity 20S proteasome in STR (not quant in other tissues) at 1 (40%) and 24h (50%) after single i.p. dose. 20S activity was reduced in STR after 5 doses (15%)</td>
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<td>Prasad et al., 2007</td>
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</table>
Uncertainties or inconsistencies

- The exact molecular link from mitochondrial dysfunction to disturbed proteostasis is not known. It is not clear which is the oxidative modification that drives the process.

- Proteostasis incidence is higher than mitochondrial dysfunction at PQ 0.5 mM (Yang and Tiffany-Castiglioni, 2007) but not at PQ 0.2 mM (Ding and Keller, 2001) at the same time point in SH-SY5Y cells. These results suggest that, in vitro, at doses higher than 0.2 mM PQ might involve mechanisms other than mitochondrial dysfunction.

- The sequence of events that link mitochondrial dysfunction to proteases inhibition is not entirely clear, proteosomal dysfunction might contribute to mitochondrial dysfunction (Ding and Keller 2001). On the other side, sublethal proteasome inhibition induces neurons to increase proteosome activity and promotes resistance to oxidative injury (Lee et al., 2004), whereas oxidative stress can increase proteasome activity early in the sequence leading to cell death in vitro (Holtz et al., 2006).

- A vicious circle is observed that make it difficult to establish an exact quantitative relationship between mitochondrial and proteosomal dysfunction. This task needs a better dose- and time-related definition of PQ effect on those two events that is actually lacking.

- Lack of evidences of the link between mitochondrial dysfunction and disturbed proteostasis in WT animals exposed to PQ.

- Distinct unfolded protein response (UPR) signaling branches could have specific and even opposite consequences on neuronal survival depending on the disease input (Hetz and Mollereau, 2014). Proteostasis impairment at the level of the endoplasmic reticulum (ER) is emerging as a driving factor of dopaminergic neuron loss in PD. ER stress engages the activation of the UPR adaptive reaction to recover proteostasis or trigger apoptosis of damaged cells. PQ may induce ER stress (Huang et al. 2012).

- A genetic screening in yeast revealed that one of the major physical targets of α-Synuclein is Rab1, an essential component of the ER-to-Golgi trafficking machinery (Cooper et al., 2006; Gitler et al., 2008). Over-expression of Rab1 in animal models of PD reduced stress levels and protected dopaminergic neurons against degeneration (Coune et al., 2011).
References


3rd KER: Impaired proteostasis leads to degeneration of DA neurons of the nigrostriatal pathway.

How this key event relationship works

See AOP 1 (p.141)

Weight of evidence for the KER

1. Biological plausibility

See AOP 1(p.141)
2. Empirical support for linkage

Large part of the empirical evidence supporting this KEs relationship comes from observational studies conducted in human affected by PD, from in vitro and in vivo studies conducted with the chemical stressors MPTP and rotenone or from experiments conducted with proteasome inhibitors. With the chemical stressor paraquat, used for the empirical support of this AOP, most of the studies where providing evidence that in the same experiments impaired proteostasis and neuronal degeneration where co-existing. Although different concentrations of paraquat when used in the in vitro assays, in vivo studies where generally conducted at fixed dose though different doses scheduling could have been applied.

Paraquat is an herbicide for which a unique sensitivity of dopaminergic neuronal cells was also observed (Uversky, 2004; McCormack et al. 2002; Brooks et al. 1999). Similarly to MPTP and Rotenone, also in paraquat treated mice, an up regulation and aggregation of α synuclein and inhibition of the proteosomal pathway was demonstrated in DA neurons in SN ( Manning-Bog et al. 2002; Wills et al.2012). Additionally, paraquat is able to reduce proteosomal function in DJ-1 deficient mice with an impaired clearance of altered proteins (Yang et al. 2007). Paraquat is clearly more toxic in aged animal or when co-administered with the fungicide maneb. (Thiruchelvam et al. 2003, McCormack et al. 2002). Duration of treatment could also impact neuronal loss (Ossowska et al. 2005).

Human evidences

- Human data from PD patients are indicative of an overall inhibition of axonal autophagy with an increased level of mTor (a major protein involved in autophagy) which was accompanied with an impairment to form autophagosome. The observed increase in m TOR levels was of 63% (Wills et al. 2012).
- Inclusion bodies in DA neurons (ie Lewy bodies), a pathological hallmark for sporadic PD, stains specifically for proteins associated with the UPS (Fornai et al. 2005, Mcnaught et al. 2002), including α-synuclein, parkin and ubiquitin; possibly indicating that failure of the UPS system represents a common step in the pathogenesis of PD and impairment of the proteasome system was found in humans affected by sporadic PD (McNaught et al. 2001, 2003).
- Lysosomal breakdown and autophagosome (AP) accumulation with co-localization of lysosomal markers in Lewy Bodies is reported to occur in PD brain samples where Lewy bodies were strongly immunoreactive for the autophagosomes markers (LC3II). (Dehay et al. 2010).
- Postmortem studies on PD patients show axonal pathology that is likely to precede the loss of neuronal bodies In this investigation, TH immunoreactive fibers had almost entirely disappeared with preservation of neuronal bodies (Orimo et al. 2005 and 2008).

Paraquat

- Paraquat (10 mg/kg ip once a week for 3 consecutive weeks) exposure in male mice (control mice and transgenic mice expressing either wild type human α-synuclein or mutant form of the human protein) induced in control mice accumulation of intracellular α-synuclein-immunoreactive deposits in 30% of dopaminergic neurons and decreases by 25-35% the number of TH positive and Nissl-stained neurons in SNpc following stereological evaluation. A protective effect (presence of intracellular protein positive deposits – 36%- with lack of neurodegeneration) was observed in animal overexpressing the wild as well as a mutated form of α-synuclein. In these animals a concomitant increase of HSP70 chaperone protein was observed (Manning-Bog et al. 2003). Heat shock proteins has been reported to play a protective role against PQ toxicity (Ding and Keller, 2001; Minois et al., 2001) and its increase may represent an adaptive change to high intraneuronal α-synuclein concentrations.
- Weekly ip injection of 10 mg/kg of paraquat for 3 weeks in male mice overexpressing α-synuclein induced loss of dopaminergic neurons in SNpc and decrease in TH optical density (slight) in the striatum which was accompanied by an increase of intracytoplasmic insoluble α-synuclein (Fernagut et al. 2007). (Similar decrease in dopaminergic neurons, without α-synuclein accumulation, was observed also in PQ-treated WT animals).
Administration of 10 mg/kg ip twice a week for 4 weeks to adult Swiss albino mice induced dopaminergic neuronal loss (ca. 40% reduction) in SN (also in FC and hippocampus) which was associated with a decrease in α-synuclein expression (ca. 50% reduction) (increased in hippocampus) and reduction of TH levels (ca. 50% reduction) in SN (and hippocampus) (Mitra et al. 2011). The reduced α-synuclein expression in SN, increased expression in hippocampus, and aggregated forms in FC might correlate with α-synuclein gene polymorphism associated with PQ-mediated neurotoxicity and the differential time frames necessary to initiate neurodegeneration in the different regions.

In male Wistar rat receiving four ip injections, separated by one day, of paraquat at 10 mg/kg/day, showed a 50% increase of α-synuclein immunoreactivity and protein level (by Western blot) in SN. The stereological count of TH-positive neurons showed that Nox 1 knockdown animals (stereotaxically injected with a viral constructed expressing Nox 1 or ablated for it) treated with Paraquat, significantly reduced PQ-elicited dopaminergic neuronal loss from 37% in the group treated with vector and PQ to 13% in the Nox 1 KO treated with PQ. Nox 1 knockdown reduced by 37% the PQ-mediated α-synuclein levels, compared to vector plus PQ, as well as α-synuclein aggregation and it was accompanied by a reduction in α-synuclein immunoreactivity and protein level as well as a decrease in α-synuclein aggregation (Cristovao et al. 2012).

Proteasome activity was investigated in dopaminergic SH-SY5Y cells treated with paraquat. Results showed that at a concentration of paraquat that reduced viability by about 60% at 48 h (0.5 mM) loss of proteasome activity occurred. Furthermore, paraquat-treated cells showed decreased protein levels of proteasome 19S subunits, but not 20S alpha or beta subunits, suggesting that the effects observed were not the result of general cytotoxicity. Paraquat also increased levels of alpha-synuclein and ubiquitinated proteins, suggesting that paraquat-induced proteasome dysfunction leads to aberrant protein accumulation (Yang et al. 2007).

Low concentration of paraquat (10 µM) induced autophagy in human neuroblastoma cells line (SH-SY5Y). Paraquat induced autophagic vacuoles (AV) and recruitment of LC3-GFP fusion protein to AV. Finally, cell death with hallmarks of apoptosis was observed. Paraquat also increased long-lived protein degradation which was blocked by the autophagy inhibitor 3-methyladenine (3-MA). While caspase inhibition retarded cell death, autophagy inhibition accelerated the apoptotic cell death induced by paraquat. (Gonzalez-Polo et al. 2007).

SH-SY5Y cell transfected with DJ-1-specific siRNA and exposed to paraquat showed additive effect on apoptotic cell death, inhibition of the cytoplasmic accumulation of autophagic vacuoles as well as recruitment of LC3 fusion protein to the vacuoles. The effect was time and dose related (25 to 500 µM); (Gonzalez-Polo et al. 2009). Apoptotic cell death was accelerated by treatment with the autophagy inhibitor 3-methyladenine (3-MA). Findings suggest an active role for DJ-1 in the autophagic response produced by Paraquat, providing evidence for the role of PD-related proteins in the autophagic degradation pathway.

Paraquat (500 µM) triggers endoplasmic reticulum stress and cell death (70% reduction in cell survival) and inhibits proteasomal activity (60-70% reduction) in a rat N27 mesencephalic dopaminergic cells system (Shankar et al. 2008).

Males C57BL/6NcrlVr mice received ip injectionsof 10 mg/kg of paraquat twice a week for 4 weeks showed a decrease in TH+ neurons of approximately 43%. This was accompanied by: increased of 133% of α-synuclein, increased by 13% (not statistically significative) in 19S proteasome function and decrease of 5% 20S proteasome function (not stat significative), increase by 43% in mTOR (autophagy inhibitor), increase by 81% of beclin-1 (autophagy inducer) and increase in Atg12 of 36% (Su et al. 2015).

Quantifiable understanding

A quantitative relationship has been established between the chemical stressor paraquat inducing impaired proteostasis and loss of DA neurons of nigrostriatal pathway. A response concordance was observed for the quoted studies; however dose and time relationship could be only established in a limited number of in-vitro studies as the in-vivo studies were conducted at single dose and single evaluation time-point.
<table>
<thead>
<tr>
<th>Impaired proteostasis</th>
<th>DA neurons degeneration</th>
<th>Treatment</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Intracellular deposit of in α-synuclein observed in 30% of DA neurons</td>
<td>Approx. 30 (25-35%) % of cell loss (TH positive cells) in SNpc</td>
<td>C57BL/6 mice treated with Paraquat once a week for 3 weeks at 10 mg/kg ip</td>
<td>Manning–Bog et al. 2003&lt;br&gt;Fernagut 2007</td>
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<tr>
<td>Increase of approx. 91% of α-synuclein inclusion (proteinase-K-resistant α-syn aggregates) only observed in α-synuclein overexpressing animals</td>
<td>Approx. 25% loss of DA neurons (stereological analysis TH-positive neurons) in both WT as well as α-synuclein overexpressing animals</td>
<td>Weekly ip administration of 10 mg/kg paraquat for 3 weeks in mice WT and overexpressing α-synuclein</td>
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<tr>
<td>Approx. 50% reduction in α-synuclein expression in SN</td>
<td>Approx. 40% loss of DA neurons (TH+ and FOX3+ neurons)</td>
<td>Paraquat 10 mg/kg ip twice a week for 4 weeks to adult Swiss albino mice</td>
<td>Mitra et al. 2011</td>
</tr>
<tr>
<td>Approx. 50% increase of α-synuclein expression (immunoreactivity and protein)</td>
<td>Paraquat significantly reduced PQ-elicited dopaminergic neuronal loss from 37%</td>
<td>Wistar rat receiving four ip injections, separated by one day, of paraquat at 10 mg/kg/day</td>
<td>Cristovao et al. 2012</td>
</tr>
<tr>
<td>Proteasome inhibition (approx.60% at 24 hours and 80% at 48h)</td>
<td>Reduction of 60% in cell viability at 48h</td>
<td>DA SH-SY5Y cells treated with paraquat 0.5mM</td>
<td>Yang et al. 2007</td>
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<tr>
<td>Increased protein levels of α-synuclein (2.3 fold at 24h and 3 fold at 48h)</td>
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<td>Increased ubiquinated protein levels (1.5 fold at 24h and 1.7 fold at 48h)</td>
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<td>Accumulation of AV (%vacuolated cell volume) at 6, 15 and 24h was 20, 40 and 45% respectively.</td>
<td>25% of nuclear apoptosis at 24h (caspase-3 maximum level)</td>
<td>DA SH-SY5Y cells treated with paraquat 10µM</td>
<td>Gonzalez-Polo et al. 2007</td>
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<tr>
<td>Inhibition of PQ-induced autophagic vacuolization and protein degradation after treatment with 3-MA</td>
<td>Apoptosis cell death was accelerated and caspase-3 activation increased after 3-MA treatment</td>
<td>DA SH-SY5Y cells treated with paraquat 10µM were then treated with prototypic autophagy inhibitor 3-MA 10 mM</td>
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<tr>
<td>SiRNA knockdown of DJ-1 has no effect alone on the formation of autophagic vacuoles.</td>
<td>SiRNA knockdown of DJ-1 induces apoptotic death (25-30%)</td>
<td>DA SH-SY5Y cells transfected with DJ-1 si RNAs and exposed to paraquat 250-500 µM</td>
<td>Gonzalez-Polo et al. 2009</td>
</tr>
<tr>
<td>In the presence of PQ (250 µM), DJ-1 knockdown significantly inhibited cytoplasmic accumulation of autophagic vacuoles, with an additive increase in apoptotic chromatin condensation</td>
<td>The combination of DJ-1 si RNA and Paraquat induces additive apoptotic death (more significant in the range 250-500 µM PQ) and caspase-3 activation.</td>
<td>DA SH-SY5Y cells transfected with DJ-1 si RNAs exposed to paraquat 250-500 µM</td>
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<tr>
<td>Uncertainties or inconsistencies</td>
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<tr>
<td>• The ability of paraquat to induce loss of DA neurons in SN in vivo is sometime equivocal. Loss</td>
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<td>of 60% of DA neurons in SN and 90% of their striatal terminals are reported (Brooks et al. 1999)</td>
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<td>following repeated treatment with paraquat but less significant evidence, or no evidence, has</td>
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<td>been reported in later studies (McCormack et al. 2002; Thiruchelvam et al. 2000). No effect of</td>
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<tr>
<td>paraquat on dopaminergic neurons has been reported by some authors (Widdowson et al. 1996;</td>
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<tr>
<td>Breckenridge et al., 2013; Minnema et al. 2014;). However, the applied dose, the treatment</td>
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<tr>
<td>scheduling, the route of administration as well as the animal age, species and strain (Tieu,</td>
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<td>2016; Jiao et al. 2012; Yin et al. 201; McCormack et al. 2002; Thiruchelvam et al. 2003) are</td>
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<td>all important factor to be considered in the evaluation of the study's outcome.</td>
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</tbody>
</table>
Dopaminergic neurons in SN and VTA seem to have a different susceptibility to the damage induced by paraquat (McCormack et al. 2006). However, whether impaired proteostasis and protein aggregation would cause the selective death of DA neurons in the SN still remain an uncertainties.

Selectivity of paraquat-induced DA neuronal cell death still remains uncertain. Similar effect on other brain region i.e. frontal cortex and hippocampus) are also affected (Mitra et al. 2011).

The vulnerability of the dopaminergic pathway still remains circumstantial. Paraquat has been proposed to pass the blood-brain-barrier by mediation of neutral amino acid transportation (Shimizu et al. 2001; McCormack et al. 2003). Accumulation of paraquat in the brain is reported to be age dependent, possibly indicating a role for the blood-brain-barrier permeability (Corasaniti et al 1991); however, paraquat is not a substrate for dopamine transporter (Richardson et al. 2005), and hence how the toxicant enters into dopaminergic neurons still remain uncertain.
References


Uversky VN. 2004. Neurotoxicant-induced animal models of Parkinson’s disease: understanding the role of otenone, maneb and paraquat in neurodegeneration. celltissueRes. 318;225-41

Widdowsen PS., Farnworth MJ., Upton R., Simpson MG. 1996. No changes in behaviour, nigro-striatal system neurochemistry or neuronal cell death following toxic multiple oral paraquat administration to rats. Human and Experimental Toxicology., 15, 583-91.


4th KER: Neuroinflammation leads to degeneration of the dopaminergic neurons of nigrostriatal pathway

See AOP1 (p. 172)

and

5th KER: Degeneration of dopaminergic neurons of the nigrostriatal pathway directly leads to neuroinflammation

See AOP1 (p. 180).

6th KER: Degeneration of DA neurons of nigrostriatal pathway leads to motor symptoms of PD

How this key event relationship works

See AOP 1 (pag193)

Weight of evidence for the KER

Biological plausibility

See AOP 1 (p.193)

Empirical support for linkage

- Paraquat treatment (10 mg/kg twice a week for 4 weeks) of young adult Sprague-Dawley rats (2 months old) induced a significant loss of nigral dopaminergic neurons (by Nissl staining and TH immunostaining) in SNpc of 15% and a mixed pattern of motor impairments (postural deficit, decrease in speed and mobility), which may have been related to early effects of nigral dopaminergic neuronal loss (Cicchetti et al, 2005).

- Adult C57 BL/6 mice treated i.p. with paraquat (5 and 10 mg/kg) showed a dose-dependent decrease in substantia nigra dopaminergic neurons (36% and 61%, respectively, assessed by Fluoro-gold prelabeling method), a decline in striatal dopamine nerve terminal density (87% and 94%, respectively, assessed by TH immunoreactivity) and neurobehavioural syndrome characterized by reduced ambulatory (locomotor) activity (Brooks et al, 1999).

- Paraquat treatment (i.p.10 mg/kg twice a week for 4 weeks) of male Swiss Albino mice, 22-14 weeks old, induced progressive motor dysfunction with severe postural instability and gait impairment. A concomitant decrease in the expression levels of TH in SN (approximately 60%), FC (frontal cortex) and hippocampus and a decrease (approx. 40%) in TH+ and FOX3+ neurons in SN were observed (stereological evaluation). As part of the toxicological evaluation of the most suitable sub-lethal dose, mice were also treated at 5 mg/kg by i.p. twice a week for 4 weeks. In addition, a decrease in DOPA-decarboxylase was observed in the SN and FC. The only endpoint measured (in addition to the general toxicity endpoints) was the neuronal count in the SN. A statistical significant decrease (approximately 15%) in TH+ and FOX3+ neurons was observed (Mitra et al, 2011).

- Male C57BL/6 mice, 6 weeks, 5 months and 18 months old, were i.p. treated with paraquat at 10 mg/kg twice a week for 3 weeks (6 injections in total). Age-dependent reduction in locomotor activity and motor coordination was observed. The 18-month old mice were the most severely affected and failed to recover 24h post treatment. Progressive reduction in dopamine metabolites and turnover were greatest in the 18-month old group of animals. Increased in striatal TH activity was observed in the 6-week-old and 5-month-old animals but not in 18-month-old mice. The number of nigrostriatal dopaminergic neurons was reduced in all age group animals but these losses, along with the decreases in striatal TH protein levels,
were progressive in 18-month-old paraquat groups between 2 weeks and 3 months post-exposure. (Thiruchelvam et al, 2003).

- Intracerebral injection of 1-5 µg paraquat in male Wistar rats (3 months old) for 16 weeks caused dose-dependent depletion DOPA in the ipsilateral striatum starting 2 weeks after treatment (long-lasting and irreversible) up to 91.5% at 3 µg paraquat. Paraquat induced marked loss of Nissl substances and severe loss of neurons at 3 µg. PQ caused dose-dependent rotational behavior in rats contralateral to the lesion side in response to apomorphine administration (inducing circling behavior) (Liou et al, 1996).

- Male Wistar rats were injected with 10 mg/kg paraquat i.p. for 4-24 weeks. Paraquat induced reduction in TH+ neurons of the SN (17% at 4-week mainly in the rostral region, up to 37% at 24 weeks expanding to the whole length of SN; evaluated by stereology). DOPA levels increased in the caudate-putamen (4-8 weeks) then returned to control values and dropped (25-30%) after 24 weeks. This seems to result from degeneration of DOPA neurons. TH level (Western blot) decreased in the caudate-putamen after 24 weeks (55%) but this effect was not reflected by the loss in TH-ir neurons (being already dropped in the rostral part of SN after 4 weeks) (Ossowska et al, 2005). Clinical signs were not recorded in this study; however the study design was considered of relevance for the evaluation of the progression of the fibing associated with neuronal loss.

- Parauquat treatment (i.p. injection 10 mg/kg bw every five days over 20 days) of Long Evans Hooded rats induced progressive (TH positive neurons stereology counted) loss in dopaminergic neurons up to 47% (end of week 8 post PQ exposure) and deficiency in behavioural motor function (horizontal beam walking test) (after 4 and 8 weeks). Ubisol-Q10 (6 mg/bw) administration after completion of paraquat injections (when the degenerative process had already began (20% TH positive neurons lost)) was effective in blocking the progression of neurodegeneration and improved motor skills. To maintain this neuroprotection, continuous Ubisol-Q10 supplementation was required. Discontinuation of treatment resulted in neuronal death, suggesting that the presence of the antioxidant was essential for blocking the pathway (Muthukumaran et al, 2014).

- In Fernagut (2007) experiment, male mice over-expressing human α-syn under the Thy 1 promoter (Thy 1-aSYN) and WT were i.p. injected PQ 10 mg/kg once a week for 3 weeks. Despite degeneration of dopaminergic neurons (densitometric measurement and stereological analysis for counting TH+ neurons) in both Thy 1-aSYN mice and WT PQ-treated mice, behavioural impaired sensimotor performance was observed in non-treated Thy 1-aSYN mice only, remaining unchanged after PQ administration. The sensimotor abnormalities in Thy 1-aSYN were observed in a previous work (Fleming et al., 2004) and the lack of behavioural deficits after PQ administration was commented by the author as not surprising in the view of small magnitude neuronal loss TH-positive terminals in striatum (25%).

**Quantifiable understanding**

**Table 15:** Quantitative understanding of the KER.

<table>
<thead>
<tr>
<th>DA neurons dedegeneration</th>
<th>Parkinsonian motor symptoms</th>
<th>Treatment</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>15% DA neuronal loss (Nissl staining and TH immunostaining) in SNpc</td>
<td>Mixed pattern of motor impairment observed for testing posture and speed but not for mobility (approx. 3 times the control, as average for total score from Fig 5)</td>
<td>Young adult Sprague-Dawley rats (2 months old) i.p. injected with PQ 10 mg/kg, twice a week for 4 weeks</td>
<td>Cicchetti et al, 2005</td>
</tr>
<tr>
<td>Decrease in SN dopaminergic neurons of 36% and 61%, respectively</td>
<td>Neurobehavioural syndrome characterized by reduced ambulatory</td>
<td>Adult C57 BL/6J mice i.p. injected with PQ 5 and 10 mg/kg, 3 doses</td>
<td>Brooks et al, 1999</td>
</tr>
<tr>
<td>(assessed by Fluoro-gold prelabeling method). Decline in striatal dopamine nerve terminal density of 87% and 94%, respectively (assessed by TH immunoreactivity)</td>
<td>(locomotor) activity 48h after final treatment (during the course of 60 min experimental session) observed at both doses (reduction approx. 45% after 60 min. Fig 5A)</td>
<td>separated by 1 week each</td>
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<tr>
<td>Differential immunolocalisation and decreased expression levels of TH in SN (60%), FC (50%) and hippocampus (30%) (only measured at 10 mg/kg) Decrease in TH+ and FOX3+ neurons in SN (stereological count) of approximately 40% at 10 mg/kg and of approximately 10-15% at 5 mg/kg</td>
<td>Motor dysfunction (only observed at 10 mg/kg) after 2 weeks of treatment (progressive over the next days) with severe postural instability and gait impairment consistent with a unilateral lesion: • Curling test (qualitative asymmetry evaluation): ipsilateral. • Gait impairment: walking footprint pathway (qualitative assessment), stride length of consecutive steps and step frequency</td>
<td>Adult male Swiss Albino mice i.p. treated with 5 and 10 mg/kg PQ twice a week for 4 weeks</td>
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<tr>
<td>Dose-dependant DA depletion in ipsilateral striatum 2 weeks after treatment. 26. 7, 60.3 and 91.5 % at 1,2 and 3µg PQ respectively. The effect lasted up to 16 wks Marked loss of Nissl substances and severe loss of neurons at 3 µg PQ (2 weeks after injection). The effect was considered moderate at 2 µg PQ (2 weeks after injection).</td>
<td>Circling behavior (direction of the lesioned side) due to the imbalance of dopaminergic activity in striata (unilateral lesion) at 3 µg PQ. Dose-dependent rotational behavior in rats contralateral to the lesion side in response to apomorphine s.c. administration 0.5 mg/kg (inducing circling behavior) at 3 µg PQ (2 weeks after injection)</td>
<td>Intracerebral (unilateral intranigral) injection of 1, 2 and 3 µg PQ in male Wistar rats for 16 weeks</td>
<td></td>
</tr>
<tr>
<td>Progressive TH positive neurons (stereology count) loss up to 47% at the end of week 8 post PQ exposure.</td>
<td>Deficiency in behavioural motor function (horizontal beam walking test) after 4 and 8 weeks.</td>
<td>Long Evans Hooded rats i.p. injected PQ 10 mg/kg bw, every five days over 20 days</td>
<td></td>
</tr>
<tr>
<td>Nigrostriatal dopaminergic neurons reduced in all age</td>
<td>Reduction in locomotor activity and motor</td>
<td>Muthukumaran et al., 2014</td>
<td></td>
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<tr>
<td></td>
<td></td>
<td>Thiruchelvam et al, 2003</td>
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</table>
groups but progressive in 18-month-old PQ groups between 2 weeks and 3 months post-exposure. coordination, age dependent with 18-month old mice most affected and failing to recover 24h post treatment 18 months old) i.p. treated with PQ 10 mg/kg twice a week for 3 weeks (6 injections in total).

