European Food Safety Authority

ZOONOSES MONITORING

HUNGARY

The Report referred to in Article 9 of Directive 2003/99/EC

TRENDS AND SOURCES OF ZOONOSES AND ZOONOTIC AGENTS IN HUMANS, FOODSTUFFS, ANIMALS AND FEEDINGSTUFFS

including information on foodborne outbreaks, antimicrobial resistance in zoonotic agents and some pathogenic microbiological agents.

IN 2013

INFORMATION ON THE REPORTING AND MONITORING SYSTEM

Country: Hungary

Reporting Year: 2013

Laboratory name	Description	Contribution
Central Agricultural Office		Responsible authority for zoonoses data collection and reporting

PREFACE

This report is submitted to the European Commission in accordance with Article 9 of Council Directive 2003/99/ EC*. The information has also been forwarded to the European Food Safety Authority (EFSA).

The report contains information on trends and sources of zoonoses and zoonotic agents in Hungary during the year 2013 .

The information covers the occurrence of these diseases and agents in humans, animals, foodstuffs and in some cases also in feedingstuffs. In addition the report includes data on antimicrobial resistance in some zoonotic agents and commensal bacteria as well as information on epidemiological investigations of foodborne outbreaks. Complementary data on susceptible animal populations in the country is also given. The information given covers both zoonoses that are important for the public health in the whole European Community as well as zoonoses, which are relevant on the basis of the national epidemiological situation.

The report describes the monitoring systems in place and the prevention and control strategies applied in the country. For some zoonoses this monitoring is based on legal requirements laid down by the Community Legislation, while for the other zoonoses national approaches are applied.

The report presents the results of the examinations carried out in the reporting year. A national evaluation of the epidemiological situation, with special reference to trends and sources of zoonotic infections, is given. Whenever possible, the relevance of findings in foodstuffs and animals to zoonoses cases in humans is evaluated.

The information covered by this report is used in the annual Community Summary Report on zoonoses that is published each year by EFSA.

Hungary - 2013

^{*} Directive 2003/ 99/ EC of the European Parliament and of the Council of 12 December 2003 on the monitoring of zoonoses and zoonotic agents, amending Decision 90/ 424/ EEC and repealing Council Directive 92/ 117/ EEC, OJ L 325, 17.11.2003, p. 31

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1. ANIMAL POPULATIONS

The relevance of the findings on zoonoses and zoonotic agents has to be related to the size and nature of the animal population in the country.

A. Information on susceptible animal population

Sources of information

Data on susceptible animal populations were taken from official publications of the Hungarian Central Statistical Office unless it was collected by the Directorate of Food Chain Safety and Animal Health of the National Food Chain Safety Office.

Dates the figures relate to and the content of the figures

The figures relate to year 2013.

National evaluation of the numbers of susceptible population and trends in these figures

Data of December 2013 show that the number of cattle continued to grow. The pig stock – after 3 years of decrease and a slightly rose in 2012 is stable. The stock of poultry increased compared to December of the previous year.

Additional information

Table Susceptible animal populations

* Only if different than current reporting year

		Number of he	erds or flocks		slaughtered nals	Livestock numbers (live animals)		Number o	f holdings
Animal species	Category of animals	Data	Year*	Data	Year*	Data	Year*	Data	Year*
Cattle (bovine animals)	- in total	17573				848338			
Ducks	- in total					5868518			
Gallus gallus (fowl)	- in total					34113345			
Goats	- in total	785				15190			
Pigs	- in total	29046				2954264			
Sheep	- in total	7001				829171			
Solipeds, domestic	horses - in total					59200			
Turkeys	- in total					4668994			

2. INFORMATION ON SPECIFIC ZOONOSES AND ZOONOTIC AGENTS

Zoonoses are diseases or infections, which are naturally transmissible directly or indirectly between animals and humans. Foodstuffs serve often as vehicles of zoonotic infections. Zoonotic agents cover viruses, bacteria, fungi, parasites or other biological entities that are likely to cause zoonoses.

2.1 SALMONELLOSIS

2.1.1 General evaluation of the national situation

A. General evaluation

History of the disease and/or infection in the country

In 1992 the Veterinary Science Committee of the Hungarian Academy of Sciences has established its Salmonella Subcommittee with the main aim to support the work of the Hungarian Ministry of Agriculture and Rural Development in the control of Salmonella with regards to poultry flocks.

This subcommittee has formed a working group with EU experts to prepare the Integrated Quality Chain System for Salmonella Control in the Hungarian Poultry Sector (Edel-Wray-Nagy et al, 1995).

This has been issued by the Ministry for use in the poultry sector and distributed to the County Animal Health and Food Control Stations in 1995. In further years the Salmonella Subcommittee has arranged several courses and lectures to distribute the booklet for wider use. The Basic Document of this Guideline contained the adaptation of Council directive 92/117/EEC. The Guidelines contained general and specific instructions for hatcheries, breeding flocks, broilers, layers, egg packaging plants, slaughterhouses and feedmills. A special chapter was devoted to disinfection and cleaning.

Based on the above Guidelines several large Hungarian poultry farming systems (Babolna, Boly, Nadudvar) have built up and started their Salmonella Reduction Programs between 1996 and 2002. Besides, the Salmonella subcommittee has agreed with the Ministry of Agriculture and Rural Development to review the situation and to propose a Hungarian Salmonella Reduction Plan for Hungary, which was published by Nagy et al. in 1997.

Directive 92/117/EEC and the basics of the above mentioned Guidelines served the basis for the first ministerial decree [49/2002. (V.24) FVM] on the control of salmonellosis in poultry flocks, which referred to Salmonella Enteritidis and S. Typhimurium in Gallus gallus. The amendment to this Directive [97/2003. (VIII.19) FVM] made the application of the Order compulsory for breeding flocks and hatcheries, and continued to define the above 2 Salmonella serovars to be regarded as Salmonella for the purposes of that decree. The amendment also made the vaccination of table egg producing laying flocks compulsory. After the accession the EC regulations became directly applicable in Hungary as well. From that time EC regulations are followed. The implementation of these regulations is regulated by Decree 180/2009. (XII.29.)of Ministry of Agriculture.

National evaluation of the recent situation, the trends and sources of infection

Significant decrease could be seen as in the prevalence of salmonella in all types of flocks under scope of national control plans as in meat, meat products, table eggs and egg products of Gallus gallus.

Recent actions taken to control the zoonoses

Vaccination is not compulsory in flocks of Gallus gallus and Meleagris gallopavo. The rules of using vaccination and treatment are laid down in Commission Regulation (EC) No 200/2010 of implementing Regulation (EC) No 2160/2003 of the European Parliament and of the Council as regards requirements for the use of specific control methods in the framework of the national programmes for the control of salmonella in poultry.

2.1.2 Salmonellosis in humans

A. Salmonellosis in humans

Reporting system in place for the human cases

There are around 80 communicable diseases notifiable in Hungary based on legal background. The physician (in primary health care, specialist care, inpatient medical institution or pathology) who first diagnoses a case of a notifiable communicable disease (even the suspicion of the disease!) immediately reports data of case to the first level of the epidemiological network (municipal institute) of National Public Health and Medical Officer's Service (NPHMOS). Data must be reported both at the beginning and at end of the illness (recovery/death, result of laboratory test). The NPHMOS has a nationwide electronic system for registering and analysing data of communicable diseases in a combined national database, so the system provides online connection amid the three levels (municipal, county and national level – National Centre of Epidemiology - NCE) of the organization. The NCE prepares reports regularly (weekly, monthly, yearly) to the Chief Medical Officer, the MoH and the Hungarian Central Statistical Office.

Case definition

Confirmed case: a clinically compatible case when the salmonella infection is laboratory confirmed. Probable case: a clinically compatible case that is not confirmed by laboratory investigation, but it has an epidemiological link to a confirmed salmonellosis outbreak.

Diagnostic/analytical methods used

Salmonella isolates are obtained by culturing the faeces samples of the patients on selective-differentiating media, followed by biochemical testing and serotyping. Since 2003 the Hungarian and the Colindale sets of phages have been parallel used for phage typing of the human S. Enteritidis isolates received by the Phage-typing and Molecular Epidemiology Department of the 'Johan Bela' National Centre for Epidemiology. For S.Typhimurium isolates the schemes of Felix and Callow as well as Anderson et al. are also in use.

Notification system in place

Human cases have been notifiable since 1959. The physician reports data of case on a "case report form" by mail to the municipal institute of NPHMOS. The specialist of the institute records data immediately in the electronic system of the NPHMOS. Hungary has also a laboratory based surveillance system, and the NPHMOS has representative dataset from most of the microbiological laboratories about the laboratory investigated cases (since 2003 antibiotic resistances have also been reported from 5 regional laboratory of NPHMOS and from a number of laboratories from universities or hospitals).

The illness is reported first as enteritis infectiosa syndrome on the basis of the symptoms. Having the results of the laboratory tests this syndrome-based diagnose is modified to etiology-based diagnose. In some cases reporting follows only the available laboratory test results.

History of the disease and/or infection in the country

Human cases have been notifiable since 1959. The isolated strains have been phage-typed since the 1960s. The number of the recorded cases has continuously increased from 1959 to 1996 (with a maximum of 28 046 reported case/year, incidence: 274,6/100 000 inhabitant/year). The number of the recorded outbreaks has also increased in a similar way (outbreak = two epidemiologically linked cases of salmonellosis, maximum number of reported outbreaks: 3450 outbreaks in 1995). Since 1996 both the number of the recorded cases and the outbreaks has continuously decreased. The mortality has

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increased only in the period of 1972-1994 (10-20 death/year, case fatality rate: 0.1-0.4%). In the other years the mortality was 5-10 death cases per year (case fatality rate: 0.03-0.09%). The age-specific incidence was the highest for the infants in all periods, and it declined with the progressing of the age. The investigation of the outbreaks mostly demonstrated a food-borne origin. The ratio of the person-to-person transmission is insignificant. In the history of human salmonellosis in Hungary there were less than 10 outbreaks caused by contaminated water.

Up to 1980 the serotype S. Typhimurium predominated, and pork was identified as the main source of infection. At that time the infection has spread by homemade foods and also by the products of food-industry. Since 1980 the serotype S. Enteritidis has become predominant and poultry has been identified as the main source of the infection. Since then the prevalence of this serotype has remained about 70-80%. Between 1975 and 1980 the S. Enteritidis phage type 7 (according to the Hungarian scheme) has predominated. In the period of 1980-1990 strains characterized with phage type 1, from 1990 to 1996 strains characterized with phage type 1, 6 and 6b (according to the Hungarian scheme) were most frequently identified. After 1997 the phage type 6 (acc. to the Hungarian scheme) has become the most frequently occurring phage type.

National evaluation of the recent situation, the trends and sources of infection

The epidemiological situation of the salmonellosis in Hungary has continuously improved till 2004. The number of cases has decreased from 11 507 to 7557 since 2000 (incidence ranged between $114,3-74,7/100\,000$ inhabitants/year), the case fatality ratio changed between 0,01-0,08%. The decrease in the number of salmonellosis cases was mainly due to the decrease in the number of cases caused by S. Enteritidis. Eighty percent of the cases were sporadic. There were 6-700 community/institutional and family acquired outbreaks recorded. The number of the outbreaks declined more significantly than that of the sporadic cases. The investigation of the outbreaks has showed that in most cases the source of the infection was poultry. Mainly poultry eggs, and foods that contained eggs used without adequate heat-treatment and that were prepared at privet home or at canteen/catering trade caused outbreaks. There were only very few outbreaks caused by foods of industrial origin in the past ten years and there were no outbreaks caused by contaminated water.

Relevance as zoonotic disease

In the outbreaks a person-to-person transmission has been detected only in very few cases (in specific communities). In most case the outbreaks were suspectedly or conformedly caused by strains originated from poultry, via contaminated food.

Additional information

At the Phage-typing and Molecular Epidemiology Department of the 'Johan Bela' National Center for Epidemiology, the phage typing reactions for S. Enteritidis and S. Typhimurium are prepared parallel both with a Hungarian and the international (Ward et al., Colindale) and the Felix-Callow as well as Anderson et al. sets of phages, respectively.

2.1.3 Salmonella in foodstuffs

A. Salmonella spp. in broiler meat and products thereof

Monitoring system

Sampling strategy

At slaughterhouse and cutting plant

The sampling strategy in the slaughterhouses is based on the previous years' data on production volume. The monitoring plan prepared by the CAO Food and Feed Safety Directorate determines the number of samples/county/month. The monitoring samples are thrown by the regional veterinary authority and are examined in the official control laboratories belonging to the Central Agricultural Office (CAO). It is a permanent monitoring scheme, data are reported by the official laboratories to CAO and the Ministry of Agriculture and Rural Development in the frame of an annual laboratory report. All the Salmonella strains isolated are serotyped by the NRL Salmonella.

At meat processing plant

The sampling strategy in processing plants is randomised based on the previous years' data on production volume. The samles are thrown by the veterinary authority and are examined in the official food control laboratory. It is a permanent monitoring scheme, data are reported by the official laboratories to the Ministry of Agriculture and Rural Development in the frame of an annual laboratory report.

At retail

Retail is also sampled by the authority on a regular basis. The total number of samples is determentd in the annual monitoring plan. About 60 % of the official control samples in a product group are taken at retail.

Frequency of the sampling

At slaughterhouse and cutting plant

Sampling distributed evenly throughout the year

At meat processing plant

Sampling distributed evenly throughout the year

At retail

Sampling distributed evenly throughout the year

Type of specimen taken

At slaughterhouse and cutting plant

Fresh meat

At meat processing plant

minced meat, meat prep., meat products

At retail

minced meat, meat prep., meat products

Methods of sampling (description of sampling techniques)

At slaughterhouse and cutting plant

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At least 500 grams of meat is sent to the laboratory. The test portion is 25 grams.

At meat processing plant

Batch sampling with 5 subsamples. Test portion is 5 x 10 or 25 grams according to Regulation 2073/2005/EC.

Definition of positive finding

At slaughterhouse and cutting plant

a sample or a batch is positive if salmonella was isolated

At meat processing plant

a sample or a batch is positive if salmonella was isolated

At retail

a sample or a batch is positive if salmonella was isolated

Diagnostic/analytical methods used

At slaughterhouse and cutting plant

Bacteriological method: ISO 6579:2002

At meat processing plant

Bacteriological method: ISO 6579:2002

At retail

Bacteriological method: ISO 6579:2002

Preventive measures in place

According to 2073/2005/EC Reg.

Measures in case of the positive findings or single cases

According to Reg.2073/2005/EC.

National evaluation of the recent situation, the trends and sources of infection

Based on the monitoring results, salmonella prevalence is high in broiler meat in Hungary. The dominance of Salmonella Infantis strains is well-known in the past years. 90 % of the isolated strains are belonging to this serovar now.

From 1995, the rate of Salmonella Infantis/Enteritidis is showing a continuous increase for Infantis (1% to 90 %), and a decreasing trend for S. Enteritidis (from 60 % to 5%).

The marked increase of Salmonella Infantis serovar in broiler meat was not caused a significant increase in human Salmonella Infantis incidence. The dominating serovar in human infections is continuously S. Enteritidis wich has been responsible for 70-80 % of the human infections for many years.

B. Salmonella spp. in pig meat and products thereof

Monitoring system

Sampling strategy

At slaughterhouse and cutting plant

The sampling strategy in the slaughterhouses is based on the previous years' data on production volume. The monitoring plan prepared by the CAO Food and Feed Safety Directorate determines the number of samples/county/month. The monitoring samples are thrown by the regional veterinary authority and are examined in the official control laboratories belonging to the Central Agricultural Office (CAO). It is a permanent monitoring scheme, data are reported by the official laboratories to CAO and the Ministry of Agricilture and Regional Development in the frame of an annual laboratory report. All the Salmonella strains isolated are serotyped by the NRL Salmonella.

At meat processing plant

The sampling strategy in processing plants is randomised based on the previous years' data on production volume. The samles are thrown by the veterinary authority and are examined in the official food control laboratory. It is a permanent monitoring scheme, data are reported by the official laboratories to the Ministry of Agricilture and Regional Development in the frame of an annual laboratory report.

Frequency of the sampling

At slaughterhouse and cutting plant

Sampling distributed evenly throughout the year

At meat processing plant

Sampling distributed evenly throughout the year

Type of specimen taken

At slaughterhouse and cutting plant

Fresh meat

At meat processing plant

Surface of carcass

Diagnostic/analytical methods used

At slaughterhouse and cutting plant

Bacteriological method: ISO 6579:2002

At meat processing plant

Bacteriological method: NMKL No 71:1999

C. Salmonella spp. in bovine meat and products thereof

Monitoring system

Sampling strategy

At slaughterhouse and cutting plant

Food business operators perform continuous sampling system determined in their HACCP plans, and nearby there is an official control system of the competent authorities with a randomised sampling as well. The data of self control processes are checked in the frame of official control of course, but are not collected to a database, therefore these are not involved in this report. The test results of samples examined by competent authorities in their own laboratories are reported, but the data collection system do not allow to report the data separately for te different stages of food chain (slaughterhouses, processing plants, retail). Based on the structure of the EU zoonosis report, the data collection system will be resturctured this year. This year all the data on fresh meat are reported in the table of slaughterhouses.

At meat processing plant

The sampling strategy is randomised and continuous, performed by the competent authorities. Food producers operate their own continuous sampling system determined in their HACCP plans as well, with the same remarks as in the case of slaughterhouses.

Frequency of the sampling

At slaughterhouse and cutting plant

Sampling distributed evenly throughout the year

At meat processing plant

Sampling distributed evenly throughout the year

At retail

Sampling distributed evenly throughout the year

Type of specimen taken

At slaughterhouse and cutting plant

Fresh meat

At meat processing plant

Surface of carcass

At retail

fresh meat and all kinds of meat products

Methods of sampling (description of sampling techniques)

At slaughterhouse and cutting plant

500 garms of sample is sent to the laboratory, the test portion is 25 grams

At meat processing plant

Batch sampling with 5 subsamples. Test portion is 10 or 25 grams determined by 2073/2005/EC Regulation.

Diagnostic/analytical methods used

At slaughterhouse and cutting plant

Bacteriological method: ISO 6579:2002

At meat processing plant

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Bacteriological method: ISO 6579:2002

At retail

Bacteriological method: ISO 6579:2002

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Sample weight	Units tested	Total units positive for Salmonella	S. Enteritidis	S. Typhimurium
Meat from broilers (Gallus gallus) - carcase - Slaughterhouse - Surveillance	NFCSO	Objective sampling	Official sampling	food sample > neck skin	Domestic	Single	25 g	213	37	0	0
Meat from broilers (Gallus gallus) - fresh - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	263	61	0	0
Meat from broilers (Gallus gallus) - fresh - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	325	106	0	0
Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	41	7	0	0
Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	80	24	0	0
Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	79	1	0	0
Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	182	0	0	0
Meat from broilers (Gallus gallus) - meat products - raw but intended to be eaten cooked - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	16	0	0	0
Meat from broilers (Gallus gallus) - meat products - raw but intended to be eaten cooked - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	112	2	0	0
Meat from broilers (Gallus gallus) - minced meat - intended to be eaten cooked - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	4	4	0	0

0

0

2

1

S. Total units Sample type Sample origin Sampling unit Source of Sampling Sample Units tested positive for S. Enteritidis Typhimurium Sampler information strategy weight Salmonella Meat from broilers (Gallus gallus) - minced meat -Official Objective intended to be eaten cooked - Retail - Surveillance **NFCSO** 2 0 0 0 food sample Single 25 g sampling sampling Objective Official food sample Meat from turkey - carcase - Slaughterhouse -**NFCSO** Domestic Single 25 g 81 12 0 0 sampling sampling Surveillance > neck skin Meat from turkey - fresh - Processing plant -Objective Official **NFCSO** Domestic 119 16 0 0 food sample Single 25 g Surveillance sampling sampling Official Objective **NFCSO** food sample 0 Meat from turkey - fresh - Retail - Surveillance Single 25 g 172 18 0 sampling sampling Official Meat from turkey - meat products - cooked, ready-to Objective **NFCSO** 0 0 food sample 25 g 74 0 Single -eat - Processing plant - Surveillance sampling sampling Meat from turkey - meat products - cooked, ready-to Objective Official **NFCSO** 0 food sample Single 25 g 177 0 0 -eat - Retail - Surveillance sampling sampling Meat from turkey - meat products - raw but intended Official Objective to be eaten cooked - Processing plant - Surveillance **NFCSO** food sample Single 25 g 12 0 0 0 sampling sampling Meat from turkey - meat products - raw but intended Official Objective **NFCSO** to be eaten cooked - Retail - Surveillance food sample Single 25 g 77 0 0 sampling sampling Official Meat from duck - carcase - Slaughterhouse -Objective **NFCSO** food sample Single 25 g 33 0 2 Surveillance sampling sampling Meat from geese - carcase - Slaughterhouse -Objective Official **NFCSO** food sample Single 25 g 21 3 0 1 Surveillance sampling sampling

Official

sampling

Official

sampling

food sample

food sample

Single

Single

25 g

25 g

118

18

10

2

Objective

sampling Objective

sampling

NFCSO

NFCSO

Meat from duck - fresh - Retail - Surveillance

Meat from geese - fresh - Retail - Surveillance

S. Total units Sample type Sample origin Sampling unit Source of Sampling Sample Sampler Units tested positive for S. Enteritidis Typhimurium information weight strategy Salmonella Official Meat from turkey - meat preparation - intended to Objective NFCSO 0 food sample Single 25 g 33 2 0 sampling be eaten cooked - in total - Surveillance sampling Meat from turkey - minced meat - intended to be Objective Official NFCSO food sample Domestic 25 g 14 0 0 0 Single eaten cooked - Processing plant - Surveillance sampling sampling Meat from turkey - minced meat - intended to be Objective Official NFCSO 0 0 99 23 food sample Single 25 g sampling eaten cooked - Retail - Surveillance sampling Meat from wild game - birds - fresh - in total -Objective Official NFCSO food sample 25 g 24 2 0 1 Single sampling Surveillance sampling

	S. 1,4,[5],12:i: -	Salmonella spp., unspecified	Other serovars	S. Bredeney	S. Infantis	S. Kentucky	S. Stanley
Meat from broilers (Gallus gallus) - carcase - Slaughterhouse - Surveillance	0	0	0	0	37	0	0
Meat from broilers (Gallus gallus) - fresh - Processing plant - Surveillance	0	0	4	0	57	0	0
Meat from broilers (Gallus gallus) - fresh - Retail - Surveillance	0	2	0	0	101	3	0
Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - Processing plant - Surveillance	0	0	1	0	6	0	0
Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - Retail - Surveillance	0	1	1	0	19	3	0

	S. 1,4,[5],12:i: -	Salmonella spp., unspecified	Other serovars	S. Bredeney	S. Infantis	S. Kentucky	S. Stanley
Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Processing plant - Surveillance	0	0	0	0	1	0	0
Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Retail - Surveillance	0	0	0	0	0	0	0
Meat from broilers (Gallus gallus) - meat products - raw but intended to be eaten cooked - Processing plant - Surveillance	0	0	0	0	0	0	0
Meat from broilers (Gallus gallus) - meat products - raw but intended to be eaten cooked - Retail - Surveillance	0	0	0	0	1	0	1
Meat from broilers (Gallus gallus) - minced meat - intended to be eaten cooked - Processing plant - Surveillance	0	0	0	0	4	0	0
Meat from broilers (Gallus gallus) - minced meat - intended to be eaten cooked - Retail - Surveillance	0	0	0	0	0	0	0
Meat from turkey - carcase - Slaughterhouse - Surveillance	0	0		1	2	7	2
Meat from turkey - fresh - Processing plant - Surveillance	0	0	3	1	8	1	3
Meat from turkey - fresh - Retail - Surveillance	0	0	6	1	3	4	4
Meat from turkey - meat products - cooked, ready-to -eat - Processing plant - Surveillance	0	0	0	0	0	0	0
Meat from turkey - meat products - cooked, ready-to -eat - Retail - Surveillance	0	0	0	0	0	0	0

	S. 1,4,[5],12:i: -	Salmonella spp., unspecified	Other serovars	S. Bredeney	S. Infantis	S. Kentucky	S. Stanley
Meat from turkey - meat products - raw but intended to be eaten cooked - Processing plant - Surveillance	0	0	0	0	0	0	0
Meat from turkey - meat products - raw but intended to be eaten cooked - Retail - Surveillance	0	0	0	0	1	0	0
Meat from duck - carcase - Slaughterhouse - Surveillance	0	0	2	0	0	0	0
Meat from geese - carcase - Slaughterhouse - Surveillance	0	0	2	0	0	0	0
Meat from duck - fresh - Retail - Surveillance	0	0	7	0	1	0	0
Meat from geese - fresh - Retail - Surveillance	0	0	1	0	0	0	0
Meat from turkey - meat preparation - intended to be eaten cooked - in total - Surveillance	0	0	0	0	1	1	0
Meat from turkey - minced meat - intended to be eaten cooked - Processing plant - Surveillance	0	0	0	0	0	0	0
Meat from turkey - minced meat - intended to be eaten cooked - Retail - Surveillance	0	1	2	1	2	4	13
Meat from wild game - birds - fresh - in total - Surveillance	1	0	0	0	0	0	0

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Sample weight	Units tested	Total units positive for Salmonella	S. Enteritidis	S. Typhimurium
Milk, cows' - raw milk - intended for direct human consumption - Farm - Surveillance	NFCSO	Objective sampling	Official sampling	food sample > milk	Domestic	Single	25 ml	194	0	0	0
Milk, goats' - raw milk - intended for direct human consumption - Farm - Surveillance	NFCSO	Objective sampling	Official sampling	food sample > milk	Domestic	Single	25 ml	4	0	0	0
Dairy products (excluding cheeses) - milk powder and whey powder - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	8	0	0	0
Dairy products (excluding cheeses) - milk powder and whey powder - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	62	0	0	0
Cheeses made from cows' milk - curd - in total - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	85	0	0	0
Cheeses made from cows' milk - fresh - made from pasteurised milk - in total - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	12	0	0	0
Cheeses made from cows' milk - hard - made from pasteurised milk - in total - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	16	0	0	0
Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - in total - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	175	0	0	0
Cheeses made from goats' milk - in total	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	7	0	0	0
Cheeses made from sheep's milk - fresh - made from pasteurised milk - in total - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	65	0	0	0
Cheeses made from sheep's milk - soft and semi- soft - made from pasteurised milk - in total - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	3	0	0	0

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Sample weight	Units tested	Total units positive for Salmonella	S. Enteritidis	S. Typhimurium
Dairy products (excluding cheeses) - dairy desserts - in total - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	119	0	0	0
Dairy products (excluding cheeses) - fermented dairy products - in total - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	28	0	0	0
Dairy products (excluding cheeses) - ice-cream - made from pasteurised milk - Catering - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	49	0	0	0
Dairy products (excluding cheeses) - ice-cream - made from pasteurised milk - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	37	0	0	0
Dairy products (excluding cheeses) - ice-cream - made from pasteurised milk - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	287	0	0	0

	S. 1,4,[5],12:i: -	Salmonella spp., unspecified
Milk, cows' - raw milk - intended for direct human consumption - Farm - Surveillance	0	0
Milk, goats' - raw milk - intended for direct human consumption - Farm - Surveillance	0	0
Dairy products (excluding cheeses) - milk powder and whey powder - Processing plant - Surveillance	0	0
Dairy products (excluding cheeses) - milk powder and whey powder - Retail - Surveillance	0	0

	S. 1,4,[5],12:i: -	Salmonella spp., unspecified
Cheeses made from cows' milk - curd - in total - Surveillance	0	0
Cheeses made from cows' milk - fresh - made from pasteurised milk - in total - Surveillance	0	0
Cheeses made from cows' milk - hard - made from pasteurised milk - in total - Surveillance	0	0
Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - in total - Surveillance	0	0
Cheeses made from goats' milk - in total	0	0
Cheeses made from sheep's milk - fresh - made from pasteurised milk - in total - Surveillance	0	0
Cheeses made from sheep's milk - soft and semi- soft - made from pasteurised milk - in total - Surveillance	0	0
Dairy products (excluding cheeses) - dairy desserts - in total - Surveillance	0	0
Dairy products (excluding cheeses) - fermented dairy products - in total - Surveillance	0	0
Dairy products (excluding cheeses) - ice-cream - made from pasteurised milk - Catering - Surveillance	0	0
Dairy products (excluding cheeses) - ice-cream - made from pasteurised milk - Processing plant - Surveillance	0	0

	S. 1,4,[5],12:i: -	Salmonella spp., unspecified
Dairy products (excluding cheeses) - ice-cream - made from pasteurised milk - Retail - Surveillance	0	0

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Sample weight	Units tested	Total units positive for Salmonella	S. Enteritidis	S. Typhimurium
Eggs - table eggs - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Batch	25 g	458	0	0	0
Eggs - raw material (liquid egg) for egg products - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 ml	49	0	0	0
Fishery products, unspecified - cooked - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	41	0	0	0
Fish - smoked - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	71	0	0	0
Crustaceans - unspecified - cooked - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	13	0	0	0
Molluscan shellfish - raw - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	9	0	0	0
Molluscan shellfish - cooked - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	51	0	0	0
Seeds, sprouted - ready-to-eat - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	1	0	0	0
Seeds, sprouted - ready-to-eat - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	69	0	0	0
Vegetables - pre-cut - ready-to-eat - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	205	0	0	0
Infant formula - dried - intended for infants below 6 months - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	50	0	0	0
Bakery products - cakes - Catering - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	172	0	0	0
Bakery products - cakes - Retail - Surveillance	NFCSO	Objective	Official sampling	food sample		Single	25 g	158	1	1	0

Total units Sample type Sample origin Sampling unit Source of Sampling Sample Units tested positive for S. Enteritidis Typhimurium Sampler information strategy weight Salmonella Cereals and meals - flakes - in total - Surveillance Objective Official NFCSO food sample 55 0 0 Sinale 25 g 0 sampling sampling Official Objective Chocolate - in total - Surveillance **NFCSO** food sample Sinale 25 a 251 0 0 0 sampling sampling Official Cocoa and cocoa preparations, coffee and tea - in Objective NFCSO food sample Single 25 g 266 0 0 0 total - Surveillance sampling sampling Official Coconut - coconut products - in total - Surveillance Objective **NFCSO** food sample Sinale 25 a 78 0 0 0 sampling sampling Official Objective **NFCSO** food sample 56 0 0 Egg products - dried - in total - Surveillance Single 25 g 1 sampling sampling Egg products - liquid - Processing plant -Objective Official **NFCSO** food sample Domestic Single 25 ml 25 0 0 0 Surveillance sampling sampling Objective Official **NFCSO** food sample 73 0 0 0 Follow-on formulae - Retail - Surveillance Sinale 25 g sampling sampling Foodstuffs intended for special nutritional uses - in Objective Official NECSO 0 0 food sample Single 25 g 32 total - Surveillance sampling sampling Foodstuffs intended for special nutritional uses -Official Objective NFCSO processed cereal-based food for infants and young 18 0 0 food sample Single 25 g 0 sampling sampling children - Retail - Surveillance Official Objective NFCSO 36 0 0 Fruits - non-pre-cut - Retail - Surveillance food sample Single 25 g 0 sampling sampling Nuts and nut products - dried - in total - Surveillance Objective Official NFCSO 0 95 0 0 food sample Single 25 g sampling sampling Other processed food products and prepared dishes Objective Official **NFCSO** 394 0 0 - meat based dishes - in total - Surveillance food sample Single 25 g 0 sampling sampling Other processed food products and prepared dishes Official Objective **NFCSO** - sandwiches - with meat - in total - Surveillance food sample Single 25 g 243 0 0 0 sampling sampling

S. Total units Sample type Sample origin Sampling unit Sampling Sample Source of Units tested Sampler positive for S. Enteritidis Typhimurium information strategy weight Salmonella Official Objective NFCSO food sample Ready-to-eat salads - in total - Surveillance Single 25 g 389 0 0 sampling sampling Objective Official Spices and herbs - dried - in total - Surveillance NFCSO food sample 25 g 0 0 0 Single 187 sampling sampling

	S. 1,4,[5],12:i: -	Salmonella spp., unspecified	S. Mbandaka
Eggs - table eggs - Retail - Surveillance	0	0	0
Eggs - raw material (liquid egg) for egg products - Processing plant - Surveillance	0	0	0
Fishery products, unspecified - cooked - Retail - Surveillance	0	0	0
Fish - smoked - Retail - Surveillance	0	0	0
Crustaceans - unspecified - cooked - Retail - Surveillance	0	0	0
Molluscan shellfish - raw - Retail - Surveillance	0	0	0
Molluscan shellfish - cooked - Retail - Surveillance	0	0	0
Seeds, sprouted - ready-to-eat - Processing plant - Surveillance	0	0	0
Seeds, sprouted - ready-to-eat - Retail - Surveillance	0	0	0
Vegetables - pre-cut - ready-to-eat - Retail - Surveillance	0	0	0

	S. 1,4,[5],12:i: -	Salmonella spp., unspecified	S. Mbandaka
Infant formula - dried - intended for infants below 6 months - Retail - Surveillance	0	0	0
Bakery products - cakes - Catering - Surveillance	0	0	0
Bakery products - cakes - Retail - Surveillance	0	0	0
Cereals and meals - flakes - in total - Surveillance	0	0	0
Chocolate - in total - Surveillance	0	0	0
Cocoa and cocoa preparations, coffee and tea - in total - Surveillance	0	0	0
Coconut - coconut products - in total - Surveillance	0	0	0
Egg products - dried - in total - Surveillance	0	0	1
Egg products - liquid - Processing plant - Surveillance	0	0	0
Follow-on formulae - Retail - Surveillance	0	0	0
Foodstuffs intended for special nutritional uses - in total - Surveillance	0	0	0
Foodstuffs intended for special nutritional uses - processed cereal-based food for infants and young children - Retail - Surveillance	0	0	0
Fruits - non-pre-cut - Retail - Surveillance	0	0	0
Nuts and nut products - dried - in total - Surveillance	0	0	0

	S. 1,4,[5],12:i: -	Salmonella spp., unspecified	S. Mbandaka
Other processed food products and prepared dishes - meat based dishes - in total - Surveillance	0	0	0
Other processed food products and prepared dishes - sandwiches - with meat - in total - Surveillance	0	0	0
Ready-to-eat salads - in total - Surveillance	0	1	0
Spices and herbs - dried - in total - Surveillance	0	0	0

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Sample weight	Units tested	Total units positive for Salmonella	S. Enteritidis	S. Typhimurium
Meat from pig - carcase - Slaughterhouse - Surveillance	NFCSO	Objective sampling	Official sampling	food sample > carcase swabs	Domestic	Single	400 cm2	239	2	0	2
Meat from pig - fresh - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	182	2	0	1
Meat from pig - fresh - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	193	4	0	1
Meat from pig - minced meat - intended to be eaten cooked - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	10 g	53	0	0	0
Meat from pig - minced meat - intended to be eaten cooked - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	10 g	172	6	0	1
Meat from pig - meat preparation - intended to be eaten cooked - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	10 g	81	3	0	0
Meat from pig - meat preparation - intended to be eaten cooked - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	10 g	103	2	0	2
Meat from pig - meat products - raw but intended to be eaten cooked - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	7	0	0	0
Meat from pig - meat products - raw but intended to be eaten cooked - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	20	0	0	0
Meat from pig - meat products - cooked, ready-to- eat - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	54	0	0	0
Meat from pig - meat products - cooked, ready-to- eat - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	116	0	0	0
Meat from bovine animals - carcase - Slaughterhouse - Surveillance	NFCSO	Objective sampling	Official sampling	food sample > carcase	Domestic	Single	400 cm2	233	0	0	0

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Sample weight	Units tested	Total units positive for Salmonella	S. Enteritidis	S. Typhimurium
Meat from bovine animals - fresh - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	97	2	0	1
Meat from bovine animals - fresh - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	190	1	0	0
Meat from bovine animals - minced meat - intended to be eaten cooked - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	10 g	14	0	0	0
Meat from bovine animals - minced meat - intended to be eaten cooked - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	10 g	133	0	0	0
Meat from bovine animals - meat products - raw but intended to be eaten cooked - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	9	0	0	0
Meat from bovine animals - meat products - raw but intended to be eaten cooked - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	48	0	0	0
Meat from bovine animals - meat products - cooked, ready-to-eat - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	2	0	0	0
Meat from bovine animals - meat products - cooked, ready-to-eat - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	42	0	0	0
Meat from sheep - fresh - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	5	0	0	0
Other products of animal origin - gelatin and collagen - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	63	0	0	0
Meat from horse - meat products - fermented sausages - in total - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	3	0	0	0

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Sample weight	Units tested	Total units positive for Salmonella	S. Enteritidis	S. Typhimurium
Meat from pig - meat products - fermented sausages - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	443	5	0	2
Meat from pig - meat products - fermented sausages - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	568	5	0	1
Meat from pig - meat products - raw and intended to be eaten raw - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	88	0	0	0
Meat from pig - meat products - raw and intended to be eaten raw - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	56	0	0	0
Meat from wild boar - fresh - in total - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	54	1	1	0
Meat from wild game - land mammals - fresh - in total - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	75	0	0	0

	S. 1,4,[5],12:i: -	Salmonella spp., unspecified	Other serovars	S. Bredeney	S. Derby
Meat from pig - carcase - Slaughterhouse - Surveillance	0	0	0	0	0
Meat from pig - fresh - Processing plant - Surveillance	0	0	0	0	1
Meat from pig - fresh - Retail - Surveillance	0	0	0	1	2
Meat from pig - minced meat - intended to be eaten cooked - Processing plant - Surveillance	0	0	0	0	0

	S. 1,4,[5],12:i: -	Salmonella spp., unspecified	Other serovars	S. Bredeney	S. Derby
Meat from pig - minced meat - intended to be eaten cooked - Retail - Surveillance	2	0	1	0	2
Meat from pig - meat preparation - intended to be eaten cooked - Processing plant - Surveillance	0	0	1	0	2
Meat from pig - meat preparation - intended to be eaten cooked - Retail - Surveillance	0	0	0	0	0
Meat from pig - meat products - raw but intended to be eaten cooked - Processing plant - Surveillance	0	0	0	0	0
Meat from pig - meat products - raw but intended to be eaten cooked - Retail - Surveillance	0	0	0	0	0
Meat from pig - meat products - cooked, ready-to- eat - Processing plant - Surveillance	0	0	0	0	0
Meat from pig - meat products - cooked, ready-to- eat - Retail - Surveillance	0	0	0	0	0
Meat from bovine animals - carcase - Slaughterhouse - Surveillance	0	0	0	0	0
Meat from bovine animals - fresh - Processing plant - Surveillance	0	0	0	0	1
Meat from bovine animals - fresh - Retail - Surveillance	0	0	0	0	1
Meat from bovine animals - minced meat - intended to be eaten cooked - Processing plant - Surveillance	0	0	0	0	0

	S. 1,4,[5],12:i: -	Salmonella spp., unspecified	Other serovars	S. Bredeney	S. Derby
Meat from bovine animals - minced meat - intended to be eaten cooked - Retail - Surveillance	0	0	0	0	0
Meat from bovine animals - meat products - raw but intended to be eaten cooked - Processing plant - Surveillance	0	0	0	0	0
Meat from bovine animals - meat products - raw but intended to be eaten cooked - Retail - Surveillance	0	0	0	0	0
Meat from bovine animals - meat products - cooked, ready-to-eat - Processing plant - Surveillance	0	0	0	0	0
Meat from bovine animals - meat products - cooked, ready-to-eat - Retail - Surveillance	0	0	0	0	0
Meat from sheep - fresh - Retail - Surveillance	0	0	0	0	0
Other products of animal origin - gelatin and collagen - Retail - Surveillance	0	0	0	0	0
Meat from horse - meat products - fermented sausages - in total - Surveillance	0	0	0	0	0
Meat from pig - meat products - fermented sausages - Processing plant - Surveillance	1	0	2	0	0
Meat from pig - meat products - fermented sausages - Retail - Surveillance	1	0	3	0	0
Meat from pig - meat products - raw and intended to be eaten raw - Processing plant - Surveillance	0	0	0	0	0

Table Salmonella in red meat and products thereof

	S. 1,4,[5],12:i: -	Salmonella spp., unspecified	Other serovars	S. Bredeney	S. Derby
Meat from pig - meat products - raw and intended to be eaten raw - Retail - Surveillance	0	0	0	0	0
Meat from wild boar - fresh - in total - Surveillance	0	0	0	0	0
Meat from wild game - land mammals - fresh - in total - Surveillance	0	0	0	0	0

2.1.4 Salmonella in animals

Table Salmonella in breeding flocks of Gallus gallus

	No of flocks under control programme	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Target Verification	Sampling unit	Units tested	Total units positive for Salmonella	S. Enteritidis
Gallus gallus (fowl) - breeding flocks, unspecified - adult - Control and eradication programmes	890	county reports	Census	Official and industry sampling	environmenta I sample > boot swabs and dust	Domestic	yes	Flock	890	17	7
Gallus gallus (fowl) - breeding flocks, unspecified - day-old chicks - Farm - Control and eradication programmes	108	county reports	Census	Industry sampling	environmenta I sample > boot swabs and dust	Domestic	no	Flock	108	0	0
Gallus gallus (fowl) - breeding flocks, unspecified - during rearing period - Farm - Control and eradication programmes	277	county reports	Census	Official and industry sampling	environmenta I sample > boot swabs and dust	Domestic	no	Flock	277	0	0

	S. Hadar	S. Infantis	S. Typhimurium	S. Virchow	S. 1,4,[5],12:i: -	Salmonella spp., unspecified
Gallus gallus (fowl) - breeding flocks, unspecified - adult - Control and eradication programmes	0	4	3	0	0	3
Gallus gallus (fowl) - breeding flocks, unspecified - day-old chicks - Farm - Control and eradication programmes	0	0	0	0	0	0
Gallus gallus (fowl) - breeding flocks, unspecified - during rearing period - Farm - Control and eradication programmes	0	0	0	0	0	0

Table Salmonella in other birds

		Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Units tested	Total units positive for Salmonella	S. Enteritidis	S. Typhimurium	S. 1,4,[5],12:i: -	
	Partridges - farmed - Farm - Monitoring	NFCSO VDD	Unspecified	Not	animal	Domestic	Animal	16	0	0	0	0	
L			·	applicable	sample								1
	Dhaanata wild	NECCO VED		Not	animal	D	A	404	24	_	_	0	
	Pheasants - wild	NFCSO VDD	Unspecified	applicable	sample	Domestic	Animal	134	34	U	U	0	
	D' F	NEODO VIDIO		Not	animal	D	A - ' I	47	_		0	0	
	Pigeons - Farm	NFCSO VDD	Unspecified	applicable	sample	Domestic	Animal	17	/	0	U	0	

