



WHAT IS THE RISK FOR XYLELLA FASTIDIOSA TO ESTABLISH IN TEMPERATE EUROPEAN REGIONS

A BELGIAN CASE STUDY

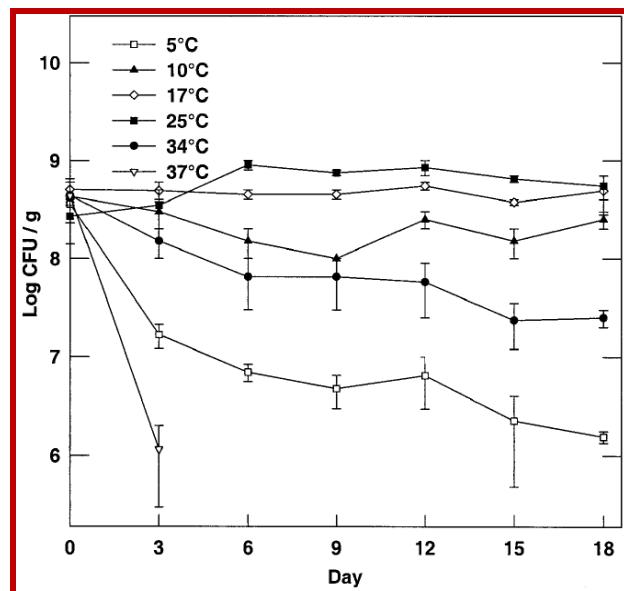
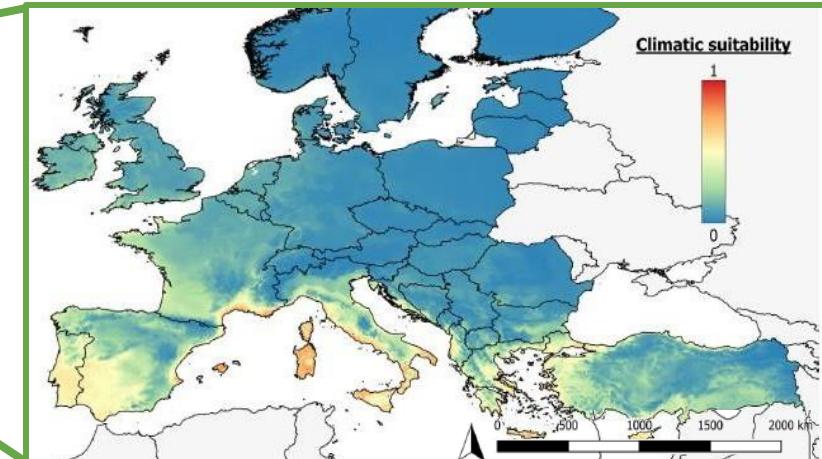
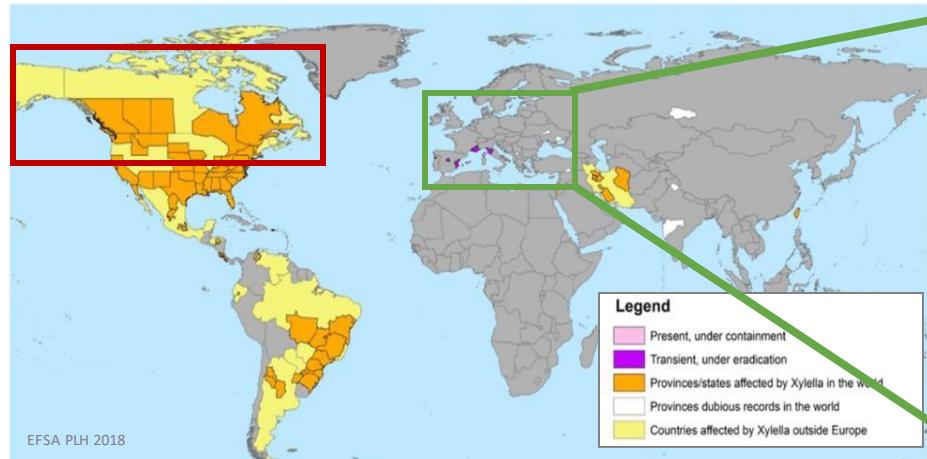
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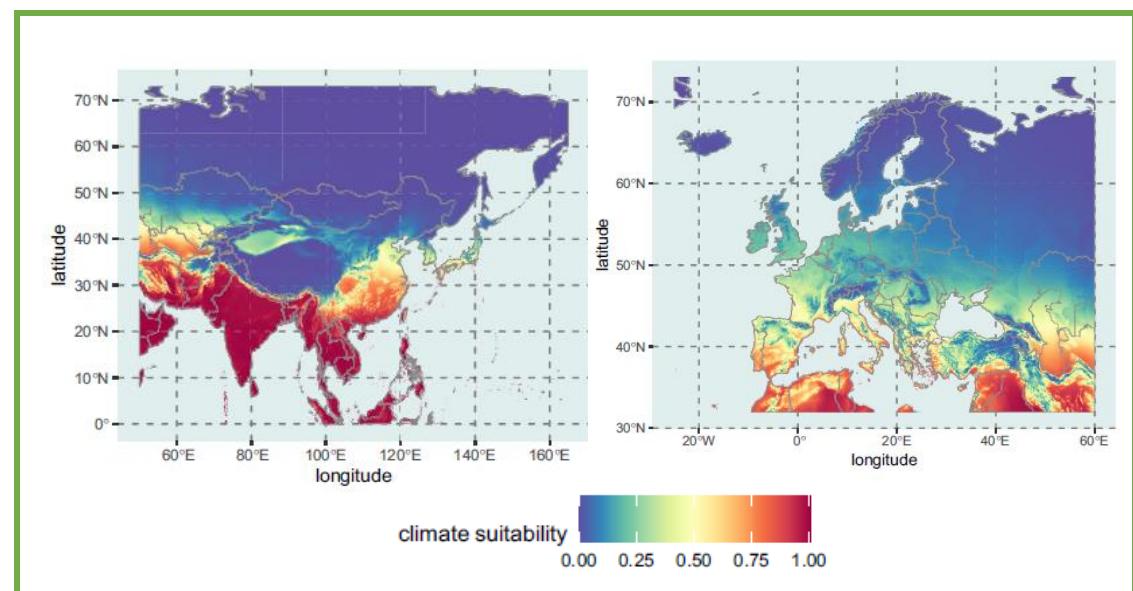
Introduction –