**Uncertainties or inconsistencies**

- Exposure to paraquat may decrease the number of nigral neurons without triggering motor impairment (Fernagut 2007). This can be consequent to the low level of DA reduction or limited neuronal loss observed following the treatment.

- The impact of paraquat upon the striatum appears to be somewhat less pronounced than the effects of the pesticide upon SNC DA neuronal soma (Mangano et al., 2012). As well, some authors have failed to find changes in striatal DA levels or behavioral impairment, even in the presence of loss of DA soma (Thiruchelvam et al., 2003). It is conceivable that compensatory/buffer downstream processes provoked by soma loss, variations in experimental design (e.g., route of administration, dosing regimen, sacrifice interval, striatal subregions tested, age of mice) can possibly contribute to some of the inconsistency observed across studies (Rojo et al., 2007; Rappold et al., 2010, Prasad et al., 2009; Kang et al., 2010).

- The effects on nigral dopaminergic neurons appear to be specific (Tieu et al, 2011). However, damage in dopaminergic cell bodies and terminal has not been consistently observed (Thiruchelvam et al., 2000b; Cicchetti et al., 2005). In addition, even in studies in which a loss of nigral dopaminergic neurons is detected, PQ does not have an effect on striatal dopamine level (Thiruchelvam et al., 2000b; McCormack et al., 2002). This lack of dopamine reduction might be related to the compensatory up-regulation of tyrosine hydroxylase activity in the striatum after PQ injection (Thiruchelvam et al. 2000b; McCormack et al. 2002; Ossowska et al. 2005, Tieu 2011)

- The repeat dose administration of 10mg/kg i.p. is likely representing the maximum tolerated dose of the chemical stressor. The observed movement disorders can, at least in part, come from systemic illness and the contribution of systemic pathological changes to the observed movement disorders cannot ruled out (Cicchetti et al. 2005).

**References**


weeks does not result in a loss of dopaminergic neurons in the substantia nigra of C57BL/6J mice.

Regulatory Toxicology and Pharmacology. 68(2):250–258.


Overall assessment of the AOP

1. Concordance in dose-response and incidence (Table 1)

Chemical toxicity mediated by redox cycling is based on acceptance of an electron by a chemical from a reductant, formation of a radical, and transfer of an electron to molecular oxygen. The process is leading to the generation of superoxide and mitochondria are one of the presumed site where the chemical is initially reduced within the cell to form superoxide. This is the chemical based mechanism of action of the herbicide paraquat (PQ), which is therefore considered a suitable chemical tool/stressor for exploring the link between the MIE and the AO. In animal models, PQ susceptibility is known to act synergistically with microglia leading to its activation (Purisai et al., 2007; Mitra et al. 2011). Microglia through plasma-membrane NADPH-oxidase may also activate the extracellular redox cycling of PQ favouring its transport within dopaminergic neurons (Rappold et al. 2011). The kinetic and metabolism of PQ is complex and the amount of PQ entering and accumulating into the brain is dependent on dose, route of administration, expression of transporters, animal age and strain.

Multiple genetic factors are also involved in host susceptibility which is likely to represent an important source of variability (Tieu 2011; Corasaniti et al. 1991, Jiao et al.2012). These elements are the possible/likely reason of lack of reproducibility of apical endpoints as observed in some studies conducted with this stressor (Breckenridge et al. 2013). Furthermore, because of these complexities, it is nearly impossible to extrapolate the concentration used in-vitro (mainly in purified neuronal cultures) with the doses applied in-vivo. The in-vivo dose response curve for PQ was limited by the general toxicity effects induced by the stressor. For practical convenience and better understanding of the dose and temporal concordance, in-vitro and in-vivo studies were kept separated in this overall assessment.

In-vivo, the commonly used single dose of 10 mg/kg administered i.p., corresponds to a plasma concentration of approximately 780 μM (Prasad et al. 2007). The same dose leads to a brain concentration of 0.78 to 5.4 μM after 3 or 18 doses. A single s.c. administration of 10 mg/kg leads to 3.88 ± 0.79 μM serum concentration after 3 hours reaching 0.36 ± 0.09 μM in the extracellular space of the striatum (Shimizu et al. 2001). At this dose level (10 mg/kg i.p.), ambiguous results in terms of neuronal loss and occurrence of parkinsonian motor symptoms are reported (McCormack et al., 2002; Prasad et al 2007 and 2009; Breckenridge et al. 2013; Thiruchelvam et al, 2000b). In a conservative approach, the range of brain concentration observed following single and/or repeated administration of 10 mg/kg i.p. or sc. of PQ can be considered only indicative and will be used in this AOP to define a possible probabilistic threshold of activation of the MIE leading to the AO. With the limited number of doses known from in-vivo studies, an intra (in the same) and inter (between) KE dose relationship can be observed. A dose response relationship in the increase in activity of ROS scavenging enzymes can be observed between 5 and 40 mg/kg i.p. of paraquat in mice; however, the link with the neuronal cell loss and the AO can only be seen up to 10 mg/kg i.p. of PQ due to the marked general toxicity observed above this dose (Mitra et al. 2011). At the dose of 10 mg/kg i.p. following multiple dose administration, all the KEs are observable, however, when present, the AO can only be observed at 10 mg/kg i.p and not at 5 mg/kg i.p. (Mitra et al. 2011). Not enough details were reported in the
consulted studies to establish incidence concordance. It is however evident that the frequency of the reported apical effect i.e. parkinsonian motor symptoms was less than for the key events. In this AOP, neuroinflammation was considered to have a direct effect on paraquat activation and on loss of DA neurons (Rappold et al. 2011, Purisai et al. 2007). However, in addition to neurodegenerative consequences, neuroinflammation can have also protective effects. Therefore, due to this complexity this key events was not included in table 1 and2.

In vitro, an intra KEs concentration relationship is evident, with some evidences of inter KEs concentration-response concordance. However, when multiple time of sampling are applied to the experimental design, the inter KEs concentration response concordance is stronger. In-vitro, a strong response concordance between ROS generation and cell death is evident (Fig 31). Overall, the concordance in dose-response and incidence was considered moderate.

**Paraquat: dose-response 24h - *vitro***

Fig.31 PQ-induced ROS and Cell death (% over controls) at 24h in neuronal cells. Points in the figure derive from the listed papers, targeted by the number associated to each symbol (1-de Oliveira 2016, 2-González-Polo 2007, 3-Lopert 2012, 4-Rodriguez-Rocha 2013, 5- Huang 2012, 6-Ding 2001, 7-Yang and Tiffany Castiglioni 2007) Data refers to different neuronal cell lines and primary cultures, and to different methods of detection. As such, single results have been calculated over their control to allow comparison between different studies.

### 2. Temporal concordance among the MIE, KEs and AO

There is a strong agreement on the sequence of pathological events linking the MIE to the adverse outcome (Fujita, et al.2014). The temporal concordance is strong when considering the chronicity and progressive nature of the pathology of parkinsonian disorders. Temporal concordance among the KEs, KE2, 3 and AO can be observed in the experimental models of PD using the chemical stressors rotenone and MPTP (Betarbet 2000 and 2006; Sherer et al. 2003, Fornai et al. 2005) which are sharing the same KEs with this AOP but are caused by a different MIE. With the chemical stressor MPTP, to trigger the KE3 (i.e. degeneration of DA neurons in SNpc with presence of intracytoplasmatic Lewy-like bodies) and motor deficits (AO), proteostasis needs to be disturbed for a minimum period of time (Fornai et al. 2005) and this is similarly expected with chemicals inducing redox cycling like PQ (Ossowska et al. 2005). In vivo, with the chemical stressor PQ, evidence of temporal concordance is limited by the study design using single time-point descriptive assessment. In vitro, evidence of temporal concordance is limited by the fact that 24 and 48 h were the most investigated time points. Nevertheless, those papers taking into account shorter time points show that a good temporal concordance exist between MIE (4h), KE2 (6h) and KE3 (12-24h) (Cantu 2011, González-Polo 2007;
Ding 2001; de Oliveira 2016). Based on the established knowledge on chronicity and progression of parkinsonian disorders, the temporal concordance is considered strong for this AOP up to the KE 3 (degeneration of DA neurons of nigrostriatal pathway). The occurrence of the AO outcome is strongly linked to the amount of DA in the striatum and to the loss of DA neurons in the SNpc. In PD or following treatment with the chemical stressor/s the key events are observed in the proposed order in this AOPr.
### Table 16: Response-Response and Temporality Concordance Table

<table>
<thead>
<tr>
<th>Concentration at the target site (Paraquat)</th>
<th>MIE</th>
<th>KE1 Mitochondrial ROS production and dysfunction</th>
<th>KE2 Impaired proteostasis</th>
<th>KE3 Degeneration of DA neurons of nigrostriatal pathway</th>
<th>AO Parkinsonian motor symptoms</th>
</tr>
</thead>
<tbody>
<tr>
<td>10 µM [1, 10, 12]</td>
<td>No data</td>
<td>± (at 24 hr)</td>
<td>+ (ALP 6h, time-dependent at 24 hr)</td>
<td>± (at 24 hr)</td>
<td></td>
</tr>
<tr>
<td>20 to 50 µM [1, 2, 10, 12]</td>
<td>No data</td>
<td>+ (at 6 and 24 hr; n.e. 3h)</td>
<td>+ (UPS at 6 hr)</td>
<td>+ (at 24 hr)</td>
<td></td>
</tr>
<tr>
<td>50 to 200 µM [1, 2, 8, 10, 13]</td>
<td>++ (at 24 and 48 hrs)</td>
<td>++ (at 6 and 24 hr)</td>
<td>++ (UPS at 6 hr)</td>
<td>++ (at 24 and 48 hr)</td>
<td></td>
</tr>
<tr>
<td>200 µM to 1 mM [3, 4, 7, 8, 9, 10, 12]</td>
<td>+++ (at 4, 6, 24 and 48 hr)</td>
<td>++ (at 4 and 6 hr)</td>
<td>+++ (UPS at 48 hr)</td>
<td>+ (ALP at 24 hr)</td>
<td>+++++ (at 18, 24 and 48 hr)</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Dose, in-vivo studies</th>
<th>MIE</th>
<th>KE1 Mitochondrial dysfunction (ROS production)</th>
<th>KE2 Impaired proteostasis</th>
<th>KE3 Degeneration of DA neurons of nigrostriatal pathway</th>
<th>AO Parkinsonian motor symptoms</th>
</tr>
</thead>
<tbody>
<tr>
<td>5 mg/kg i.p twice a week for 4 weeks [15] or once a week for 3 weeks [16]</td>
<td>+ (4 wks)</td>
<td>Increase activity in ROS-scavenging enzymes + (4 wk)</td>
<td>No data</td>
<td>Decrease in number of TH+ in SN + (at 4 wk)</td>
<td>No locomotor deficit</td>
</tr>
<tr>
<td>10 mg/kg ip twice a week for 1, 2, 3, 4, 6, 9 weeks [6, 13, 14, 15]</td>
<td>++ (4 wks)</td>
<td>Increased lipid peroxidation - (1 wk) +/+++ (2 wk) +++ (6-9 wk)</td>
<td>Impaired proteostasis and autophagy ++</td>
<td>Decrease in number of TH+ in SN - (1 wk) ++ (at 2 to 4 wk)</td>
<td>Locomotor deficit ±</td>
</tr>
<tr>
<td>10 mg/kg ip once a week for 1, 2 weeks (14) or 3 weeks (16); twice a week for 4, 6, 9 weeks (6, 13, 15)</td>
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</table>


+, ++, +++ are intended only to demonstrate intra and inter KEs relationships.
3. Strength, reproducibility of the experimental evidence, and specificity of association of AO and MIE

There is a strong agreement that ROS production and mitochondrial dysfunction can lead to neurodegeneration and motor symptoms of parkinsonian disorders and familial PD genes are also implicated in ROS production by mitochondria (Fujita et al. 2014, Yao et al. 2011, Gandhi et al. 2009). PQ is a well-known substance with a toxicity primarily mediated by redox cycling (Tieu 2011; Day et al. 1999). With PQ, ROS production and oxidative stress, impaired proteostasis, and loss of nigral dopaminergic neurons are reported (Brooks et al. 1999, McCormack et al. 2002, Mitra et al. 2011, Su et al. 2015). Some uncertainties on the initial mitochondrial involvement in triggering PQ redox-cycle in vivo exists due to the prolonged (consistent with a generalized oxidative stress) and repeated exposure and the use of general indicators of oxidative stress like lipid and protein oxidation. Non-reproducibility of DA neuronal loss is reported by some authors (Breckenridge et al. 2013; Thiruchelvam et al. 2000b; Cicchetti et al. 2005, Minnema et al. 2014) and in studies with loss of dopaminergic neurons, PQ was not showing an effect on striatal dopamine levels (Thiruchelvam et al. 2000b, McCormack et al. 2002). Although this can be due to the activation of compensatory effects or compensatory up-regulation of TH activity in the striatum following PQ treatment (Thiruchelvam et al. 2000b, McCormack et al. 2002, Ossowska et al. 2005), the role and influence of the animal species, strain, age, route of administration, dose scheduling and susceptibility of neuronal population to the noxa on the outcome of the studies cannot be completely ruled out. However, when considering the amount of positive studies vs. negative studies, there is a clear prevalence of positive studies supporting the KE 3(degeneration of DA neuronal cells of the nigrostriatal pathway). The occurrence of parkinsonian motor symptoms was not consistently reported for the chemical stressor PQ. Evidence on the occurrence of the AO can however be observed with PQ following unilateral intranigral administration where loss of neuronal cells was marked (ca. 90%). For most of the studies conducted with the tool chemical PQ administered by i.p. the amount of DA neuronal loss was relatively limited (e.g. 20-30%) and AO i.e. parkinsonian motor symptoms, was not consistently reported. These observations are in line with the human evidence that parkinsonian motor symptoms are only evident in PD when striatal DA drops approximately 80% (corresponding to a 60% DA neuronal cells loss (Jellinger et al. 2009). Considering the relevance of ROS production and oxidative damage in Parkinson’s models, it is expected that the specificity of this AOP would be high. However, with the use of PQ as a unique chemical stressor supporting the empirical evidence, judging specificity was not possible. Overall, the strength linking the MIE to the AO was considered high for this AOP and using PQ as a chemical tool, the reproducibility of experimental evidence and the specificity of the association was considered moderate.

4. Weight of Evidence (WoE)

4.1 Biological plausibility, analogy between chemical stressors, and species consistency of the experimental evidence

ROS generation and deregulation of ROS management by dysfunctional mitochondria is known to be a crucial event in neurodegeneration in general and for dopaminergic neurons in SNpc in particular when considering the unique susceptibility of these neurons (Fujita et al. 2014). Familial forms of PD include genes (i.e. PINK1 and DJ1) that are implicated in ROS management by mitochondria resulting in mitochondrial DNA damage and inflammation as a downstream effect (Fujita et al. 2014, Gandhi et al. 2009, Yao et al. 2011). The biological plausibility for the KEs relationship linking the MIE to the AO is strong based on the existing knowledge of PD pathogenesis. As PQ is the only tool compound so far analysed and comprehensively studied, analogy is considered moderate as the KE relationship is only plausible based on the supporting analogy with PD, but a scientific understanding on the relationship between a chemically induced redox cycler and parkinsonian motor deficits is not completely established. ROS generation is mechanistically recognized as a cause of PD. However, epidemiological studies linking exposure to the tool compound PQ and PD are not definitive due to the multiple intrinsic limitations of the studies. Mouse and rat are the most frequently used animal models to support this AOP using the tool compound PQ. The same pattern of effects has been observed in a different test species i.e. drosophila. Overall the consistency of this AOP was considered moderate to high.
<table>
<thead>
<tr>
<th>1 Support for Biological Plausibility of KERs</th>
<th>Defining Question</th>
<th>High (Strong)</th>
<th>Moderate</th>
<th>Low(Weak)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Is there a mechanistic (i.e. structural or functional) relationship between KE up and KE down consistent with established biological knowledge?</td>
<td>Extensive understanding of the KER based on extensive previous documentation and broad acceptance</td>
<td>The KER is plausible based on analogy to accepted biological relationships, but scientific understanding is not completely established</td>
<td>There is empirical support for a statistical association between KEs but the structural or functional relationship between them is not understood</td>
<td></td>
</tr>
</tbody>
</table>

**MIE to KE1**

**KE1 to KE2**

**KE2 to KE3**

**KE1 to KE2**

**Mitochondrial dysfunction (ROS production) to impaired proteostasis**

Moderate

The weight of evidence supporting the biological plausibility behind the relationship between mitochondrial dysfunction and impaired proteostasis, including the impaired function of UPS and ALP that results in decreased protein degradation and increase protein aggregation is well documented but not fully understood. It is well established that the two main mechanisms that normally remove abnormal proteins (UPS and ALP) rely on physiological mitochondrial function. The role of oxidative stress, due to mitochondrial dysfunction, burdens the proteostasis with oxidized proteins and impairs the chaperone and the degradation systems. This leads to a vicious circle of oxidative stress inducing further mitochondrial impairment (Powers et al., 2009; Zaltieri et al., 2015; McNaught and Jenner, 2001, Moore et al., 2005). Therefore, the interaction of mitochondrial dysfunction and UPS / ALP deregulation plays a pivotal role in the pathogenesis of PD (Dagda et al., 2013; Pan et al., 2008; Fornai et al., 2005; Sherer et al., 2002).

**KE2 to KE3**

**Impaired proteostasis leads to degeneration of DA neurons of the nigrostriatal pathway**

Moderate

It is well known that impaired proteostasis refers to misfolded and aggregated proteins including alpha-synuclein, autophagy, deregulated axonal transport of mitochondria and impaired trafficking of cellular organelles. Evidences are linked to PD and experimental PD models as well as from genetic studies (McNaught et al. 2001, 2003, 2004; Matsuda and Tanaka, 2010; Tieu et al. 2014; Rappold et al. 2014). Strong evidence for degeneration of the nigrostriatal pathway comes from the experimental manipulations that directly induce the same disturbances of proteostasis as observed in PD patients (e.g. viral mutated alpha-synuclein expression). However, a clear mechanistic proof for the understanding of the exact event triggering cell death is lacking.
The fact that reactive glial cells (microglia and astrocytes) may kill neurons is well accepted. The mechanisms underlying this effect may include the release of cytotoxic signals (e.g. cytokines) or production of ROS and RNS (Chao et al., 1995; Brown and Bal-Price, 2003; Kraft and Harry, 2011; Taetzsch and Block, 2013). However, the studied mediators differ from model to model. The fact that neuronal injury/death can trigger neuroinflammation is supported by evidence in human and experimental models. The evidence that neuroinflammation triggered by neuronal damage can cause neuronal death (vicious circle), is mostly indirect or by analogy (Hirsch and Hunot, 2009; Tansey and Goldberg, 2009; Griffin et al., 1998; McGeer and Mc Geer, 1998; Blasko et al., 2004; Cacquevel et al., 2004; Rubio-Perez and Morillas-Ruiz, 2012; Thundiyil and Lim, 2014; Barbeito et al., 2010).

Degeneration of DA neurons of the nigrostriatal pathway leads to parkinsonian motor symptoms

The mechanistic understanding of the regulatory role of striatal DA in the extrapyramidal motor control system is well established. The loss of DA in the striatum is characteristic of all aetiologies of PD and is not observed in other neurodegenerative diseases (Bernheimer et al. 1973; Reynolds et al. 1986). Characteristic motor symptoms such as bradykinesia, tremor, or rigidity are manifested when more than 80 % of striatal DA is depleted as a consequence of SNpc DA neuronal degeneration (Koller et al. 1992), possibly corresponding to approximately 60% of neuronal cell loss (Jellinger et al. 2009). However, when considering these quantitative thresholds, experimental evidences with the tool chemical paraquat are largely inconsistent with only limited evidence of this KE, indicating that, at least quantitatively, the scientific understanding is not complete.

4.2 Essentiality

Direct essentiality evidence is coming from experiments conducted with antioxidant agents or following manipulation of the biological systems protecting from or regulating ROS production and oxidative stress. Manifestation of motor symptoms differs in rodents and human and for this reason their value should depend upon its relationship to striatal dopaminergic function. Study designed to demonstrate recovery of clinical signs following DA replacement are lacking with the chemical tool paraquat and most of the time the level of DA neuronal loss or drop in DA is not enough to trigger specific behaviour symptoms. Evidence of essentiality is however indirectly provided following unilateral intranigral injection of paraquat or in drosophila models. The overall WoE for the essentiality is strong.

Table 18: Essentiality of the KE; WoE analysis

<table>
<thead>
<tr>
<th>2 Support for Essentiality of KEs</th>
<th>Defining Question</th>
<th>High (Strong)</th>
<th>Moderate</th>
<th>Low(Weak)</th>
</tr>
</thead>
<tbody>
<tr>
<td>MIE</td>
<td>Strong</td>
<td>Direct evidence from specifically designed experimental studies illustrating essentiality for at least one of the important KEs (e.g. stop/reversibility studies, antagonism, knock out models, etc.)</td>
<td>Indirect evidence that sufficient modification of an expected modulating factor attenuates or augments a KE leading to increase in KE down or AO</td>
<td>No or contradictory experimental evidence of the essentiality of any of the KEs</td>
</tr>
</tbody>
</table>

Overexpressing enzymes involved in O₂ dismutation specifically located at the mitochondria prevents neuronal cell...
Redox-cycling (of a chemical) initiated by electrons of the mitochondrial respiratory chain
death in vitro (Rodriguez-Rocha et al., 2013; Filograna et al., 2016). Accordingly, depletion of mitochondrial SOD2 exacerbate PQ-toxicity in Drosophila (Kirby et al., 2002), while mitochondrial enzymes activity is restored and neuronal death in cortex reduced in SOD2 knock out animals treated with antioxidants (Hinerfeld et al., 2004). Mitochondrial aconitase knock down attenuated PQ induced H2O2 production and respiratory capacity deficiency in neuronal cells (Cantu 2011).

**KE1**
Mitochondrial reactive oxygen species (ROS) formation and dysfunction

In vitro, PQ toxicity both in terms of ROS production, mitochondrial dysfunction and neuronal death is rescued by several antioxidants (Peng et al. 2005; Hinerfeld et al., 2004; McCarthy et al., 2004; Chau et al., 2009, 2010; de Oliveira 2016). Most of these drugs, like synthetic superoxide dismutase/catalase mimetics or SOD-fusion proteins also protect against PQ-induced oxidative damage and/or DA neurons degeneration in vivo (Peng et al. 2005; Choi 2006; Hinerfeld et al., 2004) improving motor skills (Muthukumaran et al., 2014; Somayajulu-Nitu et al. 2009). Overexpression of antioxidant enzymes specifically at the mitochondria protects Drosophila and yeast from PQ-toxicity at low doses (Mockett et al., 2003; Tien Nguyen-nhu and Knoops, 2003). On the other hand, depletion of antioxidant systems exacerbates PQ toxicity both in vitro and in vivo (Lopert et al., 2012; Liang et al. 2013; Van Remmen et al 2004). USP30, a deubiquitinase localized to mitochondria, antagonizes mitophagy. Overexpression of USP30 removes ubiquitin in damaged mitochondria and blocks mitophagy. Reducing USP30 activity enhances mitochondrial degradation in neurons. Knockdown of USP30 in dopaminergic neurons protects flies against paraquat toxicity in vivo, ameliorating defects in dopamine levels, motor function and organismal survival in Drosophila (Bingol et al., 2014). More in general, overexpression of Sods in DA neurons counteracts PQ-induced oxidative damage and reduces motor dysfunction in Drosophila (Filograna et al., 2016). Similarly, the use of antioxidants also restores PQ-induced motor activity in Drosophila (Jimenez-Del-Rio et al., 2010).

**KE2**
Impaired proteostasis

Most of the experimental evidence supporting the essentiality is coming from experiments conducted in transgenic animals or studies conducted with the chemical stressor Rotenone and MPTP, known chemical toxins used to mimic PD (Fornai et al., 2005; Dauer et al., 2002; Kirk, 2002; Klein, 2002; Lo Blanco, 2002; Lauwers, 2003; Kirk, 2003).

Exposure to the chemical stressor paraquat of mice with inducible overexpression of familial PD-linked mutant α-syn in dopaminergic neurons of the olfactory bulb exacerbate the increase of soluble and insoluble α-syn expression, accumulation of α-syn at the dendritic terminals, reduction of auto-lysosomal clearance, mitochondrial condensation and damage. None of these effects occurs in PQ-treated mice with suppressed α-syn expression. Loss of DA neurons in the olfactory bulb is evident in PQ-treated mutant mice but not in both PQ-treated mice with suppressed α-syn expression (after doxycycline administration) and untreated mutant mice (Nuber et al., 2014).

In vitro system overexpressing the neuroprotective molecular chaperone human DJ-1, showed more resistance to the proteasome impairment induced by paraquat. Similarly, preservation was observed in the same system following treatment with a known proteasome inhibitors (epoxomycin) (Ding and Keller, 2001). However, although evidence exists to support some essentiality of impaired proteostasis, a single molecular chain of events cannot be established.

**KE3**
Degeneration of DA neurons of nigrostriatal pathway

Clinical and experimental evidences show that the pharmacological replacement of the DA neurofunction by allografting fetal ventral mesencephalic tissues is successfully replacing degenerated DA neurons resulting in the total reversibility of motor deficits in an animal model and a partial effect is observed in human PD patients (Han et al., 2015; Widner et al., 1992; Henderson et al., 1991; López-Lozano et al., 1991; Freed et al., 1990;
Concomitant administration of selective type B monoamine oxidase inhibitor slowed the progression parkinsonian motor symptoms induced by unilateral intranigral injection of paraquat which is expected to induce approximately 90% of neuronal loss. It provides a protective effect on the moderate injury elicited by PQ toxicity. A post-treatment administration of apomorphine, a DA agonist, induced contralateral circling behaviour which correlated well with the decrease of striatal DA (Liou 1996 and 2001). However, for most of the experiments conducted with paraquat, the amount of DA neuronal loss and drop in striatal DA was not consistent or below the threshold for triggering motor symptoms In addition, studies showing an altered behavior resulting from striatal dopaminergic degeneration lacked a DA replacement strategy.

