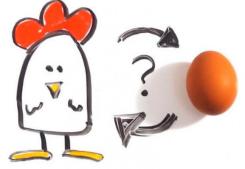




Working Group 2: In vitro methods for predicting sensitization

Daniel Lozano-Ojalvo, PhD

A chicken-and-egg situation:



What makes a food protein an allergen? Does digestion of food proteins affect their allergenicity?

On the cutting edge of reactions triggered by food:

Availability, strengths, and limitations of in vitro methods for allergenic sensitization assessment. What's about the celiac disease? And the GMOs?





Dr. Joana Costa Protein stability



Dr. Alan Mackie Protein digestion



Dr. Katrine Bogh In vitro approaches for allergenicity



Dr. Frits Koning Binding affinity in celiac disease



Dr. Andre Silvanovich Allergenicity assessment of GMOs – CropLife Europe

Novel protein-rich foods derived from plants and animals





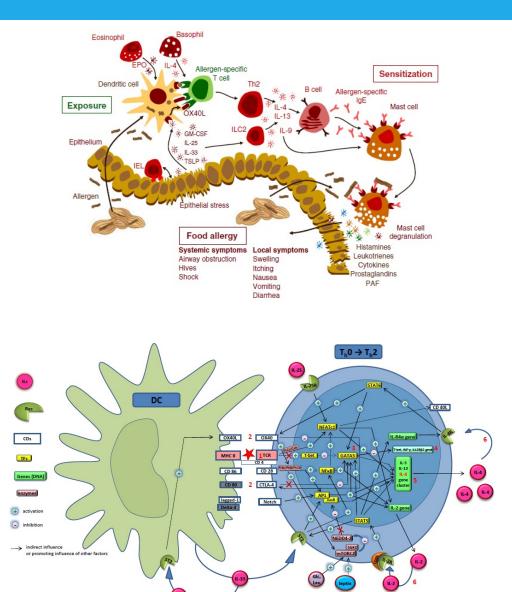


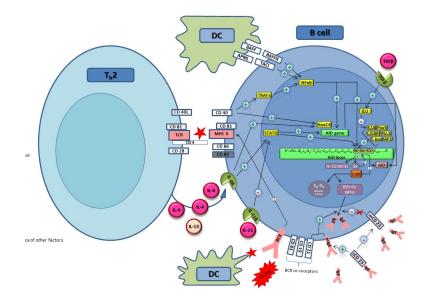




RISK ASSESSMENT: ALLERGENICITY

Biological events involved in sensitization to food proteins







Working Group for In Vitro Methods to Predict Sensitization

(Adapted from Smit et al., 2015, Drug Discov. Today, 17-19, 63-69.Van Bilsen et al., 2017, Clin. Transl. Allergy, 7, 18)

van Bilsen et al. Clin Transl Allergy (2017) 7:13 DOI 10.1186/s13601-017-0152-0 Clinical and Translational Allergy