Introduction – Risk of *Xylella* in northern European temperate regions should not be underestimated

(EFSA PLH, 2019)



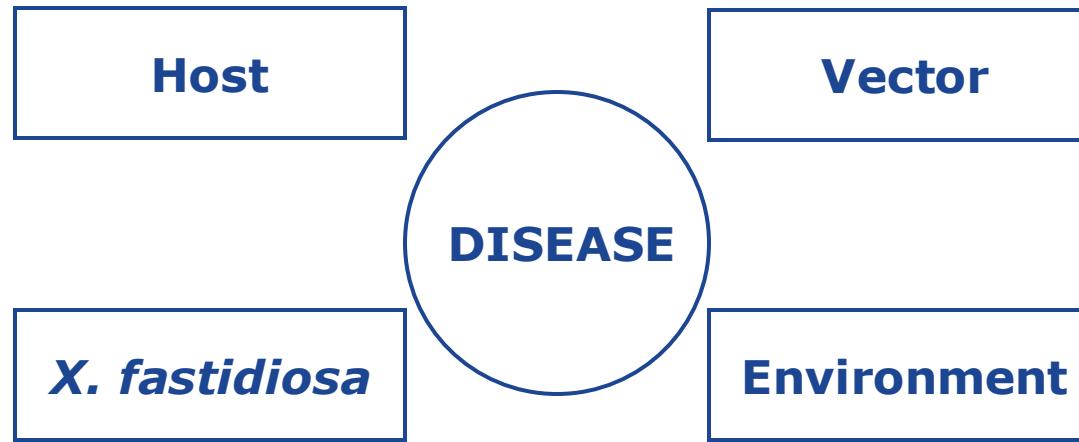
(Feil & Purcell, 2001)



(Godefroid et al., 2022)



Introduction – Risk of *Xylella* in northern European temperate regions should not be underestimated



Introduction – Risk of *Xylella* in northern European temperate regions should not be underestimated

J. Clark - University of California, Berkeley USA



Grapevine
Vitis vinifera

Mike Lewis, University of California, Riverside



L. J. Friesen - University of California, Berkeley USA



PIERCE'S
DISEASE

X. fastidiosa
subsp. *fastidiosa*
ST1

Graphocephala
atropunctata
Homalodisca
vitripennis

California,
Florida, etc.

Introduction – Risk of *Xylella* in northern European temperate regions should not be underestimated