Protection from neuronal cell loss following treatment with 10 mg/kg bw of paraquat was observed in interferon-gamma KO animals or blockade of iNOS, NF-κB or p38 MAPK. In both cases a decrease of microglial reactivity or prevention of microglia activation was observed (Mangano et al. 2012, Yadav et al. 2012). Minocycline or silencing of NADPH oxidase prevented DA neurodegeneration subsequent to the administration of 10 mg/kg bw of paraquat (Purisai et al. 2007). Essentiality of microglial NADPH oxidase for mediating DA neurodegeneration was observed in vitro in neuron-glia co-cultures prepared from NADPH oxidase-deficient mice (Wu et al. 2005). However, inhibition was different in different models and considered as an indirect evidence of essentiality.

4.3 Empirical support

The empirical support provides evidence that the KE up is linked to KE down. With PQ, as the only available chemical tool, the strength of this relationship is limited by the fact that the large majority of studies are conducted at fixed doses and single time-point descriptive assessment. This affected the dose response and incidence concordance analysis and the overall concordance for empirical support was considered moderate. The empirical support in-vivo is mainly provided by studies conducted with PQ in rodents species and drosophila aimed to model PD. In vitro the concentration-response concordance was more evident.
<table>
<thead>
<tr>
<th>Pathway</th>
<th>Grade</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>MIE to KE1</strong></td>
<td>Moderate</td>
<td>With the chemical tool paraquat, studies are mainly conducted at fixed doses and dose relationships studies are very limited for the O₂ production, which is relevant for the intra MIE dose relationship (De Oliveira et al., 2016; Rodriguez-Rocha et al., 2013; Cantu et al., 2011; Huang et al., 2012; Dranka et al., 2012, Mitra et al. 2011). However, intra KE1 dose relationship is observable for ROS production/lipid peroxidation using the same stressor compound (de Oliveira et al., 2016; Lopert et al., 2012; McCormack et al., 2005, Mitra et al. 2011). In-vitro, high concentrations of PQ showing activation of the MIE are showing the most pronounced ROS production indicating that a concordance in dose and response relationship exists between the MIE and KE1 and cell death (Rodriguez-Rocha et al. 2013; de Oliveira et al., 2016; Lopert et al., 2012; Chau et al., 2009). Temporal relationship between MIE and KE1 is indistinguishable due to the fast conversion of O₂ to H₂O₂ and other ROS species (Cohen and Doherty, 1987). However, when considering cell death as the observational end point, a dose response and time concordance exists. PQ (0.1-1 mM) induces O₂° and H₂O₂ production within minutes in isolated mitochondria and mitochondrial brain fraction (Cochemé and Murphy, 2008; Castello et al., 2007), while in cells this process is detectable after 4-6h from the exposure (Rodriguez-Rocha et al., 2013; Cantu et al., 2011, Huang et al, 2012, Dranka et al. 2012). At these time points no death is generally detected. In-vivo, there is limited evidence of intra MIE dose relationship with paraquat and temporal concordance cannot be defined as the experiments are conducted at single time point descriptive assessment (Mitra et al. 2011). However, circumstantial evidences are supported by the knowledge on the chronic and progressive nature of parkinsonian syndromes.</td>
</tr>
<tr>
<td><strong>KE1 to KE2</strong></td>
<td>Low</td>
<td>Mitochondrial dysfunction (ROS production) results in impaired proteostasis Evidence is provided that exposure to PQ and deficiency of DJ-1 might cooperatively induce mitochondrial dysfunction resulting in ATP depletion and contribute to proteasome dysfunction in mouse brain (Yang et al., 2007). Moreover, exacerbation of Paraquat effect on the autophagic degradation pathway is observed in an in vitro system with silenced DJ-1 (González-Polo et al., 2009). In C57BL/6J mice 10 mg/kg i.p. for 1 to 5 doses, increased level of lipid peroxides in ventral midbrain was associated impaired proteostasis (Prasad et al. 2007) Temporal and dose concordance cannot be elaborated from in vivo studies as they are conducted at the same dose and observational time-point. However, in vitro studies are indicative of a temporal and concentration concordance, evidencing concentration-and/or time-dependent effects on mitochondrial and proteasome functions (Ding and Keller, 2001; Yang and Tiffany-Castiglioni, 2007).</td>
</tr>
<tr>
<td><strong>KE2 to KE3</strong></td>
<td>Moderate</td>
<td>Impaired proteostasis leads to degeneration of DA neurons of the nigrostriatal pathway The empirical support linking impaired proteostasis with degeneration of DA neurons of the nigrostriatal pathway comes from post-mortem human evidences in PD patients supporting a causative link between the two key events. With paraquat, a response concordance was observed in multiple in vivo studies (Manning-Bog 2003, Fearnatt 2007, Mitra 2011).Temporal and dose concordance cannot be elaborated from these studies as they are conducted at the same dose and observational time-point. Some inconsistencies were observed, ie. partial effect on proteasomal inhibition which is likely due to compensatory effects and/or lower toxicity of PQ when compared to other chemical stressor (e.g. rotenone, MPTP). In vivo studies with Paraquat are showing a more relevant effect on the ALP and α-synuclein overexpression with a less evident effect on proteasome inhibition. A dose and temporal concordance was more consistently observed in in vitro studies (Chinta 2010; Gonzalez-Polo 2009).</td>
</tr>
<tr>
<td><strong>KE3 &lt;= KE4</strong></td>
<td>Moderate</td>
<td>Neuroinflammation Multiple in vivo and in vitro experiments support the link between neuroinflammation and degeneration of DA neurons in the nigrostriatal pathway as well as vice versa. The observation of concomitant presence of glial and astrocytic cells and degenerated/degenerating DA neurons is also reported in many studies (Cicchetti et al. 2005,</td>
</tr>
</tbody>
</table>
Mitra et al. 2011, Purisai et al. 2007, Mangano et al. 2012, Wu et al. 2005). A similar relationship was observed with compounds like rotenone and MPTP.

| KE3 to AO Degeneration of DA neurons of nigrostriatal pathway leads to parkinsonian motor symptoms | Low | PQ is reported to induce motor deficits and loss of nigral dopaminergic neurons in a dose- (Brooks et al., 1999) and age (Thiruchelvam et al., 2003) dependent manner. The concomitant observation of dopaminergic neuronal loss and parkinsonian motor symptoms has been confirmed by other authors (Cicchetti et al. 2005, Prasad et al. 2009, Mitra et al. 2011). However, with similar doses and experimental design a number of inconsistencies or lack of reproducibility were noted and described in the uncertainties. In human (and animal models using rotenone and MPTP), motor symptoms are expected to be clinically visible when striatal dopamine levels drop of approximately 80%, corresponding to a DA neuronal loss of approximately 60% (Jellinger et al. 2009, Lloyd et al. 1975; Hornykiewicz et al. 1986; Bernheimer et al. 1973). This threshold of pathological changes was only achieved when paraquat was administered directly in the SN and the link between neuronal loss and clinical symptoms was empirically consolidated by the following treatment with apomorphine or the concomitant treatment with the MAOB inhibitors (Liou 1996 and 2001). When different routes of administration were applied, neuronal loss was below this pathological threshold, not consistently related to drop in striatal DA and motor symptoms were not observed or not clearly separated from clinical signs of general toxicity. |
5. Uncertainties and Inconsistencies

- No direct evidence exists in the literature of PQ-induced $O_2^\cdot$ production in vivo. The involvement of $O_2^\cdot$ production is deduced by the efficacy of superoxide dismutase analogues to prevent/reduce PQ neurotoxicity.
- Besides mitochondria, cellular NADPH-oxidases (Cristóvão et al., 2012; Rappold et al., 2011) also contribute to PQ-induced ROS production. Furthermore, *in vitro* data suggest that for time points of exposure longer than 48h oxidative stress occurs both at mitochondria and cytosol. This makes it difficult to discriminate the source of PQ-induced ROS and the early involvement of mitochondria *in vivo* due to the extensive treatments and to the indirect detection of oxidative stress mainly by mean of lipid peroxidation and/or protein oxidation.
- Mitochondrial involvement is suggested by *ex-vivo* studies (Czerniczyniec et al., 2013; 2015).
- The exact molecular link from mitochondrial dysfunction to disturbed proteostasis is still unclear (Malkus et al 2009; Zaltieri et al. 2015). Furthermore, whether impaired proteostasis and protein aggregation would cause the selective death of DA neurons in the SN still remains an uncertainty.
- The role of α-synuclein in neuronal degeneration is still unclear as well as the mechanisms leading to its aggregation.
- Priority of the pattern leading to cell death could depend on concentration, time of exposure and species/strain sensitivity; these factors have to be taken into consideration for the interpretation of the study’s result and extrapolation of potential low-dose chronic effect as this AOP refers to chronic exposure.
- The model of striatal DA loss and its influence on motor output ganglia does not allow to explain specific motor abnormalities observed in PD (e.g. resting tremor vs bradykinesia) (Obeso et al. 2000). Other neurotransmitters (ACh) may play additional roles in other brain areas like the olfactory bulb. Transfer to animal models of PD symptoms also represents an uncertainty.
- The role of neuronal plasticity in intoxication recovery and resilience is unclear.
- This AOP is a linear sequence of KEs. However, ROS production, mitochondrial dysfunction and impaired proteostasis are influencing each other and these are considered as uncertainties (Malkus et al. 2009).
- When measurement of loss of mitochondrial membrane potential is performed together with cell viability, the former is detected only in dead cells in PQ treated cells (on the contrary for MPTP and Rotenone is detected in alive cells suggesting this event precedes cell death). It is suggested that decrease PQ-induced neuronal death is independent on mitochondrial membrane potential (mmp) decrease. As such overexpression of Mn SOD (in mitochondria) prevents PQ-induced cell death but not mmp decrease (Rodriguez-Rocha 2013).
- The ability of paraxquat to induce loss of DA neurons in SN in vivo is sometime equivocal. Loss of 60% of DA neurons in SN and 90% of their striatal terminals are reported (Brooks et al. 1999) following repeated treatment with paraxquat but less significant evidence, or no evidence, has been reported in later studies (McCormack et al. 2002; Thiruchelvam et al. 2000; Cicchetti et al., 2005). No effect of paraxquat on dopaminergic neurons has been reported by some authors (Widdowson et al. 1996; Breckenridge et al., 2013; Minnema et al. 2014). However, the applied dose, the treatment scheduling, the route of administration as well as the animal age species and strain (Tieu, 2011; Jiao et al. 2012; Yin et al. 2011; McCormack et al. 2002; Thiruchelvam et al. 2003) are all important factor to be considered in the evaluation of the study’s outcome.
- Paraxquat induced neurotoxicity could affect a sub-population of DA neurons. This might explain why, once the maximum effect is reached, no further neuronal death occurs after supplementary exposures (McCormack et al. 2005). Another possibility is the development of defensive mechanisms, which preserve neurons from further toxicity. This hypothesis is consistent with the *in vitro* observation of an increased transcripational activation of redox-
sensitive antioxidant response elements and NF--B, specifically induced from paraquat but not from rotenone and MPTP (Rodriguez-Rocha et al. 2013).

- The impact of paraquat upon the striatum appears to be somewhat less pronounced than the effects upon SN dopaminergic neuronal soma (Mangano et al., 2012). In addition, some authors have failed to find changes in striatal DA levels or behavioral impairment, even in the presence of loss of dopaminergic soma (Thiruchelvam et al., 2003). It is conceivable that compensatory/buffer downstream processes provoked by soma loss or variations in experimental design can possibly contribute to some of the inconsistencies observed across studies (Rojo et al., 2007; Rappold et al., 2011, Prasad et al., 2009; Kang et al., 2010).

- Dopaminergic neurons in SN and VTA seem to have a different susceptibility to the damage induced by paraquat (McCormack et al. 2006).

- Few hypothesis have been put forward to explain the selective vulnerability of the SN pc dopaminergic pathway, although a defined molecular mechanism remains elusive. Elevated iron content in this region, that increase sensitivity to redox-damage catalyzing the generation of ROS (Liddell et al., 2013) and regional distribution of transporters able to uptake PQ (i.e. DAT and Oct3) in combination with a high microglia population in the nigra (Rappold et al., 2011) have been evoked.

- The vulnerability of the dopaminergic pathway still remains circumstantial. Paraquat has been proposed to pass the blood-brain-barrier by mediation of neutral amino acid transportation (Shimizu et al. 2001; McCormack and DiMonte 2003). Accumulation of paraquat in the brain is reported to be age dependent, possibly indicating a role for the blood-brain-barrier permeability (Corasaniti et al 1991); Di=cation paraquat has been reported not to be a substrate for dopamine transporter (Richardson et al. 2005). Nevertheless, Rappold et al. (2011), demonstrated that radical paraquat is transported by DAT and hence how the toxicant enters into dopaminergic neurons is still unclear. One possibility is extracellular paraquat reduction by membrane-bound NADPH oxidase with the formed paraquat monocation radical entering DA neurons by neuronal DAT (Rappold et al. 2011).

- Exposure to paraquat may decrease the number of nigral neurons without triggering motor impairment (Fernagut 2007). This can be consequent to the low level of DA reduction or limited neuronal loss observed following the treatment.

- The repeat dose administration of 10mg/kg i.p. is likely representing the maximum tolerated dose of the chemical stressor. The observed movement disorders can, at least in part, come from systemic illness and the contribution of systemic pathological changes to the observed movement disorders cannot ruled out (Cicchetti et al. 2005).

- There is uncertainty on what is the real brain concentration that is triggering this AOP. In addition, because of the complexity of the kinetic e metabolism of the used tool compound, extrapolation of the in vitro concentration to in vivo scenario is an uncertainty.

6. Quantitative Considerations

The quantitative understanding of this AOP includes evidence of response-response relationship and the identification of a potential threshold effect. However, this threshold should be taken into consideration with caution as the triggering effect at MIE level was explored in only few studies and the repeatability of the KE 3 is questionable for some authors. More evidence exists that an increase from 200 to 600% of lipid peroxidation (endpoint of KE1) in DA neuronal cells can be used as a probabilistic threshold triggering the degeneration of DA neurons of the nigrostriatal pathway. In line with others chemical tools that can induce DA neuronal loss through different MIE (i.e. rotenone and MPTP), for the identification of the AO the design of the in-vivo studies should be tailored as to a MIE which leads to a long-lasting perturbation of the KEs. This provides the most specific and definite context to trigger neuronal death. A major hurdle for this AOP is represented by the AO. With PQ, the low level of reported DA neuronal loss (ca.20-30%) is not expected to induce parkinsonian motor symptoms and no essentiality data (i.e recovery of motor symptoms following treatment with DA) are...
available. Moving from a qualitative AOP to quantitative AOP would need a clear understanding of effect thresholds for the different KEs.

Table 20: Concordance table for the tool compound paraquat

<table>
<thead>
<tr>
<th>Dose/Concentration at the target site</th>
<th>MIE</th>
<th>KE1 Mitochondrial dysfunction (ROS production)</th>
<th>KE2 Impaired proteostasis</th>
<th>KE3 Degeneration of DA neurons of nigrostriatal pathway</th>
<th>AO Parkinsonian motor symptoms</th>
</tr>
</thead>
<tbody>
<tr>
<td>10 mg/kg/bw ip single dose (non-cumulative concentration), corresponding to 0.78 µM brain concentration [1 and 2]</td>
<td>No data</td>
<td>200% increase in lipid peroxidation[4]</td>
<td>No data</td>
<td>No effect [4]</td>
<td>No data</td>
</tr>
<tr>
<td>5mk/kg/bw (intended as a cumulative concentration; 8 doses) [8]</td>
<td>42% increase in SOD activity[8]</td>
<td>No data</td>
<td>10 % decrease in TH+ neurons [8]</td>
<td>No data</td>
<td></td>
</tr>
<tr>
<td>(10mg/kg/bw i.p. corresponding to 5.4 µM (cumulative concentration after multiple doses) [1 and 2]</td>
<td>75% increase in SOD activity[8]</td>
<td>500-600% (cumulative effect) lipid peroxidation [2, 4]</td>
<td>50% increase in 20S proteasome fraction at 24 hrs [2] Intracellular deposits of α-synuclein in 30% of DA neurons [6] 50% increase in α-synuclein expression [8]</td>
<td>30-50% (cumulative effect) decrease in TH+ neurons [2, 4, 6, 7, 8]</td>
<td>Motor impairment [2, 8, 10]</td>
</tr>
</tbody>
</table>

[1] Breckenridge 2013
7. Applicability of the AOP

This proposed AOP is neither sex-dependent nor associated with certain life stage; however, aged animals are considered more sensitive. The relevance of this AOP during the developmental period has not been investigated.

In vivo testing has no species restriction. However, host susceptibility is likely to have a relevant impact on the outcome of the studies and in this context, elements of stress and animal strain could have a profound impact on the outcome of the studies (Jones et al. 2014, Jiao et al. 2012). The mouse was the species most commonly used in the experimental models conducted with the chemical stressor paraquat and the C57BL/6J is considered the most sensitive mouse strain (Jiao et al. 2012). However, animal models (rodents in particular) would have limitations as they are poorly representative of the long human life-time as well as of the human long-time exposure to the potential toxicants. Human cell-based models would likely have better predictivity for humans than animal cell models if biologically relevant by means of being able of recapitulate the key events in the toxicology and pathology pathway providing robust and repeatable results predictive of the chemical concentration that lead to a particular outcome. In this case, toxicokinetics information from in-vivo studies would be essential to test the respective concentrations in-vitro on human cells.

8. Regulatory considerations

The AOP is a conceptual framework to mechanistically understand apical hazards. The AO, parkinsonian motor symptoms, is an apical endpoint that can be explored and quantified in the regulatory toxicology studies conducted in experimental laboratory animals. However, it is noteworthy that decrease in neuronal cell count is also an apical regulatory endpoint explorabale and quantifiable in the regulatory toxicology studies conducted in-vivo; if the appropriate areas of the brain are sampled and properly evaluated. A statistically significant decrease in DA neuronal cell count is considered an adverse event, regardless of the concomitant presence of motor symptoms. This has to be taken into consideration for the potential regulatory applications of this AOP and for the sensibility of the method applied to capture the KE/apical endpoint/hazard. If the intention is to use this AOP for defining the link between the MIE and the degeneration of DA neuronal cells of the nigrostriatal pathway, the WoE is considered strong; however, in the case of defining the link between the MIE and parkinsonian motor symptoms, the WoE should account for the biology and complexity linking disruption of the nigrostriatal pathway and occurrence of motor symptoms. Because of the potential different uses of this AOP, keeping the parkinsonian motor symptoms as AO was considered relevant. It is also foreseen that for potential additional uses, like defining a testing strategy or properly design an in-vitro or an in-vivo study, or evaluation of mixture of chemicals, degeneration of DA neuronal cells of nigrostriatal pathway should be considered as AO.

9. Potential application of this AOP

This AOP was developed in order to evaluate the biological plausibility that the adverse outcome i.e. parkinsonian motor deficits, is linked to the selected MIE. By means of using a human health outcome from epidemiological studies and metaanalysis, the authors intend to embed the AO in the process of hazard identification. This AOP can be used to support the biological plausibility during the process of evaluation of epidemiological studies when a chemical substance is known to interfere with the proposed pathway.

In addition, this AOP can be used to support identification of data gap that can be required or explored when a chemical substance is affecting the pathway or provide recommendation on the most adequate study design that can be applied to investigate the apical endpoints. It is important to note that, although the AO is defined in this AOP as parkinsonian motor deficits, degeneration of DA neurons is already per se an adverse event even in situations where is not leading to parkinsonian motor deficits or clinical signs indicative of a central effect, and this should be taken into consideration for the regulatory applications of this AOP. In addition, this AOP can inform on the identifications of in vitro methods that can be developed for an integrated approach to testing and assessment (IATA) based on in vitro neurotoxicity assays complementary to in vivo assays.
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Appendix B – AOPs developed for Infant Leukaemia and Childhood Leukaemia

Adverse Outcome Pathway (AOP) 3: In utero DNA topoisomerase II poisons leading to infant leukaemia

Introduction

Infant leukaemia is a rare haematological disease (1 in $10^6$ newborns, accounting for 10% of all childhood acute lymphoblastic leukaemias (ALL)) manifesting soon after birth (<1 year) and having a poor prognosis (Sanjuan-Pla et al 2015). Compared to the more frequent childhood leukaemia, infant leukaemia show distinct features:

- An early neonatal onset linked to its plausible origin as a ‘intrauterine developmental disease’ (Greaves 2015; Sanjuan-Pla et al 2015);
- Rearrangements of the mixed-lineage leukaemia (MLL; KMT2A) gene on the q23 band of chromosome 11, as the hallmark genetic abnormality (Joannides and Grimwade 2010);
- However, MLL is not the only translocation gene; for infant ALL, about 60-80% carry an MLL rearrangement (Sam et al.2012; Jansen et al.2007) and the percentage for infant acute myeloid leukaemia (AML) is about 40 %;
- The MLL rearrangement at an early stage of development; the likely target cells (still unidentified) are the hematopoietic stem and progenitor cells (HSPC) in fetal liver and/or earlier (mesenchymal) stem cells in embryonic mesoderm (Bueno et al 2009; Menendez et al 2009);
- The infant MLL-rearranged leukaemia carries less somatic mutations (1.3 vs 6.5/case) than the childhood disease (Andersson et al 2015; Dobbins et al 2013), pointing to the lack of a “second hit” and suggesting a “one big hit” origin.

Overall, based on the available evidence, infant leukaemia pathogenesis originates from a single, severe hit to a target cell during early intrauterine development. Whereas the limited epidemiological studies do not allow any firm conclusion on a possible role for chemicals in infant leukaemia (Pombo-de-Oliveira et al 2006; Ferreira et al 2013), exposures to chemicals able to induce MLL rearrangements through topoisomerase II (TopoII) “poison”, particularly etoposide and other TopoII “poisons”, including some bioflavonoids, have been suggested as agents promoting the driver genetic oncogenic event. Experimental models for infant leukaemia have been developed, but a wholly satisfactory model reproducing the phenotype and latency is not yet available.

Nevertheless, the anticancer drug etoposide can be considered as a model chemical for DNA topoisomerase II "poison". Acute leukaemia is an adverse effect recorded in etoposide-treated patients, showing MLL rearrangements that are in many ways analogous to those in infant leukaemia (Bueno et al 2009; Joannides et al 2010, 2011). Therefore the proposed AOP is supported by a number of convincing inferential evidences by means of using etoposide as a tool compound to empirically support the linkage between the proposed molecular initiating event (MIE) and the adverse outcome (AO). In the meanwhile, this AOP identifies several knowledge gaps, the main ones being the identification of the initiating cell and the investigation of TopoII poisons in a robust model; thus, the present AOP may be modified in future on the basis of new evidence.
References


Ferreira JD, Couto AC, Pombo-de-Oliveira MS, Koifman S; Brazilian Collaborative Study Group of Infant Acute Leukemia. In utero pesticide exposure and leukemia in Brazilian children < 2 years of age. Environ Health Perspect 2013 Feb;121(2):269-75. doi: 10.1289/ehp.1103942.


Abbreviations: TopoII, DNA topoisomerase II; HSPC, hematopoietic stem and progenitor cell; t-AL, therapy-associated acute leukaemia;
AOP

Adverse Outcome Pathway (AOP): In utero DNA topoisomerase II inhibition leading to infant leukaemia

Fig 32: AOP scheme

MIE: In utero exposure to DNA topoisomerase II “poisons”

How this MIE works

DNA topoisomerase (Top) II enzyme “poisons” disturb the normal TopoII enzyme function and cause a ‘hanging double strand break (DSB)’ at a specified DNA sequence. The above description of the MIE is of significance because there are 3 different kinds of “poisons” of TopoII enzyme, out of which competitive inhibitors prevent the function of the enzyme and cause cell death, whereas other interfacial and covalent inhibitors may cause – depending on the situation – other consequences of DNA damage response including chromosomal rearrangements (Pendleton et L 2014; Lu et al 2015).

A further prerequisite for the specific outcome, i.e. creation of chromosomal rearrangement, is that TopoII “poison” has to occur in an especially vulnerable and correct hot spot in the MLL locus in the right target cell vulnerable to transformation.

TopoII enzymes have several crucial functions in DNA replication, transcription, repair and chromatin remodelling, i.e. TopoII enzymes take care of DNA integrity and topology. Because the enzyme functions by passing an intact double helix through a transient double-stranded break, any disturbances in its function, e.g. by chemical inhibitors, could have a profound effect on genomic stability, resulting in DNA repair response, gene and chromosomal damage, initiation of apoptosis and ultimate cell death. A double-strand break and error-prone non-homologous end-joining (NHEJ) DNA repair mechanism may lead to gene rearrangements; chromosomal translocations and consequently fusion genes (see Figure 33). A comprehensive description of TopoII enzymes and their functions and derangements could be found in recent review articles (Cowell and Austin 2012; Pendleton et al 2014; Ketron and Osheroff 2014).
**Fig. 33:** TOP2 Poisons, downstream events. TOP2 poisons inhibit the religation step of the TOP2 reaction cycle, leading to accumulation of covalent TOP2-DNA cleavage complexes. These lesions are cytotoxic and lead to activation of the DNA damage response and potentially apoptosis. Alternatively these lesions are repaired, largely through the non-homologous end-joining pathway. Translocations observed in therapy-related leukemia are presumed to occur as a result of mis-repair, joining two heterologous ends. (from Cowell and Austin 2012)

**How it is measured or detected**

The identification and measurement of the inhibition of TopoII enzymes is made more difficult by the presence of different molecular mechanisms (see above). However, some assays are used in pharmacological research to screen TopoII “poisons”, including cell-free decatenation assay (Schroeter et al., 2015). The most important mode, the cleavage activity of TopoII can be studied in vitro, by using a human recombinant enzyme and an appropriate double-stranded plasmid as a target to quantitate double-strand breaks (Fortune and Osheroff 1998). A cleavage can also be indirectly detected by measuring various indicators of DNA damage response, such as ATM activity, p53 expression, γH2AX or Comet assay (Li et al 2014, Schroeter et al., 2015, Castano et al 2016).

It is useful to note that several chemicals identified as TopoII “poisons” do require metabolic oxidation to become active inhibitors. Etoposide itself is converted via the catechol metabolite to etoposide 3-quinone, which is a covalent TopoII poison (Smith et al 2014), whereas etoposide and its catechol are interfacial inhibitors. Curcumin is also an active TopoII poison due to its oxidized metabolites (Gordon et al 2015). This fact deserves consideration if a screening for TopoII inhibition is envisaged.