Working Group for In Vitro Methods to Predict Sensitization

REVIEV

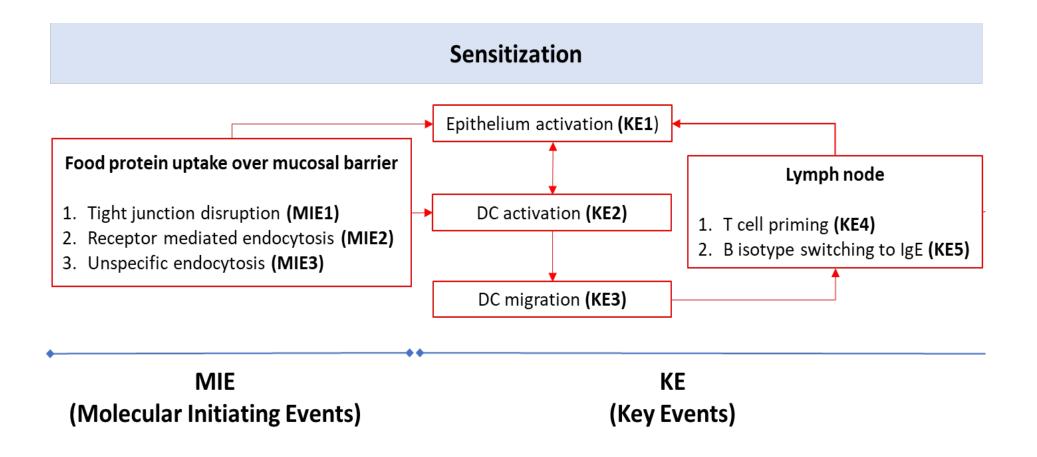


Application of the adverse outcome pathway (AOP) concept to structure the available in vivo and in vitro mechanistic data for allergic sensitization to food proteins

Jolanda H. M. van Bilsen^{1*}, Edyta Sienkiewicz-Szłapka², Daniel Lozano-Ojalvo³, Linette E. M. Willemsen⁴, Celia M. Antunes⁵, Elena Molina³, Joost J. Smit⁴, Barbara Wróblewska⁶, Harry J. Wichers⁷, Edward F. Knol⁸, Gregory S. Ladics⁹, Raymond H. H. Pieters⁴, Sandra Denery-Papini¹⁰, Yvonne M. Vissers¹¹, Simona L. Bavaro¹², Colette Larré¹⁰, Kitty C. M. Verhoeckx¹ and Erwin L. Roggen¹³

What is an Adverse Outcome Pathway (AOP)?

- An adverse outcome pathway (AOP) is a structured representation of biological events leading to adverse effects.
- A stepwise approach of events (one event leads to another event)
- It helps us to understand the mechanisms of adverse effects at a biological level of organization



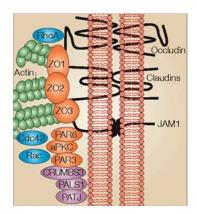


Working Group for In Vitro Methods to Predict Sensitization

We aimed to provide the state-of-the-art of existing *in vitro* approaches for assessing sensitizing potential of food proteins based on the identified MIE and KE proposed in the AOP for allergic sensitization to food proteins.

For this propose, we clustered and structured the existing *in vitro* models that are suitable to study the major events involved in allergic sensitization, focusing on major read-outs as well as strengths and limitations of these assays.

In vitro models to assess tight junction disruption (MIE1)



In vitro models:

- Caco-2
- HT-29
- T84
- IPEC-J2

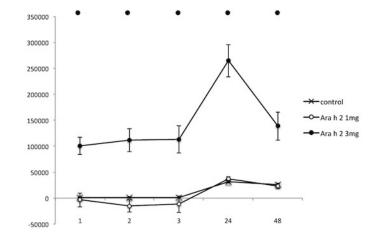
Disruption evaluation:

- Transepithelial Electrical Resistance (TEER)
- Blue Dextran leakage assay
- Lucifer Yellow assay

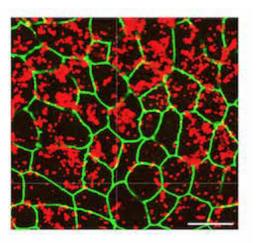
Cleavage of tight junctions:

- Western blotting
- Confocal microscopy
- RT-qPCR
- Mass spectrometry (tight junctions proteolysis)





Transport of Ara h 2



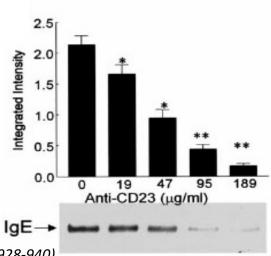
Modifications of Occludin, JAM-A, claudin-1, and ZO-1

In vitro models to assess receptor-mediated (MIE2) and unspecific endocytosis (MIE3)

MIE2: The role of CD23

In vitro models:

- Caco-2
- HT-29
- T84



(Tu et al., 2005, Gastroentorology, 129, 928-940)