	Salmonella spp., unspecified
Partridges - farmed - Farm - Monitoring	0
Pheasants - wild	34
Pigeons - Farm	7

Table Salmonella in other animals

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Units tested	Total units positive for Salmonella	S. Enteritidis	S. Typhimurium	S. 1,4,[5],12:i: -
Cattle (bovine animals) - Farm - Clinical investigations	NFCSO VDD	Suspect sampling	Not applicable	animal sample	Domestic	Animal	73	25	1	13	0
Pigs - Farm - Clinical investigations	NFCSO VDD	Suspect sampling	Not applicable	animal sample	Domestic	Animal	138	52	0	11	0

	Salmonella spp., unspecified
Cattle (bovine animals) - Farm - Clinical investigations	11
Pigs - Farm - Clinical investigations	41

Table Salmonella in other poultry

	No of flocks under control programme	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Target Verification	Sampling unit	Units tested	Total units positive for Salmonella	S. Enteritidis
Gallus gallus (fowl) - laying hens - day-old chicks - Control and eradication programmes	14	county reports	Census	Industry sampling	environmenta I sample > delivery box liner	Domestic	no	Flock	14	2	0
Gallus gallus (fowl) - laying hens - during rearing period - Control and eradication programmes	53	county reports	Census	Official and industry sampling	environmenta I sample > boot swabs and dust	Domestic	no	Flock	53	1	1
Gallus gallus (fowl) - laying hens - adult - Farm - Control and eradication programmes	1055	county reports	Census	Official and industry sampling	environmenta I sample > boot swabs and dust	Domestic	yes	Flock	1055	68	19
Gallus gallus (fowl) - broilers - day-old chicks - Control and eradication programmes	7873	county reports	Unspecified	Industry sampling	environmenta I sample > delivery box liner	Domestic	yes	Flock	109	5	2
Gallus gallus (fowl) - broilers - before slaughter - Farm - Control and eradication programmes	7873	county reports	Census	Official and industry sampling	environmenta I sample > boot swabs and dust	Domestic	yes	Flock	7873	1274	3
Turkeys - breeding flocks, unspecified - during rearing period - Farm - Control and eradication programmes	41	county reports	Census	Official and industry sampling	environmenta I sample > boot swabs and dust	Domestic	no	Flock	41	0	0
Turkeys - breeding flocks, unspecified - adult - Farm - Control and eradication programmes	212	county reports	Census	Official and industry sampling	environmenta I sample > boot swabs and dust	Domestic	yes	Flock	212	51	0
Turkeys - fattening flocks - before slaughter - Farm - Control and eradication programmes	2456	county reports	Census	Official and industry sampling	environmenta I sample > boot swabs and dust	Domestic	yes	Flock	2456	876	1

Table Salmonella in other poultry

	S. Typhimurium	S. 1,4,[5],12:i: -	Salmonella spp., unspecified
Gallus gallus (fowl) - laying hens - day-old chicks - Control and eradication programmes	2	0	0
Gallus gallus (fowl) - laying hens - during rearing period - Control and eradication programmes	0	0	0
Gallus gallus (fowl) - laying hens - adult - Farm - Control and eradication programmes	2	0	47
Gallus gallus (fowl) - broilers - day-old chicks - Control and eradication programmes	3	0	0
Gallus gallus (fowl) - broilers - before slaughter - Farm - Control and eradication programmes	4	0	1267
Turkeys - breeding flocks, unspecified - during rearing period - Farm - Control and eradication programmes	0	0	0
Turkeys - breeding flocks, unspecified - adult - Farm - Control and eradication programmes	0	0	51
Turkeys - fattening flocks - before slaughter - Farm - Control and eradication programmes	0	0	875

2.1.5 Salmonella in feedingstuffs

Table Salmonella in compound feedingstuffs

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Sample weight	Units tested	Total units positive for Salmonella	S. Enteritidis	S. Typhimurium
Compound feedingstuffs for cattle - final product - Feed mill - Surveillance	NFCSO FFSD	Objective sampling	Official sampling	feed sample	Unknown	Batch	1 kg	33	1	0	0
Compound feedingstuffs for pigs - final product - Feed mill - Surveillance	NFCSO FFSD	Objective sampling	Official sampling	feed sample	Unknown	Batch	1 kg	183	2	1	0
Compound feedingstuffs for poultry (non specified) - final product - Feed mill - Surveillance	NFCSO FFSD	Objective sampling	Official sampling	feed sample	Unknown	Batch	1 kg	62	0	0	0
Compound feedingstuffs for poultry - breeders - final product - Feed mill - Surveillance	NFCSO FFSD	Objective sampling	Official sampling	feed sample	Unknown	Batch	1 kg	6	0	0	0
Compound feedingstuffs for poultry - laying hens - final product - Feed mill - Surveillance	NFCSO FFSD	Objective sampling	Official sampling	feed sample	Unknown	Batch	1 kg	47	2	0	0
Compound feedingstuffs for poultry - broilers - final product - Feed mill - Surveillance	NFCSO FFSD	Objective sampling	Official sampling	feed sample	Unknown	Batch	1 kg	71	0	0	0

	S. 1,4,[5],12:i: -	Salmonella spp., unspecified	S. Lille	S. Loenga	S. Senftenberg
Compound feedingstuffs for cattle - final product - Feed mill - Surveillance	0	0	0	1	0
Compound feedingstuffs for pigs - final product - Feed mill - Surveillance	0	0	0	0	1

Table Salmonella in compound feedingstuffs

	S. 1,4,[5],12:i: -	Salmonella spp., unspecified	S. Lille	S. Loenga	S. Senftenberg
Compound feedingstuffs for poultry (non specified) - final product - Feed mill - Surveillance	0	0	0	0	0
Compound feedingstuffs for poultry - breeders - final product - Feed mill - Surveillance	0	0	0	0	0
Compound feedingstuffs for poultry - laying hens - final product - Feed mill - Surveillance	0	0	1	0	1
Compound feedingstuffs for poultry - broilers - final product - Feed mill - Surveillance	0	0	0	0	0

S. Sample type Sample origin Sampling unit Total units Source of Sampling Sample Units tested positive for S. Enteritidis Typhimurium Sampler strategy information weight Salmonella **NFCSO** Official Feed material of land animal origin - dairy products -Objective feed sample Unknown 0 0 0 Batch 1 kg 1 Feed mill - Surveillance FFSD sampling sampling **NFCSO** Official Feed material of land animal origin - meat meal -Objective feed sample 0 Unknown Batch 1 kg 13 0 0 sampling sampling Feed mill - Surveillance **FFSD** Feed material of land animal origin - animal fat -**NFCSO** Objective Official 0 0 feed sample Unknown Batch 1 kg 1 0 Feed mill - Surveillance **FFSD** sampling sampling **NFCSO** Official Feed material of marine animal origin - fish meal -Objective feed sample Unknown Batch 1 kg 3 0 0 0 Feed mill - Surveillance **FFSD** sampling sampling

	S. 1,4,[5],12:i: -	Salmonella spp., unspecified
Feed material of land animal origin - dairy products - Feed mill - Surveillance	0	0
Feed material of land animal origin - meat meal - Feed mill - Surveillance	0	0
Feed material of land animal origin - animal fat - Feed mill - Surveillance	0	0
Feed material of marine animal origin - fish meal - Feed mill - Surveillance	0	0

Table Salmonella in feed material of animal origin

Table Salmonella in other feed matter

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Sample weight	Units tested	Total units positive for Salmonella	S. Enteritidis	S. Typhimurium
Feed material of cereal grain origin - wheat derived - Feed mill - Surveillance	NFCSO FFSD	Objective sampling	Official sampling	feed sample	Unknown	Batch	1 kg	12	0		
Feed material of cereal grain origin - maize derived - Feed mill - Surveillance	NFCSO FFSD	Objective sampling	Official sampling	feed sample	Unknown	Batch	1 kg	22	0		
Feed material of oil seed or fruit origin - rape seed derived - Feed mill - Surveillance	NFCSO FFSD	Objective sampling	Official sampling	feed sample	Unknown	Batch	1 kg	1	1		
Feed material of oil seed or fruit origin - soya (bean) derived - Feed mill - Surveillance	NFCSO FFSD	Objective sampling	Official sampling	feed sample	Unknown	Batch	1 kg	4	0		
Feed material of oil seed or fruit origin - sunflower seed derived - Feed mill - Surveillance	NFCSO FFSD	Objective sampling	Official sampling	feed sample	Unknown	Batch	1 kg	4	1	0	0
Feed material of oil seed or fruit origin - other oil seeds derived - Feed mill - Surveillance	NFCSO FFSD	Objective sampling	Official sampling	feed sample	Unknown	Batch	1 kg	2	0		
Other feed material - forages and roughages - Feed mill - Surveillance	NFCSO FFSD	Objective sampling	Official sampling	feed sample	Unknown	Batch	1 kg	1	0		

	S. 1,4,[5],12:i: -	Salmonella spp., unspecified	S. Lille	S. Thompson
Feed material of cereal grain origin - wheat derived - Feed mill - Surveillance				
Feed material of cereal grain origin - maize derived - Feed mill - Surveillance				
Feed material of oil seed or fruit origin - rape seed derived - Feed mill - Surveillance				1

Table Salmonella in other feed matter

	S. 1,4,[5],12:i: -	Salmonella spp., unspecified	S. Lille	S. Thompson
Feed material of oil seed or fruit origin - soya (bean) derived - Feed mill - Surveillance				
Feed material of oil seed or fruit origin - sunflower seed derived - Feed mill - Surveillance	0	0	1	0
Feed material of oil seed or fruit origin - other oil seeds derived - Feed mill - Surveillance				
Other feed material - forages and roughages - Feed mill - Surveillance				

2.1.6 Antimicrobial resistance in Salmonella isolates

A. Antimicrobial resistance in Salmonella in poultry

Sampling strategy used in monitoring

Methods used for collecting data

Testing and data collection was the task of the NRL Salmonella.

Laboratory methodology used for identification of the microbial isolates

ISO 6579 - isolation, biochemical and serological confirmation. ISO 6579 - isolation, biochemical and serological confirmation.

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Disc diffusion method according to NCCLS is used. The inhibitive zone diameters are measured by a computerised system.

Results of the investigation

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B. Antimicrobial resistance in Salmonella in foodstuff derived from poultry

Sampling strategy used in monitoring

Frequency of the sampling

Frequency: as described previously in prevalence tables. As only Salmonella Enteritidis and Typhimurium strains are involved in the resistence monitoring program in foodstuff, and the number of isolates belonging to these serovars is very limited because of the 90% dominance of Salmonella Infantis in broiler chicken, only a limited number of isolates are available for the tests.

Type of specimen taken

Fresh meat at slaughterhouses, minced meat, meat preparations, meat products at processing level and at the market. There is no direct sampling program for antimicrobial resistance, it is connected to prevalence monitoring.

Methods of sampling (description of sampling techniques)

As described earlier.

Procedures for the selection of isolates for antimicrobial testing

S. Enteritidis and Salmonella Infantis strains are selected. All the S. Enteritidis strains of broiler origin were tested. As S. Infantis shows a characteristic dominance in Hungary, the number of the strains available is just 2000. Therefore only 10 % of the isolates were selected for testing.

Methods used for collecting data

All the strains isolated from food are serotyped in the NRL Salmonella. Antimicrobial resistence testing is performed in the NRL.

Laboratory methodology used for identification of the microbial isolates

ISO 6579 - isolation, biochemical and serological confirmation.

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Disc diffusion method according to NCCLS is used. The inhibitive zone diameters are measured by a computerised system.

Preventive measures in place

There are no specific preventive measures in place.

National evaluation of the recent situation, the trends and sources of infection

Because of the very low number of Salmonella Enteritidis isolates the information available is limited. There is no significant change in level of resistance in the past four years.

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S. Enteritidis												Gallus g	allus (fo	owl) - lay	ring hens	s											_
Isolates out of a monitoring program (yes/no)																Hungary											
Number of isolates available in the laboratory		I													1												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	2013
Aminoglycosides - Gentamicin	2	20	0									9	9	2													
Aminoglycosides - Streptomycin	16	20	0												6	14											Report
Amphenicols - Chloramphenicol	16	20	0												1	5	12	2									on tre
Cephalosporins - Cefotaxime	0.5	20	0							10	9	1															ends
Fluoroquinolones - Ciprofloxacin	0.064	20	1				5		13	1			1														and
Penicillins - Ampicillin	8	20	1										1	2	15	1			1								(A)
Quinolones - Nalidixic acid	16	20	1													17	2			1							ources
Tetracyclines - Tetracycline	8	20	1											6	12	1				1							of zo
Trimethoprim	2	20	1										19						1								zoonos
Sulfonamides - Sulfamethoxazole	256	20	2																3	10	5			2			ses

S. Enter	itidis	Gallus gallus (fowl) - laying hens							
	Isolates out of a monitoring program (yes/no)								
	Number of isolates available in the laboratory								
Antimicrob	oials:	lowest	highest						
Aminoglycosides	s - Gentamicin	0.25	32						
Aminoglycosides	2	128							
Amphenicols - C	hloramphenicol	2	64						

Table Antimicrobial susceptibility testing of S. Enteritidis in Gallus gallus (fowl) - laying hens - quantitative data [Dilution method]

S. Enteritidis	(fowl) -	gallus laying ns
Isolates out of a monitoring program (yes/no)		
Number of isolates available in the laboratory	2	0
Antimicrobials:	lowest	highest
Cephalosporins - Cefotaxime	0.06	4
Fluoroquinolones - Ciprofloxacin	0.008	8
Penicillins - Ampicillin	0.5	32
Quinolones - Nalidixic acid	4	64
Tetracyclines - Tetracycline	1	64
Trimethoprim	0.5	32
Sulfonamides - Sulfamethoxazole	8	1024

							<u> </u>	, , .																		
S. Havana												Gallus	gallus	(fowl) - b	roilers											
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory														1												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	1	0									1														
Aminoglycosides - Streptomycin	16	1	0													1										
Amphenicols - Chloramphenicol	16	1	0														1									
Cephalosporins - Cefotaxime	0.5	1	0								1															
Fluoroquinolones - Ciprofloxacin	0.064	1	0						1																	
Penicillins - Ampicillin	8	1	0												1											
Quinolones - Nalidixic acid	16	1	0													1										
etracyclines - Tetracycline	8	1	0												1											
Frimethoprim	2	1	0										1													
Sulfonamides - Sulfamethoxazole	256	1	0																		1					

S. Hava	na		gallus broilers
		ı	
Antimicrob	oials:	lowest	highest
Aminoglycosides	- Gentamicin	0.25	32
Aminoglycosides	2	128	
Amphenicols - C	hloramphenicol	2	64

Table Antimicrobial susceptibility testing of S. Havana in Gallus gallus (fowl) - broilers -

 quantitative data 	Dilution	method
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S. Hava	na		gallus broilers						
	Isolates out of a monitoring program (yes/no)								
		1							
Antimicro	in the laboratory Antimicrobials:								
Cephalosporins	- Cefotaxime	0.06	4						
Fluoroquinolone	s - Ciprofloxacin	0.008	8						
Penicillins - Amp	picillin	0.5	32						
Quinolones - Na	lidixic acid	4	64						
Tetracyclines - T	etracycline	1	64						
Trimethoprim		0.5	32						
Sulfonamides - S	Sulfamethoxazole	8	1024						

Concentration (µg/ml)	number of isolates with a concentration of inhibition equal to)

S. Mbandaka												Gallus g	allus (fo	wl) - lay	ing hens	;										
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory													4	4												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	4	0										4													
Aminoglycosides - Streptomycin	16	4	0														3	1								
Amphenicols - Chloramphenicol	16	4	0														4									
Cephalosporins - Cefotaxime	0.5	4	0							2	2															
Fluoroquinolones - Ciprofloxacin	0.064	4	0				4																			
Penicillins - Ampicillin	8	4	0											3	1											
Quinolones - Nalidixic acid	16	4	0													4										
Tetracyclines - Tetracycline	8	4	0												4											
Trimethoprim	2	4	0										4													
Sulfonamides - Sulfamethoxazole	256	4	0																	3	1					

S. Mban	daka	Gallus (fowl) - he	
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory	4	1
Antimicrob	oials:	lowest	highest
Aminoglycosides	- Gentamicin	0.25	32
Aminoglycosides	- Streptomycin	2	128
Amphenicols - Ch	nloramphenicol	2	64

Table Antimicrobial susceptibility testing of S. Mbandaka in Gallus gallus (fowl) - laying hens - quantitative data [Dilution method]

S. Mbandaka	(fowl) -	gallus laying ns
Isolates out of a monitoring program (yes/no)		
Number of isolates available in the laboratory	4	1
Antimicrobials:	lowest	highest
Cephalosporins - Cefotaxime	0.06	4
Fluoroquinolones - Ciprofloxacin	0.008	8
Penicillins - Ampicillin	0.5	32
Quinolones - Nalidixic acid	4	64
Tetracyclines - Tetracycline	1	64
Trimethoprim	0.5	32
Sulfonamides - Sulfamethoxazole	8	1024

Concentration (μ g/ml), number of isolates with a concentration of inhibition equal to

S. Anatum		Gallus gallus (fowl) - laying hens																								
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory														1												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	1	0										1													
Aminoglycosides - Streptomycin	16	1	0															1								
Amphenicols - Chloramphenicol	16	1	0														1									
Cephalosporins - Cefotaxime	0.5	1	0							1																
Fluoroquinolones - Ciprofloxacin	0.064	1	0						1																	
Penicillins - Ampicillin	8	1	0											1												
Quinolones - Nalidixic acid	16	1	0													1										
Tetracyclines - Tetracycline	8	1	0												1											
Trimethoprim	2	1	0										1													
Sulfonamides - Sulfamethoxazole	256	1	0																		1					

S. Anatu	ım	Gallus (fowl) - he	
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory		1
Antimicrob	oials:	lowest	highest
Aminoglycosides	- Gentamicin	0.25	32
Aminoglycosides	- Streptomycin	2	128
Amphenicols - Ch	hloramphenicol	2	64

Table Antimicrobial susceptibility testing of S. Anatum in Gallus gallus (fowl) - laying hens - quantitative data [Dilution method]

S. Anatu	ım	(fowl) -	gallus laying ns							
	Isolates out of a monitoring program (yes/no)									
	Number of isolates available in the laboratory		1							
Antimicrob	Antimicrobials:									
Cephalosporins -	- Cefotaxime	0.06	4							
Fluoroquinolones	s - Ciprofloxacin	0.008	8							
Penicillins - Amp	icillin	0.5	32							
Quinolones - Nal	lidixic acid	4	64							
Tetracyclines - T	etracycline	1	64							
Trimethoprim		0.5	32							
Sulfonamides - S	Sulfamethoxazole	8	1024							

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S. Indiana												Gallus g	allus (fo	wl) - layi	ing hens	;										
Isolates out of a monitoring program (yes/no)																										-
Number of isolates available in the laboratory														1												,
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	1	0										1													
Aminoglycosides - Streptomycin	16	1	0															1								
Amphenicols - Chloramphenicol	16	1	0														1									
Cephalosporins - Cefotaxime	0.5	1	0								1															
Fluoroquinolones - Ciprofloxacin	0.064	1	0						1																	
Penicillins - Ampicillin	8	1	0											1												
Quinolones - Nalidixic acid	16	1	0													1										
Tetracyclines - Tetracycline	8	1	0												1											
Trimethoprim	2	1	0										1													
Sulfonamides - Sulfamethoxazole	256	1	0																	1						

Concentration (µg/ml), number of isolates with a concentration of inhibition equal to

S. Indiar	na	Gallus (fowl) - he	0
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory		1
Antimicrob	oials:	lowest	highest
Aminoglycosides	- Gentamicin	0.25	32
Aminoglycosides	- Streptomycin	2	128
Amphenicols - C	2	64	

Table Antimicrobial susceptibility testing of S. Indiana in Gallus gallus (fowl) - laying hens - quantitative data [Dilution method]

S. Indiar	าล	(fowl) -	gallus laying ns
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory		1
Antimicrob	oials:	lowest	highest
Cephalosporins -	- Cefotaxime	0.06	4
Fluoroquinolones	s - Ciprofloxacin	0.008	8
Penicillins - Amp	icillin	0.5	32
Quinolones - Nal	lidixic acid	4	64
Tetracyclines - T	etracycline	1	64
Trimethoprim		0.5	32
Sulfonamides - S	Sulfamethoxazole	8	1024

S. Kentucky												Turk	eys - fa	ttening f	locks												
Isolates out of a monitoring program (yes/no) Number of isolates available														25													Hungary
in the laboratory			1			1			1			1	-			1		1		1		1	1		1		- '2
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	2013
Aminoglycosides - Gentamicin	2	25	22										3					20	2								
Aminoglycosides - Streptomycin	16	25	22														2	1	8	13	1						Report
Amphenicols - Chloramphenicol	16	25	0													7	18										on tre
Cephalosporins - Cefotaxime	0.5	25	0							3	19	3															ends
Fluoroquinolones - Ciprofloxacin	0.064	25	25														25										and
Penicillins - Ampicillin	8	25	25																25								source
Quinolones - Nalidixic acid	16	25	25																	25							rces
Tetracyclines - Tetracycline	8	25	22											1	2					22							of zo
Trimethoprim	2	25	0										25														onc
Sulfonamides - Sulfamethoxazole	256	25	22																3					22			ses

S. Kentu	cky	Turk fattenin	eys - g flocks
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory	2	5
Antimicrob	ials:	lowest	highest
Aminoglycosides	- Gentamicin	0.25	32
Aminoglycosides	- Streptomycin	2	128
Amphenicols - Ch	nloramphenicol	2	64

Table Antimicrobial susceptibility testing of S. Kentucky in Turkeys - fattening flocks - quantitative data [Dilution method]

S. Kentu	ıcky		eys - g flocks
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory	2	5
Antimicrob	oials:	lowest	highest
Cephalosporins -	- Cefotaxime	0.06	4
Fluoroquinolones	s - Ciprofloxacin	0.008	8
Penicillins - Amp	icillin	0.5	32
Quinolones - Nal	idixic acid	4	64
Tetracyclines - T	etracycline	1	64
Trimethoprim		0.5	32
Sulfonamides - S	Sulfamethoxazole	8	1024

Concentration (µg/ml), number of isolates with a concentration of inhibition equal to

S. Bovismorbificans		Turkeys - fattening flocks																								
Isolates out of a monitoring program (yes/no)		Hungary																								
Number of isolates available in the laboratory		5 5 7 1 2 4 8 16 32 64 128 256 512 1024 2048 >4096 1 2 1 2 1 2 1 2 1 2 1 2 2 2 2 2 2 2 2																								
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	5	0									1	4													
Aminoglycosides - Streptomycin	16	5	0														5									
Amphenicols - Chloramphenicol	16	5	0													4		1								
Cephalosporins - Cefotaxime	0.5	5	0							3	2															
Fluoroquinolones - Ciprofloxacin	0.064	5	0				1		3	1																
Penicillins - Ampicillin	8	5	0											4	1											
Quinolones - Nalidixic acid	16	5	0													4		1								
Tetracyclines - Tetracycline	8	5	0											4	1											
Trimethoprim	2	5	0										5													
Sulfonamides - Sulfamethoxazole	256	5	0																	1	4					

S. Bovis	morbificans	Turkeys - fattening flock					
	Isolates out of a monitoring program (yes/no)						
	Number of isolates available in the laboratory	ţ	5				
Antimicrob	oials:	lowest highest					
Aminoglycosides	- Gentamicin	0.25	32				
Aminoglycosides	- Streptomycin	2	128				
Amphenicols - C	hloramphenicol	2	64				

Table Antimicrobial susceptibility testing of S. Bovismorbificans in Turkeys - fattening flocks - quantitative data [Dilution method]

S. Bovis	S. Bovismorbificans								
		5							
Antimicrob	lowest	highest							
Cephalosporins -	0.06	4							
Fluoroquinolones	s - Ciprofloxacin	0.008	8						
Penicillins - Amp	icillin	0.5	32						
Quinolones - Nal	idixic acid	4	64						
Tetracyclines - T	etracycline	1	64						
Trimethoprim		0.5	32						
Sulfonamides - S	Sulfamethoxazole	8	1024						

Concentration (µg/ml),	number of isolates with a	concentration of inhibition equal to
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S. Bredeney		Gallus gallus (fowl) - broilers																									
Isolates out of a monitoring program (yes/no)		Hungay																									
Number of isolates available in the laboratory		ı ı																									
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	2013
Aminoglycosides - Gentamicin	2	3	0										3														
Aminoglycosides - Streptomycin	16	3	0														1	2									Report
Amphenicols - Chloramphenicol	16	3	0														3										on tr
Cephalosporins - Cefotaxime	0.5	3	0								3																trends
Fluoroquinolones - Ciprofloxacin	0.064	3	3									3															and
Penicillins - Ampicillin	8	3	3																3								
Quinolones - Nalidixic acid	16	3	3																	3							sources
Tetracyclines - Tetracycline	8	3	3																	3							of zo
Trimethoprim	2	3	0										3														zoonoses
Sulfonamides - Sulfamethoxazole	256	3	0																3								ses

S. Brede	S. Bredeney						
	Isolates out of a monitoring program (yes/no)						
	Number of isolates available in the laboratory	÷	3				
Antimicrob	oials:	lowest highest					
Aminoglycosides	- Gentamicin	0.25	32				
Aminoglycosides	- Streptomycin	2	128				
Amphenicols - C	hloramphenicol	2	64				

Table Antimicrobial susceptibility testing of S. Bredeney in Gallus gallus (fowl) - broilers - quantitative data [Dilution method]

S. Bredeney	Gallus gallus (fowl) - broilers							
Isolates out of a monitoring program (yes/no)								
Number of isolates available in the laboratory								
Antimicrobials:	lowest	highest						
Cephalosporins - Cefotaxime	0.06	4						
Fluoroquinolones - Ciprofloxacin	0.008	8						
Penicillins - Ampicillin	0.5	32						
Quinolones - Nalidixic acid	4	64						
Tetracyclines - Tetracycline	1	64						
Trimethoprim	0.5	32						
Sulfonamides - Sulfamethoxazole	8	1024						

Concentration (µg/ml), number of isolates with a concentration of inhibition equal to

S. Newport		Gallus gallus (fowl) - laying hens																								
Isolates out of a monitoring program (yes/no)		Hungary																								
Number of isolates available in the laboratory		2 2 2 2 3 3 4 8 16 32 64 128 256 512 1024 2048 >4096 21 3 3 4 8 16 32 64 128 256 512 1024 2048 3 4096 21 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3																								
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	2	0									1	1													
Aminoglycosides - Streptomycin	16	2	0													1	1									
Amphenicols - Chloramphenicol	16	2	0														2									
Cephalosporins - Cefotaxime	0.5	2	0							2																
Fluoroquinolones - Ciprofloxacin	0.064	2	2										2													
Penicillins - Ampicillin	8	2	2																2							
Quinolones - Nalidixic acid	16	2	1															1	1							
Tetracyclines - Tetracycline	8	2	2																	2						
Trimethoprim	2	2	0										2													
Sulfonamides - Sulfamethoxazole	256	2	0																	2						

S. Newp	S. Newport						
	Number of isolates available in the laboratory		2				
Antimicrob	pials:	lowest highes					
Aminoglycosides	- Gentamicin	0.25	32				
Aminoglycosides	- Streptomycin	2	128				
Amphenicols - Ch	nloramphenicol	2	64				

Table Antimicrobial susceptibility testing of S. Newport in Gallus gallus (fowl) - laying hens - quantitative data [Dilution method]

S. Newp	S. Newport								
	2	2							
Antimicrob	lowest	highest							
Cephalosporins -	0.06	4							
Fluoroquinolones	s - Ciprofloxacin	0.008	8						
Penicillins - Amp	icillin	0.5	32						
Quinolones - Nal	idixic acid	4	64						
Tetracyclines - T	etracycline	1	64						
Trimethoprim		0.5	32						
Sulfonamides - S	Sulfamethoxazole	8	1024						

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Gallus gallus (fowl) - broiler					
5	5				
lowest highes					
0.25	32				
2	128				
2	64				
	owest 0.25				

Table Antimicrobial susceptibility testing of S. Kottbus in Gallus gallus (fowl) - broilers - quantitative data [Dilution method]

S. Kottbus	S. Kottbus								
	olates out of a monitoring ogram (yes/no)								
Ni in	,	5							
Antimicrobia	ls:	lowest	highest						
Cephalosporins - Ce	fotaxime	0.06	4						
Fluoroquinolones - C	Ciprofloxacin	0.008	8						
Penicillins - Ampicilli	n	0.5	32						
Quinolones - Nalidixi	ic acid	4	64						
Tetracyclines - Tetra	cycline	1	64						
Trimethoprim		0.5	32						
Sulfonamides - Sulfa	amethoxazole	8	1024						

Concentration (µg/ml), number of isolates with a concentration of inhibition equal to

							<u> </u>	<u> </u>																		
S. Lille	Gallus gallus (fowl) - laying hens																									
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory														2												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	2	0										1	1												
Aminoglycosides - Streptomycin	16	2	0														2									
Amphenicols - Chloramphenicol	16	2	0														2									
Cephalosporins - Cefotaxime	0.5	2	0								2															
Fluoroquinolones - Ciprofloxacin	0.064	2	0				2																			
Penicillins - Ampicillin	8	2	0											1	1											
Quinolones - Nalidixic acid	16	2	0													2										
Tetracyclines - Tetracycline	8	2	0												2											
Trimethoprim	2	2	0										2													
Sulfonamides - Sulfamethoxazole	256	2	0																			2				

S. Lille	Gallus gallus (fowl) - laying hens				
	Isolates out of a monitoring program (yes/no)				
	Number of isolates available in the laboratory				
Antimicrob	lowest	highest			
Aminoglycosides	0.25	32			
Aminoglycosides	2	128			
Amphenicols - C	2	64			

Table Antimicrobial susceptibility testing of S. Lille in Gallus gallus (fowl) - laying hens - quantitative data [Dilution method]

S. Lille	Gallus gallus (fowl) - laying hens				
	Isolates out of a monitoring program (yes/no)				
	2				
Antimicrob	lowest	highest			
Cephalosporins -	0.06	4			
Fluoroquinolones	0.008	8			
Penicillins - Amp	0.5	32			
Quinolones - Nal	4	64			
Tetracyclines - T	1	64			
Trimethoprim	0.5	32			
Sulfonamides - S	8	1024			

	Concentration (u-	g/ml), number of isolates	with a concentration	of inhibition equal to
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S. Stanley												Turk	eys - fat	tening fl	ocks												-
Isolates out of a monitoring program (yes/no)																											9
Number of isolates available in the laboratory																											
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	
Aminoglycosides - Gentamicin	2	25	0										24	1													1
Aminoglycosides - Streptomycin	16	25	1														18	6	1								2
Amphenicols - Chloramphenicol	16	25	0													10	15										
Cephalosporins - Cefotaxime	0.5	25	0							19	5		1														
Fluoroquinolones - Ciprofloxacin	0.064	25	25								8	14	3														2
Penicillins - Ampicillin	8	25	7											12	5	1			7								
Quinolones - Nalidixic acid	16	25	25																	25							0
Tetracyclines - Tetracycline	8	25	0											9	14	2											2
Trimethoprim	2	25	0										25														
Sulfonamides - Sulfamethoxazole	256	25	3															1	5	16				3			000

S. Stanle	еу	Turk fattenin	eys - g flocks					
	Isolates out of a monitoring program (yes/no)							
	· · · · · · · · · · · · · · · · · · ·							
Antimicrob								
Aminoglycosides	s - Gentamicin	0.25	32					
Aminoglycosides	s - Streptomycin	2	128					
Amphenicols - C	hloramphenicol	2	64					

Table Antimicrobial susceptibility testing of S. Stanley in Turkeys - fattening flocks - quantitative data [Dilution method]

S. Stanley	Turk fattenin	eys - g flocks
Isolates out of a monitoring program (yes/no)		
Number of isolates available in the laboratory	2	5
Antimicrobials:	lowest	highest
Cephalosporins - Cefotaxime	0.06	4
Fluoroquinolones - Ciprofloxacin	0.008	8
Penicillins - Ampicillin	0.5	32
Quinolones - Nalidixic acid	4	64
Tetracyclines - Tetracycline	1	64
Trimethoprim	0.5	32
Sulfonamides - Sulfamethoxazole	8	1024

S. Thompson		Turkeys - fattening flocks Tungan																								
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory														1												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	1	0									1														
Aminoglycosides - Streptomycin	16	1	0														1									
Amphenicols - Chloramphenicol	16	1	0														1									
Cephalosporins - Cefotaxime	0.5	1	0							1																
Fluoroquinolones - Ciprofloxacin	0.064	1	0				1																			
Penicillins - Ampicillin	8	1	0												1											
Quinolones - Nalidixic acid	16	1	0													1										
Tetracyclines - Tetracycline	8	1	0												1											
Trimethoprim	2	1	0										1													
Sulfonamides - Sulfamethoxazole	256	1	0																	1						

S. Thom	pson	Turk fattenin	eys - g flocks						
	Isolates out of a monitoring program (yes/no)								
	Number of isolates available in the laboratory		1						
Antimicrob									
Aminoglycosides	ntimicrobials:								
Aminoglycosides	- Streptomycin	2	128						
Amphenicols - Ch	nloramphenicol	2	64						

Table Antimicrobial susceptibility testing of S. Thompson in Turkeys - fattening flocks - quantitative data [Dilution method]

S. Thompson	Turk fattenin	eys - g flocks
Isolates out of a monitoring program (yes/no)		
Number of isolates available in the laboratory		1
Antimicrobials:	lowest	highest
Cephalosporins - Cefotaxime	0.06	4
Fluoroquinolones - Ciprofloxacin	0.008	8
Penicillins - Ampicillin	0.5	32
Quinolones - Nalidixic acid	4	64
Tetracyclines - Tetracycline	1	64
Trimethoprim	0.5	32
Sulfonamides - Sulfamethoxazole	8	1024

Concentration (µg/ml), number of iso	lates with a concentration of inhibition equal to
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							- 1	, ,																		
S. Senftenberg												Turk	eys - fat	tening fl	ocks											
Isolates out of a monitoring program (yes/no)																										•
Number of isolates available in the laboratory														1												'
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	1	0										1													
Aminoglycosides - Streptomycin	16	1	0														1									
Amphenicols - Chloramphenicol	16	1	0														1									
Cephalosporins - Cefotaxime	0.5	1	0								1															
Fluoroquinolones - Ciprofloxacin	0.064	1	0				1																			
Penicillins - Ampicillin	8	1	0											1												
Quinolones - Nalidixic acid	16	1	0													1										
Tetracyclines - Tetracycline	8	1	0												1											
Trimethoprim	2	1	0										1													
Sulfonamides - Sulfamethoxazole	256	1	0																	1						

S. Senft	enberg	Turk fattenin	eys - g flocks						
	Isolates out of a monitoring program (yes/no)								
	· · · · · · · · · · · · · · · · · · ·								
Antimicrob									
Aminoglycosides	- Gentamicin	0.25	32						
Aminoglycosides	- Streptomycin	2	128						
Amphenicols - C	hloramphenicol	2	64						

Table Antimicrobial susceptibility testing of S. Senftenberg in Turkeys - fattening flocks - quantitative data [Dilution method]

S. Senftenberg	Turk fattenin	eys - g flocks
Isolates out of a monitoring program (yes/no)		
Number of isolates available in the laboratory		1
Antimicrobials:	lowest	highest
Cephalosporins - Cefotaxime	0.06	4
Fluoroquinolones - Ciprofloxacin	0.008	8
Penicillins - Ampicillin	0.5	32
Quinolones - Nalidixic acid	4	64
Tetracyclines - Tetracycline	1	64
Trimethoprim	0.5	32
Sulfonamides - Sulfamethoxazole	8	1024

Table Antimicrobial susceptibility testing of S. enterica subsp. salamae in Gallus gallus (fowl) - laying hens - quantitative data [Dilution method]

							\1 ·	J. /																		
S. enterica subsp. salamae												Gallus g	allus (fo	wl) - lay	ing hens	3										
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory		off N n ==0.002 ==0.004 0.008 0.015 0.016 0.03 0.06 0.12 0.25 0.5 1 2 4 8 16 32 64 128 256 512 1.024 2048 24096																								
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	1	0										1													
Aminoglycosides - Streptomycin	16	1	0														1									
Amphenicols - Chloramphenicol	16	1	0													1										
Cephalosporins - Cefotaxime	0.5	1	0								1															
Fluoroquinolones - Ciprofloxacin	0.064	1	0				1																			
Penicillins - Ampicillin	8	1	0												1											
Quinolones - Nalidixic acid	16	1	0													1										
Tetracyclines - Tetracycline	8	1	0											1												
Trimethoprim	2	1	0										1													
Sulfonamides - Sulfamethoxazole	256	1	0																	1						

S. enteri	ica subsp.	(fowl) -	gallus laying					
	Number of isolates available in the laboratory		1					
Antimicrob	oials:	lowest	highest					
Aminoglycosides	s - Gentamicin	0.25	32					
Aminoglycosides	Aminoglycosides - Streptomycin							
Amphenicols - C	2	64						

Table Antimicrobial susceptibility testing of S. enterica subsp. salamae in Gallus gallus (fowl) - laying hens - quantitative data [Dilution method]

S. enterica subsp. salamae	Gallus gallus (fowl) - laying hens							
Isolates out of a program (yes/no								
Number of isola in the laboratory	1							
Antimicrobials:		lowest	highest					
Cephalosporins - Cefotaxime		0.06	4					
Fluoroquinolones - Ciprofloxacin		0.008	8					
Penicillins - Ampicillin		0.5	32					
Quinolones - Nalidixic acid		4	64					
Tetracyclines - Tetracycline		1	64					
Trimethoprim	0.5	32						
Sulfonamides - Sulfamethoxazole		8	1024					

Concentration	(µg/mi),	number	of isolates	with a	concentration	of inhibition	equal to

S. Tennessee												Gallus	gallus	(fowl) - b	oroilers												Ŧ
Isolates out of a monitoring program (yes/no)																											Hungary
Number of isolates available in the laboratory														3													1
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	2013
Aminoglycosides - Gentamicin	2	3	0										3														
Aminoglycosides - Streptomycin	16	3	0															3									Report
Amphenicols - Chloramphenicol	16	3	0														3										on tr
Cephalosporins - Cefotaxime	0.5	3	0								3																trends
Fluoroquinolones - Ciprofloxacin	0.064	3	0				2		1																		and
Penicillins - Ampicillin	8	3	0											2	1												
Quinolones - Nalidixic acid	16	3	0													3											sources
Tetracyclines - Tetracycline	8	3	0												3												of zo
Trimethoprim	2	3	0										3														zoonos
Sulfonamides - Sulfamethoxazole	256	3	0																1		2						ses

S. Tenne	essee		gallus broilers						
	Isolates out of a monitoring program (yes/no)								
	Number of isolates available in the laboratory	·	3						
Antimicrob	oials:	lowest	highest						
Aminoglycosides	- Gentamicin	0.25	32						
Aminoglycosides	- Streptomycin	2	128						
Amphenicols - Ch	2	64							

Table Antimicrobial susceptibility testing of S. Tennessee in Gallus gallus (fowl) - broilers - quantitative data [Dilution method]

S. Tennessee	Gallus gallus (fowl) - broilers					
Isolates out of a monitoring program (yes/no)						
Number of isolates available in the laboratory	3					
Antimicrobials:	lowest	highest				
Cephalosporins - Cefotaxime	0.06	4				
Fluoroquinolones - Ciprofloxacin	0.008	8				
Penicillins - Ampicillin	0.5	32				
Quinolones - Nalidixic acid	4	64				
Tetracyclines - Tetracycline	1	64				
Trimethoprim	0.5	32				
Sulfonamides - Sulfamethoxazole	8	1024				

S. Tennessee												Turk	eys - fa	ttening f	ocks												ェ
Isolates out of a monitoring program (yes/no)																											Hungary
Number of isolates available in the laboratory														3													1
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	2013
Aminoglycosides - Gentamicin	2	3	0										3														Re
Aminoglycosides - Streptomycin	16	3	0														1	2									Report
Amphenicols - Chloramphenicol	16	3	0														3										on tr
Cephalosporins - Cefotaxime	0.5	3	0							2	1																trends
Fluoroquinolones - Ciprofloxacin	0.064	3	1				2					1															and
Penicillins - Ampicillin	8	3	0											3													S
Quinolones - Nalidixic acid	16	3	1													2				1							ources
Tetracyclines - Tetracycline	8	3	1											1	1					1							of zo
Trimethoprim	2	3	0										3														zoonoses
Sulfonamides - Sulfamethoxazole	256	3	0																	3							ses

S. Tennessee			eys - g flocks
Isolates out of a monitoring program (yes/no)			
Number of isolates available in the laboratory	е	·	3
Antimicrobials:		lowest	highest
Aminoglycosides - Gentamicin		0.25	32
Aminoglycosides - Streptomycin		2	128
Amphenicols - Chloramphenicol		2	64

Table Antimicrobial susceptibility testing of S. Tennessee in Turkeys - fattening flocks

- quantitative data [Dilution method]

S. Tennessee	Turkeys - fattening flocks				
Isolates out of a monitoring program (yes/no)					
Number of isolates available in the laboratory	;	3			
Antimicrobials:	lowest	highest			
Cephalosporins - Cefotaxime	0.06	4			
Fluoroquinolones - Ciprofloxacin	0.008	8			
Penicillins - Ampicillin	0.5	32			
Quinolones - Nalidixic acid	4	64			
Tetracyclines - Tetracycline	1	64			
Trimethoprim	0.5	32			
Sulfonamides - Sulfamethoxazole	8	1024			

							<u> </u>	<u> </u>																		
S. Virchow		Gallus gallus (fowl) - laying hens Hungary																								
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory														1												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	1	0										1													
Aminoglycosides - Streptomycin	16	1	1																	1						
Amphenicols - Chloramphenicol	16	1	1																	1						
Cephalosporins - Cefotaxime	0.5	1	0							1																
Fluoroquinolones - Ciprofloxacin	0.064	1	1								1															
Penicillins - Ampicillin	8	1	0											1												
Quinolones - Nalidixic acid	16	1	1																	1						
Tetracyclines - Tetracycline	8	1	1																	1						
Trimethoprim	2	1	1																1							
Sulfonamides - Sulfamethoxazole	256	1	1																					1		