**Evidence supporting taxonomic applicability (tissue type, taxa, life stage, sex)**

DNA topoisomerases are ubiquitous enzymes, which control the integrity of double-stranded DNA. They are thus key enzymes at all levels of living organisms. The available evidence suggest that important differences in sensitivity to topoisomerase inhibition might exist among different cell types, depending on the amount of proliferative burden, of the TopoII enzymes and on physiological repair processes. Mesodermal precursor or hematopoietic stem and progenitor cells (HSPCs) are rapidly dividing cells with a high content of TopoII and for these reasons they can be a sensitive target during a critical developmental window (Hernandez and Menendez 2016). In addition, evidence from micronuclei assay studies conducted in untreated and chemical-treated foetuses and newborns show that both the baseline and chemically induced micronuclei frequencies are higher in the foetuses and infants than in adults (Udroiu et al 2016). This is possibly indicating a greater sensitivity to genotoxic insult during development which can be due to the higher proliferation rate and lower ability of DNA repair of the hematopoietic stem cells. However, the role that the different microenvironments (foetal liver, infant bone marrow and adult bone marrow) during ontogenesis can exert on cell sensitivity...
cannot be ruled out (Udroiu et al. 2016). The existence of relevant interspecies differences is unknown, but it cannot be ruled out presently.

**Evidence for Chemical Initiation of this Molecular Initiating Event (MIE)**

A number of drugs, environmental chemicals and natural substances are identified as TopoII "poisons" (Pendleton et al 2014) (Table 21). A well investigated example is the anticancer drug etoposide; also bioflavonoids, e.g. genistein, (Barjesteh van Waalwijk van Doorn-Khosrovani et al 2007; Azarova et al 2010) bind to TopoII enzymes, induce cleavage in the MLL gene and produce a fusion gene (and its product) in human cells. The organophosphate pesticide chlorpyrifos has been shown to inhibit ('poison') the enzyme *in vitro* (Lu et al 2015).

**Table 21.** TopoII poisons

<table>
<thead>
<tr>
<th>Chemical class</th>
<th>Examples</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Anticancer agents</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Epipodophyllotoxins</td>
<td>etoposide, teniposide</td>
<td>Montecucco et al 2015</td>
</tr>
<tr>
<td>Anthracyclines</td>
<td>doxorubicin, epirubicin, daunorubicin, idarubicin, aclarubicin</td>
<td>Cowell and Austin 2012</td>
</tr>
<tr>
<td>Anthracenedione</td>
<td>Mitoxantrone</td>
<td>Cowell and Austin 2012</td>
</tr>
<tr>
<td>Acridines</td>
<td>Amsacrine</td>
<td>Cowell and Austin 2012</td>
</tr>
<tr>
<td><strong>Bioflavonoids</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Flavones</td>
<td>luteolin, apigenin, diosmetin</td>
<td>Ketron and Osheroff 2014</td>
</tr>
<tr>
<td>Flavonols</td>
<td>myricetin, quercetin, kaempferol, fisetin</td>
<td>Ketron and Osheroff 2014</td>
</tr>
<tr>
<td>Isoflavones</td>
<td>Genistein</td>
<td>Ketron and Osheroff 2014</td>
</tr>
<tr>
<td>Catechins</td>
<td>EGCG, ECG, EGC, EC</td>
<td>Ketron and Osheroff 2014</td>
</tr>
<tr>
<td>Isothiocyanates</td>
<td>benzyl-isothiocyanate, phenethyl-isothiocyanate, sulforaphane</td>
<td>Ketron and Osheroff 2014</td>
</tr>
<tr>
<td>Other phytochemicals</td>
<td>Curcumin</td>
<td>Ketron and Osheroff 2014</td>
</tr>
<tr>
<td><strong>Environmental chemicals</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Aromatic compounds</td>
<td>benzene, PAHs</td>
<td></td>
</tr>
<tr>
<td>Nitrosamines</td>
<td>Diethylnitrosamine</td>
<td>Thys et al 2015</td>
</tr>
<tr>
<td>Organophosphates</td>
<td>Chlorpyrifos</td>
<td>Lu et al 2015</td>
</tr>
</tbody>
</table>

Etoposide

Much of the relevant, albeit indirect, evidence to support this AOP come from the studies on etoposide, an anticancer drug TopoII "poison", which is known to induce therapy-associated acute leukaemia (t-AL) in adults (Cowell and Austin 2012; Pendleton et al 2014). It is of interest that the
latency of t-AL is <2 years between the treatment of the primary malignancy and the clinical diagnosis of the secondary disease and that the prognosis of t-AL is poor (Pendleton et al 2014). t-AL is characterized by the MLL rearrangements and it is practically certain that these fusion genes are caused by etoposide or anthracyclines treatment; because MLL rearrangements have not been detected in bone marrow samples banked before the start of the treatment of the first malignancy. Also the breakpoints in MLL or partner genes fall within a few base pairs of a drug-induced enzyme-mediated DNA cleavage site (Pendleton et al 2014).

Etoposide can induce MLL rearrangements in different cell types; interestingly, embryonic stem cells and their hematopoietic derivatives are much more sensitive than cord blood-derived CD34+ cells to etoposide induced MLL rearrangements; in addition, undifferentiated human embryonic stem cells (hESCs) were concurrently liable to acute cell death (Bueno et al., 2009). These findings suggest that the MIE should be put into evidence in target cell models with appropriate sensitivity.

Bioflavonoids

Bioflavonoids are natural polyphenolic compounds in a large variety of plant-derived food items. TopoII-mediated DNA cleavage has been linked to genistein, kaempferol, luteolin, myricetin and apigenin (Strick et al 2000; Bandele and Osheroff 2007; Azarova et al 2010; Lopez-Lazaro et al 2010), although the concentrations in in vitro studies have been quite high. It has also been demonstrated that several bioflavonoids are capable of inducing the cleavage of the MLL gene in human cell lines (Strick et al 2000; van Doorn-Khosrovani et al 2007). The in vitro effects of bioflavonoids suggested a possible link between dietary intake and infant leukemia (e.g., Azarova et al., 2010; Lanoue et al., 2010); however until now, epidemiological evidence existing to support or refute such a hypothesis is based on small studies (Ross et al 1996; Spector et al 2005).

Chlorpyrifos

Chlorpyrifos is a widely used organophosphate insecticide, which has been suspected as a risk factor for infant and childhood leukaemia after the household exposure of pregnant women (r). According to Lu et al (2015), chlorpyrifos and its metabolite chlorpyrifos oxon exhibit an inhibitory effect on in vitro TopoII activity. Chlorpyrifos causes DNA double strand breaks as measured by the neutral Comet assay and induces MLL gene rearrangements in human fetal liver-derived CD34+ hematopoietic stem cells via TopoII ‘poisoning’ as detected by the FISH assay and in vitro isolated TopoII inhibition assay, respectively (Lu et al 2015). Chlorpyrifos also stabilizes the TopoII-DNA cleavage complex. Etoposide was used a positive reference compound in these studies and it performed as expected. The lowest concentration of chlorpyrifos used was 1 µM and it gave a statistically significant effect in many in vitro assays. The point of departure of etoposide, which was calculated to be 0.01 to 0.1 µM (Li et al 2014), is at least 10-fold lower than that of chlorpyrifos.
References


KE1: In utero MLL chromosomosomal translocation

How this key event works

Chromosomal rearrangements of the mixed-lineage leukaemia (MLL) gene, located on the q23 band of chromosome 11 (11q23), are the genetic hallmark of most infant leukaemias (Meyer et al 2013; Sanjuan-Pla et al 2015). MLL is located within the fragile site FRA11G; chromosomal fragile sites are regions of the genome susceptible to breakage under conditions of replication stress; interference with TopoII may promote fragile site instability. MLL encodes a protein homologous to the Drosophila trithorax gene, which has relevant functions in embryogenesis and hematopoiesis (Ernest et al 2004, Hess et al 1997).

There are many translocation and fusion partners for MLL; DNA breakage within MLL can lead to rearrangement with over 120 partner genes (Meyer et al 2013). In principle all MLL fusion genes are potential initiating drivers, although clinical studies have shown a preponderance with infant leukaemia for only a few of these rearrangements. For infants diagnosed with ALL, approximately 60-80% carry an MLL rearrangement (Sam et al 2012; Jansen et al 2007), with predominant fusion partners being AF4 (41%), ENL (18%), AF9 (11%) or another partner gene (10%). In particular, the fusion gene MLL-AF4 shows a specific and consistent relationship with the disease (Menendez et al., 2009): however, it has been difficult to reproduce a manifest disease resulting from this rearrangement in in vivo animal models. For AML, about 30% of the patients carry an MLL rearrangement.

The occurrence of MLL rearrangements at a very early fetal development is highly probable on the basis of neonatal blood spot analysis and by the high concordance rate of infant leukaemia in monozygotic twins (Ford et al 1993; Gale et al 1997; Sanjuan-Pla 2015). Menendez et al (2009) showed that MLL-AF4 fusion gene is present in bone marrow mesenchymal stem cells in infant leukaemia patients, but not in patients of childhood leukaemia, suggesting that the origin of the fusion gene is probably prehaematopoietic. Consequently, the affected cell, the so called leukaemia-initiating cell, may be an early prehaematopoietic mesodermal precursor, a hematopoietic stem cell or hematopoietic progenitor cell residing mainly in the liver (Greaves 2015; Sanjuan-Pla et al 2015).

MLL protein (complexed with a large number of other protein factors) serves as a transcriptional activator or repressor via the binding to promoter regions of active genes, marking these regions by covalent histone modifications (Sanjuan-Pla et al 2015). Translocation and creation of fusion genes and products destroys the intrinsic control mechanisms of the MLL protein. The resulting ‘ectopic’ functions involve promoter hyper-activation and re-acquiring stem cell features (Sanjuan-Pla et al 2015). A schematic presentation of the drastic changes of the MLL product is depicted in figure 34.

Report title


Fig 34: Proposed model for the oncogenic conversion of MLL fusions: A. Physiological situation and B: Chromosomal translocation, which leads to the intrinsic regulatory mechanism of MLL being destroyed. (Sanjuan-Pla et al 2015.)

MLL translocation sites (breakpoint sequences) in the therapy-related leukaemia fall within a few base pairs of etoposide-induced enzyme-mediated DNA cleavage site (r). Although rearrangements associated with infant leukaemias are often more complex than those observed in treatment-related leukaemias, many are nevertheless associated with stable TopoII-mediated DNA cut sites. Although all these findings are indirect regarding infant leukaemia, they are nevertheless rather persuasive in this respect.

Growing scientific evidence, including the stable genome of the patients, suggests that infant leukaemia originates from one “big-hit” occurring during a critical developmental window of stem cell vulnerability (Andersson et al 2013; Greaves 2015). Therefore, the totality of evidence suggests the essential role of the formation of MLL-AF4 (and other partner) fusion gene and product in causing pleiotropic effects in the affected cell and directing it to the obligatory pathway to the adverse outcome of leukaemia (see KER2).

How it is measured

- The presence and structure of a fusion gene can be identified with PCR or related techniques. Mapping of cleavage sites in the gene needs genomic DNA. In cells or tissues, the detection of a fusion gene is possible by appropriate immunofluorescent techniques.

- Assays measuring chromosomal aberrations, micronuclei or DNA and chromosome damage (Comet assay) may indirectly identify the KE through its consequences in experimental systems in vitro and in vivo; the degree of accuracy of such identification cannot be evaluated presently.
**Taxonomic applicability**

Although the KE deals with the general process of DNA integrity, the available evidence do not allow for evaluating whether any significant difference occurs among cell types or species. It has been shown that the mouse has an analogous fusion gene *mll*-af4. A recent study has shown that in utero exposure to etoposide induces *mll* translocations in in Atm-knockout mice, which are defective in the DNA damage response, albeit not in wild-type mice; moreover, fetal liver hematopoietic stem cells were more susceptible to etoposide than maternal bone marrow mononuclear cells, pointing out the life stage-related susceptibility in regards to TopoII “poison” also in the mouse (Nanya et al., 2015).

**References**


Short title

Adverse Outcome (AO) Infant leukaemia

How this key event works

Symptoms of leukaemia – thrombocytopenia resulting in sensitivity to bruising and bleeding, anaemia with pallor and fatigue, neutropenia associated with increased susceptibility to infections – are principally due to the displacement of the normal haematopoiesis by expansion of leukaemia cells. Leukemic infiltration of the brain is common at diagnosis of the infant leukaemia (Hunger and Mullighan, 2015).

How it is measured

Haematological methods – identification of leukaemia cells and routine blood cell counts; observations of clinical symptoms.

Following clinical diagnosis, methods for refined diagnosis include bone marrow aspirates for immunophenotypic analyses and cytogenetic assays for molecular stratification.

The Cancerogenicity assays and the extended one generation test (OECD 443) include endpoints that can potentially explore the AO; however, considerations should be made on the specificity of the disease to humans.

Taxonomic applicability

Infant leukaemia is a paediatric leukaemia likely resulting from gene-environmental interactions. The limited data available suggest that dietary and environmental exposure to substances targeting topoisomerases together with reduced ability of the foetus or their mother to detoxify such compounds because of the polymorphic variants of given genes could contribute to the development of this AO (Hernadez et al. 2016).

In animals the disease is not known and artificial animal models able to reproduce the disease have limitations. Bardini et al (2015) has however developed a xenograph mouse model with patient MLL-AF4-involving leukoblasts transplanted.

Regulatory relevance of the AO

Genotoxicity in general and carcinogenicity are apical endpoints in established regulatory guideline study. TopoII poisoning has been listed as one of the potential mechanisms of genotoxicity and carcinogenicity in the ICH M7 guideline for human medicines. It is also known that some manifestations of genotoxicity in tests measuring chromosomal aberrations, micronuclei or DNA and chromosome damage (Comet assay) are partially due to double-strand breaks created by the disturbed action of TopoII enzymes.

The extended one generation test (OECD 443) includes a developmental immunotoxicity cohort. At present the cohort may identify post-natal effects of prenatal and neonatal exposures on the immune tissues and white blood cells population. However, each regulatory guideline study has potential limitations e.g. no specific parameters are in place to identify a pattern relevant to infant leukemia in humans in the extended one generation test, no treatment is occurring during the early in-utero development phase in the carcinogenicity assay and no considerations on the possible higher sensitivity of the HSC are in place for the genotoxicity assays.

Epidemiological evidences are linking pesticide exposure to infant leukaemia, also suggesting that pesticide exposure may have a greater impact on children than adults; though, almost all of the available evidence are not making a distinction between infant and childhood leukaemia. However, most epidemiological studies are limited because no specific pesticides have been directly associated with the risk of leukaemia, but rather the broad term “pesticide exposure” (Hernandez and Menendez 2016). In this perspective, this AOP would provide a regulatory relevant support for understanding the potential of a chemical to be involved in this toxicological pathway.
References


1st KER: In utero DNA topoisomerase II inhibition (KE up) leading to In utero MLL chromosomal translocation (KE down)

How this Key Event Relationship works

Certain TopoII poisons stabilize the intermediate cleavage complex and prevent the religation with appropriate DNA strands. Covalently DNA end-bound TopoII protein is digested and a hanging end is created. The same process happens in the translocation partner gene. Hanging ends of both genes are processed and subsequently joined by non-homologous end joining (Cowell and Austin 2012).

There is evidence that this inappropriate joining of ‘hanging ends’ happens in the same transcriptional factory (hub), and the result is a fusion gene and ultimately protein product (Cowell & Austin 2012; Pendleton et al 2014; Sanjuan-Pla et al 2015). The first part of this description has not been shown in the putative target cell, which is still not unequivocally identified, but for the second part there is ample evidence of formation of MLL-AF4 fusion product that has been a result of a very early chromosomal translocation and rejoining. It is of interest that the simultaneously induced specific DSBs in the MLL gene and two different translocation partners (AF4 and AF9) by engineered nucleases in human HSPCs resulted in specific ‘patient-like’ chromosomal translocations (Breese et al 2016).

Weight of Evidence

Evidence supporting the causal relationship between etoposide-induced TopoII inhibition and the MLL rearrangement leading to the fusion gene is strong regarding treatment-related acute leukemia (*Cowell and Austin 2012; *Pendleton et al 2014). The bioflavonoid-rich diet in pregnant women has been suggested to initiate infant leukaemia by an analogous causality between in utero inhibition of TopoII enzymes and creation of the fusion gene. However, there is no direct evidence in humans and it is also difficult or impossible to study. Power of epidemiological studies is relatively weak in the case of a very rare disease and case-control or spatiotemporal cluster studies have been barely suggestive to indicate a causal relationship between exposures and disease.

Biological plausibility

The KER as such is biologically plausible. Type II topoisomerases are ubiquitous enzymes which are essential for a number of fundamental DNA processes. As they generate DNA strand breaks, they can potentially fragment the genome. Indeed, while these enzyme are essential for the survival of proliferating cells they can also have significant genotoxic effects by means of accumulation of DNA strand breaks that, if not resulting in cell death may lead to chromosomal translocation in the surviving cell population (McClendon et al. 2007). DNA breaks and MLL rearrangements by etoposide and bioflavonoids have been demonstrated in human fetal liver haematopoietic stem cells, in human embryonic stem cells and in human prehaematopoietic mesenchymal stem cells as well as in cord blood mononuclear cells (Ishii et al 2002; Blanco et al 2004; Moneypenny et al 2006; Bueno et al 2009; Menendez et al 2009), which clearly shows that TopoII-associated MLL rearrangements are produced in appropriate human cells in utero.
Empirical support for linkage

There are animal models for infant leukaemia which recapitulate at least some salient aspects of the disease (Sanjuan-Pla et al 2015). However, for example the MLL-AF4 knock-in mice develop leukaemia only after a prolonged latency (Chen et al 2006), thus not recapitulating the ‘pathognomonic’ feature of infant leukaemia.

Etoposide treatment in vivo in mice at day 13.5 of pregnancy induces MLL breakage in fetal liver haematopoietic stem cells in utero, but MLL-rearranged fusion mRNAs were detected only in mice which were defective in the DNA damage response, i.e. atm knockout mice. A fusion gene analogous to MLL-AF4 was not detectable in the wild type mice. In this study, an intraperitoneal injection of 10 mg/kg of etoposide into pregnant mice at day 13.5 of pregnancy resulted in a maximum fetal liver concentration of about 5 µM. A dose of 0.5 mg/kg did not result in a measurable concentration. A statistically significant increase (about 6-fold) in DSBs in the MLL gene of isolated fetal liver haematopoietic stem cells was observed after a single dose of 1 mg/kg to pregnant mice6. A clear activation of DNA damage response was observed at the dose of 10 mg/kg (Nanya et al. 2016).

There is a lot of information about the interaction of etoposide with TopoII enzymes and MLL chromosomal translocation at the cell culture level and in connection with treatment-related leukaemia.

Molecular dose-response modelling of etoposide-induced DNA damage response, based on comprehensive in vitro high content imaging in the HT1080 cell model, was developed by Li et al (2014). The model was based on the hypothesis that cells are capable of clearing low-level DNA damage with existing repair capacity, but when the number of DSBs exceeds a certain value; ATM and p53 become fully activated through reversible mechanism, leading to elevated repair capacity. The model was able to capture quantitatively the dose-response relationships of a number of markers observed with etoposide. Especially interesting are the dose-response relationships for activation of p53 and the formation of micronuclei in the target cell model, which indicate point-of-departure concentrations of etoposide in the range of 0.01 to 0.1 µM (Li et al. 2014). This range is in agreement with the finding that in human fetal liver CD34+ cells an increase in DSBs was observed at a concentration of 0.14 µM and MLL translocations were detectable by FISH or flow cytometry at higher concentrations (Moneypenny et al 2006).

Uncertainties and Inconsistencies

- A prerequisite for the specific outcome, i.e. creation of chromosomal rearrangement, is that TopoII inhibition has to occur in an especially vulnerable and correct hot spot in the MLL locus; however, details of this process and how it happens are not clear.
- A target cell, i.e. leukaemia-initiating cell, has not been identified with sufficient confidence and consequently there is no target cell model to recapitulate the linkage between TopoII inhibition (‘poisoning’) and the production of DSB in an appropriate target. Recently, by the expression of engineered nucleases (TALENs) to induce simultaneous patient specific double strand breaks in the MLL gene and two different known translocation partners (AF4 and AF9), Breese et al (2015) were able to produce specific chromosomal translocations in K562 cells and in primary HSPCs.
- In-utero etoposide-treatment failed to induce leukaemogenesis (Nanya et al 2015). Consequently, the envisaged linkage has not been empirically supported or rejected. However, it should be kept in mind that, whereas etoposide does induce a large number of MLL rearrangements, most of them occur within non-coding regions, therefore not eliciting any direct oncogenic consequence. A MLL-AF4 in frame fusion is a rare event that needs to occur in a target cell within a relatively small and spatially restricted cell population during the appropriate, epigenetically plastic, developmental window; thus it may be difficult to empirically support this process.
- Dose-response relationships between etoposide and treatment-related leukaemia are difficult to unravel, but risk of leukaemia seems to increase with larger total exposure to etoposide.

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6 Hypothetically, based on linear extrapolation from the dose of 10 mg/kg, the concentration would be of the order of 0.5 µM.
However, comparison of exposures or kinetics of etoposide between leukaemia patients and non-leukemic treated subjects did not reveal any significant differences (Relling et al 1998). Also, it is not known whether the etoposide (or metabolite) concentrations during the treatment are of significance. In child and adult chemotherapy, concentrations are extremely variable between individuals; the lowest through plasma concentrations of etoposide have been of the order of 1 µM and peak concentrations very much higher. For example, in a study of Relling et al (1998), the maximum plasma concentration of etoposide was about 90 µM and that of etoposide catechol about 100-times less, below 1 µM. In another high dose chemotherapy study (Stremetzne et al 1997), the etoposide concentration was 170 µM and that of the catechol metabolite 5.8 µM maximally. However, it is not straightforward to juxtapose plasma concentrations and the tissue or cell concentration which TopoII enzyme ‘sees’. Penetration of etoposide or its metabolite through plasma membrane is probably rather slow and it has been shown that the brain cancer tissue (metastasis or glioma) to plasma ratio for etoposide is only 0.1 (Pitz et al 2011). Blood-brain barrier is not necessarily a good model for cross-membrane distribution, but may give some idea about the general distributional behaviour of a drug. Even if the active target concentration of etoposide is only 10 % of the plasma concentration, it is still in the same range as the effective concentrations in cellular studies (see above). A final note on relevant concentrations: etoposide concentrations resulting in DSB and fusion gene are probably within a relatively restricted range. The concentration resulting in a proper fusion gene should be in a range which gives rise to a partially repaired insult and cells bypassing death and accumulating the abnormality.
References


2nd KER: In utero MLL chromosomal translocation (KE up) leading to Infant leukaemia (KE down)

How this Key Event Relationship works

Propagation of a leukaemic cell clone is based on both blockage of differentiation to more mature cells and ability to expand in an uncontrolled way. Formation of the MLL-rearranged fusion genes and their protein products are intimately involved in both the blocked differentiation of HSPCs and the expansion of the fusion gene-carrying clone. It is believed that the fusion gene product block cell differentiation by inhibiting the normal transcriptional programs and recruiting repressor molecules such as histone deacetylase enzymes (Greaves 2002; Teitell and Pandolfi 2009). Furthermore, the fusion gene product activates other key target genes, which ultimately lead to the propagation of transformed cell lines without normal restrictions (Greaves 2015; Sanjuan-Pla et al 2015). Therefore, the potential of both differentiation blockage and clonal expansion are inherent properties of the MLL-rearranged fusion product, based on the preservation of some original functions, even if in a modified form, and on the gain of some other functions due to the sequences from the new fusion partner gene (Marschalek 2010; Sanjuan-Pla et al 2015).

Molecular mechanisms

The MLL is the most common translocation gene in infant leukaemia. The N-terminal part of MLL becomes fused in frame to one of a large number of fusion partners, but in most cases, this fusion occurs between the N-terminal MLL and either AF4, AF6, AF9, AF10, or ENL (Krivtsov and Armstrong 2007). Due to the DNA-binding properties of the N-terminal MLL motif, these fusion proteins are always nuclear and bind to target genes controlled by MLL irrespective of the normal location of the C-terminal partner.

Many fusion proteins have been shown to recruit DOT1L (catalyzing methylation of histone H3K79) to the promoters of MLL target genes and this recruitment seems to be a common feature of many oncogenic MLL fusion proteins. Although DOT1L is not genetically altered in the disease per se, its mislocated enzymatic activity is a direct consequence of the chromosomal translocation. Thus, DOT1L has been proposed to be a catalytic driver of leukemogenesis (Chen and Armstrong 2015). The enzymatic activity of DOT1L is critical to the pathogenesis of MLL, because methyltransferase-deficient Dot1L is capable of suppressing growth of MLL-rearranged cells. A small-molecule inhibitor of DOT1L inhibits cellular H3K79 methylation, blocks leukemogenic gene expression, and selectively kills cultured cells bearing MLL translocations (Chen and Armstrong 2015). One of the target gene of DOT1L is BCL-2, belonging to a family of anti-apoptotic genes, which maintains the survival of the MLL-rearranged cells (Benito et al 2015). Expression of BCL-2 is high in human MLL-AF4 leukemia cells from a large number of patients. A specific BCL-2 inhibitor, ABT-199 is capable of killing MLL-AF4 leukaemia cells and prevents cell proliferation in xenograph mouse leukaemia models (Benito et al 2015). Furthermore, a MLL-AF4 cell line is sensitive to a combination of ABT-199 and DOT1L inhibitors. Fig 35 provides a schematic representation of the molecular pathway.
Fig 35: MLL-rearranged acute lymphoblastic leukemias activate BCL-2 through H3K79 methylation and are sensitive to the BCL-2-specific antagonist ABT-199 (Benito et al, Cell Rep 2015).

Possible facilitating mutated genes

Recurrent activating mutations in the components of the PI3K-RAS signalling pathway have been detected in almost half of the tested MLL-rearranged ALLs in one study (Andersson et al 2015). Prenatal origin of RAS mutations have been demonstrated also in other studies of infant leukaemia with frequencies of about 15-25% of cases (Driessen et al 2013; Prelle et al 2013; Emerenciano et al 2015). Emerenciano et al (2015) are of the opinion that RAS mutations seem not to be driver mutations, but may aid disease onset by accelerating the initial expansion of cells.