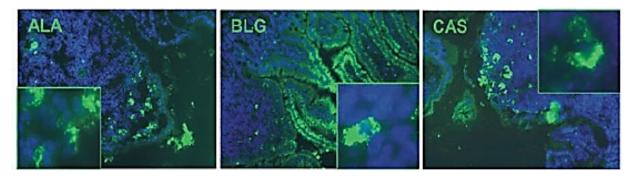
MIE3: Unspecific endocytosis

In vitro models:

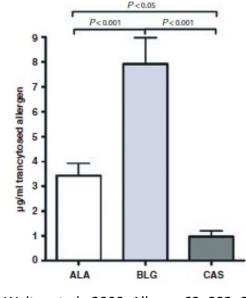
- Caco-2
- HT-29
- Murine M cells

Quantification of allergen passage:

- ¹⁴C-radiolabelled allergens
- Fluorophore-labelled allergens
- ELISA, SDS-PAGE and Western blotting



Transport mediated by endocytosis



(Roth-Walter et al., 2008, Allergy, 63, 882-890)

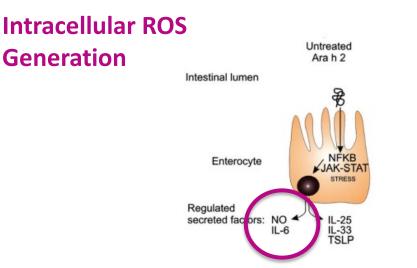
In vitro models to assess epithelium activation (KE1)

In vitro models:

- Caco-2
- HT-29
- T84

Evaluation of the epithelial activation:

- Intracellular ROS generation
- Cytokine production
- Mucus secretion
- Ca²⁺-signaling pathways

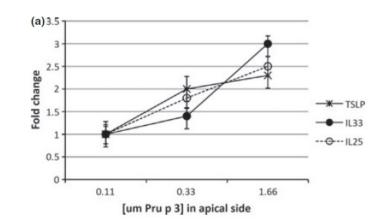


(Starkl et al., 2011, Open Allergy J., 4, 24-34)

***** Cow's Milk proteins (*α*-lactalbumin and β-lactoglobulin)

- 💠 Peanut (Ara h 2)
- Egg (ovomucoid and ovalbumin)
- Peach (Pru p 3)
- ***** Wheat proteins (β-, γ-, ω 1,2-, ω 5-gliadins)
- Soybean (P34)
- Brazil nuts (Ber e 1)
- Sesame seeds (Ses i 1)

Cytokine production



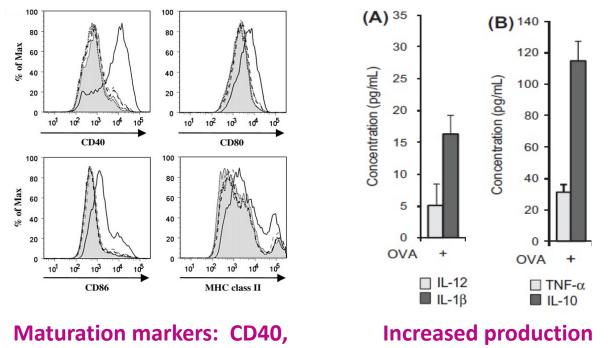
(Tordesillas et al., 2013, Clin. Exp. Allergy, 43, 1374-1383)

In vitro models:

- THP-1-derived DCs
- Human monocyte-derived DCs (moDCs)
- Murine bone marrow-derived DCs (BM-DCs)

Evaluation of the DC activation:

- Surface markers
- Cytokine production
- Allergen uptake



CD80, CD86, and MHC-II

of IL-1β and IL-10

Cow's Milk proteins (Bos d 5)

- Peanut (Agglutinin)
- ***** Egg (ovalbumin)
- Peach (Pru p 3)
- ***** Hazelnut (Cor a 8)
- Carrot (Dau c 1)
- Celery (Api g 1)
- Apple (Mad d 1)

