

ZOONOSES MONITORING

Portugal

TRENDS AND SOURCES OF ZOONOSES AND ZOONOTIC AGENTS IN FOODSTUFFS, ANIMALS AND FEEDINGSTUFFS

including information on foodborne outbreaks, antimicrobial resistance in zoonotic and indicator bacteria and some pathogenic microbiological agents

IN 2017

Portugal - 2017

PREFACE

This report is submitted to the European Commission in accordance with Article 9 of Council Directive 2003/99/EC*. The information has also been forwarded to the European Food Safety Authority (EFSA).

The report contains information on trends and sources of zoonoses and zoonotic agents in Portugal during the year 2017.

The information covers the occurrence of these diseases and agents in animals, foodstuffs and in some cases also in feedingstuffs. In addition the report includes data on antimicrobial resistance in some zoonotic agents and indicator bacteria as well as information on epidemiological investigations of foodborne outbreaks. Complementary data on susceptible animal populations in the country is also given. The information given covers both zoonoses that are important for the public health in the whole European Union as well as zoonoses, which are relevant on the basis of the national epidemiological situation.

The report describes the monitoring systems in place and the prevention and control strategies applied in the country. For some zoonoses this monitoring is based on legal requirements laid down by the European Union legislation, while for the other zoonoses national approaches are applied.

The report presents the results of the examinations carried out in the reporting year. A national evaluation of the epidemiological situation, with special reference to trends and sources of zoonotic infections, is given. Whenever possible, the relevance of findings in foodstuffs and animals to zoonoses cases in humans is evaluated. The information covered by this report is used in the annual European Union Summary Reports on zoonoses and antimicrobial resistance that are published each year by EFSA.

The national report contains two parts: tables summarising data reported in the Data Collection Framework and the related text forms. The text forms were sent by email as pdf files and they are incorporated at the end of the report.

^{*} Directive 2003/ 99/ EC of the European Parliament and of the Council of 12 December 2003 on the monitoring of zoonoses and zoonotic agents, amending Decision 90/ 424/ EEC and repealing Council Directive 92/ 117/ EEC, OJ L 325, 17.11.2003, p. 31

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| Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Madelia Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meat from sheep - Processing plant - Monitoring - Official and industry sampling - AMR MON Salmonella Mbandaka Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official sampling - AMR MON Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official sampling - AMR MON Meat from brollers (Gallus gallus) - carcase - Slaughterhouse - Monitoring - OTHER AMR MON Salmonella Mikawasima Gallus gallus (fow) - laying hens - Hatchery - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Molade Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Monitorial - Official - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Monitorial - Cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Monitorial - Corton and eradication programmes - Official sampling - AMR MON Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official sampling - AMR MON Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Muenchen Meat from pig - meat products - Processing plant - Monitoring - Official sampling - AMR MON Turkeys - Farm - Clinical insetty sampling - OTHER AMR MON Gallus gallus (fow) - broilers - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Roughus (fow) - broilers - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Oliva - Farm - Control and eradication programmes - Official | 137 138 139 140 140 141 142 143 144 145 146 147 147 147 150 151 152 152 152 155 156 156 157 157 |
| Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official and lindustry sampling - AMR MON Salmonella Madelia Gallus gallus (fow) - broilers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meat from sheep - Processing plant - Monitoring - Official and industry sampling - AMR MON Salmonella Mandaka Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official sampling - AMR MON Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official sampling - AMR MON Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meat from broilers (Gallus gallus) - carcase - Staughterhouse - Monitoring - HACCP and own check - OTHER AMR MON Salmonella Mikawasima Gallus gallus (fow) - Jaying hens - Hatchery - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Montevideo Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Montevideo Gallus gallus (fow) - Jaying hens - Farm - Control and eradication programmes - Official and industry sampling - OTHER AMR MON Gallus gallus (fow) - Jaying hens - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Montevideo Gallus gallus (fow) - Jaying hens - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Montevideo Gallus gallus (fow) - Jaying hens - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Official - Jaying hens - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Official - Jaying hens - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Official - Jaying hens - Farm - Control and erad | 137 138 139 140 140 141 142 142 143 144 145 146 147 147 147 150 151 152 152 153 154 155 155 156 157 157 |
| Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Madelia Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meat from sheep - Processing plant - Monitoring - Official and industry sampling - AMR MON Salmonella Mbandaka Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official sampling - AMR MON Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official sampling - AMR MON Meat from brollers (Gallus gallus) - carcase - Slaughterhouse - Monitoring - OTHER AMR MON Salmonella Mikawasima Gallus gallus (fow) - laying hens - Hatchery - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Molade Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Monitorial - Official - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Monitorial - Cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Monitorial - Corton and eradication programmes - Official sampling - AMR MON Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official sampling - AMR MON Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Muenchen Meat from pig - meat products - Processing plant - Monitoring - Official sampling - AMR MON Turkeys - Farm - Clinical insetty sampling - OTHER AMR MON Gallus gallus (fow) - broilers - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Roughus (fow) - broilers - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Oliva - Farm - Control and eradication programmes - Official | 137 138 139 140 140 141 142 142 143 144 145 146 147 147 147 150 151 152 152 153 154 155 155 156 157 157 |
| Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official and lindustry sampling - AMR MON Salmonella Madelia Gallus gallus (fow) - broilers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meat from sheep - Processing plant - Monitoring - Official and industry sampling - AMR MON Salmonella Mandaka Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official sampling - AMR MON Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official sampling - AMR MON Gallus gallus (fow) - laying hens - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meat from broilers (Gallus gallus) - carcase - Staughterhouse - Monitoring - HACCP and own check - OTHER AMR MON Salmonella Mikawasima Gallus gallus (fow) - Jaying hens - Hatchery - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Montevideo Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Montevideo Gallus gallus (fow) - Jaying hens - Farm - Control and eradication programmes - Official and industry sampling - OTHER AMR MON Gallus gallus (fow) - Jaying hens - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Montevideo Gallus gallus (fow) - Jaying hens - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Montevideo Gallus gallus (fow) - Jaying hens - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Official - Jaying hens - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Official - Jaying hens - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Official - Jaying hens - Farm - Control and erad | 137 138 139 140 140 141 142 142 143 144 145 146 147 147 147 150 151 152 152 153 154 155 155 156 157 157 |
| Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official am industry sampling - AMR MON Salmonella Madelia Gallus gallus (fow) - broilers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meta from sheep - Processing plant - Monitoring - Official and industry sampling - AMR MON Salmonella Mbandaka Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official sampling - AMR MON Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meta from broilers (Gallus gallus) - (control and eradication programmes - Official and industry sampling - AMR MON Meta from broilers (Gallus gallus) - (control and eradication programmes - Official and industry sampling - AMR MON Salmonella Milawasiama Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - OTHER AMR MON Salmonella Milawasiama Gallus gallus (fow) - laying hers - Hatchey - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Mollade Feed material of coreal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Muenchen Meat from pig - meat products - Processing plant - Monitoring - Official and industry sampling - AMR MON Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official sampling - OTHER AMR MON Salmonella Otho Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Otho Salmonella Reading Meat from pig - carcase - Slaughterhouse - M | 137 138 139 140 140 141 142 143 144 145 146 146 147 147 147 148 149 150 151 151 152 153 154 155 156 156 157 158 158 158 159 |
| Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Madella Gallus gallus (fow) - broilers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meat from sheep - Processing plant - Monitoring - Official and industry sampling - AMR MON Salmonella Mandaka Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meat from broilers (Sallus gallus) - carcase - Saughterhouse - Monitoring - Official and industry sampling - AMR MON Salmonella Milavassima Gallus gallus (fow) - laying hers - Hatchery - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Milavassima Gallus gallus (fow) - laying hers - Farm - Control and industry sampling - OTHER AMR MON Salmonella Montervicico Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Mutenchen Meat from pig - meat products - Processing plant - Monitoring - Official and industry sampling - AMR MON Turkeys - Farm - Clinical investigations - Industry sampling - OTHER AMR MON Salmonella Neutrone Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official sampling - AMR MON Turkeys - Farm - Clinical investigations - Industry sampling - OTHER AMR MON Salmonella Neutrone Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Neutrone Meat from pig - carc | 137 138 139 140 140 141 142 142 143 144 145 146 147 147 148 148 149 150 151 152 152 153 154 155 156 157 157 158 158 159 159 |
| Gallus gallus (fow) - laying here - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Modelia Gallus gallus (fow) - broilers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meat from sheep - Processing plant - Monitoring - Official and industry sampling - AMR MON Salmonella Mbandaka Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Gallus gallus (fow) - laying here - Farm - Control and eradication programmes - Official sampling - AMR MON Gallus gallus (fow) - laying here - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meat from broilers (Gallus gallus) - carcase - Slaughterhouse - Monitoring - HACCP and own check - OTHER AMR MON Salmonella Milasus (fow) - laying here - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Mollade Feed material of creat grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Mollade Feed material of creat grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Gallus gallus (fow) - laying here - Ferm - Control and eradication programmes - Official sampling - AMR MON Gallus gallus (fow) - laying here - Farm - Control and eradication programmes - Official sampling - AMR MON Gallus gallus (fow) - laying here - Farm - Control and eradication programmes - Official sampling - AMR MON Gallus gallus (fow) - laying here - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Numer. Meat from pip - meat products - Processing plant - Monitoring - Official sampling - AMR MON Meat from pip - meat products - Processing plant - Monitoring - Official sampling - AMR MON Salmonella Numer. Gallus gallus (fow) - laying here - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Numer. Gallus gallus (fow) - laying here - Farm - Control and era | 137 138 139 140 140 141 142 142 143 144 145 146 147 148 149 150 151 151 152 152 152 153 154 155 156 156 157 157 158 158 159 160 161 |
| Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Madella Gallus gallus (fow) - broilers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meat from sheep - Processing plant - Monitoring - Official and industry sampling - AMR MON Salmonella Mandaka Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meat from broilers (Sallus gallus) - carcase - Saughterhouse - Monitoring - Official and industry sampling - AMR MON Salmonella Milavassima Gallus gallus (fow) - laying hers - Hatchery - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Milavassima Gallus gallus (fow) - laying hers - Farm - Control and industry sampling - OTHER AMR MON Salmonella Montervicico Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Mutenchen Meat from pig - meat products - Processing plant - Monitoring - Official and industry sampling - AMR MON Turkeys - Farm - Clinical investigations - Industry sampling - OTHER AMR MON Salmonella Neutrone Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official sampling - AMR MON Turkeys - Farm - Clinical investigations - Industry sampling - OTHER AMR MON Salmonella Neutrone Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Neutrone Meat from pig - carc | 137 138 139 140 140 141 142 142 143 144 145 146 147 147 148 148 149 150 151 152 152 153 154 155 156 157 157 158 158 159 159 |
| Gallus gallus (fow) - juying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Madelia Gallus gallus (fow) - broilers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meat from sheep - Processing plant - Monitoring - Official and industry sampling - AMR MON Salmonella Mbandakia Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - AMR MON Gallus gallus (fow) - juying hers - Farm - Control and eradication programmes - Official sampling - AMR MON Gallus gallus (fow) - juying hers - Farm - Control and eradication programmes - Official sampling - AMR MON Gallus gallus (fow) - juying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Milassis (fow) - juying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Milassis (fow) - juying hers - Farm - Control and eradication programmes - Official and industry sampling - OTHER AMR MON Salmonella Milassis (fow) - juying hers - Hatchery - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Milassis (fow) - juying hers - Farm - Control and eradication programmes - Official sampling - OTHER AMR MON Gallus gallus (fow) - juying hers - Farm - Control and eradication sampling - OTHER AMR MON Salmonella Milassis (fow) - juying hers - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Milassis (fow) - juying hers - Farm - Control and eradication programmes - Official and industry sampling - OTHER AMR MON Salmonella Neuerhen Meat from pig - meat products - Processing plant - Monitoring - Official sampling - AMR MON Salmonella Neuerhen Meat from pig - meat products - Processing plant - Monitoring - Official sampling - OTHER AMR MON Salmonella Neuerhen Meat from pig - racrase - Saughterhouse - Monitoring - HACCP and own check - AMR MON Meat from pig - carcase - Saughterhouse - Mon | 137 138 139 140 140 141 142 142 143 144 145 146 147 148 149 150 151 151 152 152 152 153 154 155 156 156 157 157 158 158 159 160 161 |
| Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official ampling - ANR MON Salmonella Madelia Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - ANR MON Meat from sheep - Processing plant - Monitoring - Official and industry sampling - ANR MON Salmonella Mibandika Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER ANR MON Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - OTHER ANR MON Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - ANR MON Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - ANR MON Salmonella Micavasima Gallus gallus (fow) - laying hers - Hatchey - Monitoring - Official and industry sampling - OTHER ANR MON Salmonella Micavasima Gallus gallus (fow) - laying hers - Hatchey - Monitoring - Official and industry sampling - OTHER ANR MON Salmonella Model Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER ANR MON Salmonella Model Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER ANR MON Salmonella Modelia (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - ANR MON Salmonella Modelia (fow) - laying hers - Farm - Control and eradication programmes - Official sampling - ANR MON Salmonella Modelia (fow) - laying hers - Farm - Control and eradication programmes - Official sampling - ANR MON Salmonella Modelia (fow) - laying hers - Farm - Control and eradication programmes - Official sampling - ANR MON Salmonella Modelia (fow) - laying hers - Farm - Control and eradication programmes - Official sampling - ANR MON Salmonella Modelia (fow) - laying hers - Farm - Control and eradication programmes - Official sampling - ANR MON Salm | 137 138 139 140 140 141 142 142 143 144 145 146 147 147 148 148 149 150 151 152 152 152 153 154 155 156 157 157 157 158 158 158 159 160 161 162 163 |
| Gallus gallus (fow) - laying hers - Farm - Control and endication programmes - Official sampling - ANR NON Salmonells Madelia Gallus gallus (fow) - bring hers - Farm - Control and endication programmes - Official and industry sampling - ANR NON Meat from sheep - Processing plant - Monitoring - Official and industry sampling - ANR NON Salmonells Mishandsa Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER ANR NON Gallus gallus (fow) - laying hers - Farm - Control and endication programmes - Official and industry sampling - OTHER ANR NON Gallus gallus (fow) - laying hers - Farm - Control and endication programmes - Official and industry sampling - ANR NON Meat from brolers (Gallus gallus) - carcase - Saluptherhouse - Monitoring - HACCP and own check - OTHER ANR NON Salmonells Miskawsima Gallus gallus (fow) - laying hers - Farm - Control and endication programmes - Official and industry sampling - ANR NON Salmonells Miskawsima Gallus gallus (fow) - laying hers - Farm - Control and endication programmes - Official and industry sampling - OTHER ANR NON Salmonells Monited (fow) - laying hers - Farm - Control and endication programmes - Official and industry sampling - OTHER ANR NON Gallus gallus (fow) - laying hers - Farm - Control and endication programmes - Official sampling - ANR NON Gallus gallus (fow) - laying hers - Farm - Control and endication programmes - Official sampling - ANR NON Gallus gallus (fow) - laying hers - Farm - Control and endication programmes - Official sampling - ANR NON Salmonella Notework - Processing plant - Monitoring - Official sampling - ANR NON Gallus gallus (fow) - laying hers - Farm - Control and endication programmes - Official sampling - ANR NON Meat from pip - meat products - Processing plant - Monitoring - Official sampling - ANR NON Salmonella Notework - Processing plant - Monitoring - Official sampling - ANR NON Meat from pip - meat products - Processing plant - Monitoring - Official and industry sampling - OTHER ANR NON Meat | 137 138 139 140 140 141 142 142 143 144 145 146 147 147 148 149 150 151 152 152 153 154 155 156 157 157 158 159 159 160 161 162 163 163 |
| Gallus gallus (fowl) - Jaying hers - Farm - Control and eradication programmes - Official and industry sampling - ANR NON Salmonella Madella (Fowl) - Jaying hers - Farm - Control and eradication programmes - Official and industry sampling - ANR NON Meat from sheep - Processing plant - Monitoring - Official and industry sampling - OTHER ANR NON Salmonella Minandiaka Feed material of creatal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER ANR MON Gallus gallus (fowl) - Jaying hers - Farm - Control and eradication programmes - Official and industry sampling - ANR NON Gallus gallus (fowl) - Jaying hers - Farm - Control and eradication programmes - Official and industry sampling - ANR NON Meat from brollens (Gallus gallus) - carcase - Slaughterhouse - Monitoring - HACCP and own check - OTHER ANR MON Salmonella Milavascima Callus gallus (fowl) - Jaying hers - Hatchery - Monitoring - Official and industry sampling - OTHER ANR MON Salmonella Molaved Feed material of creatal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER ANR MON Salmonella Monitored Gallus gallus (fowl) - Jaying hers - Farm - Control and eradication programmes - Official sampling - OTHER ANR MON Salmonella Monitored Feed material of creatal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER ANR MON Gallus gallus (fowl) - Jaying hers - Farm - Control and eradication programmes - Official sampling - OTHER ANR MON Gallus gallus (fowl) - Jaying hers - Farm - Control and eradication programmes - Official sampling - OTHER ANR MON Salmonella Monitor (fowl) - Jaying hers - Farm - Control and eradication programmes - Official sampling - ANR MON Salmonella Monitor (fowl) - Jaying hers - Farm - Control and eradication programmes - Official sampling - OTHER ANR MON Gallus gallus (fowl) - Jaying hers - Farm - Control and eradication programmes - Official sampling - OTHER ANR MON Salmonella Monitor (fowl) - Jaying hers - Farm - Control and eradication programmes - Official samp | 137 138 139 140 140 141 142 142 143 144 145 146 146 147 147 148 149 150 151 151 152 153 154 155 156 156 157 158 158 158 159 159 160 161 162 163 163 164 |
| Gallus gallus (fow) - laying hers - Farm - Control and endication programmes - Official sampling - ANR NON Salmonells Madelia Gallus gallus (fow) - bring hers - Farm - Control and endication programmes - Official and industry sampling - ANR NON Meat from sheep - Processing plant - Monitoring - Official and industry sampling - ANR NON Salmonells Mishandsa Feed material of cereal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER ANR NON Gallus gallus (fow) - laying hers - Farm - Control and endication programmes - Official and industry sampling - OTHER ANR NON Gallus gallus (fow) - laying hers - Farm - Control and endication programmes - Official and industry sampling - ANR NON Meat from brolers (Gallus gallus) - carcase - Saluptherhouse - Monitoring - HACCP and own check - OTHER ANR NON Salmonells Miskawsima Gallus gallus (fow) - laying hers - Farm - Control and endication programmes - Official and industry sampling - ANR NON Salmonells Miskawsima Gallus gallus (fow) - laying hers - Farm - Control and endication programmes - Official and industry sampling - OTHER ANR NON Salmonells Monited (fow) - laying hers - Farm - Control and endication programmes - Official and industry sampling - OTHER ANR NON Gallus gallus (fow) - laying hers - Farm - Control and endication programmes - Official sampling - ANR NON Gallus gallus (fow) - laying hers - Farm - Control and endication programmes - Official sampling - ANR NON Gallus gallus (fow) - laying hers - Farm - Control and endication programmes - Official sampling - ANR NON Salmonella Notework - Processing plant - Monitoring - Official sampling - ANR NON Gallus gallus (fow) - laying hers - Farm - Control and endication programmes - Official sampling - ANR NON Meat from pip - meat products - Processing plant - Monitoring - Official sampling - ANR NON Salmonella Notework - Processing plant - Monitoring - Official sampling - ANR NON Meat from pip - meat products - Processing plant - Monitoring - Official and industry sampling - OTHER ANR NON Meat | 137 138 139 140 140 141 142 142 143 144 145 146 147 147 148 149 150 151 152 152 153 154 155 156 157 157 158 159 159 160 161 162 163 163 |
| Gallus gallus (fowl) - Juying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Mtadelia Gallus gallus (fowl) - brughers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meat from sheep - Processing plant - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Mtadelia Feed material of creeal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Gallus gallus (fowl) - Juying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meat from brolles (Gallus gallus (fowl) - Juying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Mtawasima Gallus gallus (fowl) - Juying hers - Hatchery - Monitoring - Official and industry sampling - AMR MON Salmonella Mtawasima Gallus gallus (fowl) - Juying hers - Hatchery - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Mtawasima Gallus gallus (fowl) - Juying hers - Hatchery - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Monitorial of creeal grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Gallus gallus (fowl) - Juying hers - Farm - Control and eradication programmes - Official sampling - AMR MON Gallus gallus (fowl) - Juying hers - Farm - Control and eradication programmes - Official sampling - AMR MON Gallus gallus (fowl) - Juying hers - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Meet from po - meet products - Processing plant - Monitoring - Official sampling - OTHER AMR MON Gallus gallus (fowl) - Juying hers - Farm - Control and eradication programmes - Official sampling - AMR MON Salmonella Newport Meet from pop - meet products - Processing plant - Monitoring - Official sampling - OTHER AMR MON Salmonella Scelea Meet from pop - meet products - Processing plant - Monitoring - Official and industry sampling - OTHER AMR MON Mee | 137 138 139 140 140 141 142 143 144 145 146 147 147 147 150 151 152 153 154 155 156 156 157 158 158 159 160 161 162 163 164 165 |
| Gallus gallus (own) - Juying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Madelia (www) - brupiers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meat from sheep - Processing plant - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Mibandiaka Feed material of createj grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Gallus gallus ((ww) - Juying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Meat from broller (Gallus gallus (own) - Juying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Mibandia ((ww) - Juying hers - Hatchery - Monitoring - Official and industry sampling - AMR MON Salmonella Mibandia ((ww) - Juying hers - Hatchery - Monitoring - Official and industry sampling - AMR MON Salmonella Mibandia ((ww) - Juying hers - Hatchery - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Mibandia ((ww) - Juying hers - Hatchery - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Monitorido Feed material of creat grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER AMR MON Salmonella Monitorido Gallus gallus ((two) - Juying hers - Farm - Control and eradication programmes - Official and industry sampling - OTHER AMR MON Salmonella Monitorido ((two) - Juying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Monitorido ((two) - Juying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Monitorido ((two) - Juying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Monitorido ((two) - Juying hers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON Salmonella Seefing ((two) - Juying hers - Farm - Control and eradic | 137 138 139 140 140 141 142 142 143 144 145 146 147 147 148 148 149 150 151 152 153 154 155 156 157 158 159 159 159 159 160 161 162 163 163 164 165 165 |
| Gallas gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - ANR NON Salmonella Madella Mate from sheep - Processing plant - Monitoring - Official and industry sampling - ANR NON Salmonella Mandanda Feed material of create grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER ANR NON Salmonella Mandanda Feed material of create grain origin - Feed mill - Monitoring - Official and industry sampling - OTHER ANR NON Gallas gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - OTHER ANR NON Gallas gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and industry sampling - ANR NON Meet from broids (Sallus gallus - creates - Saluptrinos) - HACC and own check - OTHER ANR NON Salmonella Milkonesiama Callas gallus (fow) - laying bress - Hatchery - Monitoring - Official and industry sampling - OTHER ANR NON Salmonella Monitorino - Monitoring - Official and industry sampling - OTHER ANR NON Salmonella (fow) - laying bress - Hatchery - Monitoring - Official and industry sampling - OTHER ANR NON Salmonella (fow) - laying bress - Hatchery - Monitoring - Official and industry sampling - OTHER ANR NON Salmonella (fow) - laying bress - Farm - Control and eradication programmes - Official and industry sampling - ANR NON Salmonella (fow) - laying bress - Farm - Control and eradication programmes - Official and industry sampling - ANR NON Salmonella (fow) - laying bress - Farm - Control and eradication programmes - Official and industry sampling - ANR NON Salmonella Musericon - Farm - Control and eradication programmes - Official and industry sampling - ANR NON Salmonella Musericon - Farm - Control and eradication programmes - Official and industry sampling - ANR NON Salmonella Salus (fow) - laying bress - Farm - Control and eradication programmes - Official and industry sampling - ANR NON Meet from povine arinask and pig - mixed eradication programmes - O | 137 138 139 140 140 141 142 142 143 144 145 146 146 147 147 148 149 150 151 152 153 154 155 156 157 158 158 158 159 159 160 161 162 163 164 165 165 165 |
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| Birds - Natural habitat - Monitoring - Not applicable - OTHER AMR MON | |
|---|------|
| Pigeons - Natural habitat - Monitoring - Official sampling - OTHER AMR MON | 4.77 |
| Gallus gallus (fowl) - broilers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON | |
| Partridges - Farm - Monitoring - Official and industry sampling - OTHER AMR MON | |
| Meat from duck - fresh - Processing plant - Monitoring - Official and industry sampling - OTHER AMR MON | 400 |
| Meat from duck - carcase - Slaughterhouse - Monitoring - Official and industry sampling - OTHER AMR MON | |
| Meat from other poultry species - Processing plant - Monitoring - Official and industry sampling - OTHER AMR MON | |
| Turkeys - fattening flocks - Farm - Control and eradication programmes - Official and industry sampling - AMR MON | 400 |
| Gallus qallus (fowl) - laying hens - Farm - Control and eradication programmes - Official sampling - AMR MON | |
| Gallus qallus (fowl) - laying hens - Farm - Control and eradication programmes - Official and industry sampling - AMR MON | |
| Meat from duck - offal - Processing plant - Monitoring - Official and industry sampling - OTHER AME MON | |
| Meat from bovine animals - carcase - Slaughterhouse - Monitoring - Official and industry sampling - OTHER AMR MON | 407 |
| Meat from pig - meat products - Processing plant - Monitoring - Official and industry sampling - OTHER AMR MON | |
| Meat from pig - fresh - Processing plant - Monitoring - Official sampling - OTHER AME MON | |
| Meat from pig - fresh - Processing plant - Monitoring - Official and industry sampling - OTHER AMR MON | |
| Salmonella Uppsala | |
| Meat from broilers (Gallus gallus) - fresh - Processing plant - Monitoring - Official and industry sampling - OTHER AMR MON | |
| Salmonella Virchow | |
| Gallus gallus (fowl) - broilers - Farm - Control and eradication programmes - Official sampling - AMR MON | 400 |
| Gallus gallus (fowl) - laying hens - Farm - Control and eradication programmes - Official sampling - AMR MON | |
| Gallus gallus (fowl) - laying hens - Farm - Control and eradication programmes - Official and industry sampling - AMR MON | |
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| Escherichia coli, non-pathogenic, unspecified | |
| Meat from bovine animals - fresh - Retail - Monitoring - Official sampling - ESBL MON pnl2 | |
| Meat from bovine animals - fresh - Retail - Monitoring - Official sampling - ESBL MON | |
| Cattle (bovine animals) - calves (under 1 year) - Slaughterhouse - Monitoring - Official sampling - AMR MON pnl2 | |
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| Cattle (bovine animals) - calves (under 1 year) - Slaughterhouse - Monitoring - Official sampling - ESBL MON | 205 |
| Pigs - fattening pigs - Slaughterhouse - Monitoring - Official sampling - AMR MON pnl2 | 207 |
| Pigs - fattening pigs - Slaughterhouse - Monitoring - Official sampling - AMR MON | 208 |
| Pigs - fattening pigs - Slaughterhouse - Monitoring - Official sampling - ESBL MON pnl2 | 210 |
| Pigs - fattening pigs - Slaughterhouse - Monitoring - Official sampling - ESBL MON | 212 |
| Pigs - fattening pigs - Slaughterhouse - Monitoring - Official sampling - AMR MON | 214 |
| Pigs - fattening pigs - Slaughterhouse - Monitoring - Official sampling - ESBL MON pnl2 | 215 |
| Pigs - Fattening pigs - Slaughterhouse - Monitoring - Official sampling - ESBL MON | 217 |
| Meat from pig - fresh - Retail - Monitoring - Official sampling - ESBL MON pnl2 | 219 |
| Meat from pig - fresh - Retail - Monitoring - Official sampling - ESBL MON | 221 |
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| ESBL | 224 |
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ANIMAL POPULATION TABLES

Table Susceptible animal population

| | | | Population | |
|-------------------------------|--|------------|-----------------------------|------------|
| Animal species | Category of animals | animal | slaughter animal (heads) | herd/flock |
| Cattle (bovine animals) | Cattle (bovine animals) | 1,666,040 | 382,413 | 40,917 |
| | Cattle (bovine animals) - calves (under 1 year) | 290,879 | | 23,637 |
| | Cattle (bovine animals) - dairy cows | 341,816 | | 10,106 |
| | Cattle (bovine animals) - meat production animals | 780,899 | | 29,659 |
| Deer | Deer - farmed | | 41 | |
| Ducks | Ducks | 746,470 | 3,959,449 | 20 |
| Gallus gallus (fowl) | Gallus gallus (fowl) | | 215,769,317 | |
| | Gallus gallus (fowl) - breeding flocks, unspecified | | 1,964,227 | |
| | Gallus gallus (fowl) - broilers | 40,804,716 | 210,209,743 | 1,501 |
| | Gallus gallus (fowl) - laying hens | 11,708,665 | 3,595,347 | 188 |
| | Gallus gallus (fowl) - parent breeding flocks, unspecified | 4,644,852 | | 110 |
| Geese | Geese | | 6 | |
| Goats | Goats | 328,232 | 107,147 | 11,719 |
| 30als | Goats - animals over 1 year | 282,713 | | 11,601 |
| | Goats - animals under 1 year | 45,519 | | 4,238 |
| | Goats - meat production animals | 247,173 | | 10,945 |
| | Goats - milk goats | 69,578 | | 772 |
| Guinea fowl | Guinea fowl | | 320 | |
| Pigs | Pigs | 2,119,532 | 1,262,215 | 4,531 |
| | Pigs - breeding animals | 223,123 | | 3,251 |
| | Pigs - breeding animals - unspecified - sows and gilts | 218,267 | | 3,229 |
| | Pigs - fattening pigs | 1,169,298 | | 2,750 |
| | Pigs - fattening pigs - unspecified - piglets | | 1,224,482 | |
| Quails | Quails | | 9,194,121 | |
| Rabbits | Rabbits - farmed | | 4,104,195 | |
| Ratites (ostrich, emu, nandu) | Ratites (ostrich, emu, nandu) - farmed | | 55 | |
| Sheep | Sheep | 2,204,763 | 803,634 | 25,042 |
| neep - | Sheep - animals over 1 year | 1,715,410 | | 24,903 |
| | Sheep - animals under 1 year (lambs) | 489,353 | | 14,072 |
| | Sheep - meat production animals | 1,911,068 | | 23,539 |
| | Sheep - milk ewes | 257,257 | | 1,508 |

| | | | Population | | | | | | |
|--------------------|---------------------|------------------|------------|------------|--|--|--|--|--|
| | | slaughter animal | | | | | | | |
| Animal species | Category of animals | animal | (heads) | herd/flock | | | | | |
| Solipeds, domestic | Solipeds, domestic | 84,760 | 1,002 | 21,307 | | | | | |
| Turkeys | Turkeys | 1,861,945 | 3,602,995 | 127 | | | | | |
| Wild boars | Wild boars - farmed | | 189 | | | | | | |