Overall the activation of the RAS pathway could support the extremely rapid progression of the infant leukaemia. Under this view the mechanism may represent a factor modulating (i.e., increasing) the progression and severity of the adverse outcome, rather than a necessary key event (second hit) for infant leukaemia. In the transgenic MLL-AF4 mouse model, activated K-RAS accelerated disease onset with a short latency (Tamai et al 2011), possibly by augmenting the upregulation of HoxA9. In a recent study of Prieto et al (2016), the activated K-RAS enhanced extramedullary haematopoiesis of MLL-AF4 expressing cell lines and cord blood-derived CD34+ hematopoietic stem/progenitor cells that was associated with leucocytosis and central nervous system infiltration, both hallmarks of infant MLL-AF4 leukaemia. However, K-RAS activation was insufficient to initiate leukaemia, supporting that the involvement of RAS pathway is an important modifying factor in infant leukemia. It has also been demonstrated that MLL-AF6 fusion product sequesters AF6 into the nucleus to trigger RAS activation in myeloid leukaemia cells and it is possible to attenuate the activation by tipifarnib, a RAS inhibitor (Manara et al 2014).

A possibility that MLL fusions render cells susceptible to additional chromosomal damage upon exposure to etoposide was studied by introducing MLL-AF4 and AF4-MLL via CRISPR/Cas9-genome editing in HEK293 cells as a model to study MLL fusion-mediated DNA-DSB formation/repair (Castano et al 2016). In short, the expression of fusion genes does neither influence DNA signaling nor DNA-DSB repair.
Weight of Evidence

The overall scientific evidence, including the stable genome of patients, suggests that infant leukemia originates from one “big-hit” occurring during a critical developmental window of stem cell vulnerability (Andersson et al 2013; Greaves 2015). Different from the “two-hit” model of the adult leukemias, the infant leukemia is a developmental disorder where the clonal expansion is a direct consequence of in utero MLL translocation.

Biological plausibility

The biological plausibility linking the MLL translocation to infant leukemia is strong. Rearrangement in the MLL gene is commonly associated with infant acute leukemia and the disease has unique clinical and biological feature (Ernest et al. 2002). An in utero initiation, an extremely rapid progression, and a silent mutational landscape of infant leukemia suggest that the MLL-translocation-associated gene fusion product is itself sufficient to spawn leukemia and no ‘second hit’ is required. Therapy-related leukemias following exposure to the topo II poisons such as etoposide are characterized by the MLL chromosomal translocation (Libura et al. 2006, Super et al.1993) and translocations involving MLL are associated with a gain of function and leukemogenic effect (Yu et al. 1998). A critical developmentally early window of stem cell vulnerability, involving perhaps lesions based on epigenetically controlled regulatory factors, has been suggested to explain a rare occurrence and an exceptionally short latency of infant leukemia (Greaves 2015; Sanjuan-Pla et al 2015). In primary HSPCs genome engineered for patient specific MLL translocations it was possible to show that this specific ‘artificial’ initiation can induce a selective advantage in survival in extended culturing and a higher clonogenic potential in colony forming assay (Breese et al. 2015).

Empirical support

A number of MLL-fusion products, such as MLL-AF9 and MLL-ENL, have shown leukemogenic potential in cord-blood stem cells. Although the MLL rearrangement is essential to develop leukemia, it alone may not be sufficient and activation of cellular proliferation might be necessary for overt leukemia (Nanya et al. 2015).

There are several animal models, in which MLL-AF4 fusion gene has been expressed (Chen et al 2006; Metzler et al 006; Krivtsov et al 2008; Bursen et al 2008; Tamai et al 2011). In all these models leukemia is ultimately developed, but latency has been very protracted. In any case, one could conclude that the expression of the MLL-AF4 fusion gene is capable of developing leukemia, but it is unknown whether facilitating or necessary changes are required during the long latency in mouse.

Gene engineered human HSPCs carrying MLL rearrangements showed that a subset of cells persisted over time and demonstrated a higher clonogenic potential in colony forming assay (Breese et al. 2015).

Transcription activator-like effecter nuclease (TALEN)-mediated genome editing generated endogenous MLL-AF9 and MLL-ENL oncogenes in primary human HSPCs derived from human umbilical cord plasma (Buechele et al 2015). Engineered HSPCs displayed altered in vitro growth potential and induced acute leukemias following transplantation in immunocompromised mice at a mean latency of 16 weeks. The leukemias displayed phenotypic and morphologic similarities with patient leukemia blasts, expressed elevated levels of crucial MLL-fusion partner target genes, displayed heightened sensitivity to DOT1L inhibition, and demonstrated increased oncogenic potential ex vivo and in secondary transplant assays.

Uncertainties and Inconsistencies

- The MLL-AF4 knock-in mice develop leukaemia only after a prolonged latency (Chen et al 2006), thus not recapitulating the ‘pathognomonic’ feature of infant leukaemia. Also other animal models have been developed with similar results. Thus, an adequate experimental model for infant leukaemia is still in need.

- The role of a reciprocal fusion gene AF4-MLL in leukemias is controversial: it has a transformation potential in animal model (Bursen et al 2010), but it is not expressed in all MLL-AF4 patients (Andersson et al 2015). The potential role of other reciprocal fusion genes has not been studied.
Beyond MLL rearrangements, activation of cellular proliferation by mutation or other (epi)genetic insults might be necessary for overt leukaemia. Further studies are necessary to fully understand which factors would contribute to convey a proliferative advantage, as observed in cells with MLL translocation, to leukaemia.

Quantifiable understanding

Relationships between different fusion genes and subsequent leukemia types are incompletely understood. Although roughly 70-80% of infant B-ALL leukemias carry MLL rearrangements, in 20-30% of the cases there are no MLL rearrangements. In AML and T-ALL leukemia cases MLL rearrangements are even rarer.
References


Greaves M. Childhood leukaemia. BMJ 2002; 324: 283-287


Overall assessment of the AOP

Infant leukaemia is a "hidden" disease quite concretely: initiation occurs in utero at an early phase of foetal development. Studies both in identical twins (Ford et al 1993) and in neonatal blood samples retrospectively (Gale et al 1997) strongly indicate in utero origin of the disease. Consequently, direct studies in pregnant humans are difficult or impossible and one has to resort to surrogate in vitro or ex vivo studies or to animal models which necessarily are associated with difficulties in interpretation and extrapolation. Thus, what is described in this overall assessment is based largely on inferences from analogous diseases using tool chemicals able to reproduce the biological basis of the disease (especially etoposide (a Topoisomerase II poison)-caused acute leukaemia in children or adults) or from cellular and animal models.

1. Concordance of dose-response relationship

The only in utero study in mice (Nanya et al 2016) has shown that the dose of 0.5 mg/kg (day 13.5 of pregnancy) does not result in measurable etoposide concentration in foetal liver HSCs whereas the dose of 10 mg/kg leads to a maximal concentration of 5 µM. A statistically significant increase in double strand break (DSBs) in MLL gene was observed at a dose of 1 mg/kg, which would result in a concentration of 0.5 µM by linear extrapolation. In treatment-related acute human leukaemia, various treatment schedules in adults and children give rise to etoposide concentrations between (roughly) <1
µM (through) to >150 µM (peak). There are no adequate experimental systems to study dose-response and response-response relationships across MIE, KEs and AO in a single model.

2. Temporal concordance among the MIE, KEs and AO

There are no serious doubts about temporal concordance among MIE, KEs and AO. It is very difficult to see any other sequence of events (among this AOP), which would bring the AO into effect. Another matter is that it has never been shown in human pregnancy (or will be reliably or robustly demonstrated in the foreseeable future). In this respect, it is difficult to envisage whether epidemiological studies that are possible in humans, would ever be able to demonstrate the link without a direct biomarker for the MIE and KE1. Available experimental models (Sanjuan-Pla et al 2015) are in conformation with the AOP, except that in experimental in vivo models a very protracted appearance of leukaemia is not in line with a very short latency of infant leukaemia in human.

It is obvious that there exists a vast gap between wide exposure to potential TopII poisons and the rarity of infant leukaemia. On the basis of studies in human adult and childhood leukemias, there are a large number of genetic, epigenetic and host factors potentially modifying the link between topII poisons and leukaemia. Because of the rarity of the disease, it is difficult to envisage an even partial proofing these factors as of importance for the infant leukaemia.
Table 22. Response-Response and Temporality Concordance for the tool compound etoposide

<table>
<thead>
<tr>
<th>Concentration of etoposide</th>
<th>MIE In utero DNA topoisomerase II inhibition</th>
<th>KE1 In utero MLL chromosomal rearrangement</th>
<th>AO Infant leukaemia</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.01 – 0.1 µM, in vitro(TopII enzymes and cells in culture)</td>
<td>+++ (DNA damage response in various cells)</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>0.1 – 1 µM, in vitro cell cultures</td>
<td>+++</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>0.5-5 µM, ex vivo, mouse fetal liver HSC concentration</td>
<td>+++ (inference from MLL cleavage)</td>
<td>+ (only MLL cleavage)</td>
<td>- (no leukemia development)</td>
</tr>
<tr>
<td>max 5 µM, ex vivo, mouse fetal liver HSC concentration</td>
<td>+++ (inference from MLL cleavage)</td>
<td>+ MLL fusions detected only in DNA repair deficient mice</td>
<td>- (no leukemia development)</td>
</tr>
<tr>
<td>Max &gt;150 µM, plasma concs in etoposide-treated patients</td>
<td>+++ (inference from MLL cleavage)</td>
<td>++ MLL-AF4 fusion gene and protein</td>
<td>+ treatment-related acute leukaemia</td>
</tr>
</tbody>
</table>

*a range of concentrations is linearly extrapolated on the basis of the concentration of 5 µM after the dose of 10 mg/kg.
*bplasma concentration of etoposide cannot be directly extrapolated to the concentration at the active site. Probably the actual active cellular concentrations of etoposide is much lower, perhaps 10% or less of the plasma concentration.

3. Strength, consistency of the experimental evidence, and specificity of association of AO and MIE

Regarding the treatment-related acute leukaemia, strength, consistency and specificity of association of AO and MIE is strong, because only etoposide and a few other TopII-poison anticancer agents (Mention!) have strong evidence for causing acute leukaemia in human via the general process of the AOP described here. Although direct observations on the initial in utero MIE in infant leukaemia are not possible, there is a lot of inferential evidence from animal and in vitro cellular studies suggesting strongly that infant leukaemia recapitulates at least at an apparent process level the treatment-related leukaemia. It is important to recognize that in therapy-related AML this has been clearly demonstrated with abnormalities affecting MLL locus. Chlorpyrifos is reported to be a Topo II poison and to induce MLL translocation in the human liver haematopoietic stem cells (Lu et al. 2015). Considering the rarity of IFL and the common exposure to Topo II poisons like bioflavonoids, specificity is low. However, this consideration is limited by lack of experimental studies conducted with other than anticancer drugs on the sensitive target cells ie the liver haematopoietic stem cell.

4. Weight of Evidence (WoE)

4.1 Biological plausibility.

The biological plausibility for this AOP is strong. The relationship between DNA double strand breaks, MLL chromosomal translocation and infant leukaemia is well established. The same pathway is reproducible in chemotherapy-induced acute leukaemia in patients following treatment with etoposide, a known Topo II poison.
**Table 23: Biological plausibility of the KERs; WoE analysis**

<table>
<thead>
<tr>
<th>MIE → KE1</th>
<th>Defining Question</th>
<th>High (Strong)</th>
<th>Moderate</th>
<th>Low (Weak)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>In utero exposure to DNA topoisomerase II poison leads to In utero MLL chromosomal translocation</strong></td>
<td>Is there a mechanistic (i.e. structural or functional) relationship between KEup and KE down consistent with established biological knowledge?</td>
<td>Extensive understanding of the KER based on extensive previous documentation and broad acceptance</td>
<td>The KER is plausible based on analogy to accepted biological relationships, but scientific understanding is not completely established</td>
<td>There is empirical support for a statistical association between KEs but the structural or functional relationship between them is not understood</td>
</tr>
</tbody>
</table>

**Rationale:** Although type II topoisomerases are essential to cell proliferation and survival, they have a significant genotoxic potential consequent to the resulting (double) strand breaks. Mis-repair of accumulated DNA double strand breaks can result in chromosomal translocations which can persist in survived cells (Mc Clendon et al. 2009).

Studies on identical twins and neonatal blood samples strongly implicate an in utero occurrence of the KER (Sanjuan-Pla et al 2015). Furthermore, a study in pregnant mice demonstrates that in utero exposure of the foetus to etoposide causes the MLL chromosomal translocation analogous to the human translocation except the principal fusion partner (Nanya et al 2015). Indirect evidence from human prehaematopoietic/mesenchymal stem cells and foetal liver haematopoietic progenitor and stem cells strengthen the plausibility. Experimental evidence in these cell lines has demonstrated that etoposide as a TopII poison causes DSBs in MLL and partner genes, which leads to the formation of fusion genes and their products (SanjuanPla et al 2015).

MLL translocation sites (breakpoint sequences) in the therapy-related leukaemia fall within a few base pairs of etoposide-induced enzyme-mediated DNA cleavage site. Although rearrangements associated with infant leukaemias are often more complex than those observed in treatment-related leukaemias, many are nevertheless associated with stable TopII-mediated DNA cut sites (Cowell and Austin 2012; Pendleton et al 2014).

<table>
<thead>
<tr>
<th>KE1 → AO</th>
<th>Defining Question</th>
<th>High (Strong)</th>
<th>Moderate</th>
<th>Low (Weak)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>In utero MLL chromosomal translocation leads to Infant leukaemia</strong></td>
<td>Is there a mechanistic (i.e. structural or functional) relationship between KEup and KE down consistent with established biological knowledge?</td>
<td>Extensive understanding of the KER based on extensive previous documentation and broad acceptance</td>
<td>The KER is plausible based on analogy to accepted biological relationships, but scientific understanding is not completely established</td>
<td>There is empirical support for a statistical association between KEs but the structural or functional relationship between them is not understood</td>
</tr>
</tbody>
</table>

**Rationale:** The basic processes underlying overt leukaemia development are well understood and accepted. There is a general understanding of the molecular and epigenetic mechanisms leading to differentiation blockage and clonal expansion and there is evidence that the principal MLL-fusion genes and proteins harbour the necessary properties to execute the pathways associated with differentiation blockage and clonal expansion (Benito et al 2015; Chen and Armstrong 2015; Chen et al 2015).
4.2 Essentiality

In line with the defining question, essentiality for this AOP is moderate. However, the actual knowledge of the IFL is supporting the evidence that IFL is a “single hit” developmental disease and MLL translocation is an essential KE based on the probability linking MLL translocation and the occurrence of the disease. Based on this the overall essentiality can be considered moderate to strong.

Table 24: Essentiality of the KEs; WoE analysis

<table>
<thead>
<tr>
<th>2 Support for Essentiality of KEs</th>
<th>Defining Question</th>
<th>High (Strong)</th>
<th>Moderate</th>
<th>Low(Weak)</th>
</tr>
</thead>
<tbody>
<tr>
<td>MIE In utero exposure to DNA topoisomerase II poison</td>
<td>Are downstream KEs and/or the AO prevented if an upstream KE is blocked?</td>
<td>Direct evidence from specifically designed experimental studies illustrating essentiality for at least one of the important KEs (e.g. stop/reversibility studies, antagonism, knock out models, etc.)</td>
<td>Indirect evidence that sufficient modification of an expected modulating factor attenuates or augments a KE leading to increase in KE down or AO</td>
<td>No or contradictory experimental evidence of the essentiality of any of the KEs</td>
</tr>
<tr>
<td>KE1 In utero MLL chromosomal translocation</td>
<td></td>
<td>Growing scientific evidence, including the stable genome of the patients, suggests that infant leukaemia originates from one “big-hit” occurring during a critical developmental window of stem cell vulnerability (Andersson et al 2013; Sanjuan-Pla et al 2015; Greaves 2015). Therefore, the totality of evidence suggests the essential role of the formation of MLL-partner fusion gene and product in causing pleiotropic effects in the affected cell and directing it to the obligatory pathway to the adverse outcome of leukaemia. The MLL-AF4 fusion gene is present in bone marrow mesenchymal stem cells in infant leukaemia patients, but not in patients of childhood leukaemia, suggesting that the origin of the fusion gene is probably prehaematopoietic and essential for development of IFL (Menendez et al 2009). TopoII ‘poisons’ etoposide and bioflavonoids (and some other chemicals) promote MLL rearrangements in in vitro prenatal cells or in utero. There are in vitro cellular and in vivo xenograph studies demonstrating that upon inhibiting signalling pathways from the fusion product on, cells can resume differentiation or clonal expansion of fusion gene-carrying cells is prevented (Benito et al 2015; Buechele et al 2015; Chen and Armstrong 2015). However, in absence of a relevant in vivo experimental model these findings are suggestive but not yet totally convincing.</td>
<td>MODERATE.</td>
<td></td>
</tr>
</tbody>
</table>

4.3 Empirical support

The overall empirical support, using the chemical tool etoposide, is moderate. In vivo and, mainly in-vitro, experiments exist but they are lacking a clear dose or concentration response relationship.
### Table 25: Empirical support of the KERs; WoE analysis

<table>
<thead>
<tr>
<th></th>
<th>Defining Question</th>
<th>High (Strong)</th>
<th>Moderate</th>
<th>Low(Weak)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Does the empirical evidence support that a change in the KEup leads to an</td>
<td>Multiple studies showing dependent change in both</td>
<td>Demonstrated dependent change in both events following exposure to a small number of specific stressors</td>
<td>Limited or no studies reporting dependent change in both events following exposure to a specific</td>
</tr>
<tr>
<td>3</td>
<td>appropriate change in the KE down? Does KEup occur at lower doses and earlier</td>
<td>exposure to a wide range of specific stressors (extensive evidence for</td>
<td>and some evidence inconsistent with expected pattern that can be explained by factors such as</td>
<td>stressor (ie endpoints never measured in the same study or not at all); and/or significant</td>
</tr>
<tr>
<td></td>
<td>time points than KE down and is the incidence of KEup higher than that for KE</td>
<td>temporal, dose-response and incidence concordance) and no or few critical</td>
<td>experimental design, technical considerations, differences among laboratories, etc.</td>
<td>inconsistencies in empirical support across taxa and species that don’t align with expected</td>
</tr>
<tr>
<td></td>
<td>down? Are inconsistencies in empirical support cross taxa, species and stressors</td>
<td>data gaps or conflicting data.</td>
<td></td>
<td>pattern for hypothesized AOP</td>
</tr>
<tr>
<td></td>
<td>that don’t align with expected pattern of hypothesized AOP?</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

|   | MIE → KE1                                                                          | MODERATE                                                                    |                                                                                                         |                                                                                               |
|   | In utero exposure to DNA topoisomerase II poison leads to In utero MLL chromosomal | Rationale: Evidence comes from in vitro studies in appropriate human cells  |                                                                                                         |                                                                                               |
|   | translocation                                                                      | and from an in vivo/ex vivo study in pregnant mice; the stressor has been   |                                                                                                         |                                                                                               |
|   |                                                                                   | etoposide in most of the experiments (Libura et al 2005; Whitmarsh et al 2003; Lovett et al 201, Nanya et al 2015). Some evidence to back this KER comes from in vitro studies with bioflavonoids, especially quercetin, genistein and kaempferol (Barjesteh et al 2007). |                                                                                                         |                                                                                               |

|   | KE1 → KE2                                                                         | MODERATE                                                                    |                                                                                                         |                                                                                               |
|   | In utero MLL chromosomal translocation leads to Infant leukaemia                   | Rationale: There are a number of factors and pathways linking the fusion    |                                                                                                         |                                                                                               |
|   |                                                                                   | products with differentiation blockage and clonal expansion                 |                                                                                                         |                                                                                               |
|   |                                                                                   | (Marschalek 2010; Sanjuan-Pia et al 2015). **MLL** encodes a protein         |                                                                                                         |                                                                                               |
|   |                                                                                   | homologous to the Drosophila trithorax gene, which has relevant functions in |                                                                                                         |                                                                                               |
|   |                                                                                   | Studies with **MLL-AF4, MLL-AF9 and MLL-ENL**                               |                                                                                                         |                                                                                               |
|   |                                                                                   | (Barabe et al 2007; Mulloy et al 2008) have clearly demonstrated how MLL     |                                                                                                         |                                                                                               |
|   |                                                                                   | chromosomal rearrangements block differentiation and enhance clonal         |                                                                                                         |                                                                                               |
|   |                                                                                   | expansion. However, there is a specific need to execute these studies in    |                                                                                                         |                                                                                               |
|   |                                                                                   | an appropriate experimental system with a proper target cell within a proper  |                                                                                                         |                                                                                               |
|   |                                                                                   | molecular and physiological environment.                                      |                                                                                                         |                                                                                               |
|   |                                                                                   | There are several animal models, in which **MLL**-rearranged fusion genes   |                                                                                                         |                                                                                               |
|   |                                                                                   | have been expressed and leukemia developed (Chen et al 2006; Metzler et al 2006; Krivtsov et al 2008; Bursen et al 2008; Tamai et al 2011). Engineered human hematopoietic stem and progenitor cell carrying an MLL rearrangement showed that a subset of cells persisted over time and demonstrated a higher clonogenic potential in colony forming assay (Breese et al. 2015). Cells engineered to carry MLL-AF9 and MLL-ENL fusions demonstrated leukemogenicity especially after ex vivo and repeated transplantation (Buechele et al 2015). |                                                                                                         |                                                                                               |
5. Uncertainties and Inconsistencies

- In utero evidence of the disease is difficult to obtain in humans and one has to resort to in vitro cellular systems, which may be inadequate to take into consideration the potential effects of proposed microenvironments, rapidly changing developmental stages and facilitating and modifying factors.

- Animal models are a possibility (e.g. Nanya et al 2015), but are naturally prone to species-specific factors.

- An important problem is to provide a convincing and experimentally justified explanation for the dilemma between the rarity of disease in the face of pervasive exposure to topoII inhibitors.

- The treatment-related AML apparently is a true surrogate for the infant leukaemia, at least mechanistically. Is it only because of etoposide as a principal chemical initiator has provided many crucial findings for understanding the infant leukaemia.

- The ’poisoning’ of the TopoII-DNA cleavage complex has not been shown in the putative target cell, which is still not unequivocally identified.

- MLL-AF4 knock-in mice develop leukaemia only after a prolonged latency (e.g. Chen et al 2006), thus not recapitulating the ‘pathognomonic’ feature of infant leukaemia.

- The inability of available in vivo models to recapitulate the whole AOP process is due to a crucial factor which has not yet been found, or to model-specific peculiarities.

- In the face of the rarity of the disease, epidemiological studies especially concerning aetiology and risk factors are not powerful enough to provide robust answers. For instance, investigating the hypothesized relationship of bioflavonoids with infant leukaemia will have to consider the gap between the widespread intake of these phytochemicals and the very rare occurrence of the disease.

- The biology of the disease (i.e. IFL) and the experimental studies conducted with etoposide, indicate in-utero exposure of hematopoietic stem cells (HSC) as the most critical, if not essential, factor for the development of the A. However, a clear comparative quantification in terms of dose response vs different time of exposure and cell systems is lacking.

- The very early embryonic structure and the liver haematopoietic stem cells in particular, are representing the target cell for this AOP. A clear understanding of a higher sensitivity of HSC vs, mature hematopoietic cells, particularly in the standard genotoxicity test battery is lacking and more chemicals and comparative assays should be tested to scientifically validate this cell system.

- What would be consequences if we say that the AOP is biologically possible, feasible, even probable, and then say that most of the evidence is impossible to get directly and has to be based on surrogates?

6. Quantitative Considerations

The WOE analysis indicates that many KEs and KERs lack especially experimental evidence, but overall the analysis supports the qualitative AOP. The strong element in the development of the qualitative AOP is the biological plausibility of the overall pathway that it can partially be based on studies in human treatment-related disease recapitulating many crucial features of the infant leukaemia. The lack of sufficient experimental data and uncertainties in quantitative information from treatment-related acute leukaemia makes it problematic to build convincing dose (concentration)-response and response-response relationships and to identify possible practical thresholds for stressors. The MIE is expected to show a dose response relationship to a certain extent. However, it is probable that the dose dependence of the formation of DSBs and fusion genes is linear only in a very restricted “window”. In too-low concentrations the outcome of the stressor is a successful repair of the break, in too-high concentrations the outcome is cell death. It should be kept in mind additionally that the
quantification of dose–responses should also consider the different sensitivity of cell systems that
should be also representative of the specific time-window of exposure (i.e. in-utero).

The most pressing future need is an adequate and robust experimental model system for the
evaluation of relationships between doses, concentrations and responses within a temporal framework
of the AOP.

7. Applicability of the AOP

The proposed AOP is strictly life stage-dependent, being linked with in utero exposure and early
embryogenesis. However, the surrogate disease (i.e. chemotherapy-related acute leukaemia) is not
life stage restricted as well as the genotoxic hazard is not expected to be life stage related.

8. Potential regulatory applications of the AOP

This AOP was initiated with the intention to use an epidemiologically proposed human health outcome
as AO and build back an AOP leading to this. Infant childhood leukaemia is a human disease and
consequently apical regulatory endpoints can only explore the hazard by means of surrogate testing.
These include carcinogenesis assays and blood cell analyses in the in vivo toxicology assessment.
Considering the unique biology of this AO, these tests are showing some technical limitations and also
the sensitivity and specificity of the available tests for the AO is lacking. Additionally, experimental
animal models replicating the AO are limited. Technical limitations of the standard regulatory tests
include: Standard carcinogenesis studies do not include an early in-utero exposure time, blood cell
analysis is not a standard requirement in the extended multi-generation reproductive toxicity study
and no cancer-related endpoints are included in this study. In addition, considering the rarity and the
complexity of the disease, the sensitivity and specificity of these tests to capture this hazard is likely to
represent a big hurdle and the regulatory tests are unlikely to represent the best way to explore this
AO.

This AOP is however indicating that the MIE and the KE1 can be measured in scientific and/or
regulatory validated test assays.

With these premises, the authors support the use of this AOP during the process of assessment of
epidemiological studies and the use of the AOP framework to support the biological plausibility of the
effects observed in the epidemiological studies when experimental and toxicological studies are
indicative that the AOP is affected and this should guide on which additional studies should be
performed, if the case, to integrate the AOP framework into the MOA framework for specific chemical
entities.