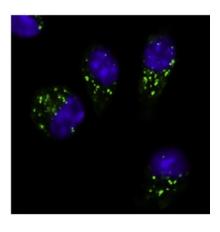
(Ilchmann et al., 2010, JACI, 125, 175-183)

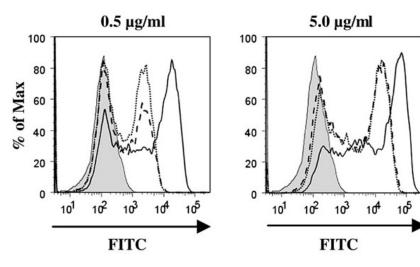
(Katayama et al., 2013, Food Chem., 138, 757-761)

DC activation

In vitro models to assess dendritic cell activation and migration (KE2 and KE3)

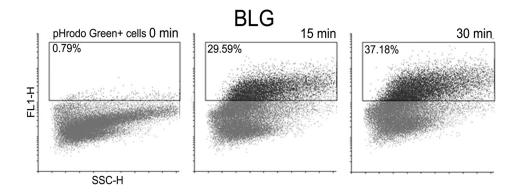
Allergen uptake



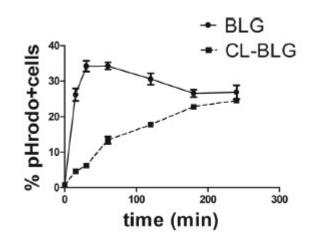


Enhanced uptake of glycosylated OVA (BM-DCs)

(Ilchmann et al., 2010, JACl, 125, 175-83)



Enhanced uptake of intact β-LG (BM-DCs)



(Stojadinovic et al., 2014, Toxicol. Sci., 140, 224-35)

Human in vitro models employed:

- Isolated PBMCs
- T cell lines
- T cell clones

Murine in vitro models employed:

- Isolated cells from spleen, MLN, and lamina propria
- Primed CD4+ T cells from sensitized mice
- CD4+ T cells from TCR trangenic mice (DO11.10 and OTII)

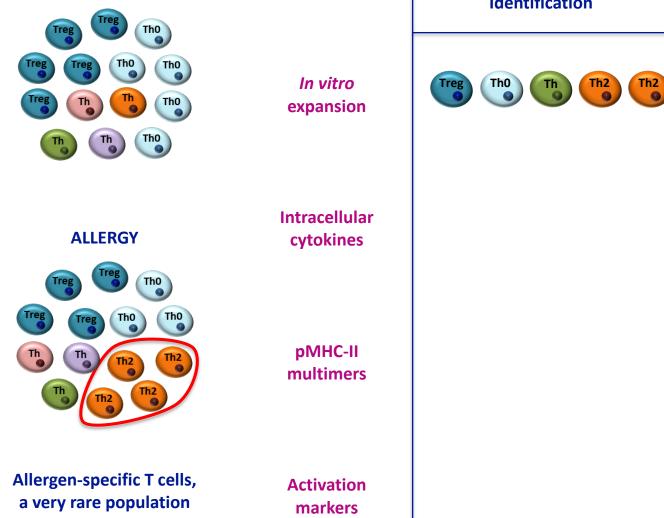
Evaluation of T cell activation :

- Proliferation
- Surface marker expression
- Cytokine secretion
- Cow's Milk proteins
- ***** (casein, *α*-lactalbumin and β-lactoglobulin)
- Peanut
- (Ara h 1, Ara h 2, Ara h 3 and Ara h 6)
- Egg (ovalbumin and ovomucoid)
- Peach (Pru p 3)
- Hazelnut (Cor a 8)
- Apple (Mal d 1)
- Walnut (Jug r 1, Jug r 2 and Jug r 3)
- Tree nuts (Ana o 1 and Ana o 2)