Grapevine
Vitis vinifera

Graphocephala
atropunctata
Homalodisca
vitripennis

PIERCE'S
DISEASE

X. fastidiosa subsp.
fastidiosa ST1

A. Purcell
Source: <http://www.invasive.org>



Orange tree
Citrus sinensis

E. di Fiori
Source: <https://inaturalist.nz/>



F. Bernasconi



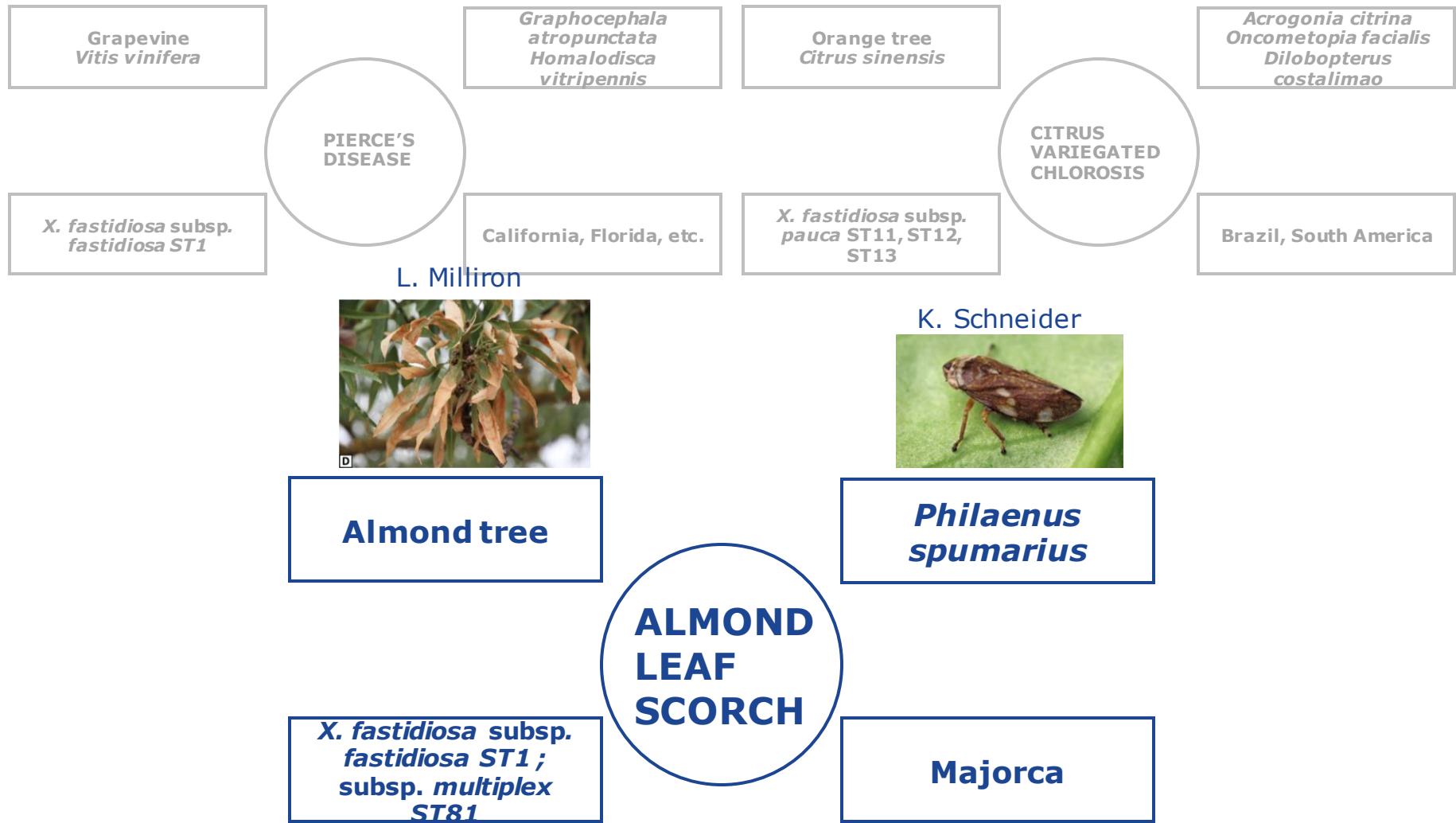
Acrogonia citrina
Oncometopia facialis
Dilobopterus costalimao

CITRUS
VARIEGATED
CHLOROSIS

X. fastidiosa
subsp. *pauca*
ST11, ST12, ST13

Brazil, South
America

Introduction – Risk of *Xylella* in northern European temperate regions should not be underestimated



Introduction – Risk of *Xylella* in northern European temperate regions should not be underestimated



Olive tree
Olea europaea

Philaenus spumarius

**OLIVE QUICK
DECLINE
SYNDROME**

***X. fastidiosa*
subsp. *pauca*
ST53**

Apulia, Italy

Introduction – Risk of *Xylella* in northern European temperate regions should not be underestimated

Van den Berk nurseries



Salicaceae

T. Murray



K. Schneider



***Philaenus spumarius*
*Aphrophora salicina***

DISEASE

***X. fastidiosa* subsp.
fastidiosa ST1
KLN59.3**

Riparian areas

Objectives -

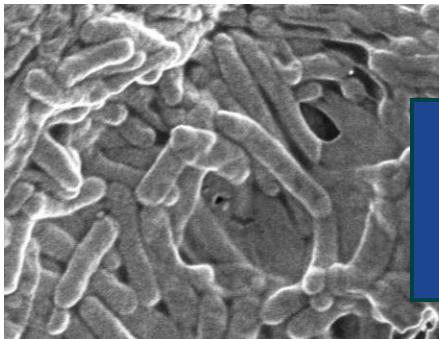
Objectives – Investigating the potential host plants and vectors

- Sentinel plantation
- Transmission test experiments and dispersion capacity
- Mechanical inoculations