In addition, this AOP should serve in guiding testing strategy. This include the exploration of Topo II
poison characteristics of a chemical and, if the genotoxicity standard regulatory testing battery is
negative, considerations should be made on the sensitivity of the cell system used in the assay
(i.e. liver HSPC).

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**AOP4: In utero induction of chromosomal rearrangements/translocations in haematopoietic stem/progenitor cells (HSPCs) followed by postnatal mutations and an aberrant immune response leads to childhood leukaemia**

**Introduction**

Leukaemia is the most common cancer in children under 15 years of age, with an annual incidence of up to 40 cases per million children in developed countries and an incidence peak between 3 and 5 years of age (Hunger and Mullighan, 2015; ENHIS, 2009). Childhood leukaemia (also termed paediatric leukaemia) is a biologically heterogeneous disease of immature haematopoietic progenitors that consists of multiple subtypes depending on the cell type and lineage involved (lymphoid or myeloid progenitors). Seventy percent of cases are comprised by acute lymphoblastic leukaemia (ALL) and the remaining 30% by acute myeloid leukaemia (AML). ALL may be of B-cell lineage (85%) or T-cell lineage (15%). However, there are some cases of biophenotypic acute leukaemias commonly harbouring Mixed Leukaemia Lineage (MLL) rearrangements, in which myeloid and lymphoid markers have been shared by the blast population (Hunger and Mullighan, 2015).

Childhood leukaemia should be distinguished from infant leukaemia, a more rare disease that manifests soon after birth (<1 year of life) and has a poorer prognosis. Infant leukaemia is considered as a ‘developmental disease’ showing different features and pathogenesis than childhood leukaemia, with more immature precursors being involved (Sanjuan-Pla et al., 2015). A remarkable difference between the two entities is that childhood leukaemia may arise as a consequence of a “2-hit” model producing two independent (epi)-genetic insults, the first one occurring in utero and the second one either before, or more often, after birth. In contrast, the natural history and genome-wide sequencing studies on infant leukaemia suggest that only a single hit occurring in utero is needed. A common initiating pathogenic event for both types of leukaemias is the occurrence of chromosomal rearrangements (i.e., chromosomal translocations) that create fusion genes encoding transcriptional factors involved in the regulation of early haematopoiesis. Chimeric fusion proteins encoded by chromosomal translocations lead to differentiation arrest of HSPCs, which represents a hallmark in childhood leukaemia. Almost half of the B-cell ALL cases exhibit aneuploidy, either hyperdiploidy or hypodiploidy with non-random chromosomal gain or loss, respectively, affecting different chromosomes. Hyperdiploidy causes chromosomal instability as a result of chromosomal translocations, duplications and deletions (Paulsson et al., 2006).

The genetic basis of ALL consists of recurrent genetic alterations, such as loss-of-function mutations involving genes regulating lymphoid development that contribute to the maturation arrest characteristic of B-ALL, mutations that inactivate tumour suppressor and cell cycle regulatory proteins, and mutations in genes encoding cytokine receptor and/or protein kinases regulating cell signalling pathways (Mullighan, 2012). For T-cell ALL, the main drivers are chromosomal translocations resulting from aberrant recombination between T-cell receptor genes and oncogenes (Mullighan, 2012), together with activating Notch mutations, a protein involved in T-cell differentiation and thymocyte development (Weng et al., 2004). For AML leukaemias to occur, cooperation is required between gene rearrangements involving haematopoietic transcription factors (i.e., AML1/ETO, MLL-related fusion genes, etc.) and activating mutations (i.e., in apical regulators of intracellular signalling cascades) (Mullighan, 2012).

Despite the rather comprehensive epidemiologic evidence linking pesticide exposures during different reproductive stages (pre-conception, pregnancy and early postnatal) with childhood leukaemia, no robust mechanisms supporting these associations have been reported so far. Pesticide exposure has not been directly linked to the development of childhood leukaemia in animal models. Although negative results for genotoxicity tests have been observed in regulatory studies on individual pesticides, there is limited experimental evidence in the open literature about the genotoxic or cancer-promoting capacities of some pesticides in cells, suggesting a potential leukaemogenic effect. However, the target cells used in these experiments are not the most appropriate for this purpose and the role played by some pesticide metabolites cannot be ruled out.

Regardless of the extensive gap in our understanding, particularly on how pesticides mechanistically interact with biological targets to trigger childhood leukaemia, the AOP proposed for this disease is supported by experimental evidence and cellular models, with the exception of the molecular initiating
event. However, as knowledge increases, the present AOP may be modified on the basis of novel supporting evidence.

**AOP: In utero induction of chromosomal translocations in haematopoietic stem/progenitor cells (HSPCs) leads to childhood leukaemia**

**Fig.36** AOP scheme

The MIE is a specialized type of KE representing the starting point of chemical interaction with a biological target leading to disruption at the molecular (including genetic) level and subsequent disease progression. Expectation is that perturbation of the MIE, if quantitatively enough, will lead to the AO. In the case of childhood leukaemia, early in utero interaction of a chemical with DNA (or DNA-related proteins/enzymes) might lead to double strand DNA breaks, which if non-repaired or mis-repaired, may result in genomic instability, leukaemic transformation or cell death.

HSPCs exposed to ionizing radiation, environmental chemicals or chemotherapeutic agents are prone to DNA breakage at sites with the potential to form leukaemia-causing gene rearrangements. Exposure to non-cytotoxic levels of environmental chemicals and chemotherapeutic agents can induce DNA damage in HSPCs without causing cell death (Thys et al., 2015). Several studies investigating the role of DNA repair systems in response to DNA damage found that human foetal liver CD34+ HSPCs are more sensitive to DNA damage than other haematopoietic precursors at different ontogeny stages (Bracker et al., 2006). Human fetal liver CD34+ HSPCs are more sensitive to oxidative stress induced by certain environmental chemicals, including many classes of pesticides than neonatal or adult CD34+ cells (Bueno et al 2009). Among environmental chemicals, the organophosphate (OP) insecticide chlorpyrifos has been reported to cause DNA double-strand breaks (DSB) and further chromosome rearrangements in human foetal liver HSPCs in part through oxidative stress (Gupta et al., 2010).

Chemical exposure may result, either directly or indirectly, in DNA damage. Three potential mechanisms are involved in this process: generation of DNA DSB, improper repair of these DNA lesions and erroneous V(D)J recombination (Hernández and Menéndez, 2016).

a) **DNA double strand break.** Exposures to ionizing radiation and numerous chemicals are capable of inducing oxidative DNA damage through the generation of reactive oxygen or nitrogen species (ROS and RNS, respectively). These highly reactive species may produce DNA base or sugar damage leading to single-strand break formation. However, under some circumstances DNA DSBs can arise, for instance: a) when two single strand breaks form close to each other on opposite strands, b) when topoisomerases cleave next to a single strand breaks on the opposite strand, and c) when either DNA replication or transcription takes place at unrepaired DNA damage.

b) **Improper DNA repair.** ROS-induced DNA DSBs in human foetal liver-derived HSPCs following maternal exposure to chemicals triggers recombination/repair pathways by non-homologous end-
joining (NHEJ), the main repair pathway for DSBs. The majority of damaged HSPCs may either successfully repair the break or fail and die through secondary activation of apoptotic pathways. In a fraction of cells, the attempt to repair the DNA DSBs within particular breakpoints cluster regions is not completed properly, so that chromosomal translocations or deletions may occur (Wiemels and Greaves, 1999). Translocation breakpoints harbour evidence of NHEJ mechanisms, but in only a few examples are the causative mechanisms of breakage evident, such as V(D)J recombinase gene activation (Wiemels, 2008).

c) Erroneous V(D)J recombination. V(D)J recombination is a process occurring in developing lymphocytes during cell maturation, where gene segments of immunoglobulin chains or T-cell receptor, known as variable (V), diversity (D) or joining (J), are rearranged to yield a wide range of immunoglobulins and T-cell receptors. The process entails the cleavage of the V(D)J gene at the flanking recombination signal sequences (RSS) by lymphocyte-specific recombination-activating gene (RAG) endonucleases and subsequent ligation of the segments via the classical NHEJ pathway (Meissner et al., 2014). In the case of childhood leukaemia, chromosomal translocations as well as gene deletions often arise as result of mistakes in V(D)J recombination, e.g. RAG can erroneously recognise and target RSS-like sequences. There is growing evidence that in vivo exposure to DNA-damaging agents can increase the frequency of V(D)J rearrangements at RSS-like sequences that are widely distributed throughout the genome (ref). However, the mechanism by which exposure to those agents increase the frequency of V(DJ)-recombinase-mediated genomic rearrangements is still unknown (Pinsoneault et al., 2007). The lack of site-specific clustering of translocations (which show a dispersed breakpoint distribution) suggests that chromosomal translocation arise in HSPCs before the expression of recombinase-activating genes (Wiemels, 2008).

Although all the above reported mechanisms can be chemically induced, a chemical tool able to initiate the triggering cascade of the proposed pathway was not identified. For this reason this AOP was considered putative and the KE 1 was used as initiator of the pathway.

However, the potential initiating events speculated above can be measured. For this reasons, technologies able to do it are reported here. Oxidative stress can be measured by a number of biomarkers such as plasma antioxidant status, lipid peroxidation products, reduced-to-oxidized glutathione (GSH:GSSG) ratio, and levels of 4-hydroxynonenal (4-HNE) and nitrotyrosine products. However, these biomarkers may only provide an indirect assessment of an increased risk of oxidative DNA damage in vivo (Badham and Winn, 2010). There is a variety of techniques and methods useful for the detection of single oxidatively generated DNA lesions like 8-oxodG, thymine glycol (Tg) and abasic (AP) sites such as high performance liquid chromatography (HPLC), liquid chromatography/tandem mass spectrometry (LC-MS/MS), alkaline filter elution, single cell gel electrophoresis (SCGE or Comet assay) and adaptations of agarose gel electrophoresis (Kryston et al., 2011).

DNA damage can be assessed by the single cell gel electrophoresis (SCGE or Comet assay), which is a simple sensitive and rapid method for the detection and quantification of DNA damage (Singh et al., 1988) and provides a direct microscopic measure of DNA single and double strand breaks.

DNA breakpoints can be determined by Southern blot, polymerase chain reaction (PCR) and DNA sequencing. Gene mutations can be comprehensively searched for by array-comparative genome hybridisation (array-CGH) or whole-genome/exome sequencing. Allele-specific restriction assay, single strand conformation polymorphism and/or direct sequencing are valid methodologies for point mutation analysis.

Increased DNA damage in leukaemia cells can be demonstrated by the formation of phosphorylated histone H2AX (γ-H2AX), a marker of DNA DSBs (Graillot et al., 2012).
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KE1: \textit{In utero} chromosomal translocations.

\textbf{How this key event works}

Early \textit{in utero} interaction of a chemical with DNA (or DNA-related proteins/enzymes) might lead to permanent DNA damage and further chromosomal translocations. Other chromosomal insults can occur as well, such as intrachromosomal rearrangements, genetic deletions or activating mutations. Altogether, these chromosomal lesions are considered initiating events in leukaemogenesis, and most likely occur prenatally as common leukaemia fusion genes have been detected in cord blood, neonatal Guthrie cards and shared by monozygotic twins (Greaves et al., 2003).

There are two functional classes of translocations; the first one relocates a proto-oncogene (or genes encoding for transcription factors or non-antigen receptors) into regulatory regions of actively transcribed genes (such as those encoding for immunoglobulin chains or T-cell receptors), causing dysregulated expression of an intact protein. The second class juxtaposes two genes to encode a chimeric protein that has distinct functions from the proteins from which it is derived (Hunger and Mulligham, 2015).

\textit{TEL-AML1} is the most common chromosomal translocation associated with B-ALL, which affects haematopoietic stem/progenitor cells (HSPCs). \textit{TEL-AML1} is a fusion gene involving \textit{AML1} (also known as \textit{RUNX1}), which controls the emergence of definitive haematopoietic stem cells in foetal haemogenic sites, and \textit{TEL (ETV6)}, responsible for adult haematopoietic stem cells survival (Teitell and Pandolfi, 2009).

It is not known to what extent chromosomal translocations/rearrangements are caused by errors in normal DNA processing or by external factors (chemicals, viruses), but translocations are 100-fold more common in the population than leukaemia, indicating that most translocations are not sufficient for disease (Wiemels, 2008).

The finding that most common translocations found in childhood leukaemia (TEL-AML1 and AML1-ETO) occur at a rate of 1\% in the normal population suggests that a significant proportion of the population carries preleukaemic clones. However, most of these clones are self-limiting and do not result in disease. During the progression of the disease multiple genetic alterations accumulate over time being selected by their potential to give fitness advantage to the new clones.

\textbf{How it is measured}

Conventional cytogenetics, fluorescence in situ hybridization (FISH, using commercially available dual colour translocation probes) and reverse transcription polymerase chain reaction (RT-PCR) methods allow the identification of specific chromosome abnormalities (fusion genes, translocations, etc.), which can be further identified by subsequent cloning and sequencing (Soszynska et al., 2008). Cytokinesis-block micronuclei assay also allows to assess chromosome damage.

Gene expression profiling defines distinct oncogenic groups in ALL related to the presence of different fusion oncogenes.

\textbf{Taxonomic applicability}

Chromosomal translocations can occur at all levels of living organisms and they have been created in murine and zebrafish models. These models can be useful for the \textit{in vivo} study of leukaemogenic potential of chemicals in immature organisms as they may recapitulate human childhood leukaemia.

Bone marrow and foetal liver cells from mice have been retrovirally transduced to express \textit{TEL-AML1} protein in an attempt to model human ALL.
Regulatory examples using this KE

The extended one generation test (OECD 443) includes a developmental immunotoxicity cohort. At present the cohort may identify post-natal effects resulting from prenatal or neonatal exposures on the immune tissues and white blood cells population. However, no specific parameters are in place to identify a pattern relevant to human childhood leukaemia in the extended one generation test. Besides, no treatment is administered in utero during the early developmental phase in the carcinogenicity assay and no considerations on the possible higher sensitivity of the HSPCs are in place for the genotoxicity assays. Thus, regulatory studies following OECD test guidelines may have potential limitations and experimental gaps eventually leading to false negative results.

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**KE2: Differentiation arrest of HSPCs.**

**How this key event works**

Chromosomal translocations create fusion genes encoding active kinases and altered transcription factors as well as hyperdiploidy. The genetic changes alter key regulatory processes by maintaining or enhancing an unlimited capacity for self-renewal, subverting the controls of normal proliferation, blocking differentiation and promoting resistance to death signals (i.e. apoptosis).

Altered self-renewal and differentiation of HSPCs can result from chimeric transcription factors, which arise from chromosome translocations that fuse portions of two different transcription factors. The aberrant proteins produced by fusion genes inhibit the normal transcriptional program and block the differentiation of B-cell and myeloid precursors by recruiting repressor molecules such as histone deacetylase enzymes, resulting in aberrant cell proliferation and survival (Greaves, 2002; Pui et al., 2004; Papaemmanuil et al., 2014; Teitell and Pandolfi, 2009). For instance, TEL-AML1 and MLL fusions in undifferentiated progenitor cells can block the differentiation phase between pro-B to pre-B cells.

Most of paediatric B-ALL with BCR-ABL fusion genes exhibits IKZF1 deletions. The gene IKZF1 encodes a transcription factor that belongs to the family of zinc-finger DNA-binding proteins associated with chromatin remodelling. The expression of this protein is restricted to the foetal and adult hemo-lymphopoietic system, and it functions as a key regulator of lymphocyte differentiation. Mice with reduced Ikaros expression exhibited partial inhibition in precursor B-cell maturation, which might be relevant in leukaemogenesis (Teitell and Pandolfi, 2009).

**How it is measured**

Arrest of B-cell differentiation can be observed by histological assessment (Sabaawy et al., 2006). Methods for detecting suppression of haematopoiesis include the assessment of cell-specific markers via immunolabelling followed by flow cytometry and/or microscopy (e.g. CD34, CD19 and IgM for stages of B-cell differentiation).

Methods for detecting epigenetic modifications include:

- DNA methylation: Combined bisulphite restriction analysis (COBRA) and bisulphite sequencing for methylation; methylation-specific PCR.
- miRNA/non-coding RNAs: miRNA/non-coding RNA isolation followed by amplification using reverse transcription-PCR; miRNA/non-coding RNA microarray profiling/analysis

**Taxonomic applicability**

Mice have been transplanted with TEL-AML1-transduced bone marrow stem cells (Tsuzuki et al., 2004). There are also zebrafish models of TEL-AML1-positive ALL (Sabaawy et al., 2006).
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KE3: Clonal expansion as a result of secondary oncogenic insults (activating mutations) and delayed infections.

How this key event works

ALL is mainly a disease of childhood that arises from recurrent genetic insults that block precursor B and T cell differentiation and drive aberrant cell proliferation and survival. A pre-leukaemic clone with self-renewal stem cell activity may acquire progressive mutations in proliferative genes (activated signalling) resulting in a frank leukaemic clone. Cancer genome re-sequencing studies have determined that most leukaemia cases harbour multiple mutations that have sequentially occurred in a single cell lineage to generate a dominant leukaemic clone (Jan and Majeti, 2013).

- (Epi)genetic modifications:

Epigenetic modifications, in particular DNA methylation leading to reduced expressions of tumour suppressor genes contribute to the pathogenesis of childhood leukaemia. The inactivation or silencing of tumour suppressor genes can result in the sustained proliferation or reduced cell death response (e.g. apoptosis) of leukaemic cells. In childhood ALL, extensive hypermethylation of tumour suppressor genes such as FHIT, DLX3, p16 and p15 resulting in gene silencing has been observed. Furthermore, epigenetic silencing of proapoptotic genes (e.g. BIM), or blockade of apoptotic activation via deregulated expression of anti-apoptotic genes (e.g., Bcl2 and BAX), inhibits activation of apoptosis and enhances survival of leukaemic cells, enabling the progression of leukaemogenesis (Bachmann et al., 2010; Sabaawy et al., 2006). Exposure to a variety of environmental agents can alter DNA methylation pattern inducing destabilizing changes in gene expression potentially eading to cell transformation and tumorigenesis. There is some evidence suggesting that epigenetic modifications may be one of the mechanisms by which pesticides may exert adverse effects on human health (Collota et al., 2013).

The inhibition or aberrant regulation of apoptosis due to gene/protein dysfunction also plays a role in the pathogenesis of childhood leukaemia. Increased expression of Ikaros isoform 6 in murine myeloid precursor cell line appears to up-regulate the expression of the anti-apoptotic protein Bcl-XL, preventing apoptosis and potentially leading to the pathogenesis of AML (Yagi et al., 2002).

- Delayed infections

The delayed-infection hypothesis of Greaves is based on a minimal two-hit model and suggests that some susceptible children with a prenatally acquired preleukaemic clone had limited exposure to common infections early in life because they lived in a very hygienic environment. Such infectious insolation results in an immune system improperly developed that further predisposes these children to develop exacerbated aberrant responses after subsequent or delayed exposure to common infections later on in life, at an age commensurate with increased lymphoid-cell proliferation (Gilham et al., 2005; Kamper-Jørgensen et al., 2008; Pui et al., 2008). This untimely and excessive inflammatory response abolishes normal haematopoiesis such that lymphocytes or myeloid progenitor cells cannot mature. Thus, the innate and adaptive immune system is not fully functional upon an immune response and promotes selective expansion of a preleukaemic clone because of proliferative advantage and an increased opportunity for the acquisition of secondary genetic changes or mutations ultimately resulting in overt leukaemic phenotype (Ford et al., 2009; Greaves, 2006; Swaminathan et al., 2015).

- Potential targets of chemical exposures

The immune system may be a target of the toxic effect of several chemicals. Chemically-induced immune alteration through altering well-regulated immune responses to tumour antigens, allergens, self-antigens and microbial antigens can contribute to predisposition to different types of disorders, including cancers (Mokarizadeh et al., 2015). Evidence suggests that children may be particularly susceptible to adverse effects from exposure to pesticides, thus rendering them susceptible to infections and other immune mediated disorders (Corsini et al., 2013). Some evidence of effects of environmental exposures to pesticides during prenatal and early postnatal development on childhood leukaemia has been reported, raising the importance of studying the effects of toxicants on the developing immune system (Duramad et al., 2007). Xenobiotics may initiate, facilitate or exacerbate aberrant immune processes by inducing mutations in genes coding for immunoregulatory factors,
modifying immune tolerance and activation pathways. Besides, various general or immune specific signalling pathways can be interfered by chemicals, resulting in changes in cytokine production, surface markers expression, cell differentiation and activation (Corsini et al., 2013).

Immunosuppression induced by pesticides may explain the relation with increased infections in humans observed in several studies. Particularly susceptible to immunotoxicity are children, as the vulnerable period for toxic insults to the developing immune system extends from early gestation to adolescence (Dietert, 2008). Background exposure to some pesticides early in life (pre- and postnatal exposure) may modulate the immune system development, increasing infection risks (Weselak et al., 2007). Furthermore, pesticides may interfere with immune surveillance, which in turn can affect recognition and destruction of abnormal cells, increasing the risk of cancer (Corsini et al., 2013).

**How it is measured**

Methods of detecting leukaemic cell proliferation include flow cytometry using cell-specific markers followed by quantitative analysis, and incorporation and detection of bromodeoxyuridine (BrdU) by proliferating cells.

Multicolor fluorescence in situ hybridization (FISH) may be used to track multiple genetic abnormalities identified in bulk ALL cells, yielding quantitative single cell resolution of the relative frequency of genetically distinct leukaemia subclones.

A novel experimental and computational single-cell sequencing approach has been used to directly measure the clonal structures of childhood ALL samples at diagnosis (Gawad et al., 2011).

Apoptosis can be measured by using plasma membrane assays, mitochondrial assays, caspase assays, nuclear apoptosis assays and flow cytometry.

**Taxonomic applicability**

Mechanisms relevant to clonal expansion may not show significant interspecies differences and potential mechanisms remain currently unclear.

Murine models with human precursor cells harbouring the TEL-AML1 fusion have been developed.

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Adverse Outcome (AO): Overt childhood leukaemia

How this key event works

Symptoms of childhood leukaemia include sensitivity to bruising and bleeding due to thrombocytopenia, pallor and fatigue from anaemia, and increased susceptibility to infections caused by neutropenia. These symptoms result from the displacement of the normal haematopoiesis by expansion of leukaemia cells. Leukaemic infiltration of the liver, spleen, lymph nodes, and mediastinum is common at diagnosis (Hunger and Mulligham, 2015).

How it is measured

Haematological methods: observations of clinical symptoms, routine blood cell count and identification of leukaemia cells (i.e., immunophenotyping by flow cytometry) in peripheral blood and bone marrow. Diagnosis stratification relies on molecular cytogenetics (FISH and karyotype).

Immunophenotyping allows the identification of pathologic cells and phenotype characterization based on specific pattern-identification of surface as well as intracellular antigen expressions in unique cell populations.

Flow cytometry is a laser-based technology that uses monoclonal antibodies for the detection of expression of a number of antigens on the cell surface, thus distinguishing between healthy and diseased cells. Flow cytometry allows the identification and quantification of subsets of the major leukocyte populations and even further sub-divisions that differ in biologic function, maturation stage, and activation (Adin-Cinar, 2013).

Taxonomic applicability

The following animal models have been developed for childhood leukaemia:

a) MLL-ENL and MLL-AF9 fusions have been proven to be oncogenic by themselves in human cord blood progenitor cells (Barebé et al., 2007).

b) TEL-AML1 and hyperdiploid primary blasts recapitulate the disease phenotype in immunodeficient mice (Rehe et al., 2014, le Viseur et al., 2008).

Regulatory relevance of the AO

Genotoxicity and cancerogenicity are standard endpoints measured in the regulatory studies performed for the risk assessment of chemicals and they are mandatory for pesticide substances. However, no treatment is occurring during the early in-utero development phase in the carcinogenicity study.

The extended one generation test (OECD 443) assesses parental fertility and reproductive function and the development of offspring to sexual maturity and also includes a developmental immunotoxicity cohort. A second generation can be triggered if any effects requiring further evaluation are identified in the first generation (OECD, 2011). The study design provides the opportunity to evaluate life stages not covered by other study types and represents a highly integrated study design that includes an assessment of developmental immunotoxicity. While the developmental immunotoxicity cohort may identify post-natal effects resulting from prenatal or neonatal exposures on the immune tissues and white blood cells population; however, no specific parameters are in place to identify a pattern relevant to human childhood leukaemia.
References


1st KER: In utero chemical exposure (KE up) leading to unrepaired/misrepaired double DNA damage and further chromosomal translocations (KE down)

How this Key Event Relationship works

DNA is highly susceptible to oxidative damage, which can result in single strand breaks (SSBs) and DSBs, base and sugar-moiety oxidation, strand crosslinks and the generation of abasic sites. DSBs are the most serious type of DNA damage because a small number of these lesions are sufficient to induce gene mutations or chromosomal aberrations (Sedelnikova et al. 2001; Woodbine et al., 2011). Oxidative molecules may either enhance the likelihood of DSB in HSPCs or interact with biological molecules disrupting the normal synthesis and repair of DNA. This disruption is primarily associated with inhibition or inactivation of antioxidant proteins as well as DNA repair enzymes (Kryston et al., 2011).

Upon DNA damage or genotoxic stress, hematopoietic stem cells differentiate to lineage-committed progenitors, which can be considered as a method to escape propagating damaged genetic information. This escape mechanism fails when hematopoietic stem cells chose DNA repair by NHEJ over differentiation, in order to maintain their self-renewal, thus thriving haematological malignancies (Weiss and Ito, 2015).

Defects in NHEJ can create chromosomal deletions and translocations. The accumulation of genetic damage through misrepair or incomplete repair of DNA may lead to mutagenesis and eventually cell transformation, particularly if combined with a deficient apoptotic pathway (Kryston et al., 2011). An impaired repair of oxidatively modified DNA, documented in children with ALL, may contribute to the genetic instability of precursor-B cells which may be linked with the development of the disease (Olsinski et al., 2014).