Challenge: To identify and characterize allergen-specific T cells

Detection of allergen-specific T cells

TOLERANCE



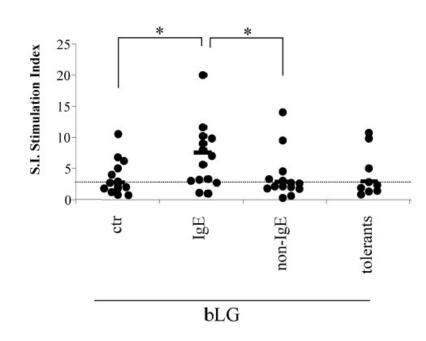
APPROACHES FOR THE STUDY OF ALLERGEN-SPECIFIC T CELLS

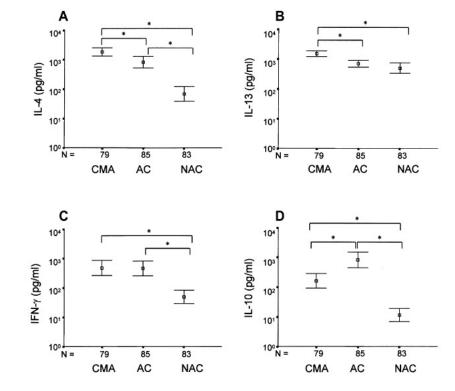
Identification

In vitro models to assess Th2 cell priming (KE4)

T cell proliferation

Cytokine production





β-LG induces proliferation in PBMCs from CMA allergic patients

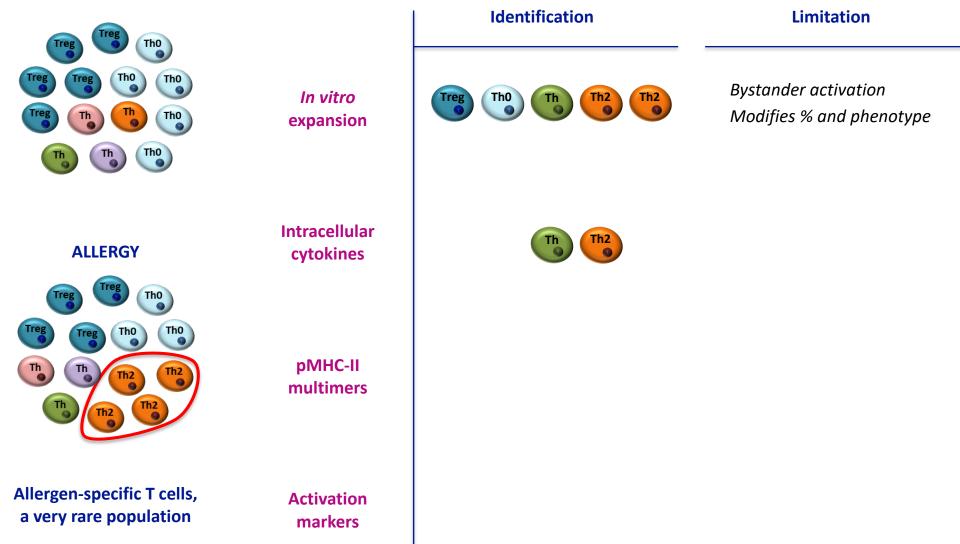
Increased production of IL4, IL13 and IFN-γ in T cell clones from CMA patients

(Vocca et al., 2011, Pediatric Res., 70, 549–554)

(Tiemessen et al., 2004, JACI., 113, 932-939)