S. Vircho	ow	Gallus (fowl) - he							
	Isolates out of a monitoring program (yes/no)								
	Number of isolates available in the laboratory		1						
Antimicrob	oials:	lowest	highest						
Aminoglycosides	- Gentamicin	0.25	32						
Aminoglycosides	- Streptomycin	2	128						
Amphenicols - Ch	2	64							

Table Antimicrobial susceptibility testing of S. Virchow in Gallus gallus (fowl) - laying hens - quantitative data [Dilution method]

S. Virch	S. Virchow								
	1								
Antimicro	oials:	lowest	highest						
Cephalosporins	- Cefotaxime	0.06	4						
Fluoroquinolones	s - Ciprofloxacin	0.008	8						
Penicillins - Amp	picillin	0.5	32						
Quinolones - Na	lidixic acid	4	64						
Tetracyclines - T	etracycline	1	64						
Trimethoprim	0.5	32							
Sulfonamides - S	8	1024							

							\1 ·	<i></i>																		
S. Infantis		Gallus gallus (fowl) - broilers																								
Isolates out of a monitoring program (yes/no)		Hungary 148																								
Number of isolates available in the laboratory		148 148 148 148 148 148 148 15 16 17 18 18 18 18 18 18 18 18 18																								
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	148	0									101	44	3												
Aminoglycosides - Streptomycin	16	148	119													6	14	9	96	21	2					
Amphenicols - Chloramphenicol	16	148	5												1	16	56	70	5							
Cephalosporins - Cefotaxime	0.5	148	0							5	58	73	12													
Fluoroquinolones - Ciprofloxacin	0.064	148	148									28	91	22	4	2	1									
Penicillins - Ampicillin	8	148	12											23	65	47	1		12							
Quinolones - Nalidixic acid	16	148	148																	148						
Tetracyclines - Tetracycline	8	148	122												7	17	2		1	121						
Trimethoprim	2	148	0										146	2												
Sulfonamides - Sulfamethoxazole	256	148	126																4	16	2			126		

S. Infant	is		gallus broilers
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory	14	48
Antimicrob	oials:	lowest	highest
Aminoglycosides	- Gentamicin	0.25	32
Aminoglycosides	- Streptomycin	2	128
Amphenicols - C	hloramphenicol	2	64

Table Antimicrobial susceptibility testing of S. Infantis in Gallus gallus (fowl) - broilers - quantitative data [Dilution method]

S. Infant	tis		gallus broilers
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory	14	18
Antimicrob	oials:	lowest	highest
Cephalosporins -	- Cefotaxime	0.06	4
Fluoroquinolones	s - Ciprofloxacin	0.008	8
Penicillins - Amp	icillin	0.5	32
Quinolones - Nal	lidixic acid	4	64
Tetracyclines - T	etracycline	1	64
Trimethoprim	0.5	32	
Sulfonamides - S	Sulfamethoxazole	8	1024

S. Infantis		Turkeys - fattening flocks														ļ											
Isolates out of a monitoring program (yes/no)		Hungary																									
Number of isolates available in the laboratory													;	33													1
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	7013
Aminoglycosides - Gentamicin	2	33	0									22	9	2													ᄀ ᇡ
Aminoglycosides - Streptomycin	16	33	31														1	1	22	7	2						01
Amphenicols - Chloramphenicol	16	33	1													3	12	17	1								
Cephalosporins - Cefotaxime	0.5	33	0							1	10	19	3														ַ מו ט
Fluoroquinolones - Ciprofloxacin	0.064	33	32			1						8	15	8		1											_ a
Penicillins - Ampicillin	8	33	2										1	2	14	13	1		2								sourc
Quinolones - Nalidixic acid	16	33	33																	33							rces
Tetracyclines - Tetracycline	8	33	33																1	32							
Trimethoprim	2	33	0										33														
Sulfonamides - Sulfamethoxazole	256	33	32																	1				32			ses

S. Infant	is		eys - g flocks					
	Isolates out of a monitoring program (yes/no)							
	Number of isolates available in the laboratory	3	3					
Antimicrob	oials:	lowest	highest					
Aminoglycosides	- Gentamicin	0.25	32					
Aminoglycosides	2	128						
Amphenicols - Cl	amphenicols - Chloramphenicol							

Table Antimicrobial susceptibility testing of S. Infantis in Turkeys - fattening flocks

- quantitative data [Dilution method]

S. Infan	tis		eys - g flocks
	Isolates out of a monitoring program (yes/no)		
	3	3	
Antimicrol	lowest	highest	
Cephalosporins	- Cefotaxime	0.06	4
Fluoroquinolone	s - Ciprofloxacin	0.008	8
Penicillins - Amp	picillin	0.5	32
Quinolones - Na	llidixic acid	4	64
Tetracyclines - T	Fetracycline	1	64
Trimethoprim	0.5	32	
Sulfonamides - S	Sulfamethoxazole	8	1024

Concentration (µg/ml), number of isolates with a concentration of inhibition	equal to
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S. Brandenburg												Gallus g	allus (fo	owl) - lay	ing hens	3										
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory														1												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	1	0											1												
Aminoglycosides - Streptomycin	16	1	0														1									
Amphenicols - Chloramphenicol	16	1	0														1									
Cephalosporins - Cefotaxime	0.5	1	0							1																
Fluoroquinolones - Ciprofloxacin	0.064	1	0				1																			
Penicillins - Ampicillin	8	1	0												1											
Quinolones - Nalidixic acid	16	1	0													1										
Tetracyclines - Tetracycline	8	1	0											1												
Trimethoprim	2	1	0										1													
Sulfonamides - Sulfamethoxazole	256	1	0																	1						

S. Brand	denburg	(fowl) -	gallus laying ns						
	Isolates out of a monitoring program (yes/no) Number of isolates available								
		1							
Antimicrob	oials:	lowest	highest						
Aminoglycosides	s - Gentamicin	0.25	32						
Aminoglycosides	s - Streptomycin	2	128						
Amphenicols - C	hloramphenicol	2	64						

S. Brandenburg	(fowl) -	gallus laying ns
Isolates out of a monitoring program (yes/no)		
Number of isolates available in the laboratory		1
Antimicrobials:	lowest	highest
Cephalosporins - Cefotaxime	0.06	4
Fluoroquinolones - Ciprofloxacin	0.008	8
Penicillins - Ampicillin	0.5	32
Quinolones - Nalidixic acid	4	64
Tetracyclines - Tetracycline	1	64
Trimethoprim	0.5	32
Sulfonamides - Sulfamethoxazole	8	1024

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S. Montevideo		Gallus gallus (fowl) - laying hens																								
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory														2												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	2	0										2													
Aminoglycosides - Streptomycin	16	2	0															2								
Amphenicols - Chloramphenicol	16	2	0													1	1									
Cephalosporins - Cefotaxime	0.5	2	0							1	1															
Fluoroquinolones - Ciprofloxacin	0.064	2	0				2																			
Penicillins - Ampicillin	8	2	0											2												
Quinolones - Nalidixic acid	16	2	0													2										
Tetracyclines - Tetracycline	8	2	0												2											
Trimethoprim	2	2	0										2													
Sulfonamides - Sulfamethoxazole	256	2	0																1	1						

S. Monte	evideo		gallus laying ns
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory		2
Antimicrob	oials:	lowest	highest
Aminoglycosides	- Gentamicin	0.25	32
Aminoglycosides	- Streptomycin	2	128
Amphenicols - C	hloramphenicol	2	64

Table Antimicrobial susceptibility testing of S. Montevideo in Gallus gallus (fowl) - laying hens - quantitative data [Dilution method]

S. Montevideo	Gallus gallus (fowl) - laying hens				
Isolates out of a monitoring program (yes/no)					
Number of isolates available in the laboratory	1	2			
Antimicrobials:	lowest	highest			
Cephalosporins - Cefotaxime	0.06	4			
Fluoroquinolones - Ciprofloxacin	0.008	8			
Penicillins - Ampicillin	0.5	32			
Quinolones - Nalidixic acid	4	64			
Tetracyclines - Tetracycline	1	64			
Trimethoprim	0.5	32			
Sulfonamides - Sulfamethoxazole	8	1024			

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	Concentration (µg/ml),	number of isolates with a	a concentration of inhibition e	equal to
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S. Szentes							·					Gallus	gallus	(fowl) - b	roilers	·										
Isolates out of a monitoring program (yes/no)		ungary																								
Number of isolates available in the laboratory																										
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	1	0										1													
Aminoglycosides - Streptomycin	16	1	0														1									
Amphenicols - Chloramphenicol	16	1	0														1									
Cephalosporins - Cefotaxime	0.5	1	0								1															
Fluoroquinolones - Ciprofloxacin	0.064	1	0				1																			
Penicillins - Ampicillin	8	1	0												1											
Quinolones - Nalidixic acid	16	1	0													1										
Tetracyclines - Tetracycline	8	1	0												1											
Trimethoprim	2	1	0										1													
Sulfonamides - Sulfamethoxazole	256	1	1																					1		

S. Szent	Gallus gallus (fowl) - broiler				
	Isolates out of a monitoring program (yes/no)				
	Number of isolates available in the laboratory		1		
Antimicrob	lowest	highest			
Aminoglycosides	- Gentamicin	0.25	32		
Aminoglycosides	- Streptomycin	2	128		
Amphenicols - Cl	nloramphenicol	2	64		

Table Antimicrobial susceptibility testing of S. Szentes in Gallus gallus (fowl) - broilers - quantitative data [Dilution method]

S. Szent	Gallus gallus (fowl) - broilers				
	Isolates out of a monitoring program (yes/no)				
		1			
Antimicrob	oials:	lowest	highest		
Cephalosporins -	Cefotaxime	0.06	4		
Fluoroquinolones	- Ciprofloxacin	0.008	8		
Penicillins - Ampi	cillin	0.5	32		
Quinolones - Nali	idixic acid	4	64		
Tetracyclines - Te	etracycline	1	64		
Trimethoprim		0.5	32		
Sulfonamides - S	ulfamethoxazole	8	1024		

S. Typhimurium		Gallus gallus (fowl) - laying hens																								
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory		5																								
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	5	0									1	4													-
Aminoglycosides - Streptomycin	16	5	0														3	2								
Amphenicols - Chloramphenicol	16	5	0													4	1									
Cephalosporins - Cefotaxime	0.5	5	0							5																
Fluoroquinolones - Ciprofloxacin	0.064	5	0				2		3																	
Penicillins - Ampicillin	8	5	0											1	4											
Quinolones - Nalidixic acid	16	5	0													5										
Tetracyclines - Tetracycline	8	5	0											1	4											
Trimethoprim	2	5	0										5													
Sulfonamides - Sulfamethoxazole	256	5	0		_															5						

S. Typhimuriui	(fowl) -	gallus laying		
Isolates program				
Number in the lal	5			
Antimicrobials:		lowest	highest	
Aminoglycosides - Gentam	icin	0.25	32	
Aminoglycosides - Streptor	mycin	2	128	
Amphenicols - Chloramphe	2	64		

S. Typhimurium	Gallus gallus (fowl) - laying hens					
Isolates out of a monitoring program (yes/no)						
Number of isolates available in the laboratory		5				
Antimicrobials:	lowest	highest				
Cephalosporins - Cefotaxime	0.06	4				
Fluoroquinolones - Ciprofloxacin	0.008	8				
Penicillins - Ampicillin	0.5	32				
Quinolones - Nalidixic acid	4	64				
Tetracyclines - Tetracycline	1	64				
Trimethoprim	0.5	32				
Sulfonamides - Sulfamethoxazole	8	1024				

S. Enteritidis		Gallus gallus (fowl) - broilers														ı											
Isolates out of a monitoring program (yes/no)		Hungary																									
Number of isolates available in the laboratory		1																									
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	2013
Aminoglycosides - Gentamicin	2	3	0									3															
Aminoglycosides - Streptomycin	16	3	0												1	2											Report
Amphenicols - Chloramphenicol	16	3	0														3										on tr
Cephalosporins - Cefotaxime	0.5	3	0							2	1																trends
Fluoroquinolones - Ciprofloxacin	0.064	3	0				1		2																		and
Penicillins - Ampicillin	8	3	0											2	1												S
Quinolones - Nalidixic acid	16	3	0													3											ources
Tetracyclines - Tetracycline	8	3	0											2	1												of zo
Trimethoprim	2	3	0										3														zoonoses
Sulfonamides - Sulfamethoxazole	256	3	0																1	2							ses

S. Enter	Gallus gallus (fowl) - broilers					
	Number of isolates available in the laboratory	;	3			
Antimicrob	lowest	highest				
Aminoglycosides	- Gentamicin	0.25	32			
Aminoglycosides	- Streptomycin	2	128			
Amphenicols - Cl	hloramphenicol	2	64			

Table Antimicrobial susceptibility testing of S. Enteritidis in Gallus gallus (fowl) - broilers - quantitative data [Dilution method]

S. Enteritidis	Gallus gallus (fowl) - broilers				
Isolates out of a monitoring program (yes/no)					
Number of isolates available in the laboratory	;	3			
Antimicrobials:	lowest	highest			
Cephalosporins - Cefotaxime	0.06	4			
Fluoroquinolones - Ciprofloxacin	0.008	8			
Penicillins - Ampicillin	0.5	32			
Quinolones - Nalidixic acid	4	64			
Tetracyclines - Tetracycline	1	64			
Trimethoprim	0.5	32			
Sulfonamides - Sulfamethoxazole	8	1024			

Concentration (µg/ml)	, number of isolates with a concentration	of inhibition equal to
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S. Enteritidis												Turk	eys - fat	tening fl	ocks											
Isolates out of a monitoring program (yes/no) Number of isolates available														1												
in the laboratory			1	_	1				1					· 	I							1				$\overline{}$
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	1	0									1														
Aminoglycosides - Streptomycin	16	1	0													1										
Amphenicols - Chloramphenicol	16	1	0														1									
Cephalosporins - Cefotaxime	0.5	1	0								1															
Fluoroquinolones - Ciprofloxacin	0.064	1	0						1																	
Penicillins - Ampicillin	8	1	0												1											
Quinolones - Nalidixic acid	16	1	0													1										
Tetracyclines - Tetracycline	8	1	0												1											
Trimethoprim	2	1	0										1													
Sulfonamides - Sulfamethoxazole	256	1	0																	1						

Turke fattenin	eys - g flocks
1	
lowest	highest
0.25	32
2	128
2	64
F	lowest 0.25

Table Antimicrobial susceptibility testing of S. Enteritidis in Turkeys - fattening flocks - quantitative data [Dilution method]

S. Enteritidis		eys - g flocks
Isolates out of a monitoring program (yes/no)		
Number of isolates available in the laboratory		1
Antimicrobials:	lowest	highest
Cephalosporins - Cefotaxime	0.06	4
Fluoroquinolones - Ciprofloxacin	0.008	8
Penicillins - Ampicillin	0.5	32
Quinolones - Nalidixic acid	4	64
Tetracyclines - Tetracycline	1	64
Trimethoprim	0.5	32
Sulfonamides - Sulfamethoxazole	8	1024

S. Mbandaka							,					Gallus		(fowl) - b												
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory														2												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	2	0										2													
Aminoglycosides - Streptomycin	16	2	0														2									
Amphenicols - Chloramphenicol	16	2	0														2									
Cephalosporins - Cefotaxime	0.5	2	0								2															
Fluoroquinolones - Ciprofloxacin	0.064	2	0				2																			
Penicillins - Ampicillin	8	2	0												2											
Quinolones - Nalidixic acid	16	2	0													2										
Tetracyclines - Tetracycline	8	2	0												2											
Trimethoprim	2	2	0										2													
Sulfonamides - Sulfamethoxazole	256	2	0																	1	1					

S. Mban	daka		gallus broilers					
	Isolates out of a monitoring program (yes/no)							
	Number of isolates available in the laboratory	2	2					
Antimicrob	ials:	lowest	highest					
Aminoglycosides	- Gentamicin	0.25	32					
Aminoglycosides	- Streptomycin	2	128					
Amphenicols - Ch	mphenicols - Chloramphenicol							

Table Antimicrobial susceptibility testing of S. Mbandaka in Gallus gallus (fowl) - broilers - quantitative data [Dilution method]

S. Mbandaka		gallus broilers
Isolates out of a monitoring program (yes/no)		
Number of isolates available in the laboratory	:	2
Antimicrobials:	lowest	highest
Cephalosporins - Cefotaxime	0.06	4
Fluoroquinolones - Ciprofloxacin	0.008	8
Penicillins - Ampicillin	0.5	32
Quinolones - Nalidixic acid	4	64
Tetracyclines - Tetracycline	1	64
Trimethoprim	0.5	32
Sulfonamides - Sulfamethoxazole	8	1024

S. Abony												Gallus g	allus (fo	owl) - lay	ing hens	5										
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory														1												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	1	0										1													
Aminoglycosides - Streptomycin	16	1	1																1							
Amphenicols - Chloramphenicol	16	1	0														1									
Cephalosporins - Cefotaxime	0.5	1	0								1															
Fluoroquinolones - Ciprofloxacin	0.064	1	0						1																	
Penicillins - Ampicillin	8	1	0											1												
Quinolones - Nalidixic acid	16	1	0													1										
Tetracyclines - Tetracycline	8	1	0												1											
Trimethoprim	2	1	0										1													
Sulfonamides - Sulfamethoxazole	256	1	0																1							

S. Abony	У	Gallus (fowl) - he	
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory		I
Antimicrob	oials:	lowest	highest
Aminoglycosides	- Gentamicin	0.25	32
Aminoglycosides	- Streptomycin	2	128
Amphenicols - Cl	hloramphenicol	2	64

Table Antimicrobial susceptibility testing of S. Abony in Gallus gallus (fowl) - laying hens - quantitative data [Dilution method]

S. Abony	(fowl) -	gallus laying
Isolates out of a monitoring program (yes/no)		
Number of isolates available in the laboratory		1
Antimicrobials:	lowest	highest
Cephalosporins - Cefotaxime	0.06	4
Fluoroquinolones - Ciprofloxacin	0.008	8
Penicillins - Ampicillin	0.5	32
Quinolones - Nalidixic acid	4	64
Tetracyclines - Tetracycline	1	64
Trimethoprim	0.5	32
Sulfonamides - Sulfamethoxazole	8	1024

Concentration (µg/ml), number of isolates with a concentration of in	hibition equal to
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S. Indiana												Gallus	gallus	(fowl) - k	oroilers												ı
Isolates out of a monitoring program (yes/no)																											Hungary
Number of isolates available in the laboratory														1													1
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	2013
Aminoglycosides - Gentamicin	2	1	0										1														
Aminoglycosides - Streptomycin	16	1	0															1									Report
Amphenicols - Chloramphenicol	16	1	0														1										on tr
Cephalosporins - Cefotaxime	0.5	1	0							1																	trends
Fluoroquinolones - Ciprofloxacin	0.064	1	0				1																				and
Penicillins - Ampicillin	8	1	0										1														S
Quinolones - Nalidixic acid	16	1	0													1											ources
Tetracyclines - Tetracycline	8	1	0												1												of zo
Trimethoprim	2	1	0										1														zoonoses
Sulfonamides - Sulfamethoxazole	256	1	0																	1							ses

S. Indiar	na		gallus broilers				
	Isolates out of a monitoring program (yes/no)						
	Number of isolates available in the laboratory	•	1				
Antimicrob	pials:	lowest	highest				
Aminoglycosides	- Gentamicin	0.25	32				
Aminoglycosides	- Streptomycin	2	128				
Amphenicols - Ch	mphenicols - Chloramphenicol						

Table Antimicrobial susceptibility testing of S. Indiana in Gallus gallus (fowl) - broilers

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S. Indiana		Gallus gallus (fowl) - broilers	
	Isolates out of a monitoring program (yes/no)		
Number of isolates available in the laboratory		1	
Antimicrobials:		lowest	highest
Cephalosporins - Cefotaxime		0.06	4
Fluoroquinolones - Ciprofloxacin		0.008	8
Penicillins - Ampicillin		0.5	32
Quinolones - Nalidixic acid		4	64
Tetracyclines - Tetracycline		1	64
Trimethoprim		0.5	32
Sulfonamides - Sulfamethoxazole		8	1024

<u>Table Antimicrobial susceptibility testing of S. Infantis in Meat from broilers (Gallus gallus)</u> - food sample - meat - quantitative data [Dilution method]

S. Infantis						ncentra	ποπ (μ	griii), ii	difficer	01 13014		Meat fro					10									
Isolates out of a monitoring program (yes/no)													ye	es												
Number of isolates available in the laboratory													14	45												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	145	0									95	35	15												
Aminoglycosides - Streptomycin	32	145	82											1		2	13	10	37	54	25	3				
Amphenicols - Chloramphenicol	16	145	0												13	45	72	15								
Cephalosporins - Cefotaxime	0.5	145	0								86	49	10													
Fluoroquinolones - Ciprofloxacin	0.064	145	145								5	32	65	34	9											
Penicillins - Ampicillin	8	145	15										12	24	37	30	27	11			4					
Quinolones - Nalidixic acid	16	145	145																		1	144				
Sulfonamides	256	145	112													1	2	3	6	8	6	7	8	104		
Tetracyclines - Tetracycline	8	145	124											10	5	6			4	31	60	29				
Trimethoprim	2	145	0									14	74	50	7											
Resistant to 2 antimicrobials		12	12	12																						
Resistant to 3 antimicrobials		15	15	15																						
Resistant to 4 antimicrobials		43	43	43																						
Resistant to >4 antimicrobials		75	75	75																						

Table Antimicrobial susceptibility testing of S. Infantis in Meat from broilers (Gallus gallus) - food sample - meat - quantitative data [Dilution method]

S. Infant	is		from (Gallus us)
	Isolates out of a monitoring program (yes/no)	ye	es
	Number of isolates available in the laboratory	14	15
Antimicrob	oials:	lowest	highest
Aminoglycosides	- Gentamicin		
Aminoglycosides	- Streptomycin		
Amphenicols - Ch	nloramphenicol		
Cephalosporins -	Cefotaxime		
Fluoroquinolones	- Ciprofloxacin		
Penicillins - Ampi	cillin		
Quinolones - Nali	idixic acid		
Sulfonamides			
Tetracyclines - Te	etracycline		
Trimethoprim			
Resistant to 2 and	timicrobials		
Resistant to 3 and	timicrobials		
Resistant to 4 and	timicrobials		
Resistant to >4 a	ntimicrobials		

	Concentration (µg/ml)	, number of isolates with a	a concentration of inhibition equal to	
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S. Infantis												ı	Meat fro	m turkey	y											
Isolates out of a monitoring program (yes/no)													ye	es												
Number of isolates available in the laboratory													1	7												
Antimicrobials:	Cut-off value	Ν	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	17	0									9	8													Į.
Aminoglycosides - Streptomycin	32	17	11														1	1	4	10		1				Į.
Amphenicols - Chloramphenicol	16	17	0												2	3	11	1								
Cephalosporins - Cefotaxime	0.5	17	0								6	9	2													
Fluoroquinolones - Ciprofloxacin	0.064	17	17									5	2	9	1											
Penicillins - Ampicillin	8	17	4											1	3	3	6	3			1					
Quinolones - Nalidixic acid	16	17	17																			17				
Sulfonamides	256	17	10													1			2		1	3	1	9		Į.
Tetracyclines - Tetracycline	8	17	16												1				2	1	8	5				
Trimethoprim	2	17	0									2	7	6	2											
Resistant to 2 antimicrobials		3	3	3																						
Resistant to 3 antimicrobials		6	6	6																						
Resistant to 4 antimicrobials		5	5	5																						
Resistant to >4 antimicrobials		3	3	3							-															

S. Infant	is	Meat turl	from
	Isolates out of a monitoring program (yes/no)	ye	es
	Number of isolates available in the laboratory	1	7
Antimicrob	ials:	lowest	highest
Aminoglycosides	- Gentamicin		
Aminoglycosides	- Streptomycin		
Amphenicols - Ch	nloramphenicol		
Cephalosporins -	Cefotaxime		
Fluoroquinolones	- Ciprofloxacin		
Penicillins - Ampi	cillin		
Quinolones - Nali	dixic acid		
Sulfonamides			
Tetracyclines - Te	etracycline		
Trimethoprim			
Resistant to 2 and	timicrobials		
Resistant to 3 and	timicrobials		
Resistant to 4 and	timicrobials		
Resistant to >4 a	ntimicrobials		

S. Kentucky							·					Gallus	gallus	(fowl) - b	oroilers												ı
Isolates out of a monitoring program (yes/no)																											Hungary
Number of isolates available in the laboratory													•	2													1
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	2013
Aminoglycosides - Gentamicin	2	2	2															2									1
Aminoglycosides - Streptomycin	16	2	2																	2							Report
Amphenicols - Chloramphenicol	16	2	0													1	1										on tr
Cephalosporins - Cefotaxime	0.5	2	0								2																trends
Fluoroquinolones - Ciprofloxacin	0.064	2	2														2										and
Penicillins - Ampicillin	8	2	2																2								S
Quinolones - Nalidixic acid	16	2	2																	2							ources
Tetracyclines - Tetracycline	8	2	2																	2							of zo
Trimethoprim	2	2	0										2														zoonoses
Sulfonamides - Sulfamethoxazole	256	2	2																					2			ses

S. Kentu	cky		gallus broilers
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory	2	2
Antimicrob	ials:	lowest	highest
Aminoglycosides	- Gentamicin	0.25	32
Aminoglycosides	- Streptomycin	2	128
Amphenicols - Ch	nloramphenicol	2	64

Table Antimicrobial susceptibility testing of S. Kentucky in Gallus gallus (fowl) - broilers - quantitative data [Dilution method]

S. Kentucky			gallus broilers
Isolates out of a monitoring program (yes/no)	g		
Number of isolates availab in the laboratory	ole	1	2
Antimicrobials:		lowest	highest
Cephalosporins - Cefotaxime		0.06	4
Fluoroquinolones - Ciprofloxacin		0.008	8
Penicillins - Ampicillin		0.5	32
Quinolones - Nalidixic acid		4	64
Tetracyclines - Tetracycline		1	64
Trimethoprim		0.5	32
Sulfonamides - Sulfamethoxazole		8	1024

S. Bovismorbificans	(fowl)	gallus laying ns
Isolates out of a monitoring program (yes/no)		
Number of isolates available in the laboratory		2
Antimicrobials:	lowest	highest
Aminoglycosides - Gentamicin	0.25	32
Aminoglycosides - Streptomycin	2	128
Amphenicols - Chloramphenicol	2	64

S. Bovismorbificans	(fowl) -	gallus laying ns
Isolates out of a monitoring program (yes/no)		
Number of isolates available in the laboratory	1	2
Antimicrobials:	lowest	highest
Cephalosporins - Cefotaxime	0.06	4
Fluoroquinolones - Ciprofloxacin	0.008	8
Penicillins - Ampicillin	0.5	32
Quinolones - Nalidixic acid	4	64
Tetracyclines - Tetracycline	1	64
Trimethoprim	0.5	32
Sulfonamides - Sulfamethoxazole	8	1024

Table Antimicrobial susceptibility testing of S. Enteritidis in All foodstuffs - food sample - quantitative data [Dilution method]

S. Enteritidis							N.	<u>, , , , , , , , , , , , , , , , , , , </u>		01 13014				dstuffs		•										
Isolates out of a monitoring program (yes/no)													r	10												
Number of isolates available in the laboratory														9												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	9	0									5	4													
Aminoglycosides - Streptomycin	32	9	0												3	3	3									
Amphenicols - Chloramphenicol	16	9	0												5	2	2									
Cephalosporins - Cefotaxime	0.5	9	0								8	1														
Fluoroquinolones - Ciprofloxacin	0.064	9	1						8			1														
Penicillins - Ampicillin	8	9	0										1	3	2	3										
Quinolones - Nalidixic acid	16	9	1												3	5						1				
Sulfonamides	256	9	1															1	2	3	2		1			
Tetracyclines - Tetracycline	8	9	0											8	1											
Trimethoprim	2	9	0									3	5	1												
Fully sensitive		7	7	7																						
Resistant to 1 antimicrobial		1	1	1																						
Resistant to 2 antimicrobials		1	1	1																						

Table Antimicrobial susceptibility testing of S. Enteritidis in All foodstuffs - food sample - quantitative data [Dilution method]

S. Enter	ritidis	All foo	dstuffs
	Isolates out of a monitoring program (yes/no)	n	0
	Number of isolates available in the laboratory	!	9
Antimicrol	bials:	lowest	highest
Aminoglycosides			
Aminoglycosides			
Amphenicols - C	chloramphenicol		
Cephalosporins	- Cefotaxime		
Fluoroquinolone	s - Ciprofloxacin		
Penicillins - Amp	picillin		
Quinolones - Na	lidixic acid		
Sulfonamides			
Tetracyclines - T	etracycline		
Trimethoprim			
Fully sensitive			
Resistant to 1 ar	ntimicrobial		
Resistant to 2 ar	ntimicrobials		

Concentration (µg/ml), number of isolates with a concentration of inhibition e	qual to
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S. Braenderup												Gallus g	allus (fo	wl) - lay	ing hens	3										
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory														3												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	3	0										3													
Aminoglycosides - Streptomycin	16	3	0														3									
Amphenicols - Chloramphenicol	16	3	0														3									
Cephalosporins - Cefotaxime	0.5	3	0							3																
Fluoroquinolones - Ciprofloxacin	0.064	3	0				3																			
Penicillins - Ampicillin	8	3	0											2	1											
Quinolones - Nalidixic acid	16	3	0													3										
Tetracyclines - Tetracycline	8	3	0												3											
Trimethoprim	2	3	0										3													
Sulfonamides - Sulfamethoxazole	256	3	0																	3						

S. Braen	derup	Gallus (fowl) - he	, ,
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory	;	3
Antimicrob	ials:	lowest	highest
Aminoglycosides	- Gentamicin	0.25	32
Aminoglycosides	- Streptomycin	2	128
Amphenicols - Ch	nloramphenicol	2	64

S. Braenderup	(fowl) -	gallus laying ns
Isolates out of a monitoring program (yes/no)		
Number of isolates available in the laboratory	;	3
Antimicrobials:	lowest	highest
Cephalosporins - Cefotaxime	0.06	4
Fluoroquinolones - Ciprofloxacin	0.008	8
Penicillins - Ampicillin	0.5	32
Quinolones - Nalidixic acid	4	64
Tetracyclines - Tetracycline	1	64
Trimethoprim	0.5	32
Sulfonamides - Sulfamethoxazole	8	1024

	Concentration (u-	g/ml), number of isolates	with a concentration	of inhibition equal to
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S. Bredeney												Turk	eys - fat	tening fl	ocks											
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory													2	3												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	23	1									1	20	1				1								
Aminoglycosides - Streptomycin	16	23	1														18	4	1							
Amphenicols - Chloramphenicol	16	23	0													7	9	7								
Cephalosporins - Cefotaxime	0.5	23	0							11	8	4														
Fluoroquinolones - Ciprofloxacin	0.064	23	23								5	10	7				1									
Penicillins - Ampicillin	8	23	22											1					22							
Quinolones - Nalidixic acid	16	23	23																	23						
Tetracyclines - Tetracycline	8	23	23																	23						
Trimethoprim	2	23	0										22	1												
Sulfonamides - Sulfamethoxazole	256	23	1															4	15	3				1		

S. Brede	eney	Turk fattenin	eys - g flocks
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory	2	3
Antimicrob	oials:	lowest	highest
Aminoglycosides	- Gentamicin	0.25	32
Aminoglycosides	- Streptomycin	2	128
Amphenicols - C	hloramphenicol	2	64

Table Antimicrobial susceptibility testing of S. Bredeney in Turkeys - fattening flocks - quantitative data [Dilution method]

S. Bredeney		eys - g flocks
Isolates out of a monitoring program (yes/no)		
Number of isolates available in the laboratory	2	3
Antimicrobials:	lowest	highest
Cephalosporins - Cefotaxime	0.06	4
Fluoroquinolones - Ciprofloxacin	0.008	8
Penicillins - Ampicillin	0.5	32
Quinolones - Nalidixic acid	4	64
Tetracyclines - Tetracycline	1	64
Trimethoprim	0.5	32
Sulfonamides - Sulfamethoxazole	8	1024

S. Newport												Turk	eys - fa	ttening fl	ocks												
Isolates out of a monitoring program (yes/no) Number of isolates available in the laboratory													1	13												_	Hungary -
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	2013
Aminoglycosides - Gentamicin	2	13	0									5	7	1													
Aminoglycosides - Streptomycin	16	13	0													2	9	2									Report
Amphenicols - Chloramphenicol	16	13	0													7	6										on tre
Cephalosporins - Cefotaxime	0.5	13	0							8	4	1															ends
Fluoroquinolones - Ciprofloxacin	0.064	13	12				1						11	1													and
Penicillins - Ampicillin	8	13	13																13								Source
Quinolones - Nalidixic acid	16	13	9													1		3	9								rces
Tetracyclines - Tetracycline	8	13	13																	13							of zo
Trimethoprim	2	13	0										12	1													oonos
Sulfonamides - Sulfamethoxazole	256	13	0																1	8	4						ses

S. Newpo	ort	Turk fattenin	eys - g flocks
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory	1	3
Antimicrob	lowest	highest	
Aminoglycosides -	- Gentamicin	0.25	32
Aminoglycosides -	- Streptomycin	2	128
Amphenicols - Ch	2	64	

Table Antimicrobial susceptibility testing of S. Newport in Turkeys - fattening flocks - quantitative data [Dilution method]

S. Newport		eys - g flocks
Isolates out of a monitoring program (yes/no)		
Number of isolates available in the laboratory	1	3
Antimicrobials:	lowest	highest
Cephalosporins - Cefotaxime	0.06	4
Fluoroquinolones - Ciprofloxacin	0.008	8
Penicillins - Ampicillin	0.5	32
Quinolones - Nalidixic acid	4	64
Tetracyclines - Tetracycline	1	64
Trimethoprim	0.5	32
Sulfonamides - Sulfamethoxazole	8	1024

							\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \	<u> </u>																		
S. Agona												Gallus g	jallus (fo	owl) - lay	ing hens	8										
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory														3												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	3	0									1	2													
Aminoglycosides - Streptomycin	16	3	0														2	1								
Amphenicols - Chloramphenicol	16	3	0														3									
Cephalosporins - Cefotaxime	0.5	3	0								2	1														
Fluoroquinolones - Ciprofloxacin	0.064	3	0				2		1																	
Penicillins - Ampicillin	8	3	0											2	1											
Quinolones - Nalidixic acid	16	3	0													2	1									
Tetracyclines - Tetracycline	8	3	0												3											
Trimethoprim	2	3	0										3													
Sulfonamides - Sulfamethoxazole	256	3	0															1	1	1						

S. Agona	а	Gallus (fowl) - he	, ,
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory	;	3
Antimicrob	oials:	lowest	highest
Aminoglycosides	- Gentamicin	0.25	32
Aminoglycosides	- Streptomycin	2	128
Amphenicols - Cl	hloramphenicol	2	64

Table Antimicrobial susceptibility testing of S. Agona in Gallus gallus (fowl) - laying hens - quantitative data [Dilution method]

S. Agona	Gallus gallus (fowl) - laying hens					
	;	3				
Antimicrob	lowest	highest				
Cephalosporins -	0.06	4				
Fluoroquinolones	s - Ciprofloxacin	0.008	8			
Penicillins - Amp	icillin	0.5	32			
Quinolones - Nal	idixic acid	4	64			
Tetracyclines - T	etracycline	1	64			
Trimethoprim		0.5	32			
Sulfonamides - S	Sulfamethoxazole	8	1024			

Concentration (µg/ml), number of isolates with a concentration of inhibition equal t	to
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S. Kottbus												Turk	eys - fat	tening fl	ocks											
Isolates out of a monitoring program (yes/no) Number of isolates available																										
in the laboratory																										
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	8	0										8													
Aminoglycosides - Streptomycin	16	8	0														4	4								
Amphenicols - Chloramphenicol	16	8	0												1	7										
Cephalosporins - Cefotaxime	0.5	8	0							7	1															
Fluoroquinolones - Ciprofloxacin	0.064	8	8									8														
Penicillins - Ampicillin	8	8	0											7	1											
Quinolones - Nalidixic acid	16	8	8																	8						
Tetracyclines - Tetracycline	8	8	0											8												
Trimethoprim	2	8	0										8													
Sulfonamides - Sulfamethoxazole	256	8	0															3	3	2						

S. Kottb	Turkeys - fattening flock							
	Isolates out of a monitoring program (yes/no)							
	Number of isolates available in the laboratory							
Antimicrob	oials:	lowest	highest					
Aminoglycosides	s - Gentamicin	0.25	32					
Aminoglycosides	s - Streptomycin	2	128					
Amphenicols - C	2	64						

Table Antimicrobial susceptibility testing of S. Kottbus in Turkeys - fattening flocks - quan

- quantitative data [Dilution method]

S. Kottbu	Turkeys - fattening flocks					
		3				
Antimicrob	lowest	highest				
Cephalosporins -	0.06	4				
Fluoroquinolones	- Ciprofloxacin	0.008	8			
Penicillins - Ampid	cillin	0.5	32			
Quinolones - Nalid	dixic acid	4	64			
Tetracyclines - Te	tracycline	1	64			
Trimethoprim		0.5	32			
Sulfonamides - Su	ulfamethoxazole	8	1024			

S. Stanley												Gallus	gallus	(fowl) - b	oroilers												Ī
Isolates out of a monitoring program (yes/no)																											Hungary
Number of isolates available in the laboratory														2													1
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	2013
Aminoglycosides - Gentamicin	2	2	0										2														Re
Aminoglycosides - Streptomycin	16	2	0														2										port
Amphenicols - Chloramphenicol	16	2	0													1	1										on tre
Cephalosporins - Cefotaxime	0.5	2	0							2																	ends
Fluoroquinolones - Ciprofloxacin	0.064	2	2									2															and
Penicillins - Ampicillin	8	2	1											1					1								sourc
Quinolones - Nalidixic acid	16	2	2																	2							es
Tetracyclines - Tetracycline	8	2	0											1	1												of zo
Trimethoprim	2	2	0										2														onoses
Sulfonamides - Sulfamethoxazole	256	2	0																1	1							ses

S. Stanle	Gallus gallus (fowl) - broilers						
	Isolates out of a monitoring program (yes/no)						
	2						
Antimicrob	pials:	lowest	highest				
Aminoglycosides	- Gentamicin	0.25	32				
Aminoglycosides	- Streptomycin	2	128				
Amphenicols - Ch	hloramphenicol	2	64				

Table Antimicrobial susceptibility testing of S. Stanley in Gallus gallus (fowl) - broilers - quantitative data [Dilution method]

S. Stanley	Gallus gallus (fowl) - broilers					
Isolates out of a monitoring program (yes/no)						
Number of isolates available in the laboratory	:	2				
Antimicrobials:	lowest	highest				
Cephalosporins - Cefotaxime	0.06	4				
Fluoroquinolones - Ciprofloxacin	0.008	8				
Penicillins - Ampicillin	0.5	32				
Quinolones - Nalidixic acid	4	64				
Tetracyclines - Tetracycline	1	64				
Trimethoprim	0.5	32				
Sulfonamides - Sulfamethoxazole	8	1024				

S. Thompson							,	<u> </u>				Gallus		(fowl) - b		·										
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory		Tunggary - 2013 Solut-off N n <=0.002 <=0.004 0.008 0.015 0.016 0.03 0.06 0.12 0.25 0.5 1 2 4 8 16 32 64 128 256 512 1024 2048 >4096 2013																								
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	3	0										3													
Aminoglycosides - Streptomycin	16	3	0														3									
Amphenicols - Chloramphenicol	16	3	0														3									
Cephalosporins - Cefotaxime	0.5	3	0							3																
Fluoroquinolones - Ciprofloxacin	0.064	3	1				2				1															
Penicillins - Ampicillin	8	3	1											1	1				1							
Quinolones - Nalidixic acid	16	3	1													1	1			1						
Tetracyclines - Tetracycline	8	3	0												3											
Trimethoprim	2	3	0										3													
Sulfonamides - Sulfamethoxazole	256	3	0																	3						

S. Thom	S. Thompson						
	3						
Antimicrob	ials:	lowest	highest				
Aminoglycosides	- Gentamicin	0.25	32				
Aminoglycosides	- Streptomycin	2	128				
Amphenicols - Ch	nloramphenicol	2	64				

Table Antimicrobial susceptibility testing of S. Thompson in Gallus gallus (fowl) - broilers - quantitative data [Dilution method]

S. Thompson	Gallus gallus (fowl) - broilers						
Isolates out of a monitoring program (yes/no)							
Number of isolates available in the laboratory	;	3					
Antimicrobials:	lowest	highest					
Cephalosporins - Cefotaxime	0.06	4					
Fluoroquinolones - Ciprofloxacin	0.008	8					
Penicillins - Ampicillin	0.5	32					
Quinolones - Nalidixic acid	4	64					
Tetracyclines - Tetracycline	1	64					
Trimethoprim	0.5	32					
Sulfonamides - Sulfamethoxazole	8	1024					

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					Co	ncentra	ation (µ	g/ml), n	umber	of isola	tes with	n a con	centrati	on of ir	hibition	equal	to									
S. Senftenberg		Gallus gallus (fowl) - broilers																								
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory		4																								
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>409
Aminoglycosides - Gentamicin	2	4	0									1	3													
Aminoglycosides - Streptomycin	16	4	0														4									
Amphenicols - Chloramphenicol	16	4	1															3	1							
Cephalosporins - Cefotaxime	0.5	4	0									3	1													
Fluoroquinolones - Ciprofloxacin	0.064	4	0						1	3																
Penicillins - Ampicillin	8	4	0												2	2										
Quinolones - Nalidixic acid	16	4	0														4									
Tetracyclines - Tetracycline	8	4	0													4										
Trimethonrim	2	1	0										1													

S. Senfte	S. Senftenberg					
	Isolates out of a monitoring program (yes/no)					
	Number of isolates available in the laboratory	4	1			
Antimicrob	Antimicrobials:					
Aminoglycosides	- Gentamicin	0.25	32			
Aminoglycosides	- Streptomycin	2	128			
Amphenicols - Ch	2	64				