For fusion genes to be effective in promoting leukaemogenesis, DNA DSB must occur simultaneously in two chromosomes in a single HSPC that does not undergo cell death, and must also be situated in the coding region of the genes to generate a functional chimeric gene product. The resulting chromosomal recombination must take place in a HSPC with a sustainable lifespan and clonal potential to propagate the chimeric gene product (Greaves and Wiemels, 2003).

A massive parallel sequencing approach performed in a cohort of twins concordant for ALL indicated that the TEL-AML1 fusion gene arises as a consequence of NHEJ as no binding motifs indicative of RAG1/2 or terminal deoxynucleotidyl transferase (TdT) activity were found. The TEL-AML1 fusion arises in a foetal HSPCs that lies upstream of B-cell lineage-restricted RAG1/2 active precursors. The pre-leukaemic clone arises and expands in the pro- or pre-B-lineage compartment in the foetal liver and then undergoes V(D)J rearrangements (Alpar et al., 2015).

Biological plausibility

In the last decades the occurrence of childhood leukaemia showed a rise that was in part attributed to an increased exposure to risk factors. Although the aetiology of ALL remains elusive, ionizing radiation, congenital genetic syndromes and in utero exposure to specific genotoxic chemicals, including household pesticides, are considered the major risk factors (Pui et al., 2008). Despite the mounting epidemiologic evidence linking pesticide exposure during pre- and postnatal life with childhood leukaemia, robust underlying pathological mechanisms remain unknown. The initiating event at the molecular level might be generation of oxidative stress by environmental exposures (including pesticides) leading directly or indirectly to DNA damage and further chromosomal damage (Hernández and Menéndez, 2016); however this still remains hypothetical.

A massive parallel sequencing approach performed in a cohort of twins concordant for ALL indicated that the TEL-AML1 fusion gene arises as a consequence of NHEJ as no binding motifs indicative of RAG1/2 or terminal deoxynucleotidyl transferase (TdT) activity were found. The TEL-AML1 fusion arises in a foetal HSPCs that lies upstream of B-cell lineage-restricted RAG1/2 active precursors. The pre-leukaemic clone arises and expands in the pro- or pre-B-lineage compartment in the foetal liver and then undergoes V(D)J rearrangements (Alpar et al., 2015).
Investigation of the DNA damage in steady state, as well as after exposure to UV light, confirmed increased DNA damage in pro-B cells lacking a functional allele of Ebf1 (a transcription factor critical for the activation of B-lineage restricted genes in the earliest B-lineage progenitors that also controls DNA repair). Reduced Ebf1 levels may contribute to malignant transformation by a combination of impaired DNA repair and increased cell survival rather than simply by a differentiation block (Prassad et al., 2015). Since Rad 51 is one of the central components of the DNA DSB repair gene, whose expression can be induced by DNA damage, a drop in leukaemic potential after Rad51 re-expression would conclusively demonstrate that loss in HR DNA repair was the main driving force of leukaemic transformation of the Ebf1\(^{+/−}\) Pax5\(^{+/−}\) B-cell precursors (Georgopoulos, 2015).

**Uncertainties and Inconsistencies**

Despite the sound epidemiological evidence linking pesticide exposure and childhood leukaemia, the first initiating molecular event(s) has not been unravelled yet. In contrast to MLL-rearranged infant leukaemia, there is no evidence at all regarding the molecular basis of how some individual pesticide or pesticide class (or functional group) can interact with biological targets to elicit DNA damage. We can speculate only with potential mechanisms, such as induction of oxidative stress in HSPCs, as DNA is highly susceptible to oxidative damage and can result in single and double strand breaks. Besides, it not clearly understood what drives damaged HSPCs to initiate DNA repair systems and when to enter the cell cycle or to keep quiescent accumulating genotoxic stress (Weiss and Ito, 2015). While, regulatory studies have consistently found lack of genotoxic effects of pesticides in many test systems, there are studies in the open literature supporting genotoxicity by using different biomarkers. In addition, some epidemiological studies on agricultural workers exposed to pesticides have reported DNA damage. These uncertainties and inconsistencies warrant further research to delineate how pesticides interact with DNA and produce genetic lesions.

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2nd KER: Title: In utero chromosomal translocations (KE up) leading to differentiation arrest of HSPCs (KE down)

How this Key Event Relationship works
There are many potential chromosomal translocations associated to childhood leukaemia, suggesting that multiple mechanisms underlie the development of the disease. Nevertheless, the major fusion genes generated by chromosome translocations are TEL/AML1 in ALL and AML1/ETO in AML. The chimeric (aberrant) proteins produced by these genes inhibit gene activity and block cell differentiation by recruiting repressor molecules such as histone deacetylase enzymes (Greaves, 2002).

TEL-AML1 and MLL fusions in undifferentiated progenitor cells can block the differentiation phase from pro-B to pre-B cells. By stalling B-cell development, subsequent recombination-activating gene (RAG)-mediated genomic rearrangements become drivers of the creation of polyclonal structures (Papaemmanuil et al., 2014). A study using whole-genome sequencing in ALL has suggested that the aberrant activity of RAG recombinases, which are highly expressed in cells harbouring TEL-AML1, can result in various oligoclonal V(D)J recombination events and further inactivation of genes required for B-lineage differentiation (Papaemmanuil et al., 2014).

Weight of evidence
The block of differentiation of HSPCs provides proliferative advantage because of conferring self-renewal properties to lymphoid progenitors. Enhanced self-renewal would promote the extended longevity of B-cell precursors to acquire and accumulate additional genomic aberrations and secondary mutations, which collaborate to fully transform these B cell precursors into leukaemia cells (Duque-Afonso et al., 2015).

The pre-leukaemic transformation conferring the in utero clonal expansion of TEL-AML1 cells occurs in an early B-cell lineage committed progenitor, most likely at the pro-B or pre-B-cell stage in the foetal liver (Alpar et al., 2015).

Biological plausibility
Under the current paradigm, the first initiating oncogenic mutation usually involves structural or numerical chromosomal alterations impairing normal cell differentiation, while secondary hits more commonly comprise mutations affecting developmentally regulated master transcription factors or membrane-proximal signalling pathways conferring proliferation and survival advantages to the differentiation-blocked clone. The development of leukaemia requires activation of cell proliferation in addition to differentiation blockade (reviewed in Hernández and Menéndez, 2016).

As a result of chromosomal translocations, aberrant chimeric proteins alter the normal transcriptional program and block normal B-cell and/or myeloid differentiation. Childhood leukemia arises from recurrent genetic insults that block differentiation of hematopoietic stem and/or progenitor cells (HSPCs), and drives uncontrolled proliferation and survival of the differentiation-blocked clone.

Epigenetic modifications to DNA affect the activity of genes and their cellular expression and include DNA methylation, histone modification, and alterations in non-coding microRNAs (miRNAs). Each of these mechanisms alters how genes are expressed or silenced without modifying the DNA sequence. Epigenetic control of transcriptional activation also plays an essential role in regulating gene expression during early development and haematopoiesis. Besides, epigenetic modifications can influence leukaemogenesis if they lead to silencing of tumor suppressor genes or activation of oncogenes (Burke and Bhatia, 2014).

Non-coding RNAs have been implicated in the pathogenesis of childhood leukaemia as their altered expression can regulate various physiological processes such as cell differentiation, proliferation and immune responses. Expression of miRNAs is triggered by epigenetic modifications, e.g. hyper/hypomethylation of CpG islands in the promoter region of genes, or by fusion proteins.

In a zebrafish model of TEL-AML1+ B-ALL, arrest of B-cell differentiation has been observed by histological assessment (Sabaawy et al., 2006). An accumulation of early pro-B cells and a
differentiation deficit after pro-B cell formation has been reported in mice transplanted with TEL-AML1-transduced bone marrow stem cells (Tsuzuki et al., 2004).

De Laurentiis et al. (2015) generated an experimental model using the murine hematopoietic stem progenitor cell line EML1 expressing the TEL-AML1 fusion protein, and analyzed its differentiation and global gene expression properties. Upon TEL-AML1 expression, EML1 cells lost the capacity to differentiate into B-cells and underwent apoptosis. TEL-AML1 expression impaired the activation of IFNα/β signalling pathway in primary murine and human HSPCs with a dramatic inhibition of IRF3 phosphorylation, a member of the IFN-regulatory transcription factor family (De Laurentiis et al. (2015). This finding is consistent with the down-regulation of genes involved in IRF3-IFN signalling as shown in gene expression data derived from blasts of ALL patients expressing TEL-AML1 (Linka et al., 2013). These data suggest that IRF3-IFNα/β signalling is involved in the block of B-cell maturation elicited by TEL-AML1 expression. Furthermore, differentiation of cells expressing the TEL-AML1 protein can be restored by treatment with IFNβ (de Laurentiis et al., 2015).

Mice with reduced Ikaros expression (a master transcription factor that regulates lymphocyte differentiation) have a partial block at the pro-B cell stage in development, suggesting a tumorigenic role by blocking B-cell maturation (Teitell and Pandolfi, 2009). In the case of T-cell ALL, aberrant regulation or genetic mutations of cell-specific transcription factors inhibit cell maturation/development, leading to increased expansion of leukemic cells. More than 50% of T-cell ALL have activating mutations involving NOTCH1, a gene encoding a transmembrane receptor that regulates normal T-cell development by enhancing the transcription of diverse responder genes in developing thymocytes, such as cyclin D1 and c-MYC (Pui et al., 2008).

The Pax5 gene encodes the B-cell lineage specific activator protein that is expressed at early, but not late stages of B-cell differentiation. The developmental block observed in Pax5-deficient leukaemia cells can be reversed on restoration of Pax5 expression, suggesting that the reduction in Pax5 function results in a reversible disruption of differentiation. Transgenic RNAi can reversibly suppress endogenous Pax5 expression in the haematopoietic compartment of mice, which cooperates with activated signal transducer and activator of transcription 5 (STAT5) to induce B-ALL (Liu et al., 2016).

Although the Ebf1 dose-dependent events in B-cell precursors are not overtly leukaemogenic, the combination with Pax5 haploinsufficiency dramatically increases leukaemic potential by stalling B-cell differentiation at a highly proliferative and recombination active stage, which allows the selection and expansion of precursors carrying appropriate DNA mutations (Georgopoulos, 2015).

Epigenetic activation (i.e. hypomethylation) of ZNF423, a protein that interferes with B-cell differentiation, interacts with the early B-cell factor 1 (EBF-1) to inhibit transcription of EBF-1-targeted genes and subsequently trigger B-cell maturation arrest (Harder et al., 2013). Silencing of some miRNAs (e.g. miR-34b) or increased level of other miRNAs (e.g. miR-155) may lead to enhanced cell proliferation of leukaemic cells and/or inhibition of cell differentiation.

Uncertainties and inconsistencies

A prerequisite for the specific AOP is the occurrence of genetic damage (i.e. chromosomal translocations) in a particularly vulnerable genetic locus, within the proper cell and in a specific time window. However, details of this entire process and how it happens are not clear.

The target leukaemia-initiating cell(s) have not been identified so far with sufficient confidence and consequently there is no faithful cell model that recapitulates the pathogenesis in humans at the molecular level.
References


13277
3rd KER: Differentiation arrest of HSPCs (KE up) leading to clonal expansion of leukaemogenic cells (KE down)

How this Key Event Relationship works

In the ‘two-hit model’ widely accepted for leukaemogenesis two types of genetic aberrations and/or mutations are required. The first one is associated with a block in differentiation/maturation through chromosomal translocations affecting transcription factors that normally promote cellular differentiation at crucial steps during early haematopoiesis (see KER2). Further mutations affect genes controlling cellular proliferation and apoptosis, classically intracellular signalling pathways (i.e., tyrosine kinases), which lead to increased proliferation and/or inhibition of apoptosis (such as FLT3, RAS, KIT, and BCR–ABL) (Eriksson et al., 2014).

Dysregulation of immune responses to common infections might promote the malignant evolution of TEL-AML1-expressing preleukaemic clones. Ford et al. (2009) linked paediatric ALL with signalling pathways involved in infection and inflammation.

The major histocompatibility genes might play a role in the linkage between patterns of infection and leukaemia risk as several HLA haplotypes have been associated with childhood leukaemia (Wiemels, 2012). However, it is possible that exposure to infections promote B-ALL in children harbouring an intrinsic genetic susceptibility (Hauer et al., 2015).

Weight of evidence

Fusion gene products may suffice to initiate but not to fully complete leukaemogenesis, and other secondary genetic lesions must occur for developing childhood leukaemia. For instance, the concordance rate for ALL in monozygotic twin children is around 5-10%, and transgenic mice expressing fusion gene products (e.g. TEL-AML1 or AML1-ETO) did not exhibit overt signs of leukaemia (Mori et al., 2002). Also, some fusion genes associated with childhood leukaemia can be detected in the blood of normal individuals, indicating that they occur ubiquitously in humans and do not necessarily lead to the disease (Greaves and Wiemels, 2003).

After the occurrence of oncogenic fusion proteins resulting from chromosomal translocations, subsequent cooperating hits define the disease latency and occur after birth, and may be of genetic, epigenetic or immune nature (i.e. delayed infection-mediated immune deregulation).

Transgenic mice expressing TEL-AML1 failed to develop leukaemia and this finding was corroborated in subsequent reports where no leukaemia was observed despite the use of differing gene promoters to express the translocation. These experiments support the need of a second genetic event is necessary for the development of leukaemia (Jacoby et al., 2014).

Biological plausibility

Dysfunctions of the immune system and delayed infections have been linked to childhood leukaemia (Greaves, 2006; Swaminathan et al., 2015). Two factors might explain this association: a) a lower repertoire of infections during early immune development, and b) an altered congenital responder status to infection resulting in functionally aberrant clinical presentation of occasional infections. Thus, an untimely and excessive inflammatory response abolishes normal haematopoiesis, promoting selective expansion of a preleukaemic clone because of proliferative advantage and increased likelihood for a second mutation required for the development of the disease to occur (Sanjuan-Pla et al., 2015). Additional support is provided by studies showing that an increased opportunity for early childhood infections as well as normal childhood vaccinations protects against leukaemia indicating that vaccination reduces risk to leukaemia (Ma et al., 2005).

The IFNα/β cytokines, whose production is impaired by TEL-AML1 expression, have been long known to modulate resistance to viral infections and enhance innate and acquired immune responses. IFNs also influence tumour growth by directly inducing the expression of genes involved in apoptosis, or indirectly by inhibiting angiogenesis and modulating immune response (reviewed in De Laurentiis et al. (2015)).
Murine models with human precursor cells harbouring the *TEL-AML1* fusion gene generated a pre-leukaemic state that only resulted in an overt leukaemic phenotype upon the acquisition of additional genetic abnormalities (Alpar et al., 2015).

- Genetic alterations

Genetic alterations that impair cell differentiation probably cooperate with a second class of mutations that alter the proliferation and survival of HSPCs. One of these mutations affects the RAS-RAF-MEK-ERK signalling cascade, important for the HSPC development, leading to enhanced cell survival/proliferation (Case et al., 2008). Mutations of the receptor tyrosine kinase (RTK)-Ras signalling pathway have been associated with the pathogenesis of childhood (and perhaps infant) leukaemia (Driessen et al., 2013; Paulsson et al., 2015; Prelle et al., 2013). Also, the lack of degradation of cell signalling proteins enhances survival and proliferation of leukaemic cells as occurs either with inactivation of E3 ubiquitin ligase (Aranaz et al., 2012; Makishima et al., 2009), or with constitutive activation of MAP kinase (e.g. JNK), leading to proteasomal degradation of proapoptotic proteins (Leung et al., 2008).

Multiple secondary changes have been proposed to cooperate with *TEL-AML1* fusion for overt B-cell ALL. Gene deletions of non-antigen receptor or cell cycle regulatory proteins can further promote the proliferation and survival of leukaemic cells (Aplan, 2006; Meissner et al., 2014; Novara et al., 2009).

*TEL-AML1*-positive pre-leukaemic clones trigger an increase in ROS, which promotes the accumulation of secondary genetic lesions by increasing genetic instability and DNA DSBs, therefore enabling these preleukaemic clones to develop into leukaemic cells (Kantner et al., 2013).

Gawad et al. (2014) sequenced a panel of single nucleotide variants (SNVs), deletions, and IgH sequences in 1,479 single tumour cells from six ALL patients. By accurately segregating groups of co-occurring mutations into distinct clonal populations, co-dominant clones were found in the majority of patients. Evaluation of intraclonal mutation patterns a) identified clone-specific cytosine mutagenesis events, b) showed that most structural variants are acquired before SNVs, c) determined that *KRAS* mutations occur late in disease development but are not sufficient for clonal dominance, and d) identified within the same patient clones arrested at varied stages in B-cell development. Most large deletions occurred before cytosine mutagenesis-driven SNV acquisition, thus providing further evidence that the majority of the SNVs were caused by an APOBEC protein. Ongoing V(D)J recombination can occur in the most evolved clones, which can have variable magnitude between clones in the same patient. The development of leukaemic cells can be promoted by rearrangement of T-cell acute lymphoblastic leukaemia 1 gene (*TAL1*), which encodes a transcription factor that regulates both embryonic and adult haematopoiesis along with the inactivation of phosphatase and tensin homolog gene (*PTEN*), encoding a tumour suppressor dual-specificity phosphatase that antagonizes the PI3K signalling pathway via microdeletions due to illegitimate RAG activity (Mendes et al., 2014).

- Role of infections and immunity

Infection can trigger a series of events that are directly involved in genome instability. Olinski et al (2014) proposed that viral infections may result in aberrant expression of the AID (activation-induced deaminase)/APOBEC (apolipoprotein B editing complex) family of DNA cytosine deaminases, which are able to insert mutations in DNA and RNA by deaminating cytidine to uridine. AID is essential for the antigen-driven diversification of already rearranged immunoglobulin genes in the adaptive immune system. Since these enzymes also participate in active DNA demethylation process, changes in DNA methylation status or aberrant methylation can occur (Olinski et al., 2014). Altogether, these processes may lead to genome instability in prenatally generated pre-leukaemic cells and the emergence of ALL.

The inflammatory cytokine TGFβ has been involved in TEL-AML1-mediated leukaemogenesis since B cell progenitor cell line and human cord blood progenitor cells expressing TEL-AML1 inhibit downstream activation of TGF-β by binding to Smad3, the main TGF-β signalling target, thus preventing the activation of target promoters. As a result, TEL-AML1-expressing cells might propagate by inhibiting the tumour-suppressive properties of TGF-β.

Wild type mice kept in a specific-pathogen-free environment from birth and then moved to common infectious environment did not develop B-ALL (Martín-Lorenzo et al., 2015). Pax5+/− mice also failed to
develop leukaemia under non-infection exposure conditions; however, when these mice were exposed to infection they acquired point mutations in the second allele, which triggered the development of pB-ALL recapitulating the clinical, histopathological and molecular features of human B-ALL (Hauer et al., 2015). These data provide evidence that delayed exposure to infection can induce human-like B-ALL in mice on the basis of inherited genetic predisposition (see Figure 37 from Hauer et al., 2015).

**Fig 37:** Exposure to infection is a causal factor in B-precursor acute lymphoblastic leukemia as a result of Pax5 inherited susceptibility.

Monoallelic loss of Pax5 promotes leukaemogenesis by creating an aberrant IL7-sensitive progenitor compartment, a pre-leukaemic pre-B cell population susceptible to malignant transformation through accumulation of secondary Jak3 mutations, which depicts a rescue mechanism of the IL7/IL7R/STAT5 signalling. Transplantation experiments demonstrate that the activating Jak3 mutations per se are sufficient to drive leukaemia (Martin-Lorenzo et al., 2015).

The mechanisms underlying the conversion of the preleukaemic clone carrying the inherited PAX5 mutations into B-ALL are not understood yet; however, the B cell-specific enzyme AID might be the predominant driver of clonal evolution in human TEL-AML1 pB-ALL (Swaminathan et al., 2015).

Besides AID, RAG1-RAG2 also drives leukaemic clonal evolution after repeated exposure to inflammatory stimuli, paralleling chronic infections in childhood. Abnormal cytokine signalling and repetitive inflammatory stimuli exacerbated susceptibility to genetic lesions during B lymphopoiesis at the transition from the large pre-BII cell stage to the small pre-BII cell stage (Swaminathan et al., 2015).

**Uncertainties and inconsistencies**

One important question in leukaemia genomics is the identity of leukaemia-initiating mutations that result in preleukaemic clones. Owing to the technical challenge of distinguishing and isolating distinct cancer subclones, many aspects of clonal evolution are poorly understood, including the diversity of different subclones in an individual cancer, the nature of the subclones contributing to relapse, and the identity of pre-cancerous mutations. Studies of paediatric ALL demonstrated that in individual patients there are multiple genetic subclones of leukaemia-initiating cells, with a complex clonal architecture which limits to build a consistent AOP.

It remains to be demonstrated that subpopulations of acute leukaemia cells exhibit epigenetic heterogeneity, but it seems very likely that epigenetic diversity contributes to subclonal heterogeneity in acute leukaemia. Such epigenetic subclones likely differ in their proliferation, self-renewal,
differentiation and response to therapy, adding an additional dimension to the functional heterogeneity of leukaemia subclones (Jan and and Majety, 2013).

A number of questions arise from the Pax5 promoted leukaemogenesis: how relevant is the timing and pattern of infectious exposure for B-ALL development, how the second hit impacts on the target cell, and what makes Pax5+−/− stem/progenitor target cells more vulnerable to malignancy (Hauer et al., 2015).

On the other hand, the adverse effect of pesticides can be produced not only at the MIE level but also by promoting the accumulation of cooperating mutations in the quiescent preleukaemic clones based on a potential oxidative damage in rapidly dividing cells during DNA replications. Additionally, pesticides can exert a developmental immunotoxic effect by the interference of the normal development of the immune cells and their strictly regulated function (Corsini et al, 2013). However, the precise nature of these potential effects is lacking and would impact the current paradigm as pesticides might act at different events of the AOP by means of different toxicological pathways.

References


Jacoby E, Chien CD, Fry TJ. Murine models of acute leukemia: important tools in current pediatric leukemia research. Front Oncol 2014; 4: 95


4th KER: Clonal expansion of leukaemogenic cells (KE up) leading to overt childhood leukaemia (KE down – AO)

How this Key Event Relationship works

Children with ALL often present with signs and symptoms that reflect bone marrow infiltration with leukaemic blasts and the extent of extramedullary disease spread. The initial manifestations of the disease are based on the expansion of leukaemogenic cells replacing normal blood cells, and involve anaemia, thrombocytopenia and neutropenia with apparent clinical signs and symptoms.

The majority of ALL cases have multiple clones with distinct genetic alterations that influence the response to treatment and the risk of recurrence. Genome-wide association studies comparing diagnosis versus relapse specimens have shown that both of them share common origin at prediagnosis or clonal ancestry, but show differences in the nature of genetic alterations (Lo Nigro et al., 2013).

Biological stress from postnatal infection in combination with a dysregulated immune response may confer a growth advantage for a preleukaemic clone leading to its rapid expansion and an increased opportunity for the occurrence of a second mutation required for the development of childhood leukaemia (Greaves, 2006).

Childhood leukaemia is a biologically heterogeneous disease represented by distinct clinical and biological subtypes. The disease consists of a multistep process requiring the acquisition of multiple somatic lesions, and the definition of such pathways is being elucidated.

Weight of evidence

Although there is scarce scientific evidence on how leukaemic clones grow and expand, the pathobiology of the disease along with the evolutionary genetic landscape, response to treatment and relapse clearly indicate a causal linkage between the expansion of leukaemic clones and the onset of clinical features.

Biological plausibility

Sequential cooperating mutations in several signaling pathways (ie., RAS) and cellular processes are selectively produced in any of the in utero subclones and originated from the same pre-leukaemic clone. Later on, an aberrant inflammatory response abolishes normal hematopoiesis promoting selective expansion of a preleukaemic clone, resulting in stochastic or microenvironment-derived cooperating drivers toward overt leukaemia.

In the cord blood of healthy newborns the prevalence of a TEL/AML1 translocation is about 1 in 100, while only 1 in 10,000 will later in life develop ALL with this translocation. This clearly indicates a multistep pathogenesis: since at least 99% of the children with this 'first hit' will not develop leukaemia, more hits are necessary to develop leukaemia.

Only few children who are born with a chromosomal translocation will develop ALL, proving that these are preleukaemic changes and that leukaemogenesis is multifactorial and depending on multiple consecutive events. The 'first hit', most likely acquired during pregnancy, will give rise to preleukaemic cells and clones being more susceptible to additional oncogenic events, the 'second hit'. Most children with ALL carry 6 up to ~20 different genetic abnormalities in their leukaemia cells.

Biological stress from postnatal infection in combination with a dysregulated immune response may confer a growth advantage for a preleukaemic clone leading to its rapid expansion and an increased opportunity for the occurrence of a second mutation required for the development of childhood leukaemia (Greaves, 2006).

Uncertainties and inconsistencies

The main challenge of developing AOPs for childhood leukaemia is the complex nature of the disease. For example, a tumour suppressor gene could be mutated or transcriptionally inactivated to trigger leukaemogenesis. Different genetic aberrations affect different subtypes of childhood leukaemia (even between cell-specific B-cell and T-cell ALL) as almost all of the evaluated human studies report...
percentages of a specific mutation found in cohorts, meaning there is no single mutation responsible for the disease.

Whole genome and transcriptome sequencing of three B-cell precursor patients (of which one carried the TEL-AML1 translocation and two lacked a known primary genetic aberration and one T-ALL patient) found that each patient had a unique genome, with a combination of well-known and previously undetected genomic aberrations (Lindqvist et al., 2015).

References


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**Overall assessment of the AOP**

Childhood leukaemia, the most common cancer affecting children, fits a two-hit cancer model. While the first hit occurs *in utero* during fetal hematopoiesis (chromosomal translocation in a HSPC with a sustainable lifespan and clonal potential to propagate the chimeric gene product), the second hit takes place postnatally and is related to aberrant immune response to delayed infections or other secondary activating mutations both leading to clonal expansion. Studies in archived Guthrie cards suggest the presence of several common chromosomal translocations on neonatal blood spots in children who contract leukaemia later, which retrospectively indicate clearly an *in utero* origin of the disease (Wiemels et al., 1999). However, chromosomal translocations are insufficient by themselves to cause disease as they are found in approximately 1% of the normal population, a frequency 100 times higher than the prevalence of acute lymphoblastic leukaemia (ALL) (Mori et al., 2002). This fact suggests that the vast majority of preleukaemic clones are self-limiting and do not result in disease (Pui et al., 2008; Wiemels, 2012). In fact, transgenic mice expressing fusion gene products did not exhibit overt signs of leukaemia (Mori et al., 2002). The variable incubation period and clinical outcome of the disease, and the 10% concordance rate of leukaemia in identical twins harbouring the same genetic abnormalities indicates that additional postnatal events are needed for the development of full-blown childhood leukaemia (Greaves and Wiemels, 2003).