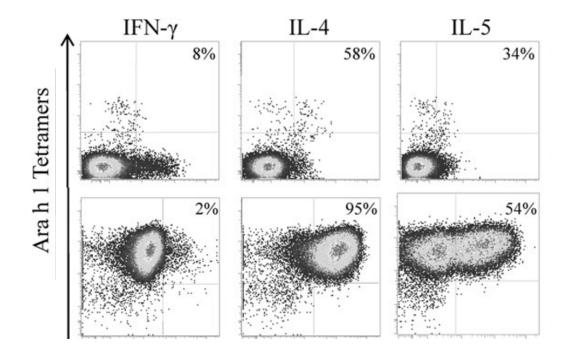
Detection of allergen-specific T cells

TOLERANCE



APPROACHES FOR THE STUDY OF ALLERGEN-SPECIFIC T CELLS

Intracellular Cytokine Production

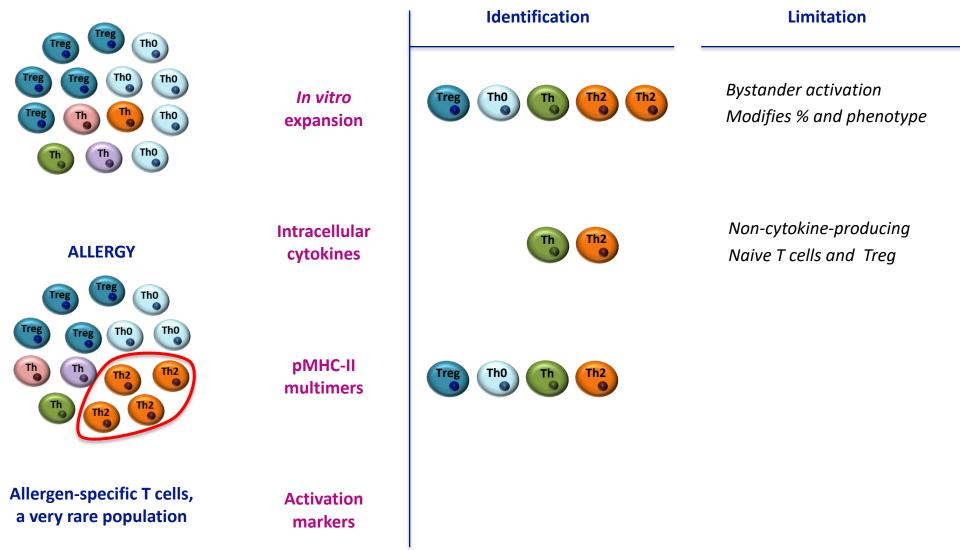


Increased production of IL4 and IL5 T cell clones from peanut patients

(DeLong et al., 2011, JACI, 127, 1211-1218)