Portugal - 2017

DISEASE STATUS TABLES

Table Bovine brucellosis - data on animals - Community co-financed eradication programmes

| Region | Total number of animals | Number of animals to be tested under the program | Number of animals tested | Number of animals tested individually | Number of positive animals | Number of positive animals slaughtered | Total number of animals slaughtered |
|--|-------------------------------|--|--------------------------|---------------------------------------|----------------------------|--|--|
| PORTUGAL | 1,624,756 | 910,874 | 900,822 | 820,044 | 343 | 359 | 371 |
| Norte | 355,567 | 179,481 | 179,118 | 141,522 | 172 | 169 | 177 |
| Centro (PT) | 166,322 | 96,398 | 96,253 | 75,081 | 0 | 0 | 0 |
| Lisboa | 186,611 | 72,261 | 71,401 | 59,608 | 1 | 3 | 3 |
| Alentejo | 710,992 | 427,962 | 423,703 | 417,251 | 156 | 174 | 174 |
| REGIÃO AUTÓNOMA DOS AÇORES (NUTS level 1) | 205,264 | 134,772 | 130,347 | 126,582 | 14 | 13 | 17 |

Table Bovine brucellosis - data on herds - Community co-financed eradication programmes

| _ | | | | | | |
|--|---------------------------------------|------------------------------------|-----------------------------|--|--|--------------------------|
| Region | Number of new positive herds | Number of depopulate d herds | Total number of herds | Number of herds under the program | Number of herds under the program tested/chec ked | Number of positive herds |
| PORTUGAL | 57 | 0 | 39,060 | 35,340 | 31,071 | 62 |
| Norte | 31 | 0 | 19,376 | 17,083 | 16,040 | 31 |
| Centro (PT) | 0 | 0 | 7,981 | 7,537 | 6,347 | 0 |
| Lisboa | 1 | 0 | 2,264 | 1,469 | 1,329 | 1 |
| Alentejo | 11 | 0 | 4,726 | 4,538 | 4,371 | 16 |
| REGIÃO AUTÓNOMA DOS AÇORES (NUTS level 1) | 14 | 0 | 4,713 | 4,713 | 2,984 | 14 |

Table Bovine brucellosis - data on status of herds at the end of the period - Community co-financed eradication programmes

| Region | Total number of herds under the program, at the end of the period | under the | Number of herds with status not free or not officially free and last check positive, at the end of the period | Number of animals with status not free or not officially free and last check positive, at the end of the period | Number of herds with status not free or not officially free and last check negative, at the end of the period | Number of animals with status not free or not officially free and last check negative, at the end of the period | Number of herds with status free or officially free suspended, at the end of the period | Number of animals with status free or officially free suspended, at the end of the period | Number of herds with status free, at the end of the period | Number of animals with status free, at the end of the period | Number of herds with status officially free, at the end of the period | Number of animals with status officially free, at the end of the period |
|--|---|-----------|--|---|--|---|---|--|---|---|---|---|
| PORTUGAL | 35,340 | 1,446,302 | 10 | 1,339 | 21 | 997 | 122 | 4,477 | 6,417 | 230,584 | 28,770 | 1,208,905 |
| Norte | 17,083 | 296,423 | 5 | 291 | 17 | 150 | 46 | 653 | 1,711 | 21,928 | 15,304 | 273,401 |
| Centro (PT) | 7,537 | 158,648 | 0 | 0 | 0 | 0 | 22 | 488 | 0 | 0 | 7,515 | 158,160 |
| Lisboa | 1,469 | 132,951 | 0 | 0 | 0 | 0 | 29 | 385 | 0 | 0 | 1,440 | 132,566 |
| Alentejo | 4,538 | 653,016 | 5 | 1,048 | 4 | 847 | 19 | 1,520 | 15 | 7,595 | 4,495 | 642,006 |
| REGIÃO AUTÓNOMA DOS AÇORES (NUTS level 1) | 4,713 | 205,264 | 0 | 0 | 0 | 0 | 6 | 1,431 | 4,691 | 201,061 | 16 | 2,772 |

Table Bovine brucellosis in countries and regions that do not receive Community co-financing for eradication programme

| Region | Number of animals serologicall y tested under investigations of suspect cases | Number of | seropositiv e animals under | positive to BST under | microbiolog ical testing under | Number of | Number of infected herds | Total number of animals | Number of herds tested under surveillance | Number of animals tested under surveillance | Total number of herds | Number of infected herds tested under surveillance | Number of herds tested under surveillance b by bulk milk | animals or pools tested under surveillance | | Number of notified abortions whatever cause | Number of isolations of Brucella infections | Number of abortions due to Brucella abortus | Number of animals tested by microbiolog y under investigatio ns of suspect cases |
|--|---|-----------|-----------------------------------|--------------------------|--------------------------------------|-----------|--------------------------------|-------------------------------|---|---|-----------------------------|---|---|--|---|---|--|---|--|
| PORTUGAL | (|) 0 |) (|) (|) (| 2,484 | 0 | 70,614 | 717 | 11,055 | 2,484 | (|) 101 | 1,798 | 0 | 4 | . 0 | (| 0 |
| Algarve (NUTS level 2) | C |) 0 |) (|) (|) (| 308 | 0 | 7,757 | 73 | 1,666 | 308 | (|) 0 | 0 | 0 | 0 | 0 | (| 0 |
| REGIÃO AUTÓNOMA DOS AÇORES (NUTS level 1) | (|) 0 | (|) (|) C | 2,176 | 0 | 62,857 | 644 | 9,389 | 2,176 | (|) 101 | 1,798 | 0 | 4 | 0 | (| 0 |

Table Ovine or Caprine brucellosis - data on animals - Community co-financed eradication programmes

| Region | Total number of animals | Number of animals to be tested under the program | Number of animals tested | Number of animals tested individually | Number of positive animals | Number of positive animals slaughtered | Total number of animals slaughtered |
|----------|-------------------------------|--|--------------------------|---------------------------------------|----------------------------|---|--|
| PORTUGAL | 2,487,280 | 1,644,739 | 1,512,820 | 1,512,820 | 1,588 | 1,389 | 1,420 |
| CONTINEN | TE 2,487,280 | 1,644,739 | 1,512,820 | 1,512,820 | 1,588 | 1,389 | 1,420 |

Table Ovine or Caprine brucellosis - data on herds - Community co-financed eradication programmes

| Region | Number of new positive herds | Number of depopulate d herds | Total number of herds | Number of herds under the program | Number of herds under the program tested/chec ked | Number of positive herds |
|------------|---------------------------------------|------------------------------------|-----------------------------|--|--|--------------------------|
| PORTUGAL | 341 | 3 | 56,348 | 56,216 | 54,550 | 396 |
| CONTINENTE | 341 | 3 | 56,348 | 56,216 | 54,550 | 396 |

Table Ovine or Caprine brucellosis - data on status of herds at the end of the period - Community co-financed eradication programmes

| = Region | the end of | Total number of animals under the program, at the end of the period | the end of | the end of | Number of herds with status not free or not officially free and last check negative, at the end of the period | negative, at the end of | | Number of animals with status free or officially free suspended, at the end of the period | Number of herds with status free, at the end of the period | Number of animals with status free, at the end of the period | Number of herds with status officially free, at the end of the period | Number of animals with status officially free, at the end of the period |
|-------------|------------|---|------------|------------|--|-------------------------|-------|--|---|---|---|---|
| PORTUGAL | 56,216 | 2,463,567 | 52 | 8,339 | 67 | 8,866 | 1,134 | 39,386 | 5,527 | 321,759 | 49,436 | 2,085,217 |
| CONTINENTE | 56,216 | 2,463,567 | 52 | 8,339 | 67 | 8,866 | 1,134 | 39,386 | 5,527 | 321,759 | 49,436 | 2,085,217 |

Table Ovine or Caprine brucellosis in countries and regions that do not receive Community co-financing for eradication programme

| Region | Number of animals serologicall y tested under investigatio ns of suspect cases | suspended herds under | e animals under | Number of animals positive in microbiolog ical testing under investigatio ns of suspect cases | Number of herds with status officially free | Number of infected herds | Total number of animals | Number of herds tested under surveillance | Number of animals tested under surveillance | Total number of herds | Number of infected herds tested under surveillance | Number of animals tested by microbiolog y under investigatio ns of suspect cases |
|--|--|--------------------------|--------------------|--|---|--------------------------------|-------------------------------|---|---|-----------------------------|---|--|
| PORTUGAL | 111 | 3 | 7 | . 0 | 788 | 0 | 9,619 | 248 | 2,691 | 788 | 0 | 7 |
| REGIÃO AUTÓNOMA DOS AÇORES (NUTS level 1) | | 3 | 7 | 0 | 788 | 0 | 9,619 | 248 | 2,691 | 788 | 0 | 7 |

DISEASE STATUS TABLES

Table Bovine tuberculosis - data on animals - Community co-financed eradication programmes

| Region | Total number of animals | Number of animals to be tested under the program | Number of animals tested | Number of animals tested individually | Number of positive animals | Number of positive animals slaughtered | Total number of animals slaughtered |
|--|-------------------------|--|--------------------------|---------------------------------------|----------------------------|--|-------------------------------------|
| PORTUGAL | 1,687,613 | 1,098,845 | 1,086,122 | 1,086,122 | 537 | 538 | 815 |
| Norte | 355,567 | 197,352 | 197,343 | 197,343 | 66 | 65 | 219 |
| Centro (PT) | 166,322 | 133,835 | 133,782 | 133,782 | 95 | 92 | 93 |
| Lisboa | 186,611 | 95,298 | 94,981 | 94,981 | 44 | 32 | 136 |
| Alentejo | 710,992 | 617,214 | 604,870 | 604,870 | 331 | 349 | 367 |
| REGIÃO AUTÓNOMA DOS AÇORES (NUTS level 1) | 268,121 | 55,146 | 55,146 | 55,146 | 1 | 0 | 0 |

Table Bovine tuberculosis - data on herds - Community co-financed eradication programmes

| Region | Number of new positive Num | ber of depopulated | Total number of herds | Number of herds under the program | Number of herds under the program tested/checked | Number of positive herds |
|--|----------------------------|--------------------|-----------------------|-----------------------------------|--|--------------------------|
| PORTUGAL | 61 | 3 | 41,236 | 37,516 | 31,552 | 87 |
| Norte | 12 | 2 | 19,376 | 17,083 | 16,653 | 14 |
| Centro (PT) | 5 | 0 | 7,981 | 7,537 | 7,312 | 12 |
| Lisboa | 4 | 1 | 2,264 | 1,469 | 1,417 | 5 |
| Alentejo | 39 | 0 | 4,726 | 4,538 | 4,431 | 55 |
| REGIÃO AUTÓNOMA DOS AÇORES (NUTS level 1) | 1 | 0 | 6,889 | 6,889 | 1,739 | 1 |

Table Bovine tuberculosis - data on status of herds at the end of the period - Community co-financed eradication programmes

| Region | Total number of herds under the program, at the end of the period | Total number of animals under the program, at the end of the period | Number of herds with status not free or not officially free and last check positive, at the end of the period | Number of animals with status not free or not officially free and last check positive, at the end of the period | Number of herds with status not free or not officially free and last check negative, at the end of the period | status not free or not | Number of herds with | | Number of herds with | |
|--|---|--|---|---|---|------------------------|----------------------|-------|----------------------|-----------|
| PORTUGAL | 37,516 | 1,509,159 | 10 | 1,728 | 58 | 9,500 | 100 | 5,187 | 37,348 | 1,492,744 |
| Norte | 17,083 | 296,423 | 0 | 0 | 9 | 365 | 18 | 172 | 17,056 | 295,886 |
| Centro (PT) | 7,537 | 158,648 | 4 | 288 | 10 | 1,761 | 29 | 1,052 | 7,494 | 155,547 |
| Lisboa | 1,469 | 132,951 | 0 | 0 | 2 | 208 | 20 | 371 | 1,447 | 132,372 |
| Alentejo | 4,538 | 653,016 | 6 | 1,440 | 37 | 7,166 | 29 | 3,332 | 4,466 | 641,078 |
| REGIÃO AUTÓNOMA DOS AÇORES (NUTS Jevel 1) | 6,889 | 268,121 | 0 | 0 | 0 | 0 | 4 | 260 | 6,885 | 267,861 |

Table Bovine tuberculosis in countries and regions that do not receive Community co-financing for eradication programme

| Region | Number of herds with status officially free | Number of infected herds | Total number of animals | Interval between routine tuberculin tests | Number of animals tested with tuberculin routine testing | Number of animals with suspicious lesions of tuberculosis examined and submitted to histopathological and bacteriological examinations | Number of animals detected positive in bacteriological examination | Total number of herds |
|------------------------|---|--------------------------|-------------------------|--|--|--|---|-----------------------|
| PORTUGAL | 307 | 1 | 7,757 | | 1,909 | 1 | 1 | 308 |
| Algarve (NUTS level 2) | 307 | 1 | 7,757 | 48 | 1,909 | 1 | 1 | 308 |

PREVALENCE TABLES

Table Brucella:BRUCELLA in food

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|------------------|---|---------------------------|------------------|-----------------------|-----------------------|--------------------------|----------------------------|----------|---------------------|
| Not Available | Cheeses, made from mixed milk from cows, sheep and/or goats - fresh - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Microbiological tests | 2 | 0 | Brucella | 0 |
| | Cheeses, made from mixed milk from cows, sheep and/or goats - soft and semi-soft - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Microbiological tests | 2 | 0 | Brucella | 0 |
| | Cheeses, made from mixed milk from cows, sheep and/or goats - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Microbiological tests | 9 | 0 | Brucella | 0 |
| | Milk, cows' - raw milk - Farm - Portugal - food sample - milk - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Millilitre | Microbiological tests | 2 | 0 | Brucella | 0 |
| | Milk, goats' - raw milk - Farm - Portugal - food sample - milk - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Millilitre | Microbiological tests | 4 | 0 | Brucella | 0 |
| | $\label{eq:milk-surveillance-official} Milk, sheep's - raw milk - Farm - Portugal - food sample - milk - Surveillance - Official sampling - Objective sampling$ | single (food/fee d) | 25 | Millilitre | Microbiological tests | 10 | 0 | Brucella | 0 |

Table Calicivirus: CALICIVIRUS in food

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | | Sample weight unit | Method | units | | Zoonoses | N of units positive |
|------------------|--|--------------------------|----|-----------------------|---------------|-------|---|-----------|------------------------|
| Not Available | Fruits - non-pre-cut - frozen - Border inspection activities - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Not Available | 5 | 0 | Norovirus | 0 |

Table Campylobacter: CAMPYLOBACTER in food

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|------------------|---|---------------------------|---------------|-----------------------|----------------------------------|--------------------------|----------------------------|--------------------------------|---------------------|
| Not Available | Meat from bovine animals - carcase - chilled - Slaughterhouse - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 10 | Gram | Detection method presence in x g | 15 | 0 | Campylobacter | 0 |
| | Meat from bovine animals - fresh - chilled - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 10 | Gram | Detection method presence in x g | 37 | 1 | Campylobacter, unspecified sp. | 1 |
| | Meat from bovine animals - meat preparation - intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 10 | Gram | Detection method presence in x g | 17 | 0 | Campylobacter | 0 |
| | Meat from bovine animals - minced meat - intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 10 | Gram | Detection method presence in x g | 9 | 0 | Campylobacter | 0 |
| | Meat from broilers (Gallus gallus) - carcase - Slaughterhouse - Portugal - food sample - Surveillance - | single | 10 | Gram | Detection method | 4 | 3 | Campylobacter coli | 1 |
| | Official sampling - Objective sampling | (food/fee d) | | | presence in x g | | | Campylobacter jejuni | 1 |
| | | | | | | | | Campylobacter, unspecified sp. | 1 |
| | Meat from broilers (Gallus gallus) - fresh - Processing plant - Portugal - food sample - Surveillance - Official | single (food/fee | 10 | Gram | Detection method | 27 | 9 | Campylobacter coli | 6 |
| | sampling - Objective sampling | d) | | | presence in x g | | | Campylobacter jejuni | 2 |
| | | · · | | | | | | Campylobacter, unspecified sp. | 1 |
| | Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 10 | Gram | Detection method presence in x g | 12 | 1 | Campylobacter coli | 1 |
| | Meat from goat - fresh - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 10 | Gram | Detection method presence in x g | 3 | 0 | Campylobacter | 0 |
| | Meat from pig - carcase - Slaughterhouse - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 10 | Gram | Detection method presence in x g | 9 | 1 | Campylobacter coli | 1 |
| | Meat from pig - fresh - Processing plant - Portugal - food sample - Surveillance - Official sampling - | single | 10 | Gram | Detection method | 29 | 3 | Campylobacter coli | 1 |
| | Objective sampling | (food/fee d) | | | presence in x g | | | Campylobacter, unspecified sp. | 2 |
| | Meat from pig - meat preparation - intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | - / | 10 | Gram | Detection method presence in x g | 22 | 2 | Campylobacter coli | 2 |
| | Meat from pig - meat products - cooked, ready-to-eat - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 10 | Gram | Detection method presence in x g | 6 | 0 | Campylobacter | 0 |
| | Meat from sheep - fresh - Slaughterhouse - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 10 | Gram | Detection method presence in x g | 6 | 1 | Campylobacter jejuni | 1 |
| | Meat from turkey - fresh - Slaughterhouse - Portugal - food sample - Surveillance - Official sampling - | single | 10 | Gram | Detection method | 11 | 3 | Campylobacter coli | 1 |
| | Objective sampling | (food/fee d) | | | presence in x g | | | Campylobacter, unspecified sp. | 2 |
| | Meat from turkey - meat preparation - intended to be eaten cooked - Slaughterhouse - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 10 | Gram | Detection method presence in x g | 7 | 2 | Campylobacter, unspecified sp. | 2 |
| | Meat, mixed meat - meat preparation - intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 10 | Gram | Detection method presence in x g | 11 | 0 | Campylobacter | 0 |

Table Cronobacter: CRONOBACTER in food

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | | Method | | Total units positive | Zoonoses | N of units positive |
|------------------|--|---------------------------|------------------|------|--|----|----------------------------|-------------|------------------------|
| Not Available | Infant formula - ready-to-eat - Hospital or medical care facility - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee d) | 10 | Gram | ISO/TS 22964:2006 (IDF/RM 210: 2006) Cronobacter spp. (Enterobacter sakazakii) | 17 | 0 | Cronobacter | 0 |

| ea of sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling un | Sample it weight | Sample weight unit | Method | total unit | s total units positive | Zoonoses | ANTH | VTX | AG | N units positive |
|----------------|--|-----------------------|------------------|--------------------------|---|------------|---------------------------|--|-----------------------------------|--|---|------------------|
| ot Available | Juice - fruit juice - unpasteurised - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/feed) | 25 | Millilitre | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 4 | 0 | Verocytotoxi genic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Meat from bovine animals - carcase - chilled - Slaughterhouse - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 11 | 1 | VTEC O157 | H7 | Verotoxin production, VT1 | eae negative | 1 |
| | Meat from bovine animals - fresh - chilled - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 23 | 1 | VTEC O103 | H-antigen unknown | Verotoxin production, toxin type unknown | Adhesion genes not investigate d | 1 |
| | Meat from bovine animals - fresh - frozen - Border inspection activities - Not Available - food sample - Surveillance - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 15 | 1 | VTEC other than O157 O26 O103 O111 O145 | H-antigen unknown | VT2, gene identified, subtype unspecified | eae negative | 1 |
| | Meat from bovine animals - meat preparation - intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | ISO/TS 13136:2012 (including the EU-RL adaptation for | 14 | 2 | VTEC O145 | H-antigen unknown | Verotoxin production, toxin type unknown | Adhesion genes not investigate d | 1 |
| | | | | | O104:H4) | | | VTEC 0157 | H-antigen unknown | Verotoxin production, toxin type unknown | Adhesion genes not investigate d | 1 |
| | Meat from bovine animals - minced meat - intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 15 | 1 | VTEC O103 | H-antigen unknown | Verotoxin production, toxin type unknown | Adhesion genes not investigate d | 1 |
| | Meat from pig - meat preparation - intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | ISO/TS 13136:2012 (including the EU-RL adaptation for | 35 | 2 | VTEC 0145 | H-antigen unknown H-antigen | Verotoxin production, toxin type unknown Verotoxin | eae negative eae | 1 |
| | | | | | O104:H4) | | | V1200137 | unknown | production, VT1 | negative | 1 |
| | Meat from pig - meat products - cooked, ready-to-eat - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 1 | 1 | VTEC O26 | H-antigen unknown | Verotoxin production, toxin type unknown | Adhesion genes not investigate d | 1 |
| | Meat from pig - minced meat - intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 7 | 0 | Verocytotoxi genic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Other processed food products and prepared dishes - unspecified - ready-to-eat foods - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 46 | 0 | Verocytotoxi genic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Other processed food products and prepared dishes - vegetable based dishes - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 16 | 0 | Verocytotoxi genic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Ready-to-eat salads - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/feed) | 25 | Gram | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 10 | 0 | Verocytotoxi genic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |

| Area of sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling uni | Sample t weight | Sample weight unit | Method | total units | total units | Zoonoses | ANTH | VTX | AG | N units positive |
|------------------|--|-----------------------|--------------------|--------------------------|---|-------------|-------------|---|------------------|------------------|------------------|------------------|
| Not Available | Seeds, sprouted - ready-to-eat - Farm - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 4 | 0 | Verocytotoxi genic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Seeds, sprouted - ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | In house real time PCR methods based on ISO/TS 13136:2012 | 5 | 0 | Verocytotoxi genic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Seeds, sprouted - ready-to-eat - Retail - Spain - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/feed) | 25 | Gram | In house real time PCR methods based on ISO/TS 13136:2012 | 5 | 0 | Verocytotoxi genic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |
| | Vegetables - pre-cut - ready-to-eat - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/feed) | 25 | Gram | ISO/TS 13136:2012 (including the EU-RL adaptation for O104:H4) | 12 | 0 | Verocytotoxi genic E. coli (VTEC) | Not Available | Not Available | Not Available | 0 |

Table FLAVIVIRUS in animal

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Vaccination status | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|------------------|---|------------------|--------------------|---|--------------------------|----------------------------|-----------------|---------------------|
| PORTUGAL | Solipeds, domestic - horses - Farm - Not Available - animal sample - blood - Monitoring - active - Official sampling - Suspect sampling | animal | No | Enzyme-linked immunosorbent assay (ELISA) | | 3 | West Nile virus | 3 |
| CONTINENTE | Solipeds, domestic - horses - Farm - Not Available - animal sample - blood - Monitoring - active - Official sampling - Suspect sampling | animal | No | Enzyme-linked immunosorbent assay (ELISA) | | 3 | West Nile virus | 3 |

| pling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|-------|--|---------------------------|------------------|--------------------------|--------------------------|----------------------------|-------------------|-----------|-------------------|---------------------|
| е | Fish - cooked - chilled - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official | batch | 10 | Gram | 9 | 0 | <= 100 | Histamine | 9 | 0 |
| | sampling - Objective sampling | (food/fee d) | | | | | >100 TO <= 200 | Histamine | 9 | 0 |
| | | | | | | | >200 | Histamine | 9 | 0 |
| | Fish - cooked - frozen - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official | batch | 10 | Gram | 9 | 0 | <= 100 | Histamine | 9 | 0 |
| | sampling - Selective sampling | (food/fee d) | | | | | >100 TO <= 200 | Histamine | 9 | 0 |
| | | | | | | | >200 | Histamine | 9 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Processing | single | 10 | Gram | 29 | 1 | <= 100 | Histamine | 29 | 8 |
| | plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | (food/fee d) | | | | | >100 TO <= 200 | Histamine | 29 | 0 |
| | | | | | | | >200 | Histamine | 29 | 1 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Selective sampling | batch | 10 | Gram | 72 | 0 | <= 100 | Histamine | 72 | 12 |
| | piant - Portugai - 1000 sample - Surveillance - based on Regulation 2073 - Official sampling - Selective sampling | (food/fee d) | | | | | >100 TO <= 200 | Histamine | 72 | 0 |
| | | | | | | | >200 | Histamine | 72 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Retail - | batch | 10 | Gram | 9 | 0 | <= 100 | Histamine | 9 | 0 |
| | Mauritius - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | (food/fee d) | | | | | >100 TO <= 200 | Histamine | 9 | 0 |
| | | | | | | | >200 | Histamine | 9 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Retail - Not | batch | 10 | Gram | 90 | 0 | <= 100 | Histamine | 90 | 6 |
| | Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | (food/fee d) | | | | | >100 TO <= 200 | Histamine | 90 | 0 |
| | | | | | | | >200 | Histamine | 90 | 0 |
| | Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - Retail - Spain | batch | 10 | Gram | 9 | 0 | <= 100 | Histamine | 9 | 0 |
| | - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | (food/fee d) | | | | | >100 TO <= 200 | Histamine | 9 | 0 |
| _ | | | | | | | >200 | Histamine | 9 | 0 |
| | Fish - raw - frozen - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official | single | 10 | Gram | 1 | 0 | <= 100 | Histamine | 1 | 0 |
| | sampling - Selective sampling | (food/fee d) | | | | | >100 TO <= 200 | Histamine | 1 | 0 |
| | | | | | | | >200 | Histamine | 1 | 0 |
| | Fish - raw - frozen - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - | batch | 10 | Gram | 9 | 9 | <= 100 | Histamine | 9 | 0 |
| | Surveillance - based on Regulation 2073 - Official sampling - Selective sampling | (food/fee d) | | | | | >100 TO <= 200 | Histamine | 9 | 0 |
| | | | | | | | >200 | Histamine | 9 | 9 |
| | Fish - raw - frozen - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - | single | 10 | Gram | 2 | 0 | <= 100 | Histamine | 2 | 0 |
| | Surveillance - Official sampling - Selective sampling | (food/fee d) | | | | | >100 TO <= 200 | Histamine | 2 | 0 |
| | | | | | | | >200 | Histamine | 2 | 0 |
| | Fish - raw - frozen - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | single | 10 | Gram | 4 | 0 | <= 100 | Histamine | 4 | 2 |
| | | (food/fee d) | | | | | >100 TO <= 200 | Histamine | 4 | 0 |
| | | | | | | | >200 | Histamine | 4 | 0 |
| | Fish - raw - frozen - Wholesale - Not Available - food sample - Surveillance - Official sampling - Selective sampling | single | 10 | Gram | 2 | 0 | <= 100 | Histamine | 2 | 0 |
| | | (food/fee d) | | | | | >100 TO <= 200 | Histamine | 2 | 0 |
| | | | | | | | >200 | Histamine | 2 | 0 |
| | Fish - raw - frozen - Wholesale - Spain - food sample - Surveillance - based on Regulation 2073 - Official sampling - | batch | 10 | Gram | 18 | 0 | <= 100 | Histamine | 18 | 1 |
| | Selective sampling | (food/fee d) | | | | | >100 TO <= 200 | Histamine | 18 | 0 |
| | | | | | | | >200 | Histamine | 18 | 0 |
| | Meat from bovine animals - fresh - frozen - Retail - Brazil - food sample - Surveillance - Official sampling - Selective sampling | single (food/fee d) | 10 | Gram | 1 | 0 | <= 100 | Histamine | 1 | 0 |
| | Meat from bovine animals - fresh - frozen - Wholesale - Brazil - food sample - Surveillance - Official sampling - Selective | single | 10 | Gram | 3 | 0 | <= 100 | Histamine | 3 | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | weight unit | units tested | units positive | Method | Zoonoses | N of units tested | N of units positive |
|------------------|--|---------------------------|------------------|----------------|-----------------|-------------------|--------|-----------|-------------------|---------------------|
| Not Available | Meat from broilers (Gallus gallus) - fresh - frozen - Wholesale - Brazil - food sample - Surveillance - Official sampling - Selective sampling | single (food/fee d) | 10 | Gram | 4 | 0 | <= 100 | Histamine | 4 | 0 |
| | Meat from poultry, unspecified - offal - unspecified - frozen - Retail - Brazil - food sample - Surveillance - Official sampling - Selective sampling | single (food/fee d) | 10 | Gram | 1 | 0 | <= 100 | Histamine | 1 | 0 |
| | Meat from poultry, unspecified - offal - unspecified - frozen - Wholesale - Brazil - food sample - Surveillance - Official sampling - Selective sampling | single (food/fee d) | 10 | Gram | 5 | 0 | <= 100 | Histamine | 5 | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|------------------|--|---------------------------|------------------|--------------------------|--------------------------|----------------------------|-------------------|---------------------------------------|-------------------|---------------------|
| Not Available | Bakery products - desserts - containing heat-treated cream - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Bakery products - desserts - containing raw eggs - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 30 | 0 | <= 100 | Listeria monocytogenes | 30 | 0 |
| | Discount to describe Described a Octoor Described a | | 05 | 0 | 1 | | >100 | Listeria monocytogenes | 30 | 0 |
| | Bakery products - desserts - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 1 | 0 | <= 100 >100 | Listeria monocytogenes | 1 | 0 |
| | Delega anadusta, descerta Detail Net Augilable, food comple. Curvillance, based on Devulation 2072. Official | batch | 25 | Gram | 60 | 0 | <= 100 | Listeria monocytogenes | 1 | 0 |
| | Bakery products - desserts - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | (food/fee d) | 25 | Gram | 60 | U | >100 | Listeria monocytogenes Listeria | 60 | 0 |
| | Cheeses made from cows' milk - fresh - made from pasteurised milk - Processing plant - Portugal - food sample - | single | 25 | Gram | 44 | 0 | <= 100 | monocytogenes Listeria | 60 | 0 |
| | Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | (food/fee d) | 25 | Giaili | 44 | U | >100 | monocytogenes Listeria | 44 | 0 |
| | Cheeses made from cows' milk - fresh - made from pasteurised milk - Processing plant - Portugal - food sample - | single | 25 | Gram | 44 | 0 | detection | monocytogenes Listeria | 44 | 0 |
| | Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | (food/fee d) | | Glaili | 77 | Ů | detection | monocytogenes | 44 | 0 |
| | Cheeses made from cows' milk - fresh - made from pasteurised milk - Retail - Germany - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from cows' milk - fresh - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 10 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | Change made from a well will, head made from restauried will. Descension plant Destroyle food comple | | 25 | Gram | 10 | 0 | >100 detection | Listeria monocytogenes Listeria | 10 | 0 |
| | Cheeses made from cows' milk - hard - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 10 | U | detection | monocytogenes | 10 | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee | 25 | Gram | 26 | 0 | <= 100 | Listeria monocytogenes | 26 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 26 | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | | Gram | 26 | 0 | detection | Listeria monocytogenes | 26 | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Retail - Italy - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 30 | 0 | <= 100 | Listeria monocytogenes | 30 | 0 |
| | | | | | | | >100 | Listeria monocytogenes | 30 | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Retail - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | 05 | 0 | _ | • | >100 | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Retail - Switzerland - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 5 | 0 | <= 100 >100 | Listeria monocytogenes | 5 | 0 |
| | Change made from equal wills, soft and easy only made from the change of | | 25 | Owe | 20 | 0 | | Listeria monocytogenes | 5 | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 26 | 0 | <= 100 >100 | Listeria monocytogenes | 26 | 0 |
| | | -, | | | | | >100 | Listeria monocytogenes | 26 | 0 |

| Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | | Sample weight unit | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|--|---------------------------|-----|--------------------------|--------------------------|----------------------------|-------------------|---------------------------------------|-------------------|---------------------|
| Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 26 | 0 | detection | Listeria monocytogenes | 26 | 0 |
| Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 30 | 0 | <= 100 | Listeria monocytogenes | 30 | 0 |
| | d) | | | | | >100 | Listeria monocytogenes | 30 | 0 |
| Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 10 | 0 | <= 100 >100 | Listeria monocytogenes Listeria | 10 | 0 |
| Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Portugal - | | 25 | Gram | 10 | 0 | detection | monocytogenes Listeria | 10 | 0 |
| food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | (food/fee d) | 20 | Ciuiii | 10 | | detection | monocytogenes | 10 | 0 |
| Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Italy - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 10 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| Characteristic Control of the Contro | d) | 0.5 | 0 | 40 | 0 | >100 | Listeria monocytogenes | 10 | 0 |
| Cheeses made from goats' milk - fresh - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 12 | U | <= 100 >100 | Listeria monocytogenes Listeria | 12 | 0 |
| Cheeses made from goats' milk - fresh - made from pasteurised milk - Processing plant - Portugal - food sample - | single | 25 | Gram | 12 | 0 | detection | monocytogenes Listeria | 12 | 0 |
| Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | (food/fee d) | | | | | | monocytogenes | 12 | 0 |
| Cheeses made from goats' milk - hard - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 2 | 0 | <= 100 | Listeria monocytogenes | 2 | 0 |
| Change made from postel will, hard, made from poster wind will. Desposing along Darluge Lind comple | | 25 | Cross | 2 | 0 | >100 | Listeria monocytogenes | 2 | 0 |
| Cheeses made from goats' milk - hard - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 2 | 0 | detection | Listeria monocytogenes | 2 | 0 |
| Cheeses made from goats' milk - hard - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 10 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | d) | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| Cheeses made from goats' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 4 | 0 | <= 100 >100 | Listeria monocytogenes Listeria | 4 | 0 |
| Cheeses made from goats' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Portugal - food | single | 25 | Gram | 4 | 0 | detection | monocytogenes Listeria | 4 | 0 |
| sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | (food/fee d) | | | · | | | monocytogenes | 4 | 0 |
| Cheeses made from goats' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 2 | 0 | <= 100 | Listeria monocytogenes | 2 | 0 |
| Cheeses made from goats' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Portugal - | | 25 | Cross | 2 | 0 | >100 detection | Listeria monocytogenes Listeria | 2 | 0 |
| food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 2 | U | detection | monocytogenes | 2 | 0 |
| Cheeses made from goats' milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | d) | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| Cheeses made from sheep's milk - fresh - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 17 | 0 | <= 100 | Listeria monocytogenes | 17 | 0 |
| Cheeses made from sheep's milk - fresh - made from pasteurised milk - Processing plant - Portugal - food sample - | single | 25 | Gram | 17 | 0 | >100 detection | Listeria monocytogenes Listeria | 17 | 0 |
| Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | (food/fee d) | 20 | Ciam | | | JOLOGIOIT | monocytogenes | 17 | 0 |
| Cheeses made from sheep's milk - hard - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee | 25 | Gram | 2 | 0 | <= 100 | Listeria monocytogenes | 2 | 0 |
| | d) | | | | | >100 | Listeria monocytogenes | 2 | 0 |

| oling Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---|-------------------------------|------------------|------|--------------------------|----------------------------|----------------|---------------------------------------|-------------------|---------------------|
| Cheeses made from sheep's milk - hard - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 2 | 0 | detection | Listeria monocytogenes | 2 | 0 |
| Cheeses made from sheep's milk - hard - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee | 25 | Gram | 3 | 0 | <= 100 | Listeria monocytogenes | 3 | 0 |
| | d) | | | | | >100 | Listeria monocytogenes | 3 | 0 |
| Cheeses made from sheep's milk - hard - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | | Gram | 3 | 0 | detection | Listeria monocytogenes | 3 | 0 |
| Cheeses made from sheep's milk - hard - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - Official sampling - Selective sampling | batch (food/fee d) | 25 | Gram | 35 | 0 | <= 100 >100 | Listeria monocytogenes Listeria | 35 | 0 |
| | ۵, | | | | | >100 | monocytogenes | 35 | 0 |
| Cheeses made from sheep's milk - hard - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - Official sampling - Selective sampling | batch (food/fee d) | 25 | Gram | 35 | 0 | detection | Listeria monocytogenes | 35 | 0 |
| Cheeses made from sheep's milk - hard - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 10 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | d) | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 55 | 2 | <= 100 >100 | Listeria monocytogenes Listeria | 55 | 2 |
| | | | | | | 1.00 | monocytogenes | 55 | 0 |
| Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 55 | 2 | detection | Listeria monocytogenes | 55 | 2 |
| Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - France - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 15 | 0 | <= 100 | Listeria monocytogenes | 15 | 0 |
| | d) | | | | | >100 | Listeria monocytogenes | 15 | 0 |
| Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - Official sampling - Selective sampling | single (food/fee d) | 25 | Gram | 1 | 0 | <= 100 >100 | Listeria monocytogenes Listeria | 1 | 0 |
| | <u> </u> | | | | | | monocytogenes | 1 | 0 |
| Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - Official sampling - Selective sampling | single (food/fee d) | 25 | Gram | 1 | 0 | detection | Listeria monocytogenes | 1 | 0 |
| Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 95 | 11 | <= 100 | Listeria monocytogenes | 95 | 2 |
| | | | | | | >100 | Listeria monocytogenes | 95 | 9 |
| Cheeses, made from mixed milk from cows, sheep and/or goats - fresh - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | - single (food/fee d) | 25 | Gram | 3 | 1 | <= 100 | Listeria monocytogenes | 3 | 1 |
| | | | | | | >100 | Listeria monocytogenes | 3 | 0 |
| Cheeses, made from mixed milk from cows, sheep and/or goats - fresh - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | - single (food/fee d) | 25 | Gram | 3 | 1 | detection | Listeria monocytogenes | 3 | 1 |
| Cheeses, made from mixed milk from cows, sheep and/or goats - hard - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | - batch (food/fee d) | 25 | Gram | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | u) | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| Cheeses, made from mixed milk from cows, sheep and/or goats - soft and semi-soft - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective | single (food/fee | 25 | Gram | 4 | 0 | <= 100 | Listeria monocytogenes | 4 | 0 |
| sampling | d) | | | | | >100 | Listeria monocytogenes | 4 | 0 |
| Cheeses, made from mixed milk from cows, sheep and/or goats - soft and semi-soft - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 4 | 0 | detection | Listeria monocytogenes | 4 | 0 |
| Cheeses, made from mixed milk from cows, sheep and/or goats - soft and semi-soft - made from pasteurised milk - Retai Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | il - batch (food/fee d) | 25 | Gram | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | u) | | | | | >100 | Listeria monocytogenes | 5 | 0 |

| Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|--|---------------------------|------------------|--------------------------|--------------------------|----------------------------|----------------|---------------------------------------|-------------------|---------------------|
| Cheeses, made from mixed milk from cows, sheep and/or goats - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective | single (food/fee | 25 | Gram | 10 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| sampling | d) | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| Cheeses, made from mixed milk from cows, sheep and/or goats - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 10 | 0 | detection | Listeria monocytogenes | 10 | 0 |
| Cheeses, made from mixed milk from cows, sheep and/or goats - soft and semi-soft - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 10 | 0 | <= 100 >100 | Listeria monocytogenes Listeria | 10 | 0 |
| Crustaceans - prawns - cooked - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official | batch | 25 | Gram | 5 | 0 | <= 100 | monocytogenes Listeria | 10 | 0 |
| sampling - Objective sampling | (food/fee d) | 20 | Olalli | J | o . | >100 | monocytogenes Listeria | 5 | 0 |
| Crustaceans - prawns - shelled, shucked and cooked - frozen - Border inspection activities - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| Crustaceans - shrimps - cooked - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - | d) single | 25 | Gram | 2 | 0 | detection | Listeria | | |
| Official sampling - Objective sampling Crustaceans - shrimps - cooked - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official | (food/fee d) batch | 25 | Gram | 20 | 0 | <= 100 | monocytogenes Listeria | 2 | 0 |
| sampling - Objective sampling | (food/fee d) | | | | | >100 | monocytogenes Listeria | 20 | 0 |
| Crustaceans - unspecified - cooked - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 3 | 0 | detection | Listeria monocytogenes | 3 | 0 |
| Dairy products (excluding cheeses) - dairy desserts - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 55 | 0 | <= 100 | Listeria monocytogenes | 55 | 0 |
| | d) | | | | | >100 | Listeria monocytogenes | 55 | 0 |
| Dairy products (excluding cheeses) - dairy desserts - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 4 | 0 | <= 100 | Listeria monocytogenes | 4 | 0 |
| | , | | | | | >100 | Listeria monocytogenes | 4 | 0 |
| Dairy products (excluding cheeses) - ice-cream - made from pasteurised milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 90 | 0 | <= 100 >100 | Listeria monocytogenes Listeria | 90 | 0 |
| Dairy products (excluding cheeses) - milk powder and whey powder - Retail - Not Available - food sample - Surveillance - | batch | 25 | Gram | 20 | 0 | <= 100 | monocytogenes Listeria | 90 | 0 |
| based on Regulation 2073 - Official sampling - Objective sampling | (food/fee d) | 25 | Ciaiii | 20 | · · | >100 | monocytogenes Listeria | 20 | 0 |
| Fish - smoked - Border inspection activities - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 5 | 0 | detection | monocytogenes Listeria monocytogenes | 5 | 0 |
| Fruits - pre-cut - ready-to-eat - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | d) single (food/fee | | Gram | 38 | 0 | <= 100 | Listeria monocytogenes | 38 | 0 |
| | d) | | | | | >100 | Listeria monocytogenes | 38 | 0 |
| Fruits - pre-cut - ready-to-eat - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee d) | 25 | Gram | 38 | 0 | detection | Listeria monocytogenes | 38 | 0 |
| Fruits - pre-cut - ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 2 | 0 | <= 100 | Listeria monocytogenes | 2 | 0 |
| | | | | | | >100 | Listeria monocytogenes | 2 | 0 |
| Fruits - pre-cut - ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 2 | 0 | detection | Listeria monocytogenes | 2 | 0 |
| Fruits - pre-cut - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 25 | 0 | <= 100 | Listeria monocytogenes | 25 | 0 |
| | u) | | | | | >100 | Listeria monocytogenes | 25 | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|------------------|--|---------------------------|------------------|--------------------------|--------------------------|----------------------------|-------------------|---------------------------------------|-------------------|---------------------|
| Not Available | Infant formula - ready-to-eat - Hospital or medical care facility - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee | 1 | Millilitre | 17 | 0 | <= 100 | Listeria monocytogenes | 17 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 17 | 0 |
| | Infant formula - ready-to-eat - Hospital or medical care facility - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee d) | 25 | Gram | 17 | 0 | detection | Listeria monocytogenes | 17 | 0 |
| | Juice - fruit juice - pasteurised - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee | 25 | Millilitre | 2 | 0 | <= 100 | Listeria monocytogenes | 2 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 2 | 0 |
| | Juice - fruit juice - pasteurised - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Millilitre | 2 | 0 | detection | Listeria monocytogenes | 2 | 0 |
| | Juice - fruit juice - unpasteurised - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee | 25 | Millilitre | 4 | 0 | <= 100 | Listeria monocytogenes | 4 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 4 | 0 |
| | Juice - fruit juice - unpasteurised - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Millilitre | 4 | 0 | detection | Listeria monocytogenes | 4 | 0 |
| | Juice - vegetable juice - unpasteurised - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee | 25 | Millilitre | 2 | 0 | <= 100 | Listeria monocytogenes | 2 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 2 | 0 |
| | Juice - vegetable juice - unpasteurised - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Millilitre | 2 | 0 | detection | Listeria monocytogenes | 2 | 0 |
| | Meat from bovine animals - meat products - raw and intended to be eaten raw - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - chilled - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 10 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | Mark from he iller (Orline at line) and and a head and the set December 1 for december 1 | | | 0 | | | >100 | Listeria monocytogenes | 10 | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 6 | 0 | <= 100 >100 | Listeria monocytogenes Listeria | 6 | 0 |
| | Most from brailers (Cellus gallus), most products, pooled, rough to get. Dressoning plant. Portugal, food gamela | | 25 | Crom | 6 | 0 | | monocytogenes Listeria | 6 | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | | | detection | monocytogenes | 6 | 0 |
| | Meat from pig - meat products - cooked ham - sliced - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee d) | | Gram | 4 | 1 | <= 100 | Listeria monocytogenes | 4 | 0 |
| | | | | | | | >100 | Listeria monocytogenes | 4 | 0 |
| | Meat from pig - meat products - cooked ham - sliced - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee d) | 25 | Gram | 4 | 1 | detection | Listeria monocytogenes | 4 | 1 |
| | Meat from pig - meat products - cooked ham - sliced - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 20 | 0 | <= 100 | Listeria monocytogenes | 20 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 20 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - chilled - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | Market and an advantage of the second | | 0.5 | 0 | 40.1 | 40 | >100 | Listeria monocytogenes | 5 | 0 |
| | Meat from pig - meat products - cooked, ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 131 | 13 | <= 100 | Listeria monocytogenes | 131 | 12 |
| | Meat from pig - meat products - cooked, ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on | | 25 | Gram | 131 | 13 | >100 detection | Listeria monocytogenes Listeria | 131 | 1 |
| | Meat from pig - meat products - cooked, feady-to-eat - Processing plant - Portugal - tood sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 20 | Giaili | 131 | 13 | uetection | monocytogenes | 131 | 13 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|------------------|--|---------------------------|------------------|--------------------------|--------------------------|----------------------------|-------------------|---------------------------------------|-------------------|---------------------|
| Not Available | Meat from pig - meat products - fermented sausages - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Selective sampling | batch (food/fee d) | 25 | Gram | 15 | 14 | detection | Listeria monocytogenes | 15 | 14 |
| | Meat from pig - meat products - fermented sausages - Processing plant - Not Available - food sample - Surveillance - Official sampling - Selective sampling | single (food/fee | 25 | Gram | 2 | 0 | <= 100 | Listeria monocytogenes | 2 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 2 | 0 |
| | Meat from pig - meat products - fermented sausages - Processing plant - Not Available - food sample - Surveillance - Official sampling - Selective sampling | single (food/fee d) | 25 | Gram | 2 | 0 | detection | Listeria monocytogenes | 2 | 0 |
| | Meat from pig - meat products - fermented sausages - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 10 | 2 | <= 100 | Listeria monocytogenes | 10 | 2 |
| | | | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| | Meat from pig - meat products - raw but intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 5 | 0 | <= 100 >100 | Listeria monocytogenes Listeria | 5 | 0 |
| | Meat from pig - meat products - raw but intended to be eaten cooked - Processing plant - Portugal - food sample - | single | 25 | Gram | 5 | 0 | detection | monocytogenes Listeria | 5 | 0 |
| | Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | (food/fee d) | | | | | | monocytogenes | 5 | 0 |
| | Meat from pig - meat products - raw ham - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 10 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | Meat from pig - meat products - unspecified, ready-to-eat - Processing plant - Not Available - food sample - Surveillance - | batch | 25 | Gram | 5 | 0 | >100 detection | Listeria monocytogenes Listeria | 10 | 0 |
| | based on Regulation 2073 - Official sampling - Selective sampling | (food/fee d) | | | | | | monocytogenes | 5 | 0 |
| | Meat from pig - meat products - unspecified, ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Meat from pig - offal - Processing plant - Not Available - food sample - Surveillance - Official sampling - Selective sampling | batch (food/fee d) | 25 | Gram | 10 | 5 | detection | Listeria monocytogenes | 10 | 5 |
| | Meat from turkey - meat products - cooked, ready-to-eat - chilled - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 40 | 0 | <= 100 | Listeria monocytogenes | 40 | 0 |
| | Markfrondular, markerdular, called radius, ast Pressain plant Patrical food carries Consillance, board | | 25 | Crom | 8 | 0 | >100 | Listeria monocytogenes | 40 | 0 |
| | Meat from turkey - meat products - cooked, ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 0 | U | <= 100 >100 | Listeria monocytogenes Listeria | 8 | 0 |
| | Meat from turkey - meat products - cooked, ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based | single | 25 | Gram | 8 | 0 | detection | monocytogenes Listeria | 8 | 0 |
| | on Regulation 2073 - Official sampling - Objective sampling | (food/fee d) | | Crare | 4 | 0 | <= 100 | monocytogenes Listeria | 8 | 0 |
| | Meat, mixed meat - meat products - cooked, ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 4 | U | >100 | monocytogenes Listeria | 4 | 0 |
| | Meat, mixed meat - meat products - cooked, ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based | single | 25 | Gram | 4 | 0 | detection | monocytogenes Listeria | 4 | 0 |
| | on Regulation 2073 - Official sampling - Objective sampling | (food/fee d) | | | | | | monocytogenes | 4 | 0 |
| | Molluscan shellfish - cooked - frozen - Border inspection activities - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 25 | 0 | detection | Listeria monocytogenes | 25 | 0 |
| | Other food of non-animal origin - Processing plant - Portugal - food sample - Surveillance - Industry sampling - Objective sampling | single (food/fee d) | 25 | Gram | 4 | 0 | detection | Listeria monocytogenes | 4 | 0 |
| | Other processed food products and prepared dishes - egg based dishes - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective | batch (food/fee | 25 | Gram | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | sampling | d) | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Other processed food products and prepared dishes - fish and seafood based dishes - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 75 | 1 | <= 100 | Listeria monocytogenes | 75 | 1 |
| | Cojouri Coumpany | u, | | | | | >100 | Listeria monocytogenes | 75 | 0 |

| ampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|---------|--|---------------------------|------------------|--------------------------|--------------------------|----------------------------|-----------|---------------------------|-------------------|---------------------|
| able | Other processed food products and prepared dishes - meat based dishes - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective | batch (food/fee | 25 | Gram | 90 | 0 | <= 100 | Listeria monocytogenes | 90 | 0 |
| | sampling | d) | | | | | >100 | Listeria monocytogenes | 90 | 0 |
| | Other processed food products and prepared dishes - meat based dishes - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - Official sampling - Selective sampling | single (food/fee | 25 | Gram | 4 | 0 | <= 100 | Listeria monocytogenes | 4 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 4 | 0 |
| | Other processed food products and prepared dishes - sandwiches - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee | | Gram | 30 | 0 | <= 100 | Listeria monocytogenes | 30 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 30 | 0 |
| | Other processed food products and prepared dishes - sandwiches - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee d) | 25 | Gram | 30 | 0 | detection | Listeria monocytogenes | 30 | 0 |
| | Other processed food products and prepared dishes - sandwiches - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 10 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| | Other processed food products and prepared dishes - sandwiches - with meat - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective | batch (food/fee | 25 | Gram | 50 | 0 | <= 100 | Listeria monocytogenes | 50 | 0 |
| | sampling | d) | | | | | >100 | Listeria monocytogenes | 50 | 0 |
| | Other processed food products and prepared dishes - unspecified - ready-to-eat foods - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee | | Gram | 14 | 0 | <= 100 | Listeria monocytogenes | 14 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 14 | 0 |
| | | | | | 1012 | 2 | <= 100 | Listeria monocytogenes | 1,012 | 0 |
| | | | | | | | >100 | Listeria monocytogenes | 1,012 | 0 |
| | Other processed food products and prepared dishes - unspecified - ready-to-eat foods - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee d) | 25 | Gram | 1012 | 2 | detection | Listeria monocytogenes | 1,012 | 2 |
| | Other processed food products and prepared dishes - unspecified - ready-to-eat foods - frozen - Processing plant - Portugal - food sample - Surveillance - Industry sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 5 | 0 | detection | Listeria monocytogenes | 5 | 0 |
| | Other processed food products and prepared dishes - unspecified - ready-to-eat foods - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Other processed food products and prepared dishes - vegetable based dishes - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee | | Gram | 249 | 3 | <= 100 | Listeria monocytogenes | 249 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 249 | 0 |
| | Other processed food products and prepared dishes - vegetable based dishes - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee d) | 25 | Gram | 249 | 3 | detection | Listeria monocytogenes | 249 | 3 |
| | Other processed food products and prepared dishes - vegetable based dishes - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective | batch (food/fee | 25 | Gram | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | sampling | d) | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Ready-to-eat salads - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee | | Gram | 110 | 3 | <= 100 | Listeria monocytogenes | 110 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 110 | 0 |
| | Ready-to-eat salads - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee d) | 25 | Gram | 110 | 3 | detection | Listeria monocytogenes | 110 | 3 |
| | Ready-to-eat salads - containing mayonnaise - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 15 | 0 | <= 100 | Listeria monocytogenes | 15 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 15 | 0 |

| npling N | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Total units tested | Total units positive | Method | Zoonoses | N of units tested | N of units positive |
|----------|--|---------------------------|------------------|--------------------------|--------------------------|----------------------------|-----------|---------------------------|-------------------|---------------------|
| | Ready-to-eat salads - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 5 | 0 | <= 100 | Listeria monocytogenes | 5 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 5 | 0 |
| | Ready-to-eat salads - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - Official sampling - Selective sampling | single (food/fee | 25 | Gram | 1 | 0 | <= 100 | Listeria monocytogenes | 1 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 1 | 0 |
| | Ready-to-eat salads - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 30 | 0 | <= 100 | Listeria monocytogenes | 30 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 30 | 0 |
| | Sauce and dressings - mayonnaise - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 35 | 0 | <= 100 | Listeria monocytogenes | 35 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 35 | 0 |
| | Sauce and dressings - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 95 | 0 | <= 100 | Listeria monocytogenes | 95 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 95 | 0 |
| | Seeds, sprouted - ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee | 25 | Gram | 4 | 0 | <= 100 | Listeria monocytogenes | 4 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 4 | 0 |
| | Seeds, sprouted - ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 4 | 0 | detection | Listeria monocytogenes | 4 | 0 |
| | Seeds, sprouted - ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 15 | 0 | <= 100 | Listeria monocytogenes | 15 | 0 |
| _ | | d) | | | | | >100 | Listeria monocytogenes | 15 | 0 |
| | Soups - ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 40 | 0 | <= 100 | Listeria monocytogenes | 40 | 0 |
| _ | | d) | | | | | >100 | Listeria monocytogenes | 40 | 0 |
| | Spices and herbs - dried - irradiated - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 10 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| _ | | d) | | | | | >100 | Listeria monocytogenes | 10 | 0 |
| | Surimi - frozen - Border inspection activities - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | 15 | 0 | detection | Listeria monocytogenes | 15 | 0 |
| | Vegetables - pre-cut - ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee | 25 | Gram | 12 | 0 | <= 100 | Listeria monocytogenes | 12 | 0 |
| | | d) | | | | | >100 | Listeria monocytogenes | 12 | 0 |
| | Vegetables - pre-cut - ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | 12 | 0 | detection | Listeria monocytogenes | 12 | 0 |
| | Vegetables - pre-cut - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee | 25 | Gram | 10 | 0 | <= 100 | Listeria monocytogenes | 10 | 0 |
| | | d) | | | | | >100 | Listeria | 10 | 0 |

Table Lyssavirus:LYSSAVIRUS in animal

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Method | Sampling unit | units | Total units positive | Zoonoses | N of units positive |
|------------------|---|---------------|------------------|-------|----------------------------|------------|---------------------|
| PORTUGAL | Dogs - Official kennel - Not Available - animal sample - Surveillance - Official sampling - Suspect sampling | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Foxes - wild - Natural habitat - Not Available - animal sample - Surveillance - Official sampling - Convenient sampling | Not Available | animal | 1 | 0 | Lyssavirus | 0 |
| | Foxes - wild - Natural habitat - Not Available - animal sample - Surveillance - Official sampling - Suspect sampling | Not Available | animal | 1 | 0 | Lyssavirus | 0 |

Table Salmonella:SALMONELLA in animal

| ing N | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | N of flocks under contro programme | Target verification | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|-------|---|----------------|--|---------------------|---------------|--------------------|----------------------------|---|--|
| | Gallus gallus (fowl) - breeding flocks for broiler production line - adult - Farm - Portugal - Not Available - Control and | herd/floc | 476 | Υ | Not Available | 476 | 3 | Salmonella 1,4,12:i:- | 2 |
| | eradication programmes - Official and industry sampling - Census | k | | | | | | Salmonella Kirkee | 1 |
| | Gallus gallus (fowl) - breeding flocks for egg production line - adult - Farm - Portugal - Not Available - Control and eradication programmes - Official and industry sampling - Census | herd/floc k | 22 | Y | Not Available | 22 | 0 | Salmonella | 0 |
| | Gallus gallus (fowl) - broilers - before slaughter - Farm - Portugal - Not Available - Control and eradication programmes - | herd/floc | 10934 | N | Not Available | 10934 | 45 | Salmonella 4,5,12:i:- | 4 |
| | Industry sampling - Census | k | | | | | | Salmonella Anatum | 1 |
| | | | | | | | | Salmonella Brandenburg | 1 |
| | | | | | | | | Salmonella Cerro | 5 |
| | | | | | | | | Salmonella Havana | 11 |
| | | | | | | | | Salmonella Kirkee | 1 |
| | | | | | | | | Salmonella Lexington | 2 |
| | | | | | | | | Salmonella Llandoff | 1 |
| | | | | | | | | Salmonella Madelia | 1 |
| | | | | | | | | Salmonella Newport | 1 |
| | | | | | | | | Salmonella Other serovars | 15 |
| | | | | | | | | Salmonella Typhimurium | 1 |
| | | | | | | | | Salmonella Virchow | 1 |
| | Gallus gallus (fowl) - broilers - before slaughter - Farm - Portugal - Not Available - Control and eradication programmes - | herd/floc | 11011 | Υ | Not Available | 11011 | 51 | Salmonella 4,5,12:i:- | 6 |
| | Official and industry sampling - Census | K | | | | | | Salmonella Anatum | 1 |
| | | | | | | | | Salmonella Brandenburg | 1 |
| | | | | | | | | Salmonella Cerro | 5 |
| | | | | | | | | Salmonella Havana | 15 |
| | | | | | | | | Salmonella Kirkee | 1 |
| | | | | | | | | Salmonella Lexington | 2 |
| | | | | | | | | Salmonella Llandoff | 1 |
| | | | | | | | | Salmonella Madelia | 1 |
| | | | | | | | | Salmonella Newport | 1 |
| | | | | | | | | Salmonella Other serovars | 15 |
| | | | | | | | | Salmonella Typhimurium Salmonella Virchow | 1 1 |
| - | Gallus gallus (fowl) - broilers - before slaughter - Farm - Portugal - Not Available - Control and eradication programmes - | herd/floc | 126 | N | Not Available | 126 | 6 | Salmonella 4,5,12:i:- | 2 |
| | Official sampling - Census | k | 120 | IN . | NOT Available | 120 | U | Salmonella Havana | 4 |
| | Gallus gallus (fowl) - laying hens - adult - Farm - Portugal - Not Available - Control and eradication programmes - Official | herd/floc | 464 | Υ | Not Available | 464 | 39 | Salmonella 4,5,12:i:- | 1 |
| | and industry sampling - Census | k | 707 | • | NOT Available | 404 | 33 | Salmonella Agona | 1 |
| | | | | | | | | Salmonella Brandenburg | 1 |
| | | | | | | | | Salmonella Bredeney | <u>.</u> 1 |
| | | | | | | | | Salmonella Enteritidis | 6 |
| | | | | | | | | Salmonella Infantis | 10 |
| | | | | | | | | Salmonella Llandoff | 2 |
| | | | | | | | | Salmonella Mbandaka | 6 |
| | | | | | | | | | |
| | | | | | | | | | |
| | | | | | | | | Salmonella Montevideo | 2 |
| | | | | | | | | | 2 |
| | | | | | | | | Salmonella Montevideo Salmonella Newport | 2 |
| | | | | | | | | Salmonella Montevideo Salmonella Newport Salmonella Ohio | 2 1 1 |
| | | | | | | | | Salmonella Montevideo Salmonella Newport Salmonella Ohio Salmonella Ouakam | 2 1 1 1 |
| | | | | | | | | Salmonella Montevideo Salmonella Newport Salmonella Ohio Salmonella Ouakam Salmonella Senftenberg | 2 1 1 1 1 |
| | | | | | | | | Salmonella Montevideo Salmonella Newport Salmonella Ohio Salmonella Ouakam Salmonella Senftenberg Salmonella Tennessee | 2 1 1 1 1 |
| | Turkeys - fattening flocks - before slaughter - Farm - Portugal - Not Available - Control and eradication programmes - | herd/floc | 1196 | N | Not Available | 1196 | 5 | Salmonella Montevideo Salmonella Newport Salmonella Ohio Salmonella Ouakam Salmonella Senftenberg Salmonella Tennessee Salmonella Typhimurium | 2 1 1 1 1 1 1 2 |
| | Turkeys - fattening flocks - before slaughter - Farm - Portugal - Not Available - Control and eradication programmes - Industry sampling - Census | herd/floc k | 1196 | N | Not Available | 1196 | 5 | Salmonella Montevideo Salmonella Newport Salmonella Ohio Salmonella Ouakam Salmonella Senftenberg Salmonella Tennessee Salmonella Typhimurium Salmonella Virchow | 2 1 1 1 1 1 2 2 |
| | | herd/floc k | 1196 | N | Not Available | 1196 | 5 | Salmonella Montevideo Salmonella Newport Salmonella Ohio Salmonella Ouakam Salmonella Senftenberg Salmonella Tennessee Salmonella Typhimurium Salmonella Virchow Salmonella | 2 1 1 1 1 1 2 2 5 |
| | Industry sampling - Census Turkeys - fattening flocks - before slaughter - Farm - Portugal - Not Available - Control and eradication programmes - | herd/floc k | 1196 | N | Not Available | 1196 | 5 | Salmonella Montevideo Salmonella Newport Salmonella Ohio Salmonella Ouakam Salmonella Senftenberg Salmonella Tennessee Salmonella Typhimurium Salmonella Virchow Salmonella Salmonella 4,5,12:i:- | 2 1 1 1 1 1 2 2 5 |
| | Industry sampling - Census | k | | | | | | Salmonella Montevideo Salmonella Newport Salmonella Ohio Salmonella Ouakam Salmonella Senftenberg Salmonella Tennessee Salmonella Typhimurium Salmonella Virchow Salmonella Salmonella 4,5,12:i- Salmonella Cerro | 2 1 1 1 1 1 2 2 5 4 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|------------------|---|---------------------------|------------------|-----------------------|---|--------------------------|----------------------------|------------------------------|---------------------|
| Not Available | Bakery products - desserts - containing raw cream - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Bakery products - desserts - containing raw eggs - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 30 | 0 | Salmonella | 0 |
| | Bakery products - desserts - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 25 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - fresh - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 14 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 26 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Retail - Italy - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 30 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Retail - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Retail - Switzerland - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 11 | 0 | Salmonella | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 10 | 0 | Salmonella | 0 |
| | Cheeses made from goats' milk - fresh - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 12 | 0 | Salmonella | 0 |
| | Cheeses made from goats' milk - hard - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 2 | 0 | Salmonella | 0 |
| | Cheeses made from goats' milk - hard - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 10 | 0 | Salmonella | 0 |
| | Cheeses made from goats' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 4 | 1 | Salmonella spp., unspecified | 1 |
| | Cheeses made from goats' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 2 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - fresh - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 17 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - hard - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 2 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - hard - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 2 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 55 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Selective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 40 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - France - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 15 | 0 | Salmonella | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 100 | 0 | Salmonella | 0 |
| | | | | | | | | | |