Sulfonamides - Sulfamethoxazole

256

Table Antimicrobial susceptibility testing of S. Senftenberg in Gallus gallus (fowl) - broilers - quantitative data [Dilution method]

S. Senftenberg	Gallus gallus (fowl) - broile			
Isolates out of a monitoring program (yes/no)				
Number of isolates available in the laboratory	4			
Antimicrobials:	lowest	highest		
Cephalosporins - Cefotaxime	0.06	4		
Fluoroquinolones - Ciprofloxacin	0.008	8		
Penicillins - Ampicillin	0.5	32		
Quinolones - Nalidixic acid	4	64		
Tetracyclines - Tetracycline	1	64		
Trimethoprim	0.5	32		
Sulfonamides - Sulfamethoxazole	8	1024		

Concentration (µg/ml),	number of isolates with a	concentration of inhibition equal to
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S. Derby												Gallus	gallus	(fowl) - b	oroilers											
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory														1												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	1	0										1													
Aminoglycosides - Streptomycin	16	1	0															1								
Amphenicols - Chloramphenicol	16	1	0														1									
Cephalosporins - Cefotaxime	0.5	1	0							1																
Fluoroquinolones - Ciprofloxacin	0.064	1	0				1																			
Penicillins - Ampicillin	8	1	0											1												
Quinolones - Nalidixic acid	16	1	0													1										
Tetracyclines - Tetracycline	8	1	0												1											
Trimethoprim	2	1	0										1													
Sulfonamides - Sulfamethoxazole	256	1	0																	1						

S. Derby	Gallus gallus (fowl) - broilers					
	Isolates out of a monitoring program (yes/no)					
	Number of isolates available in the laboratory	,	1			
Antimicrob	lowest	highest				
Aminoglycosides	- Gentamicin	0.25	32			
Aminoglycosides	- Streptomycin	2	128			
Amphenicols - Cl	nloramphenicol	2	64			

Table Antimicrobial susceptibility testing of S. Derby in Gallus gallus (fowl) - broilers - quantitative data [Dilution method]

S. Derby	S. Derby							
	Isolates out of a monitoring program (yes/no)							
	1							
Antimicrob	lowest	highest						
Cephalosporins -	0.06	4						
Fluoroquinolones	s - Ciprofloxacin	0.008	8					
Penicillins - Amp	icillin	0.5	32					
Quinolones - Nal	idixic acid	4	64					
Tetracyclines - T	etracycline	1	64					
Trimethoprim		0.5	32					
Sulfonamides - S	Sulfamethoxazole	8	1024					

S. Saintpaul		Turkeys - fattening flocks																								
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory		Topic N n <=0.002 <=0.004 0.008 0.015 0.016 0.03 0.06 0.12 0.25 0.5 1 2 4 8 16 32 64 128 256 512 1024 2048 >4096 2018																								
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	10	0									2	7	1												
Aminoglycosides - Streptomycin	16	10	9															1	8	1						
Amphenicols - Chloramphenicol	16	10	0														9	1								
Cephalosporins - Cefotaxime	0.5	10	0							4	6															
Fluoroquinolones - Ciprofloxacin	0.064	10	0				1		9																	
Penicillins - Ampicillin	8	10	1											1	7	1			1							
Quinolones - Nalidixic acid	16	10	0													9	1									
Tetracyclines - Tetracycline	8	10	10																	10						
Trimethoprim	2	10	1										9						1							
Sulfonamides - Sulfamethoxazole	256	10	10																					10		

S. Saintpa	S. Saintpaul					
	solates out of a monitoring program (yes/no)					
N ir	10					
Antimicrobia	lowest	highest				
Aminoglycosides - 0	Gentamicin	0.25	32			
Aminoglycosides - \$	Streptomycin	2	128			
Amphenicols - Chlo	oramphenicol	2	64			

Table Antimicrobial susceptibility testing of S. Saintpaul in Turkeys - fattening flocks - quantitative data [Dilution method]

S. Saint	oaul	Turk fattenin	eys - g flocks
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory	1	0
Antimicrob	oials:	lowest	highest
Cephalosporins -	Cefotaxime	0.06	4
Fluoroquinolones	- Ciprofloxacin	0.008	8
Penicillins - Ampi	cillin	0.5	32
Quinolones - Nali	idixic acid	4	64
Tetracyclines - Te	etracycline	1	64
Trimethoprim		0.5	32
Sulfonamides - S	ulfamethoxazole	8	1024

S. Tenne	essee	. ,	gallus laying ns
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory	1	3
Antimicrob	oials:	lowest	highest
Aminoglycosides	- Gentamicin	0.25	32
Aminoglycosides	- Streptomycin	2	128
Amphenicols - C	hloramphenicol	2	64

Table Antimicrobial susceptibility testing of S. Tennessee in Gallus gallus (fowl) - laying hens - quantitative data [Dilution method]

S. Tennessee	Gallus gallus (fowl) - laying hens			
Isolates out of a monitoring program (yes/no)				
Number of isolates available in the laboratory	13			
Antimicrobials:	lowest	highest		
Cephalosporins - Cefotaxime	0.06	4		
Fluoroquinolones - Ciprofloxacin	0.008	8		
Penicillins - Ampicillin	0.5	32		
Quinolones - Nalidixic acid	4	64		
Tetracyclines - Tetracycline	1	64		
Trimethoprim	0.5	32		
Sulfonamides - Sulfamethoxazole	8	1024		

Concentration	g/ml), number of isolates with a concentration of inhibition equal to	

S. Veneziana												Gallus g	allus (fo	wl) - lay	ing hens	S											
Isolates out of a monitoring program (yes/no)																											ungary
Number of isolates available in the laboratory														1													
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	2013
Aminoglycosides - Gentamicin	2	1	0									1															7
Aminoglycosides - Streptomycin	16	1	0													1											0
Amphenicols - Chloramphenicol	16	1	0														1										
Cephalosporins - Cefotaxime	0.5	1	0							1																	מומ
Fluoroquinolones - Ciprofloxacin	0.064	1	0				1																				٩
Penicillins - Ampicillin	8	1	0											1													SOU
Quinolones - Nalidixic acid	16	1	0													1											G
Tetracyclines - Tetracycline	8	1	0												1												2
Trimethoprim	2	1	0										1														
Sulfonamides - Sulfamethoxazole	256	1	0																1								Coes

S. Venez	ziana	Gallus gallus (fowl) - laying hens					
	Number of isolates available in the laboratory	1					
Antimicrob	lowest	highest					
Aminoglycosides	- Gentamicin	0.25	32				
Aminoglycosides	- Streptomycin	2	128				
Amphenicols - Cl	hloramphenicol	2	64				

Table Antimicrobial susceptibility testing of S. Veneziana in Gallus gallus (fowl) - laying hens - quantitative data [Dilution method]

S. Venez	Gallus gallus (fowl) - laying hens				
	Isolates out of a monitoring program (yes/no)				
	1				
Antimicrob	lowest	highest			
Cephalosporins -	0.06	4			
Fluoroquinolones	0.008	8			
Penicillins - Ampi	0.5	32			
Quinolones - Nali	dixic acid	4	64		
Tetracyclines - Te	etracycline	1	64		
Trimethoprim		0.5	32		
Sulfonamides - S	8	1024			

							т. (р.	9,,,		0. 10010		i a com				. oqua.										
S. 4,5:i:-													Meat f	rom pig												
Isolates out of a monitoring program (yes/no)														no												
Number of isolates available in the laboratory														4												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	4	0										4													
Aminoglycosides - Streptomycin	32	4	3																1		1	2				
Amphenicols - Chloramphenicol	16	4	1													2	1				1					
Cephalosporins - Cefotaxime	0.5	4	0								4															
Fluoroquinolones - Ciprofloxacin	0.064	4	1						3				1													
Penicillins - Ampicillin	8	4	3													1					3					
Quinolones - Nalidixic acid	16	4	1													2	1					1				
Sulfonamides	256	4	3																	1				2	1	
Tetracyclines - Tetracycline	8	4	3											1						1	2					
Trimethoprim	2	4	0									2	2													
Fully sensitive		1	1	1																						
Resistant to 4 antimicrobials		2	2	2																						
Resistant to >4 antimicrobials		1	1	1																						

Table Antimicrobial susceptibility testing of S. 4,5:i:- in Meat from pig - food sample - meat - quantitative data [Dilution method]

S. 4,5:i:-	Meat from pig						
	Isolates out of a monitoring program (yes/no)	no					
	Number of isolates available in the laboratory	4	1				
Antimicrob	oials:	lowest	highest				
Aminoglycosides	- Gentamicin						
Aminoglycosides							
Amphenicols - Ch	nloramphenicol						
Cephalosporins -	Cefotaxime						
Fluoroquinolones	- Ciprofloxacin						
Penicillins - Ampi	cillin						
Quinolones - Nali	idixic acid						
Sulfonamides							
Tetracyclines - Te	etracycline						
Trimethoprim							
Fully sensitive							
Resistant to 4 and	timicrobials						
Resistant to >4 a	ntimicrobials						

					Co	ncentra	ation (µ	g/ml), n	umber	of isola	tes with	a con	centrat	on of ir	hibition	n equal	to										_
S. Virchow		Turkeys - fattening flocks															_										
Isolates out of a monitoring program (yes/no)																											Hungary
Number of isolates available in the laboratory														1													1
Antimicrobials:	Cut-off value	Ν	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	2013
Aminoglycosides - Gentamicin	2	1	0										1														Re
Aminoglycosides - Streptomycin	16	1	1																	1							Report
Amphenicols - Chloramphenicol	16	1	1																	1							on tra
Cephalosporins - Cefotaxime	0.5	1	0							1																	trends
Fluoroquinolones - Ciprofloxacin	0.064	1	1								1																and
Penicillins - Ampicillin	8	1	0												1												S
Quinolones - Nalidixic acid	16	1	1																	1							ources
Tetracyclines - Tetracycline	8	1	1																	1							of zo
Trimethoprim	2	1	1																1								zoonos
Sulfonamides - Sulfamethoxazole	256	1	1																					1			ses

S. Vircho	ow .	Turkeys - fattening flocks					
	Isolates out of a monitoring program (yes/no)						
	Number of isolates available in the laboratory		1				
Antimicrob	oials:	lowest	highest				
Aminoglycosides	- Gentamicin	0.25	32				
Aminoglycosides	- Streptomycin	2	128				
Amphenicols - Ch	Amphenicols - Chloramphenicol						

Table Antimicrobial susceptibility testing of S. Virchow in Turkeys - fattening flocks - quantitative data [Dilution method]

S. Vircho	Turkeys - fattening flocks			
	Isolates out of a monitoring program (yes/no)			
	Number of isolates available in the laboratory		1	
Antimicrob	oials:	lowest	highest	
Cephalosporins -	- Cefotaxime	0.06	4	
Fluoroquinolones	s - Ciprofloxacin	0.008	8	
Penicillins - Amp	icillin	0.5	32	
Quinolones - Nal	idixic acid	4	64	
Tetracyclines - T	etracycline	1	64	
Trimethoprim		0.5	32	
Sulfonamides - S	8	1024		

Concentration (µg/ml), nun	ber of isolates with a	concentration of inhibition equ	ual to
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S. Infantis												Gallus g	allus (fo	wl) - lay	ing hens	5											ı.
Isolates out of a monitoring program (yes/no) Number of isolates available														6												_	Hungary
in the laboratory			1						1			1		· 													- 2(
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	013
Aminoglycosides - Gentamicin	2	6	0									5	1														Re
Aminoglycosides - Streptomycin	16	6	5														1		4	1							port
Amphenicols - Chloramphenicol	16	6	0														3	3									on tre
Cephalosporins - Cefotaxime	0.5	6	0								2	4															ends
Fluoroquinolones - Ciprofloxacin	0.064	6	5						1			1	4														and
Penicillins - Ampicillin	8	6	0											1	2	3											
Quinolones - Nalidixic acid	16	6	5													1				5							sources
Tetracyclines - Tetracycline	8	6	5												1					5							of zo
Trimethoprim	2	6	0										6														onos
Sulfonamides - Sulfamethoxazole	256	6	5																	1				5			ses

S. Infant	is	Gallus gallu (fowl) - layin hens		
	Isolates out of a monitoring program (yes/no)			
	Number of isolates available in the laboratory	6	6	
Antimicrob	oials:	lowest	highest	
Aminoglycosides	- Gentamicin	0.25	32	
Aminoglycosides	- Streptomycin	2	128	
Amphenicols - Cl	hloramphenicol	2	64	

Table Antimicrobial susceptibility testing of S. Infantis in Gallus gallus (fowl) - laying hens - quantitative data [Dilution method]

S. Infant	S. Infantis							
	Isolates out of a monitoring program (yes/no)							
	Number of isolates available in the laboratory	(6					
Antimicrob	oials:	lowest	highest					
Cephalosporins -	Cefotaxime	0.06	4					
Fluoroquinolones	s - Ciprofloxacin	0.008	8					
Penicillins - Amp	icillin	0.5	32					
Quinolones - Nal	idixic acid	4	64					
Tetracyclines - T	etracycline	1	64					
Trimethoprim		0.5	32					
Sulfonamides - S	8	1024						

Concentration (µg/ml), number of isolates with a	concentration of inhibition equal to
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S. Typhimurium							4.	<u> </u>						rom pig		•										
Isolates out of a monitoring program (yes/no)													г	10												
Number of isolates available in the laboratory													1	13												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	13	0									4	7	2												
Aminoglycosides - Streptomycin	32	13	9															2	2		2	7				
Amphenicols - Chloramphenicol	16	13	2												2	8		1		1	1					
Cephalosporins - Cefotaxime	0.5	13	0								13															
Fluoroquinolones - Ciprofloxacin	0.064	13	0						13																	
Penicillins - Ampicillin	8	13	11											1		1				1	10					
Quinolones - Nalidixic acid	16	13	0												4	5	2	2								
Sulfonamides	256	13	10																1		2		2	7	1	
Tetracyclines - Tetracycline	8	13	11											2					2	2	3	4				
Trimethoprim	2	13	2									3	4	4				2								
Fully sensitive		1	1	1																						
Resistant to 1 antimicrobial		2	2	2																						
Resistant to 4 antimicrobials		8	8	8																						
Resistant to >4 antimicrobials		2	2	2																						

S. Typhi	Meat from pig									
	Isolates out of a monitoring program (yes/no)									
	Number of isolates available in the laboratory	1	3							
Antimicrob	oials:	lowest	highest							
Aminoglycosides	- Gentamicin									
Aminoglycosides	- Streptomycin									
Amphenicols - Ch	nloramphenicol									
Cephalosporins -	Cefotaxime									
Fluoroquinolones	- Ciprofloxacin									
Penicillins - Ampi	cillin									
Quinolones - Nali	idixic acid									
Sulfonamides										
Tetracyclines - Te	etracycline									
Trimethoprim										
Fully sensitive										
Resistant to 1 an	timicrobial									
Resistant to 4 an	timicrobials									
Resistant to >4 a	ntimicrobials									

and

sources

of zoonoses

S. Kedo	ugou	Turk fattenin	eys - g flocks
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory		1
Antimicrob	oials:	lowest	highest
Aminoglycosides	- Gentamicin	0.25	32
Aminoglycosides	- Streptomycin	2	128
Amphenicols - Ch	hloramphenicol	2	64

Table Antimicrobial susceptibility testing of S. Kedougou in Turkeys - fattening flocks - quantitative data [Dilution method]

S. Kedougou		eys - ig flocks
Isolates out of a monitori program (yes/no)	ng	
Number of isolates availate in the laboratory	able	1
Antimicrobials:	lowest	highest
Cephalosporins - Cefotaxime	0.06	4
Fluoroquinolones - Ciprofloxacin	0.008	8
Penicillins - Ampicillin	0.5	32
Quinolones - Nalidixic acid	4	64
Tetracyclines - Tetracycline	1	64
Trimethoprim	0.5	32
Sulfonamides - Sulfamethoxazole	8	1024

Table Antimicrobial susceptibility testing of S. Typhimurium in Meat from other poultry species - food sample - quantitative data [Dilution method]

Concentration (µg/ml), number of isolates with a concentration of inhibition equal to

S. Typhimurium						ncentra	шоп (р	g/1111), 11	umber	oi isola		Meat fro					to									
Isolates out of a monitoring program (yes/no)													n	0												
Number of isolates available in the laboratory														9												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	9	0									2	4	3												
Aminoglycosides - Streptomycin	32	9	2														1	3	3	2						
Amphenicols - Chloramphenicol	16	9	0												3	5	1									
Cephalosporins - Cefotaxime	0.5	9	0								9															
Fluoroquinolones - Ciprofloxacin	0.064	9	1						8			1														
Penicillins - Ampicillin	8	9	1											7		1					1					
Quinolones - Nalidixic acid	16	9	1												1	4	3					1				
Sulfonamides	256	9	4																1	4				4		
Tetracyclines - Tetracycline	8	9	1											2	5	1					1					
Trimethoprim	2	9	0									2	6	1												
Fully sensitive		5	5	5																						
Resistant to 1 antimicrobial		2	2	2																						
Resistant to 2 antimicrobials		1	1	1																						
Resistant to >4 antimicrobials		1	1	1																						

Table Antimicrobial susceptibility testing of S. Typhimurium in Meat from other poultry species - food sample - quantitative data [Dilution method]

S. Typhimurium			om other species
Isolates out of a mo program (yes/no)	onitoring	r	10
Number of isolates in the laboratory	available		9
Antimicrobials:		lowest	highest
Aminoglycosides - Gentamicin			
Aminoglycosides - Streptomycin			
Amphenicols - Chloramphenicol			
Cephalosporins - Cefotaxime			
Fluoroquinolones - Ciprofloxacin			
Penicillins - Ampicillin			
Quinolones - Nalidixic acid			
Sulfonamides			
Tetracyclines - Tetracycline			
Trimethoprim			
Fully sensitive			
Resistant to 1 antimicrobial			
Resistant to 2 antimicrobials			
Resistant to >4 antimicrobials			

S. Montevideo							·					Gallus	gallus	(fowl) - b	oroilers	·											ı
Isolates out of a monitoring program (yes/no)																											Hungary
Number of isolates available in the laboratory													•	2													1
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	2013
Aminoglycosides - Gentamicin	2	2	0										2														Re
Aminoglycosides - Streptomycin	16	2	0														1	1									Report
Amphenicols - Chloramphenicol	16	2	0														2										on tr
Cephalosporins - Cefotaxime	0.5	2	0							2																	trends
Fluoroquinolones - Ciprofloxacin	0.064	2	0				2																				and
Penicillins - Ampicillin	8	2	0											1	1												S
Quinolones - Nalidixic acid	16	2	0													2											ources
Tetracyclines - Tetracycline	8	2	0												2												of zo
Trimethoprim	2	2	0										2														zoonoses
Sulfonamides - Sulfamethoxazole	256	2	0																	2							ses

S. Montevi	deo		gallus broilers					
	plates out of a monitoring ogram (yes/no)							
	mber of isolates available the laboratory	2	2					
Antimicrobial	ls:	lowest	highest					
Aminoglycosides - Ge	entamicin	0.25	32					
Aminoglycosides - St	reptomycin	2	128					
Amphenicols - Chlora	mphenicols - Chloramphenicol							

Table Antimicrobial susceptibility testing of S. Montevideo in Gallus gallus (fowl) - broilers

 quantitative data [Dilution method]

S. Montevideo		gallus broilers
Isolates out of a monitoring program (yes/no)		
Number of isolates available in the laboratory	:	2
Antimicrobials:	lowest	highest
Cephalosporins - Cefotaxime	0.06	4
Fluoroquinolones - Ciprofloxacin	0.008	8
Penicillins - Ampicillin	0.5	32
Quinolones - Nalidixic acid	4	64
Tetracyclines - Tetracycline	1	64
Trimethoprim	0.5	32
Sulfonamides - Sulfamethoxazole	8	1024

Table Antimicrobial susceptibility testing of S. Typhimurium in Meat from bovine animals - food sample - meat - quantitative data [Dilution method]

S. Typhimurium												Meat	t from bo	ovine an	imals											
Isolates out of a monitoring program (yes/no)													y	es												
Number of isolates available in the laboratory														1												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	1	0										1													
Aminoglycosides - Streptomycin	32	1	0																1							
Amphenicols - Chloramphenicol	16	1	0													1										
Cephalosporins - Cefotaxime	0.5	1	0								1															
Fluoroquinolones - Ciprofloxacin	0.064	1	0						1																	
Penicillins - Ampicillin	8	1	0												1											
Quinolones - Nalidixic acid	16	1	0												1											
Sulfonamides	256	1	0																		1					
Tetracyclines - Tetracycline	8	1	0											1												
Trimethoprim	2	1	0										1													
Fully sensitive		1	1	1																						

S. Typhimurium		Meat bovine	from animals
Isolates out of a program (yes/n	•	ує	es
Number of isola in the laborator		1	ı
Antimicrobials:		lowest	highest
Aminoglycosides - Gentamicin			
Aminoglycosides - Streptomycin			·

Table Antimicrobial susceptibility testing of S. Typhimurium in Meat from bovine animals - food sample - meat - quantitative data [Dilution method]

S. Typhi	murium	Meat bovine	from animals						
	Isolates out of a monitoring program (yes/no)	ye	es						
	Number of isolates available in the laboratory		ı						
Antimicrob	oials:	lowest	highest						
Amphenicols - C	hloramphenicol								
Cephalosporins -	- Cefotaxime								
Fluoroquinolones	s - Ciprofloxacin								
Penicillins - Amp	icillin								
Quinolones - Nal	idixic acid								
Sulfonamides									
Tetracyclines - T	etracycline								
Trimethoprim									
Fully sensitive	Fully sensitive								

								3. //								- 1											
S. Ohio												Gallus	gallus	(fowl) - b	oroilers												=
Isolates out of a monitoring program (yes/no)																											idigaly
Number of isolates available in the laboratory														1													
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	70
Aminoglycosides - Gentamicin	2	1	0										1														
Aminoglycosides - Streptomycin	16	1	0													1											Cool
Amphenicols - Chloramphenicol	16	1	0															1									-
Cephalosporins - Cefotaxime	0.5	1	0										1														נוכונט
Fluoroquinolones - Ciprofloxacin	0.064	1	0							1																	2
Penicillins - Ampicillin	8	1	0													1											
Quinolones - Nalidixic acid	16	1	0														1										
Tetracyclines - Tetracycline	8	1	0													1											2
Trimethoprim	2	1	0										1														20010303
Sulfonamides - Sulfamethoxazole	256	1	0																	1							

Concentration (µg/ml), number of isolates with a concentration of inhibition equal to

S. Ohio			gallus broilers
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory	,	1
Antimicrob	pials:	lowest	highest
Aminoglycosides	- Gentamicin	0.25	32
Aminoglycosides	- Streptomycin	2	128
Amphenicols - Cl	hloramphenicol	2	64

Table Antimicrobial susceptibility testing of S. Ohio in Gallus gallus (fowl) - broilers - quantitative data [Dilution method]

S. Ohio		Gallus gallus (fowl) - broiler								
	Isolates out of a monitoring program (yes/no)									
	Number of isolates available in the laboratory		1							
Antimicrol	Antimicrobials:									
Cephalosporins	0.06	4								
Fluoroquinolone	es - Ciprofloxacin	0.008	8							
Penicillins - Amp	picillin	0.5	32							
Quinolones - Na	ılidixic acid	4	64							
Tetracyclines - T	Tetracycline	1	64							
Trimethoprim		0.5	32							
Sulfonamides - S	Sulfamethoxazole	8	1024							

Table Antimicrobial susceptibility testing of S. Typhimurium, monophasic in Meat from other poultry species - food sample - meat - quantitative data [Dilution method]

Concentration (µg/ml), number of isolates with a concentration of inhibition equal to

S. Typhimurium, monophasic		Meat from other poultry species																								
Isolates out of a monitoring program (yes/no)													n	0												
Number of isolates available in the laboratory														1												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	1	0											1												
Aminoglycosides - Streptomycin	32	1	1																			1				
Amphenicols - Chloramphenicol	16	1	0														1									
Cephalosporins - Cefotaxime	0.5	1	0								1															
Fluoroquinolones - Ciprofloxacin	0.064	1	0						1																	
Penicillins - Ampicillin	8	1	1																		1					
Quinolones - Nalidixic acid	16	1	0													1										
Sulfonamides	256	1	1																					1		
Tetracyclines - Tetracycline	8	1	1																		1					
Trimethoprim	2	1	0										1													
Resistant to 4 antimicrobials		1	1	1																						

5 I vonimumimim	Meat fro				
Isolates out of a monitoring program (yes/no)	no				
Number of isolates available in the laboratory	1	ı			
Antimicrobials:	lowest	highest			
Aminoglycosides - Gentamicin					
Aminoglycosides - Streptomycin					

Table Antimicrobial susceptibility testing of S. Typhimurium, monophasic in Meat from other poultry species - food sample - meat - quantitative data [Dilution method]

	S. Typhimurium, monophasic								
	r	10							
		1							
Antimicrob	lowest	highest							
Amphenicols - Ch									
Cephalosporins -									
Fluoroquinolones	- Ciprofloxacin								
Penicillins - Ampi	cillin								
Quinolones - Nali	dixic acid								
Sulfonamides									
Tetracyclines - Te	etracycline								
Trimethoprim									
Resistant to 4 and	timicrobials								

	Concentration (µg/ml), number of isolates with a concentration of inhibition equal to																									
S. Derby													Meat fr	om pig												
Isolates out of a monitoring program (yes/no)													ує	es												
Number of isolates available in the laboratory		7																								
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	7	0									2	3	1	1											
Aminoglycosides - Streptomycin	32	7	3															1	3		2	1				
Amphenicols - Chloramphenicol	16	7	3													2	1	1	1		2					
Cephalosporins - Cefotaxime	0.5	7	0								6	1														
Fluoroquinolones - Ciprofloxacin	0.064	7	1						6				1													
Penicillins - Ampicillin	8	7	2											1	3	1			1		1					
Quinolones - Nalidixic acid	16	7	1											1	1	3	1					1				
Sulfonamides	256	7	4																1	1	1			4		
Tetracyclines - Tetracycline	8	7	5														2			2	2	1				
Trimethoprim	2	7	3										1	2	1	1		2								
Fully sensitive		1	1	1																						
Resistant to 1 antimicrobial		2	2	2																						
Resistant to 4 antimicrobials		1	1	1																						
Resistant to >4 antimicrobials		3	3	3									_	_												

Table Antimicrobial susceptibility testing of S. Derby in Meat from pig - food sample - meat - quantitative data [Dilution method]

S. Derby	1	Meat fr	om pig
	Isolates out of a monitoring program (yes/no)	ye	es
	Number of isolates available in the laboratory	:	7
Antimicrob	oials:	lowest	highest
Aminoglycosides			
Aminoglycosides			
Amphenicols - Cl			
Cephalosporins -			
Fluoroquinolones			
Penicillins - Ampi	icillin		
Quinolones - Nal	idixic acid		
Sulfonamides			
Tetracyclines - To	etracycline		
Trimethoprim			
Fully sensitive			
Resistant to 1 an	timicrobial		
Resistant to 4 an	timicrobials		
Resistant to >4 a	ntimicrobials		

S. Typhimurium		Gallus gallus (fowl) - broilers																									
Isolates out of a monitoring program (yes/no)																											Hungary
Number of isolates available in the laboratory														1													1
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	2013
Aminoglycosides - Gentamicin	2	1	0										1														Rep
Aminoglycosides - Streptomycin	16	1	0														1										Report
Amphenicols - Chloramphenicol	16	1	0													1											on tr
Cephalosporins - Cefotaxime	0.5	1	0							1																	trends
Fluoroquinolones - Ciprofloxacin	0.064	1	0						1																		and
Penicillins - Ampicillin	8	1	0												1												S
Quinolones - Nalidixic acid	16	1	0													1											ources
Tetracyclines - Tetracycline	8	1	0												1												of zo
Trimethoprim	2	1	0										1														zoonoses
Sulfonamides - Sulfamethoxazole	256	1	0																1								ses

S. Typhir	nurium		gallus broilers
	Isolates out of a monitoring program (yes/no)		
		1	
Antimicrob	ials:	lowest	highest
Aminoglycosides -	- Gentamicin	0.25	32
Aminoglycosides -	2	128	
Amphenicols - Chi	2	64	

Table Antimicrobial susceptibility testing of S. Typhimurium in Gallus gallus (fowl) - broilers - quantitative data [Dilution method]

S. Typhi	murium	Gallus gallus (fowl) - broiler										
	Isolates out of a monitoring program (yes/no)											
	Number of isolates available in the laboratory		1									
Antimicrob	Antimicrobials:											
Cephalosporins -	0.06	4										
Fluoroquinolones	s - Ciprofloxacin	0.008	8									
Penicillins - Ampi	icillin	0.5	32									
Quinolones - Nali	idixic acid	4	64									
Tetracyclines - Te	etracycline	1	64									
Trimethoprim		0.5	32									
Sulfonamides - S	iulfamethoxazole	8	1024									

S. Kentucky	Meat from turkey yes																									
Isolates out of a monitoring program (yes/no)													у	es												
Number of isolates available in the laboratory														16												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	16	16														5	11								
Aminoglycosides - Streptomycin	32	16	15																1	1	10	4				
Amphenicols - Chloramphenicol	16	16	0													16										
Cephalosporins - Cefotaxime	0.5	16	0								14	2														
Fluoroquinolones - Ciprofloxacin	0.064	16	16														10	6								
Penicillins - Ampicillin	8	16	16																		16					
Quinolones - Nalidixic acid	16	16	16																			16				
Sulfonamides	256	16	16																					15	1	
Tetracyclines - Tetracycline	8	16	16																	1	12	3				
Trimethoprim	2	16	0									1	7	8												
Resistant to >4 antimicrobials		16	16	16																						

S. Kentucky		Meat turl	
Isolates out of a program (yes/no	уe	es	
Number of isolate in the laboratory	1	6	
Antimicrobials:		lowest	highest
Aminoglycosides - Gentamicin			
Aminoglycosides - Streptomycin			

S. Kentucky			from							
	ites out of a monitoring ram (yes/no)	ye	es							
	Number of isolates available in the laboratory									
Antimicrobials	s:	lowest	highest							
Amphenicols - Chloran										
Cephalosporins - Cefot										
Fluoroquinolones - Cip										
Penicillins - Ampicillin										
Quinolones - Nalidixic	acid									
Sulfonamides										
Tetracyclines - Tetracy	cline									
Trimethoprim										
Resistant to >4 antimic	robials									

Table Antimicrobial susceptibility testing of S. Kentucky in Meat from broilers (Gallus gallus) - food sample - meat - quantitative data [Dilution method]

Concentration (µg/ml), number of isolates with a concentration of inhibition equal to

							<u> </u>	<i>,</i> , .																		
S. Kentucky		Meat from broilers (Gallus gallus)																								
Isolates out of a monitoring program (yes/no)		yes																								
Number of isolates available in the laboratory													1	0												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	10	10														2	7	1							
Aminoglycosides - Streptomycin	32	10	10																	3	6	1				
Amphenicols - Chloramphenicol	16	10	0													10										
Cephalosporins - Cefotaxime	0.5	10	0								10															
Fluoroquinolones - Ciprofloxacin	0.064	10	10														7	3								
Penicillins - Ampicillin	8	10	10																		10					
Quinolones - Nalidixic acid	16	10	10																			10				
Sulfonamides	256	10	10																					10		
Tetracyclines - Tetracycline	8	10	10																	1	9					
Trimethoprim	2	10	0									3	2	5												
Resistant to >4 antimicrobials		10	10	10										_					_							

S. Kentuck	у	Meat from broilers (Gallus gallus)			
Isolates out of a monitoring program (yes/no)		yes			
	imber of isolates available the laboratory	10			
Antimicrobia	ls:	lowest	highest		
Aminoglycosides - Go	entamicin				
Aminoglycosides - St	reptomycin	·			

Table Antimicrobial susceptibility testing of S. Kentucky in Meat from broilers (Gallus gallus) - food sample - meat - quantitative data [Dilution method]

S. Kentu	cky	broilers	from (Gallus lus)					
	Isolates out of a monitoring program (yes/no)	yı	es					
	Number of isolates available in the laboratory							
Antimicrob	lowest	highest						
Amphenicols - Ch								
Cephalosporins -								
Fluoroquinolones	- Ciprofloxacin							
Penicillins - Ampi	cillin							
Quinolones - Nali	dixic acid							
Sulfonamides								
Tetracyclines - Te	etracycline							
Trimethoprim								
Resistant to >4 a	ntimicrobials							

Concentration (µg/ml)	, number of isolates with	a concentration of inhil	bition equal to
-----------------------	---------------------------	--------------------------	-----------------

S. Stanley		Meat from turkey																								
Isolates out of a monitoring program (yes/no)		yes																								
Number of isolates available in the laboratory		26																								
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	26	0									11	10	5												
Aminoglycosides - Streptomycin	32	26	1														6	12	7	1						
Amphenicols - Chloramphenicol	16	26	0												4	11	11									
Cephalosporins - Cefotaxime	0.5	26	0								26															
Fluoroquinolones - Ciprofloxacin	0.064	26	25							1	16	8	1													
Penicillins - Ampicillin	8	26	8										5	5	3	5		1			7					
Quinolones - Nalidixic acid	16	26	26																	1	4	21				
Sulfonamides	256	26	5														2	1	7	6	5		1	3	1	
Tetracyclines - Tetracycline	8	26	0											15	11											
Trimethoprim	2	26	0									4	13	9												
Resistant to 1 antimicrobial		1	1	1																						
Resistant to 2 antimicrobials		14	14	14																						
Resistant to 3 antimicrobials		8	8	8																						
Resistant to 4 antimicrobials		3	3	3																						

S. Stanle	èλ	Meat turl			
	Isolates out of a monitoring program (yes/no)	yes			
	Number of isolates available in the laboratory	26			
Antimicrob	ials:	lowest	highest		
Aminoglycosides	- Gentamicin				
Aminoglycosides	- Streptomycin				
Amphenicols - Ch	nloramphenicol				
Cephalosporins -	Cefotaxime				
Fluoroquinolones					
Penicillins - Ampi	cillin				
Quinolones - Nali	dixic acid				
Sulfonamides					
Tetracyclines - Te	etracycline				
Trimethoprim					
Resistant to 1 and	timicrobial				
Resistant to 2 and	timicrobials				
Resistant to 3 and	timicrobials				
Resistant to 4 and	timicrobials				

Table Cut-off values for antibiotic resistance testing of Salmonella in Animals

Test Method Used	Standard methods used for testing

			Concentration (microg/ml)	Zone diameter (mm)
		Standard	Resistant >	Resistant <=
Aminoglycosides	Gentamicin		2	
	Streptomycin		32	
Amphenicols	Chloramphenicol		16	
Cephalosporins	Cefotaxime		0.5	
	Ceftazidime		2	
Fluoroquinolones	Ciprofloxacin		0.064	
Penicillins	Ampicillin		8	
Quinolones	Nalidixic acid		16	
Sulfonamides	Sulfonamides		256	
Tetracyclines	Tetracycline		8	
Trimethoprim	Trimethoprim		2	

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Table Cut-off values for antibiotic resistance testing of Salmonella in Feed

Test Method Used	Standard methods used for testing

			Concentration (microg/ml)	Zone diameter (mm)
		Standard	Resistant >	Resistant <=
Aminoglycosides	Gentamicin		2	
	Streptomycin		32	
Amphenicols	Chloramphenicol		16	
Cephalosporins	Cefotaxime		0.5	
	Ceftazidime		2	
Fluoroquinolones	Ciprofloxacin		0.064	
Penicillins	Ampicillin		8	
Quinolones	Nalidixic acid		16	
Sulfonamides	Sulfonamides		256	
Tetracyclines	Tetracycline		8	
Trimethoprim	Trimethoprim		2	

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Table Cut-off values for antibiotic resistance testing of Salmonella in Food

Test Method Used	Standard
Broth dilution	EFSA

Standard methods used for testing	
EFSA	

			Concentration (microg/ml)	Zone diameter (mm)
		Standard	Resistant >	Resistant <=
Aminoglycosides	Gentamicin	EFSA	2	
	Streptomycin	EFSA	32	
Amphenicols	Chloramphenicol	EFSA	16	
Cephalosporins	Cefotaxime	EFSA	0.5	
	Ceftazidime	EFSA	2	
Fluoroquinolones	Ciprofloxacin	EFSA	0.064	
Penicillins	Ampicillin	EFSA	8	
Quinolones	Nalidixic acid	EFSA	16	
Sulfonamides	Sulfonamides	EFSA	256	
Tetracyclines	Tetracycline	EFSA	8	
Trimethoprim	Trimethoprim	EFSA	2	_

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2.2 CAMPYLOBACTERIOSIS

2.2.1 General evaluation of the national situation

A. Thermophilic Campylobacter general evaluation

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

The main source of human campylobacter infections in Hungary is raw meat especially poultry meat. The seasonal prevalence of campylobacters in raw chicken meat shows a strong correlation with the seasonal distribution of human cases. The prevalence in raw milk is low, but it can mean a possible source in some cases. As typing of Campylobacter of food origin is not performed at a large scale, PFGE and other molecular based methods are used mainly for outbreak invetigations and in small scale regional studies, the identification of sources should be improved in the future.

Recent actions taken to control the zoonoses

Actions specifically used for the control of campylobacters are not implemented in Hungary. Hygienic measurements used in the primary production (all in -all out systems, cleaning, desinfection, pest control)HACCP and GHP systems at slaughterhouses, improvement of the packaging of raw meat, labelling the minced meat and meat preparations with the requirement of heat treatment before consumption are the main actions in use.

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2.2.2 Campylobacteriosis in humans

A. Thermophilic Campylobacter in humans

Reporting system in place for the human cases

There are around 80 communicable diseases notifiable in Hungary based on legal background. The physician (in primary health care, specialist care, inpatient medical institution or pathology) who first diagnoses a case of a notifiable communicable disease (even the suspicion of the disease!) immediately reports data of case to the first level of the epidemiological network (municipal institute) of National Public Health and Medical Officer's Service (NPHMOS). Data must be reported both at the beginning and at end of the illness (recovery/death, result of laboratory test). The NPHMOS has a nationwide electronic system for registering and analysing data of communicable diseases in a combined national database, so the system provides online connection between the three levels (municipal, regional and national level – National Centre of Epidemiology - NCE) of the organization. The NCE prepares reports regularly (weekly, monthly, yearly) to the Chief Medical Officer, the MoH and the Hungarian Central Statistical Office.

Case definition

Confirmed case: a clinically compatible case when the campylobacter infection is laboratory confirmed. Probable case: a clinically compatible case that is not confirmed by laboratory investigation, but it has an epidemiological link to a confirmed campylobacter outbreak.

Diagnostic/analytical methods used

Campylobacter isolates are obtained by culturing the faeces samples of the patients on selectivedifferentiating media, using reduced oxigen tension and special incubation temperature, followed by biochemical tests.

Notification system in place

The laboratories of NPHMOS have been able to identify campylobacters since 1987. Human cases have been notifiable since 1998. The physician reports data of case on a "case report form" by mail to the municipal institute of NPHMOS. The specialist of the institute records data immediately in the electronic system of the NPHMOS. Hungary has also a laboratory based surveillance system, and the NPHMOS has representative dataset from most of the microbiological laboratories about the laboratory investigated cases (since 2003 antibiotic resistances have also been reported from 5 regional laboratories of NPHMOS and from a number of laboratories of universities or hospitals).

The illness is reported first as enteritis infectiosa syndrome on the basis of the symptoms. Having the results of the laboratory tests this syndrome-based diagnose is modified to etiology-based diagnose. In some cases the reporting follows only the available laboratory test results.

History of the disease and/or infection in the country

The laboratories of NPHMOS have been able to identify campylobacters since 1987. In 1990 the National Centre for Epidemiology prepared a guideline on campylobacter enteritis, and then the collection of data on campylobacteriosis was started on this basis. The number of isolates increased from 5 500/year in 1990 to 12 000/year in 1996. Since 1998 this number has varied between 9 500 – 11 500 /year. Human cases have been notifiable since 1998, so the laboratory and clinical surveillance have been linked in this year.

The number of registered cases remained around 8 300-9 200 between 1998 and 2004 (incidence: 81,6 -

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91,0 /100 000 inhabitants/year).

Altogether four death cases were registered between 1998 and 2004 (case fatality rate ranged between 0.0 - 0.02%/year). The highest age-specific incidence was observed among children under five years in all periods, and the incidence has declined with the progressing of the age.

The 95% of cases were sporadic, widespread outbreaks were observed very rarely; outbreaks mostly appeared in families (200 – 300/year). The most of the outbreaks were caused by poultry prepared with inadequate heat treatment or additionally contaminated food. There has not been any evidence in Hungary for outbreaks caused by ready-to-eat foods of industrial origin.

[In 1998 a single outbreak was investigated that occurred among consumers exposed to non-pasteurised milk (cow) consumed on a livestock market and exhibition (51 cases)]

75-80% of isolated strains were C.jejuni, around 10% were C.coli, 4-5% were C.lari, the distribution of campylobacter specieses did not changed significantly during the last five years.

Relevance as zoonotic disease

It is supposed that person-to-person transmission of campylobacter occur only in very few cases (infants, etc). Most of the outbreaks originated from poultry, via contaminated food. However, this facts have not based on statistical or laboratory evidences in Hungary.

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2.2.3 Campylobacter in foodstuffs

A. Thermophilic Campylobacter in Broiler meat and products thereof

Monitoring system

Sampling strategy

At slaughterhouse and cutting plant

There is an annual monitoring program based on the production capacity of the region. The monitoring plan is prepared by the central authority. The samples are taken by the regional authorities. Only one sample unit is taken from a batch, 25 grams are examined in the laboratory. These official samples are examined in the NRL Campylobacter with a presence-absence test followed by species identification and antimicrobial resistance.

At retail

To be reported via ECDC.

Frequency of the sampling

At slaughterhouse and cutting plant

Sampling distributed evenly throughout the year

Type of specimen taken

At slaughterhouse and cutting plant

Fresh meat

Methods of sampling (description of sampling techniques)

At slaughterhouse and cutting plant

At least 500 grams of fresh meat is sampled in a sterile plastic bag. The sample is transported to the laboratory in a cool box by courier.

Definition of positive finding

At slaughterhouse and cutting plant

When a strain of thermophilic Campylobacter is isolated from the sample (25g) after enrichment.