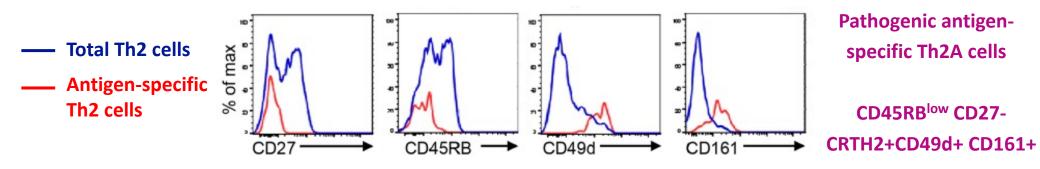
Detection of allergen-specific T cells

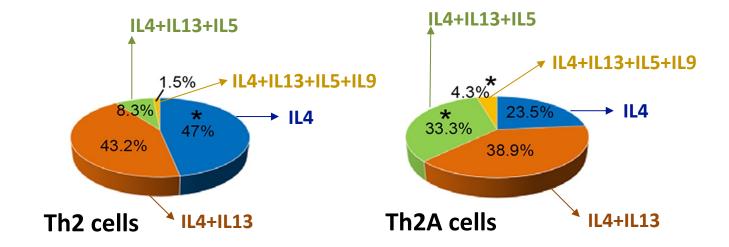
TOLERANCE



APPROACHES FOR THE STUDY OF ALLERGEN-SPECIFIC T CELLS

Identification of peanut-specific Th2 cells by pMHC-II TETRAMERS



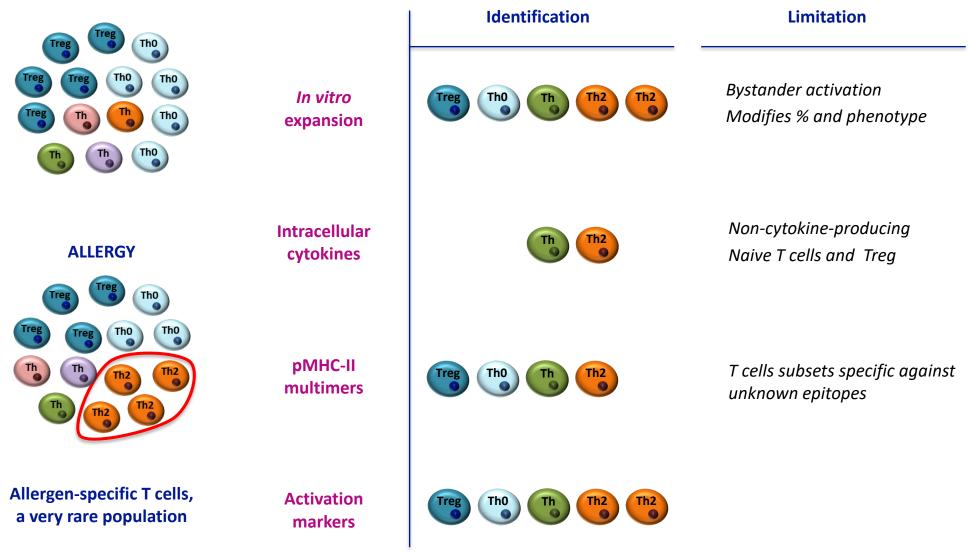


Polyfunctionality of Th2A cells with simultaneous expression of multiple Th2 effector cytokines

(Adapted from Wambre et al., 2017, Sci Transl Med, 9, eaam9171)

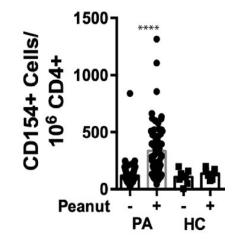
Detection of allergen-specific T cells

TOLERANCE

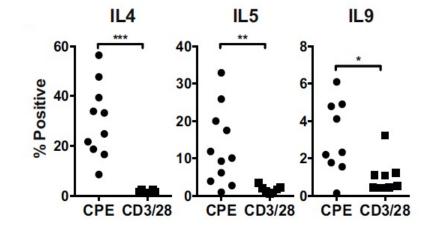


APPROACHES FOR THE STUDY OF ALLERGEN-SPECIFIC T CELLS

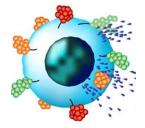
Identification of allergen-specific Th2 cells by ACTIVATION MARKERS



(Chiang et al., 2018, JACI, 141:2107-2120)

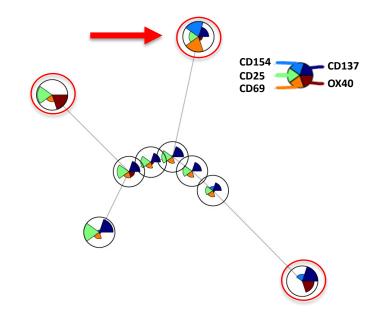


CD154 identified highly differentiated allergen-specific Th2 cells

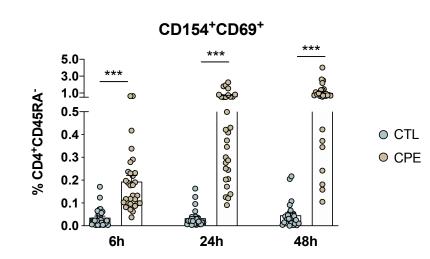


Induced <u>ACTIVATION MARKERS</u> for the identification of allergen- specific T cells:

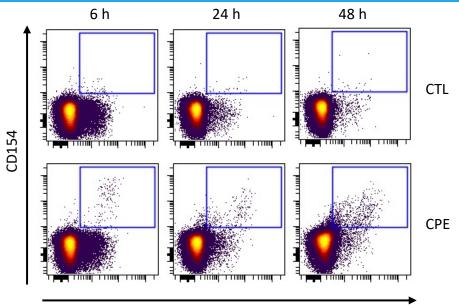
- Kinetics of expression of activation markers
- Combined expression of activation