Salicaceae

Aphrophoridae



***subsp.
fastidiosa ST1***

**Riparian areas
Salicaceae
corridors**



Methods and results – Sentinel plantation

Methods and results – Sentinel plantation



Casarín et al., 2023

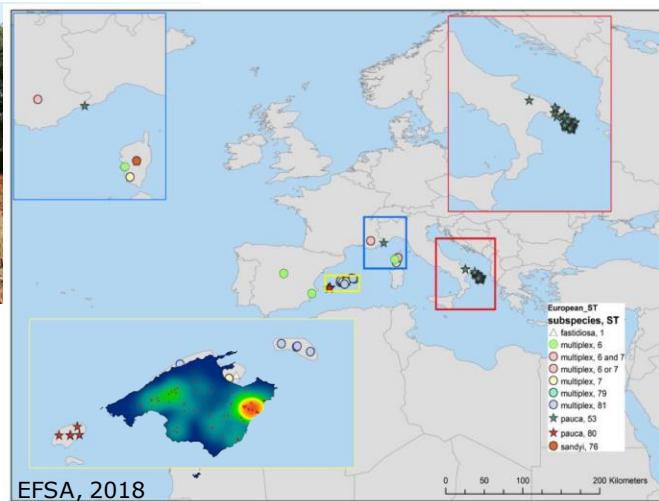
NeoBiota



27 *Salix alba*

27 *Quercus petraea*

27 *Prunus domestica* cv. Opal
from Belgium

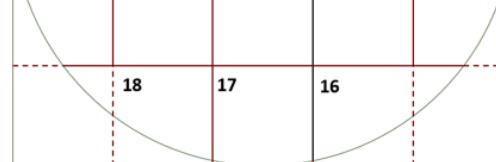


To the campus of UIB
(Palma de Mallorca)

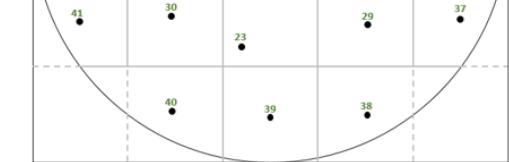
Floristic inventory



Insect vector sampling 25 blocks



Plantation of 32 « Spy plants » *Rosmarinus officinalis*



Methods and results – Sentinel plantation



Casarín et al., 2023

NeoBiota

Useful informations on how to set up a sentinel plantation

27 *Salix alba*

27 *Quercus petraea*

27 *Prunus*
from Be

Florist

Interesting knowledge to add sentinel plantation
in surveillance routine

Thanks to UIB team for helping us in this set up

Symptoms but no bacterial detection by PCR



Methods and results – Dispersion capacity and transmission test

Methods and results – Dispersion capacity



A. salicina > P. spumarius

Mark-release-recapture



Flight-mill



Collection of wild insects (50-60 ind. for each period) and sex identification

Flight-mill recording for 2h30
FIRST FLIGHT SESSION

Break: 24h feeding & resting on a host plant

Flight-mill recording for 2h30
SECOND FLIGHT SESSION



Casarín et al., 2023

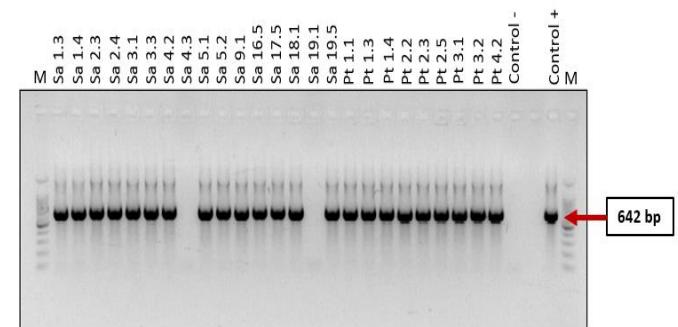
Journal of Pest Science



Transmission test

- 20 *S. alba* & 20 *P. tremula*
- 2 branches/tree
- 5-10 *P. spumarius* collected in infected parts of the Islands/branch
- Inoculation period: 4 to 5 days

- Insects tested by qPCR (Harper et al., 2010)
- Prevalence rate of infection: 17%
- MLST performed on positive samples (Yuan et al., 2010)





Methods – Mechanical inoculations reveal the potential of *Salicaceae* as host plants



Methods – Mechanical inoculations reveal the potential of Salicaceae as host plants



Pinprick inoculation (Hill & Purcell, 1995; Almeida et al., 2001)
in biosafety greenhouse