The initiating hit usually occurs *in utero* and commonly leads to the expression of oncogenic fusion proteins. Subsequent cooperating hits occurring after birth define the disease latency and may be of a genetic, epigenetic or immune nature (i.e., delayed infection-mediated immune deregulation). However, currently available information does not suggest a strong association of an exogenous agent(s) with a particular exposure window for childhood ALL. Although prenatal initiation of ALL might be a result of spontaneous developmental errors normally occurring through endogenous oxidative stress in the absence of an exogenous DNA-damaging exposure, the likelihood of this formation may be modified by other factors including exogenous ones. Different mechanisms of cellular responses pertinent to ALL induction are expected for the different classes of agents (e.g., chemicals, radiation and infection) (Kim et al., 2006). The effect of infection is postulated to be caused by a delayed immunological challenge associated with dysregulated proliferative and/or apoptotic stress to the ‘preleukaemic’ bone marrow, indicating a promotional effect of infection on childhood ALL, which could be considered a “classical” second hit, i.e. tumour promotion.

More recently, a 3-step model of leukaemia pathogenesis has been suggested, which postulates that an initiating genetic lesion (diverse chromosomal translocations leading to gene fusions) confers self-renewal properties to foetal liver HSPCs. A second lesion, normally affecting essential transcription factors for lymphoid cell development, causes differentiation block at the progenitor cell level. A third class of cooperating mutations accumulate and are needed to fully transform leukaemia cells. These secondary mutations affect pathways such as cell cycle, cytokine receptors and associated kinases, RAS signalling or several other transcription factors or epigenetic regulators (Duque-Afonso et al., 2015). While mutations of RAS in HSPCs have been demonstrated, K-RAS mutations are not sufficient to produce overt leukaemia, but requires additional genetic mutation(s) in most likely lineage-committed progenitors (Zhang et al., 2009). Besides, NOD/scid mice with transplanted human bone marrow leukaemic blasts at different maturation stages isolated from paediatric ALL patients developed the complete leukaemic phenotype in vivo. This suggests that B precursor blasts at different maturation stages have capability of self-renewal as a means of maintaining their malignancies *in vivo* (Le Viseur et al., 2008).
Pediatric leukaemia is phenotypically and genetically heterogeneous with an obscure aetiology. The interaction between genetic factors and environmental agents represents a potential etiological driver. Despite the multifactorial causal mechanism and a heterogeneous biological composition, the timing of environmental exposures and genetic changes associated with childhood leukaemia must be considered (Buffler et al., 2005). However, its genetic diversity limits investigation into the molecular pathogenesis of disease. As a result of the peculiar natural history of childhood leukaemia, direct studies in pregnant women are not possible and there is a need to rely on surrogate in vitro or ex vivo studies or on animal models which entail difficulties in the interpretation and extrapolation of results.

Over the last 3 decades, significant progress has been made through the identification of recurrent genetic alterations and translocations in leukaemic blast populations, and their subsequent functional characterization in cell lines and/or mouse models. Recently, primary human hematopoietic cells have emerged as a complementary means to characterize leukaemic oncogenes (Kennedy and Barabé, 2008). Accordingly, this overall assessment is based largely on empirical evidence found in cases of childhood leukaemia or from cellular and animal models.

1. Concordance of dose-response relationship

In contrast to infant leukaemia, the lack of a known etiological (chemical) agent directly related to the onset of the disease has prevented the conduct of experimental studies in animals, so a dose-response relationship is lacking so far. In addition, there are no adequate experimental systems in which dose-response and response-response relationships can be studied across MIE, KEs and AO.

Conversely, models of radiation-induced leukaemia risk derived from leukaemia mortality among Japanese survivors of atomic bombs adopted a linear dose–response relationship in the low-dose (<100 mGy) region (Wakeford et al., 2010). A dose-response relationship was demonstrated for childhood leukaemia based on number of X-ray films taken and from the observation that the excess risk was greater among twins for whom X-ray pelvimetry was far more frequent than among singletons (Boice, 2006).

2. Temporal concordance among the MIE, KEs and AO

There is no doubt about temporal concordance among MIE, KEs and AO for childhood leukaemia. Key molecular events leading to childhood leukaemia are chromosomal translocations, and mis-repaired DNA DSBs are prerequisites for their occurrence. Most of DNA lesions in foetal liver HSPCs are properly repaired and only persist in case of a failure in the DNA repairing system. Chromosomal translocations ultimately result in the deregulation of key cellular proteins, especially those encoded by proto-oncogenes and tumour suppressor genes, which are critical functional regulators of cells.

Recurrent genetic insults leading to differentiation arrest of HSPCs are needed to drive uncontrolled proliferation and survival of the differentiation-blocked clone. A study using transgenic mice with the TEL-AML1 transgene has demonstrated that expression of the fusion gene alone is not sufficient to induce leukaemia, but following prenatal initiation a postnatal second event is necessary for ALL to be manifested (Andreasson et al., 2001).

Regardless of the gap of knowledge on the chemical(s) involved in the MIE, and the molecular mechanisms underlying this interaction, it is clear that chromosomal aberrations represent a necessary but not suffice cause occurring in utero. A second-hit is required for the expansion of quiescent leukaemic clones and this occurs during postnatal life in a subset of vulnerable children because of an immunological system improperly developed owing to low exposure to common infections early in life.

The separation of a clonal antecedent preleukaemic cell population from frank leukaemic cells has been identified in a monochorionic twin pair with one ‘leukaemic’ and one ‘healthy’ twin (Hong et al., 2008). The ‘healthy’ twin shared TEL-AML1 transduction in normal cord blood suggested that the founding chromosomal translocation was likely sufficient to induce the preleukaemic population found in the ‘healthy’ twin. A follow-up study in the same twin pair used genome-wide copy number alterations (CNA) profiling to identify three potential ‘driver’ CNA in the leukaemic cells. FISH analysis did not detect these three CNA in the ‘healthy’ twin’s preleukaemic cells, supporting the hypothesis that the pre-leukaemic cells diverged genetically after the initiating chromosomal translocation, with subsequent events leading to the clonal evolution of the affected twin’s leukaemia (Bateman et al., 2010). A further whole genome sequencing study assessed
the genomic profiles of monozygotic twins with ALL, and found that while twins share the first
initiating lesion (occurring in utero), each twin then acquire distinct non-coding mutational changes
postnatally that drive leukaemogenesis (Ma et al., 2013).

Current epidemiological studies have limitations for the demonstration of the AOP in pregnant women
even in the absence of a clear characterization of exposure. While experimental models are in
accordance with the AOP, the available evidence shows a large number of (epi)genetic and host
factors potentially modifying the pathogenesis of childhood leukaemia. The translational biology of B
cell precursor ALL has been investigated using comparative genomics and functional approaches
(Duque-Afonso et al., 2015), which has allowed to recapitulate experimentally the multistep
pathogenesis of ALL previously inferred from genomic analyses and highlight key cooperating
oncogenic pathways.

When known mutations occur in non-stem cells, they will quickly be lost from the hematopoietic pool
due to the natural course of differentiation and cell death. In contrast, a mutation in a stem cell may
persist, and the mutated clone may expand, facilitating further clonal progression until a leukaemic
stem cell with extensive self-renewal ability develops (Jan and Majeti, 2013).

**Table 26:** Temporality concordance table

<table>
<thead>
<tr>
<th>Model (concentration)</th>
<th>MIE</th>
<th>KE1 In utero induction of chromosomal translocations</th>
<th>KE2 Differentiation blockage</th>
<th>KE3 Clonal expansion</th>
<th>AO Childhood leukaemia</th>
</tr>
</thead>
<tbody>
<tr>
<td>Conditional activation of E2A-PBX1 in the B cell compartment of mice¹</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>ALL development (similar to human E2A-PBX1⁺ leukaemias)</td>
<td></td>
</tr>
<tr>
<td>Transplantation of TEL-AML1 transduced human cord blood cells into NOD/scid mice²</td>
<td></td>
<td>+</td>
<td>+</td>
<td>Mice exhibited features of preleukaemic phase of pre-B cell ALL²</td>
<td></td>
</tr>
<tr>
<td>Human peripheral lymphocytes exposed to 0.1-10 µg/mL isofenphos for 1 h</td>
<td></td>
<td>+</td>
<td>-</td>
<td>Human peripheral lymphocytes exposed to 0.1-10 µg/mL isofenphos for 1 h could lead to genomic instability and leukaemogenesis³</td>
<td></td>
</tr>
<tr>
<td>Human K562 cells exposed to 0.1 µM of diazinon</td>
<td></td>
<td>+</td>
<td>-</td>
<td>Human K562 cells exposed to 0.1 µM of diazinon hypermethylated genes involved in cell cycle arrest (cyclin-dependent kinase inhibitor 1A and CDKN1C) as well as tumor suppressor genes (p53 and PTEN)⁴</td>
<td></td>
</tr>
<tr>
<td>1-100 µM chlorpyrifos for up to 24 hours</td>
<td></td>
<td>+</td>
<td>-</td>
<td>1-100 µM chlorpyrifos for up to 24 hours induced double-strand DNA breaks in HSCs (and MLL rearrangements)⁵</td>
<td></td>
</tr>
<tr>
<td>Human T-cell leukaemia Jurkat cells exposed to</td>
<td></td>
<td>-</td>
<td>-</td>
<td>Human T-cell leukaemia Jurkat cells exposed to increased double DNA breaks (possibly due to…</td>
<td></td>
</tr>
</tbody>
</table>

¹ Duque-Afonso et al., 2015
² Jan and Majeti, 2013
³ Human peripheral lymphocytes exposed to 0.1-10 µg/mL isofenphos for 1 h could lead to genomic instability and leukaemogenesis
⁴ Human K562 cells exposed to 0.1 µM of diazinon hypermethylated genes involved in cell cycle arrest (cyclin-dependent kinase inhibitor 1A and CDKN1C) as well as tumor suppressor genes (p53 and PTEN)
⁵ 1-100 µM chlorpyrifos for up to 24 hours induced double-strand DNA breaks in HSCs (and MLL rearrangements)
⁶ Human T-cell leukaemia Jurkat cells exposed to increased double DNA breaks (possibly due to…
### Table: Studies on the Effects of Methyl-Pyrazole Insecticides on Human Cells

<table>
<thead>
<tr>
<th>Study Description</th>
<th>Results</th>
<th>Notes</th>
</tr>
</thead>
<tbody>
<tr>
<td>CEM x 174 cell line, (a hybrid of human T and B cells), exposed to 50 µM heptachlor, chlordane or toxaphene for 24-36 h</td>
<td>Decrease in levels of the tumour suppressors p53 and Rb&lt;sup&gt;6&lt;/sup&gt;</td>
<td></td>
</tr>
<tr>
<td>Human peripheral lymphocytes exposed to a commercial fungicide karathane (dinocap) at 20 µg/mL for 24 h</td>
<td>Increased chromosomal aberrations, formation of sister chromatid exchanges and decreased mitotic index&lt;sup&gt;9&lt;/sup&gt;</td>
<td></td>
</tr>
<tr>
<td>Studies on the Kasumi-1 cell line, which harbors an AML1–ETO translocation,</td>
<td>The differentiation block induced by AML1–ETO is due in part to its ability to physically bind to and inactivate the master myeloid transcription factor PU.1&lt;sup&gt;10&lt;/sup&gt;</td>
<td></td>
</tr>
</tbody>
</table>

<sup>1</sup> A conditional E2A-PBX1 fusion gene (Duque-Afonso et al., 2015).
<sup>6</sup> Graillot V, Tomasetti F, Cravedi JP, Audebert M. Evidence of the in vitro genotoxicity of methyl-pyrazole pesticides in human cells. Mutation Research-Genetic Toxicol Environ Mutag 2012; 748: 8-16
<sup>7</sup> Rought SE, Yau PM, Chuan LF, Doo RH, Chuan RY. Effect of the chlorinated hydrocarbons heptachlor, chlordane, and toxaphene on retinoblastoma tumor suppressor in human lymphocytes. Toxicol Lett 1999; 104: 127-135

### 3. Strength, consistency, and specificity of association of AO and MIE

Regarding the experimental models and genome-wide association studies on childhood leukaemia, strength, consistency and specificity of association of AO and MIE is rather strong in spite of the gap of knowledge on the etiological factors involved. Although direct observations on the initial <i>in utero</i> MIE are not possible, there is inferential evidence from animal and <i>in vitro</i> cellular studies suggesting...
strongly that childhood leukaemia recapitulates to a large extent the development of the human
disease.

Under the current paradigm, the first initiating oncogenic mutation usually involves structural or
numerical chromosomal alterations, impairing normal cell differentiation, while secondary hits more
commonly comprise mutations affecting developmentally-regulated master transcription factors or
membrane-proximal signalling pathways conferring proliferation and survival advantages to the
differentiation-blocked clone. The development of leukaemia requires the activation of cell
proliferation in addition to differentiation blockage.

The consistent finding of a number of chromosomal translocations across studies indicates that they
are needed for the development of the disease, although not enough by themselves. There is no
alternative explanation for this finding because a reasonable confidence for chance or confounding is
lacking. Besides, the identified chromosomal damage (and no other) has to occur in a particular cell
(foetal liver HSPCs) and in a particular time window, as otherwise the disease will not develop.
### 4. Weight of Evidence (WoE)

#### 4.1 Biological plausibility, coherence, and consistency of the experimental evidence

**Table 27: Biological plausibility of the KERs; WoE analysis**

<table>
<thead>
<tr>
<th>Support for Biological Plausibility of KERs</th>
<th>Defining Question</th>
<th>High (Strong)</th>
<th>Moderate</th>
<th>Low (Weak)</th>
</tr>
</thead>
<tbody>
<tr>
<td>MIE → KE1</td>
<td>Low</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Unknown</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

#### KE1 → KE2

Chromosomal translocations lead to differentiation arrest of HSPCs *in utero*

<table>
<thead>
<tr>
<th>Defining Question</th>
<th>High (Strong)</th>
<th>Moderate</th>
<th>Low (Weak)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Is there a mechanistic (i.e. structural or functional) relationship between KEup and KE down consistent with established biological knowledge?</td>
<td>Extensive understanding of the KER based on extensive previous documentation and broad acceptance</td>
<td>The KER is plausible based on analogy to accepted biological relationships, but scientific understanding is not completely established</td>
<td>There is empirical support for a statistical association between KEs but the structural or functional relationship between them is not understood</td>
</tr>
</tbody>
</table>

Rationale: there is convincing evidence to indicate that chromosomal translocation leading to formation of TEL-AML1 is the initiating event in most B-precursor ALL, the most frequent childhood leukaemia.

DNA sequencing analysis has revealed that TEL-AML1 translocations occur by imprecise and error-prone DNA repair process after DNA double-strand breaks and not by aberrant topoisomerase activity (Wiemels and Greaves, 1999).

Studies on identical twins and neonatal blood samples strongly indicate an *in utero* occurrence of the KER. The TEL-AML1 fusion gene usually arises before birth, inducing persistent self-renewing of pro-B cells in mice (covert preleukaemic clone). Aberrant proteins produced by fusion genes are responsible of cell differentiation arrest.

<table>
<thead>
<tr>
<th>Low (Weak)</th>
</tr>
</thead>
<tbody>
<tr>
<td>KE2 → KE3</td>
</tr>
</tbody>
</table>

Cooperative mutations and Delayed infections in HSPCs with a differentiation blockage lead to

<table>
<thead>
<tr>
<th>Defining Question</th>
<th>High (Strong)</th>
<th>Moderate</th>
<th>Low (Weak)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Is there a mechanistic (i.e. structural or functional) relationship between KEup and KE down consistent with established biological knowledge?</td>
<td>Extensive understanding of the KER based on extensive previous documentation and broad acceptance</td>
<td>The KER is plausible based on analogy to accepted biological relationships, but scientific understanding is not completely established</td>
<td>There is empirical support for a statistical association between KEs but the structural or functional relationship between them is not understood</td>
</tr>
</tbody>
</table>

Rationale: Covert preleukaemic clones may convert to precursor B-cell leukaemia following the accumulation of secondary genetic hits. 1 TEL-AML1+ cells differentiate terminally in the long term, providing a "window" period that may allow secondary genetic hits to accumulate and lead to leukaemia. In childhood leukaemia, altered differentiation and self-renewal of haematopoietic stem cells or early progenitor cells might occur due to the presence of chimeric transcription factors that alter the regulation of genes required
clonal expansion

for the proper differentiation of haematopoietic stem cells (Pui et al., 2004).

There is a general understanding of the mechanisms leading to differentiation arrest and clonal expansion and there is evidence that the principal fusion product TEL-AML1 protein harbours the necessary properties to execute the necessary pathways. However, the inability of available in vivo models to recapitulate the whole AOP process is an important limitation.

The longer latency observed in childhood leukaemia unequivocally indicates that the initiating chromosomal translocation itself is unlikely to convert a preleukaemic clone into an overt disease, thus suggesting the need for secondary cooperating (epi)-genetic events.

KE3 → AO

Clonal expansion leads to childhood leukaemia

STRONG

Rationale: The basic processes underlying overt leukaemia development are well understood and accepted.

13733 Tsuzuki S, Seto M. TEL (ETV6)-AML1 (RUNX1) initiates self-renewing fetal pro-B cells in association with a transcriptional program shared with embryonic stem cells in mice. Stem Cells 2013; 31: 236-247

13734

4.2 Essentiality

Table 28: Essentiality of the KEs; WoE analysis

<table>
<thead>
<tr>
<th>KE1</th>
<th>STRONG</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Experimental models and genome wide association studies have consistently demonstrated that in the absence of chromosomal damage there is no chance for the leukaemia to occur. The reverse is also true, as the</td>
</tr>
</tbody>
</table>
**In utero** chromosomal translocations  | presence of fusion genes per se are not enough for fully developing the disease.

**KE2**  
Blockage of HSPCs differentiation *in utero*  | The developmental block observed in Pax5-deficient leukemia cells can be reversed on restoration of Pax5 expression, suggesting that the reduction in Pax5 function results in a reversible disruption of differentiation. Transgenic RNAi can reversibly suppress endogenous Pax5 expression in the haematopoietic compartment of mice, which cooperates with activated signal transducer and activator of transcription 5 (STAT5) to induce B-ALL (Liu et al., 2016).

**KE3**  
Expansion of preleukaemic clones as a result of cooperative mutations and delayed infections.  | There are no relevant inhibitors for the clonal expansion of preleukaemic clones.

---

### 4.3 Empirical support

**Table 29:** Essentaility of the KERs; WoE analysis

<table>
<thead>
<tr>
<th>3 Empirical support for KERs</th>
<th>Defining Question</th>
<th>High (Strong)</th>
<th>Moderate</th>
<th>Low(Weak)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Does the empirical evidence support that a change in the KEup leads to an appropriate change in the KE down? Does KEup occur at lower doses and earlier time points than KE down and is the incidence of KEup higher than that for KE down? Are inconsistencies in empirical support across taxa, species and stressors that don’t align with expected pattern for hypothesized AOP?</td>
<td>Multiple studies showing dependent change in both events following exposure to a small number of specific stressors and some evidence inconsistent with expected pattern that can be explained by factors such as experimental design, technical considerations, differences among laboratories, etc.</td>
<td>Demonstrated dependent change in both events following exposure to a small range of specific stressors (extensive evidence for temporal, dose-response and incidence concordance) and no or few critical data gaps or conflicting data.</td>
<td>Limited or no studies reporting dependent change in both events following exposure to a specific stressor (ie endpoints never measured in the same study or not at all); and/or significant inconsistencies in empirical support across taxa and species that don’t align with expected pattern for hypothesized AOP.</td>
<td></td>
</tr>
</tbody>
</table>

**MIE → KE1**  
LOW
| KE1 → KE2 | STRONG | Rationale: A transgenic zebrafish model expressing TEL-AML1-positive ALL either ubiquitously or in lymphoid progenitors showed B-cell differentiation arrest. TEL-AML1 expression in all lineages, but not lymphoid-restricted expression, led to progenitor cell expansion that evolved into oligoclonal B-lineage ALL in 3% of the transgenic zebrafish\textsuperscript{2}.
| | | The strongest evidence comes from experimental models and genome wide association studies.
| KE2 → KE3 | Moderate | Rationale: In principle, there are a large number of factors and pathways linking the fusion product and differentiation blockage with clonal expansion, recurrent cooperative mutations and delayed infections.
| KE3 → AO | Moderate | Rationale: The accumulation of recurrent mutations in preleukaemic clones and dysfunction of the immune system following delayed infections have been linked to the development of childhood leukaemia.

5. Uncertainties and Inconsistencies

- Although the causes of the specific genetic events leading to ALL formation are not known, numerous exposure-dependent risk factors for childhood ALL have been proposed, including pesticide exposure. These risk factors can be classified based on their relationship to potentially critical exposure windows (i.e., preconceptional, prenatal, and postnatal stages). [KER1]

- As in utero evidence is difficult to obtain in humans, mouse models are used instead.

- Limitations of transgenic animal models faithfully recapitulating all the aspects of human pB-ALL need to be recognized. The reproducibility and accuracy of these models for human responses have yet to be validated, so their application to elucidate postnatal exposure effects of exogenous agents on childhood ALL should be cautiously approached. Mouse models in which the initiating oncogenic alteration(s) is not directed to the right cell-of-origin are unlikely to accurately recapitulate the etiology of the human disease and will originate an inaccurate model of human leukaemia (Hauer et al., 2014). [Experimental models]

- One important question in leukaemia genomics is the identity of leukaemia-initiating mutations that result in preleukaemic clones. Owing to the technical challenge of distinguishing and isolating distinct cancer subclones, many aspects of clonal evolution are poorly understood, including the diversity of different subclones in an individual cancer, the nature of the subclones contributing to relapse, and the identity of pre-cancerous mutations. Studies of paediatric ALL demonstrated that in individual patients there are multiple genetic subclones of leukaemia-initiating cells, with a complex clonal architecture which limits to build a consistent AOP. [KER3]

- The main challenge of developing AOPs for childhood leukaemia is the complex nature of the disease. For example, a tumour suppressor gene could be mutated or transcriptionally inactivated to trigger leukaemogenesis. Different genetic aberrations affect different subtypes of childhood leukaemia (even between cell-specific B-cell and T-cell ALL) as almost all of the evaluated human studies report percentages of a specific mutation found in cohorts, meaning there is no single mutation responsible for the disease.

- It remains to be demonstrated that subpopulations of acute leukaemia cells exhibit epigenetic heterogeneity, but it seems very likely that epigenetic diversity contributes to subclonal heterogeneity in acute leukaemia. Such epigenetic subclones likely differ in their proliferation, self-renewal, differentiation and response to therapy, adding an additional dimension to the functional heterogeneity of leukaemia subclones (Jand and Majety, 2013). [Clones]

- While there are emerging studies implicating epigenetics as an influential factor in childhood leukaemia, it is not clear at which point epigenetics influences childhood leukaemia pathogenesis, i.e. as a MIE or later as an intermediate event. For example, DNA hypermethylation of tumour suppressor genes leading to their decreased expressions can occur early in childhood leukaemia pathogenesis to facilitate the growth of leukaemic cells, or altered expressions of microRNAs might be influenced by earlier events (e.g.), resulting in alterations in haematopoiesis or inhibition of apoptosis. Therefore, the putative relevance of epigenetics needs to be further evaluated before it can be considered in the AOP development for childhood leukaemia. [Epigenetics]

- A detailed understanding of leukaemogenesis requires the development of experimental models that can accurately model this process. As a complement to work in cell lines and in mice, there is a need for oncogenes and chromosomal translocations to be studied in the appropriate cellular context, that of primary human hematopoietic cells. Retroviral-mediated transduction of primary human hematopoietic cells followed by their transplantation in vivo has emerged as a feasible approach to study the process of human leukaemogenesis (Kennedy and Barabe, 2008). [Experimental models]
6. Quantitative Considerations

The WOE analysis indicates that some KEs and KERs lack especially experimental evidence, but overall the analysis supports the qualitative AOP. The strong element in the development of the qualitative AOP is that it can partially be based on animal models recapitulating many crucial features of childhood leukaemia and genome-wide association studies. The absence of a MIE clearly defined is a major limitation. The lack of sufficient experimental data and uncertainties in quantitative information from available studies makes it difficult to build convincing dose (concentration)-response and response-response relationships and to identify possible practical thresholds for stressors.

There is a need for an adequate and robust experimental model system where relationships between doses, concentrations and responses can be evaluated within a temporal framework of the AOP.

7. Applicability of the AOP

Even in the absence of a mechanistic understanding in regard to the MIE, the proposed AOP might be applied to pesticide-related leukaemia not only in children but also in adults, although in the latter case chromosomal translocations are acquired in the postnatal life (the persistence of prenatal chromosomal translocations does not play a role in adult leukaemogenesis). Based on the rather consistent epidemiological evidence on human exposure to pesticides and the risk of childhood leukaemia, it is possible that at least a subset of acute childhood leukaemias may be caused by environmental exposure to pesticides. Consequently, the proposed AOP may be partially applicable to these situations, but should be supported with an understanding of the mechanistic processes underlying the direct or indirect interaction of pesticides (or their metabolites) with DNA.

References

Andreasson P, Schwaller J, Anastasiadou E, Aster J, Gilliland DG. The expression of ETV6/CBFA2 (TEL/AML1) is not sufficient for the transformation of hematopoietic cell lines in vitro or the induction of hematologic disease in vivo. Cancer Genet Cytogenet 2001; 130: 93-104


Glossary [and/or] Abbreviations

Glossary: an alphabetical list of words relating to a specific subject with explanations; a brief dictionary.

Abbreviation: a shortened form of a word or phrase (such as Mr, Prof). It also includes acronyms (a group of initial letters used as an abbreviation for a name or expression, each letter being pronounced separately – such as DVD, FDA – or as a single word – such as EFSA, NATO).

XXX  Dsadsadsadsa
YYY  Sdsdsadsad
ZZZ  Fdfsafasdf