Diagnostic/analytical methods used

At slaughterhouse and cutting plant

Bacteriological method: ISO 10272:1995

National evaluation of the recent situation, the trends and sources of infection

Thermophilic Campylobacter - as in many countries - shows a high prevalence in broiler meat with a marked sesonal disribution of 30 % in winter to more than 60% in the summer months.

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Table Campylobacter in other food

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Sample weight	Units tested	Total units positive for Campylobact er	C. coli	C. jejuni
Meat from pig - fresh - Slaughterhouse		Objective sampling	Official sampling	food sample > meat		Single	25 grams	33	5	4	1
Meat from pig - fresh - Processing plant		Objective sampling	Official sampling	food sample > meat		Single	25 grams	151	3	1	2
Meat from pig - fresh - Retail		Objective sampling	Official sampling	food sample > meat		Single	25 grams	94	2	1	1
Milk, goats' - raw milk for manufacture - intended for manufacture of raw or low heat-treated products - Processing plant		Objective sampling	Official sampling	food sample > milk		Single	25 ml	3	0	0	0
Meat from bovine animals - fresh - Processing plant - Monitoring - active		Objective sampling	Official sampling	food sample > meat		Single	25 grams	51	3	3	0
Meat from bovine animals - fresh - Retail - Monitoring - active		Objective sampling	Official sampling	food sample > meat		Single	25 grams	19	0	0	0
Meat from bovine animals - fresh - Slaughterhouse - Monitoring - active		Selective sampling	Official sampling	food sample > meat		Single	25 grams	19	1	1	0
Milk, cows' - raw milk - Retail		Objective sampling	Official sampling	food sample > milk		Single	25 ml	55	0	0	0
Milk, cows' - raw milk for manufacture - intended for manufacture of raw or low heat-treated products - Monitoring - active		Objective sampling	Official and industry sampling	food sample > milk		Single	25 ml	132	0	0	0

	C. lari	C. upsaliensis	Thermophilic Campylobact er spp., unspecified
Meat from pig - fresh - Slaughterhouse	0	0	0
Meat from pig - fresh - Processing plant	0	0	0

Table Campylobacter in other food

	C. lari	C. upsaliensis	Thermophilic Campylobact er spp., unspecified
Meat from pig - fresh - Retail	0	0	0
Milk, goats' - raw milk for manufacture - intended for manufacture of raw or low heat-treated products - Processing plant	0	0	0
Meat from bovine animals - fresh - Processing plant - Monitoring - active	0	0	0
Meat from bovine animals - fresh - Retail - Monitoring - active	0	0	0
Meat from bovine animals - fresh - Slaughterhouse - Monitoring - active	0	0	0
Milk, cows' - raw milk - Retail	0	0	0
Milk, cows' - raw milk for manufacture - intended for manufacture of raw or low heat-treated products - Monitoring - active	0	0	0

Table Campylobacter in poultry meat

Total units Sample type Sample origin Sampling unit Source of Sampling Sample positive for Units tested C. jejuni Sampler C. coli information strategy weight Campylobact Meat from broilers (Gallus gallus) - fresh -Official Objective food sample 25 grams 243 60 37 19 Single sampling Processing plant sampling > meat Objective Official food sample Meat from broilers (Gallus gallus) - fresh - Retail 66 39 Single 25 grams 280 15 sampling sampling > meat Objective Official food sample Meat from duck - fresh - Retail Single 25 grams 38 4 3 1 sampling sampling > meat Objective Official food sample Meat from geese - fresh - Retail 4 0 0 0 Single 25 grams sampling sampling > meat Official Objective food sample Meat from duck - fresh - Processing plant Single 25 grams 25 10 3 5 sampling sampling > meat Objective Official food sample Meat from geese - fresh - Processing plant 24 6 0 3 Single 25 grams sampling sampling > meat food sample Meat from turkey - fresh - Processing plant -Objective Official 8 7 Single 25 grams 112 22 Monitoring - active sampling sampling > meat Objective Official food sample Meat from turkey - fresh - Retail - Monitoring -77 Single 25 grams 14 8 4

> meat

sampling

sampling

	C. lari	C. upsaliensis	Thermophilic Campylobact er spp., unspecified
Meat from broilers (Gallus gallus) - fresh - Processing plant			4
Meat from broilers (Gallus gallus) - fresh - Retail			12
Meat from duck - fresh - Retail			
Meat from geese - fresh - Retail	0	0	0

active

Table Campylobacter in poultry meat

	C. lari	C. upsaliensis	Thermophilic Campylobact er spp., unspecified
Meat from duck - fresh - Processing plant	0	0	2
Meat from geese - fresh - Processing plant	0	0	3
Meat from turkey - fresh - Processing plant - Monitoring - active	0	0	7
Meat from turkey - fresh - Retail - Monitoring - active	0	0	2

2.2.4 Campylobacter in animals

Table Campylobacter in animals

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Units tested	Total units positive for Campylobact er	C. coli	C. jejuni	C. lari
Pigs - fattening pigs - Slaughterhouse - Monitoring	NFCSO VDD	Unspecified	Not applicable	animal sample	Domestic	Holding	163	73	70	3	0
Gallus gallus (fowl) - broilers - Slaughterhouse - Monitoring	NFCSO VDD	Unspecified	Not applicable	animal sample	Domestic	Flock	159	118	53	65	0

	C. upsaliensis	Thermophilic Campylobact er spp., unspecified
Pigs - fattening pigs - Slaughterhouse - Monitoring	0	0
Gallus gallus (fowl) - broilers - Slaughterhouse - Monitoring	0	0

2.2.5 Antimicrobial resistance in Campylobacter isolates

A. Antimicrobial resistance in Campylobacter jejuni and coli in foodstuff derived from poultry

Sampling strategy used in monitoring

Frequency of the sampling

Isolates derive from monitoring system performed for measurement of prevalence of campylobacters in fresh poultry meat. The sampling is random, performed by the regional competent authorities. The samples are taken in slaughterhouses, and is a part of a permanent monitoring scheme.

Type of specimen taken

500 grams of fresh poultry meat.

Procedures for the selection of isolates for antimicrobial testing

Almost every isolated strains are tested.

Methods used for collecting data

All the tests are performed by the NRL.

Laboratory methodology used for identification of the microbial isolates

Disc diffusion method on horseblood agar plates. Control strains are used.

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Table Antimicrobial susceptibility testing of C. coli in Pigs - fattening pigs - quantitative data [Dilution method]

C. coli												Р	igs - fatt	ening pi	gs											
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory		60																								
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	60	1								1	2	5	42	9			1								
Aminoglycosides - Streptomycin	4	60	49												1	10	3	46								
Fluoroquinolones - Ciprofloxacin	0.5	60	31							2	19	6	2			31										
Tetracyclines - Tetracycline	2	60	56									2	2					56								
Macrolides - Erythromycin	8	60	9										19	20	9	2	1	1	8							

C. coli		Pigs - fa	attening gs
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory	6	0
Antimicro	oials:	lowest	highest
Aminoglycosides	s - Gentamicin	0.12	16
Aminoglycosides	s - Streptomycin	1	16
Fluoroquinolone	s - Ciprofloxacin	0.06	4
Tetracyclines - T	etracycline	0.25	16
Macrolides - Ery	thromycin	0.5	32

Table Antimicrobial susceptibility testing of C. jejuni in Pigs - fattening pigs

- quantitative data [Dilution method]

C. jejuni												Р	igs - fatte	ening pi	gs											
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory		1																								
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	1	0											1												
Aminoglycosides - Streptomycin	4	1	0													1										
Fluoroquinolones - Ciprofloxacin	0.5	1	0										1													
Tetracyclines - Tetracycline	1	1	0											1												
Macrolides - Erythromycin	4	1	0											1												

C. jejuni		Pigs - fa	
	Isolates out of a monitoring program (yes/no)		
	Number of isolates available in the laboratory		1
Antimicrob	oials:	lowest	highest
Aminoglycosides	s - Gentamicin	0.12	16
Aminoglycosides	s - Streptomycin	1	16
Fluoroquinolones	s - Ciprofloxacin	0.06	4
Tetracyclines - T	etracycline	0.25	16
Macrolides - Eryt	thromycin	0.5	32

Table Antimicrobial susceptibility testing of C. coli in Gallus gallus (fowl) - broilers

- quantitative data [Dilution method]

C. coli												Gallus	gallus (fowl) - b	roilers											
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory		51																								
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	51	0								7	1	18	24	1											
Aminoglycosides - Streptomycin	4	51	8											6	10	27	2	6								
Fluoroquinolones - Ciprofloxacin	0.5	51	47							1	2	1				47										
Tetracyclines - Tetracycline	2	51	27									15	6	2	1			27								
Macrolides - Erythromycin	8	51	0										38	9	4											

C. coli			gallus broilers							
	Isolates out of a monitoring program (yes/no)									
	Number of isolates available in the laboratory	5	1							
Antimicrol	bials:	lowest	highest							
Aminoglycosides	s - Gentamicin	0.12	16							
Aminoglycosides	s - Streptomycin	1	16							
Fluoroquinolone	s - Ciprofloxacin	0.06	4							
Tetracyclines - T	Fetracycline	0.25	16							
Macrolides - Ery	Isolates out of a monitoring program (yes/no) Number of isolates available									

Table Antimicrobial susceptibility testing of C. jejuni in Gallus gallus (fowl) - broilers

quantitative data [Dilution method]

C. jejuni												Gallus	gallus ((fowl) - b	oroilers											
Isolates out of a monitoring program (yes/no)																										
Number of isolates available in the laboratory																										
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	56	0								5	7	35	8	1											
Aminoglycosides - Streptomycin	4	56	7											17	22	10	2	5								
Fluoroquinolones - Ciprofloxacin	0.5	56	48							2	4	2				48										
Tetracyclines - Tetracycline	1	56	28									18	9	1				28								
Macrolides - Erythromycin	4	56	0										51	5												

C. jejuni	Isolates out of a monitoring program (yes/no) Number of isolates available in the laboratory imicrobials: oglycosides - Gentamicin oglycosides - Streptomycin oquinolones - Ciprofloxacin										
		5	6								
Antimicrob	Isolates out of a monitoring program (yes/no) Number of isolates available in the laboratory Intimicrobials: Intimicrobial										
Aminoglycosides	program (yes/no) Number of isolates available in the laboratory ntimicrobials: inoglycosides - Gentamicin inoglycosides - Streptomycin proquinolones - Ciprofloxacin										
Aminoglycosides	inoglycosides - Gentamicin inoglycosides - Streptomycin										
Fluoroquinolones	s - Ciprofloxacin	0.06	4								
Tetracyclines - T	Isolates out of a monitoring program (yes/no) Number of isolates available in the laboratory Itimicrobials: noglycosides - Gentamicin noglycosides - Streptomycin roquinolones - Ciprofloxacin acyclines - Tetracycline										
Macrolides - Eryt	Isolates out of a monitoring program (yes/no) Number of isolates available										

Table Antimicrobial susceptibility testing of C. jejuni in Meat from broilers (Gallus gallus) - Objective sampling - Official sampling - food sample - quantitative data [Dilution method]

	Meat from broilers (Gallus gallus) yes																									
C. jejuni											ı	Meat fro	m broile	rs (Gallu	ıs gallus)										
Isolates out of a monitoring program (yes/no)													ye	es												
Number of isolates available in the laboratory														4												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	24	0								9	9	5	1												
Aminoglycosides - Streptomycin	4	24	2											9	6	7	1	1								
Amphenicols - Chloramphenicol	16	24	0											3	11	7	3									
Fluoroquinolones - Ciprofloxacin	0.5	24	18								2	4	0	0	2	6	10									
Quinolones - Nalidixic acid	16	24	19													2	3	0	2	9	8					
Tetracyclines - Tetracycline	1	24	13										5	6	3	0	1	2	6	1						
Fully sensitive		4	4	4																						
Macrolides - Erythromycin	4	24	0										11	9	4											
Resistant to 1 antimicrobial		2	2	2																						
Resistant to 2 antimicrobials		6	6	6																						
Resistant to 3 antimicrobials		10	10	10																						
Resistant to 4 antimicrobials		2	2	2																						

C. jejuni			from (Gallus us)
	Isolates out of a monitoring program (yes/no)	ye	es
	Number of isolates available in the laboratory	2	4
Antimicrobi	ials:	lowest	highest
Aminoglycosides -	Gentamicin		

Table Antimicrobial susceptibility testing of C. jejuni in Meat from broilers (Gallus gallus) - Objective sampling - Official sampling - food sample - quantitative data [Dilution method]

quant	itativo data [Diic	1011	
C. jejuni			from (Gallus lus)
	Isolates out of a monitoring program (yes/no)	уe	es
	Number of isolates available in the laboratory	2	4
Antimicrob	ials:	lowest	highest
Aminoglycosides	- Streptomycin		
Amphenicols - Ch	nloramphenicol		
Fluoroquinolones	- Ciprofloxacin		
Quinolones - Nali	dixic acid		
Tetracyclines - Te	etracycline		
Fully sensitive			
Macrolides - Erytl	hromycin		
Resistant to 1 ant	timicrobial		
Resistant to 2 and	timicrobials		
Resistant to 3 and	timicrobials		
Resistant to 4 ant	timicrobials		

Table Antimicrobial susceptibility testing of C. coli in Meat from broilers (Gallus gallus) - food sample - meat - quantitative data [Dilution method]

						TIOOTII G	ιιιοτί (μ	9,,,,,,	di i i bei	01 10010	VIII	i a com	2011111111	011 01 11		roquar										
C. coli												Meat fro	m broile	rs (Gallu	ıs gallus)										
Isolates out of a monitoring program (yes/no)													y	es												
Number of isolates available in the laboratory													5	6												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	56	2								25	17	8	4			2									-
Aminoglycosides - Streptomycin	4	56	13										1	25	13	4	3	6	1	3						
Amphenicols - Chloramphenicol	16	56	1											8	26	16	3	2	1							
Fluoroquinolones - Ciprofloxacin	0.5	56	48								5	1	2	1	3	12	32									
Quinolones - Nalidixic acid	16	56	47													2	4	3	10	14	23					
Tetracyclines - Tetracycline	2	56	32									1	12	9	2	2	3	5	22							
Fully sensitive		3	3	3																						
Macrolides - Erythromycin	8	56	2									1	27	21	3	2			2							
Resistant to 1 antimicrobial		6	6	6																						
Resistant to 2 antimicrobials		12	12	12																						
Resistant to 3 antimicrobials		29	29	29																						
Resistant to 4 antimicrobials		2	2	2																						
Resistant to >4 antimicrobials		4	4	4																						

Table Antimicrobial susceptibility testing of C. coli in Meat from broilers (Gallus gallus) - food sample - meat - quantitative data [Dilution method]

C. coli	-		from (Gallus lus)
	Isolates out of a monitoring program (yes/no)	ye	es
	Number of isolates available in the laboratory	5	6
Antimicrob	oials:	lowest	highest
Aminoglycosides	- Gentamicin		
Aminoglycosides	- Streptomycin		
Amphenicols - C	hloramphenicol		
Fluoroquinolones	s - Ciprofloxacin		
Quinolones - Nal	idixic acid		
Tetracyclines - T	etracycline		
Fully sensitive			
Macrolides - Eryt	hromycin		
Resistant to 1 an	timicrobial		
Resistant to 2 an	timicrobials		
Resistant to 3 an	timicrobials		
Resistant to 4 an	timicrobials		
Resistant to >4 a	ntimicrobials		

Table Antimicrobial susceptibility testing of C. jejuni in Meat from turkey - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

C. jejuni							ų.	<u>. </u>			NOS WILL		Meat fro			·										
Isolates out of a monitoring program (yes/no)													ye	es												
Number of isolates available in the laboratory													1	0												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	10	0								5	3	1	1												
Aminoglycosides - Streptomycin	4	10	2										1	4	3	0	1	1								
Amphenicols - Chloramphenicol	16	10	0												3	4	1	2								
Fluoroquinolones - Ciprofloxacin	0.5	10	8									1	1	1		3	4									
Quinolones - Nalidixic acid	16	10	7													1	2	0	1	2	4					
Tetracyclines - Tetracycline	1	10	5										2	3	1	3			1							
Fully sensitive		1	1	1																						
Macrolides - Erythromycin	4	10	0										6	3	1											
Resistant to 1 antimicrobial		2	2	2																						
Resistant to 2 antimicrobials		2	2	2																						
Resistant to 3 antimicrobials		4	4	4																						
Resistant to 4 antimicrobials		1	1	1																						

C. jejuni		Meat turl	from
Isolates out of a monitoring program (yes/no)		ye	es
Number of isolates available in the laboratory	9	1	0
Antimicrobials:		lowest	highest
Aminoglycosides - Gentamicin		·	

Table Antimicrobial susceptibility testing of C. jejuni in Meat from turkey - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

900	ativo data [Bilat		
C. jejuni			from key
	Isolates out of a monitoring program (yes/no)	yı	es
	Number of isolates available in the laboratory	1	0
Antimicrob	oials:	lowest	highest
Aminoglycosides	s - Streptomycin		
Amphenicols - C	hloramphenicol		
Fluoroquinolones	s - Ciprofloxacin		
Quinolones - Nal	idixic acid		
Tetracyclines - T	etracycline		
Fully sensitive			
Macrolides - Eryt	thromycin		
Resistant to 1 an	ıtimicrobial		
Resistant to 2 an	itimicrobials		
Resistant to 3 an	timicrobials		
Resistant to 4 an	timicrobials		

Table Antimicrobial susceptibility testing of C. coli in Meat from turkey - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

							'' '	, ,.																		
C. coli													Meat fro	m turkey	/											
Isolates out of a monitoring program (yes/no)													y	es												
Number of isolates available in the laboratory													1	11												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	11	0								5	3	2	1												
Aminoglycosides - Streptomycin	4	11	3											4	2	2	1	1	1							
Amphenicols - Chloramphenicol	16	11	0											1	4	3	3									
Fluoroquinolones - Ciprofloxacin	0.5	11	8								2		1			2	6									
Quinolones - Nalidixic acid	16	11	6													2		3		2	4					
Tetracyclines - Tetracycline	2	11	5										2	2	2	1	0	0	3	0	1					
Fully sensitive		3	3	3																						
Macrolides - Erythromycin	8	11	1										2	5	3				1							
Resistant to 2 antimicrobials		2	2	2																						
Resistant to 3 antimicrobials		5	5	5																						
Resistant to 4 antimicrobials		1	1	1														_								

C. coli		Meat turl	
	Isolates out of a monitoring program (yes/no)	ye	es
	Number of isolates available in the laboratory	1	1
Antimicrob	oials:	lowest	highest
Aminoglycosides	- Gentamicin		
Aminoglycosides	- Streptomycin	·	

Table Antimicrobial susceptibility testing of C. coli in Meat from turkey - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

C. coli	Meat from turkey						
	Isolates out of a monitoring program (yes/no)	y	es				
	Number of isolates available in the laboratory	1	1				
Antimicrol	oials:	lowest	highest				
Amphenicols - C	hloramphenicol						
Fluoroquinolone	s - Ciprofloxacin						
Quinolones - Na	lidixic acid						
Tetracyclines - T	etracycline						
Fully sensitive							
Macrolides - Ery	thromycin						
Resistant to 2 ar	ntimicrobials						
Resistant to 3 ar							
Resistant to 4 ar							

<u>Table Antimicrobial susceptibility testing of C. jejuni in Meat from pig - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]</u>

Concentration (µg	/ml), number of	isolates with a	concentration of	of inhibition ed	ual to
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C. jejuni													Meat f	rom pig													ᆵ
Isolates out of a monitoring program (yes/no)													у	es													Hungary
Number of isolates available in the laboratory														3													07
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	2
Aminoglycosides - Gentamicin	2	3	0									2	1														Report
Aminoglycosides - Streptomycin	4	3	1												1	1				1							on
Amphenicols - Chloramphenicol	16	3	0												2		1										rends
Fluoroquinolones - Ciprofloxacin	0.5	3	1								1	1					1										l ds
Quinolones - Nalidixic acid	16	3	1														1	1		1							ands
Tetracyclines - Tetracycline	1	3	3													1		1	1								ources
Macrolides - Erythromycin	4	3	1											1		1			1								es of
Resistant to 1 antimicrobial		1	1	1																							
Resistant to 2 antimicrobials		1	1	1																							Zoonoses
Resistant to 4 antimicrobials		1	1	1) ř.

C. jejuni		Meat fr	om pig			
	yes					
	Number of isolates available in the laboratory	;	3			
Antimicrob	oials:	lowest	highest			
Aminoglycosides	- Gentamicin					
Aminoglycosides	- Streptomycin					
Amphenicols - C	·					

Table Antimicrobial susceptibility testing of C. jejuni in Meat from pig - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

C. jejun	Meat from pig						
	yes						
	Number of isolates available in the laboratory	:	3				
Antimicro	bials:	lowest	highest				
Fluoroquinolone	es - Ciprofloxacin						
Quinolones - Na	alidixic acid						
Tetracyclines -	Tetracycline						
Macrolides - Er	ythromycin						
Resistant to 1 a	ıntimicrobial						
Resistant to 2 a							
Resistant to 4 a							

Concentration (µg/m), number of isolates v	with a concentration of	of inhibition equal to
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								, ,																		
C. coli		Meat from pig																								
Isolates out of a monitoring program (yes/no)													ye	es												
Number of isolates available in the laboratory														6												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	6	0								1	1	2	2												
Aminoglycosides - Streptomycin	4	6	4											1		1			1	3						
Amphenicols - Chloramphenicol	16	6	0											1	4	1										
Fluoroquinolones - Ciprofloxacin	0.5	6	4								1	1			1	1	2									
Quinolones - Nalidixic acid	16	6	4														1	1	1	1	2					
Tetracyclines - Tetracycline	2	6	5											1				1	4							
Macrolides - Erythromycin	8	6	1										2	2	1			1								
Resistant to 1 antimicrobial		1	1	1																						
Resistant to 2 antimicrobials		2	2	2																						
Resistant to 4 antimicrobials		2	2	2																						
Resistant to >4 antimicrobials		1	1	1																						

C. coli	C. coli								
	Isolates out of a monitoring program (yes/no)	ye	es						
	Number of isolates available in the laboratory	•	6						
Antimicrob	oials:	lowest	highest						
Aminoglycosides	- Gentamicin								
Aminoglycosides	- Streptomycin								

Table Antimicrobial susceptibility testing of C. coli in Meat from pig - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

C. coli	Meat fi	om pig									
	Isolates out of a monitoring program (yes/no)										
	Number of isolates available in the laboratory		6								
Antimicrob	oials:	lowest	highest								
Amphenicols - C	hloramphenicol										
Fluoroquinolones	s - Ciprofloxacin										
Quinolones - Nal	idixic acid										
Tetracyclines - T	etracycline										
Macrolides - Eryt	thromycin										
Resistant to 1 an	timicrobial										
Resistant to 2 an	itimicrobials										
Resistant to 4 an											
Resistant to >4 a	ntimicrobials										

Table Antimicrobial susceptibility testing of C. jejuni in Meat from other poultry species - Objective sampling - Official sampling - food sample meat - quantitative data [Dilution method]

C. jejuni							·					Meat fro	om other	r poultry	species	.										
Isolates out of a monitoring program (yes/no)													y	es												
Number of isolates available in the laboratory														6												
Antimicrobials:	Cut-off value	Z	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	6	0								1	3	1	1												
Aminoglycosides - Streptomycin	4	6	2											2		2				2						
Amphenicols - Chloramphenicol	16	6	0											1	2	3										
Fluoroquinolones - Ciprofloxacin	0.5	6	4									2				2	2									
Quinolones - Nalidixic acid	16	6	5														1		2		3					
Trimethoprim		6	6											1			1	1	3							
Macrolides - Erythromycin	4	6	0										3	1	2											
Resistant to 2 antimicrobials		2	2	2																						
Resistant to 3 antimicrobials		2	2	2																						
Resistant to 4 antimicrobials		2	2	2																						

C. jejuni			om other species
	Isolates out of a monitoring program (yes/no)	ye	es
	Number of isolates available in the laboratory	(6
Antimicrob	oials:	lowest	highest
Aminoglycosides	- Gentamicin		
Aminoglycosides	- Streptomycin		
Amphenicols - Ch			

Table Antimicrobial susceptibility testing of C. jejuni in Meat from other poultry species - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

C. jejuni	i		m other species	
	Isolates out of a monitoring program (yes/no)	y	es	
	Number of isolates available in the laboratory			
Antimicrol	lowest	highest		
Fluoroquinolone				
Quinolones - Na	lidixic acid			
Trimethoprim				
Macrolides - Ery	thromycin			
Resistant to 2 ar	ntimicrobials			
Resistant to 3 ar				
Resistant to 4 ar	ntimicrobials			

Table Antimicrobial susceptibility testing of C. coli in Meat from other poultry species - Objective sampling - Official sampling - food sample meat - quantitative data [Dilution method]

Concentration (µg/ml)	number of isolates with a concentration of	of inhibition equal to

C. coli		Meat from other poultry species																								
Isolates out of a monitoring program (yes/no)													ye	es												
Number of isolates available in the laboratory													4	4												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	4	0								2	2														
Aminoglycosides - Streptomycin	4	4	0											2	2											
Amphenicols - Chloramphenicol	16	4	0											2	1	1										
Fluoroquinolones - Ciprofloxacin	0.5	4	3									1				1	2									
Quinolones - Nalidixic acid	16	4	3													1				2	1					
Tetracyclines - Tetracycline	2	4	2										1	1				1	1							
Fully sensitive		1	1	1																						
Macrolides - Erythromycin	8	4	0											4												
Resistant to 2 antimicrobials		1	1	1																						
Resistant to 3 antimicrobials		2	2	2																						

C. coli	Meat from other poultry species			
	Isolates out of a monitoring program (yes/no)			
	Number of isolates available in the laboratory	4		
Antimicrob	lowest	highest		
Aminoglycosides	s - Gentamicin			
Aminoglycosides				
Amphenicols - C				

Table Antimicrobial susceptibility testing of C. coli in Meat from other poultry species - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

	•			
C. coli	Meat from other poultry species			
	Isolates out of a monitoring program (yes/no)	yes		
	Number of isolates available in the laboratory	,	1	
Antimicrol	lowest	highest		
Fluoroquinolone				
Quinolones - Na	lidixic acid			
Tetracyclines - T	etracycline			
Fully sensitive				
Macrolides - Ery	thromycin			
Resistant to 2 ar				
Resistant to 3 ar	ntimicrobials			

Table Cut-off values used for antimicrobial susceptibility testing of C. coli in Animals

Test Method Used	Standard methods used for testing

			Concentration (microg/ml)	Zone diameter (mm)
		Standard	Resistant >	Resistant <=
Aminoglycosides	Gentamicin		2	
	Streptomycin		4	
Fluoroquinolones	Ciprofloxacin		0.5	
Macrolides	Erythromycin		8	
Quinolones	Nalidixic acid		16	
Tetracyclines	Tetracycline		2	

Table Cut-off values used for antimicrobial susceptibility testing of C. coli in Feed

Test Method Used	Standard methods used for testing	

			Concentration (microg/ml)	Zone diameter (mm)
		Standard	Resistant >	Resistant <=
Aminoglycosides	Gentamicin		2	
	Streptomycin		4	
Fluoroquinolones	Ciprofloxacin		0.5	
Macrolides	Erythromycin		8	
Quinolones	Nalidixic acid		16	
Tetracyclines	Tetracycline		2	

Table Cut-off values used for antimicrobial susceptibility testing of C. coli in Food

Test Method Used	Standard methods used for testing
Broth dilution	EFSA

			Concentration (microg/ml)	Zone diameter (mm)
		Standard	Resistant >	Resistant <=
Aminoglycosides	Gentamicin	EFSA	2	
	Streptomycin	EFSA	4	
Fluoroquinolones	Ciprofloxacin	EFSA	0.5	
Macrolides	Erythromycin	EFSA	8	
Quinolones	Nalidixic acid	EFSA	16	
Tetracyclines	Tetracycline	EFSA	2	

Table Cut-off values used for antimicrobial susceptibility testing of C. jejuni in Animals

Test Method Used	Standard methods used for testing

			Concentration (microg/ml)	Zone diameter (mm)
		Standard	Resistant >	Resistant <=
Aminoglycosides	Gentamicin		2	
	Streptomycin		4	
Fluoroquinolones	Ciprofloxacin		0.5	
Macrolides	Erythromycin		4	
Quinolones	Nalidixic acid		16	
Tetracyclines	Tetracycline		1	

Table Cut-off values used for antimicrobial susceptibility testing of C. jejuni in Feed

Test Method Used	Standard methods used for testing

			Concentration (microg/ml)	Zone diameter (mm)
		Standard	Resistant >	Resistant <=
Aminoglycosides	Gentamicin		2	
	Streptomycin		4	
Fluoroquinolones	Ciprofloxacin		0.5	
Macrolides	Erythromycin		4	
Quinolones	Nalidixic acid		16	
Tetracyclines	Tetracycline		1	

Table Cut-off values used for antimicrobial susceptibility testing of C. jejuni in Food

Test Method Used	Standard methods used for testing
Broth dilution	EFSA

			Concentration (microg/ml)	Zone diameter (mm)
		Standard	Resistant >	Resistant <=
Aminoglycosides	Gentamicin	EFSA	2	
	Streptomycin	EFSA	4	
Fluoroquinolones	Ciprofloxacin	EFSA	0.5	
Macrolides	Erythromycin	EFSA	4	
Quinolones	Nalidixic acid	EFSA	16	
Tetracyclines	Tetracycline	EFSA	1	

2.3 LISTERIOSIS

2.3.1 General evaluation of the national situation

A. Listeriosis general evaluation

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

Testing of ready-to-eat products for the presence/and/or the determination of the number of Listeria monocytogenes is obligatory for food business operators based on Reg.2073/2005/EC. The official monitoring program concentrates to take samples from these products on a risk based approach as well. Only the data of official control are presented in this report, because only these data are collected in the database of the authority. The legislative background has changed a lot, because before 2006 only milk and milk products were regularly tested for Listeria monocytogenes and only by presence absence tests. In the frame of USDA-FSIS monitoring obligatory for US exporting establishments raw cured products were tested as well with presence-abscence tests and MPN based method suitable for enumeration of low numbers of the microorganism

From 2006, those RTE products that not support the growth of Listeria, are examined by the enumeration method ISO 11290:2 (e.g.salami, raw smoked ham). If the product is able to support the growth of the pathogen, presence-abscence test is used as a first step (ISO 11290:1), or the two method run paralel (depending on the expiry date, the amount of sample is enough to perform an enumeration test if the first test is positive). The pathogen is enumerated from all the positive samples.

Based on the past decade's USDA Listeria monitoring data, Listeria monocytogenes can be frequently isolated from traditional raw and smoked meat products as salami and sausages, but the highest contamination level was 2.3 cells (MPN method)/gram. Therefore this product group certainly does not play an important role in human infections.

Listeria monocytogenes can be isolated from mixes salads as well, but because of low pH and preservatives charateristic for this product group generally do not support the growth of the pathogen, and only level of <10 cells per gram was measured from the positive samples.

Milk products are characteristically made of pasteurised milk in Hungary, therefore these types of foodstuff are practically free from Listeria.

Consumers show an increasing interest to by raw milk for consumption in the past few years. Despite of the obligatory labelling to call the consumers' attention for heat treating of raw milk, this product can be considered as a potential source of infection in the future.

Recent actions taken to control the zoonoses

Based on Reg. 2073/2005/EC.

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2.3.2 Listeriosis in humans

A. Listeriosis in humans

Reporting system in place for the human cases

There are around 80 communicable diseases notifiable in Hungary based on legal background. The physician (in primary health care, specialist care, inpatient medical institution or pathology) who first diagnoses a case of a notifiable communicable disease (even the suspicion of the disease!) immediately reports data of case to the first level of the epidemiological network (municipal institute) of National Public Health and Medical Officer's Service (NPHMOS). Data must be reported both at the beginning and at end of the illness (recovery/death, result of laboratory test). The NPHMOS has a nationwide electronic reporting system for registering and analysing data of communicable diseases in a combined national database, so the system provides online connection amid the three levels (municipal, regional and national level – National Centre of Epidemiology - NCE) of the organization. The NCE prepares reports regularly (weekly, monthly, yearly) to the Chief Medical Officer, the MoH and the Hungarian Central Statistical Office.

Case definition

Confirmed case: Clinical picture of an invasive illness (meningitis purulenta, sepsis, stillbirth etc.), and L.monocytogenes has been isolated from invasive sample (liquor, blood, amniotic fluid etc.)

Diagnostic/analytical methods used

The samples are cultivated on enriched medium. The isolation is followed by the biochemical tests, and antimicrobial susceptibility testing.

Notification system in place

Listeriosis has been notifiable since 1998 in Hungary. The physician reports data of case on a "case report form" by mail to the municipal institute of NPHMOS. The specialist of the institute records data immediately in the electronic system of the NPHMOS. Hungary also has a laboratory based surveillance system, and the NPHMOS has representative dataset from most of the microbiological laboratories about the investigated cases (since 2003 antibiotic resistances has also been reported from 20 county institutes and 12 laboratories from universities or hospitals).

The illness is reported first as meningitis purulenta syndrome on the basis of the symptoms. Having the results of the laboratory tests this syndrome-based diagnose is modified to etiology-based diagnose (listeriosis).

History of the disease and/or infection in the country

Listeriosis has been notifiable since 1998 in Hungary, there have been 91 cases registered since then. The number of yearly registered cases ranged between 4-25 (incidence $0.04-0.2/100\ 000$ inhabitants/year; median: 14 cases), the case fatality rate ranged between 0-50% (median 22,2%). The age-distribution of cases: 12% infants, 1-14 year 3,4%, 15-19 year 0%, 20-49 year 20%, 50-59 year 20%, >60 year 43%. Most of the cases are meningitis, less of them are sepsis.

Relevance as zoonotic disease

Listeriosis is underreported in Hungary. No evidence has been found for a food-borne case based on laboratory tests in Hungary.

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2.3.3 Listeria in foodstuffs

A. Listeria in Food

```
Monitoring system
Sampling strategy
monitoring, objective sampling
Type of specimen taken
At the production plant
RTE
At retail
RTE
```

Methods of sampling (description of sampling techniques)

At the production plant

At retail

single sample

single sample

Definition of positive finding

At the production plant

Listeria monocytogenes is isolated

At retail

Listeria monocytogenes is isolated

Diagnostic/analytical methods used

At the production plant

ISO 11290-1, ISO 11290-2

At retail

ISO 11290-1, ISO 11290-2

Total units Listeria Units tested Sample type Sample origin Sampling unit positive for L monocytogen with detection Source of Sampling Sample monocytogen Sampler Units tested es presence information strategy weight method es in x q Milk. cows' - raw milk - intended for direct human Objective Official NFCSO 0 food sample Domestic Single 25 ml 185 0 185 consumption - Farm - Surveillance sampling sampling Official Objective Milk, cows' - pasteurised milk - Processing plant -NFCSO food sample Domestic 25 ml 150 0 150 0 Single Surveillance sampling sampling Milk, cows' - pasteurised milk - Retail - Surveillance Objective Official NFCSO Domestic 20 0 20 0 food sample Single 25 ml sampling sampling Official Milk, goats' - raw milk - intended for direct human Objective **NFCSO** food sample Domestic Single 25 ml 1 1 0 consumption - Farm - Surveillance sampling sampling Cheeses made from cows' milk - soft and semi-soft -Official Objective made from pasteurised milk - Processing plant -**NFCSO** food sample Domestic Sinale 25 a 43 0 43 0 sampling sampling Surveillance Cheeses made from cows' milk - soft and semi-soft -Official Objective NFCSO 2 made from pasteurised milk - Retail - Surveillance 25 g 92 2 90 food sample Unknown Single sampling sampling Cheeses made from cows' milk - hard - made from Objective Official **NFCSO** pasteurised milk - Processing plant - Surveillance food sample Domestic Single 25 g 4 0 4 0 sampling sampling Cheeses made from cows' milk - hard - made from Objective Official NFCSO 0 food sample Unknown 25 g 11 0 11 Single pasteurised milk - Retail - Surveillance sampling sampling Cheeses made from sheep's milk - soft and semi-Objective Official soft - made from pasteurised milk - Processing plant **NFCSO** food sample Domestic Single 25 g 2 0 2 0 sampling sampling - Surveillance Cheeses made from sheep's milk - soft and semi-Official Objective **NFCSO** soft - made from pasteurised milk - Retail -Domestic 25 g 2 0 2 0 food sample Single sampling sampling

Surveillance

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Sample weight		Total units positive for L. monocytogen es	Units tested with detection method	Listeria monocytogen es presence in x g
Dairy products (excluding cheeses) - butter - made from pasteurised milk - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	30	0	30	0
Dairy products (excluding cheeses) - butter - made from pasteurised milk - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Unknown	Single	25 g	56	0	56	0
Dairy products (excluding cheeses) - cream - made from pasteurised milk - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	4	0	4	0
Dairy products (excluding cheeses) - cream - made from pasteurised milk - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Unknown	Single	25 g	6	0	6	0
Cheeses made from cows' milk - curd - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	37	0	37	0
Cheeses made from cows' milk - curd - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Unknown	Single	25 g	38	1	37	1
Cheeses made from cows' milk - unspecified - made from pasteurised milk - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample > carcase swabs	Domestic	Single	25 g	7	0	7	0
Cheeses made from cows' milk - unspecified - made from pasteurised milk - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Unknown	Single	25 g	5	0	5	0
Cheeses made from goats' milk - unspecified - made from pasteurised milk - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	4	0	4	0
Cheeses made from sheep's milk - fresh - made from pasteurised milk - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	8	0	8	0

Dairy products (excluding cheeses) - milk powder

and whey powder - Processing plant - Surveillance

Dairy products (excluding cheeses) - milk powder

and whey powder - Retail - Surveillance

Total units Listeria Units tested positive for L Imonocytogen Sample type Sample origin Sampling unit Source of Sampling Sample with detection Units tested monocytogen Sampler es presence information strategy weight method es in x g Cheeses made from sheep's milk - fresh - made Objective Official NFCSO 25 g 62 2 60 2 food sample Unknown Single sampling from pasteurised milk - Retail - Surveillance sampling Dairy products (excluding cheeses) - dairy desserts -Objective Official chilled - Processing plant - Surveillance **NFCSO** 23 23 food sample Domestic Single 25 g 0 0 sampling sampling Objective Official Dairy products (excluding cheeses) - dairy desserts -**NFCSO** 25 g 87 0 87 0 food sample Single chilled - Retail - Surveillance sampling sampling Objective Official Dairy products (excluding cheeses) - fermented **NFCSO** food sample Domestic Single 25 g 105 0 105 0 dairy products - Processing plant - Surveillance sampling sampling Dairy products (excluding cheeses) - fermented Objective Official **NFCSO** food sample Single 25 g 104 0 104 0 dairy products - Retail - Surveillance sampling sampling Dairy products (excluding cheeses) - ice-cream -Objective Official made from pasteurised milk - Catering - Surveillance **NFCSO** 25 g 36 0 36 0 food sample Domestic Sinale sampling sampling Dairy products (excluding cheeses) - ice-cream -Objective Official made from pasteurised milk - Processing plant -NFCSO Domestic 20 0 20 0 food sample Single 25 g sampling sampling Surveillance Dairy products (excluding cheeses) - ice-cream -Objective Official made from pasteurised milk - Retail - Surveillance **NFCSO** food sample Single 25 g 233 232 1 sampling sampling

food sample

food sample

Domestic

Single

Single

25 g

25 g

15

49

0

0

15

49

0

0

Objective

sampling

Objective

sampling

NFCSO

NFCSO

Official

sampling

Official

sampling

	Units tested with enumeration method	> detection limit but <= 100 cfu/g	L. monocytogen es > 100 cfu/g
Milk, cows' - raw milk - intended for direct human consumption - Farm - Surveillance	0	0	0
Milk, cows' - pasteurised milk - Processing plant - Surveillance	0	0	0
Milk, cows' - pasteurised milk - Retail - Surveillance	0	0	0
Milk, goats' - raw milk - intended for direct human consumption - Farm - Surveillance	0	0	0
Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Surveillance	0	0	0
Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Retail - Surveillance	2	2	0
Cheeses made from cows' milk - hard - made from pasteurised milk - Processing plant - Surveillance	0	0	0
Cheeses made from cows' milk - hard - made from pasteurised milk - Retail - Surveillance	0	0	0
Cheeses made from sheep's milk - soft and semi- soft - made from pasteurised milk - Processing plant - Surveillance	0	0	0
Cheeses made from sheep's milk - soft and semi- soft - made from pasteurised milk - Retail - Surveillance	0	0	0

	Units tested with enumeration method	> detection limit but <= 100 cfu/g	L. monocytogen es > 100 cfu/g
Dairy products (excluding cheeses) - butter - made from pasteurised milk - Processing plant - Surveillance	0	0	0
Dairy products (excluding cheeses) - butter - made from pasteurised milk - Retail - Surveillance	0	0	0
Dairy products (excluding cheeses) - cream - made from pasteurised milk - Processing plant - Surveillance	0	0	0
Dairy products (excluding cheeses) - cream - made from pasteurised milk - Retail - Surveillance	0	0	0
Cheeses made from cows' milk - curd - Processing plant - Surveillance	0	0	0
Cheeses made from cows' milk - curd - Retail - Surveillance	1	1	0
Cheeses made from cows' milk - unspecified - made from pasteurised milk - Processing plant - Surveillance	0	0	0
Cheeses made from cows' milk - unspecified - made from pasteurised milk - Retail - Surveillance	0	0	0
Cheeses made from goats' milk - unspecified - made from pasteurised milk - Processing plant - Surveillance	0	0	0
Cheeses made from sheep's milk - fresh - made from pasteurised milk - Processing plant - Surveillance	0	0	0

	Units tested with enumeration method	> detection limit but <= 100 cfu/g	L. monocytogen es > 100 cfu/g
Cheeses made from sheep's milk - fresh - made from pasteurised milk - Retail - Surveillance	2	1	1
Dairy products (excluding cheeses) - dairy desserts - chilled - Processing plant - Surveillance	0	0	0
Dairy products (excluding cheeses) - dairy desserts - chilled - Retail - Surveillance	0	0	0
Dairy products (excluding cheeses) - fermented dairy products - Processing plant - Surveillance	0	0	0
Dairy products (excluding cheeses) - fermented dairy products - Retail - Surveillance	0	0	0
Dairy products (excluding cheeses) - ice-cream - made from pasteurised milk - Catering - Surveillance	0	0	0
Dairy products (excluding cheeses) - ice-cream - made from pasteurised milk - Processing plant - Surveillance	0	0	0
Dairy products (excluding cheeses) - ice-cream - made from pasteurised milk - Retail - Surveillance	1	1	0
Dairy products (excluding cheeses) - milk powder and whey powder - Processing plant - Surveillance	0	0	0
Dairy products (excluding cheeses) - milk powder and whey powder - Retail - Surveillance	0	0	0