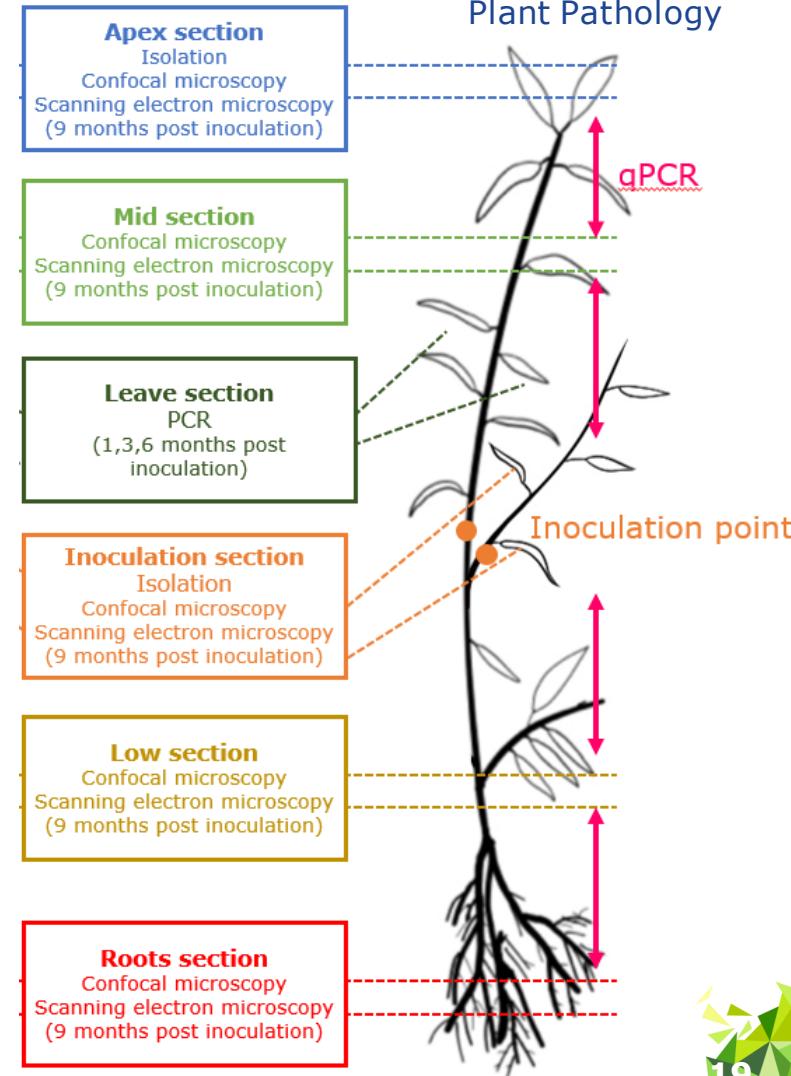
Casarín et al., 2022
European Journal of
Plant Pathology



- *P. canescens*, *P. tremula*,
S. alba



- 2 branches/plant,
- 22° C and 28° C

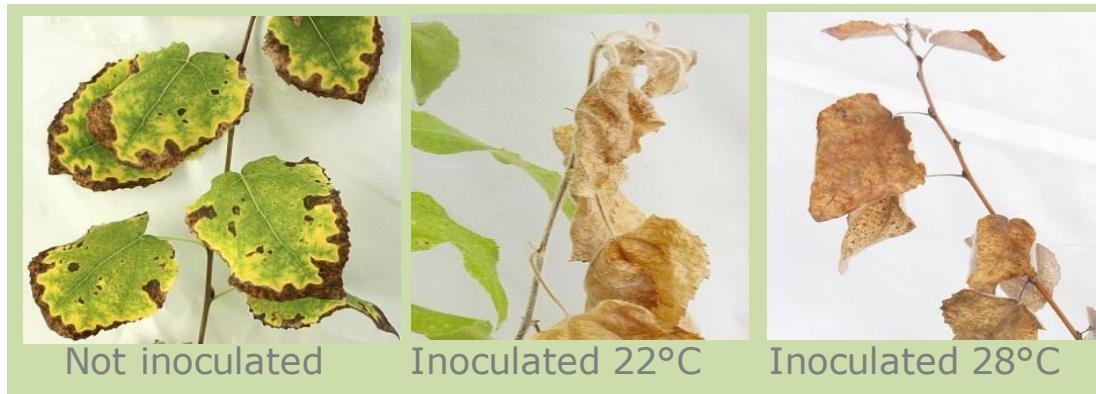




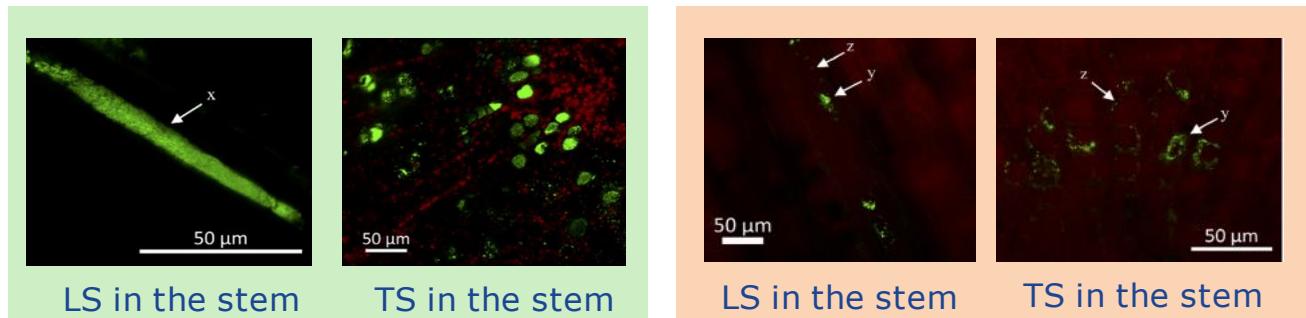
Results – Symptoms, propagation, and confocal microscopy for a look inside



Casarín et al., 2022
European Journal of
Plant Pathology



	<i>Populus tremula</i>		<i>Salix alba</i>		<i>Populus canescens</i>	
	22°C	28°C	22°C	28°C	22°C	28°C
Inoculation point (IP) detection	3/3	4/4	3/3	3/3	1/3	2/3
Apex detection	3/3	3/4	0/3	0/3	0/3	0/3
Root detection	1/1	3/3	0/1	1/1	NA	NA
Maximum spread from the IP	29 cm above the IP (stem) 53 cm below the IP (roots)	65 cm above the IP (stem) 70 cm below the IP (roots)	93 cm above the IP (stem) 15 cm below the IP (stem)	82 cm above the IP (stem) 23 cm below the IP (roots)	no spread	no spread