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| a of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|---------------|--|---------------------------|---------------|-----------------------|---|--------------------------|----------------------------|----------------------------|---------------------|
| t Available | Cheeses, made from mixed milk from cows, sheep and/or goats - fresh - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 3 | 0 | Salmonella | 0 |
| | Cheeses, made from mixed milk from cows, sheep and/or goats - hard - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Cheeses, made from mixed milk from cows, sheep and/or goats - soft and semi-soft - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 4 | 0 | Salmonella | 0 |
| | Cheeses, made from mixed milk from cows, sheep and/or goats - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 10 | 1 | Salmonella IIIb 61:k:1,5,7 | 1 |
| | Cheeses, made from mixed milk from cows, sheep and/or goats - soft and semi-soft - made from raw or low heat-treated milk - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 15 | 0 | Salmonella | 0 |
| | Crustaceans - prawns - cooked - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Crustaceans - prawns - shelled, shucked and cooked - frozen - Border inspection activities - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | ISO 6579:2002/Amd 1:2007 | 5 | 0 | Salmonella | 0 |
| | Crustaceans - shrimps - cooked - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 2 | 0 | Salmonella | 0 |
| | Crustaceans - unspecified - cooked - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 3 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - dairy desserts - chilled - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - dairy desserts - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - Official sampling - Selective sampling | single (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 1 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - milk powder and whey powder - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 5 | 0 | Salmonella | 0 |
| | Dairy products (excluding cheeses) - milk powder and whey powder - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 20 | 0 | Salmonella | 0 |
| | Egg products - liquid - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 6 | 0 | Salmonella | 0 |
| | Egg products - ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 15 | 0 | Salmonella | 0 |
| | Eggs - table eggs - Packing centre - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 45 | 0 | Salmonella | 0 |
| | Eggs - table eggs - whole - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 10 | 0 | Salmonella | 0 |
| | Fruits - pre-cut - ready-to-eat - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002/Amd 1:2007 | 38 | 0 | Salmonella | 0 |
| | Fruits - pre-cut - ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 2 | 0 | Salmonella | 0 |
| | Infant formula - ready-to-eat - Hospital or medical care facility - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002/Amd 1:2007 | 17 | 0 | Salmonella | 0 |
| | Juice - fruit juice - pasteurised - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Millilitre | ISO 6579:2002 Salmonella | 2 | 0 | Salmonella | 0 |
| | Juice - fruit juice - unpasteurised - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Millilitre | ISO 6579:2002 Salmonella | 4 | 0 | Salmonella | 0 |
| | Juice - vegetable juice - unpasteurised - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Millilitre | ISO 6579:2002 Salmonella | 2 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|------------------|---|---------------------------|------------------|-----------------------|---|--------------------------|----------------------------|------------------------------|---------------------|
| Not Available | Live bivalve molluscs - mussels - depurated - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 2 | 0 | Salmonella | 0 |
| | Live bivalve molluscs - mussels - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 1 | 0 | Salmonella | 0 |
| | Live bivalve molluscs - oysters - depurated - Processing plant - Portugal - food sample - Monitoring - HACCP and own check - Objective sampling | batch (food/fee d) | 25 | Gram | ISO 6579:2002/Amd 1:2007 | 5 | 0 | Salmonella | 0 |
| | Live bivalve molluscs - oysters - depurated - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 2 | 0 | Salmonella | 0 |
| | Live bivalve molluscs - oysters - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 4 | 0 | Salmonella | 0 |
| | Live bivalve molluscs - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 35 | 0 | Salmonella | 0 |
| | Live bivalve molluscs - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Selective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 35 | 0 | Salmonella | 0 |
| | Live bivalve molluscs - Retail - Not Available - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 51 | 1 | Salmonella spp., unspecified | 1 |
| | Live bivalve molluscs - Retail - Not Available - food sample - Surveillance - Official sampling - Selective sampling | single (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 1 | 0 | Salmonella | 0 |
| | Live bivalve molluscs - unspecified - depurated - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 8 | 0 | Salmonella | 0 |
| | Meat from bovine animals - carcase - chilled - Slaughterhouse - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 22 | 0 | Salmonella | 0 |
| | Meat from bovine animals - fresh - chilled - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 48 | 0 | Salmonella | 0 |
| | Meat from bovine animals - fresh - frozen - Wholesale - Brazil - food sample - Surveillance - Official sampling - Selective sampling | single (food/fee d) | 10 | Gram | Real-Time PCR (qualitative or quantitative) | 4 | 0 | Salmonella | 0 |
| | Meat from bovine animals - meat preparation - intended to be eaten cooked - chilled - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Selective sampling | batch (food/fee d) | 10 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Meat from bovine animals - meat preparation - intended to be eaten cooked - chilled - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 10 | Gram | Real-Time PCR (qualitative or quantitative) | 25 | 0 | Salmonella | 0 |
| | Meat from bovine animals - meat preparation - intended to be eaten cooked - frozen - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Selective sampling | batch (food/fee d) | 10 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Meat from bovine animals - meat preparation - intended to be eaten cooked - frozen - Retail - Not Available | batch | 10 | Gram | Real-Time PCR | 5 | 2 | Salmonella | 0 |
| | - food sample - Surveillance - based on Regulation 2073 - Official sampling - Selective sampling | (food/fee d) | | | (qualitative or quantitative) | | | Salmonella spp., unspecified | 2 |
| | Meat from bovine animals - meat preparation - intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 23 | 1 | Salmonella Dublin | 1 |
| | Meat from bovine animals - meat products - raw and intended to be eaten raw - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Meat from bovine animals - meat products - raw but intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 1 | 0 | Salmonella | 0 |
| | Meat from bovine animals - minced meat - intended to be eaten cooked - chilled - Processing plant - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Selective sampling | batch (food/fee d) | 10 | Gram | Real-Time PCR (qualitative or quantitative) | 10 | 0 | Salmonella | 0 |
| | Meat from bovine animals - minced meat - intended to be eaten cooked - chilled - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 10 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Meat from bovine animals - minced meat - intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 15 | 0 | Salmonella | 0 |

| g I | | Sampling unit | Sample weight | Sample weight unit | Method | Total units tested | Total units positive | Zoonoses | N of uni positiv |
|---------|---|---------------------------|---------------|-----------------------|---|--------------------------|----------------------------|------------------------------|---------------------|
| П | Meat from bovine animals and pig - meat preparation - intended to be eaten cooked - Retail - Not Available | batch | 10 | Gram | Real-Time PCR | 95 | 16 | Salmonella | 0 |
| | - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | (food/fee d) | | | (qualitative or quantitative) | | | Salmonella 4,5,12:i:- | 1 |
| | | u) | | | quantitative) | | | Salmonella Rissen | 5 |
| | | | | | | | | Salmonella Typhimurium | 10 |
| | Meat from broilers (Gallus gallus) - carcase - Slaughterhouse - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 7 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - fresh - chilled - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 35 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - fresh - frozen - Wholesale - Brazil - food sample - Surveillance - Official sampling - Selective sampling | single (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 4 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - fresh - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 46 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - chilled - Retail - Not | batch | 25 | Gram | Real-Time PCR | 70 | 2 | Salmonella | 0 |
| | Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | (food/fee d) | | | (qualitative or quantitative) | | | Salmonella Typhimurium | 2 |
| | Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 15 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - chilled - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 20 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 8 | 0 | Salmonella | 0 |
| | Meat from broilers (Gallus gallus) - meat products - raw but intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 9 | 0 | Salmonella | 0 |
| | - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch | 25 | Gram | ISO 6579:2002/Amo | 135 | 10 | Salmonella Heidelberg | 9 |
| | | (food/fee d) | | | 1:2007 | | | Salmonella Newport | 1 |
| | Meat from duck - meat products - raw and intended to be eaten raw - chilled - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Meat from goat - carcase - Slaughterhouse - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 1 | 0 | Salmonella | 0 |
| | Meat from goat - fresh - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 7 | 0 | Salmonella | 0 |
| | Meat from pig - carcase - Slaughterhouse - Portugal - food sample - carcase swabs - Surveillance - based on Regulation 2073 - HACCP and own check - Objective sampling | single (food/fee d) | 400 | Square centimetre | Not Available | 3233 | 39 | Salmonella spp., unspecified | 39 |
| | Meat from pig - carcase - Slaughterhouse - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 17 | 1 | Salmonella Typhimurium | 1 |
| | Meat from pig - fresh - Processing plant - Portugal - food sample - Surveillance - Official sampling - | single | 25 | Gram | ISO 6579:2002 | 55 | 4 | Salmonella 4,5,12:i:- | 2 |
| | Objective sampling | (food/fee | | | Salmonella | | | Salmonella Derby | 1 |
| | | d) | | | | | | Salmonella Typhimurium | 1 |
| | Meat from pig - meat preparation - intended to be eaten cooked - chilled - Retail - Not Available - food | batch | 10 | Gram | Real-Time PCR | 70 | 1 | Salmonella | 0 |
| | sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | (food/fee d) | | | (qualitative or quantitative) | | | Salmonella Derby | 1 |
| | Meat from pig - meat preparation - intended to be eaten cooked - chilled - Wholesale - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 10 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| - [V | Meat from pig - meat preparation - intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 41 | 1 | Salmonella Muenchen | 1 |
| | Meat from pig - meat products - cooked ham - sliced - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002/Amd 1:2007 | 4 | 0 | Salmonella | 0 |
| | Meat from pig - meat products - cooked ham - sliced - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 20 | 0 | Salmonella | 0 |

| mpling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|--------|--|---------------------------|---------------|-----------------------|---|--------------------------|----------------------------|---|---------------------|
| ble | Meat from pig - meat products - cooked, ready-to-eat - chilled - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| , | Meat from pig - meat products - cooked, ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee | 25 | Gram | ISO 6579:2002 Salmonella | 138 | 8 | Salmonella 4,12:i:- Salmonella Derby | 2 |
| | | d) | | | | | | Salmonella Muenchen Salmonella Rissen | 1 |
| | | | | | | | | Salmonella spp., unspecified Salmonella Tennessee | 1 |
| | Meat from pig - meat products - fermented sausages - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 30 | 0 | Salmonella | 0 |
| | Meat from pig - meat products - meat specialities - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 20 | 0 | Salmonella | 0 |
| · | Meat from pig - meat products - raw and intended to be eaten raw - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Meat from pig - meat products - raw but intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 72 | 0 | Salmonella | 0 |
| | Meat from pig - meat products - raw ham - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 10 | 0 | Salmonella | 0 |
| | Meat from pig - meat products - unspecified, ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Meat from pig - minced meat - intended to be eaten cooked - chilled - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 15 | 0 | Salmonella | 0 |
| | Meat from pig - minced meat - intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 7 | 0 | Salmonella | 0 |
| | Meat from poultry, unspecified - meat preparation - intended to be eaten cooked - chilled - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 45 | 0 | Salmonella | 0 |
| | Meat from poultry, unspecified - meat preparation - intended to be eaten cooked - frozen - Border inspection activities - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | ISO 6579:2002/Amd 1:2007 | 5 | 0 | Salmonella | 0 |
| • | Meat from poultry, unspecified - meat products - cooked, ready-to-eat - chilled - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Meat from poultry, unspecified - offal - unspecified - frozen - Retail - Brazil - food sample - Surveillance - Official sampling - Selective sampling | single (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 2 | 0 | Salmonella | 0 |
| | Meat from poultry, unspecified - offal - unspecified - frozen - Wholesale - Brazil - food sample - Surveillance - Official sampling - Selective sampling | single (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 4 | 0 | Salmonella | 0 |
| | Meat from sheep - carcase - Slaughterhouse - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 8 | 0 | Salmonella | 0 |
| | Meat from sheep - fresh - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 8 | 0 | Salmonella | 0 |
| : : | Meat from turkey - carcase - Slaughterhouse - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 1 | 0 | Salmonella | 0 |
| | Meat from turkey - fresh - chilled - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Meat from turkey - fresh - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 16 | 0 | Salmonella | 0 |
| | Meat from turkey - meat preparation - intended to be eaten cooked - chilled - Processing plant - Not | batch | 25 | Gram | Real-Time PCR | 5 | 1 | Salmonella | 0 |
| | Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Selective sampling | (food/fee d) | | | (qualitative or quantitative) | | | Salmonella Kentucky | 1 |

| npling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Method | Total units tested | Total units positive | Zoonoses | N of uni positiv |
|--------|--|---------------------------|---------------|-----------------------|---|--------------------------|----------------------------|------------------------------|---------------------|
| le | Meat from turkey - meat preparation - intended to be eaten cooked - chilled - Retail - Not Available - food | batch | 25 | Gram | Real-Time PCR | 90 | 13 | Salmonella | 0 |
| | sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | (food/fee | | | (qualitative or | | | Salmonella Coeln | 5 |
| | | d) | | | quantitative) | | | Salmonella Newport | 5 |
| | | | | | | | | Salmonella spp., unspecified | 3 |
| | Meat from turkey - meat preparation - intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 11 | 0 | Salmonella | 0 |
| | Meat from turkey - meat products - cooked, ready-to-eat - chilled - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 25 | 0 | Salmonella | 0 |
| | Meat from turkey - meat products - cooked, ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 8 | 0 | Salmonella | 0 |
| | Meat from turkey - meat products - raw but intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 1 | 0 | Salmonella | 0 |
| | Meat from turkey - meat products - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 20 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat preparation - intended to be eaten cooked - chilled - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat preparation - intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 9 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat products - cooked, ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 4 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat products - meat specialities - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 40 | 0 | Salmonella | 0 |
| | Meat, mixed meat - meat products - raw but intended to be eaten cooked - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 4 | 0 | Salmonella | 0 |
| | Molluscan shellfish - cooked - frozen - Border inspection activities - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | ISO 6579:2002/Amd 1:2007 | 25 | 0 | Salmonella | 0 |
| | Other food of non-animal origin - Processing plant - Portugal - food sample - Surveillance - Industry sampling - Objective sampling | batch (food/fee d) | 25 | Gram | ISO 6579:2002/Amd 1:2007 | 5 | 0 | Salmonella | 0 |
| | | single (food/fee d) | 25 | Gram | ISO 6579:2002/Amd 1:2007 | 14 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - egg based dishes - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - fish and seafood based dishes - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 20 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - meat based dishes - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 40 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - meat based dishes - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - Official sampling - Selective sampling | (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 3 | 0 | Salmonella | 0 |
| _ | | single (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 1 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - pasta based dishes - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 10 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - Processing plant - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 27 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - sandwiches - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002/Amd 1:2007 | 32 | 0 | Salmonella | 0 |

| rea of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|-----------------|---|---------------------------|------------------|-----------------------|---|--------------------------|----------------------------|------------|---------------------|
| Not Available | Other processed food products and prepared dishes - sandwiches - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 20 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - sandwiches - with meat - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 30 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - unspecified - ready-to-eat foods - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002/Amd 1:2007 | 1030 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - unspecified - ready-to-eat foods - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - vegetable based dishes - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002/Amd 1:2007 | 249 | 0 | Salmonella | 0 |
| | Other processed food products and prepared dishes - vegetable based dishes - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Ready-to-eat salads - Catering - Portugal - food sample - Surveillance - HACCP and own check - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002/Amd 1:2007 | 110 | 0 | Salmonella | 0 |
| | Ready-to-eat salads - containing mayonnaise - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 15 | 0 | Salmonella | 0 |
| | Ready-to-eat salads - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - Official sampling - Selective sampling | single (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 1 | 0 | Salmonella | 0 |
| | Ready-to-eat salads - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Sauce and dressings - mayonnaise - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Sauce and dressings - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 5 | 0 | Salmonella | 0 |
| | Seeds, sprouted - ready-to-eat - Farm - Portugal - food sample - Surveillance - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 4 | 0 | Salmonella | 0 |
| | Seeds, sprouted - ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 15 | 0 | Salmonella | 0 |
| | Vegetables - pre-cut - ready-to-eat - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 12 | 0 | Salmonella | 0 |
| | Vegetables - pre-cut - ready-to-eat - Restaurant or Cafe or Pub or Bar or Hotel or Catering service - Not Available - food sample - Surveillance - Official sampling - Selective sampling | single (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 1 | 0 | Salmonella | 0 |
| | Vegetables - pre-cut - ready-to-eat - Retail - Not Available - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 40 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|------------------|---|--------------------------|------------------|-----------------------|---|--------------------------|----------------------------|------------------------------|---------------------|
| Not Available | Compound feedingstuffs for cattle - final product - Feed mill - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 25 | 2 | Salmonella spp., unspecified | 2 |
| | Compound feedingstuffs for horses - final product - Feed mill - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 1 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for pigs - final product - Feed mill - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | ISO 6579:2002 Salmonella | 34 | 3 | Salmonella spp., unspecified | 3 |
| | Compound feedingstuffs for poultry (non specified) - final product - Feed mill - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 3 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for poultry, broilers - final product - Feed mill - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 20 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for poultry, laying hens - final product - Feed mill - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 24 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for rabbits - final product - Feed mill - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 12 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for sheep - final product - Feed mill - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 17 | 0 | Salmonella | 0 |
| | Compound feedingstuffs for turkeys - final product - Feed mill - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 1 | 0 | Salmonella | 0 |
| | Compound feedingstuffs, not specified - final product - Feed mill - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 2 | 0 | Salmonella | 0 |
| | Feed material of cereal grain origin - barley derived - Processing plant - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 1 | 0 | Salmonella | 0 |
| | Feed material of cereal grain origin - maize derived - Processing plant - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 11 | 0 | Salmonella | 0 |
| | Feed material of cereal grain origin - other cereal grain derived - by-products of brewing and distilling - Processing plant - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 1 | 0 | Salmonella | 0 |
| | Feed material of cereal grain origin - other cereal grain derived - Processing plant - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 12 | 0 | Salmonella | 0 |
| | Feed material of cereal grain origin - rice derived - Processing plant - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 2 | 0 | Salmonella | 0 |
| | Feed material of land animal origin - feather meal - Processing plant - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 1 | 0 | Salmonella | 0 |
| | Feed material of oil seed or fruit origin - palm kernel derived - Processing plant - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 1 | 0 | Salmonella | 0 |
| | Feed material of oil seed or fruit origin - soya (bean) derived - Processing plant - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 2 | 0 | Salmonella | 0 |
| | Feed material of oil seed or fruit origin - sunflower seed derived - Processing plant - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 1 | 0 | Salmonella | 0 |
| | Other feed material - forages and roughages - Processing plant - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 1 | 0 | Salmonella | 0 |
| | Other feed material - miscellaneous - Processing plant - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 8 | 0 | Salmonella | 0 |
| | Other feed material - other plants - Processing plant - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 1 | 0 | Salmonella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | | Method | | Total units positive | Zoonoses | N of units positive |
|------------------|---|--------------------------|------------------|------|---|---|----------------------------|------------|------------------------|
| Not Available | Other feed material - yeast - Processing plant - Portugal - feed sample - Surveillance - Official sampling - Objective sampling | batch (food/fee d) | 25 | Gram | Real-Time PCR (qualitative or quantitative) | 1 | 0 | Salmonella | 0 |

Table Staphylococcal enterotoxins:STAPHYLOCOCCAL ENTEROTOXINS in food

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Sampling unit | Sample weight | Sample weight unit | Method | Total units tested | Total units positive | Zoonoses | N of units positive |
|------------------|--|---------------------------|---------------|-----------------------|---|--------------------------|----------------------------|-----------------------------|---------------------|
| Not Available | Cheeses made from cows' milk - fresh - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Immunofluorenscen ce assay tests (IFA) | 11 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from cows' milk - hard - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Immunofluorenscen ce assay tests (IFA) | 1 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Immunofluorenscen ce assay tests (IFA) | 11 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Immunofluorenscen ce assay tests (IFA) | 3 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from goats' milk - fresh - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Immunofluorenscen ce assay tests (IFA) | 12 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from goats' milk - hard - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Immunofluorenscen ce assay tests (IFA) | 2 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from goats' milk - hard - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Immunofluorenscen ce assay tests (IFA) | 1 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from goats' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Immunofluorenscen ce assay tests (IFA) | 4 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from goats' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Immunofluorenscen ce assay tests (IFA) | 2 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from sheep's milk - fresh - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Immunofluorenscen ce assay tests (IFA) | 17 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from sheep's milk - hard - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Immunofluorenscen ce assay tests (IFA) | 2 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from sheep's milk - hard - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Immunofluorenscen ce assay tests (IFA) | 3 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Immunofluorenscen ce assay tests (IFA) | 1 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses made from sheep's milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Immunofluorenscen ce assay tests (IFA) | 53 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses, made from mixed milk from cows, sheep and/or goats - fresh - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Immunofluorenscen ce assay tests (IFA) | 2 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses, made from mixed milk from cows, sheep and/or goats - soft and semi-soft - made from pasteurised milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Immunofluorenscen ce assay tests (IFA) | 4 | 0 | Staphylococcal enterotoxins | 0 |
| | Cheeses, made from mixed milk from cows, sheep and/or goats - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Immunofluorenscen ce assay tests (IFA) | 9 | 0 | Staphylococcal enterotoxins | 0 |
| | Dairy products (excluding cheeses) - milk powder and whey powder - Processing plant - Portugal - food sample - Surveillance - based on Regulation 2073 - Official sampling - Objective sampling | single (food/fee d) | 25 | Gram | Immunofluorenscen ce assay tests (IFA) | 5 | 0 | Staphylococcal enterotoxins | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of unit |
|------------------|--|---|------------------|--------------------------|----------------------------|---------------------|-----------|
| Not Available | Pigs - breeding animals - not raised under controlled housing conditions - sows and boars - Slaughterhouse - Spain - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 71 | 0 | Trichinella | 0 |
| | Pigs - breeding animals - raised under controlled housing conditions - sows and boars - Slaughterhouse - Spain - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 41 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Spain - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 12041 1 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Belgium - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 732 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Netherlands - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 157 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Spain - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 89148 9 | 0 | Trichinella | 0 |
| PORTUGAL | Pigs - breeding animals - not raised under controlled housing conditions - sows and boars - Slaughterhouse - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 2175 | 0 | Trichinella | 0 |
| | Pigs - breeding animals - raised under controlled housing conditions - sows and boars - Slaughterhouse - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 18317 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 61498 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 32885 23 | 0 | Trichinella | 0 |
| | Solipeds, domestic - Slaughterhouse - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 1001 | 0 | Trichinella | 0 |
| | Wild boars - wild - Game handling establishment - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 189 | 0 | Trichinella | 0 |
| | Wild boars - wild - Hunting - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 54 | 3 | Trichinella britovi | 3 |
| | Wild boars - wild - Hunting - Portugal - animal sample - organ/tissue - Surveillance - Private sampling - Objective sampling | Magnetic stirrer method for pooled sample digestion | animal | 32 | 0 | Trichinella | 0 |
| ONTINENTE | Pigs - breeding animals - not raised under controlled housing conditions - sows and boars - Slaughterhouse - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 2037 | 0 | Trichinella | 0 |
| | Pigs - breeding animals - raised under controlled housing conditions - sows and boars - Slaughterhouse - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 18208 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 46690 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 32360 58 | 0 | Trichinella | 0 |
| | Solipeds, domestic - Slaughterhouse - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 1001 | 0 | Trichinella | 0 |
| | Wild boars - wild - Game handling establishment - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 189 | 0 | Trichinella | 0 |
| | Wild boars - wild - Hunting - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 51 | 0 | Trichinella | 0 |
| | Wild boars - wild - Hunting - Portugal - animal sample - organ/tissue - Surveillance - Private sampling - Objective sampling | Magnetic stirrer method for pooled sample digestion | animal | 32 | 0 | Trichinella | 0 |

| Area of Sampling | Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy | Method | Sampling unit | Total units tested | Total units positive | Zoonoses | N of units positive |
|--|--|---|------------------|--------------------------|----------------------------|---------------------|---------------------|
| REGIÃO AUTÓNOMA DOS AÇORES | Pigs - breeding animals - not raised under controlled housing conditions - sows and boars - Slaughterhouse - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 138 | 0 | Trichinella | 0 |
| (NUTS level 1) | Pigs - breeding animals - raised under controlled housing conditions - sows and boars - Slaughterhouse - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 99 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 14807 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 51816 | 0 | Trichinella | 0 |
| REGIÃO AUTÓNOMA DA MADEIRA (NUTS | Pigs - breeding animals - raised under controlled housing conditions - sows and boars - Slaughterhouse - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 10 | 0 | Trichinella | 0 |
| level 1) | Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 1 | 0 | Trichinella | 0 |
| | Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 649 | 0 | Trichinella | 0 |
| Terras de Trás- os-Montes | Wild boars - wild - Hunting - Portugal - animal sample - organ/tissue - Surveillance - Official sampling - Census | Magnetic stirrer method for pooled sample digestion | animal | 3 | 3 | Trichinella britovi | 3 |

FOODBORNE OUTBREAKS TABLES

Foodborne Outbreaks: summarized data

| | Outbre strengi | | Stron | ng | | | Wea | k | |
|------------------------------|--|-------------|---------------|-------------------|----------|-------------|---------------|-------------------|----------|
| Causative agent | Food vehicle | N outbreaks | N human cases | N hospitalized | N deaths | N outbreaks | N human cases | N hospitalized | N deaths |
| B. cereus enterotoxins | Mixed food | 1 | 40 | 5 | 0 | | | | |
| Bacillus cereus | Mixed food | 3 | 72 | 69 | 0 | 1 | 8 | 8 | 0 |
| Clostridium botulinum toxins | Pig meat and products thereof | 1 | 4 | 4 | 0 | | | | |
| Clostridium perfringens | Mixed food | 3 | 60 | 27 | 0 | | | | |
| Norovirus | Unknown | | | | | 1 | 6 | 6 | 0 |
| Staphylococcal enterotoxins | Mixed food | 2 | 19 | 12 | 0 | | | | |
| Staphylococcus aureus | Vegetables and juices and other products there | of | | | | 1 | 21 | 0 | 0 |
| Unspecified | Unknown | | | | | 5 | 93 | 14 | 0 |

Strong Foodborne Outbreaks: detailed data

| Causative agent | Other Causative Agent | FBO nat. code | Outbreak type | Food vehicle | More food vehicle info | Nature of evidence | Setting | Place of origin of problem | Origin of food vehicle | Contributory factors | Comment | N outbreak | N humar s cases | | N p. deaths |
|--|-----------------------------|---------------------|------------------|-------------------------------|---|--|---|--|------------------------|---------------------------------------|--|---------------|-----------------------|----|----------------|
| B. cereus enterotoxi ns | Not Available | PT- 2017_ 08 | General | Mixed food | Grilled turkey steak with carrot rice | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomon ic to causative agent | Canteen or workplac e catering | Canteen or workplace catering | Portugal | Storage time/temperat ure abuse | B. cereus diarrheal enterotoxin producer | 1 | 40 | 5 | 0 |
| Bacillus cereus | Clostridium perfringens | PT- 2017_ 12 | General | Mixed food | Stewed beef meat | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomon ic to causative agent | Resident ial institution (nursing home or prison or boarding school) | Residentia I institution (nursing home or prison or boarding school) | Portugal | Storage time/temperat ure abuse | N_A | 1 | 60 | 60 | 0 |
| | Not Available | PT- 2017_ 11 | General | Mixed food | Codfish, potatoes, pepper, parsley | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomon ic to causative agent | Restaur ant or Cafe or Pub or Bar or Hotel or Catering service | Restaurant or Cafe or Pub or Bar or Hotel or Catering service | Portugal | Storage time/temperat ure abuse | N_A | 1 | 6 | 6 | 0 |
| | | PT- 2017_ 14 | General | Mixed food | Mixed foods various including fish and peas rice | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomon ic to causative agent | Resident ial institution (nursing home or prison or boarding school) | Residentia I institution (nursing home or prison or boarding school) | Portugal | Storage time/temperat ure abuse | N_A | 1 | 6 | 3 | 0 |
| Clostridiu m botulinum toxins | Not Available | PT- 2017_ 05 | General | Pig meat and products thereof | Smoked cured raw ham | Detection of causative agent in food vehicle or its component - Detection of indistinguisha ble causative agent in humans | Househ old | Household | Portugal | Unknown | Botulinum toxin B | 1 | 4 | 4 | 0 |

| Causative agent | Other Causative Agent | FBO nat. code | Outbreak type | Food vehicle | More food vehicle info | Nature of evidence | Setting | Place of origin of problem | Origin of food vehicle | Contributory factors | Comment | N outbreak | N human s cases | | N p. deaths |
|--|-----------------------------|---------------------|------------------|--------------|--|--|---|--|------------------------|--|---|---------------|-----------------------|----|----------------|
| Clostridiu m perfringen s | Not Available | PT- 2017_ 01 | General | Mixed food | Stewed meat with white rice | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomon ic to causative agent | School or kinderga rten | School or kindergart en | Portugal | Storage time/temperat ure abuse | N_A | 1 | 30 | 0 | 0 |
| | | PT- 2017_ 03 | General | Mixed food | Vegetable cream - soup | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomon ic to causative agent | Resident ial institution (nursing home or prison or boarding school) | Residentia I institution (nursing home or prison or boarding school) | Portugal | Storage time/temperat ure abuse | N_A | 1 | 25 | 25 | 0 |
| | | PT- 2017_ 10 | General | Mixed food | Poultry rice | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomon ic to causative agent | Multiple places of exposur e in one country | Residentia I institution (nursing home or prison or boarding school) | Portugal | Storage time/temperat ure abuse | N_A | 1 | 5 | 2 | 0 |
| Staphyloc occal enterotoxi ns | Bacillus | PT- 2017_ 07 | General | Mixed food | Mixed foods various including rice, mashed potatoes, fish and poultry | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomon ic to causative agent | Restaur ant or Cafe or Pub or Bar or Hotel or Catering service | Restaurant or Cafe or Pub or Bar or Hotel or Catering service | Portugal | Storage time/temperat ure abuse;Cross- contamination | Staphylococcus enterotoxin producer. "Other agent" - Bacillus subtilis Group (including B. subtilis, B. licheniformis, B. pumilus e B. amyloliquefaciens) | 1 | 7 | 7 | 0 |
| | Not Available | PT- 2017_ 02 | General | Mixed food | Sliced cheese and dry porc sausage, used to coat duck rice | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomon ic to causative agent | Multiple places of exposur e in one country | Canteen or workplace catering | Portugal | Storage time/temperat ure abuse;Cross- contamination | Staphylococcus enterotoxin A and D producer | 1 | 12 | 5 | 0 |