Total units Listeria Units tested positive for L monocytogen Sample origin Sampling unit with detection Source of Sampling Sample monocytogen Units tested Sampler es presence information strategy weight method es in x q Meat from broilers (Gallus gallus) - meat products -Official Objective food sample cooked, ready-to-eat - Processing plant -**NFCSO** Domestic Sinale 25 a 79 78 1 sampling sampling > meat Surveillance Meat from broilers (Gallus gallus) - meat products -Objective Official cooked, ready-to-eat - Retail - Surveillance NFCSO 175 food sample Unknown 25 g 176 1 Single sampling sampling Official Meat from pig - meat products - cooked, ready-to-Objective **NFCSO** food sample Domestic Single 25 g 39 38 eat - Processing plant - Surveillance sampling sampling Objective Official Meat from pig - meat products - cooked, ready-to-NECSO food sample Unknown Single 25 g 102 101 1 eat - Retail - Surveillance sampling sampling Official Meat from bovine animals - meat products - cooked. Objective **NFCSO** food sample Unknown Sinale 25 a 32 0 32 0 ready-to-eat - Retail - Surveillance sampling sampling Official Objective **NFCSO** 9 50 9 Fish - smoked - Retail - Surveillance food sample Unknown Single 25 g 59 sampling sampling Official Crustaceans - unspecified - cooked - Retail -Objective **NFCSO** food sample Sinale 25 g 20 0 20 0 Surveillance sampling sampling Official Molluscan shellfish - cooked - Retail - Surveillance Objective **NFCSO** food sample Single 25 g 60 0 60 0 sampling sampling Objective Official **NFCSO** 25 g 0 0 Infant formula - Retail - Surveillance food sample Single 201 201 sampling sampling Official Other processed food products and prepared dishes Objective **NFCSO** 234 3 231 3 food sample Sinale 25 g - sandwiches - Retail - Surveillance sampling sampling Objective Official Vegetables - pre-cut - ready-to-eat - Retail -NFCSO food sample Single 25 g 167 30 137 30 Surveillance sampling sampling Official Objective Bakery products - cakes - Catering - Surveillance **NFCSO** food sample Sinale 25 a 168 0 168 0

sampling

sampling

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Sample weight	Units tested	Total units positive for L. monocytogen es	Units tested with detection method	Listeria monocytogen es presence in x g
Bakery products - cakes - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	145	1	144	1
Cereals and meals - flakes - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	10	0	10	0
Cereals and meals - flakes - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	55	0	55	0
Chocolate - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample > carcase swabs	Domestic	Single	25 g	28	0	28	0
Chocolate - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	119	0	119	0
Cocoa and cocoa preparations, coffee and tea - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	101	0	101	0
Foodstuffs intended for special nutritional uses - ready-to-eat - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	25	0	25	0
Fruits - non-pre-cut - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	42	0	42	0
Meat from pig - meat products - fermented sausages - in total - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	803	27	776	27
Meat from pig - meat products - raw ham - in total - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	197	4	193	4
Meat from turkey - meat products - cooked, ready-to -eat - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	49	1	48	1
Meat from turkey - meat products - cooked, ready-to -eat - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	175	0	175	0
Nuts and nut products - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	67	0	67	0
Other processed food products and prepared dishes - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	57	1	56	1

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Sample weight		Total units positive for L. monocytogen es	Units tested with detection method	imonocytogeni
Other processed food products and prepared dishes - meat based dishes - Catering - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Domestic	Single	25 g	205	1	204	1
Other processed food products and prepared dishes - meat based dishes - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	178	3	175	3
Other processed food products and prepared dishes - muesli - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	63	0	63	0
Ready-to-eat salads - in total - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	399	3	396	3
Seeds, sprouted - ready-to-eat - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	74	0	74	0
Vegetables - non-pre-cut - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	25 g	24	0	24	0

	Units tested with enumeration method	> detection limit but <= 100 cfu/g	L. monocytogen es > 100 cfu/g
Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Processing plant - Surveillance	1	1	0
Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Retail - Surveillance	1	0	1
Meat from pig - meat products - cooked, ready-to- eat - Processing plant - Surveillance	1	1	0

	Units tested with enumeration method	> detection limit but <= 100 cfu/g	L. monocytogen es > 100 cfu/g
Meat from pig - meat products - cooked, ready-to- eat - Retail - Surveillance	1	1	0
Meat from bovine animals - meat products - cooked, ready-to-eat - Retail - Surveillance	0	0	0
Fish - smoked - Retail - Surveillance	9	8	1
Crustaceans - unspecified - cooked - Retail - Surveillance	0	0	0
Molluscan shellfish - cooked - Retail - Surveillance	0	0	0
Infant formula - Retail - Surveillance	0	0	0
Other processed food products and prepared dishes - sandwiches - Retail - Surveillance	3	3	0
Vegetables - pre-cut - ready-to-eat - Retail - Surveillance	30	28	2
Bakery products - cakes - Catering - Surveillance	0	0	0
Bakery products - cakes - Retail - Surveillance	1	1	0
Cereals and meals - flakes - Processing plant - Surveillance	0	0	0
Cereals and meals - flakes - Retail - Surveillance	0	0	0
Chocolate - Processing plant - Surveillance	0	0	0
Chocolate - Retail - Surveillance	0	0	0
Cocoa and cocoa preparations, coffee and tea - Retail - Surveillance	0	0	0

	Units tested with enumeration method	> detection limit but <= 100 cfu/g	L. monocytogen es > 100 cfu/g
Foodstuffs intended for special nutritional uses - ready-to-eat - Retail - Surveillance	0	0	0
Fruits - non-pre-cut - Retail - Surveillance	0	0	0
Meat from pig - meat products - fermented sausages - in total - Surveillance	27	27	0
Meat from pig - meat products - raw ham - in total - Surveillance	4	3	1
Meat from turkey - meat products - cooked, ready-to -eat - Processing plant - Surveillance	1	1	0
Meat from turkey - meat products - cooked, ready-to -eat - Retail - Surveillance	0	0	0
Nuts and nut products - Retail - Surveillance	0	0	0
Other processed food products and prepared dishes - Retail - Surveillance	1	1	0
Other processed food products and prepared dishes - meat based dishes - Catering - Surveillance	1	1	0
Other processed food products and prepared dishes - meat based dishes - Retail - Surveillance	3	2	1
Other processed food products and prepared dishes - muesli - Retail - Surveillance	0	0	0
Ready-to-eat salads - in total - Surveillance	3	3	0
Seeds, sprouted - ready-to-eat - Retail - Surveillance	0	0	0

	Units tested with enumeration method	> detection limit but <= 100 cfu/g	L. monocytogen es > 100 cfu/g
Vegetables - non-pre-cut - Retail - Surveillance	0	0	0

Comments:

2.3.4 Listeria in animals

Table Listeria in animals

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Units tested	Total units positive for Listeria	L. monocytogen es	Listeria spp., unspecified
Sheep - Farm - Clinical investigations	NFCSO VDD	Suspect sampling	Not applicable	animal sample	Domestic	Animal	13	13	0	13

2.4 E. COLI INFECTIONS

2.4.1 General evaluation of the national situation

A. Verotoxigenic Escherichia coli infections general evaluation

Additional information

E. coli- microbiological examination of food according to ISO 16654 (E. coli O157) identification by antisera

2.4.2 Escherichia coli, pathogenic in foodstuffs

Table VT E. coli in food

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Analytical Method	Sampling unit	Sample weight	Units tested	Total units positive for Verotoxigenic E. coli (VTEC)	Verotoxigenic E. coli (VTEC) - VTEC O157
Meat from bovine animals - fresh - Processing plant - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Unknown	ISO 16654:2001	Single	25 g	74	0	0
Meat from bovine animals - fresh - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample > meat	Unknown	ISO 16654:2001	Single	25 g	6	0	0
Milk, cows' - raw milk - intended for direct human consumption - Farm - Surveillance	NFCSO	Objective sampling	Official sampling	food sample > milk	Domestic	ISO 16654:2001	Single	25 ml	107	1	1
Milk, goats' - raw milk - intended for direct human consumption - Farm - Surveillance	NFCSO	Unspecified	Official sampling	food sample > milk	Domestic	ISO 16654:2001	Single	25 ml	2	0	0
Vegetables - pre-cut - ready-to-eat - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Unknown	ISO 16654:2001	Single	25 g	111	0	0
Seeds, sprouted - ready-to-eat - Processing plant - Surveillance	NFCSO	Unspecified	Industry sampling	food sample	Unknown	ISO/PRF TS 13136	Batch	25 g	24	0	0
Seeds, sprouted - ready-to-eat - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Unknown	ISO 16654:2001	Single	25 g	59	0	0
Meat from bovine animals - meat products - raw but intended to be eaten cooked - frozen - in total - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Unknown	ISO 16654:2001	Single	25 g	6	0	0
Meat from bovine animals - minced meat - intended to be eaten cooked - in total - Surveillance	NFCSO	Objective sampling	Official sampling	food sample	Unknown	ISO 16654:2001	Single	25 g	15	0	0

Table VT E. coli in food

	Verotoxigenic E. coli (VTEC) - VTEC non- O157	Verotoxigenic E. coli (VTEC) - VTEC, unspecified
Meat from bovine animals - fresh - Processing plant - Surveillance		
Meat from bovine animals - fresh - Retail - Surveillance		
Milk, cows' - raw milk - intended for direct human consumption - Farm - Surveillance		
Milk, goats' - raw milk - intended for direct human consumption - Farm - Surveillance		
Vegetables - pre-cut - ready-to-eat - Retail - Surveillance		
Seeds, sprouted - ready-to-eat - Processing plant - Surveillance	0	0
Seeds, sprouted - ready-to-eat - Retail - Surveillance		
Meat from bovine animals - meat products - raw but intended to be eaten cooked - frozen - in total - Surveillance		
Meat from bovine animals - minced meat - intended to be eaten cooked - in total - Surveillance		

2.4.3 Escherichia coli, pathogenic in animals

A. Verotoxigenic Escherichia coli in cattle (bovine animals)

Monitoring system

Sampling strategy

Monitoring, Official sampling, objective sampling

Frequency of the sampling

Animals at farm

Sampling distributed evenly throughout the year

Animals at slaughter (herd based approach)

Sampling distributed evenly throughout the year

Type of specimen taken

Animals at slaughter (herd based approach)

meat, minced meat

Methods of sampling (description of sampling techniques)

Animals at slaughter (herd based approach)

500 gram meat sample is taken (from one animal), the weight of test portion is 25 grams (cutted from the surface of meat).

The samples are examined by ISO 16654:2001 Standard. Immuno-magnetic concentration is used for the detection of the most important serotype O157. If a strain belongig to the O 157 serotype is isolated, the toxin production is detected by a latex based agglutination test.

Case definition

Animals at slaughter (herd based approach)

The sample is considered to be positive if E. coli O157 was isolated, and the strain produces verotoxin (VT-1, VT-2 or both)

Diagnostic/analytical methods used

Animals at slaughter (herd based approach)

Bacteriological method: ISO 16654:2001

2.5 TUBERCULOSIS, MYCOBACTERIAL DISEASES

2.5.1 General evaluation of the national situation

A. Tuberculosis general evaluation

History of the disease and/or infection in the country

In bovine populations, eradication measures for tuberculosis started in 1962. The eradication of bovine tuberculosis was considered to be completed at the end of 1980. Since then, only sporadic cases occur.

As regards of tuberculosis in man, the favourable tendency which could be observed from the 1950s in the epidemiology of tuberculosis seemed to stop and getting worse in 1990. (Incidence raised by 19% between 1990 and 1995.)In order to lower the incidence and improve the situation, a National Tuberculosis Programme was adopted in 1994 which also incorporated a national surveillance programme based on a central, computerised database.

Recent actions taken to control the zoonoses

Regular screening of the human population is provided. All farm workers have to be checked by the competent public health authority for their compliance with the rules set for persons dealing with animals and food intended for human consumption. The documents proving their compliance are subject to on farm checks performed by the veterinary service. Each county veterinary authority has the right to set further health requirements for persons dealing with animals kept on small size farms.

2.5.2 Mycobacterium in animals

A. Mycobacterium bovis in bovine animals

Status as officially free of bovine tuberculosis during the reporting year

The entire country free

The nationwide program for eradication of bovine tuberculosis in Hungary has successfully been completed by 31 December 1980 and the tuberculosis free status of the country were declared to the OIE. Since then no evidence of the presence of infection in more than 0,1 % of our herds has been found. The Commission Implementing Decision 2014/91/EU recognized Hungary's freedom from the disease.

Monitoring system

Sampling strategy

Post mortem inspections

According to the meat inspection rules in force in Hungary, based on a tradition of at least a century, each animal for slaughter is to be checked individually ante and post mortem. Technical methods applied at meat inspection is suitable to detect even the slightest tuberculotic lesions. The legal provisions for tuberculosis require that the organs, together with the lymphnodes belonging to them, shall be sent to the National Food Chain Safety Office, Veterinary Diagnostic Directorate (former Central Veterinary Institute) for further laboratory examination, if during post mortem inspection of a slaughtered animal the tuberculotic lesions are revealed. In case of animals ordered to be slaughtered for establishing the reason for unclarified positive or inconclusive reactions during intradermal tuberculin testing, a set of lymph nodes belonging to several organs and systems, as listed in the Annex 3 of the Decree No. 65/2002. (VIII. 9.) FVM and in the Technical Guideline, shall be sent to the National Food Chain Safety Office, Veterinary Diagnostic Directorate.

Intradermal tuberculin testing

Together with the post mortem control program, the compulsory intradermal tuberculin testing with a yearly interval of the whole Hungarian cattle population (older than six weeks), as well as case by case testing of animals moved from one herd to another, has been maintained and executed.

Frequency of the sampling

See above.

Methods of sampling (description of sampling techniques)

According to the Annex 3 of the Decree No. 65/2002. (VIII.9) FVM the rules of taking samples are the followings:

All-samples taken from animals with a large body (cattle, swine) must include the organs showing signs of the disease and the adjacent lymphatic glands, in case of birds and smaller animals the sample must be an entire carcass:

All-samples used for confirming paraallergic reaction must include the tonsils, pharyngal, mesenteric and portal lymphatic glands of the slaughtered animal;

All the purpose of detecting the presence of mycobacteria from the feedingstuffs, litter, soil etc. 20-50 gramm samples must be taken, 20 gramm samples from faeces, 50cm3 from urine and 5 litres from drinking water. The samples must be sent to the CVI with a view to carry out tests to detect tuberculosis

and confirm the presence of mycobacteria.

Case definition

An animal is considered a positive case, if the presence of tuberculosis is confirmed by the isolation of M. bovis from its lymph node(s) or parenchymatous organs on laboratory examination.

Suspension or withdrawal of the free status of a herd is based upon the analysis of the results of the intradermal tuberculin tests (if necessary, repeated and completed by simultaneous testing), post mortem examinations and laboratory tests. According to the Annex 1 of the Decree No. 65/2002. (VIII.9) the officially tuberculosis -free status of the herd have to be withdrawn if the presence of tuberculosis is confirmed by the isolation of M. bovis on laboratory examination.

Diagnostic/analytical methods used

The identification of Mycobacterium bovis is carried out only the National Food Chain Safety Office, Veterinary Diagnostic Directorate(VDD) (Budapest). The VDD works according to the OIE Manual of Standards for Diagnostic tests and Vaccines, Forth Edition, Chapter 2.3.3. (bovine tuberculosis).

Annex 7. of the Decree No. 65/2002. (VIII.9) FVM contains the standards for the tuberculin (bovine and avian) to be used during the intradermal tests. These rules are fully compatible with Annex B point 2.1. of Council Directive 64/432/EEC.

Annex 2., which contains the standards for the test procedures is fully compatible with Council Directive 64/432/EEC.

Vaccination policy

Preventive vaccination against M. bovis is prohibited by Decree No. 65/2002. (VIII. 9.) FVM.

Control program/mechanisms

The control program/strategies in place

The whole cattle population is continuously monitored for bovine tuberculosis on a yearly basis by the intradermal tuberculine tests and by post-mortem inspections.

For measures taken in case of single cases, see "Measures in case of the positive findings or single cases".

Recent actions taken to control the zoonoses

Guidelines have been issued first by the Ministry of Agriculture and Rural Development and later by the Central Agricultural Office (the currently valid guideline was issued in March 2010) about the carrying out the tuberculin test in cattle herds taking into consideration the fals positive or interference reactions as well as the data collection, and reporting by the regional authorities.

Measures in case of the positive findings or single cases

When an animal is considered to be a positive reactor in the intradermal tests, it is removed from the herd and slaughtered. The post-mortem, laboratory and epidemiological examinations shall be carried out. The status of the herd will remain suspended until the all laboratory examinations have been completed. If the presence of tuberculosis is not confirmed, the suspension of the officially tuberculosis-free status may be lifted following a test of all animals over six weeks of age with negative results at least 42 days after the removal of the reactor animal.

According to the Annex 1 of the Decree No. 65/2002. (VIII.9) the officially tuberculosis -free status of the herd have to be withdrawn if the presence of tuberculosis is confirmed by the isolation of M. bovis on laboratory examination.

The district chief veterinarian may initiate a procedure to withdraw the tuberculosis-free status of the herd, and the animal health and food control station may withdraw the status, if

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Â-the conditions for retention of the officially free status are not complied with, or

·classical lesions of tuberculosis are seen at post-mortem examination,

A an epidemiological enquiry establishes the likelihood of infection,

·it is deemed necessary to control of bovine tuberculosis in the herd for any other reason.

Notification system in place

Bovine tuberculosis is compulsory notifiable by virtue of the Veterinary Act No CLXXVI. of 2005, which replaced the Veterinary Act No XCI of 1995, from 1 September 2008 by the Decree No 113/2008 (VIII. 30.) of the Ministry of Agriculture and Rural Development (MARD) on notification of animal diseases. The detailed rules regarding bovine tuberculosis are laid down by the Decree No. 65/2002. (VIII.9) FVM of the Minister of Agriculture and Rural Development, which texts replaced the relevant parts of the Zoo-Sanitary Code implemented by the Decree No 41/1997. (V. 28.) FM of the Minister of Agriculture. As regards keeping and movements of the bovine animals the Zoosanitary Code is applied further. Before the 1st of July of 1997 the Decree No. 28/1981. (XII. 30.) MEM of the Minister of Agriculture and Alimentation contained the rules for the bovine tuberculosis and keeping or movements of the bovine animals. It is very important that the former legislative rules were essentially the same as the current ones.

Results of the investigation

During the past consecutive eight years the rate of herds infected with bovine tuberculosis has never reached 0,1 % and at least 99,9% of herds have achieved officially tuberculosis free status each year during this period.

National evaluation of the recent situation, the trends and sources of infection Hungary is free of bovine tuberculosis. However, sporadic cases are reported.

Table Tuberculosis in other animals

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Analytical Method	Sampling unit	Units tested	Total units positive for Mycobacteriu m	M. bovis	M. tuberculosis
Sheep	NFCSO VDD	Suspect sampling		animal sample > lymph nodes	Domestic		Animal	2	0	0	0
Goats	NFCSO VDD	Suspect sampling		animal sample > lymph nodes	Domestic		Animal	17	4	0	0
Pigs	NFCSO VDD	Suspect sampling		animal sample > lymph nodes	Domestic		Animal	34	2	0	0
Badgers - wild	NFCSO VDD	Suspect sampling		animal sample > lymph nodes	Domestic		Animal	4	0	0	0
Deer - wild - red deer - Monitoring - active	NFCSO VDD	Convenience sampling		animal sample > lymph nodes	Domestic		Animal	42	7	0	0
Foxes - wild	NFCSO VDD	Suspect sampling		animal sample > lymph nodes	Domestic		Animal	15	4	0	0
Wild boars - wild - Monitoring - active	NFCSO VDD	Convenience sampling		animal sample > lymph nodes	Domestic		Animal	461	137	0	0

	Mycobacteriu m spp., unspecified	M. avium	M. avium complex - M. avium subsp. paratuberculo sis	
Sheep	0	0	0	0
Goats	3	0	1	0
Pigs	0	2	0	0

Table Tuberculosis in other animals

	Mycobacteriu m spp., unspecified	ivi. avium	M. avium complex - M. avium subsp. paratuberculo sis	IVI caprae I
Badgers - wild	0	0	0	0
Deer - wild - red deer - Monitoring - active	3	4	0	0
Foxes - wild	2	1	0	1
Wild boars - wild - Monitoring - active	57	24	2	54

Table Bovine tuberculosis in countries and regions that do not receive Community co-financing for eradication programmes

If present, the row "Total -1" refers to analogous data of the previous year.

	Total number of	f existing bovine	Officially f	ree herds	Infecte	d herds	Routine tube	rculin testing	Number of tuberculin tests carried out before the introduction	Number of animals with suspicious lesions of	Number of animals detected
Region	Herds	Animals	Number of herds	%	Number of herds	%	Interval between routine tuberculin tests	Number of animals tested	into the herds (Annex A(I)(2)(c) third indent (1) of Directive 64/432/EEC)	tuberculosis examined and submitted to histopathological and bacteriological	positive in bacteriological examination
Baranya	459	30392	457	99.56	1	.22	once a year	24547	193	74	2
Borsod-Abaúj-Zemplén	951	48808	950	99.89	1	.11	once a year	43540	2816	82	2
Budapest	33	1076	33	100	0	0	once a year	983	26	0	0
Bács-Kiskun	2156	75535	2156	100	0	0	once a year	61471	529	31	0
Békés	1565	67358	1565	100	0	0	once a year	57909	3118	6	0
Csongrád	1397	44210	1396	99.93	0	0	once a year	34421	1248	12	0
Fejér	524	48231	524	100	0	0	once a year	41294	3570	9	0
Győr-Moson-Sopron	841	54576	838	99.64	1	.12	once a year	52621	2912	70	1
Hajdú-Bihar	2395	105015	2393	99.92	2	.08	once a year	89601	1763	19	5
Heves	331	15707	331	100	0	0	once a year	13546	936	4	0
Jász-Nagykun-Szolnok	1405	60162	1405	100	0	0	once a year	46357	925	2	0
Komárom-Esztergom	291	14772	291	100	0	0	once a year	13418	554	10	0

Table Bovine tuberculosis in countries and regions that do not receive Community co-financing for eradication programmes

Magyarország	17573	848338	17566	99.96	6	.03	once a year	745468	83744	466	13
Nógrád	350	18505	350	100	0	0	once a year	14725	283	0	0
Pest	1264	54499	1264	100	0	0	once a year	50355	9606	26	0
Somogy	540	37511	532	98.52	1	.19	once a year	35116	27560	95	3
Szabolcs-Szatmár- Bereg	1080	44737	1080	100	0	0	once a year	39015	2084	2	0
Tolna	478	26422	478	100	0	0	once a year	27785	5113	17	0
Vas	582	29749	582	100	0	0	once a year	27409	786	0	0
Veszprém	447	44838	447	100	0	0	once a year	38995	4493	7	0
Zala	484	26235	484	100	0	0	once a year	32360	15229	0	0
Total :	35146	1696676	35122	99.93	12	.03	N.A.	1490936	167488	932	26

Comments:

1) N.A.

Footnote:

Please note that the row "Total" IS NOT VALID as the numbers are exactly duplicated, because of the new row "Magyarország" (Hungary) that contains all the data of the regions.

2.6 BRUCELLOSIS

2.6.1 General evaluation of the national situation

A. Brucellosis general evaluation

History of the disease and/or infection in the country

Hungary is practically free of Brucellosis in bovine, ovine and caprine populations. For detailed information, please refer to the specific texts.

2.6.2 Brucellosis in humans

A. Brucellosis in humans

Reporting system in place for the human cases

1. Reporting system in place for the human cases:

There are around 80 communicable diseases notifiable in Hungary based on legal background. The physician (in primary health care, specialist care, inpatient medical institution or pathology) who first diagnoses a case of a notifiable communicable disease (even the suspicion of the disease!) immediately reports data of case to the first level of the epidemiological network (municipal institute) of National Public Health and Medical Officer's Service (NPHMOS). Data must be reported both at the beginning and at end of the illness (recovery/death, result of laboratory test). The NPHMOS has a nationwide electronic system for registering and analysing data of communicable diseases in a combined national database, so the system makes online connection amid the three levels (municipal, county and national level – National Centre of Epidemiology - NCE) of the organization. The NCE prepares reports regularly (weekly, monthly, yearly) to the Chief Medical Officer, the MoH and the Hungarian Central Statistical Office.

Case definition

Confirmed case: a clinically compatible case and the infection is laboratory confirmed.

Diagnostic/analytical methods used

A serological test (Widal type tube agglutination) is used to confirm the brucellosis diagnose in Hungary. The test preparation is a TTC stained B. melitensis biovar. abortus HNCMB 93007 strain (internationally used diagnostic strain). Result is positive: titre 1:80; uncertain: titre 1:40; negative titre between 1:20 - 1:10. The acute illness is confirmed by the increasing titre of paired sera.

Notification system in place

The disease has been notifiable since 1950 in Hungary. The physician reports data of case on a "case report form" by mail to the municipal institute of NPHMOS. The specialist of the institute records data immediately in the electronic system of the NPHMOS. Hungary has also a laboratory based surveillance system, and the NPHMOS has representative dataset from most of the microbiological laboratories about the cases investigated by the laboratory

History of the disease and/or infection in the country

The disease has been notifiable since 1950 in Hungary. The annual number of reported cases ranged between 0-132 (incidence: $0-1.3/100\ 000$ inhabitants/year, median 21 case/year $-0.2/100\ 000$ inhabitant/year). In the 1950s and 1960s the number of registered cases was about 40-60/year. The most cases were registered between 1970 and 1975 (110-135 cases/year - incidence: $1.1-1.3/100\ 000$ inhabitant/year). Between 1976 and 1986 the number of registered cases decreased to 10 cases/year. 11 death cases occurred between 1950 and 1978. The case fatality rate ranged between 0-6.5% (median 0%).

National evaluation of the recent situation, the trends and sources of infection

There were five cases registered in 2000 - 2001 (2000: 1, 2001: 4 cases), no case was reported between 2002 and 2004, in 2006 and 2008, and only 1-1 case was identified in 2005 and 2007 in Hungary. (The data of laboratory surveillance: 2000 – 4 800 tests, 23 positive; 2001 - 4 900 tests, 30 positive; between 2002 and 2003: about 3 900 tests/year, 6 – 9 /year positive.) No death was registered in this period. One case in 2001 was imported from abroad, in the four other cases between 2000-2001 the place and source of infection could not be identified. Cases registered in 2005 and 2007 were imported cases. No domestic

Hungary - 2013 Report on trends and sources of zoonoses case was reported since then.

2.6.3 Brucella in animals

A. Brucella abortus in bovine animals

Status as officially free of bovine brucellosis during the reporting year

The entire country free

The nationwide programme for eradication of bovine brucellosis in Hungary has successfully been completed by the 31st of August 1985. and the brucellosis free status of the country were declared to the OIE. Since then no evidence of the presence of infection in more than 0,2 % of our herds has been found.

Monitoring system

Sampling strategy

Together with the random blood sampling of the Hungarian cattle population, as well as case-by-case testing of animals moved from one herd to another, a system of checking abortions and irregular parturition has been maintained.

Frequency of the sampling

The whole cattle population in Hungary is subject to regular checks. Investigation of abortion and related cases is the key point of the system. Random, yearly serological testing is a complementary element. 10 % of cows in herds containing 50 or more animals shall be tested yearly, after calving. If necessary, the district veterinary officer is entitled to extend the testing to the whole herd.

Small herds are serologically tested every three years, linked to the EBL screening.

Type of specimen taken

Blood

Methods of sampling (description of sampling techniques)

Blood, milk and semen samples are taken at farm. In case of abortion, the aborted fetus, its chorions and a blood sample from the aborted cattle shall be sent to the laboratory.

Case definition

An animal is considered to be infected with B. abortus, when

- it shows clinical signs of the disease and pathological lesions can be detected on its internal organs or on its fetus or on the chorions; or
- bacteria of B. abortus could be isolated from its body fluids, its chorions or from the organs of the fetus, or
- it was suspected to be infected with B. abortus and the serological or bacteriological investigations were positive for that animal.

Diagnostic/analytical methods used

For the diagnosis of B. abortus the following diagnostic methods are used:

- -pathology
- -bacteriology
- -immunology (CFT, ELISA, SAT)

Vaccination policy

Preventive vaccination against B. abortus is prohibited in the whole territory of Hungary.

Control program/mechanisms

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Recent actions taken to control the zoonoses

Continuous monitoring of bovine herds and investigation of aborted fetuses as well as pre-movement checks are continued.

Measures in case of the positive findings or single cases

Infected male animals are

to be killed as soon as possible but not later than five days or,

to be castrated and placed under movement prohibition until it is slaughtered.

Female animals must be placed under breeding prohibition and movement control. They must be slaughtered within 15 days after the acute period or the recovery after the abortion.

Notification system in place

Bovine brucellosis (B. abortus) is compulsorily notifiable by virtue of the Act on Food Chain Safety and its official control No XLVI of 2008 that is effective since 1 September 2008 and the Decree of the Minister of Agriculture No 12/2008 (II. 14.) on detailed rules of the protection regarding certain Brucella species. Notification, as well as investigation of cases of abortion is compulsory. In case of abortion or irregular parturition, the veterinarian in charge has to send a set of samples, listed in the decree mentioned above, for further laboratory examination. Until thorough clarification of the case, the animal is kept separated and, if necessary, repeatedly tested.

Results of the investigation

Since 1985 no infection of B. abortus has been found.

B. Brucella melitensis in goats

Status as officially free of caprine brucellosis during the reporting year

The entire country free

Ovine and caprine brucellosis (B. melitensis) has been a compulsorily notifiable animal disease in Hungary since 1982. Further to the existing rules laid down in the Zoo-Sanitary Code, the recent legal provisions give the power to the Ministry of Agriculture to introduce any additional measures, should an outbreak of a disease caused by B. melitensis occur in our country.

Neither a single clinical case, nor any positive serological or bacteriological test result for B. melitensis has ever occurred in Hungary. The Commission Decision 93/52/EEC recognized Hungary's freedom from the disease.

Monitoring system

Sampling strategy

Given, that B. melitensis is not an agent which can be spread under Hungary's geographical and climatic conditions, furthermore no sign of the disease has ever been revealed, there was no scientifically based reason for an extended serological survey. Since 2007, all caprine animals tested for B. melitensis were negative.

Frequency of the sampling

Approximately 5% of the caprine population is sampled and tested for B. melitensis.

Type of specimen taken

Blood

Methods of sampling (description of sampling techniques)

Blood samples are taken at farm.

Case definition

An animal is considered to be infected with B. melitensis, when

- it shows clinical signs of the disease and pathological lesions can be detected on its internal organs or on its fetus or on the chorions; or
- bacteria of B. melitensis could be isolated from its body fluids, its chorions or from the organs of the fetus, or
- it was suspected to be infected with B. melitensis and the serological or bacteriological investigations were positive for that animal.

Diagnostic/analytical methods used

For the diagnosis of B. melitensis in goats, the CFT is used.

Vaccination policy

Vaccines for B. melitensis have never been registered in Hungary and the using of vaccines without the registration is banned in the country. Therefore no vaccination against this disease has ever been practised in the territory of Hungary.

Control program/mechanisms

The control program/strategies in place

Hungary is free of B. melitensis. However, monitoring of ovine and caprine populations is continuously done.

Measures in case of the positive findings or single cases

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In case of positive findings the positive animals have to be killed without delay. The herd containing the positive animal is subject to movement control. The further measures affecting the herd shall be decided following screening of the animals and epidemiological investigation.

Notification system in place

Ovine and caprine brucellosis (B. melitensis) are compulsorily notifiable by virtue of the Veterinary Act No CLXXVI. of 2005 (which replaced the Veterinary Act No XCI of 1995) and the Zoo-Sanitary Code implemented by the Decree No 41/1997. (V. 28.) FM of the Minister of Agriculture. These legal texts replaced the former regulations, namely Law Decree No 3. of 1981. and Decree No. 28/1981. (XII. 30.) MEM of the Minister of Agriculture and Alimentation, which have contained the same provisions for the diseases mentioned above. Therefore we can declare that ovine and caprine brucellosis is compulsory since 1 January 1982 on the basis of Decree No. 28/1981. (XII. 30.) MEM of the Minister of Agriculture and Alimentation.

Results of the investigation

No evidence of infection with B. melitensis was ever found.

C. Brucella melitensis in sheep

Status as officially free of ovine brucellosis during the reporting year

The entire country free

Ovine and caprine brucellosis (B. melitensis) has been a compulsorily notifiable animal disease in Hungary since 1982. Further to the existing rules laid down in the Zoo-Sanitary Code, the recent legal provisions give the power to the Ministry of Agriculture to introduce any additional measures, should an outbreak of a disease caused by B. melitensis occur in our country.

Neither a single clinical case, nor any positive serological or bacteriological test result for B. melitensis has ever occurred in Hungary. The Commission Decision 93/52/EEC recognized Hungary's freedom from the disease.

Monitoring system

Sampling strategy

Given, that B. melitensis is not an agent which can be spread under Hungary's geographical and climatic conditions, furthermore no sign of the disease has ever been revealed, there was no scientifically based reason for an extended serological survey. However, between 1997 and 2000 a limited serological screening was carried out and all results were negative. Since 2001 an extended serological survey has been started to demonstrate the B. melitensis free status of Hungary. During 2001, 2002 and 2003 more than 10% of the ovine animals over six months of age were tested serologically for B. melitensis and all results were negative. All ovine animals tested for B. melitensis were negative.

Frequency of the sampling

Approximately 5% of the ovine population were tested.

Type of specimen taken

Blood

Methods of sampling (description of sampling techniques)

Blood samples are taken at farm.

Case definition

An animal is considered to be infected with B. melitensis, when

- it shows clinical signs of the disease and pathological lesions can be detected on its internal organs or on its fetus or on the chorions; or
- bacteria of B. melitensis could be isolated from its body fluids, its chorions or from the organs of the fetus, or
- it was suspected to be infected with B. melitensis and the serological or bacteriological investigations were positive for that animal.

Diagnostic/analytical methods used

For the diagnostic serological tests of B. melitensis the CFT is used.

Vaccination policy

Vaccines for B. melitensis have never been registered in Hungary and the using of vaccines without the registration is banned in the country. Therefore no vaccination against this disease has ever been practised in the territory of Hungary.

Control program/mechanisms

The control program/strategies in place

Hungary is free of B. melitensis. However, monitoring of ovine and caprine populations is continuously

done.

Measures in case of the positive findings or single cases

In case of positive findings the positive animals have to be killed without delay. The herd containing the positive animal is subject to movement control. The further measures affecting the herd shall be decided following screening of the animals and epidemiological investigation.

Notification system in place

Ovine and caprine brucellosis (B. melitensis) are compulsorily notifiable by virtue of the Veterinary Act No CLXXVI. of 2005 (which replaced the Veterinary Act No XCI of 1995) and the Zoo-Sanitary Code implemented by the Decree No 41/1997. (V. 28.) FM of the Minister of Agriculture. These legal texts replaced the former regulations, namely Law Decree No 3. of 1981. and Decree No. 28/1981. (XII. 30.) MEM of the Minister of Agriculture and Alimentation, which have contained the same provisions for the diseases mentioned above. Therefore we can declare that ovine and caprine brucellosis is compulsory since 1 January 1982 on the basis of Decree No. 28/1981. (XII. 30.) MEM of the Minister of Agriculture and Alimentation.

Results of the investigation

No evidence of infection with B. melitensis were found.

Table Ovine or Caprine Brucellosis in countries and regions that do not receive Community co-financing for eradication programme

If present, the row "Total -1" refers to analogous data of the previous year.

	Total number	er of existing	Officially free herds		Infected herds		Surveillance			Investigations of suspect cases					
Region	Herds	Animals	Number of herds	%	Number of herds	%	Number of herds tested	Number of animals tested	Number of infected herds	Number of animals tested with serological blood tests	Number of animals positive serologically	Number of animals examined microbio logically	Number of animals positive microbio logically	Number of suspended herds	
Baranya	58	15706	58	100	0	0	51	1223	0	0	0	0	0	0	
Borsod-Abaúj-Zemplén	276	60429	276	100	0	0	149	2308	0	0	0	0	0	0	
Bács-Kiskun	1194	154803	1194	100	0	0	270	8315	0	0	0	0	0	0	
Békés	866	42447	866	100	0	0	125	1954	0	0	0	0	0	0	
Csongrád	667	31774	667	100	0	0	134	1979	0	0	0	0	0	0	
Fejér	252	36982	252	100	0	0	73	1849	0	0	0	0	0	0	
Győr-Moson-Sopron	91	6916	91	100	0	0	14	328	0	0	0	0	0	0	
Hajdú-Bihar	1319	122550	1319	100	0	0	379	9825	0	0	0	0	0	0	
Heves	279	14834	279	100	0	0	78	864	0	0	0	0	0	0	
Jász-Nagykun-Szolnok	689	43450	689	100	0	0	90	2099	0	0	0	0	0	0	
Komárom-Esztergom	85	8744	85	100	0	0	17	422	0	0	0	0	0	0	
Magyarország	7786	844361	7786	100	0	0	2420	47305	0	0	0	0	0	0	
Nógrád	313	15016	313	100	0	0	44	838	0	0	0	0	0	0	

Table Ovine or Caprine Brucellosis in countries and regions that do not receive Community co-financing for eradication programme

Pest	223	40797	223	100	0	0	134	2697	0	0	0	0	0	0
Somogy	348	21800	348	100	0	0	18	912	0	0	0	0	0	0
Szabolcs-Szatmár- Bereg	659	139927	659	100	0	0	659	7240	0	0	0	0	0	0
Tolna	105	27897	105	100	0	0	61	1648	0	0	0	0	0	0
Vas	83	3337	83	100	0	0	5	133	0	0	0	0	0	0
Veszprém	166	46289	166	100	0	0	102	2181	0	0	0	0	0	0
Zala	113	10663	113	100	0	0	17	490	0	0	0	0	0	0
Total:	15572	1688722	15572	100	0	0	4840	94610	0	0	0	0	0	0

Comments:

1) N.A.

Footnote:

Please note that the row "Total" IS NOT VALID as the numbers are exactly duplicated, because of the new row "Magyarország" (Hungary) that contains all the data of the regions.

Table Bovine brucellosis in countries and regions that do not receive Community co-financing for eradication programme

If present, the row "Total -1" refers to analogous data of the previous year.

	Total nu	umber of	Officially t	free herds	lasta atau				Surve	illance		· ·				Investigation	ons of susp	pect cases	3		
	existing	g bovine			Infected	a neras	Se	rological te	ests	Examir	nation of b	ulk milk	Info	ormation al	oout		Epid	lemiologica	al investig	ation	
							Number of	Number of	Number of	Number of	Number of	Number of	Number of notified	Number of isolations	Number of abortions	Number of animals tested with	Number of		of positive mals	Number of animals	Number of animals
	Herds	Animals	Number of herds	%	Number of herds	%	bovine herds tested	animals tested	infected herds	bovine herds tested	animals or pools tested	infected herds	abortions whatever	of Brucella infection	due to Brucella	serological blood tests	suspended herds	Sero logically	BST	examined microbio	positive microbio
Region													cause		abortus			logically		logically	logically
Baranya	459	30392	459	100	0	0	321	10660	0	1	374	0	17	0	0	0	0	0	0	0	0
Borsod-Abaúj-Zemplén	951	48808	951	100	0	0	694	23909	0	2	645	0	50	0	0	0	0	0	0	0	0
Budapest	33	1076	33	100	0	0	28	495	0	0	0	0	3	0	0	0	0	0	0	0	0
Bács-Kiskun	2156	75535	2156	100	0	0	1657	26801	0	18	1128	0	70	0	0	0	0	0	0	0	0
Békés	1565	67358	1565	100	0	0	1565	25452	0	0	0	0	241	0	0	0	0	0	0	0	0
Csongrád	1397	44210	1397	100	0	0	801	17633	0	46	658	0	72	0	0	0	0	0	0	0	0
Fejér	524	48231	524	100	0	0	524	22625	0	9	2312	0	42	0	0	0	0	0	0	0	0
Győr-Moson-Sopron	841	54576	841	100	0	0	472	21571	0	10	1432	0	128	0	0	0	0	0	0	0	0
Hajdú-Bihar	2395	105015	2395	100	0	0	1804	49308	0	0	0	0	142	0	0	0	0	0	0	0	0
Heves	331	15707	331	100	0	0	331	8551	0	0	0	0	26	0	0	0	0	0	0	0	0
Jász-Nagykun-Szolnok	1405	60162	1404	99.93	0	0	898	25459	0	1	212	0	62	0	0	0	0	0	0	0	0
Komárom-Esztergom	291	14772	291	100	0	0	251	12016	0	2	632	0	31	0	0	0	0	0	0	0	0

Table Bovine brucellosis in countries and regions that do not receive Community co-financing for eradication programme

Magyarország	17573	848338	17566	99.96	0	0	13806	373913	0	111	11989	0	1079	0	0	0	0	0	0	0	0
Nógrád	350	18505	350	100	0	0	328	8802	0	0	0	0	12	0	0	0	0	0	0	0	0
Pest	1264	54499	1264	100	0	0	1031	29159	0	7	3756	0	14	0	0	0	0	0	0	0	0
Somogy	540	37511	534	98.89	0	0	520	13338	0	0	0	0	27	0	0	0	0	0	0	0	0
Szabolcs-Szatmár- Bereg	1080	44737	1080	100	0	0	1080	18876	0	0	0	0	18	0	0	0	0	0	0	0	0
Tolna	478	26422	478	100	0	0	231	8705	0	15	840	0	15	0	0	0	0	0	0	0	0
Vas	582	29749	582	100	0	0	481	15159	0	0	0	0	25	0	0	0	0	0	0	0	0
Veszprém	447	44838	447	100	0	0	447	22616	0	0	0	0	69	0	0	0	0	0	0	0	0
Zala	484	26235	484	100	0	0	342	12778	0	0	0	0	15	0	0	0	0	0	0	0	0
Total:	35146	1696676	35132	99.96	0	0	27612	747826	0	222	23978	0	2158	0	0	0	0	0	0	0	0

Comments:

1) N.A.