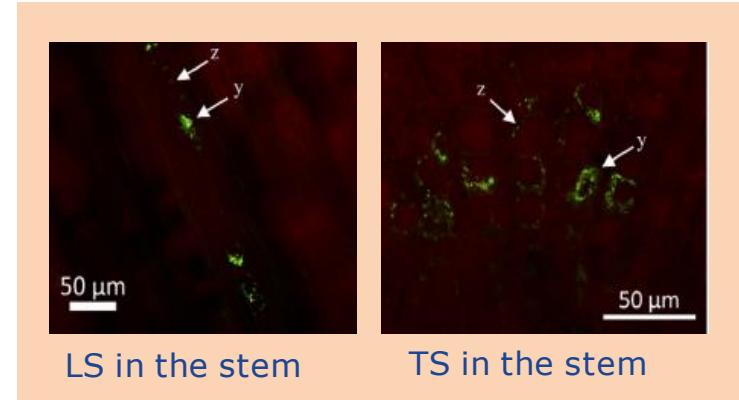
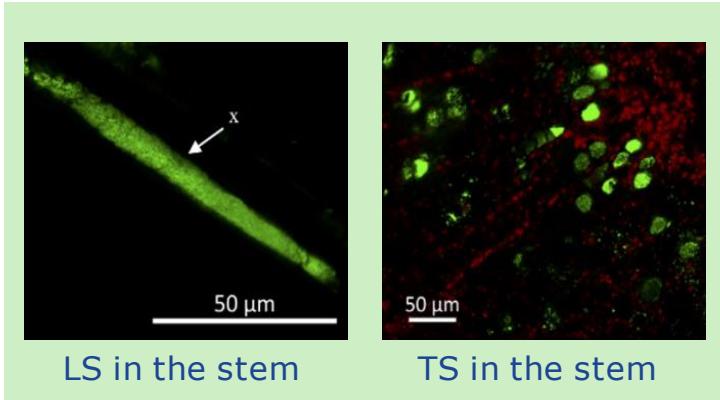
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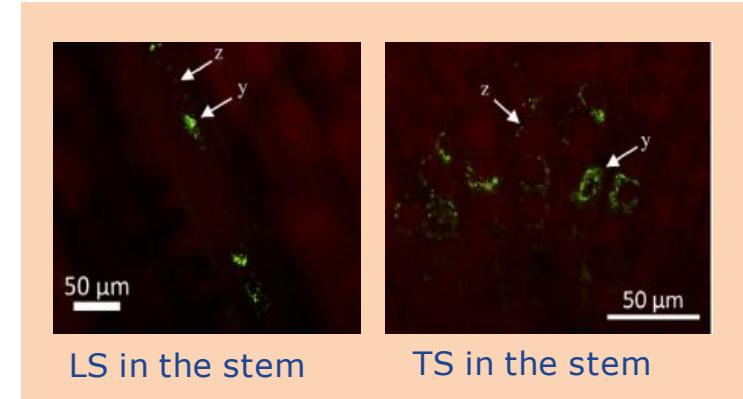
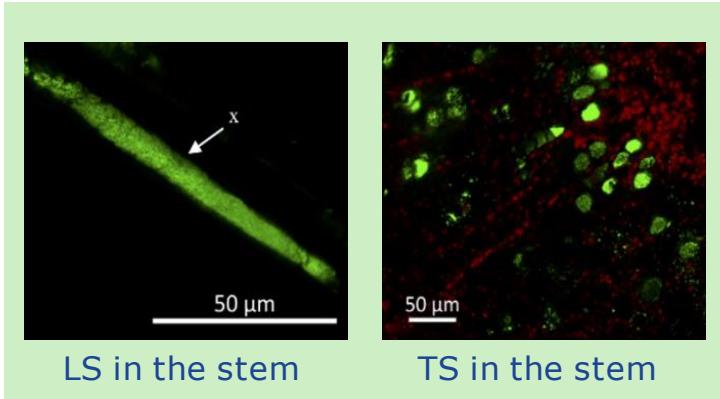
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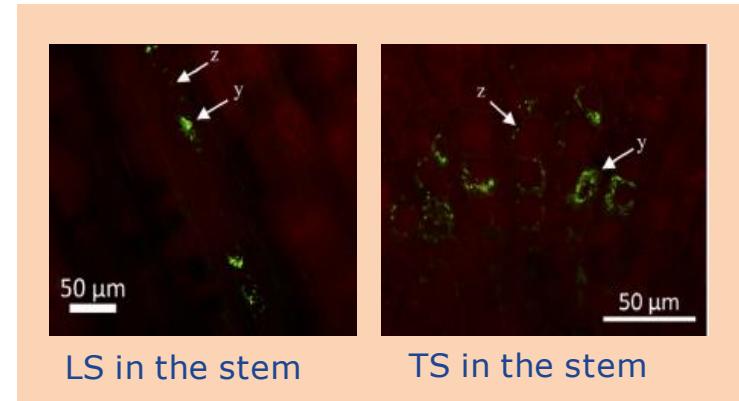
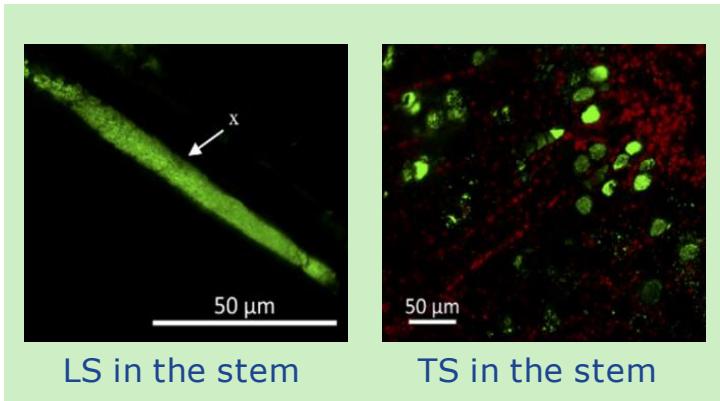
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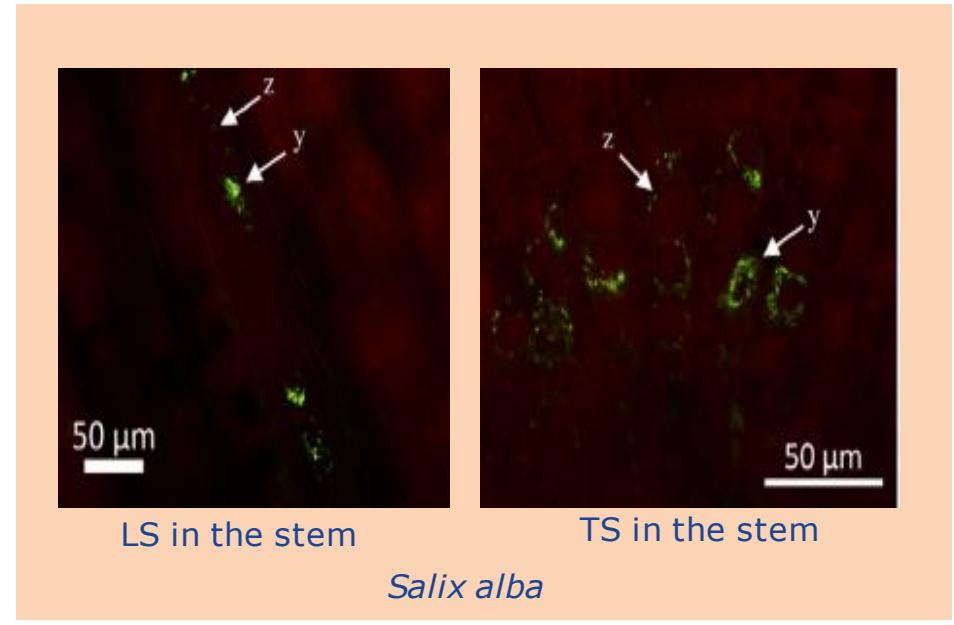
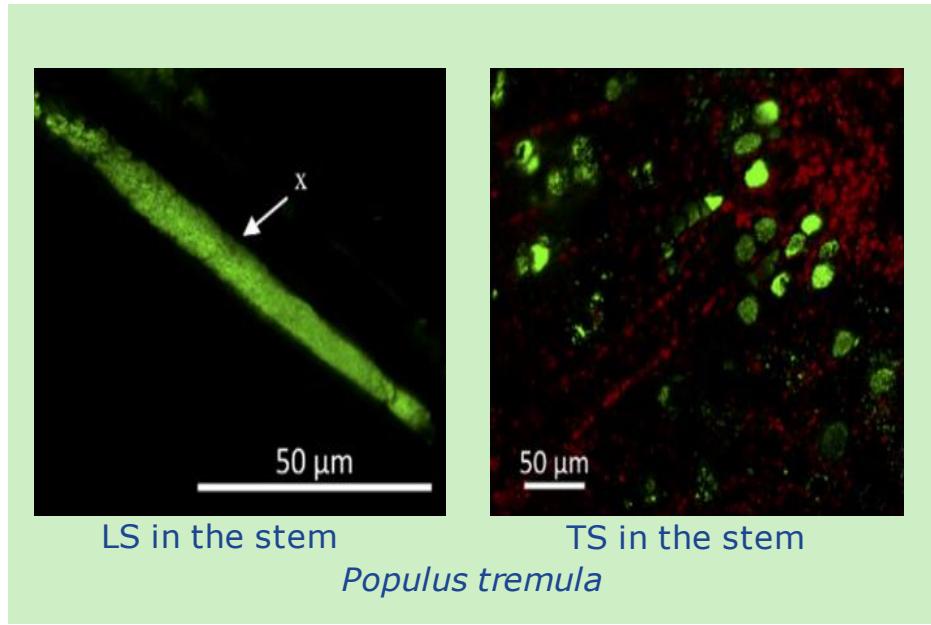