Weak Foodborne Outbreaks: detailed data

| | ausative | Other Causative Agent | FBO nat. code | Outbreak type | Food vehicle | More food vehicle info | Nature of evidence | Setting | Place of origin of problem | Origin of food vehicle | Contributory factors | Comment | N outbreaks | N human cases | | N p. deaths |
|---|------------------------------|-----------------------------|---------------------|------------------|--|--|--|--|--|------------------------|--|---------------|----------------|---------------------|---|----------------|
| | Bacillus cereus | Not Available | PT- 2017_ 13 | General | Mixed food | Mixed foods various including beans, meat, rice, tuna and potatoes | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent | Residentia I institution (nursing home or prison or boarding school) | Residentia I institution (nursing home or prison or boarding school) | Portugal | Storage time/tempera ture abuse | N_A | 1 | 8 | 8 | 0 |
| | Norovirus | Not Available | PT- 2017_ 06 | General | Unknown | N_A | Unknown | School or kindergart en | School or kindergart en | Unknown | Unknown | Norovirus GII | 1 | 6 | 6 | 0 |
| (| Staphyloc occus aureus | Not Available | PT- 2017_ 09 | General | Vegetables and juices and other products thereof | Corn and letuce salad | Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent | School or kindergart en | School or kindergart en | Portugal | Storage time/tempera ture abuse;Cross- contaminatio n | N_A | 1 | 21 | 0 | 0 |
| | Unspecifi ed | Not Available | PT- 2017_ 04 | General | Unknown | N_A | Unknown | Restauran t or Cafe or Pub or Bar or Hotel or Catering service | Restauran t or Cafe or Pub or Bar or Hotel or Catering service | Unknown | Unknown | N_A | 1 | 27 | 2 | 0 |
| | | | PT- 2017_ 15 | General | Unknown | N_A | Unknown | Restauran t or Cafe or Pub or Bar or Hotel or Catering service | Restauran t or Cafe or Pub or Bar or Hotel or Catering service | Unknown | Unknown | N_A | 1 | 3 | 3 | 0 |
| | | | PT- 2017_ 16 | General | Unknown | N_A | Unknown | Multiple places of exposure in one country | School or kindergart en | Unknown | Unknown | N_A | 1 | 21 | 0 | 0 |

| Causative agent | Other Causative Agent | | Outbreak type | Food vehicle | More food vehicle info | Nature of evidence | Setting | | Origin of food | I Contributory factors | Comment | N outbreaks | N human cases | N hosp | N o. deaths |
|--------------------|-----------------------------|--------------------|------------------|--------------|------------------------|--------------------|--|--|----------------|------------------------|---------|----------------|---------------------|-----------|----------------|
| Unspecifi ed | Not Available | PT- 2017_ 17 | General | Unknown | N_A | Unknown | Residentia I institution (nursing home or prison or boarding school) | Residentia I institution (nursing home or prison or boarding school) | Unknown | Unknown | N_A | 1 | 29 | 5 | 0 |
| | | PT- 2017_ 18 | General | Unknown | N_A | Unknown | Residentia I institution (nursing home or prison or boarding school) | Residentia I institution (nursing home or prison or boarding school) | Unknown | Unknown | N_A | 1 | 13 | 4 | 0 |

ANTIMICROBIAL RESISTANCE TABLES FOR CAMPYLOBACTER

Table Antimicrobial susceptibility testing of Campylobacter coli in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Processing plant Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ciprofloxacin | Erythromycin | Gentamicin | Nalidixic acid | Streptomycin | Tetracycline |
|--------|-------------------------|---------------|--------------|------------|----------------|--------------|--------------|
| | ECOFF | 0.5 | 8 | 2 | 16 | 4 | 2 |
| | Lowest limit | 0.12 | 1 | 0.12 | 1 | 0.25 | 0.5 |
| | Highest limit | 16 | 128 | 16 | 64 | 16 | 64 |
| | N of tested isolates | 8 | 8 | 8 | 8 | 8 | 8 |
| MIC | N of resistant isolates | 8 | 3 | 0 | 7 | 0 | 8 |
| <=0.12 | | | | 1 | | | |
| 0.25 | | | | 5 | | | |
| 0.5 | | | | 1 | | | |
| <=1 | | | 5 | | | | |
| 1 | | | | | | 6 | |
| 2 | | 2 | | 1 | | 2 | |
| 4 | | 2 | | | | | |
| 8 | | 2 | | | | | |
| 16 | | 2 | | | 1 | | 1 |
| 32 | | | | | 4 | | 3 |
| 64 | | | | | 1 | | 2 |
| >64 | | | | | 2 | | 2 |
| >128 | | | 3 | | | | |

Table Antimicrobial susceptibility testing of Campylobacter coli in Meat from pig - carcase

Sampling Stage: Processing plant Sampling Type: food sample - carcase swabs Sampling Context: Monitoring

Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ciprofloxacin | Erythromycin | Gentamicin | Nalidixic acid | Streptomycin | Tetracycline |
|--------|-------------------------|---------------|--------------|------------|----------------|--------------|--------------|
| | ECOFF | 0.5 | 8 | 2 | 16 | 4 | 2 |
| | Lowest limit | 0.12 | 1 | 0.12 | 1 | 0.25 | 0.5 |
| | Highest limit | 16 | 128 | 16 | 64 | 16 | 64 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 1 | 1 |
| <=0.12 | | 1 | | | | | |
| 0.25 | | | | 1 | | | |
| <=1 | | | 1 | | | | |
| 2 | | | | | 1 | | |
| >16 | | | | | | 1 | |
| 32 | | | | | | | 1 |

Table Antimicrobial susceptibility testing of Campylobacter coli in Meat from turkey - fresh

Sampling Stage: Processing plant Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ciprofloxacin | Erythromycin | Gentamicin | Nalidixic acid | Streptomycin | Tetracycline |
|------|-------------------------|---------------|--------------|------------|----------------|--------------|--------------|
| | ECOFF | 0.5 | 8 | 2 | 16 | 4 | 2 |
| | Lowest limit | 0.12 | 1 | 0.12 | 1 | 0.25 | 0.5 |
| | Highest limit | 16 | 128 | 16 | 64 | 16 | 64 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 1 | 1 | 0 | 1 | 0 | 1 |
| 0.25 | | | | 1 | | | |
| 1 | | | | | | 1 | |
| 4 | | 1 | | | | | |
| 32 | | | | | | | 1 |
| 64 | | | | | 1 | | |
| >128 | | | 1 | | | | |

Table Antimicrobial susceptibility testing of Campylobacter coli in Meat from pig - meat products

Sampling Stage: Processing plant Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ciprofloxacin | Erythromycin | Gentamicin | Nalidixic acid | Streptomycin | Tetracycline |
|------|-------------------------|---------------|--------------|------------|----------------|--------------|--------------|
| | ECOFF | 0.5 | 8 | 2 | 16 | 4 | 2 |
| | Lowest limit | 0.12 | 1 | 0.12 | 1 | 0.25 | 0.5 |
| | Highest limit | 16 | 128 | 16 | 64 | 16 | 64 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 1 | 1 | 0 | 1 | 1 | 1 |
| 0.25 | | | | 1 | | | |
| 4 | | 1 | | | | | |
| >16 | | | | | | 1 | |
| 64 | | | | | 1 | | 1 |
| 128 | | | 1 | | | | |

Table Antimicrobial susceptibility testing of Campylobacter coli in Meat from pig - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ciprofloxacin | Erythromycin | Gentamicin | Nalidixic acid | Streptomycin | Tetracycline |
|--------|-------------------------|---------------|--------------|------------|----------------|--------------|--------------|
| | ECOFF | 0.5 | 8 | 2 | 16 | 4 | 2 |
| | Lowest limit | 0.12 | 1 | 0.12 | 1 | 0.25 | 0.5 |
| | Highest limit | 16 | 128 | 16 | 64 | 16 | 64 |
| | N of tested isolates | 3 | 3 | 3 | 3 | 3 | 3 |
| MIC | N of resistant isolates | 2 | 2 | 2 | 2 | 3 | 3 |
| <=0.12 | | 1 | | | | | |
| 1 | | | | 1 | | | |
| 2 | | | 1 | | | | |
| 4 | | 1 | | | | | |
| 8 | | | | | 1 | | |
| 16 | | 1 | | | | | |
| >16 | | | | 2 | | 3 | |
| 64 | | | | | 1 | | 1 |
| >64 | | | | | 1 | | 2 |
| 128 | | | 1 | | | | |
| >128 | | | 1 | | | | |

Table Antimicrobial susceptibility testing of Campylobacter jejuni in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ciprofloxacin | Erythromycin | Gentamicin | Nalidixic acid | Streptomycin | Tetracycline |
|--------|-------------------------|---------------|--------------|------------|----------------|--------------|--------------|
| | ECOFF | 0.5 | 4 | 2 | 16 | 4 | 1 |
| | Lowest limit | 0.12 | 1 | 0.12 | 1 | 0.25 | 0.5 |
| | Highest limit | 16 | 128 | 16 | 64 | 16 | 64 |
| | N of tested isolates | 4 | 4 | 4 | 4 | 4 | 4 |
| MIC | N of resistant isolates | 4 | 1 | 0 | 4 | 0 | 3 |
| <=0.12 | | | | 2 | | | |
| <=0.25 | | | | | | 2 | |
| 0.25 | | | | 2 | | | |
| <=1 | | | 3 | | | | |
| 1 | | | | | | 2 | 1 |
| 4 | | 2 | | | | | |
| 8 | | 1 | | | | | |
| 16 | | 1 | | | | | 1 |
| 32 | | | 1 | | 2 | | 1 |
| >64 | | | | | 2 | | 1 |

Table Antimicrobial susceptibility testing of Campylobacter jejuni in Meat from broilers (Gallus gallus) - fresh - with skin

Sampling Stage: Processing plant Sampling Type: food sample - neck skin Sampling Context: Monitoring

Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ciprofloxacin | Erythromycin | Gentamicin | Nalidixic acid | Streptomycin | Tetracycline |
|--------|-------------------------|---------------|--------------|------------|----------------|--------------|--------------|
| | ECOFF | 0.5 | 4 | 2 | 16 | 4 | 1 |
| | Lowest limit | 0.12 | 1 | 0.12 | 1 | 0.25 | 0.5 |
| | Highest limit | 16 | 128 | 16 | 64 | 16 | 64 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.12 | | 1 | | | | | |
| <=0.5 | | | | | | | 1 |
| 0.5 | <u> </u> | <u> </u> | <u> </u> | 1 | <u> </u> | <u> </u> | <u> </u> |
| <=1 | | | 1 | | | | |
| 2 | | | | | 1 | 1 | |

ANTIMICROBIAL RESISTANCE TABLES FOR SALMONELLA

Table Antimicrobial susceptibility testing of Salmonella 4,[5],12:i:- in Meat from bovine animals and pig - meat products

Sampling Stage: Processing plant Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official and industry sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | | 1 |
| <=1 | | | | | | | | 1 | | | | | | | |
| 1 | | | | | | | | | | | | | | 1 | |
| <=4 | | | | | | | | | | | 11 | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 64 | | | | | | 1 | | | | | | | | | |
| >64 | | 1 | | | | | | | | | | | 1 | | |
| >1024 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella 4,[5],12:i:- in Meat from pig - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 |
| міс | N of resistant isolates | 2 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 2 | 3 | 0 | 0 |
| <=0.03 | | | | | | | | | | 3 | | | | | |
| 0.064 | | | | | | | 3 | | | | | | | | |
| <=0.25 | | | | 3 | | | | | | | | | | | 2 |
| <=0.5 | | | | | 3 | | | | 3 | | | | | | |
| 0.5 | | | | | | | | - | | | | | | 3 | 1 |
| <=1 | | <u> </u> | | | | | | 3 | | | | | | | |
| 2 | | 1 | | | | 2 | | | | | | | | | |
| <=8 8 | | | 3 | | | 3 | | | | | 3 | | | | |
| 32 | | | | | | | | | | | <u> </u> | 1 | | | |
| >64 | | 2 | | | | | | | | | | ' | 3 | | |
| >1024 | | | | | | | | | | | | 2 | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella 4,[5],12:i:- in Birds

Sampling Stage: Natural habitat

Sampling Type: animal sample - cloacal swab

Sampling Context: Monitoring

Sampler: Not applicable

Sampling Strategy: Other

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 1 |
| <=1 | | | | | | | | 1 | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| >64 | | 1 | | | | | | | | | | | 11 | | |
| >1024 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella 4,[5],12:i:- in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampler: Official sampling

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| міс | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 0.03 | | | | | | | 2 | | | | | | | | |
| 0.064 | | | | | | | | | | 2 | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | 2 | |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| 0.5 | | | | | | | | 1 | | | | | | | 2 |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | 1 | | | | | | 1 | | | | | 2 | | |
| <=4 | | ı | | | | | | ı | | | 2 | | | | |
| <=8 | | | | | | 2 | | | | | | | | | |
| 8 | | | 2 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |
| 32 | | | | | | | | | | | | 1 | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella 4,[5],12:i:- in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|-----------------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | | 4 | | | | | 1 | | | | | | | |
| <u>4</u> <=8 | | | 1 | | | 1 | | | | | | | | | |
| 8 | | | | | | ı | | | | | 1 | | | | |
| >64 | | 1 | | | | | | | | | | | 1 | | |
| >1024 | | · | | | | | | | | | | 1 | • | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella 4,[5],12:i:- in Meat from poultry, unspecified - meat products

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 |
| 0.03 | | | | | | | 1 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | | | | | | | 1 | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | 1 | | | | |
| >64 | | 1 | | | | | | | | | | | 1 | | |
| >1024 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella 4,[5],12:i:- in Meat from turkey

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| MIC | N of resistant isolates | 2 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 2 | 2 | 0 | 0 |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | 1 | 1 |
| <=1 | | | | | | | | 2 | | | | | | | |
| <=4 | | | | | | 2 | | | | | 1 | | | | |
| <=8 | | | 2 | | | 2 | | | | | | | | | |
| 32 | | | | | | | | | | | 1 | | | | |
| >64 | | 2 | | | | | | | | | | | 2 | | |
| >1024 | | | | | | | | | | | | 2 | | | |
| 1021 | | | | | | | | | | | | _ | | | |

Table Antimicrobial susceptibility testing of Salmonella 4,[5],12:i:- in Meat from turkey - meat products

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 1 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | 1 | |
| <=1 | | | | | | | | 1 | | | | | | | _ |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | 1 | | | | |
| >64 | | 1 | | | | | | | | | | | | | |
| >1024 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella 4,[5],12:i:- in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampler: Official sampling

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sufamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|-----------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| MIC | N of resistant isolates | 2 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 2 | 2 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | 1 | 2 |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 11 | |
| <=1 | | | | | | | | 2 | | | 4 | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 8 | | | 2 | | | 2 | | | | | 1 | | | | |
| >64 | | 2 | | | | | | | | | | | 2 | | |
| >1024 | | | | | | | | | | | | 2 | | | |
| 7 1024 | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella 4,[5],12:i:- in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| MIC | N of resistant isolates | 2 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 2 | 2 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| 0.064 | | | | | | | | | | 2 | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | 1 | 2 |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | | | | | | | 2 | | | | | | | |
| <=4 | | | | | | | | | | | 2 | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 2 | | | | | | | | | |
| 16 | | | 1 | | | | | | | | | | | | |
| >64 | | 2 | | | | | | | | | | | 2 | | |
| >1024 | | | | | | | | | | | | 2 | | | |

Table Antimicrobial susceptibility testing of Salmonella 4,[5],12:i:- in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 11 |
| <=0.5 | | | | | 1 | | | | | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| 1 | | | | | | | | | 1 | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella 4,[5],12:i:- in Meat from pig - meat products

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | | | | | | | 1 | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | 4 | | | 1 | | | | | | | | | |
| 8 >64 | | 1 | 1 | | | | | | | | | | 1 | | |
| >1024 | | ı | | | | | | | | | | 1 | | | |
| -1024 | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella 4,[5],12:i:- in Meat from pig - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| MIC | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 2 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | | 2 |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 2 | |
| <=1 | | 11 | | | | | | 2 | | | | | | | |
| <=4 | | | | | | | | | | | 2 | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 2 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |
| >64 | | 1 | | | | | | | | | | | 2 | | |
| >1024 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella 4,[5],12:i:- in Meat from pig - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 |
| 0.03 | | | | | | | 1 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | 1 | | | | |
| 16 | | | | | | | | | | | | 1 | | | |
| >64 | | | | | | | | | | | | | 1 | | |

Table Antimicrobial susceptibility testing of Salmonella 4,12:i:- in Meat from pig - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| МІС | N of resistant isolates | 2 | 0 | 0 | 0 | 2 | 0 | 0 | 0 | 0 | 0 | 2 | 2 | 0 | 2 |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | | |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 2 | |
| <=1 | | | | | | | | 2 | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 8 | | | 1 | | | | | | | | 1 | | | | |
| 16 | | | 1 | | | | | | | | | | | | |
| >32 | | | | | | | | | | | | | | | 2 |
| >64 | | 2 | | | | 4 | | | | | | | 2 | | |
| 128 | | | | | | 1 | | | | | | | | | |
| >128 | | | | | | 11 | | | | | | 2 | | | |
| >1024 | | | | | | | | | | | | 2 | | | |

Table Antimicrobial susceptibility testing of Salmonella 4,12:i:- in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampler: Official sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 1 |
| <=1 | | | | | | | | 1 | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| >64 | | 1 | | | | | | | | | | | 1 | | |
| >1024 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella 4,12:i:- in Meat from bovine animals - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 1 |
| <=1 | | | | | | | | 1 | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| >64 | | 1 | | | | | | | | | | | 1 | | |
| >1024 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella 4,12:i:- in Meat from pig - meat products

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | | | | | | | 1 | | | | | | | |
| <=4 | | | | | | <u> </u> | | | | | 1 | | | | |
| <=8 | | | 4 | | | 1 | | | | | | | | | |
| 8 >64 | | 1 | 1 | | | | | | | | | | 1 | | |
| >1024 | | ı | | | | | | | | | | 1 | | | |
| 71024 | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella 4,12:i:- in Meat from pig - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 |
| 0.064 | | | | | | | 1 | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | | | | | | | 1 | | | | | | | |
| 8 | | | 1 | | | | | | | | 1 | | | | |
| 16 | | | | | | 1 | | | | | | | | | |
| >64 | | 1 | | | | | | | | | | | 1 | | |
| >1024 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Abaetetuba in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|------------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 1 |
| <=2 | | | | | | | | 1 | | | | | 11 | | |
| 2 <=4 | | | | | | | | 1 | | | 4 | | | | |
| <=4 <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | ı ı | | | | | | | | | |
| 32 | | | · | | | | | | | | | 1 | | | |
| >64 | | 1 | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Albany in Feed material of cereal grain origin

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| 0.5 | | - | | | | | | | | | | | | 1 | 1 |
| <=1 | | 2 | | | | | | 11 | | | | | | | |
| <=2 | | | | | | | | 1 | | | | | 2 | | |
| 2 <=4 | | | | | | | | | | | 2 | | | | |
| <=8 | | | | | | 2 | | | | | | | | | |
| 8 | | | 2 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Anatum in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampler: Official sampling

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|------------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 |
| MIC | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 |
| <=0.03 | | | | | | | | | | 3 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| 0.064 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 3 | | | | | | | | | | 1 | 2 |
| <=0.5 | | | | | 3 | | | | 3 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 2 | |
| <=1 | | 1 | | | | | | 3 | | | | | | | |
| <=2 | | 4 | | | | | | | | | | | 3 | | |
| 2 <=4 | | 1 | | | | | | | | | 2 | | | | |
| <=4 <=8 | | | | | | 3 | | | | | 3 | | | | |
| 8 | | | 2 | | | <u> </u> | | | | | | | | | |
| 16 | | | 1 | | | | | | | | | | | | |
| 32 | | | • | | | | | | | | | 2 | | | |
| >32 | | | | | | | | | | | | | | | 1 |
| >64 | | 1 | | | | | | | | | | | | | |
| >1024 | | | | | | | | | | | | 1 | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Anatum in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|------------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 |
| МІС | N of resistant isolates | 3 | 0 | 0 | 0 | 0 | 0 | 0 | 2 | 0 | 0 | 1 | 0 | 0 | 11 |
| <=0.03 | | | | | | | | | | 6 | | | | | |
| 0.03 | | | | | | | 6 | | | | | | | | |
| <=0.25 | | | | 6 | | | | | | | | | | 3 | 5 |
| <=0.5 | | | | | 6 | | | | 3 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 3 | |
| <=1 | | 3 | | | | | | 6 | <u> </u> | | | | | | |
| 1 | | | | | | | | | 1 | | | | • | | |
| <=2 <=4 | | | | | | | | | | | | | 6 | | |
| <=4 | | | | | | 6 | | | | | 6 | | | | |
| 8 | | | 6 | | | 6 | | | | | | | | | |
| 32 | | | | | | | | | | | | 5 | | | |
| >32 | | | | | | | | | 2 | | | | | | 1 |
| >64 | | 3 | | | | | | | _ | | | | | | |
| >1024 | | | | | | | | | | | | 1 | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Anatum in Feed material of cereal grain origin

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Anatum in Meat from duck - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Berta in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 11 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | 4 |
| >32 | | 1 | | | | | | | | | | | | | 1 |
| >1024 | | I | | | | | | | | | | 1 | | | |
| 71024 | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Brandenburg in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | 1 | | | | | | | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | | | | | | | 1 | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | 1 | | | 1 | | | | | | | | | |
| 16 | | | 1 | | | | | | | | | 1 | | | |
| 32 | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Bredeney in Meat from pig - carcase

Sampling Stage: Slaughterhouse Sampling Type: food sample - carcase swabs Sampling Context: Monitoring

Sampler: HACCP and own check Sampling Strategy: Objective sampling Programme Code: AMR MON pnl2

Analytical Method:

| Anai | ytical Method: | | | | | | | | | | |
|----------|----------------------------|---------------|---------------|------------------------------|---------------------------------------|---------------|-------------------------------|---------------|---------------|---------------|---------------|
| Cour | ntry of Origin: | Portugal | | | | | | | | | |
| | AM substance | Cefepime | Cefotaxim | Cefotaxime + Clavulanic acid | Cefoxitin | Ceftazidim | Ceftazidime + Clavulanic acid | Ertapenem | Imipenem | Meropenem | Temocillin |
| | Cefotaxime synergy test | Not Available | Not Available | Negative/Abs ent | Not Available | Not Available | Not Available | Not Available | Not Available | Not Available | Not Available |
| | Ceftazidime synergy test | Not Available | Not Available | Not Available | Not Available | Not Available | Negative/Abs ent | Not Available | Not Available | Not Available | Not Available |
| | ECOFF | 0.125 | 0.5 | 0.5 | 8 | 2 | 2 | 0.06 | 1 | 0.125 | 32 |
| | Lowest limit | 0.064 | 0.25 | 0.064 | 0.5 | 0.25 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| | Highest limit | 32 | 64 | 64 | 64 | 128 | 128 | 2 | 16 | 16 | 128 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 0 | 0 | 0 |
| 0.03 | | | | | | | | 1 | | | |
| 0.064 | | | | | | | | | | 1 | |
| 0.25 | | | | | | | | | 1 | | |
| 2 | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | 1 | | | | 1 |
| 16 32 | | | I | 1 | 1 | 1 | | | | | |
| - 52 | | | | | · · · · · · · · · · · · · · · · · · · | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Bredeney in Meat from pig - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| MIC | N of resistant isolates | 1 | 0 | 1 | 1 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 1 |
| <=1 | | 11 | | | | | | 1 | | | | | | | |
| 1 | | | | | | | | | 1 | | | | | | |
| <=2 | | | | | | | | 4 | | | | | 1 | | |
| 2 <=4 | | | | | | | | 1 | | | 2 | | | | |
| >4 | | | | 1 | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 2 | | | ' | | | | | | | | | |
| >8 | | | | | 1 | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | _ |
| >64 | | 1 | | | | | | | | | | | 1 | | |
| 128 | | | | | | 1 | | | | | | | | | |
| >1024 | | | | | | | | | | | | 1 | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Bredeney in Birds

Sampling Stage: Natural habitat Sampling Type: animal sample - cloacal swab Sampling Context: Monitoring

Sampler: Not applicable Sampling Strategy: Other Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sufamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|-----------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 1 | 0 | 1 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | | | | | | | |
| <=1 | | 11 | | | | | | 1 | | | | | | | |
| 1 | | | | | | | | | | | | | | 1 | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 8 | | | 1 | | | | | | <u> </u> | | | | | | |
| 16 | | | | | | | | | 1 | | | | | | |
| >32 | | | | | | | | | | | | | 1 | | 1 |
| >64 | | | | | | 4 | | | | | | | 1 | | |
| 128 | | | | | | 1 | | | | | | 1 | | | |
| >1024 | | | | | | | | | | | | l | | | |

Table Antimicrobial susceptibility testing of Salmonella Bredeney in Meat from turkey - meat preparation - intended to be eaten cooked

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| міс | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | 1 | | | | | | 1 | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Bredeney in Meat from turkey

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| міс | N of resistant isolates | 1 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 1 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | | | | | | | 1 | | | | | | | |
| 1 | | | | | | | 1 | | | | | | | | 1 |
| 2 | | | | | | <u> </u> | | | | | | <u> </u> | | 1 | |
| 16 | | | 4 | | | 1 | | | | | | 1 | | | |
| 32 | | 1 | 1 | | | | | | | | | | 4 | | |
| >64 | | 7 | | | | | | | | | 1 | | 1 | | |
| >128 | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Bredeney in Meat from pig - meat products - fresh raw sausages

Sampling Stage: Processing plant

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | 11 | | | | | | | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 4 | | | | | | | | 1 | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | 4 | | | |
| 16 | | | | | | | | | | | | 11 | | | |

Table Antimicrobial susceptibility testing of Salmonella Bredeney in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampler: Official sampling

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | 1 | | | | | | | | | 1 | | | | |
| <=4 | | | | | | 4 | | | | | 1 | | | | |
| <=8 | | | 1 | | | 1 | | | | | | | | | |
| 8 32 | | | ' | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Cerro in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampler: Official sampling

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 |
| MIC | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 2 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 |
| <=0.03 | | | | | | | | | | 3 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 3 | | | | | | | | | | 2 | 2 |
| <=0.5 | | | | | 3 | | | | 3 | | | | | | |
| 0.5 | | | | | | | 2 | | | | | | | 1 | |
| <=1 | | 2 | | | | | | 3 | | | | | | | |
| <=2 | | | | | | | | | | | | | 2 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 4 | | | | | | 2 | | | | | | | 1 | | |
| <=8 8 | | | 3 | | | 3 | | | | | | | | | |
| 16 | | | 3 | | | | | | | | 2 | 2 | | | |
| >32 | | | | | | | | | | | | 2 | | | 1 |
| >64 | | 1 | | | | | | | | | | | | | ' |
| >1024 | | ' | | | | | | | | | | 1 | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Cerro in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | | |
| <=1 | | | | | | | | 1 | | | | | | | |
| <=2 | | | | | | <u> </u> | | | | | | | 1 | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | 4 | | | |
| 16 | | | | | | | | | | | | 1 | | | |
| >32 | | | | | | | | | | | 1 | | | | 1 |
| >64 | | 1 | | | | | | | | | | | | | ı |

Table Antimicrobial susceptibility testing of Salmonella Coeln in Meat from turkey

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Derby in Meat from pig - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sufamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|-----------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 18 | 18 | 18 | 18 | 18 | 18 | 18 | 18 | 18 | 18 | 18 | 18 | 18 | 18 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 |
| <=0.015 | | | | | | | 15 | | | | | | | | |
| <=0.03 | | | | | | | | | | 18 | | | | | |
| 0.03 | | | | | | | 3 | | | | | | | | |
| <=0.25 | | | | 18 | | | | | | | | | | 15 | 16 |
| <=0.5 | | | | | 18 | | | | 17 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 3 | 2 |
| <=1 | | 17 | | | | | | 17 | | | | | | | |
| 1 | | | | | | | | | 1 | | | | | | |
| <=2 | | | | | | | | | | | | | 17 | | |
| 2 | | 1 | | | | | | 11 | | | | | | | |
| <=4 | | | | | | | | | | | 17 | | | | |
| <=8 | | | | | | 18 | | | | | | | | | |
| 8 16 | | | 18 | | | | | | | | 1 | | | | |
| 32 | | | | | | | | | | | | 8 | | | |
| | | | | | | | | | | | | 8 | | | |
| 64 | | | | | | | | | | | | 1 | 1 | | |
| >64 >1024 | | | | | | | | | | | | 1 | 1 | | |
| >1024 | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Derby in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| 0.25 | | | | | | | 1 | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | 1 | | | | | | | | | | 1 | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | | | | | | | | | 1 | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Derby in Meat from poultry, unspecified - meat products

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 1 |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 4 | | | | | | | | | | | | | 1 | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Derby in Meat from pig - meat products - fresh raw sausages

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 11 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | | | | | | | 1 | | | | | | | |
| 2 | | 1 | | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | | | | | | | | | 1 | | | | |
| 16 | | | 1 | | | | | | | | | 1 | | | |
| >64 | | | | | | | | | | | | | 1 | | |

Table Antimicrobial susceptibility testing of Salmonella Derby in Meat from bovine animals - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | 1 | | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | 1 | | | | |
| 64 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Derby in Meat from pig - meat products

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 1 |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | 1 | | | | |
| 32 | | | | | | | | | | | | 1 | | | |
| 64 | | | | | | | | | | | | | 1 | | |