Footnote:

Please note that the row "Total" IS NOT VALID as the numbers are exactly duplicated, because of the new row "Magyarország" (Hungary) that contains summarized of the regions.

2.7 YERSINIOSIS

2.7.1 General evaluation of the national situation

A. Yersinia enterocolitica general evaluation

Additional information

diagnostic methods: bacteriological examination and PCR

2.7.2 Yersiniosis in humans

A. Yersinosis in humans

Reporting system in place for the human cases

There are around 80 communicable diseases notifiable in Hungary based on legal background. The physician (in primary health care, specialist care, inpatient medical institution or pathology) who first diagnoses a case of a notifiable communicable disease (even the suspicion of the disease!) immediately reports data of case to the first level of the epidemiological network (municipal institute) of National Public Health and Medical Officer's Service (NPHMOS). Data must be reported both at the beginning and at end of the illness (recovery/death, result of laboratory test). The NPHMOS has a nationwide electronic system for registering and analysing data of communicable diseases in a combined national database, so the system makes online connection between the three levels (municipal, regional and national level – National Centre of Epidemiology - NCE) of the organization. The NCE prepares reports regularly (weekly, monthly, yearly) to the Chief Medical Officer, the MoH and the Hungarian Central Statistical Office.

Case definition

Confirmed case: a clinically compatible case when the Yersinia infection is laboratory confirmed.

Diagnostic/analytical methods used

Yersinia isolates are obtained by culturing the faeces samples of the patients on selective-differentiating media, which is followed by biochemical tests and serotyping. Earlier the sera of the patient was tested by Widal-typed method, beside this test the ELISA method has been also in use since 2003.

Notification system in place

Human cases have been notifiable since 1998. The physician reports data of case on a "case report form" by mail to the municipal institute of NPHMOS. The specialist of the institute records data immediately in the electronic system of the NPHMOS. Hungary has also a laboratory based surveillance system, and the NPHMOS has representative dataset from most of the microbiological laboratories about the laboratory investigated cases (since 2003 antibiotic resistances has also been reported from 20 county institutes and 12 laboratories from universities or hospitals).

The illness is reported firstly as enteritis infectiosa syndrome on the basis of the symptoms. Having the results of the laboratory tests this syndrome-based diagnose is modified to etiology-based diagnose. There is a part of the cases which are reported only subsequently when the result of the laboratory test is available.

History of the disease and/or infection in the country

The human cases have been notifiable since 1998. The number of cases varied between 68 - 176/year (incidence: $0.7 - 1.7/100\ 000$ inhabitant/year, median 125 cases/year - $1.3/100\ 000$ inhabitant/year). There was no death registered. A few number of family outbreaks were investigated, community or institutional outbreaks did not occur. Laboratory or epidemiological evidences are not available to assess the source of infection.

National evaluation of the recent situation, the trends and sources of infection

Yersiniosis do not influence significantly the epidemiological situation of the human acut gastroenteritis caused by zoonotic agents. Between 2000 -2004 the dominant serotype is Y.enterocolitica O3. It is confirmed also by the results of culture and serologic methods.

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2.7.3 Yersinia in animals

Table Yersinia in animals

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Units tested	Total units positive for Yersinia	Y. enterocolitica		Yersinia spp., unspecified
Cattle (bovine animals) - Clinical investigations	NFCSO VDD	Unspecified	Not applicable	animal sample > organ/tissue		Animal	1	0	0	0	0
Mouflons - wild - Natural habitat - Unspecified	NFCSO VDD	Unspecified	Not applicable	animal sample > organ/tissue	Domestic	Animal	12	0	0	0	0
Pigs - breeding animals - unspecified - Slaughterhouse - Unspecified	NFCSO VDD	Unspecified		animal sample > organ/tissue		Animal	20	1	1	0	0
Wild ducks - Natural habitat - Clinical investigations (Bucephala clangula)	NFCSO VDD	Unspecified	Not applicable	animal sample > organ/tissue	Domestic	Animal	1	1	0	1	0

	Y. enterocolitica - O:3	Y. enterocolitica - O:9	Y. enterocolitica - unspecified
Cattle (bovine animals) - Clinical investigations	0	0	0
Mouflons - wild - Natural habitat - Unspecified	0	0	0
Pigs - breeding animals - unspecified - Slaughterhouse - Unspecified	0	0	1
Wild ducks - Natural habitat - Clinical investigations (Bucephala clangula)	0	0	0

Table Yersinia in animals

Comments:

1) foetus, uterus, lymphonodes

Footnote:

The duck's and the mouflons' samples sent in to the laboratory were whole dead bodies. Whole bodies were sent in in the case of pigs too.

2.8 TRICHINELLOSIS

2.8.1 General evaluation of the national situation

A. Trichinellosis general evaluation

History of the disease and/or infection in the country

In Hungary, mandatory testing for Trichinella spp. is in place since 1960. Slaughtered susceptible animals intended to be placed on the market are subject to mandatory testing for Trichinella spp.

National evaluation of the recent situation, the trends and sources of infection

Trichinellosis was a significant zoonotic disease in Hungary in the 1950s and 1960s. Due to the introduction of control strategies, the average annual incidence of trichinellosis decreased to 0-0.7 cases per 100,000 for the early 1990s. In the past 15 years, the annual incidence dropped to 0-0.07 cases per 100,000, and no mortality in men caused by the parasite was observed in the same period. The decrease of incidence observed in men is similar to that of prevalence seen in swine at slaughterhouses. Nevertheless, some increasing trends of incidence might be observed in both men and swine in the past years. Trichinella spiralis still persists in the southern and eastern border region of the country. Sporadic Trichinella infections (in average few cases per year) were also detected in wild boars and in less than 1.8% of red foxes. In wild boars, both T. spiralis and Trichinella britovi were detected. In foxes, Trichinella britovi is the dominant species; nevertheless, Trichinella spiralis and Trichinella pseudospiralis were also reported from this species.

Recent actions taken to control the zoonoses

Mandatory testing during meat inspection in all susceptible cases (swine, horse, nutria, wild boar).

2.8.2 Trichinellosis in humans

A. Trichinellosis in humans

Reporting system in place for the human cases

There are about 80 communicable diseases notifiable in Hungary based on legal background. The physician (in primary health care, specialist care, inpatient medical institution or pathology) who first diagnoses a case of a notifiable communicable disease (even the suspicion of the disease!) immediately reports data of case to the first level of the epidemiological network (municipal institute) of National Public Health and Medical Officer's Service (NPHMOS). Data must be reported both at the beginning and at end of the illness (recovery/death, result of laboratory test). The NPHMOS has a nationwide electronic system for registering and analysing data of communicable diseases in a combined national database, so the system provides online connection between the three levels (municipal, regional and national level – National Centre of Epidemiology - NCE) of the organization. The NCE prepares reports regularly (weekly, monthly, yearly) to the Chief Medical Officer, the MoH and the Hungarian Central Statistical Office.

Case definition

Confirmed case: a clinically compatible case when the Trichinella infection is laboratory confirmed. Probable case: a clinically compatible case that is not confirmed by laboratory investigation, but it has an epidemiological link to a confirmed trichinellosis outbreak.

Diagnostic/analytical methods used

Microprecipitic test on live larvae as diagnostic method has been used since 1983 in the Helmithozoonotic Reference Laboratory of the National Centre of Epidemiology. Parallel with this test an ELISA test (NOVATEC TRICHINELLA SPIRALIS IgG-ELISA, NovaTec Immundiagnostica, Germany) was introduced in 2002. The positive results of the previously mentioned tests have been confirmed by WB (TRICHINELLA WESTERN BLOT IgG, Ldbio Diagnostics, France) since 2004.

Notification system in place

Human cases have been notifiable since 1960. The physician reports data of case on a "case report form" by mail to the municipal institute of NPHMOS. The specialist of the institute records data immediately in the electronic system of the NPHMOS.

History of the disease and/or infection in the country

Human cases have been notifiable since 1960. The number of cases varied between 0-121 (incidence $0-1,2/100\ 000$ inhabitants/year – the highest one was registered in 1964). Between 1960 and 2004 the 85% of cases had epidemiological link to an outbreak. Only one death case has been registered during the Hungarian history of trichinellosis.

Between 1960 and 1975 the swine were the source of infection in 18 outbreaks (83% of all outbreaks) and wild boar in 17% of outbreaks. The significance of swine as the source of infection decreased between 1976 and 1995: 3 outbreaks (23%) were caused by swine, and 10 outbreaks (77%) were associated with consumption of wild boar meat. (Indigenous swine were the source of two outbreaks in 1978 and 1990, and swine imported from Romania and processed at home were the source of one outbreak in 1995).

National evaluation of the recent situation, the trends and sources of infection

In the last ten years the number of reported cases ranged between 0 - 7/year (incidence 0 - 0.07/100~000 inhabitants/year), there was no death in this period. All cases linked to family outbreaks and most of sporadic cases were imported from the neighbouring counties. The indigenous cases were linked to the

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consumption of indigenous wild boar meat. All human cases were caused by T.spiralis.

2.8.3 Trichinella in animals

A. Trichinella in pigs

Monitoring system

Sampling strategy

Trichinella sampling and testing is mandatory for all pigs intended to be placed on the market.

Frequency of the sampling

Every slaughtered animal is sampled

Type of specimen taken

Diaphragm muscle

Methods of sampling (description of sampling techniques)

Methods specified in Regulation 2075/2005/EC

Case definition

Animal with one or more Trichinella larva in the official examination.

Diagnostic/analytical methods used

Artificial digestion method of collective samples.

Vaccination policy

None.

Control program/mechanisms

The control program/strategies in place

See above.

Measures in case of the positive findings or single cases

Positive cases are considered not to be eligible for human consumption.

Results of the investigation

All slaughtered swine are investigated. There was no positive finding for Trichinella.

B. Trichinella in horses

Monitoring system

Sampling strategy

Trichinella testing is mandatory, all animal is sampled.

Frequency of the sampling

Every slaughtered animal is sampled

Type of specimen taken

Diaphragm muscle

Methods of sampling (description of sampling techniques)

2075/2005/EC regulation

Case definition

Animal with one or more Trichinella larva in the official examination

Diagnostic/analytical methods used

Artificial digestion method of collective samples

Vaccination policy

None.

Measures in case of the positive findings or single cases

Positive cases are considered not to be eligible for human consumption.

Results of the investigation

All the slaughtered horses (as all other susceptible animals) are investigated. There was no positive finding for trichinella.

National evaluation of the recent situation, the trends and sources of infection

Trichinella infection has never been detected in horses in Hungary.

Table Trichinella in animals

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Units tested	Total units positive for Trichinella	T. spiralis	Trichinella spp., unspecified	T. britovi
Pigs - fattening pigs	Veterinary Diagnostic Directorate	Census	Official sampling	animal sample > organ/tissue	Domestic	Animal	4025314	0	0	0	0
Solipeds, domestic - horses - Slaughterhouse - Surveillance	VDD	Census	Official sampling	animal sample > organ/tissue	Domestic	Animal	1116	0	0	0	0
Wild boars - wild - Surveillance	VDD	Census	Official sampling	animal sample > organ/tissue	Domestic	Animal	57618	11	1	1	9
Foxes - wild	VDD	Objective sampling	Official sampling	animal sample > organ/tissue	Domestic	Animal	575	17	4	6	7
Jackals - wild	VDD	Objective sampling	Official sampling	animal sample > organ/tissue	Domestic	Animal	18	1	1	0	0
Rats - wild - Monitoring	VDD	Unspecified	Official sampling	animal sample > organ/tissue	Domestic	Animal	1	0	0	0	0
Rodents - wild - Monitoring	VDD	Unspecified	Official sampling	animal sample > organ/tissue	Domestic	Animal	1	0	0	0	0

2.9 ECHINOCOCCOSIS

2.9.1 General evaluation of the national situation

A. Echinococcus spp. general evaluation

History of the disease and/or infection in the country

Echinococcus granulosus

Cystic echinococcosis caused by E. granulosus was a significant zoonosis in Hungary in the 1960s and 1970s. Due to the introduction of integrated control strategies, the average annual incidence of human cystic echinococcosis decreased to 0.08-0.2 case per 100,000 population for the early 1990s. The decrease of incidence observed in man is almost parallel with that of overall prevalence seen in swine, sheep and cattle at slaughterhouses.

Echinococcus multilocularis was not detected in man or animals in Hungary until 2002.

National evaluation of the recent situation, the trends and sources of infection

Echinococcus granulosus

In the past decade, the annual incidence was 0.05-0.1 case per 100,000 human population. The prevalence was under 0.2% in sheep, cattle and swine at slaughterhouses. Genotype identification of slaughterhouse isolates was intitiated in 2010.

Echinococcus multilocularis

E. multilocularis was first detected in red foxes (Vulpes vulpes) in Hungary in the northern border area in 2002. Between 2002 and 2004, the parasite was described in 7 northern counties with low overall prevalence (8.7%) in foxes. In the study carried out in 2009, E. multilocularis was detected in foxes of 16 out of the 19 Hungarian counties and in the suburban areas of the capital, Budapest. The prevalence of infection was significantly higher in the north-western half (16.2%) than in the south-eastern half (4.2%) of the country. The multi-locus microsatellite analysis of the isolates indicate that Hungary should be considered as a peripheral area of a single European focus, where the dispersal movement of foxes resulted in the spreading of E. multilocularis within a time period short enough to avoid a substantial genetic drift.

2.9.2 Echinococcosis in humans

A. Echinococcus spp. in humans

Reporting system in place for the human cases

There are about 80 communicable diseases notifiable in Hungary based on legal background. The physician (in primary health care, specialist care, inpatient medical institution or pathology) who first diagnoses a case of a notifiable communicable disease (even the suspicion of the disease!) immediately reports data of case to the first level of the epidemiological network (municipal institute) of National Public Health and Medical Officer's Service (NPHMOS). Data must be reported both at the beginning and at end of the illness (recovery/death, result of laboratory test). The NPHMOS has a nationwide electronic system for registering and analysing data of communicable diseases in a combined national database, so the system provides online connection amid the three levels (municipal, regional and national level – National Centre of Epidemiology - NCE) of the organization. The NCE prepares reports regularly (weekly, monthly, yearly) to the Chief Medical Officer, the MoH and the Hungarian Central Statistical Office.

Case definition

Confirmed case: a clinically compatible case when the Echinococcus infection is laboratory confirmed

Diagnostic/analytical methods used

The punctatum originated from cyst or sample from extracted cyst is investigated by microscopic methods. IHA (CELLOGNOST ECHINOCOCCOSIS for IHA, Dade Behring, Germany) and ELISA (HYDATIDOSIS ELISA IgG, Vircell, Spain) screening methods have been used parallel since 2002 in the Helminthozoonoses Reference Laboratory in 'Johan Béla' National Centre for Epidemiology. The positive results are confirmed by Western blot method (WB) (ECHINOCOCCUS WESTERN BLOT IgG, Ldbio Diagnostics, France).

Notification system in place

The disease has been notifiable since 1950 in Hungary. The physician reports data of case on a "case report form" by mail to the municipal institute of NPHMOS. The specialist of the institute records data immediately in the electronic system of the NPHMOS.

History of the disease and/or infection in the country

Complement-fixed test has been used since 1934 in Hungary to identify the presence of anti-Echinococcus antibody titre. The human cases have been notifiable since 1960. The "home made" indirect hemagglutination (IHA) was introduced in 1985, and the "home made" ELISA method in 1987. The number of registered cases ranged between 0 - 18 /year (more then 10 cases registered in the 1980s only), the incidence varied between 0 - 0.2 cases/100 000 inhabitants/year. There were 0 - 4 death cases reported yearly (the median of case fatality rate: 20%). Since 1991 there has not been any death case with this diagnosis.

National evaluation of the recent situation, the trends and sources of infection

The number of annually reported cases varied between 5 and 13 in the last five years, there was no death registered. All the reported cases were caused by E. granulosus confirmed in the reference laboratory by Western immunoblot method. In Hungary, autochtonous human case has not been identified as E. multilocularis infection.

2.9.3 Echinococcus in animals

Table Echinococcus in animals

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Region	Units tested	Total units positive for Echinococcus	E. granulosus	E. multilocularis
Cattle (bovine animals) - Slaughterhouse - Surveillance	VDD	Unspecified	Official sampling	animal sample > organ/tissue	Domestic	Animal		524	0	0	0
Sheep - Slaughterhouse - Surveillance	VDD	Unspecified	Official sampling	animal sample > organ/tissue	Domestic	Animal		384	2	0	0
Goats - Slaughterhouse - Surveillance	VDD	Unspecified	Official sampling	animal sample > organ/tissue	Domestic	Animal		71	0	0	0
Pigs - Slaughterhouse - Surveillance	VDD	Unspecified	Official sampling	animal sample > organ/tissue	Domestic	Animal		1144	1	0	0
Solipeds, domestic - horses - Slaughterhouse - Surveillance	VDD	Unspecified	Official sampling	animal sample > organ/tissue	Domestic	Animal		24	0	0	0
Foxes - wild	VDD	Objective sampling	Not applicable	animal sample > organ/tissue	Domestic	Animal		365	17	17	0
Jackals - wild	VDD	Objective sampling	Not applicable	animal sample > organ/tissue	Domestic	Animal		16	1	1	0

	Echinococcus spp., unspecified
Cattle (bovine animals) - Slaughterhouse - Surveillance	0

Table Echinococcus in animals

	Echinococcus spp., unspecified
Sheep - Slaughterhouse - Surveillance	2
Goats - Slaughterhouse - Surveillance	0
Pigs - Slaughterhouse - Surveillance	1
Solipeds, domestic - horses - Slaughterhouse - Surveillance	0
Foxes - wild	0
Jackals - wild	0

Footnote:

VDD = Veterinary Diagnostic Directorate

2.10 TOXOPLASMOSIS

2.10.1 General evaluation of the national situation

2.10.2 Toxoplasmosis in humans

A. Toxoplasmosis in humans

Reporting system in place for the human cases

There are around 80 communicable diseases notifiable in Hungary based on legal background. The physician (in primary health care, specialist care, inpatient medical institution or pathology) who first diagnoses a case of a notifiable communicable disease (even the suspicion of the disease!) immediately reports data of case to the first level of the epidemiological network (municipal institute) of National Public Health and Medical Officer's Service (NPHMOS). Data must be reported both at the beginning and at end of the illness (recovery/death, result of laboratory test). The NPHMOS has a nationwide electronic system for registering and analysing data of communicable diseases in a combined national database, so the system makes online connection amid the three levels (municipal, regional and national level – National Centre of Epidemiology - NCE) of the organization. The NCE prepares reports regularly (weekly, monthly, yearly) to the Chief Medical Officer, the MoH and the Hungarian Central Statistical Office.

Case definition

Confirmed case: a clinically compatible case when the Toxoplasma infection is laboratory confirmed.

Diagnostic/analytical methods used

The anti-Toxoplasma ELISA IgG and IgM methods (TOXONOSTIKA IgG, TOXONOSTIKA IgM, Organon Teknika, Hollandia) are used in the everyday diagnostic work since 1986 in Hungary. Today the specific anti-Toxoplasma IgG (PLATELIA® Toxo IgG, Bio-Rad, France), IgM (PLATELIA® Toxo IgM, Bio-Rad, France), IgG avidity identification (VIDAS, BioMérieux S/A, France) is used to test for the anti-Toxoplasma serologic profile.

The PCR method (classical: PRODECT TOXO B1, Bioanalisi Centro Sud s.n.c., Italy; and the light cycler method: LIGHTCYCLER FASTSTART DNA MASTERPLUS HYBRIDIZATION PROBES, Roche (Hungary) Ltd.), further the IgG/IgM Western blot test comparing the immunprofile of mother and child (TOXOPLASMA WESTERN BLOT IgG/IgM, Ldbio Diagnostics, France) are applied. For quality assurance purposes the Toxoplasma Reference Laboratory participate twice in a year in proficiency test, and the Reference Laboratory also organise proficiency tests for laboratory of NPHMOS.

History of the disease and/or infection in the country

Anti-Toxoplasma antibody assay (Sabin-Feldman dye test) has been in use since 1958 in Hungary. The human cases have been notifiable since 1967. The "home made" complement-fixed assay and indirect hemagglutination methods (IHA) were introduced in 1969.

The annual number of registered cases ranged between 0-333 (median: 136 case/year), so the incidence varied $0-3.1/100\ 000$ inhabitants/year (median 1.3/100 000/year). Between 1970 and 1985 the highest number of death cases reported was 1-5 deaths/year (max. case fatality rate 10%). Only two death cases occurred between 1985 and 2004.

National evaluation of the recent situation, the trends and sources of infection

Hungary - 2013 Report on trends and sources of zoonoses

The number of annually registered cases ranged between 292 - 107 /year (incidence $2.9 - 1.1/100\ 000$ inhabitant/year – median $1.8/100\ 000$ inhabitant/year), the trend of the incidence is decreasing. There was no death registered in this period. It was a seroprevalence survey performed by Helmithozoonotic Reference Laboratory of National Centre for Epidemiology in 2001. 6 985 persons without sings or symptoms were tested by serologic method for the presence of Toxoplasma antibodies. The proportion of positive persons ranged between 22.8% - 41.3% by county. The proportion of positive persons was 75% among pupils aged more than 60 years.

2.11 RABIES

2.11.1 General evaluation of the national situation

A. Rabies general evaluation

History of the disease and/or infection in the country

At the beginning of the twentieth century, rabies predominantly occurred in Hungary in its urban form and was transmitted to humans mainly by dogs. Therefore, in the 1930's strict animal health regulations were introduced, the main elements of these remained unchanged till recent days. These measures included nationwide mandatory regular vaccination of dogs over three months of age.

During World War II, epidemiological actions were hindered, which resulted in a re-emergence of urban rabies in 1946-47.

The re-introduction of regulatory measures as well as mandatory preventive vaccination, urban rabies seems to be sporadic in Hungary. The register of the annual vaccination of dogs show that around 1.5 Million of dogs are vaccinated every year.

In recent days, together with the disappearing of rabies from dogs, rabies in cats is considered to be of high importance. Preventive vaccination of cats against rabies is recommended but not mandatory and special epidemiological aspects are to be considered. (The movement of animals is hard to control and there is a relative large number of semi-wild living animals of this species.)

Sylvatic rabies reached the North-Eastern part of Hungary in the year 1954. Until 1966 cases remained sporadic (a total of 97 foxes, 16 badgers and wild cats confirmed positive for rabies). In the same timeframe, 35 dogs and 96 domestic cats were confirmed positive for the disease.

In 1967, sylvatic rabies crossed the Danube and by 1971 the whole country was infected. At this time, intensive attempts were executed in order to lower the number of foxes, with minimum results. These actions were suspended in 1987.

Between 1988 and 1996 around 1000 rabies cases in foxes were diagnosed per year. Oral vaccination of foxes was introduced in Hungary in 1992. From that year, the rabies cases in foxes decreased year by year, as the vaccination zone was extended from the western part of the country to the whole territory of Hungary. From 1988, rabies cases in foxes decreased by 90%.

The efficacy of the oral immunization of foxes can be demonstrated by the considerable decrease of rabies cases in the country. During the recent years the number of the detected positive cases remained under ten cases. In the calendar years 2005 only 9, in 2006 only 3, in 2007 only 4, in 2008 only 7 and in 2009 only 2 positive cases could be detected for the whole territory of the country. In 2010 fox rabies cases happened in Hungary: from this 6 cases in county Csongrád (close to the border of the country) and 1 dog in the same county, 1 case in county Hajdú-Bihar, 2 cases in county Szabolcs-Szatmár-Bereg. In 2011 two(2) rabies cases in bats were proved. In 2012 one (1) rabies case was confirmed in bat.

National evaluation of the recent situation, the trends and sources of infection

After a period of lack of rabies cases, in the autumn of 2013 24 cases were diagnosed, mainly in foxes. Therefore the oral vaccination of foxes is continued in a definied part of the country.

Recent actions taken to control the zoonoses

In order to eradicate rabies from Hungary and to protect public health, regulatory measures on domestic animals are in place. Regular preventive vaccination of dogs is mandatory two times between 3 months of age and under 1 year of age with monovalent vaccine. Later repeated annually. Stray dogs are removed from public areas and are vaccinated against the disease. Oral vaccination of foxes is done in a part of Hungary's territory.

Hungary - 2013 Report on trends and sources of zoonoses

2.11.2 Rabies in humans

A. Rabies in humans

Reporting system in place for the human cases

There are around 80 communicable diseases notifiable in Hungary based on legal background. The physician (in primary health care, specialist care, inpatient medical institution or pathology) who first diagnoses a case of a notifiable communicable disease (even the suspicion of the disease!) immediately reports data of case to the first level of the epidemiological network (municipal institute) of National Public Health and Medical Officer's Service (NPHMOS). The suspicion of the human lyssa is obligatory to be reported immediately also by telephone. Data must be reported both at the beginning and at end of the illness (recovery/death, result of laboratory test). The NPHMOS has a nationwide electronic system for registering and analysing data of communicable diseases in a combined national database, so the system provides online connection between the three levels (municipal, regional and national level – National Centre of Epidemiology - NCE) of the organization. The NCE prepares reports regularly (weekly, monthly, yearly) to the Chief Medical Officer, the MoH and the Hungarian Central Statistical Office.

Case definition

Confirmed case: Clinical picture compatible with human lyssa and the antigen/genetic material/specific antibodies are identified or viruses have been isolated from appropriate sample.

Suspected case: Clinical picture compatible with human lyssa and the patient has anamnestic data about exposure by a rabies suspected animal

Diagnostic/analytical methods used

The identification of the virus in vivo from cornea imprint of the patient by immunofluorescence method, or to determine the specific antibody titre of the blood or liquor by immunofluorescence method during the second week of the illness. Post mortem: detection of the Negri-body in the brain tissue, or the antigen by immunofluorescence method, or identification of the viral genetic material by PCR, or isolation of the virus in mouse.

Notification system in place

Human cases have been notifiable since 1950 in Hungary, injury suspected to lyssa-infection has been notifiable since 1964. The physician reports data of case on a "case report form" by mail to the municipal institute of NPHMOS. The suspicion of the human lyssa is obligatory to be reported immediately also by telephone. The specialist of the institute records data immediately in the electronic system of the NPHMOS.

History of the disease and/or infection in the country

Human cases have been notifiable since 1950 in Hungary, injury suspect to human lyssa-infection has been notifiable since 1964. 8 human lyssa cases have been reported since 1950 in Hungary. Seven cases were indigenous; only one case was presumably imported from Africa. Cat was the source of infection in four of the cases, fox in two cases, and one case was caused by a dog. The origin of the imported case remained unknown. The vaccine based on brain-extract was used for post exposure prophylaxis in Hungary until 1989. Since then the cell cultured vaccine has been used. The change in the vaccine used and not in the epidemiological situation of lyssa is reflected in the statistics of vaccinated persons (1985 – 1988.: 2000 – 3000 person vaccinated/year, 1994 – 1998. 8000 – 10 500/year, 1999 - 2003.: 9 500 – 11 000/year).

National evaluation of the recent situation, the trends and sources of infection

Hungary - 2013 Report on trends and sources of zoonoses

No human lyssa case has been registered since 1994 in Hungary.

2.11.3 Lyssavirus (rabies) in animals

A. Rabies in dogs

Vaccination policy

Obligatory vaccination of dogs, once a year.

Measures in case of the positive findings or single cases

There were no positive cases since 2010 (in dogs).

Notification system in place

Notifiable disease.

Table Rabies in animals

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Region	Units tested	Total units positive for Lyssavirus (rabies)	Rabies virus (RABV)	EBLV-1
Cattle (bovine animals)	NFCSO VDD	Unspecified		animal sample		Animal		26	2	2	0
Sheep	NFCSO VDD	Unspecified		animal sample		Animal		21	0	0	0
Goats	NFCSO VDD	Unspecified		animal sample		Animal		5	0	0	0
Foxes - wild - Monitoring	NFCSO VDD	Objective sampling		animal sample		Animal		1799	2	2	0
Bats - wild - Unknown - Clinical investigations	NFCSO VDD	Suspect sampling		animal sample		Animal		3	0	0	0
Bats - zoo animal - Zoo - Unspecified	NFCSO VDD	Unspecified		animal sample		Animal		10	0	0	0
Cats	NFCSO VDD	Unspecified		animal sample		Animal		350	0	0	0
Deer - wild - red deer - Unspecified	NFCSO VDD	Suspect sampling		animal sample		Animal		17	0	0	0
Dogs	NFCSO VDD	Unspecified		animal sample		Animal		240	0	0	0
Foxes - wild - Monitoring - passive	NFCSO VDD	Suspect sampling		animal sample		Animal		1382	20	20	0
Jackals - wild	NFCSO VDD	Suspect sampling		animal sample		Animal		27	0	0	0

	EBLV-2	Lyssavirus (unspecified virus)
Cattle (bovine animals)	0	0
Sheep	0	0
Goats	0	0

Table Rabies in animals

	EBLV-2	Lyssavirus (unspecified virus)
Foxes - wild - Monitoring	0	0
Bats - wild - Unknown - Clinical investigations	0	0
Bats - zoo animal - Zoo - Unspecified	0	0
Cats	0	0
Deer - wild - red deer - Unspecified	0	0
Dogs	0	0
Foxes - wild - Monitoring - passive	0	0
Jackals - wild	0	0

2.12 STAPHYLOCOCCUS INFECTION

2.12.1 General evaluation of the national situation

2.12.2 Staphylococcus in animals

Table Staphylococcus in Animals

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Sample weight	Units tested	Total units positive for Staphylococc us	S. aureus, meticillin resistant (MRSA)	S. aureus, meticillin resistant (MRSA) - spa -type t011
Gallus gallus (fowl) - unspecified	NECSO VDD 1	Unspecified	Not	animal		Animal		79	68	0	0
			applicable	sample							-
Pigs - unspecified	NECSO VDD	DD Unspecified	Not	animal		Animal		7	3	0	0
rigs - unspecified	IVI COO VDD	Orispedifica	applicable	sample		Allillai		,	3	O	
Sheep	NECCO VIDIO	NECSO VDD	Unspecified	Not	animal	Animal		3	2	0	0
Sheep IN C3O VD	INI COO VDD	Orispecified	applicable	sample		Allillai		3		U	
Turkeys - unspecified NFCSO VDD	NECCO VDD	Linenseified	Not	animal		A mine al		148	101	0	0
	NECSO VDD	Unspecified	applicable	sample		Animal		140	101	0	0

	S. aureus, meticillin resistant (MRSA) - spa -type t108	S. aureus, meticillin resistant (MRSA) - spa -type t034	S. aureus, meticillin resistant (MRSA) - MRSA, unspecified	S. aureus
Gallus gallus (fowl) - unspecified	0	0	0	68
Pigs - unspecified	0	0	0	3
Sheep	0	0	0	2
Turkeys - unspecified	0	0	0	101

Table Staphylococcus in Animals

Footnote:

None of S. aureus was MRSA.

2.13 Q-FEVER

2.13.1 General evaluation of the national situation

A. Coxiella burnetii (Q-fever) general evaluation

National evaluation of the recent situation, the trends and sources of infection

In May 2013 a human Q fever epidemic occurred in Baranya county. The investigation carried out in cooperation of the human and animal health authorities identified as possible source of the disease a sheep farm. During the investigation 1379 tests were carried out from samples taken in sheep, goat and cattle farms in the area around the farm. 72 bovine, 1 caprine and 34 ovine samples were positive.

Additional information

Diagnostic methods: Complement fixation test (CFT) and immunohistochemical test

2.13.2 Coxiella (Q-fever) in animals

Table Coxiella burnetii (Q fever) in animals

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Analytical Method	Sampling unit	Units tested	Total units positive for Coxiella (Q- fever)	C. burnetii	No of clinically affected herds
Cattle (bovine animals) - Farm - Unspecified	NFCSO VDD	Suspect sampling	Official sampling	animal sample > blood	Domestic	PCR	Animal	55	0	0	0
Cattle (bovine animals) - Farm - Unspecified	NFCSO VDD	Suspect sampling	Official sampling	animal sample > blood	Domestic	Immuno Histo Chemistry (IHC)	Animal	5	0	0	0
Cattle (bovine animals) - Farm - Unspecified	NFCSO VDD	Suspect sampling	Official sampling	animal sample > blood	Domestic	Isolation (Cell, Egg, Mouse)	Animal	13	0	0	0
Cattle (bovine animals) - Farm - Unspecified	NFCSO VDD	Suspect sampling	Official sampling	animal sample > milk	Domestic	PCR	Animal	86	4	4	0
Cattle (bovine animals) - Farm - Unspecified	NFCSO VDD	Suspect sampling	Official sampling	animal sample > blood	Domestic	Complement fixation test (CFT)	Animal	542	53	53	0
Cattle (bovine animals) - Farm - Unspecified	NFCSO VDD	Suspect sampling	Official sampling	animal sample > blood	Domestic	ELISA	Animal	103	35	35	0
Cattle (bovine animals) - Farm - Unspecified	NFCSO VDD	Suspect sampling	Official sampling	animal sample > faeces	Domestic	PCR	Animal	55	0	0	0
Sheep and goats - Farm - Unspecified	NFCSO VDD	Suspect sampling	Official sampling	animal sample > blood	Domestic	Complement fixation test (CFT)	Animal	298	19	19	0
Sheep and goats - Farm - Unspecified	NFCSO VDD	Suspect sampling	Official sampling	animal sample > blood	Domestic	PCR	Animal	59	0	0	0
Sheep and goats - Farm - Unspecified	NFCSO VDD	Suspect sampling	Official sampling	animal sample	Domestic	Immuno Histo Chemistry (IHC)	Animal	3	0	0	0

Table Coxiella burnetii (Q fever) in animals

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Analytical Method	Sampling unit	Units tested	Total units positive for Coxiella (Q- fever)	C. burnetii	No of clinically affected herds
Sheep and goats - Farm - Unspecified	NFCSO VDD	Suspect sampling	Official sampling	animal sample > blood	Domestic	ELISA	Animal	128	31	31	0
Sheep and goats - Farm - Unspecified	NFCSO VDD	Suspect sampling	Official sampling	environmenta I sample > dust	Domestic	PCR	Holding	161	112	112	0
Sheep and goats - Farm - Unspecified	NFCSO VDD	Suspect sampling	Official sampling	animal sample > milk	Domestic	PCR	Animal	33	4	4	0

Footnote:

Both animal and environmental samples were taken.

In the column "Total units positive for Q-fever" all the units positive are shown, altough a part of them originate from the same sample (to be tested with different analytical methods). 107 animals were positive, 72 bovine, 1 caprine and 34 ovine.

2.14 WEST NILE VIRUS INFECTIONS

2.14.1 General evaluation of the national situation

2.14.2 West Nile Virus in animals

A. West Nile Virus in Animals

Vaccination policy

In case of equine animals vaccination for West Nile Virus is on a voluntary basis.

Notification system in place

In case of animals West Nile Virus is not a notifiable disease.

Additional information

In 2004 goshawks in Hungary (Accipiter gentilis) showed symptoms of lethal encephalitis. West Nile virus nucleic acid and antigens were detected in the brain of the animals. The complete genome analysis indicated that the strain belonged to the lineage 2 of WNV. The same lineage was detected in 2005 in four goshawks and one sparrowhawk. Furthermore in 2007 the virus was detected in geese and in red-footed falcons as well. The first human case was confirmed in 2008.

Table West Nile Virus in Animals

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Vaccination status	Analytical Method	Sampling unit	Region	Units tested	Total units positive for West Nile Virus
Corvids, unspecified - wild - Natural habitat	NFCSO VDD	Suspect sampling	Not applicable	animal sample > blood	Domestic			Animal		1	1
Eagle - wild - Natural habitat	NFCSO VDD	Suspect sampling	Not applicable	animal sample > blood	Domestic		Immuno Histo Chemistry (IHC)	Animal		1	0
Falcons - wild - Natural habitat	NFCSO VDD	Suspect sampling	Not applicable	animal sample > blood	Domestic			Animal		4	1
Geese - unspecified - Farm	NFCSO VDD	Suspect sampling	Not applicable	animal sample > brain	Domestic		PCR	Animal		1	0
Owls - zoo animals - Zoo	NFCSO VDD	Suspect sampling	Not applicable	animal sample > brain	Domestic			Animal		1	1
Passeriformes, unspecified - wild - Natural habitat	NFCSO VDD	Suspect sampling	Not applicable	animal sample > blood	Domestic		Immuno Histo Chemistry (IHC)	Animal		2	0
Penguin - zoo animals - Zoo	NFCSO VDD	Suspect sampling	Not applicable	animal sample	Domestic		Immuno Histo Chemistry (IHC)	Animal		1	0
Solipeds, domestic - donkeys - Farm	NFCSO VDD	Suspect sampling	Not applicable	animal sample > blood	Domestic		ELISA	Animal		1	0
Solipeds, domestic - horses - Farm	NFCSO VDD	Suspect sampling	Not applicable	animal sample > blood	Domestic			Animal		25	1

3. INFORMATION ON SPECIFIC INDICATORS OF ANTIMICROBIAL RESISTANCE

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3.1 ESCHERICHIA COLI, NON-PATHOGENIC

- 3.1.1 General evaluation of the national situation
- 3.1.2 Antimicrobial resistance in Escherichia coli, non-pathogenic

Table Antimicrobial susceptibility testing of E. coli in Meat from bovine animals

Escherichia coli, non- pathogenic	patho	, non- genic, ecified				
Isolates out of a monitoring program (yes/no)	yes					
Number of isolates available in the laboratory	2	3				
Antimicrobials:	N	n				
Aminoglycosides - Gentamicin	23	0				
Aminoglycosides - Streptomycin	23	6				
Amphenicols - Chloramphenicol	23	4				
Cephalosporins - 3rd generation cephalosporins	23	0				
Fluoroquinolones - Ciprofloxacin	23	0				
Penicillins - Ampicillin	23	10				
Quinolones - Nalidixic acid	23	0				
Sulfonamides	23	6				
Tetracyclines - Tetracycline	23	8				
Trimethoprim	23	2				
Fully sensitive	23	11				
Resistant to 1 antimicrobial	23	3				
Resistant to 2 antimicrobials	23	3				

Table Antimicrobial susceptibility testing of E. coli in Meat from bovine animals

	Escherichia coli, non- pathogenic								
t I	yes								
1 i	23								
Antimicrobia	als:	Ν	n						
Resistant to 3 antin	nicrobials	23	1						
Resistant to 4 antin	nicrobials	23	2						
Resistant to >4 ant	23	3							

Table Antimicrobial susceptibility testing of E. coli in Meat from pig

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Escherichia coli, non- pathogenic	E.coli, non- pathogenic, unspecified						
Isolates out of a monitoring program (yes/no)	yes						
Number of isolates available in the laboratory	12						
Antimicrobials:	N	n					
Aminoglycosides - Gentamicin	12	0					
Aminoglycosides - Streptomycin	12	3					
Amphenicols - Chloramphenicol	12	2					
Cephalosporins - 3rd generation cephalosporins	12	0					
Fluoroquinolones - Ciprofloxacin	12	3					
Penicillins - Ampicillin	12	6					
Quinolones - Nalidixic acid	12	1					
Sulfonamides	12	3					
Tetracyclines - Tetracycline	12	6					
Trimethoprim	12	5					
Fully sensitive	12	4					
Resistant to 1 antimicrobial	12	1					
Resistant to 2 antimicrobials	12	1					
Resistant to 3 antimicrobials	12	2					
Resistant to 4 antimicrobials	12	1					
Resistant to >4 antimicrobials	12	3					

Table Antimicrobial susceptibility testing of E. coli in Meat from broilers (Gallus gallus)

Escherichia coli, non- pathogenic	E.coli, non- pathogenic, unspecified						
Isolates out of a monitoring program (yes/no)	yes						
Number of isolates available in the laboratory	46						
Antimicrobials:	N	n					
Aminoglycosides - Gentamicin	46	4					
Aminoglycosides - Streptomycin	46	11					
Amphenicols - Chloramphenicol	46	5					
Cephalosporins - 3rd generation cephalosporins	46	4					
Fluoroquinolones - Ciprofloxacin	46	37					
Penicillins - Ampicillin	46	34					
Quinolones - Nalidixic acid	46	35					
Sulfonamides	46	20					
Tetracyclines - Tetracycline	46	27					
Trimethoprim	46	11					
Fully sensitive	46	2					
Resistant to 1 antimicrobial	46	2					
Resistant to 2 antimicrobials	46	7					
Resistant to 3 antimicrobials	46	5					
Resistant to 4 antimicrobials	46	12					
Resistant to >4 antimicrobials	46	18					

Footnote:

One ESBL strain was detected.