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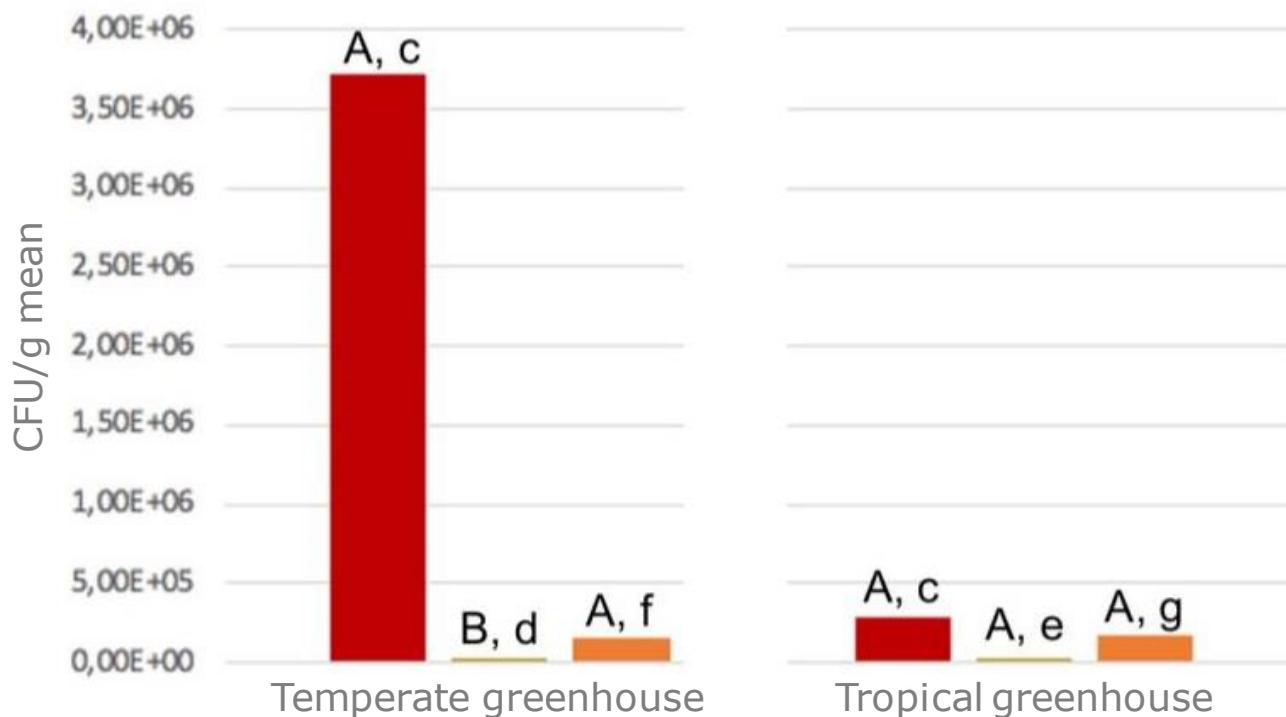
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CFU/g	<i>P. tremula</i>	<i>P. canescens</i>	<i>S. alba</i>	Total
Temperate	$3,72 \cdot 10^6$	$2,23 \cdot 10^3$	$1,53 \cdot 10^5$	$1,29 \cdot 10^6$
Tropical	$2,60 \cdot 10^5$	$1,76 \cdot 10^4$	$1,62 \cdot 10^5$	$1,47 \cdot 10^5$
Total	$1,99 \cdot 10^6$	$9,90 \cdot 10^3$	$1,58 \cdot 10^5$	