Table Antimicrobial susceptibility testing of Salmonella Derby in Meat from pig - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | 2 | |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| 0.5 | | | | | | | | | | | | | | | 2 |
| <=1 | | 2 | | | | | | 2 | | | | | | | |
| <=2 | | | | | | | | | | | | | 2 | | |
| <=4 | | | <u> </u> | | | | | | | | 2 | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 2 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | 1 | | | |
| 16 32 | | | | | | | | | | | | 1 | | | |
| 32 | | | | | | | | | | | | l | | | |

Table Antimicrobial susceptibility testing of Salmonella Derby in Meat from pig - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sufamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|-----------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | | 1 |
| <=1 | | 1 | | | | | | 11 | | | | | | <u> </u> | |
| 1 | | | | | | | | | | | | | | 1 | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | 1 | | | 1 | | | | | | | | | |
| 8 >64 | | | ı | | | | | | | | | | 1 | | |
| >1024 | | | | | | | | | | | | 1 | l e | | |
| 7 1024 | | | | | | | | | | | | · · | | | |

Table Antimicrobial susceptibility testing of Salmonella Dublin in Meat from bovine animals - minced meat - intended to be eaten cooked

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | 4 | | | 1 | | | | |
| 4 | | | | | | 4 | | 1 | | | | 1 | | | |
| <=8 8 | | | 1 | | | 1 | | | | | | 1 | | | |
| 0 | | | 1 | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella enterica, subspecies enterica in Ducks

Sampling Stage: Farm

Sampling Type: animal sample - organ/tissue

Sampling Context: Clinical investigations

Sampler: Industry sampling

Sampling Strategy: Suspect sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | 1 | | | |
| 8 | | | 1 | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella enterica, subspecies salamae in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 5 | | | | | |
| 0.03 | | | | | | | 7 | | | | | | | | |
| 0.064 | | | | | | | | | | 2 | | | | | |
| <=0.25 | | | | 7 | | | | | | | | | | 6 | 2 |
| <=0.5 | | | | | 7 | | | | 7 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 5 |
| <=1 | | 2 | | | | | | 7 | | | | | | | |
| <=2 | | | | | | | | | | | | | 6 | | |
| 2 | | 5 | | | | | | | | | | | | | |
| <=4 | | | | | | | | | | | 6 | | 4 | | |
| 4 <=8 | | | | | | 7 | | | | | | 0 | 1 | | |
| 8 | | | 4 | | | 1 | | | | | 1 | 2 | | | |
| 16 | | | 3 | | | | | | | | ı | 1 | | | |
| 32 | | | J | | | | | | | | | 4 | | | |
| - 02 | | | | | | | | | | | | <u> </u> | | | |

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|-----------------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 2 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 3 | | | | | |
| 0.03 | | | | | | | 3 | | | | | | | | |
| <=0.25 | | | | 3 | | | | | | | | | | | |
| <=0.5 | | | | | 3 | | | | 3 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 3 | 2 |
| 1 | | | | | | | | | | | | | | | 1 |
| <=2 | | - | | | | | | | | | | | 2 | | |
| 2 | | 3 | | | | | | 1 | | | | | | | |
| <=4 | | | | | | | | • | | | 3 | | 4 | | |
| <u>4</u> <=8 | | | | | | 3 | | 2 | | | | | 1 | | |
| 8 | | | 2 | | | <u> </u> | | | | | | | | | |
| 16 | | | 1 | | | | | | | | | | | | |
| 32 | | | <u>'</u> | | | | | | | | | 2 | | | |
| 64 | | | | | | | | | | | | 1 | | | |
| | | | | | | | | | | | | · · | | | |

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Meat from turkey - meat preparation - intended to be eaten cooked

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | | | | | | | | |
| 1 | | | | | | | | | | | | | | 1 | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 4 | | | | | | | | 1 | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | 1 | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampling Context: Control and eradication

Sampler: Official sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | | 1 |
| <=1 | | | | | | | | 11 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | 1 | | | | | | | | | 1 | | | | |
| <=4 | | | | | | 1 | | | | | 1 | | | | |
| <=8 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | ' | | | | | | | | | 1 | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Feed material of cereal grain origin

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 0.03 | | | | | | | 1 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | | | | | | | 11 | | | | | | | |
| <=2 | | 1 | | | | | | | | | | | 1 | | |
| 2 <=4 | | 1 | | | | | | | | | 1 | | | | |
| 4 | | | 1 | | | | | | | | ı | | | | |
| <=8 | | | ' | | | 1 | | | | | | | | | |
| 32 | | | | | | • | | | | | | 1 | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampler: Official sampling

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 2 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 2 |
| 1 | | | | | | | | | 1 | | | | | | |
| <=2 | | | | | | | | | | | | | 2 | | |
| 2 | | 2 | | | | | | 1 | | | | | | | |
| <=4 | | | | | | | | | | | 2 | | | | |
| 4 | | | | | | | | 1 | | | | | | | |
| <=8 | | | | | | 2 | | | | | | | | | |
| 8 | | | 2 | | | | | | | | | <u>.</u> | | | |
| 16 | | | | | | | | | | | | 1 | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 3 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| <=0.25 | | | | 3 | | | | | | | | | | 2 | 1 |
| 0.25 | | | | | | | 1 | | | | | | | | |
| <=0.5 | | | | | 3 | | | | 3 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 2 |
| <=1 | | 1 | | | | | | 2 | | | | | | | |
| <=2 | | | | | | | | | | | | | 3 | | |
| 2 | | 2 | | | | | | | | | | | | | |
| <=4 | | | 1 | | | | | 1 | | | 2 | | | | |
| 4 <=8 | | | | | | 2 | | | | | | | | | |
| 8 | | | 2 | | | 3 | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |
| 32 | | | | | | | | | | | | 2 | | | |
| >128 | | | | | | | | | | | 1 | | | | |
| 120 | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| _1 | | | | | | | | | | | | | | | 1 |
| 2 | | 1 | | | | | | 1 | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 4 | | | | | | | | | | | | | 1 | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Meat from pig - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | | 1 |
| <=1 | | 1 | | | | | | | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | 1 | | | 1 | | | | |
| 4 | | | | | | 1 | | 1 | | | | | | | |
| <=8 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | ' | | | | | | | | | 1 | | | |
| 10 | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Hadar in Meat, mixed meat - meat products

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| міс | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | 1 | |
| <=1 | | 11 | | | | | | 1 | | | | | | | |
| <=8 | | | | | | 11 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | 1 | | | | |
| 32 | | | | | | | | | | | | 1 | | | |
| 64 | | | | | | | | | | | | | 1 | | |

Table Antimicrobial susceptibility testing of Salmonella Havana in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 1 |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Havana in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampling Context: Control and eradication

Sampler: Official sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 6 | 0 | 0 | 0 | 3 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 7 | | | | | |
| 0.064 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 7 | | | | | | | | | | | 4 |
| <=0.5 | | | | | 7 | | | | 6 | | | | | | |
| 0.5 | | | | | | | 4 | | | | | | | 6 | 3 |
| <=1 | | 4 | | | | | | 7 | | | | | | | |
| 1 | | | | | | | 2 | | 1 | | | | | 1 | |
| <=2 | | _ | | | | | | | | | | | 6 | | |
| 2 | | 3 | | | | | | | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 4 <=8 | | | | | | ^ | | | | | | | 1 | | |
| 8 | | | 6 | | | 6 | | | | | | | | | |
| 16 | | | 1 | | | 1 | | | | | 3 | 6 | | | |
| 32 | | | ı | | | | | | | | 3 | 1 | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Havana in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 9 | 9 | 9 | 9 | 9 | 9 | 9 | 9 | 9 | 9 | 9 | 9 | 9 | 9 |
| MIC | N of resistant isolates | 1 | 0 | 0 | 0 | 1 | 3 | 0 | 0 | 0 | 2 | 1 | 0 | 0 | 1 |
| <=0.03 | | | | | | | | | | 9 | | | | | |
| 0.03 | | | | | | | 4 | | | | | | | | |
| 0.064 | | | | | | | 2 | | | | | | | | |
| <=0.25 | | | | 9 | | | | | | | | | | 5 | 5 |
| <=0.5 | | | | | 8 | | | | 9 | | | | | | |
| 0.5 | | | | | | | 3 | | | | | | | 2 | 3 |
| <=1 | | 5 | | | | | | 9 | | | | | | | |
| 1 | | | | | 1 | | | | | | | | | 2 | |
| <=2 | | ^ | | | | | | | | | | | 7 | | |
| 2 <=4 | | 3 | | | | | | | | | 2 | | | | |
| 4 | | | | | | | | | | | 3 | | 2 | | |
| <=8 | | | | | | 5 | | | | | | | 2 | | |
| 8 | | | 7 | | | <u> </u> | | | | | 2 | | | | |
| 16 | | | 2 | | | 3 | | | | | 2 | 5 | | | |
| 32 | | | _ | | | 1 | | | | | 2 | 2 | | | |
| >32 | | | | | | | | | | | | | | | 1 |
| 64 | | | | | | | | | | | | 1 | | | |
| >64 | | 1 | | | | | | | | | | | | | |
| >1024 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella IIIa in Feed material of cereal grain origin

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 0.03 | | | | | | | 1 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | | | | | | | 1 | | | | | | | _ |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 64 | | | | | | | | | | | | 1 | | | |
| >64 | | 1 | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella IIIb 61:-:1,5,7 in Cheeses, made from mixed milk from cows, sheep and/or goats - fresh

Sampling Stage: Unspecified

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | | 11 |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| _16 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella IIIb, group 0:47 in Snakes - zoo animal

Sampling Stage: Zoo

Sampling Type: animal sample - organ/tissue

Sampling Context: Clinical investigations

Sampler: Not applicable

Sampling Strategy: Suspect sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | 1 | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella IIIb, group 0:47 in Reptiles - zoo animal

Sampling Stage: Zoo

Sampling Type: animal sample - faeces

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Not specified

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella IIIb, group 0:58 in Snakes - zoo animal

Sampling Stage: Zoo Sampling Type: animal sample - faeces Sampling Context: Monitoring

Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2 Sampler: Industry sampling

Analytical Method:

| anic acid | | | | |
|---|-------------------------------|---------------|---------------|---------------|
| Cefotaxim Cofotaxim Cefotaxim Cofotaxim Cefotaxim Cofotaxim Cofotaxim Cefotaxim Cofotaxim Cefotaxim Cefotaxim Cofotaxim | Ertapenem | lmipenem | Meropenem | Temocillin |
| Cefotaxime synergy test Not Available Not Available Positive/Pres ent Not Available | | | | |
| Ceftazidime synergy test Not Available Not Available Not Available Not Available Not Available Positive/P | ^{Pres} Not Available | Not Available | Not Available | Not Available |
| ECOFF 0.125 0.5 0.5 8 2 2 | 0.06 | 1 | 0.125 | 32 |
| Lowest limit 0.064 0.25 0.064 0.5 0.25 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| Highest limit 32 64 64 64 128 128 | 2 | 16 | 16 | 128 |
| N of tested isolates 1 1 1 1 1 1 1 | 1 | 1 | 1 | 1 |
| N of resistant | 0 | 0 | 0 | 0 |
| <=0.015 | 1 | | | |
| <=0.03 | | | 1 | |
| <=0.064 1 | | | | |
| 0.25 | | 1 | | |
| 2 1 1 | | | | |
| 4 | | | | 1 |
| 8 1 | | | | _ |
| 64 1 | | | | |

Table Antimicrobial susceptibility testing of Salmonella IIIb, group 0:58 in Snakes - zoo animal

Sampling Stage: Zoo

Sampling Type: animal sample - faeces

Sampling Context: Monitoring

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| 2 0.25 32 |
|-----------------|
| |
| 32 |
| |
| 1 |
| 0 |
| |
| |
| 1 |
| |
| |
| |
| |
| |
| |
| |
| |
| |
| |
| |

Table Antimicrobial susceptibility testing of Salmonella Indiana in Meat from duck - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 1 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | | | | | | | 1 | | | | | | | |
| 1 | | | | | | | | | | | | | | 1 | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 11 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | 4 |
| >32 | | 1 | | | | | | | | | | | 1 | | 1 |
| >1024 | | ı | | | | | | | | | | 1 | ı | | |
| 71024 | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sufamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|------------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|-----------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 1 | 1 | 0 | 1 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.5 | | | | | | | | | 1 | | | | | | |
| 0.5 | | | | 1 | | | | | | | | | | | |
| <=1 | | | | | | | | 1 | | | | | | | |
| 1 | | | | | 1 | | 1 | | | | | | | 1 | |
| 4 | | 1 | | | | | | | | | | | | | |
| 16 | | | | | | 1 | | | | | | | | | |
| 32 | | | 1 | | | | | | | | | | | | 1 |
| >32 >64 | | | | | | | | | | | | | 1 | | 1 |
| >128 | | | | | | | | | | | 1 | | 1 | | |
| 1024 | | | | | | | | | | | · · | 1 | | | |
| 1021 | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from broilers (Gallus gallus) - minced meat - intended to be eaten cooked

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sufamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|-----------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 1 | 1 | 0 | 1 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | | | | | | | 1 | | | | | | | |
| 1 | | | | | | | 1 | | | | | | | 1 | |
| 4 | | 1 | | | | 4 | | | | | | | | | |
| 16 32 | | | 1 | | | 11 | | | | | | | | | |
| >32 | | | ı | | | | | | | | | | | | 1 |
| >64 | | | | | | | | | | | | | 1 | | ' |
| >128 | | | | | | | | | | | 1 | | | | |
| >1024 | | | | | | | | | | | | 1 | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Infantis in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census Sampler: Official sampling

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|------------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 10 | 10 | 10 | 10 | 10 | 10 | 10 | 10 | 10 | 10 | 10 | 10 | 10 | 10 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 10 | | | | | |
| 0.03 | | | | | | | 10 | | | | | | | | |
| <=0.25 | | | | 10 | | | | | | | | | | 7 | |
| <=0.5 | | | | | 9 | | | | 10 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 3 | 9 |
| <=1 | | 6 | | | <u> </u> | | | 10 | | | | | | | |
| 1 | | | | | 1 | | | | | | | | 10 | | 1 |
| <=2 | | 4 | | | | | | | | | | | 10 | | |
| 2 <=4 | | 4 | | | | | | | | | 10 | | | | |
| <=4 <=8 | | | | | | 9 | | | | | 10 | | | | |
| 8 | | | 8 | | | | | | | | | | | | |
| 16 | | | 2 | | | 1 | | | | | | 3 | | | |
| 32 | | | | | | ' | | | | | | 5 | | | |
| 64 | | | | | | | | | | | | 2 | | | |
| | | | | | | | | | | | | <u> </u> | | | |

Table Antimicrobial susceptibility testing of Salmonella Infantis in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 4 | | | | | |
| 0.03 | | | | | | | 4 | | | | | | | | |
| <=0.25 | | | | 4 | | | | | | | | | | 3 | 1 |
| <=0.5 | | | | | 3 | | | | 2 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 3 |
| <=1 | | 4 | | | | | | 4 | | | | | | | |
| | | | | | 1 | | | | 1 | | | | | | |
| <=2 | | | | | | | | | | | | | 4 | | |
| 2 | | | | | | | | | 1 | | | | | | |
| <=4 | | | | | | | | | | | 4 | | | | |
| <=8 | | | | | | 3 | | | | | | | | | |
| 8 | | | 4 | | | <u>.</u> | | | | | | | | | |
| 16 | | | | | | 1 | | | | | | | | | |
| 32 | | | | | | | | | | | | 4 | | | |

Table Antimicrobial susceptibility testing of Salmonella Kedougou in Feed material of cereal grain origin

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | 1 | | | | | | | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 11 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Kentucky in Meat from broilers (Gallus gallus) - offal

Sampling Stage: Processing plant

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 1 | 1 | 1 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=1 | | | | | | | | 1 | | | | | | | |
| 1 | | | | | 1 | | | | | | | | | 1 | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| >8 | | | | | | | 1 | | | | | | | | |
| 32 | | | | | | | | | 1 | | | | | | |
| >64 | | 1 | | | | | | | | | | | 1 | | |
| >128 | | | | | | | | | | | 1 | 1 | | | |
| >1024 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Kentucky in Meat from turkey - meat products

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 1 | 1 | 1 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | | | | | | | 1 | | | | | | | |
| 1 | | | | | 1 | | | | | | | | | | |
| <=8 | | | 1 | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | 1 | | | | | | | | |
| >8 16 | | | | | | | 1 | | 1 | | | | | | |
| >64 | | 1 | | | | | | | ' | | | | 1 | | |
| >128 | | • | | | | | | | | | 1 | | | | |
| >1024 | | | | | | | | | | | • | 1 | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Kimuenza in Feed material of cereal grain origin

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | | | | | | | 1 | 1 | | | | | | |
| <=2 | | | | | | | | | 1 | | | | 1 | | |
| 2 | | 1 | | | | | | | | | | | 1 | | |
| <=4 | | ı | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 16 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Kirkee in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Context: Control and eradication

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 1 | | | | | | | | | | |
| 0.5 | | | | | | | | | | | | | | | 1 |
| <=1 | | | | | | | | 1 | | | | | | | |
| 1 | | | | | | | | | 1 | | | | 4 | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 <=4 | | ı | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | ı | | | | |
| 8 | | | 1 | | | ' | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |
| | | | | | | | | | | | | • | | | |

Table Antimicrobial susceptibility testing of Salmonella Kirkee in Gallus gallus (fowl) - breeding flocks for broiler production line

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Objective sampling

Sampling Context: Control and eradication

programmes

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | | 11 |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Lexington in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | | | | | | | 1 | | | |
| 8 | | | 1 | | | 1 | | | | | | | | | |
| 16 | | | | | | 1 | | | | | | | | | - |

Table Antimicrobial susceptibility testing of Salmonella Lexington in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|-----------------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| міс | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | 2 | 2 |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| <=1 | | 2 | | | | | | 2 | | | | | | | |
| <=2 | | | | | | | | | | | | | 2 | | |
| <=4 | | | 0 | | | | | | | | 2 | | | | |
| <u>4</u> <=8 | | | 2 | | | 2 | | | | | | | | | |
| 16 | | | | | | | | | | | | 2 | | | |
| 10 | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Llandoff in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and $% \left(\frac{1}{2}\right) =0$

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|------------------------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | | | | | | | |
| <=1 | | | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | <u> </u> | | 11 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | 1 | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | 1 | | | |
| >32 | | | | | | | | | 1 | | | 1 | | | |
| >64 | | 1 | | | | | | | ı | | | | | | |
| ~∪ + | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Llandoff in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampler: Official sampling

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 0.03 | | | | | | | 1 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | 1 | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Llandoff in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

Sampler: Official and industry sampling

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Madelia in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 0.064 | | | | | | | 1 | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | | 1 |
| <=1 | | | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | 1 | | | | | | | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | 1 | | | | | | | | | |
| 64 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Madelia in Meat from sheep

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 0.03 | | | | | | | 1 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | | 11 |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | 1 | | | | | | 1 | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 16 | | | 1 | | | | | | | | | | | | |
| 64 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Mbandaka in Feed material of cereal grain origin

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | | 1 |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 64 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Mbandaka in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampler: Official sampling

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 4 | | | | | | | | |
| <=0.03 | | | | | | | | | | 4 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 5 | | | | | | | | | | 4 | 3 |
| <=0.5 | | | | | 5 | | | | 5 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 2 |
| <=1 | | 5 | | | | | | 5 | | | | | | | |
| <=2 | | | | | | | | | | | | | 5 | | |
| <=4 | | | | | | | | | | | 5 | | | | |
| <=8 | | | | | | 5 | | | | | | | | | |
| 8 | | | 3 | | | | | | | | | | | | |
| 16 | | | 2 | | | | | | | | | - | | | |
| 32 | | | | | | | | | | | | 5 | | | |

Table Antimicrobial susceptibility testing of Salmonella Mbandaka in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 2 | | | | | | | | |
| <=0.03 | | | | | | | | | | 3 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 3 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 3 | | | | 3 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 2 | 2 |
| <=1 | | 3 | | | | | | 3 | | | | | | | |
| <=2 | | | | | | | | | | | | | 3 | | |
| <=4 | | | | | | | | | | | 3 | | | | |
| <=8 | | | | | | 3 | | | | | | | | | |
| 8 | | | 3 | | | | | | | | | 0 | | | |
| 32 | | | | | | | | | | | | 2 | | | |
| 64 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Mbandaka in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Mikawasima in Gallus gallus (fowl) - laying hens - day-old chicks

Sampling Stage: Hatchery

Sampling Type: environmental sample - delivery box liner

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | | 1 |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Molade in Feed material of cereal grain origin

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | | 1 |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 64 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Montevideo in Feed material of cereal grain origin

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | | 2 |
| <=0.5 | | | | | 2 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 2 | |
| <=1 | | 2 | | | | | | 2 | <u> </u> | | | | | | |
| 1 | | | | | | | | | 1 | | | | | | |
| <=2 | | | | | | | | | | | | | 2 | | |
| <=4 | | | | | | 2 | | | | | 2 | | | | |
| <=8 8 | | | 2 | | | 2 | | | | | | | | | |
| 32 | | | | | | | | | | | | 2 | | | |
| 32 | | | | | | | | | | | | 2 | | | |

Table Antimicrobial susceptibility testing of Salmonella Montevideo in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampler: Official sampling

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Montevideo in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampling Context: Control and eradication programmes

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Muenchen in Meat from pig - meat products

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 2 |
| <=1 | | 1 | | | | | | 2 | | | | | | | |
| <=2 | | 4 | | | | | | | | | | | 2 | | |
| 2 <=4 | | 1 | | | | | | | | | 1 | | | | |
| 4 | | | 1 | | | | | | | | <u> </u> | | | | |
| <=8 | | | ı | | | 2 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | 1 | | | | |
| 32 | | | , | | | | | | | | | 2 | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Newport in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampler: Official sampling

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | 11 | | | | | | | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Newport in Turkeys

Sampling Stage: Farm

Sampling Type: animal sample - organ/tissue

Sampling Context: Clinical investigations

Sampler: Industry sampling

Sampling Strategy: Suspect sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 11 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 4 | | | 11 | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Newport in Meat from turkey - meat products

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | 1 | 1 |
| <=1 | | | | | | | | 1 | | | | | | | |
| 2 | | | | | | | | | 1 | | | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 32 | | | | | | | | | | | 1 | 1 | | | |
| >64 | | 1 | | | | | | | | | | | 1 | | |

Table Antimicrobial susceptibility testing of Salmonella Newport in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 11 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Ohio in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and $% \left(\frac{1}{2}\right) =0$

Sampling Context: Control and eradication

Sampler: Official sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | 11 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Ouakam in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampler: Official sampling

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | 1 | | | | | | 11 | <u> </u> | | | | | | |
| 1 | | | | | | | | | 1 | | | | | | |
| <=2 | | | | | | | | | | | 4 | | 1 | | |
| <=4 | | | | | | 4 | | | | | 1 | | | | |
| <=8 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | ' | | | | | | | | | 1 | | | |
| UL. | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Reading in Meat from pig - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|-------------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 3 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 3 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| <=0.25 | | | | 3 | | | | | | | | | | 3 | 1 |
| <=0.5 | | | | | 3 | | | | 3 | | | | | | |
| 0.5 | | | | | | | | | | | | | | | 2 |
| <=1 | | 3 | | | | | | 3 | | | • | | | | |
| <=4 | | | | | | | | | | | 3 | | | | |
| 4 | | | 2 | | | 2 | | | | | | | | | |
| <=8 8 | | | 1 | | | 3 | | | | | | | | | |
| 16 | | | ' | | | | | | | | | 3 | | | |
| 64 | | | | | | | | | | | | | 3 | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Rissen in Meat from bovine animals and pig - minced meat

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 1 | 0 | 0 | 0 | 1 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | 1 | |
| <=1 | | | | | | | | 1 | | | | | | | |
| 8 | | | 1 | | | | | | | | 1 | | | | |
| 16 | | | | | | | | | | | | 1 | | | |
| >64 | | 1 | | | | | | | | | | | 1 | | |
| >128 | | | | | | 1 | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Rissen in Meat from pig - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|-----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 |
| MIC | N of resistant isolates | 4 | 4 | 0 | 0 | 2 | 0 | 0 | 0 | 0 | 0 | 4 | 4 | 0 | 4 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 4 | | | | | |
| 0.03 | | | | | | | 3 | | | | | | | | |
| <=0.25 | | | | 4 | | | | | | | | | | | |
| <=0.5 | | | | | 4 | | | | 3 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | | | | | | | 4 | | | | | | | |
| 1 | | | | | | | | | 1 | | | | | 3 | |
| <=4 | | | | | | 0 | | | | | 4 | | | | |
| <=8 32 | | | 1 | | | 2 | | | | | | | | | |
| >32 | | | ' | | | | | | | | | | | | 4 |
| 64 | | | 2 | | | | | | | | | | | | 4 |
| >64 | | 4 | 1 | | | | | | | | | | 4 | | |
| >128 | | | • | | | 2 | | | | | | | | | |
| >1024 | | | | | | | | | | | | 4 | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Rissen in Meat from bovine animals - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| MIC | N of resistant isolates | 2 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 2 | 1 | 0 | 2 |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| <=1 | | | | | | | | 2 | | | | | | | |
| 1 | | | | | | | | | | | | | | 1 | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 2 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 16 | | | 1 | | | | | | | | | | | | - |
| >32 | | | | | | | | | | | | | | | 2 |
| 64 | | | <u> </u> | | | 1 | | | | | | | <u> </u> | | |
| >64 | | 2 | 1 | | | | | | | | | • | 1 | | |
| >1024 | | | | | | | | | | | | 2 | | | |

Table Antimicrobial susceptibility testing of Salmonella Rissen in Meat from pig - meat products

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|-----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 1 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 |
| 0.064 | | | | | | | 1 | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | <u> </u> | | | | | | 1 | |
| <=1 | | | | | | | | 1 | | | | | | | |
| <=4 64 | | | | | | | | | | | 1 | | | | |
| >64 | | 1 | ' | | | | | | | | | | 1 | | |
| >128 | | 1 | | | | 1 | | | | | | | 1 | | |
| >1024 | | | | | | • | | | | | | 1 | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Senftenberg in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | | 1 |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 4 | | | 1 | | | | | | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | | | | | | | | | 1 | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Senftenberg in Turkeys - unspecified - day-old chicks

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | |
| 0.25 | | | | | | | 1 | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | | 1 |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |
| >128 | | | | | | | | | | | 1 | | | | |

Table Antimicrobial susceptibility testing of Salmonella Szentes in Feed material of cereal grain origin

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 11 |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Tennessee in Meat from turkey - meat preparation - intended to be eaten cooked

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 1 |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Tennessee in Feed material of cereal grain origin

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 1 |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Tennessee in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and $% \left(\frac{1}{2}\right) =0$

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 11 |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Tennessee in Meat from pig - meat products

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 11 |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|-----------------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 |
| MIC | N of resistant isolates | 2 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 1 | 2 | 0 | 0 |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| 0.064 | | | | | | | 1 | | | 1 | | | | | |
| <=0.25 | | | | 3 | | | | | | | | | | | 2 |
| <=0.5 | | | | | 3 | | | | 2 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 3 | 1 |
| <=1 | | | | | | | | 3 | | | | | | | |
| 1 | | | | | | | | | 1 | | | | | | |
| 2 | | 1 | 4 | | | | | | | | | | | | |
| <u>4</u> <=8 | | | 1 | | | 3 | | | | | | 1 | 1 | | |
| 8 | | | 1 | | | <u> </u> | 1 | | | | 2 | ı | | | |
| 16 | | | 1 | | | | I | | | | 2 | 1 | | | |
| >64 | | 2 | <u> </u> | | | | | | | | | <u> </u> | 2 | | |
| >128 | | _ | | | | | | | | | 1 | | | | |
| >1024 | | | | | | | | | | | • | 1 | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from duck - carcase - frozen

Sampling Stage: Processing plant

Sampling Type: food sample - carcase swabs

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Sampling Context: Monitoring

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 0.064 | | | | | | | 1 | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| 2 | | 1 | | | | | | 1 | | | | | | | |
| 4 | | | | | | | | | | | | | 1 | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | 1 | | | | |
| 16 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from bovine animals and pig - minced meat

Sampling Stage: Processing plant Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official and industry sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

A - - | - - | M - + | - - |

| Anal | | | | | | | | | | | |
|---------------------|--|---------------------------------|------------------------------|--|---------------------------------|-------------------------------------|---|-------------------------|----------------------------------|--------------------------------|------------------------------|
| Cour | ntry of Origin: | Portugal | | | | | | | | | |
| | AM substance | Cefepime | Cefotaxim | Cefotaxime + Clavulanic acid | Cefoxitin | Ceftazidim | Ceftazidime + Clavulanic acid | Ertapenem | Imipenem | Meropenem | Temocillin |
| | | | | | | | | | | | |
| | Cefotaxime synergy test | Not Available | Not Available | Positive/Pres ent | Not Available | Not Available N | lot Available | Not Available | Not Available | Not Available | Not Avai |
| | synergy test | | | Positive/Pres ent Not Available | | | | | | Not Available | |
| | synergy test Ceftazidime | | | ent | | | | | | | Not Ava |
| | synergy test Ceftazidime synergy test | Not Available | Not Available | ent Not Available | Not Available | Not Available 6 | Negative/Abs ent | Not Available | Not Available | Not Available | Not Ava |
| | Synergy test Ceftazidime synergy test ECOFF | Not Available | Not Available 0.5 | Not Available | Not Available | Not Available 6 | Negative/Abs ent | Not Available 0.06 | Not Available | Not Available | Not Ava 32 0.5 |
| | Synergy test Ceftazidime synergy test ECOFF Lowest limit | Not Available 0.125 0.064 | Not Available 0.5 0.25 | Not Available 0.5 0.064 | Not Available 8 0.5 | Not Available 6 | Negative/Abs | 0.06 0.015 | Not Available 1 0.12 | Not Available 0.125 0.03 | Not Ava 32 0.5 |
| | Synergy test Ceftazidime synergy test ECOFF Lowest limit Highest limit N of tested | 0.125 0.064 32 | 0.5 0.25 64 | Not Available 0.5 0.064 64 | Not Available 8 0.5 64 | Not Available 6 2 0.25 128 | Negative/Abs ent 2 0.12 128 | 0.06 0.015 2 | Not Available 1 0.12 16 | 0.125 0.03 16 | Not Ava 32 0.5 128 |
| | Synergy test Ceftazidime synergy test ECOFF Lowest limit Highest limit N of tested isolates N of resistant | 0.125 0.064 32 | 0.5 0.25 64 | ent Not Available 0.5 0.064 64 | Not Available 8 0.5 64 | Not Available 2 0.25 128 | 2 0.12 128 | 0.06 0.015 2 | Not Available 1 0.12 16 | 0.125 0.03 16 | Not Ava 32 0.5 128 |
| .03 | Synergy test Ceftazidime synergy test ECOFF Lowest limit Highest limit N of tested isolates N of resistant | 0.125 0.064 32 | 0.5 0.25 64 | ent Not Available 0.5 0.064 64 1 0 | Not Available 8 0.5 64 | Not Available 2 0.25 128 | 2 0.12 128 | 0.06 0.015 2 1 | Not Available 1 0.12 16 | 0.125 0.03 16 | Not Ava 32 0.5 128 |
| .03 .064 | Synergy test Ceftazidime synergy test ECOFF Lowest limit Highest limit N of tested isolates N of resistant | 0.125 0.064 32 | 0.5 0.25 64 | ent Not Available 0.5 0.064 64 | Not Available 8 0.5 64 | Not Available 2 0.25 128 | 2 0.12 128 1 | 0.06 0.015 2 1 | Not Available 1 0.12 16 1 | 0.125 0.03 16 1 | Not Avai 32 0.5 128 |
| .03 .064 | Synergy test Ceftazidime synergy test ECOFF Lowest limit Highest limit N of tested isolates N of resistant | 0.125 0.064 32 | 0.5 0.25 64 | ent Not Available 0.5 0.064 64 1 0 | Not Available 8 0.5 64 | 2 0.25 128 1 | 2 0.12 128 | 0.06 0.015 2 1 | Not Available 1 0.12 16 | 0.125 0.03 16 1 | Not Avai 32 0.5 128 |
| .015 .03 .064 | Synergy test Ceftazidime synergy test ECOFF Lowest limit Highest limit N of tested isolates N of resistant | 0.125 0.064 32 | 0.5 0.25 64 | ent Not Available 0.5 0.064 64 1 0 | Not Available 8 0.5 64 | Not Available 2 0.25 128 | 2 0.12 128 1 | 0.06 0.015 2 1 | Not Available 1 0.12 16 1 | 0.125 0.03 16 1 | Not Avai 32 0.5 128 |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from bovine animals and pig - minced meat