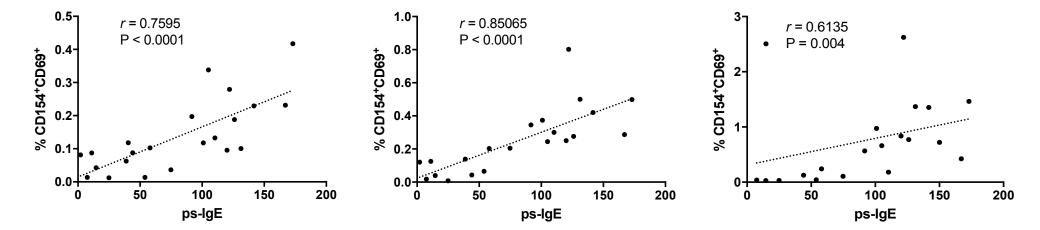
Identification of allergen-specific Th2 cells by ACTIVATION MARKERS



CD154/CD69 allow the early identification of allergen-specific T cells

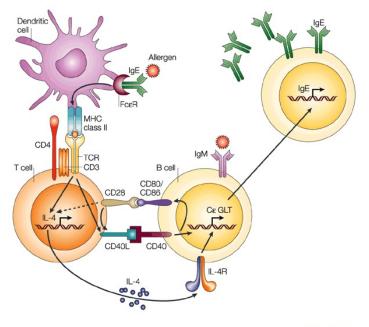


CD69



(Lozano-Ojalvo et al., 2021. Submitted)

Lack of *in vitro* methods for investigating B cell class-switching induced by food allergens



Nature Reviews | Immunology

Challenge: To identify methods

CONCLUSIONS

Trends in Food Science & Technology 85 (2019) 307-319



Contents lists available at ScienceDirect

Trends in Food Science & Technology

journal homepage: www.elsevier.com/locate/tifs



Review

Applying the adverse outcome pathway (AOP) for food sensitization to support *in vitro* testing strategies



Daniel Lozano-Ojalvo^{a,*}, Sara Benedé^b, Celia M. Antunes^c, Simona L. Bavaro^d, Grégory Bouchaud^e, Ana Costa^c, Sandra Denery-Papini^e, Araceli Díaz-Perales^f, María Garrido-Arandia^f, Marija Gavrovic-Jankulovic^g, Simone Hayen^h, Mónica Martínez-Blancoⁱ, Elena Molinaⁱ, Linda Monaci^d, Raymond H.H. Pieters^j, Clelia Villemin^e, Harry J. Wichers^k, Barbara Wróblewska^l, Linette E.M. Willemsen^m, Erwin L. Roggenⁿ, Jolanda H.M. van Bilsen^o

- * In vitro testing strategies for predicting allergenicity has been recently proposed
- B cell methods should be deeply investigated

Validate the in vitro methods proposed and harmonize a strategy for predicting sensitization

PERSPECTIVES

EU Risk Assessment Agenda (EU RAA) database



Content of a harmonized strategy for the *in vitro* **assessment of food allergenicity** (Spain)

Harmonized *in vitro* methods to evaluate dietary protein quality for human nutrition (France)

EFSA Strategy topics:

- *SO1.2* Make documentation on information gathering and the evaluation process available
- S03.2 Growing the EU and international RA community
- SO4.2 Develop and implement harmonized methodologies and guidance documents for risk assessment across the EU and internationally
- Determinations of allergen thresholds, measurements of allergens in food (Germany)
- Allergenicity of processed and raw food allergens and related analytical detectability (Belgium)
- Nanoplastic contamination in food: new factor affecting food allergy (Spain)



Sinai



Working Group 2: In vitro methods for predicting sensitization

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