■ *P. tremula* ■ *P. canescens* ■ *S. alba*





Take home message – *Salicaceae* – potential host for *Xylella fastidiosa*
in Northern Europe



Take home message – Salicaceae – potential host for *Xylella fastidiosa* in Northern Europe

- Form a closed-mesh network of potential hosts for *Xylella*
- Could be symptomatic or asymptomatic
- Could act as reservoir

- Are found in riparian areas
- Feed on potential host plants
- *A. salicina* > *P. spumarius*



- Allows confocal microscopy visualization
- What about other strains or ST?

- Form *Salicaceae* corridors
- Could act as reservoir for potential host plants and insect vectors

Contributions

All:

Noemi Casarin (UCLouvain)
Séverine Hasbroucq (ULB)

Supervisors: Claude Bragard (UCLouvain)
Jean-Claude Grégoire (ULB)

Mechanical inoculations:

Lena Pesenti (UCLouvain)
Amandine Gérardin (UCLouvain)
Amélie Emond (UCLouvain)

Sentinel plantations:

Júlia
Miguel
Allison

And thank you for your
attention !

Dispersion tests:

Audrey Glibert (UCLouvain)
Gabriel Carestia (UCLouvain)

Distribution of potential vectors:

Ewelina Czwienczek (UCLouvain, EFSA)

And thank you to all who provided material :

Dr. S. Lindow for KLN59.3 strain
BCCM LMG collection
ILVO team



service public fédéral

**SANTE PUBLIQUE,
SECURITE DE LA CHAINE ALIMENTAIRE
ET ENVIRONNEMENT**



Poster n°21



**DECIPHERING XYLELLA POTENTIAL
PATHOSYSTEMS IN THE BELGIAN FLORA:**

A study on northern European temperate regions with emphasis on riparian areas

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