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|-------------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| MIC | N of resistant isolates | 2 | 0 | 1 | 0 | 2 | 0 | 0 | 0 | 0 | 0 | 2 | 2 | 0 | 0 |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 2 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 2 | 1 |
| <=1 | | | | | | | | 2 | | | | | | | |
| 1 | | | | 1 | 1 | | | | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 8 | | 2 | 2 | | | | | | | | 1 | | 2 | | |
| >64 >128 | | 2 | | | | 2 | | | | | | | 2 | | |
| >128 | | | | | | | | | | | | 2 | | | |
| 7 1024 | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from pig - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| MIC | N of resistant isolates | 2 | 0 | 0 | 0 | 2 | 1 | 0 | 1 | 0 | 0 | 2 | 2 | 1 | 1 |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| 0.12 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | | |
| <=0.5 | | | | | 2 | | | | | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 1 |
| <=1 | | | | | | | | 2 | | | | | | | |
| 1 | | | | | | | | | 1 | | | | | | |
| 2 | | | | | | | | | | | | | | 1 | |
| <=4 | | | | | | | | | | | 1 | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 16 | | | 1 | | | | | | <u> </u> | | 1 | | | | |
| >32 | | | | | | | | | 1 | | | | • | | 1 |
| >64 | | 2 | | | | | | | | | | | 2 | | |
| 128 | | | | | | 1 | | | | | | | | | |
| >128 | | | | | | 1 | | | | | | | | | |
| >1024 | | | | | | | | | | | | 2 | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Birds - wild

Sampling Stage: Zoo

Sampling Type: animal sample - faeces

Sampling Context: Monitoring

Sampler: Industry sampling

Sampling Strategy: Not specified

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | <u> </u> | | | | | | | 1 | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | 1 | | | |
| 32 | | | | | | | | | | | 1 | 1 | | | |
| >128 | | | | | | | | | | | T T | | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Birds

Sampling Stage: Natural habitat

Sampling Type: animal sample - cloacal swab

Sampling Context: Monitoring

Sampler: Not applicable

Sampling Strategy: Other

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 1 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | | | | | | | 1 | | | | | | | |
| 1 | | | | | | | | | | | | | | 1 | |
| <=4 | | | | | | 4 | | | | | 1 | | | | |
| <=8 8 | | | 1 | | | 1 | | | | | | | | | |
| >32 | | | ı ı | | | | | | | | | | | | 1 |
| >64 | | 1 | | | | | | | | | | | 1 | | |
| >1024 | | • | | | | | | | | | | 1 | • | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Pigeons

Sampling Stage: Natural habitat

Sampling Type: animal sample - organ/tissue

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|-------------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 0.03 | | | | | | | 1 | | | | | | | | |
| 0.064 | | | | | | | 1 | | | 2 | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | 1 | 2 |
| <=0.5 | | | | | 2 | | | | 2 | | | | | <u> </u> | |
| 0.5 | | 0 | | | | | | • | | | | | | 1 | |
| <=1 <=2 | | 2 | | | | | | 2 | | | | | 2 | | |
| <= <u>2</u> | | | | | | | | | | | 2 | | | | |
| <=8 | | | | | | 2 | | | | | 2 | 2 | | | |
| 8 | | | 2 | | | | | | | | | | | | |
| | | | _ | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 |
| <=0.015 | | | | | | | 1 | | | | | | | | |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | 1 | | | | | | | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | | | | | | | 1 | | | | | | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | 1 | | | | | | |
| >1024 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Partridges

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|------------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | 1 | |
| <=1 | | | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | 1 | | | | | | | | | | | | | |
| <=8 | | | <u> </u> | | | 11 | | | | | | | | | |
| 16 | | | 1 | | | | | | | | | 4 | | | |
| 32 >128 | | | | | | | | | | | 4 | 1 | | | |
| -128 | | | | | | | | | | | 1 | | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from duck - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 | 4 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| 0.064 | | | | | | | 2 | | | 3 | | | | | |
| <=0.25 | | | | 4 | | | | | | | | | | 2 | 4 |
| <=0.5 | | | | | 4 | | | | 4 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 2 | |
| <=1 | | 11 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 3 | | |
| 2 | | 3 | | | | | | 3 | | | | | | | |
| 4 | | | | | | | | | | | | | 11 | | |
| <=8 | | | | | | 4 | | | | | | | | | |
| 8 | | | 2 | | | | | | | | 4 | | | | |
| 16 | | | 2 | | | | | | | | | 3 | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from duck - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sufamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|-----------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.064 | | | | | | | 2 | | | | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 2 | 1 |
| 2 | | 2 | | | | | | 2 | | | | | | | |
| 4 | | | | | | | | | | | | | 2 | | |
| <=8 | | | | | | 2 | | | | | | | | | |
| 8 | | | 2 | | | | | | | | 2 | | | | |
| 32 | | | | | | | | | | | | 2 | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from other poultry species

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 2 | 0 | 0 | 0 | 2 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 3 | | | | | | | | | | 2 | 2 |
| <=0.5 | | | | | 3 | | | | 2 | | | | | | |
| 0.5 | | | | | | | 2 | | | | | | | 1 | 1 |
| <=1 | | 1 | | | | | | 2 | | | | | | | |
| 1 | | | | | | | | | 1 | | | | | | |
| <=2 | | ^ | | | | | | 4 | | | | | 2 | | |
| 2 <=4 | | 2 | | | | | | 1 | | | | | | | |
| 4 | | | 1 | | | | | | | | 1 | | 1 | | |
| <=8 | | | | | | 3 | | | | | | | - 1 | | |
| 8 | | | 2 | | | 3 | | | | | | | | | |
| 16 | | | 2 | | | | | | | | | 1 | | | |
| 32 | | | | | | | | | | | | 2 | | | |
| >128 | | | | | | | | | | | 2 | 2 | | | |
| - 120 | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Turkeys - fattening flocks - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 |
| 0.064 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | 1 | |
| <=1 | | | | | | | | 1 | | | | | | | |
| 2 | | 1 | | | | | | | | | | | | | |
| 4 | | | | | | | | | | | | | 1 | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |
| >128 | | | | | | | | | | | 1 | | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampler: Official sampling

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 6 | 0 | 0 | 0 | 6 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 3 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| 0.064 | | | | | | | | | | 4 | | | | | |
| <=0.25 | | | | 7 | | | | | | | | | | 1 | 7 |
| 0.25 | | | | | | | 11 | | | | | | | | |
| <=0.5 | | | | | 7 | | _ | | 6 | | | | | | |
| 0.5 | | | | | | | 5 | 7 | | | | | | 6 | |
| <=1 1 | | 5 | | | | | | 1 | 1 | | | | | | |
| <=2 | | | | | | | | | ' | | | | 7 | | |
| 2 | | 2 | | | | | | | | | | | ı | | |
| <=4 | | _ | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 7 | | | | | | | | | |
| 8 | | | 4 | | | | | | | | | | | | |
| 16 | | | 3 | | | | | | | | | 1 | | | |
| 32 | | | | | | | | | | | | 6 | | | |
| >128 | | | | | | | | | | | 6 | | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from duck - offal - liver

Sampling Stage: Processing plant

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| міс | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 0.03 | | | | | | | 2 | | | | | | | | |
| 0.064 | | | | | | | | | | 2 | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | | 11 |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 2 | 1 |
| <=1 | | | | | | | | 11 | | | | | | | |
| <=2 | | - | | | | | | | | | | | 2 | | |
| 2 | | 2 | | | | | | 1 | | | | | | | |
| <=8 | | | 0 | | | 2 | | | | | 0 | | | | |
| 16 | | | 2 | | | | | | | | 2 | 1 | | | |
| 32 | | | | | | | | | | | | 1 | | | |
| JZ | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from bovine animals - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Sampling Context: Monitoring

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | 1 | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from pig - meat products

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | 1 | | | | | | 1 | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | 1 | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from pig - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| МІС | N of resistant isolates | 1 | 0 | 0 | 0 | 2 | 0 | 1 | 0 | 0 | 0 | 2 | 2 | 0 | 1 |
| 0.03 | | | | | | | 2 | | | | | | | | |
| 0.064 | | | | | | | | | | 2 | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 1 |
| <=1 | | | | | | | | 1 | | | | | | | |
| 2 | | 1 | | | | | | | | | | | | | |
| <=4 | | | | | | | | | | | 2 | | | | |
| 8 | | | 2 | | | | | 1 | | | | | | | |
| >32 | | | | | | | | | | | | | | | 1 |
| 64 | | | | | | | | | | | | | 1 | | |
| >64 | | 1 | | | | <u> </u> | | | | | | | 1 | | |
| >128 | | | | | | 2 | | | | | | | | | |
| >1024 | | | | | | | | | | | | 2 | | | |

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from pig - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | 1 |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Uppsala in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| >32 | | | | | | | | | | | | | | | 1 |
| >1024 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Virchow in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampler: Official sampling

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 11 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| міс | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | 1 | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | | 1 |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | 1 | | | | | | 11 | | | | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 32 | | | | | | | | | | | | 1 | | | |
| >128 | | | | | | | | | | | 1 | | | | |

Table Antimicrobial susceptibility testing of Salmonella Virchow in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs and

Sampler: Official sampling

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|--------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| МІС | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 1 | | | | | |
| 0.03 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 1 | | | | | | | | | | | |
| <=0.5 | | | | | 1 | | | | 1 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 1 | 11 |
| <=1 | | 1 | | | | | | 1 | | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| <=4 | | | | | | | | | | | 1 | | | | |
| <=8 | | | | | | 1 | | | | | | | | | |
| 8 | | | 1 | | | | | | | | | | | | |
| 64 | | | | | | | | | | | | 1 | | | |

Table Antimicrobial susceptibility testing of Salmonella Virchow in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampler: Official and industry sampling

Sampling Type: environmental sample - boot swabs and

Sampling Strategy: Census

Sampling Context: Control and eradication

programmes Programme Code: AMR MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.5 | 2 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 256 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| MIC | N of resistant isolates | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| <=0.03 | | | | | | | | | | 2 | | | | | |
| 0.03 | | | | | | | 2 | | | | | | | | |
| <=0.25 | | | | 2 | | | | | | | | | | | |
| <=0.5 | | | | | 2 | | | | 2 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 2 | 2 |
| <=1 | | 1 | | | | | | 2 | | | | | | | |
| <=2 | | 1 | | | | | | | | | | | 2 | | |
| 2 <=4 | | 1 | | | | | | | | | 2 | | | | |
| <=8 | | | | | | 2 | | | | | | | | | |
| 8 | | | 2 | | | 2 | | | | | | | | | |
| 32 | | | | | | | | | | | | 2 | | | |
| | | | | | | | | | | | | | | | |

ANTIMICROBIAL RESISTANCE TABLES FOR INDICATOR ESCHERICHIA COLI

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from bovine animals - fresh

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: ESBL MON pnl2

Analytical Method:

| | AM substance | | | | • Clavulanic acid | | | | + Clavulanic acid | | | | | |
|--------|--------------------------|---------------|---------------|----------------------|-------------------|------------------|-----------------|-------------|----------------------|--------------------|------------------|---------------|---------------|-------------|
| | | Cefepime | Cefotaxim | | Cefotaxime · | Cefoxitin | Ceftazidim | | Ceffazidime | | Ertapenem | Imipenem | Meropenem | Temocillin |
| | Cefotaxime synergy test | Not Available | Not Available | Positive/Pres ent | Negative/Al | Not Available | Not Available | | Not Available | | Not Available | Not Available | Not Available | Not Availab |
| | Ceftazidime synergy test | Not Available | Not Available | Not Available | Not Availab | le Not Available | Not Available N | ot Availabl | Positive/Pres ent | Negative/Al ent | OS Not Available | Not Available | Not Available | Not Availab |
| | ECOFF | 0.125 | 0.25 | 0.25 | 0.25 | 8 | 0.5 | 0.5 | 0.5 | 0.5 | 0.064 | 0.5 | 0.125 | 32 |
| | Lowest limit | 0.064 | 0.25 | 0.064 | 0.064 | 0.5 | 0.25 | 0.12 | 0.12 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| | Highest limit | 32 | 64 | 64 | 64 | 64 | 128 | 128 | 128 | 128 | 2 | 16 | 16 | 64 |
| | N of tested isolates | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 |
| | N of resistant isolates | 25 | 26 | 3 | 3 | 4 | 25 | 3 | 3 | 3 | 1 | 0 | 0 | 0 |
| 0.015 | | | | | | | | | | | 17 | | | |
| 0.03 | | | | | | | | | | | | | 25 | |
| 3 | | | | | | | | | | | 7 | | | |
| 0.064 | | 1 | | 16 | | | | | | | | | | |
| 64 | | | | | | | | | | | 11 | | 1 | |
|).12 | | | | | | | | 1 | 13 | | 1 | 6 | | |
| 2 5 | | 1 | | 7 | | | | | 6 | 1 | 1 | 15 | | |
| 3 | | 5 | | | | | 1 | | 2 | I | | 5 | | |
| | | | | | | | 7 | | | | | <u> </u> | | |
| | | | | | | 1 | 3 | | | | | | | 2 |
| | | | | | | | | | | | | | | |
| | | 6 | 3 | | | 10 | 1 | | | | | | | 5 |

| | AM substance | Cefepime | Cefotaxim | | | Cefoxitin | Ceftazidim | | Ceftazidime + Clavulanic acid | | Ertapenem | Imipenem | Meropenem | Temocillin |
|-----|----------------------------|---------------|---------------|----------------------|--------------------|------------------|-----------------|---------------|-------------------------------|-------------------|------------------|---------------|---------------|---------------|
| | Cefotaxime synergy test | Not Available | Not Available | Positive/Pres ent | Negative/Ab ent | S Not Available | Not Available | | Not Available | | | | Not Available | |
| | Ceftazidime synergy test | Not Available | Not Available | Not Available | Not Availab | le Not Available | Not Available I | Not Available | Positive/Pres N ent | egative/Al ent | bs Not Available | Not Available | Not Available | Not Available |
| | ECOFF | 0.125 | 0.25 | 0.25 | 0.25 | 8 | 0.5 | 0.5 | 0.5 | 0.5 | 0.064 | 0.5 | 0.125 | 32 |
| | Lowest limit | 0.064 | 0.25 | 0.064 | 0.064 | 0.5 | 0.25 | 0.12 | 0.12 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| | Highest limit | 32 | 64 | 64 | 64 | 64 | 128 | 128 | 128 | 128 | 2 | 16 | 16 | 64 |
| | N of tested isolates | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 |
| MIC | N of resistant isolates | 25 | 26 | 3 | 3 | 4 | 25 | 3 | 3 | 3 | 1 | 0 | 0 | 0 |
| 16 | | 6 | 3 | | | 1 | 8 | | | 1 | | | | 5 |
| 32 | | 2 | 6 | | | | 3 | | | | | | | |
| >32 | | 1 | | | | | | | | | | | | |
| 64 | | | 5 | | | 3 | | | | | | | | |
| >64 | | | 7 | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from bovine animals - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|---------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.25 | 0.5 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 64 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 |
| МІС | N of resistant isolates | 26 | 7 | 26 | 23 | 12 | 18 | 4 | 5 | 0 | 18 | 20 | 19 | 0 | 14 |
| <=0.015 | | | | | | | 2 | | | | | | | | |
| <=0.03 | | | | | | | | | | 26 | | | | | |
| 0.03 | | | | | | | 5 | | | | | | | | |
| 0.064 | | | | | | | 1 | | | | | | | | |
| 0.12 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | | | | | | | | | | | 21 | 7 |
| 0.25 | | | | | | | 3 | | | | | | | | |
| <=0.5 | | | | | 3 | | | | 6 | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | 5 | 3 |
| <=1 | | | | | | | | 21 | | | | | | | - |
| 1 | | | | | 8 | | | | 13 | | | | | | 2 |
| <=2 | | | 2 | | | | | | | | | | 4 | | |
| 2 | | | | | 1 | | | 1 | 2 | | | | | | |
| <=4 4 | | | 8 | 3 | | | | 4 | 1 | | 7 | | 3 | | |
| >4 | | | 0 | 23 | | | | 4 | <u> </u> | | | | <u> </u> | | |
| <=8 | | | | 23 | | 13 | | | | | | 5 | | | |
| 8 | | | 8 | | 3 | 15 | 1 | | 1 | | | <u> </u> | | | |
| >8 | | | <u> </u> | | 11 | | 12 | | • | | | | | | |
| 16 | | | 1 | | | 1 | | | | | 1 | 1 | | | |
| 32 | | | 2 | | | 1 | | | 2 | | | | | | |
| >32 | | | | | | | | | <u>_</u> 1 | | | | | | 14 |
| 64 | | | 3 | | | | | | | | 1 | | 6 | | |
| | | | | | | | | | | | | | | | |

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|-------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.25 | 0.5 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 64 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 | 26 |
| MIC | N of resistant isolates | 26 | 7 | 26 | 23 | 12 | 18 | 4 | 5 | 0 | 18 | 20 | 19 | 0 | 14 |
| >64 | | 26 | 2 | | | | | | | | | | 13 | | |
| 128 | | | | | | 8 | | | | | 3 | | | | |
| >128 | | | | | | 3 | | | | | 14 | | | | |
| >1024 | | | | | | | | | | | | 20 | | | |

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Cattle (bovine animals) - calves (under 1 year)

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON pnl2

Analytical Method:

| | AM substance | Cefepime | Cefotaxim | - - - | Cetotaxime + Clavinanic acid + Clavinanic acid + Clavinanic acid + Clavinanic acid | Cefoxitin | Ceftazidim | | Centazionne + Cravulaine acio | Ertapenem | lmipenem | Meropenem | Temocillin |
|---------|-----------------------------|---------------|---------------|---------------|--|---------------|-----------------|---------------------|-------------------------------|---------------|---------------|---------------|---------------|
| | synergy test | Not Available | Not Available | ent | ent | Not Available | Not Available | | ailable | | | Not Available | |
| | Ceftazidime synergy test | Not Available | Not Available | Not Available | Not Available | Not Available | Not Available P | ositive/Pres ent | Negative/All ent | Not Available | Not Available | Not Available | Not Available |
| | ECOFF | 0.125 | 0.25 | 0.25 | 0.25 | 8 | 0.5 | 0.5 | 0.5 | 0.064 | 0.5 | 0.125 | 32 |
| | Lowest limit | 0.064 | 0.25 | 0.064 | 0.064 | 0.5 | 0.25 | 0.12 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| | Highest limit | 32 | 64 | 64 | 64 | 64 | 128 | 128 | 128 | 2 | 16 | 16 | 64 |
| | N of tested isolates | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 |
| MIC | N of resistant isolates | 5 | 6 | 1 | 1 | 1 | 6 | 1 | 1 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | | | | 5 | | | |
| <=0.03 | | | | | | | | | | | | 6 | |
| 0.03 | | | | | | | | | | 1 | | | |
| <=0.064 | | | | 5 | | | | | | | | | |
| <=0.12 | | | | | | | | 4 | | | | | |
| 0.12 | | 1 | | | | | | | | | | | |
| 0.25 | | | | | | | | 11 | | | 5 1 | | |
| 1 | | | 1 | | 1 | | | | | | ı | | |
| 2 | | | <u>'</u> | | <u>'</u> | 2 | 1 | | 1 | | | | 1 |
| 4 | | 1 | | | | 3 | 4 | | | | | | 2 |
| 8 | | 2 | | | | | 1 | | | | | | 2 |
| 16 | | 1 | | | | 1 | | | | | | | 1 |
| 32 | | | 2 | | | | | | | | | | |
| >32 | | 1 | | | | | | | | | | | |
| 64 | | | 2 | | | | | | | | | | |
| | | | | | | | | | | | | | |

| | AM substance | Cefepime | Cefotaxim | | Cefotaxime + Clavulanic acid | Cefoxitin | Ceftazidim | | Ceffazidime + Clavulanic acid | Ertapenem | Ітірепет | Meropenem | Temocillin |
|-----|--------------------------|---------------|---------------|---------------|------------------------------|---------------|------------------|--------------------|-------------------------------|-----------------|---------------|---------------|---------------|
| | Cefotaxime synergy test | Not Available | Not Available | Positive/Pres | s Negative/Abs ent | Not Available | Not Available | Not A | Available | Not Available | Not Available | Not Available | Not Available |
| | Ceftazidime synergy test | Not Available | Not Available | Not Available | e Not Available | Not Available | Not Available Po | ositive/Pre ent | s Negative/Ab | S Not Available | Not Available | Not Available | Not Available |
| | ECOFF | 0.125 | 0.25 | 0.25 | 0.25 | 8 | 0.5 | 0.5 | 0.5 | 0.064 | 0.5 | 0.125 | 32 |
| | Lowest limit | 0.064 | 0.25 | 0.064 | 0.064 | 0.5 | 0.25 | 0.12 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| | Highest limit | 32 | 64 | 64 | 64 | 64 | 128 | 128 | 128 | 2 | 16 | 16 | 64 |
| | N of tested isolates | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 |
| MIC | N of resistant isolates | 5 | 6 | 1 | 1 | 1 | 6 | 1 | 1 | 0 | 0 | 0 | 0 |
| >64 | | | 1 | | | | | · | | | | | |

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Cattle (bovine animals) - calves (under 1 year)

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|-----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.25 | 0.5 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 64 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 181 | 181 | 181 | 181 | 181 | 181 | 181 | 181 | 181 | 181 | 181 | 181 | 181 | 181 |
| MIC | N of resistant isolates | 18 | 0 | 6 | 6 | 8 | 6 | 0 | 3 | 0 | 6 | 24 | 42 | 0 | 14 |
| <=0.015 | | | | | | | 134 | | | | | | | | |
| <=0.03 | | | | | | | | | | 180 | | | | | |
| 0.03 | | | | | | | 41 | | | | | | | | |
| 0.064 | | | | | | | | | | 1 | | | | | |
| 0.12 | | | | | | | 2 | | | | | | | | |
| <=0.25 | | | | 175 | | | | | | | | | | 161 | 84 |
| 0.25 | | | | | | | 4 | | | | | | | | |
| <=0.5 | | | | | 175 | | | | 111 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 20 | 76 |
| <=1 | | 10 | | | | | | 181 | | | | | | | |
| 1 | | | | 1 | | | | | 61 | | | | | | 7 |
| <=2 | | | 14 | | | | | | | | | | 133 | | |
| 2 | | 68 | | | | | | | 6 | | | | | | |
| <=4 | | | | | | | | | | | 171 | | | | |
| 4 | | 84 | 76 | | 5 | | | | | | | | 5 | | |
| >4 | | | | 5 | | | | | | | | | | | |
| <=8 | | | | | | 171 | | | | | | 136 | | | |
| 16 | | 1 | 83 | | 1 | | | | | | 4 | 4- | 1 | | |
| | | 1 | 8 | | | 2 | | | 2 | | | 17 | 3 | | |
| 32 | | 1 | | | | | | | 4 | | | 4 | | | 4.4 |
| >32 | | | | | | | | | 1 | | 4 | | 40 | | 14 |
| 64 >64 | | 10 | | | | | | | | | 1 | | 13 26 | | |
| >04 | | 16 | | | | | | | | | | | ∠6 | | |

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|-------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.25 | 0.5 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 64 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 181 | 181 | 181 | 181 | 181 | 181 | 181 | 181 | 181 | 181 | 181 | 181 | 181 | 181 |
| МІС | N of resistant isolates | 18 | 0 | 6 | 6 | 8 | 6 | 0 | 3 | 0 | 6 | 24 | 42 | 0 | 14 |
| 128 | | | | | | 5 | | | | | 1 | 1 | | | |
| >128 | · | | | | | 3 | | | | | 4 | | | | |
| >1024 | | | | | | | | | | | | 23 | | | |

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Cattle (bovine animals) - calves (under 1 year)

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method:

| | AM substance | Cefepime | Cefotaxim | | Cefotaxime + Clavulanic acid | Cefoxitin | Ceftazidim | | Ceffazidime + Clavulanic acid | Ertapenem | Imipenem | Meropenem | Temocillin |
|---------|----------------------------|---------------|---------------|----------------------|------------------------------|---------------|-----------------|---------------------|-------------------------------|---------------|---------------|---------------|---------------|
| | Cefotaxime synergy test | Not Available | Not Available | Positive/Pres ent | Negative/Abs ent | Not Available | Not Available | Not A | ailable | Not Available | Not Available | Not Available | Not Available |
| | Ceftazidime synergy test | Not Available | Not Available | Not Available | Not Available | Not Available | Not Available P | ositive/Pres ent | Negative/Ab | Not Available | Not Available | Not Available | Not Available |
| | ECOFF | 0.125 | 0.25 | 0.25 | 0.25 | 8 | 0.5 | 0.5 | 0.5 | 0.064 | 0.5 | 0.125 | 32 |
| | Lowest limit | 0.064 | 0.25 | 0.064 | 0.064 | 0.5 | 0.25 | 0.12 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| | Highest limit | 32 | 64 | 64 | 64 | 64 | 128 | 128 | 128 | 2 | 16 | 16 | 64 |
| | N of tested isolates | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 |
| МІС | N of resistant isolates | 107 | 107 | 1 | 1 | 4 | 105 | 1 | 1 | 1 | 0 | 0 | 0 |
| <=0.015 | | | | | | | | | | 77 | | | |
| <=0.03 | | | | | | | | | | | | 105 | |
| 0.03 | | | | | | | | | | 25 | | | |
| <=0.064 | | | | 93 | | | | | | | | | |
| 0.064 | | | | | | | | | | 4 | | 2 | |
| <=0.12 | | | | | | | | 65 | | | 39 | | |
| 0.12 | | | | 13 | | | | | | 1 | | | |
| 0.25 | | | | | | | | 37 | | | 58 | | |
| 0.5 | | 1 | | | | 1 | 2 | 4 | | | 10 | | |
| 2 | | 6 | | | | 1 17 | 10 | | | | | | 7 |
| 4 | | 17 | | | 1 | 61 | 46 | | | | | | 34 |
| 8 | | 27 | 1 | | I | 24 | 35 | | 1 | | | | 57 |
| 16 | | 21 | 5 | | | 3 | 11 | | | | | | 8 |
| 32 | | 26 | 30 | | | | 1 | | | | | | 1 |
| >32 | | 9 | | | | | | | | | | | |
| | | | | | | | | | | | | | |

| | AM substance | Cefepime | Cefotaxim | | Cefotaxime + Clavulanic acid | Cefoxitin | Ceftazidim | | Ceftazidime + Clavulanic acid | Ertapenem | Imipenem | Meropenem | Temocillin |
|-----|--------------------------|---------------|---------------|----------------------|------------------------------|-----------------|---------------|---------------|-------------------------------|---------------|---------------|---------------|---------------|
| | Cefotaxime synergy test | Not Available | Not Available | Positive/Pres ent | Negative/Ab ent | S Not Available | Not Available | Not A | vailable | Not Available | Not Available | Not Available | Not Available |
| | Ceftazidime synergy test | Not Available | Not Available | Not Available | Not Availabl | e Not Available | Not Available | Positive/Pres | Negative/Ab | Not Available | Not Available | Not Available | Not Available |
| | ECOFF | 0.125 | 0.25 | 0.25 | 0.25 | 8 | 0.5 | 0.5 | 0.5 | 0.064 | 0.5 | 0.125 | 32 |
| | Lowest limit | 0.064 | 0.25 | 0.064 | 0.064 | 0.5 | 0.25 | 0.12 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| | Highest limit | 32 | 64 | 64 | 64 | 64 | 128 | 128 | 128 | 2 | 16 | 16 | 64 |
| | N of tested isolates | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 |
| міс | N of resistant isolates | 107 | 107 | 1 | 1 | 4 | 105 | 1 | 1 | 1 | 0 | 0 | 0 |
| 64 | | | 45 | | | 1 | | | | | | | |
| >64 | | | 26 | | | · | | | | · | | | |

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Cattle (bovine animals) - calves (under 1 year)

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.25 | 0.5 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 64 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 |
| MIC | N of resistant isolates | 107 | 9 | 107 | 105 | 37 | 49 | 1 | 14 | 0 | 26 | 66 | 91 | 0 | 55 |
| <=0.015 | | | | | | | 42 | | | | | | | | |
| <=0.03 | | | | | | | | | | 105 | | | | | |
| 0.03 | | | | | | | 15 | | | | | | | | |
| 0.064 | | | | | | | 1 | | | 1 | | | | | |
| 0.12 | | | | | | | 1 | | | 1 | | | | | |
| <=0.25 | | | | | | | | | | | | | | 84 | 16 |
| 0.25 | | | | | | | 18 | | | | | | | | |
| <=0.5 | | | | | 2 | | | | 65 | | | | | | |
| 0.5 | | | | | | | 8 | | | | | | | 23 | 33 |
| <=1 | | | | | | | | 106 | | | | | | | |
| 1 | | | | | 4 | | 2 | | 23 | | | | | | 3 |
| <=2 | | | 1 | | | | | | | | | | 14 | | |
| 2 | | | | | 11 | | | | 5 | | | | | | |
| <=4 | | | 20 | | 40 | | | | | | 59 | | - | | |
| 4 >4 | | | 32 | 107 | 46 | | | | 2 | | | | 2 | | |
| <=8 | | | | 107 | | 69 | | | | | | 33 | | | |
| | | | 58 | | 28 | 69 | 3 | 1 | | | 16 | აა | | | |
| 8 >8 | | | 36 | | 16 | | 17 | | | | 10 | | | | |
| 16 | | | 7 | | 10 | 1 | 17 | | 3 | | 6 | 7 | | | |
| 16 32 | | | 1 | | | I | | | 4 | | 0 | , | 3 | | |
| >32 | | | • | | | | | | 5 | | | | | | 55 |
| 64 | | | 5 | | | 3 | | | | | | 1 | 20 | | |
| | | | | | | • | | | | | | • | | | |