E.coli, non-Escherichia coli, nonpathogenic, pathogenic unspecified Isolates out of a monitoring program (yes/no) Number of isolates available 9 in the laboratory Antimicrobials: Aminoglycosides - Gentamicin 9 0 9 Aminoglycosides - Streptomycin 2 Amphenicols - Chloramphenicol Cephalosporins - 3rd generation cephalosporins Fluoroquinolones - Ciprofloxacin 6 Penicillins - Ampicillin 9 6 9 Quinolones - Nalidixic acid 9 4 Sulfonamides Tetracyclines - Tetracycline 9 5 9 Trimethoprim 9 2 Resistant to 1 antimicrobial Resistant to 2 antimicrobials 9 9 Resistant to 3 antimicrobials 9 2 Resistant to 4 antimicrobials

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Table Antimicrobial susceptibility testing of E. coli in Meat from other poultry species

Resistant to >4 antimicrobials

Table Antimicrobial susceptibility testing of E.coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

E.coli, non-pathogenic, unspecified		Meat from broilers (Gallus gallus)																								
Isolates out of a monitoring program (yes/no) Number of isolates available		yes 946																								
in the laboratory Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	46	4								7	7	17	9	2	4										
Aminoglycosides - Streptomycin	16	46	11												1	13	16	5	2	1	2	6				
Amphenicols - Chloramphenicol	16	46	5												8	25	8			2	3					
Cephalosporins - Cefotaxime	0.25	46	4								40	2	1	1	2											
Fluoroquinolones - Ciprofloxacin	0.064	46	37						8	1	2	11	3	3	1	3	14									
Penicillins - Ampicillin	8	46	34											1	4	5	2	1			33					
Quinolones - Nalidixic acid	16	46	35											3	3	2	1	2	1	2	8	24				
Sulfonamides	256	46	20												1	5	2	8	8	2			3	15	2	
Tetracyclines - Tetracycline	8	46	27											11	7	1				8	17	2				
Trimethoprim	2	46	11								3	13	17	2				11								

E.coli, n	broilers	from (Gallus lus)			
	yes				
	46				
Antimicrob	oials:	lowest	highest		
Aminoglycosides	s - Gentamicin				
Aminoglycosides	s - Streptomycin				
Amphenicols - C					

Table Antimicrobial susceptibility testing of E.coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

E.coli, n	Meat broilers gal	(Gallus	
	ye	es	
	4	6	
Antimicrob	oials:	lowest	highest
Cephalosporins	- Cefotaxime		
Fluoroquinolones	s - Ciprofloxacin		
Penicillins - Amp	icillin		
Quinolones - Nal	idixic acid		
Sulfonamides			
Tetracyclines - T			
Trimethoprim			

Table Antimicrobial susceptibility testing of E.coli, non-pathogenic, unspecified in Meat from turkey - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

							(р.,	3, , , ,		250.0	******	5011		011 01 11												
E.coli, non-pathogenic, unspecified		Meat from turkey																								
Isolates out of a monitoring program (yes/no)		yes																								
Number of isolates available in the laboratory														9												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	9	0									4	5													
Aminoglycosides - Streptomycin	16	9	2												1	4	2		1		1					
Amphenicols - Chloramphenicol	16	9	1												2	5	1				1					
Cephalosporins - Cefotaxime	0.25	9	1								8		1													
Fluoroquinolones - Ciprofloxacin	0.064	9	6						2	1	1	1	3		1											
Penicillins - Ampicillin	8	9	6											1	1		1				6					
Quinolones - Nalidixic acid	16	9	4											1	1	1	1	1		1	1	2				
Sulfonamides	256	9	4													2			3				3	1		
Tetracyclines - Tetracycline	8	9	5											4						4	1					
Trimethoprim	2	9	1									4	4					1								

E.coli, n unspeci	Meat from turkey				
	ye	es			
	9				
Antimicro	bials:	lowest	highest		
Aminoglycoside	s - Gentamicin				
Aminoglycoside	s - Streptomycin				
Amphenicols - 0					

Table Antimicrobial susceptibility testing of E.coli, non-pathogenic, unspecified in Meat from turkey - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

E.coli, n	Meat from turkey					
	yes					
	ç	9				
Antimicrol	bials:	lowest	highest			
Cephalosporins	- Cefotaxime					
Fluoroquinolone	s - Ciprofloxacin					
Penicillins - Amp	picillin					
Quinolones - Na	lidixic acid					
Sulfonamides						
Tetracyclines - T						
Trimethoprim						

Table Antimicrobial susceptibility testing of E.coli, non-pathogenic, unspecified in Meat from bovine animals - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

							(р.,	9, , , , , , ,	G11110 C1	0. 10010	1100 1111	1 4 0011	301111 (411	011 01 11		. oqua.										
E.coli, non-pathogenic, unspecified		Meat from bovine animals																								
Isolates out of a monitoring program (yes/no)		yes																								
Number of isolates available in the laboratory													2	3												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	2	23	0								2	3	11	7												
Aminoglycosides - Streptomycin	16	23	6												1	5	8	3	2	1	2	1				
Amphenicols - Chloramphenicol	16	23	4												5	12	2			3	1					
Cephalosporins - Cefotaxime	0.25	23	0								22	1														
Fluoroquinolones - Ciprofloxacin	0.064	23	0						23																	
Penicillins - Ampicillin	8	23	10													5	8	2			8					
Quinolones - Nalidixic acid	16	23	0											1	15	7										
Sulfonamides	256	23	5												1	5	3	2	6			1	1	4		
Tetracyclines - Tetracycline	8	23	8											7	7	1				3	4	1				
Trimethoprim	2	23	2									8	11	1	1			2								

E.coli, n unspecif		from animals			
	yes				
	23				
Antimicro	oials:	lowest	highest		
Aminoglycosides	s - Gentamicin				
Aminoglycosides	s - Streptomycin				
Amphenicols - C					

Table Antimicrobial susceptibility testing of E.coli, non-pathogenic, unspecified in Meat from bovine animals - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

	<u> </u>										
E.coli, n unspecit	on-pathogenic, fied		from animals								
	Isolates out of a monitoring program (yes/no)	ye	es								
	Number of isolates available in the laboratory ntimicrobials: phalosporins - Cefotaxime										
Antimicrol	in the laboratory ntimicrobials:										
Cephalosporins	<u>'</u>										
Fluoroquinolone	Dephalosporins - Cefotaxime										
Penicillins - Amp	icillin										
Quinolones - Na	lidixic acid										
Sulfonamides											
Tetracyclines - T	etracycline										
Trimethoprim											

Table Antimicrobial susceptibility testing of E.coli, non-pathogenic, unspecified in Meat from pig - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

E.coli, non-pathogenic, unspecified		Meat from pig																								
Isolates out of a monitoring program (yes/no)													y	es												
Number of isolates available in the laboratory		12																								
Antimicrobials:	Cut-off value																									
Aminoglycosides - Gentamicin	2	12 0 5 2 5																								
Aminoglycosides - Streptomycin	16	12	3													5	4		1	1		1				
Amphenicols - Chloramphenicol	16	 																								
Cephalosporins - Cefotaxime	0.25	12	0								11	1														
Fluoroquinolones - Ciprofloxacin	0.064	12	3						8	1	2		1													
Penicillins - Ampicillin	8	12	6											1	2	3					6					
Quinolones - Nalidixic acid	16	12	1											4	3	3	1					1				
Sulfonamides	256	12	3													1	4	3	1				1	2		
Tetracyclines - Tetracycline	8	12	6											4	2					5	1					
Trimethoprim	2	12	5								1	2	3	1				5								

	Inspecified Isolates out of a monitoring program (yes/no) Number of isolates available in the laboratory Itimicrobials:							
	•	ye	es					
	program (yes/no) Number of isolates availabl		2					
Antimicrob	program (yes/no) Number of isolates available in the laboratory ntimicrobials:							
Aminoglycosides	s - Gentamicin							
Aminoglycosides	s - Streptomycin							
Amphenicols - C	hloramphenicol							

Table Antimicrobial susceptibility testing of E.coli, non-pathogenic, unspecified in Meat from pig - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

E.coli, n	on-pathogenic, ïed	Meat fr	om pig							
	in the laboratory									
	Number of isolates available in the laboratory Antimicrobials:									
Antimicrob	in the laboratory antimicrobials: ephalosporins - Cefotaxime									
Cephalosporins -	ephalosporins - Cefotaxime									
Fluoroquinolones	luoroquinolones - Ciprofloxacin									
Penicillins - Amp	Cephalosporins - Cefotaxime Cluoroquinolones - Ciprofloxacin Cenicillins - Ampicillin									
Quinolones - Nal	idixic acid									
Sulfonamides										
Tetracyclines - T	etracycline									
Trimethoprim										

Table Antimicrobial susceptibility testing of E.coli, non-pathogenic, unspecified in Cattle (bovine animals) - calves (under 1 year) quantitative data [Dilution method]

E.coli, non-pathogenic, unspecified		Cattle (bovine animals) - calves (under 1 year)														1 year)									
Isolates out of a monitoring program (yes/no)																									
Number of isolates available in the laboratory		51																							
Antimicrobials:	Cut-off value	e N N C=0.002 C=0.004 0.008 0.015 0.016 0.03 0.06 0.12 0.25 0.5 1 2 4 8 16 32 64 128 256 512 1024 2048 24096														>4096									
Aminoglycosides - Gentamicin	2	51 1 2 36 12 1																							
Aminoglycosides - Streptomycin	16	5 51 5 9 36 1 2 2 1																							
Amphenicols - Chloramphenicol	16	 																							
Cephalosporins - Cefotaxime	0.25																								
Fluoroquinolones - Ciprofloxacin	0.064	51	2			5	42		2			1					1								
Penicillins - Ampicillin	8	51	1											3	15	29	3		1						
Quinolones - Nalidixic acid	16	51	2													48	1			2					
Tetracyclines - Tetracycline	8	51	5											11	34		1		1	4					
Trimethoprim	2	51	1										47	3					1						
Sulfonamides - Sulfamethoxazole	64	51	11														4	16	9	11	7		4		

E.coli, no unspecif	on-pathogenic, ïed		•					
	Isolates out of a monitoring program (yes/no)							
	Number of isolates available in the laboratory	5	1					
Antimicrob	ntimicrobials:							
Aminoglycosides	- Gentamicin	0.25	32					
Aminoglycosides	- Streptomycin	2	128					
Amphenicols - Cl	hloramphenicol	2	64					

Table Antimicrobial susceptibility testing of E.coli, non-pathogenic, unspecified in Cattle (bovine animals) - calves (under 1 year) quantitative data [Dilution method]

1											
E.coli, non-pathounspecified	ogenic,	anim	under 1								
	Isolates out of a monitoring program (yes/no) Number of isolates available in the laboratory										
	in the laboratory										
Antimicrobials:	ntimicrobials:										
Cephalosporins - Cefotaxime	phalosporins - Cefotaxime										
Fluoroquinolones - Ciprofloxac	· ·										
Penicillins - Ampicillin		0.5	32								
Quinolones - Nalidixic acid		4	64								
Tetracyclines - Tetracycline		1	64								
Trimethoprim		0.5	32								
Sulfonamides - Sulfamethoxaz	cole	8 1024									

Table Antimicrobial susceptibility testing of E.coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers - quantitative data [Dilution method]

	Concentration (pg/m/), names or isolates with a concentration of milibrion equal to																							
E.coli, non-pathogenic, unspecified		Gallus gallus (fowl) - broilers																						
Isolates out of a monitoring program (yes/no)																								
Number of isolates available in the laboratory		152																						
Antimicrobials:	Cut-off value	lue N n <=0.002 <=0.004 0.008 0.015 0.016 0.03 0.06 0.12 0.25 0.5 1 2 4 8 16 32 64 128 256 512 1024 2048 >4096																						
Aminoglycosides - Gentamicin	2	152 3 3 107 37 2 1 2																						
Aminoglycosides - Streptomycin	16	3 152 36 34 77 5 9 4 23																						
Amphenicols - Chloramphenicol	16	 																						
Cephalosporins - Cefotaxime	0.25																							
Fluoroquinolones - Ciprofloxacin	0.064	152	104			3	36		9		5	27	13	15	12	5	27							
Penicillins - Ampicillin	8	152	67											4	36	42	3		67					
Quinolones - Nalidixic acid	16	152	96													49	3	4		96				
Tetracyclines - Tetracycline	8	152	57											26	69			1	1	55				
Trimethoprim	2	2 152 35 113 3 1 35																						
Sulfonamides - Sulfamethoxazole	64	152	51														15	36	27	23		51		

E.coli, n unspeci	on-pathogenic, fied		gallus broilers							
	Isolates out of a monitoring program (yes/no)									
	Number of isolates available in the laboratory	15	52							
Antimicrol	in the laboratory									
Aminoglycosides	s - Gentamicin	0.25	32							
Aminoglycosides	s - Streptomycin	2	128							
Amphenicols - C	chloramphenicol	2	64							

Table Antimicrobial susceptibility testing of E.coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers - quantitative data [Dilution method]

E.coli, n	on-pathogenic, fied		gallus broilers									
	Isolates out of a monitoring program (yes/no)											
	Number of isolates available in the laboratory	15	52									
Antimicro	ntimicrobials:											
Cephalosporins	phalosporins - Cefotaxime											
Fluoroquinolone	uoroquinolones - Ciprofloxacin											
Penicillins - Amp	picillin	0.5	32									
Quinolones - Na	lidixic acid	4	64									
Tetracyclines - T	etracycline	1	64									
Trimethoprim	imethoprim											
Sulfonamides - S	Sulfamethoxazole	8 1024										

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Concentration (µg/m	I), number of isolates with	th a concentration	of inhibition equal to
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E.coli, non-pathogenic, unspecified		Pigs - fattening pigs																							
Isolates out of a monitoring program (yes/no)																									
Number of isolates available in the laboratory													15	52											
Antimicrobials:	Cut-off value	N n <=0.002 <=0.004 0.008 0.015 0.016 0.03 0.06 0.12 0.25 0.5 1 2 4 8 16 32 64 128 256 512 1024 2048 >4096																							
Aminoglycosides - Gentamicin	2	152 3 6 97 43 3 3																							
Aminoglycosides - Streptomycin	16	152 80 3 24 40 5 9 24 47																							
Amphenicols - Chloramphenicol	16	152 27 2 37 81 5 3 24																							
Cephalosporins - Cefotaxime	0.25	152																							
Fluoroquinolones - Ciprofloxacin	0.064	152	14			21	102		15		3	4	2	1		1	3								
Penicillins - Ampicillin	8	152	63											2	34	49	4		63						
Quinolones - Nalidixic acid	16	152	9													140	2	1		9					
Tetracyclines - Tetracycline	8	152	98											12	41		1		4	94					
Trimethoprim	2	152	32										115	5		1			31						
Sulfonamides - Sulfamethoxazole	64	152	62														26	30	28	6	2		60		

E.coli, n unspecif	on-pathogenic, fied	Pigs - fattenin pigs					
	Isolates out of a monitoring program (yes/no)						
	Number of isolates available in the laboratory	1:	52				
Antimicro	oials:	lowest	highest				
Aminoglycosides	s - Gentamicin	0.25	32				
Aminoglycosides	s - Streptomycin	2	128				
Amphenicols - C	hloramphenicol	2	64				

E.coli, non-pathogenic, unspecified	Pigs - fattening pigs					
Isolates out of a monitoring program (yes/no)						
Number of isolates available in the laboratory	15	52				
Antimicrobials:	lowest	highest				
Cephalosporins - Cefotaxime	0.06	4				
Fluoroquinolones - Ciprofloxacin	0.008	8				
Penicillins - Ampicillin	0.5	32				
Quinolones - Nalidixic acid	4	64				
Tetracyclines - Tetracycline	1	64				
Trimethoprim	0.5	32				
Sulfonamides - Sulfamethoxazole	8	1024				

Table Cut-off values used for antimicrobial susceptibility testing of Escherichia coli, non-pathogenic in Animals

Test Method Used	Standard methods used for testing

			Concentration (microg/ml)	Zone diameter (mm)
		Standard	Resistant >	Resistant <=
Aminoglycosides	Gentamicin		2	
	Streptomycin		16	
Amphenicols	Chloramphenicol		16	
Cephalosporins	Cefotaxime		0.25	
	Ceftazidime		0.5	
Fluoroquinolones	Ciprofloxacin		0.064	
Penicillins	Ampicillin		8	
Quinolones	Nalidixic acid		16	
Sulfonamides	Sulfonamides		256	
	Sulfamethoxazole		64	
Tetracyclines	Tetracycline		8	
Trimethoprim	Trimethoprim		2	

Table Cut-off values used for antimicrobial susceptibility testing of Escherichia coli, non-pathogenic in Feed

Test Method Used	Standard methods used for testing

			Concentration (microg/ml)	Zone diameter (mm)
		Standard	Resistant >	Resistant <=
Aminoglycosides	Gentamicin		2	
	Streptomycin		16	
Amphenicols	Chloramphenicol		16	
Cephalosporins	Cefotaxime		0.25	
	Ceftazidime		0.5	
Fluoroquinolones	Ciprofloxacin		0.064	
Penicillins	Ampicillin		8	
Quinolones	Nalidixic acid		16	
Sulfonamides	Sulfonamides		256	
	Sulfamethoxazole		64	
Tetracyclines	Tetracycline		8	
Trimethoprim	Trimethoprim		2	

Table Cut-off values used for antimicrobial susceptibility testing of Escherichia coli, non-pathogenic in Food

Test Method Used	Standard methods used for testing
Broth dilution	EFSA

			Concentration (microg/ml)	Zone diameter (mm)
		Standard	Resistant >	Resistant <=
Aminoglycosides	Gentamicin	EFSA	2	
	Streptomycin	EFSA	16	
Amphenicols	Chloramphenicol	EFSA	16	
Cephalosporins	Cefotaxime	EFSA	0.25	
	Ceftazidime	EFSA	0.5	
Fluoroquinolones	Ciprofloxacin	EFSA	0.064	
Penicillins	Ampicillin	EFSA	8	
Quinolones	Nalidixic acid	EFSA	16	
Sulfonamides	Sulfonamides	EFSA	256	
	Sulfamethoxazole	EFSA	64	
Tetracyclines	Tetracycline	EFSA	8	
Trimethoprim	Trimethoprim	EFSA	2	

3.2 ENTEROCOCCUS, NON-PATHOGENIC

- 3.2.1 General evaluation of the national situation
- 3.2.2 Antimicrobial resistance in Enterococcus, non-pathogenic isolates

Table Antimicrobial susceptibility testing of E. faecalis in Meat from bovine animals - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

E. faecalis							11	, , , , , , , , , , , , , , , , , , ,						ovine an		·										
Isolates out of a monitoring program (yes/no)													у	es												
Number of isolates available in the laboratory													2	21												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	32	21	0												1	1	5	8	6							
Aminoglycosides - Kanamycin		0	0																0							
Aminoglycosides - Streptomycin	512	21	2																3	8	4		4	1	1	
Amphenicols - Chloramphenicol	32	21	0												4	17										
Penicillins - Ampicillin	4	21	0										1	16	4											
Tetracyclines - Tetracycline	4	21	12										7	1		1	2		1	4	5					
Fully sensitive		9	9	9																						
Glycopeptides (Cyclic peptides, Polypeptides) - Vancomycin	4	21	0											10	9	2										
Macrolides - Erythromycin	4	21	0									1	18	2												
Oxazolidines - Linezolid	4	21	0											6	15											
Resistant to 1 antimicrobial		10	10	10																						
Resistant to 2 antimicrobials		2	2	2																						

Table Antimicrobial susceptibility testing of E. faecalis in Meat from bovine animals - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

moat	quantitutivo au	دم رک	natio
E. faeca	lis		t from animals
	Isolates out of a monitoring program (yes/no)	у	es
	Number of isolates available in the laboratory	2	21
Antimicrob	oials:	lowest	highest
Aminoglycosides	- Gentamicin		
Aminoglycosides	- Kanamycin		
Aminoglycosides	- Streptomycin		
Amphenicols - Cl	hloramphenicol		
Penicillins - Ampi	icillin		
Tetracyclines - To	etracycline		
Fully sensitive			
Glycopeptides (C Vancomycin	Cyclic peptides, Polypeptides) -		
Macrolides - Eryt	hromycin		
Oxazolidines - Li	nezolid		
Resistant to 1 an	timicrobial		
Resistant to 2 an	timicrobials		

Table Antimicrobial susceptibility testing of E. faecalis in Meat from broilers (Gallus gallus) - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

E. faecalis							4	<i>3</i> . ,,				Meat fro	m broile	rs (Gallu	ıs gallus	;)										
Isolates out of a monitoring program (yes/no)													y	es												
Number of isolates available in the laboratory													3	15												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	32	35	3													2	12	15	3	1	2					-
Aminoglycosides - Streptomycin	512	35	9																1	6	10	3	6	3	6	
Amphenicols - Chloramphenicol	32	35	0												7	22	5		1							
Penicillins - Ampicillin	4	35	0										7	14	14											
Tetracyclines - Tetracycline	4	35	28									6	1					4	4	7	13					
Fully sensitive		6	6	6																						
Glycopeptides (Cyclic peptides, Polypeptides) - Vancomycin	4	35	2											19	14		2									
Macrolides - Erythromycin	4	35	14								3	1	14	3				1	3	1	7	2				
Oxazolidines - Linezolid	4	35	0											13	22			0	0	0	0	0				
Resistant to 1 antimicrobial		13	13	13																						
Resistant to 2 antimicrobials		6	6	6																						
Resistant to 3 antimicrobials		9	9	9																						
Resistant to 4 antimicrobials		1	1	1																						

Table Antimicrobial susceptibility testing of E. faecalis in Meat from broilers (Gallus gallus) - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

E. faeca	lis	broilers	from (Gallus lus)					
	Isolates out of a monitoring program (yes/no)							
	Number of isolates available in the laboratory	3	5					
Antimicrob	oials:	lowest	highest					
Aminoglycosides	- Gentamicin							
Aminoglycosides	- Streptomycin							
Amphenicols - Ch	nloramphenicol							
Penicillins - Ampi	cillin							
Tetracyclines - Te	etracycline							
Fully sensitive								
Glycopeptides (C Vancomycin	cyclic peptides, Polypeptides) -							
Macrolides - Eryt	hromycin							
Oxazolidines - Lii	nezolid							
Resistant to 1 an	timicrobial							
Resistant to 2 an	timicrobials							
Resistant to 3 an								
Resistant to 4 an	timicrobials							

Table Antimicrobial susceptibility testing of E. faecalis in Meat from turkey - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

								, ,.																		
E. faecalis		Meat from turkey																								
Isolates out of a monitoring program (yes/no)		yes																								
Number of isolates available in the laboratory														6												
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096
Aminoglycosides - Gentamicin	32	6	0														1	5								
Aminoglycosides - Streptomycin	512	6	2																		1	1	2		2	
Amphenicols - Chloramphenicol	32	6	1												1	2	1		1	1						
Penicillins - Ampicillin	4	6	0										2	4												
Tetracyclines - Tetracycline	4	6	6																	1	5					
Glycopeptides (Cyclic peptides, Polypeptides) - Vancomycin	4	6	0											3	1	2										
Macrolides - Erythromycin	4	6	6															1	1	2	2					
Oxazolidines - Linezolid	4	6	0											1	5											
Resistant to 2 antimicrobials		4	4	4																						
Resistant to 3 antimicrobials		1	1	1																						
Resistant to 4 antimicrobials		1	1	1																						

E. faecal	Meat from turkey				
	yes				
	6				
Antimicrob	ials:	lowest	highest		
Aminoglycosides					
Aminoglycosides	- Streptomycin	·			

Table Antimicrobial susceptibility testing of E. faecalis in Meat from turkey - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

9 010111111	ativo data [Bilat				
E. faecali	Meat from turkey				
	yes				
	Number of isolates available in the laboratory	6			
Antimicrobi	lowest	highest			
Amphenicols - Chl					
Penicillins - Ampic					
Tetracyclines - Tet	tracycline				
Glycopeptides (Cy Vancomycin	clic peptides, Polypeptides) -				
Macrolides - Erythi	romycin				
Oxazolidines - Line	ezolid				
Resistant to 2 antii	microbials				
Resistant to 3 antii	microbials				
Resistant to 4 antii	microbials				

Table Antimicrobial susceptibility testing of E. faecalis in Meat from pig - Objective sampling - Official sampling - food sample - meat quantitative data [Dilution method]

E. faecalis		Meat from pig																									
Isolates out of a monitoring program (yes/no)		Meat from pig yes Hungary																									
Number of isolates available in the laboratory														6													- 20
Antimicrobials:	Cut-off value	N	n	<=0.002	<=0.004	0.008	0.015	0.016	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>4096	C
Aminoglycosides - Gentamicin	32	6	0											1		2	1	1	1								Zebor
Aminoglycosides - Streptomycin	512	6	0														1			2	1	1	1				9
Amphenicols - Chloramphenicol	32	6	0												5	1											uellas
Penicillins - Ampicillin	4	6	0									1	2	1	2												S
Tetracyclines - Tetracycline	4	6	3										3						2		1						allo
Fully sensitive		3	3	3																							Sources
Glycopeptides (Cyclic peptides, Polypeptides) - Vancomycin	4	6	0											5	1												S
Macrolides - Erythromycin	4	6	0										5		1												
Oxazolidines - Linezolid	4	6	0											2	4												200110868
Resistant to 1 antimicrobial		3	3	3																							Ċ.

E. faeca	Meat from pig				
	yes				
	6				
Antimicrol	oials:	lowest	highest		
Aminoglycosides	s - Gentamicin				
Aminoglycosides	s - Streptomycin				
Amphenicols - C	Chloramphenicol				

Table Antimicrobial susceptibility testing of E. faecalis in Meat from pig - Objective sampling - Official sampling - food sample - meat - quantitative data [Dilution method]

E. faeca	Meat from pig				
	уe	es			
	6				
Antimicrol	lowest	highest			
Penicillins - Amp					
Tetracyclines - T	etracycline				
Fully sensitive					
Glycopeptides (0 Vancomycin	Cyclic peptides, Polypeptides) -				
Macrolides - Ery	thromycin				
Oxazolidines - L	inezolid				
Resistant to 1 ar	ntimicrobial				

Table Cut-off values for antibiotic resistance of E. faecalis in Animals

Test Method Used	Standard methods used for testing

			Concentration (microg/ml)	Zone diameter (mm)
		Standard	Resistant >	Resistant <=
Aminoglycosides	Gentamicin		32	
	Streptomycin		512	
Amphenicols	Chloramphenicol		32	
Fluoroquinolones	Ciprofloxacin		4	
Glycopeptides (Cyclic peptides, Polypeptides)	Vancomycin		4	
Macrolides	Erythromycin		4	
Oxazolidines	Linezolid		4	
Penicillins	Ampicillin		4	
Tetracyclines	Tetracycline		4	

Table Cut-off values for antibiotic resistance of E. faecalis in Feed

Test Method Used	Standard methods used for testing

			Concentration (microg/ml)	Zone diameter (mm)
		Standard	Resistant >	Resistant <=
Aminoglycosides	Gentamicin		32	
	Streptomycin		512	
Amphenicols	Chloramphenicol		32	
Fluoroquinolones	Ciprofloxacin		4	
Glycopeptides (Cyclic peptides, Polypeptides)	Vancomycin		4	
Macrolides	Erythromycin		4	
Oxazolidines	Linezolid		4	
Penicillins	Ampicillin		4	
Tetracyclines	Tetracycline		4	

Table Cut-off values for antibiotic resistance of E. faecalis in Food

Test Method Used	Standard methods used for testing

			Concentration (microg/ml)	Zone diameter (mm)
		Standard	Resistant >	Resistant <=
Aminoglycosides	Gentamicin		32	
	Streptomycin		512	
Amphenicols	Chloramphenicol		32	
Fluoroquinolones	Ciprofloxacin		4	
Glycopeptides (Cyclic peptides, Polypeptides)	Vancomycin		4	
Macrolides	Erythromycin		4	
Oxazolidines	Linezolid		4	
Penicillins	Ampicillin		4	
Tetracyclines	Tetracycline		4	

Table Cut-off values for antibiotic resistance of E. faecium in Animals

Test Method Used	Standard methods used for testing

			Concentration (microg/ml)	Zone diameter (mm)
		Standard	Resistant >	Resistant <=
Aminoglycosides	Gentamicin		32	
	Streptomycin		128	
Amphenicols	Chloramphenicol		32	
Fluoroquinolones	Ciprofloxacin		4	
Glycopeptides (Cyclic peptides, Polypeptides)	Vancomycin		4	
Macrolides	Erythromycin		4	
Oxazolidines	Linezolid		4	
Penicillins	Ampicillin		4	
Streptogramins	Quinupristin/Dalfopristin		1	
Tetracyclines	Tetracycline		4	

Table Cut-off values for antibiotic resistance of E. faecium in Feed

Test Method Used	Standard methods used for testing

			Concentration (microg/ml)	Zone diameter (mm)
		Standard	Resistant >	Resistant <=
Aminoglycosides	Gentamicin		32	
	Streptomycin		128	
Amphenicols	Chloramphenicol		32	
Fluoroquinolones	Ciprofloxacin		4	
Glycopeptides (Cyclic peptides, Polypeptides)	Vancomycin		4	
Macrolides	Erythromycin		4	
Oxazolidines	Linezolid		4	
Penicillins	Ampicillin		4	
Streptogramins	Quinupristin/Dalfopristin		1	
Tetracyclines	Tetracycline		4	

Table Cut-off values for antibiotic resistance of E. faecium in Food

Test Method Used	Standard methods used for testing

			Concentration (microg/ml)	Zone diameter (mm)
		Standard	Resistant >	Resistant <=
Aminoglycosides	Gentamicin		32	
	Streptomycin		128	
Amphenicols	Chloramphenicol		32	
Fluoroquinolones	Ciprofloxacin		4	
Glycopeptides (Cyclic peptides, Polypeptides)	Vancomycin		4	
Macrolides	Erythromycin		4	
Oxazolidines	Linezolid		4	
Penicillins	Ampicillin		4	
Streptogramins	Quinupristin/Dalfopristin		1	
Tetracyclines	Tetracycline		4	

4. INFORMATION ON SPECIFIC MICROBIOLOGICAL AGENTS

4.1 CRONOBACTER

4.1.1 General evaluation of the national situation

4.1.2 Cronobacter in foodstuffs

Table Cronobacter in food

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Sample weight	Units tested	Total units positive for Cronobacter	Cronobacter sakazakii	Cronobacter spp, unspecified
Infant formula - dried - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	10 g	114	0	0	0

4.2 HISTAMINE

4.2.1 General evaluation of the national situation

4.2.2 Histamine in foodstuffs

Table Histamine in food

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Sample weight	Units tested	Total units in non-conformity	<= 100 mg/kg	>100 - <= 200 mg/kg
Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Retail - Surveillance	NFCSO	Objective sampling	Official sampling	food sample		Single	5 g	43	0	43	0
Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Catering	NFCSO	Suspect sampling	Official sampling	food sample		Single	5 g	1	0	1	0

	>200 - <= 400 mg/kg	> 400 mg/kg
Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Retail - Surveillance	0	0
Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Catering	0	0

4.3 STAPHYLOCOCCAL ENTEROTOXINS

4.3.1 General evaluation of the national situation

4.3.2 Staphylococcal enterotoxins in foodstuffs

A. Staphylococcal enterotoxins in foodstuffs

Monitoring system

Sampling strategy

There is no direct sampling strategy, samples containing more than 100.000 coagulase positive staphyloccocci/gram are tested for the presence of enterotoxin.

Only those product groups are routinely tested for coagulase positive staphyloccocci, for which there is a criterion in 2073/2005/EC.

Type of specimen taken

milk products, in case of supposed human cases other food as meat products, prepared dishes are sampled

Definition of positive finding

If ELFA test shows a positive result, the product is considered to be positive.

Diagnostic/analytical methods used

Validated detection method of the EU-RL based on VIDAS enterotoxin test is used.

Measures in case of the positive findings or single cases

products are withdrawn from the market.

Table Staphylococcal enterotoxins in food

	Source of information	Sampling strategy	Sampler	Sample type	Sample origin	Sampling unit	Sample weight	Units tested	Total units positive for Staphylococc al enterotoxins
Meat from bovine animals and pig - minced meat - Processing plant - Clinical investigations	NFCSO	Suspect sampling	Official sampling	food sample	Domestic	Batch	75 g	2	0
Meat from pig - meat products - Retail - Clinical investigations	NFCSO	Suspect sampling	Official sampling	food sample		Single	25 g	5	0
Other processed food products and prepared dishes - Catering - Clinical investigations	NFCSO	Suspect sampling	Official sampling	food sample	Domestic	Single	25 g	11	0

Comments:

¹⁾ 3 * 25 g

5. FOODBORNE

Foodborne outbreaks are incidences of two or more human cases of the same disease or infection where the cases are linked or are probably linked to the same food source. Situation, in which the observed human cases exceed the expected number of cases and where a same food source is suspected, is also indicative of a foodborne outbreak.

A. Foodborne outbreaks

System in place for identification, epidemological investigations and reporting of foodborne outbreaks

Data on foodborne outbreaks have been collected in Hungary by legal background at the Public Health Authority since 1931. There are two surveillance systems in Hungary since 1st January 2007. One of them is for collection of communicable diseases included the human data of foodborne outbreaks (based on the obligatory reports of a physician and microbiological laboratories). The reporting system of human cases belongs to the institutes of the National Public Health and Medical Officers' Service (National Center for Epidemiologie = NCE and National Institute for Food and Nutrition Science = NIFNS). The other surveillance system is operated by the Central Agricultural Office, (since 15 March 2012 it's name is National Food Chain Safety Office = NFCSO), which is working under the supervision of Ministry of Agriculture. This system based on the reports of the food business operators, the drinking water suppliers and the data of the communicable disease reporting system. The role of the NFCSO is in this topic to investigate which food was the sourse of the outbreaks, collection and analysis of obtained data – in all events if the outbreak was general or the supposed product is produced by the food industry and/or catering, and not located to a household. The household outbreaks are investigated by the Public Health Authority. The investigation of an outbreak is usually initiated with the information about the human cases provided by the public health service. The two authorities cooperate in the whole process of investigation.

Description of the types of outbreaks covered by the reporting:

Outbreak: At least two cases with epidemiological link (exposed by the same food).

Household outbreak: At least two related cases in the same household.

General outbreak: At least two related cases in a community (school, kindergarten, hospital, events etc.).

National evaluation of the reported outbreaks in the country:

Trends in numbers of outbreaks and numbers of human cases involved

Altogether there were 119 general and household outbreaks verified as foodborne in 2013 (2012:114) in Hungary. 1145 cases (2012: 1414) were linked to the outbreaks, among them 136 (11,9%) hospitalised cases (2012: 206 (14,6%). Nobody died. Although the number of the outbreakes did not changed significantly and the number of cases and the rate of hospitalisation decreased.

The surveillance based on results of laboratories and the reports of physitians. The epidemiological investigation was carred out by Public Health Services. If it has been suspected the outbreake was foodborne, the investigation at the food chain was conducted by National Food Chain Safety Office.

The number of foodborne outbreaks registrated by National Food Chain Safety Office was less than in 2012, but the number of cases increased compared to the previous year. In 2013 there were 26 general food-borne events, there were 1023 human cases.

The proportion of causative agents:

26,9 % (7) of the outbreaks were caused by Salmonella enteritidis, 15,4 % (4) Norovirus, 3,8 % (1) Clostridium perfringens, 3,8 % (1) Staphylococcus aureus, 19,2 % (5) high microbial count and 30,8 % (8) outbreaks had unknown etiology. The proportion of Salmonella etiology have fallen compared to 2012 (2013: 26,9 %, 2012: 42,9%). 1/3 of the outbreaks were caused by facultative agents.

There was no major change in the type of food vehicles. The most foodborne outbreaks (65,4 %) were caused by mixed foods. A number of cases were caused by broiler meats and products thereof, decreased. Only one outbreaks caused by egg products in restaurants.

The most food borne events occurred in public canteens and the number of events decreased compared to 2012 (2013: 57,7 %, 2012: 58,1 %). 42,3 % of the outbreaks occured in catering services (restaurant, bar, cafe, etc.), the number of cases slumped compared to the previous year. We did not register outbreaks which were caused by the food industry or by small producers' products.

Table Foodborne Outbreaks: summarised data

	Weak	evidence or n	o vehicle outb	oreaks		
	Number of outbreaks	Human cases	Hospitalized	Deaths	Strong evidence Number of Outbreaks	Total number of outbreaks
Salmonella - S. Typhimurium	8	28	12	0	0	8
Salmonella - S. Enteritidis	73	329	95	0	5	78
Salmonella - Other serovars	12	29	6	0	0	12
Campylobacter	0	0	0	0	0	0
Listeria - Listeria monocytogenes	0	0	0	0	0	0
Listeria - Other Listeria	0	0	0	0	0	0
Yersinia	0	0	0	0	0	0
Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0	0	0	0	0	0
Bacillus - B. cereus	0	0	0	0	0	0
Bacillus - Other Bacillus	0	0	0	0	0	0
Staphylococcal enterotoxins	0	0	0	0	1	1
Clostridium - Cl. botulinum	0	0	0	0	0	0
Clostridium - Cl. perfringens	0	0	0	0	1	1

	Weak evidence or no vehicle outbreaks					
	Number of outbreaks	Human cases	Hospitalized	Deaths	Strong evidence Number of Outbreaks	Total number of outbreaks
Clostridium - Other Clostridia	0	0	0	0	0	0
Other Bacterial agents - Brucella	0	0	0	0	0	0
Other Bacterial agents - Shigella	0	0	0	0	0	0
Other Bacterial agents - Other Bacterial agents	5	329	1	0	0	5
Parasites - Trichinella	0	0	0	0	0	0
Parasites - Giardia	0	0	0	0	0	0
Parasites - Cryptosporidium	0	0	0	0	0	0
Parasites - Anisakis	0	0	0	0	0	0
Parasites - Other Parasites	0	0	0	0	0	0
Viruses - Norovirus	3	159	17	0	1	4
Viruses - Hepatitis viruses	0	0	0	0	0	0
Viruses - Other Viruses	0	0	0	0	0	0
Other agents - Histamine	0	0	0	0	0	0
Other agents - Marine biotoxins	0	0	0	0	0	0
Other agents - Other Agents	0	0	0	0	0	0

Hungary - 2013 Report on trends and sources of zoonoses

Weak	evidence or n	o vehicle outb	oreaks		
Number of outbreaks	Human cases	Hospitalized	Deaths	Strong evidence Number of Outbreaks	Total number of outbreaks
9	271	5	0	1	10

Unknown agent

Table Foodborne Outbreaks: detailed data for Clostridium

Please use CTRL for multiple selection fields

C. perfringens

Value

FBO Code	ÉTbl_7
Number of outbreaks	1
Number of human cases	66
Number of hospitalisations	0
Number of deaths	0
Food vehicle	Bovine meat and products thereof
More food vehicle information	
Nature of evidence	Descriptive epidemiological evidence
Outbreak type	General
Setting	Others
Place of origin of problem	Restaurant or Cafe or Pub or Bar or Hotel or Catering service
Origin of food vehicle	Domestic
Contributory factors	Cross-contamination
Mixed Outbreaks (Other Agent)	
Additional information	

Table Foodborne Outbreaks: detailed data for Salmonella

Please use CTRL for multiple selection fields

S. Enteritidis - PT 2

Value

FBO Code	OEK_2
Number of outbreaks	1
Number of human cases	21
Number of hospitalisations	0
Number of deaths	0
Food vehicle	Eggs and egg products
More food vehicle information	Tiramishu
Nature of evidence	Analytical epidemiological evidence
Outbreak type	General
Setting	School or kindergarten
Place of origin of problem	Household
Origin of food vehicle	Unknown
Contributory factors	Inadequate heat treatment
Mixed Outbreaks (Other Agent)	
Additional information	

Value

FBO Code	ÉTbi_24
Number of outbreaks	1
Number of human cases	116
Number of hospitalisations	12
Number of deaths	0
Food vehicle	Other foods
More food vehicle information	
Nature of evidence	Detection of causative agent in food chain or its environment - Symptoms and onset of illness pathognomonic to causative agent
Outbreak type	General
Setting	School or kindergarten
Place of origin of problem	Restaurant or Cafe or Pub or Bar or Hotel or Catering service
Origin of food vehicle	Domestic
Contributory factors	Unknown
Mixed Outbreaks (Other Agent)	
Additional information	

Value

FBO Code	OEK_1
Number of outbreaks	1
Number of human cases	23
Number of hospitalisations	0
Number of deaths	0
Food vehicle	Eggs and egg products
More food vehicle information	Vegetable salad with mayones
Nature of evidence	Analytical epidemiological evidence
Outbreak type	Household
Setting	Household
Place of origin of problem	Household
Origin of food vehicle	Unknown
Contributory factors	Inadequate heat treatment
Mixed Outbreaks (Other Agent)	
Additional information	

Value

FBO Code	OEK_3
Number of outbreaks	1
Number of human cases	6
Number of hospitalisations	2
Number of deaths	0
Food vehicle	Eggs and egg products
More food vehicle information	Pancake filled with cottage cheese cream contained raw eggs
Nature of evidence	Descriptive epidemiological evidence
Outbreak type	General
Setting	Household
Place of origin of problem	Household
Origin of food vehicle	Unknown
Contributory factors	Inadequate heat treatment
Mixed Outbreaks (Other Agent)	
Additional information	

Value

FBO Code	ÉTbl_23
Number of outbreaks	1
Number of human cases	2
Number of hospitalisations	0
Number of deaths	0
Food vehicle	Eggs and egg products
More food vehicle information	
Nature of evidence	Detection of causative agent in food chain or its environment - Symptoms and onset of illness pathognomonic to causative agent
Outbreak type	General
Setting	Canteen or workplace catering
Place of origin of problem	Canteen or workplace catering
Origin of food vehicle	Domestic
Contributory factors	Unknown
Mixed Outbreaks (Other Agent)	
Additional information	

Table Foodborne Outbreaks: detailed data for Staphylococcal enterotoxins

Please use CTRL for multiple selection fields

Enterotoxin, unspecified

Value

FBO Code	ÉTbl_17
Number of outbreaks	1
Number of human cases	17
Number of hospitalisations	13
Number of deaths	0
Food vehicle	Other foods
More food vehicle information	
Nature of evidence	Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent
Outbreak type	General
Setting	Restaurant or Cafe or Pub or Bar or Hotel or Catering service
Place of origin of problem	Unknown
Origin of food vehicle	Domestic
Contributory factors	Inadequate heat treatment
Mixed Outbreaks (Other Agent)	
Additional information	

Table Foodborne Outbreaks: detailed data for Unknown agent

Please use CTRL for multiple selection fields

Unknown

Value

FBO Code	ÉTbl_12
Number of outbreaks	1
Number of human cases	34
Number of hospitalisations	0
Number of deaths	0
Food vehicle	Broiler meat (Gallus gallus) and products thereof
More food vehicle information	
Nature of evidence	Analytical epidemiological evidence
Outbreak type	General
Setting	Temporary mass catering (fairs or festivals)
Place of origin of problem	Restaurant or Cafe or Pub or Bar or Hotel or Catering service
Origin of food vehicle	Domestic
Contributory factors	Cross-contamination
Mixed Outbreaks (Other Agent)	
Additional information	

Table Foodborne Outbreaks: detailed data for Viruses

Please use CTRL for multiple selection fields

Calicivirus - norovirus (Norwalk-like virus)

Value

FBO Code	ÉTbl_21
Number of outbreaks	1
Number of human cases	124
Number of hospitalisations	0
Number of deaths	0
Food vehicle	Other foods
More food vehicle information	
Nature of evidence	Analytical epidemiological evidence
Outbreak type	General
Setting	School or kindergarten
Place of origin of problem	Canteen or workplace catering
Origin of food vehicle	Domestic
Contributory factors	Unknown
Mixed Outbreaks (Other Agent)	
Additional information	