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|-------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.25 | 0.5 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 64 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 | 107 |
| MIC | N of resistant isolates | 107 | 9 | 107 | 105 | 37 | 49 | 1 | 14 | 0 | 26 | 66 | 91 | 0 | 55 |
| >64 | | 107 | 3 | | | | | | | | | | 68 | | |
| 128 | | | | | | 15 | | | | | 1 | | | | |
| >128 | | | | | | 19 | | | | | 25 | | | | |
| >1024 | | | | | | | | | | | | 66 | | | |

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Pigs - fattening pigs

Sampling Stage: Slaughterhouse Sampling Type: animal sample - caecum Sampling Context: Monitoring

Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: AMR MON pnl2

Analytical Method:

Country of Origin: Portugal

| | AM substance | Cefepime | Cefotaxim | Cefotaxime + Clavulanic acid | Cefoxitin | Ceftazidim | Ceftazidime + Clavulanic acid | Ertapenem | Imipenem | Meropenem | Temocillin |
|---------|-----------------------------|---------------|---------------|------------------------------|---------------|---------------|-------------------------------|---------------|---------------|---------------|---------------|
| | Cefotaxime synergy test | Not Available | Not Available | Positive/Pres ent | Not Available | Not Available | | | | | |
| | Ceftazidime synergy test | Not Available | Not Available | Not Available | Not Available | Not Available | Positive/Pres ent | Not Available | Not Available | Not Available | Not Available |
| | ECOFF | 0.125 | 0.25 | 0.25 | 8 | 0.5 | 0.5 | 0.064 | 0.5 | 0.125 | 32 |
| | Lowest limit | 0.064 | 0.25 | 0.064 | 0.5 | 0.25 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| | Highest limit | 32 | 64 | 64 | 64 | 128 | 128 | 2 | 16 | 16 | 64 |
| | N of tested isolates | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 |
| MIC | N of resistant isolates | 5 | 5 | 0 | 0 | 5 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | | 4 | | | |
| <=0.03 | | | | | | | | | | 5 | |
| 0.03 | | | | | | | | 1 | | | |
| <=0.064 | | | | 5 | | | | | | | |
| <=0.12 | | | | | | | 4 | | | | |
| 0.25 | | 1 | | | | | 1 | | 5 | | |
| 0.5 | | 11 | | | | | | | | | |
| 2 | | | | | | 1 | | | | | 1 |
| 4 | | | 1 | | 2 | 1 | | | | | 2 |
| 8 | | 1 | 1 | | 3 | 1 | | | | | 2 |
| 16 | | 2 | | | | 1 | | | | | |
| 32 | | | 0 | | | 1 | | | | | |
| 64 | | | 2 | | | | | | | | |
| >64 | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Pigs - fattening pigs

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.25 | 0.5 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 64 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 142 | 142 | 142 | 142 | 142 | 142 | 142 | 142 | 142 | 142 | 142 | 142 | 142 | 142 |
| MIC | N of resistant isolates | 85 | 23 | 5 | 5 | 63 | 30 | 3 | 4 | 0 | 17 | 97 | 127 | 0 | 80 |
| <=0.015 | | | | | | | 87 | | | | | | | | |
| <=0.03 | | | | | | | _ | | | 141 | | | | | |
| 0.03 | | | | | | | 21 | | | | | | | | |
| 0.064 | | | | | | | 4 | | | 1 | | | | | |
| 0.12 | | | | | | | 7 | | | | | | | | |
| <=0.25 | | | | 137 | | | | | | | | | | 111 | 34 |
| 0.25 | | | | | | | 10 | | | | | | | | |
| <=0.5 | | | | | 137 | | | | 72 | | | | | | |
| 0.5 | | | | | | | 6 | | | | | | | 30 | 24 |
| <=1 | | 4 | | | | | | 136 | | | | | | | |
| 1 | | | | | | | | | 61 | | | | | 1 | 3 |
| <=2 | | | 9 | | | | | | | | | | 11 | | |
| 2 | | 20 | | | | | | 3 | 5 | | | | | | 1 |
| <=4 | | | | | | | | | | | 119 | | | | |
| 4 | | 31 | 50 | | 2 | | | 2 | | | | | 4 | | |
| >4 | | | | 5 | | | | | | | | | | | |
| <=8 | | | | | | 72 | | | | | | 40 | | | |
| 8 | | 2 | 55 | | 1 | | 2 | 1 | | | 5 | | | | |
| >8 | | | | | 2 | | 5 | | | | | | | | |
| 16 | | | 5 | | | 7 | | | 2 | | 1 | 4 | 1 | | |
| 32 | | 2 | 2 | | | 11 | | | 2 | | 1 | 1 | 1 | | |
| >32 | | | | | | | | | | | | | | | 80 |
| 64 | | | 3 | | | 21 | | | | | 2 | | 24 | | |

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|-------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.25 | 0.5 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 64 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 142 | 142 | 142 | 142 | 142 | 142 | 142 | 142 | 142 | 142 | 142 | 142 | 142 | 142 |
| MIC | N of resistant isolates | 85 | 23 | 5 | 5 | 63 | 30 | 3 | 4 | 0 | 17 | 97 | 127 | 0 | 80 |
| >64 | | 83 | 18 | | | | | | | | | | 101 | | |
| 128 | | | | | | 15 | | | | | 2 | | | | |
| >128 | | | | | | 16 | | | | | 12 | | | | |
| 1024 | | | | | | | | | | | | 1 | | | |
| >1024 | | | | | | | | | | | | 96 | | | |

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Pigs - fattening pigs

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method:

| | AM substance | Cefepime | Cefotaxim | | Cefotaxime + Clavulanic acid | Cefoxitin | Ceftazidim | | Ceftazidime + Clavulanic acid | | Ertapenem | Imipenem | Meropenem | Temocillin |
|------------|----------------------------|---------------|---------------|----------------------|------------------------------|------------------|-----------------|--------------|-------------------------------|--------------------|------------------|---------------|---------------|---------------|
| | Cefotaxime synergy test | Not Available | Not Available | Positive/Pres ent | Negative/Ab ent | S Not Available | Not Available | | Not Available | | Not Available | Not Available | Not Available | Not Available |
| | Ceftazidime synergy test | Not Available | Not Available | Not Available | Not Availabl | le Not Available | Not Available N | ot Available | Positive/Pres N | legative/Ab ent | OS Not Available | Not Available | Not Available | Not Available |
| | ECOFF | 0.125 | 0.25 | 0.25 | 0.25 | 8 | 0.5 | 0.5 | 0.5 | 0.5 | 0.064 | 0.5 | 0.125 | 32 |
| | Lowest limit | 0.064 | 0.25 | 0.064 | 0.064 | 0.5 | 0.25 | 0.12 | 0.12 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| | Highest limit | 32 | 64 | 64 | 64 | 64 | 128 | 128 | 128 | 128 | 2 | 16 | 16 | 64 |
| | N of tested isolates | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 |
| MIC | N of resistant isolates | 148 | 151 | 22 | 22 | 32 | 145 | 22 | 22 | 22 | 10 | 0 | 0 | 0 |
| <=0.015 | | | | | | | | | | | 94 | | | |
| <=0.03 | | | | | | | | | | | | | 137 | |
| 0.03 | | | | | | | | | | | 33 | | | |
| <=0.064 | | 1 | | 101 | | | | | | | | | | |
| 0.064 | | | | | | | | | | | 14 | | 14 | |
| <=0.12 | | | | | | | | 5 | 50 | 1 | | 25 | | |
| 0.12 | | 2 | | 26 | | | | | | | 5 | | | |
| 0.25 | | 7 | | 2 | | | | | 64 | 4 | 4 | 109 | | |
| 0.5 | | 12 | | | | | 6 | | 5 | | | 17 | | |
| 1 | | 5 | | | | | 20 | | | | 1 | | | |
| 2 | | 9 | 2 | | 2 | 9 | 39 | | , | 2 | | | | 5 |
| 4 | | 21 | - | 1 | | 67 | 19 | | 1 | 1 | | | | 46 |
| 8 | | 41 | 5 | | 9 | 43 | 27 | | | 6 | | | | 81 |
| 16 32 | | 34 14 | 25 43 | | 5 5 | 9 4 | 22 14 | | | 10 | | | | 17 2 |
| >32 | | 5 | 43 | | j j | 4 | 14 | | | | | | | |
| -32 | | 5 | | | | | | | | | | | | |

| | AM substance | Cefepime | Cefotaxim | | Cerotaxime + Ciavulanic acid | Cefoxitin | Ceftazidim | | Ceftazidime + Clavulanic acid | | Ertapenem | Imipenem | Meropenem | Temocillin |
|-----|--------------------------|---------------|---------------|----------------------|------------------------------|----------------------------|-----------------|---------------|-------------------------------|-------------------|---------------|---------------|---------------|---------------|
| | Cefotaxime synergy test | Not Available | Not Available | Positive/Pres ent | Negative/Ab ent | ^S Not Available | Not Available | | Not Available | | Not Available | Not Available | Not Available | Not Available |
| | Ceftazidime synergy test | Not Available | Not Available | Not Available | Not Availabl | e Not Available | Not Available I | Not Available | Positive/Pres N ent | egative/Ab ent | Not Available | Not Available | Not Available | Not Available |
| | ECOFF | 0.125 | 0.25 | 0.25 | 0.25 | 8 | 0.5 | 0.5 | 0.5 | 0.5 | 0.064 | 0.5 | 0.125 | 32 |
| | Lowest limit | 0.064 | 0.25 | 0.064 | 0.064 | 0.5 | 0.25 | 0.12 | 0.12 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| | Highest limit | 32 | 64 | 64 | 64 | 64 | 128 | 128 | 128 | 128 | 2 | 16 | 16 | 64 |
| | N of tested isolates | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 |
| MIC | N of resistant isolates | 148 | 151 | 22 | 22 | 32 | 145 | 22 | 22 | 22 | 10 | 0 | 0 | 0 |
| 64 | | | 48 | | | 9 | 2 | | | 2 | | | | |
| >64 | | | 28 | | | 10 | | | | | | | | |
| 128 | | | | | | | 2 | | | | | | | |

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Pigs - fattening pigs

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method:

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.25 | 0.5 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 64 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 |
| MIC | N of resistant isolates | 151 | 44 | 151 | 144 | 70 | 90 | 15 | 11 | 0 | 68 | 119 | 131 | 1 | 106 |
| <=0.015 | | | | | | | 35 | | | | | | | | |
| <=0.03 | | | | | | | | | | 146 | | | | | |
| 0.03 | | | | | | | 24 | | | | | | | | |
| 0.064 | | | | | | | 2 | | | 5 | | | | | |
| 0.12 | | | | | | | 4 | | | | | | | | |
| <=0.25 | | | | | | | | | | | | | | 100 | 23 |
| 0.25 | | | | | | | 29 | | | | | | | | |
| <=0.5 | | | | | 7 | | | | 96 | | | | | | |
| 0.5 | | | | | | | 8 | | | | | | | 34 | 19 |
| <=1 | | | | | | | | 134 | | | | | | | |
| 1 | | | | 2 | 27 | | 4 | | 42 | | | | | 16 | 3 |
| <=2 | | | 4 | | | | | | | | | | 16 | <u> </u> | |
| 2 | | | | 2 | 38 | | | 2 | 2 | | | | | 1 | |
| <=4 | | | | | | | | | | | 68 | | | | |
| 4 | | | 39 | 1 112 | 19 | | | 15 | 2 | | | | 3 | | |
| >4 | | | | 146 | | 05 | | | | | | | | | |
| <=8 | | | 59 | | 26 | 65 | 7 | | 1 | | 10 | 26 | 1 | | 1 |
| 8 >8 | | | 59 | | 34 | | 38 | | 1 | | 10 | | 1 | | 1 |
| 16 | | 1 | E | | 34 | 16 | 38 | | 1 | | E | 4 | | | 1 |
| 32 | | ı | 5 2 | | | 9 | | | | | 5 4 | <u>4</u> 1 | 1 | | 1 |
| >32 | | | | | | <u> </u> | | | 7 | | 4 | 1 | ' | | 104 |
| 64 | | | 4 | | | 14 | | | ı | | 2 | 1 | 23 | | 104 |
| | | | | | | 17 | | | | | | 1 | 23 | | |

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|-------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.25 | 0.5 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 64 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 | 151 |
| MIC | N of resistant isolates | 151 | 44 | 151 | 144 | 70 | 90 | 15 | 11 | 0 | 68 | 119 | 131 | 1 | 106 |
| >64 | | 150 | 38 | | | | | | | | | | 107 | | |
| 128 | | | | | | 18 | | | | | 8 | | | | |
| >128 | | | | | | 29 | | | | | 54 | | | | |
| >1024 | | | | | | | | | | | | 119 | | | |

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Pigs - fattening pigs

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method:

Country of Origin: Spain

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|-----------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.25 | 0.5 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 64 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 |
| MIC | N of resistant isolates | 6 | 0 | 0 | 0 | 3 | 3 | 0 | 0 | 0 | 0 | 4 | 5 | 0 | 3 |
| <=0.015 | | | | | | | 2 | | | | | | | | |
| <=0.03 | | | | | | | | | | 6 | | | | | |
| 0.064 | | | | | | | 1 | | | | | | | | |
| <=0.25 | | | | 6 | | | | | | | | | | 4 | 2 |
| 0.25 | | | | | | | 2 | | | | | | | | |
| <=0.5 | | | | | 6 | | | | 3 | | | | | | |
| 0.5 | | | | | | | 1 | | | | | | | 2 | 1 |
| <=1 | | | | | | | | 6 | | | | | | | |
| 1 | | | | | | | | | 1 | | | | | | |
| <=2 | | | | | | | | | | | | | 1 | | |
| 2 | | | | | | | | | 2 | | | | | | |
| <=4 | | | | | | | | | | | 6 | | | | |
| 4 <=8 | | | 2 | | | 3 | | | | | | 1 | | | |
| 8 | | | 4 | | | <u> </u> | | | | | | | | | |
| 16 | | | | | | | | | | | | 1 | | | |
| 32 | | | | | | 1 | | | | | | | | | |
| >32 | | | | | | | | | | | | | | | 3 |
| | | | | | | 1 | | | | | | | 1 | | |
| 64 >64 | | 6 | | | | | | | | | | | 4 | | |
| >128 | | • | | | | 1 | | | | | | | | | |
| >1024 | | | | | | | | | | | | 4 | | | |
| | | | | | | | | | | | | | | | |

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Pigs - fattening pigs

Sampling Stage: Slaughterhouse Sampling Type: animal sample - caecum Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: ESBL MON pnl2

Analytical Method:

Country of Origin: Spain

| | AM substance | Cefepime | Cefotaxim | Cefotaxime + Clavulanic acid | Cefoxitin | Ceftazidim | Ceftazidime + Clavulanic acid | Ertapenem | lmipenem | Meropenem | Temocillin |
|---------|----------------------------|---------------|---------------|------------------------------|---------------|---------------|-------------------------------|---------------|---------------|---------------|---------------|
| | Cefotaxime synergy test | Not Available | Not Available | Positive/Pres ent | Not Available | Not Available | Not Available | Not Available | Not Available | Not Available | Not Available |
| | Ceftazidime synergy test | Not Available | Not Available | Not Available | Not Available | Not Available | Positive/Pres ent | Not Available | Not Available | Not Available | Not Available |
| | ECOFF | 0.125 | 0.25 | 0.25 | 8 | 0.5 | 0.5 | 0.064 | 0.5 | 0.125 | 32 |
| | Lowest limit | 0.064 | 0.25 | 0.064 | 0.5 | 0.25 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| | Highest limit | 32 | 64 | 64 | 64 | 128 | 128 | 2 | 16 | 16 | 64 |
| | N of tested isolates | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 |
| MIC | N of resistant isolates | 11 | 11 | 0 | 5 | 11 | 0 | 0 | 0 | 0 | 0 |
| <=0.015 | | | | | | | | 6 | | | |
| <=0.03 | | | | | | | | | | 11 | |
| 0.03 | | | | | | | | 3 | | | |
| <=0.064 | | | | 6 | | | | | | | |
| 0.064 | | | | | | | | 2 | | | |
| <=0.12 | | | | | | | 4 | | 2 | | |
| 0.12 | | | | 5 | | | 6 | | 8 | | |
| 0.25 | | 2 | | | | | 1 | | 1 | | |
| 1 | | | | | | 1 | <u>'</u> | | <u>'</u> | | |
| 2 | | 1 | | | | 3 | | | | | |
| 4 | | · | 1 | | 3 | 2 | | | | | 4 |
| 8 | | 4 | 1 | | 3 | 1 | | | | | 5 |
| 16 | | 1 | | | 5 | 3 | | | | | 2 |
| 32 | | 2 | 1 | | | 1 | | | | | |
| >32 | | 1 | | | | | | | | | |
| | | | | | | | | | | | |

| | AM substance | Cefepime | Cefotaxim | Cefotaxime + Clavulanic acid | Cefoxitin | Ceftazidim | Ceftazidime + Clavulanic acid | Ertapenem | Imipenem | Meropenem | Temocillin |
|-----|--------------------------|---------------|---------------|------------------------------|---------------|---------------|-------------------------------|---------------|---------------|---------------|---------------|
| | Cefotaxime synergy test | Not Available | Not Available | Positive/Pres ent | Not Available | Not Available | Not Available | Not Available | Not Available | Not Available | Not Available |
| | Ceftazidime synergy test | Not Available | Not Available | Not Available | Not Available | Not Available | Positive/Pres ent | Not Available | Not Available | Not Available | Not Available |
| | ECOFF | 0.125 | 0.25 | 0.25 | 8 | 0.5 | 0.5 | 0.064 | 0.5 | 0.125 | 32 |
| | Lowest limit | 0.064 | 0.25 | 0.064 | 0.5 | 0.25 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| | Highest limit | 32 | 64 | 64 | 64 | 128 | 128 | 2 | 16 | 16 | 64 |
| | N of tested isolates | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 |
| міс | N of resistant isolates | 11 | 11 | 0 | 5 | 11 | 0 | 0 | 0 | 0 | 0 |
| 64 | | | 6 | | | | | | | | |
| >64 | | | 2 | | | | | | | | |

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Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Pigs - fattening pigs

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method:

Country of Origin: Spain

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sufamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|-----------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.25 | 0.5 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 64 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 |
| MIC | N of resistant isolates | 11 | 4 | 11 | 11 | 7 | 6 | 0 | 1 | 0 | 6 | 10 | 9 | 0 | 9 |
| <=0.015 | | | | | | | 2 | | | | | | | | |
| <=0.03 | | | | | | | | | | 11 | | | | | |
| 0.03 | | | | | | | 3 | | | | | | | | |
| <=0.25 | | | | | | | | | | | | | | 8 | |
| 0.25 | | | | | | | 1 | | | | | | | | |
| <=0.5 | | | | | | | | | 5 | | | | | | |
| 0.5 | | | | | | | | | | | | | | 3 | 1 |
| <=1 | | | | | 1 | | | 11 | 4 | | | | | | 1 |
| <=2 | | | | | 1 | | | | 4 | | | | 1 | | ' |
| 2 | | | | | 1 | | | | 1 | | | | | | |
| <=4 | | | | | ı | | | | <u>'</u> | | 5 | | | | |
| 4 | | | 1 | 2 | 3 | | | | | | | | 1 | | |
| >4 | | | | 9 | - | | | | | | | | | | |
| <=8 | | | | - | | 4 | | | | | | 1 | | | |
| 8 | | | 5 | | 1 | | | | | | | | | | |
| >8 | | | | | 5 | | 5 | | | | | | | | |
| 16 | | | 1 | | | | | | | | | | | | |
| 32 | | | 1 | | | 1 | | | | | | | | | |
| >32 | | | | | | | | | 1 | | | | | | 9 |
| 64 | | | 1 | | | 1 | | | | | | | 2 | | |
| >64 | | 11 | 2 | | | | | | | | | | 7 | | |
| 128 | | | | | | | | | | | 1 | | | | |

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|-------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.25 | 0.5 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 64 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 | 11 |
| MIC | N of resistant isolates | 11 | 4 | 11 | 11 | 7 | 6 | 0 | 1 | 0 | 6 | 10 | 9 | 0 | 9 |
| >128 | | | | | | 5 | | | | | 5 | | | | |
| >1024 | | • | | • | | | | | | | | 10 | | • | |

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Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from pig - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method:

Country of Origin: Portugal

| | AM substance | Cefepime | Cefotaxim | | Cefotaxime + Clavulanic acid | Cefoxitin | Ceftazidim | | Ceftazidime + Clavulanic acid | | Ertapenem | Ітірепет | Meropenem | Temocillin |
|---------|-----------------------------|---------------|-----------------|---------------------|------------------------------|----------------------------|-----------------|-------------|-------------------------------|--------------------|----------------------------|---------------|---------------|---------------|
| | Cefotaxime synergy test | Not Available | Not Available | Positive/Pre ent | s Negative/Ab ent | ^S Not Available | Not Available | | Not Available | | | | Not Available | |
| | Ceftazidime synergy test | Not Available | Not Available I | Not Availabl | e Not Availabl | e Not Available | Not Available N | ot Availabl | e Positive/Pres ent | Negative/Ab ent | ^S Not Available | Not Available | Not Available | Not Available |
| | ECOFF | 0.125 | 0.25 | 0.25 | 0.25 | 8 | 0.5 | 0.5 | 0.5 | 0.5 | 0.064 | 0.5 | 0.125 | 32 |
| | Lowest limit | 0.064 | 0.25 | 0.064 | 0.064 | 0.5 | 0.25 | 0.12 | 0.12 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| | Highest limit | 32 | 64 | 64 | 64 | 64 | 128 | 128 | 128 | 128 | 2 | 16 | 16 | 64 |
| | N of tested isolates | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 |
| MIC | N of resistant isolates | 22 | 23 | 6 | 6 | 7 | 22 | 5 | 5 | 5 | 1 | 0 | 0 | 0 |
| <=0.015 | | | | | | | | | | | 15 | | | |
| <=0.03 | | | | | | | | | | | | | 22 | |
| 0.03 | | | | | | | | | | | 5 | | | |
| <=0.064 | | | | 17 | | | | | | | | | | |
| 0.064 | | | | | | | | | | | 2 | | 1 | |
| <=0.12 | | | | | | | | 1 | 8 | | | 10 | | |
| 0.12 | | 1 | | | | | | | | | 1 | | | |
| 0.25 | | 2 | | | | | | | 7 | 2 | | 11 | | |
| 0.5 | | 2 | | | | | 1 | | | | | 2 | | |
| 1 | | | | | 1 | 1 | 6 | | | | | | | |
| 2 | | 5 | 1 | | | | 1 | | | | | | | 3 |
| 4 | | 2 | 1 | 1 | 1 | 8 | 5 | | | 1 | | | | 3 |
| 8 | | 6 | 1 | | 2 | 7 | 4 | | | 1 | | | | 15 |
| 16 | | 3 | 4 | | 1 | 1 | 4 | | | 3 | | | | 1 |
| 32 | | 2 | 6 | | | 11 | 2 | | | | | | | 1 |
| 64 | | | 7 | | | 3 | | | | | | | | |

| | AM substance | Cefepime | Cefotaxim | | Cefotaxime + Clavulanic acid | Cefoxitin | Ceftazidim | | Ceftazidime + Clavulanic acid | | Ertapenem | Imipenem | Meropenem | Temocillin |
|-----|--------------------------|---------------|---------------|---------------|------------------------------|----------------------------|-----------------|---------------|-------------------------------|--------------------|----------------------------|---------------|---------------|---------------|
| | Cefotaxime synergy test | Not Available | Not Available | Positive/Pres | Negative/Abs | ^S Not Available | Not Available | | Not Available | | | | Not Available | |
| | Ceftazidime synergy test | Not Available | Not Available | Not Available | Not Available | e Not Available | Not Available N | lot Available | Positive/Pres N | legative/Ab ent | ^S Not Available | Not Available | Not Available | Not Available |
| | ECOFF | 0.125 | 0.25 | 0.25 | 0.25 | 8 | 0.5 | 0.5 | 0.5 | 0.5 | 0.064 | 0.5 | 0.125 | 32 |
| | Lowest limit | 0.064 | 0.25 | 0.064 | 0.064 | 0.5 | 0.25 | 0.12 | 0.12 | 0.12 | 0.015 | 0.12 | 0.03 | 0.5 |
| | Highest limit | 32 | 64 | 64 | 64 | 64 | 128 | 128 | 128 | 128 | 2 | 16 | 16 | 64 |
| | N of tested isolates | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 |
| МІС | N of resistant isolates | 22 | 23 | 6 | 6 | 7 | 22 | 5 | 5 | 5 | 1 | 0 | 0 | 0 |
| >64 | | - | 3 | | | 2 | - | | - | | - | | | |

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Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from pig - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method:

Country of Origin: Portugal

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|---------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.25 | 0.5 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 64 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 |
| MIC | N of resistant isolates | 23 | 6 | 23 | 20 | 5 | 15 | 1 | 2 | 0 | 13 | 17 | 19 | 0 | 13 |
| <=0.015 | | | | | | | 4 | | | | | | | | |
| <=0.03 | | | | | | | | | | 23 | | | | | |
| 0.03 | | | | | | | 4 | | | | | | | | |
| <=0.25 | | | | | | | | | | | | | | 20 | 4 |
| 0.25 | | | | | | | 3 | | | | | | | | |
| <=0.5 | | | | | 3 | | | | 16 | | | | | | |
| 0.5 | | | | | | | 4 | | | | | | | 2 | 6 |
| <=1 | | | | | | | | 21 | | | | | | | |
| 1 | | | | | 3 | | 1 | | 4 | | | | | 1 | |
| <=2 | | | 1 | | | | | | | | | | 2 | | |
| 2 | | | | 1 | 4 | | | 1 | 1 | | | | | | |
| <=4 | | | | | | | | | | | 8 | | | | |
| 4 | | | 10 | 1 | 4 | | | 1 | | | | | 2 | | 1 |
| >4 | | | | 21 | | | | | | | | | | | |
| <=8 | | | | | | 15 | | | | | | 2 | | | |
| 8 | | | 5 | | 4 | | 1 | | | | 1 | | | | |
| >8 | | | | | 5 | | 6 | | | | | | | | |
| 16 | | | 1 | | | 3 | | | | | 1 | 2 | | | |
| 32 | | | 1 | | | 2 | | | 2 | | 1 | 2 | 2 | | |
| >32 | | | | | | | | | | | | | | | 12 |
| 64 | | | 2 | | | 2 | | | | | | | 3 | | |
| >64 | | 23 | 3 | | | | | | | | | | 14 | | |
| 128 | | | | | | 1 | | | | | 2 | | | | |

| | AM substance | Ampicillin | Azithromycin | Cefotaxim | Ceftazidim | Chloramphenicol | Ciprofloxacin | Colistin | Gentamicin | Meropenem | Nalidixic acid | Sulfamethoxazole | Tetracycline | Tigecycline | Trimethoprim |
|-------|-------------------------|------------|--------------|-----------|------------|-----------------|---------------|----------|------------|-----------|----------------|------------------|--------------|-------------|--------------|
| | ECOFF | 8 | 16 | 0.25 | 0.5 | 16 | 0.064 | 2 | 2 | 0.125 | 16 | 64 | 8 | 1 | 2 |
| | Lowest limit | 1 | 2 | 0.25 | 0.5 | 8 | 0.015 | 1 | 0.5 | 0.03 | 4 | 8 | 2 | 0.25 | 0.25 |
| | Highest limit | 64 | 64 | 4 | 8 | 128 | 8 | 16 | 32 | 16 | 128 | 1024 | 64 | 8 | 32 |
| | N of tested isolates | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 | 23 |
| МІС | N of resistant isolates | 23 | 6 | 23 | 20 | 5 | 15 | 1 | 2 | 0 | 13 | 17 | 19 | 0 | 13 |
| >128 | | | | | | | | | | | 10 | | | | |
| >1024 | | | | | | | | · | | | | 17 | | | |

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OTHER ANTIMICROBIAL RESISTANCE TABLES

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Specific monitoring of ESBL-/AmpC-/carbapenemase-producing bacteria and specific monitoring of carbapenemase-producing bacteria, in the absence of isolate detected

| Programme Code | Matrix Detailed | Zoonotic Agent Detailed | Sampling Strategy | Sampling Stage | Sampling Details | Sampling Context | Sampler | Sample Type | Sampling Unit Type | Sample Origin | Comment | Total Units Tested | Total Units Positive |
|-------------------|--|---|----------------------|--------------------|---------------------|---------------------|--------------------------|------------------------------|---------------------------|---------------|---------|--------------------------|----------------------------|
| CARBA MON | Cattle (bovine animals) - calves (under 1 year) | Escherichia coli, non- pathogenic, unspecified | Objective sampling | Slaughte rhouse | N_A | Monitorin g | Official samplin g | animal sample - caecum | slaughter animal batch | Portugal | N_A | 289 | 0 |
| | Meat from bovine animals - fresh | Escherichia coli, non-pathogenic, unspecified | Objective sampling | Retail | N_A | Monitorin g | Official samplin g | food sample - meat | single (food/feed) | Portugal | N_A | 220 | 0 |
| | Meat from pig - fresh | Escherichia coli, non- pathogenic, unspecified | Objective sampling | Retail | N_A | Monitorin g | Official samplin g | food sample - meat | single (food/feed) | Portugal | N_A | 220 | 0 |
| | Pigs - fattening pigs | Escherichia coli, non- pathogenic, unspecified | Objective sampling | Slaughte rhouse | N_A | Monitorin g | Official samplin g | animal sample - caecum | slaughter animal batch | Portugal | N_A | 254 | 0 |



Latest Transmission set

Last submitted

| Table Name | dataset transmission date |
|--------------------------|---------------------------|
| Antimicrobial Resistance | 23-Jul-2018 |
| Esbl | 23-Jul-2018 |
| Animal Population | 23-Jul-2018 |
| Disease Status | 23-Jul-2018 |
| Food Borne Outbreaks | 23-Jul-2018 |
| Prevalence | 23-Jul-2018 |

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| 15. flo c | General evaluation: Salmonella in animal - Gallus gallus (fowl) - breeding cks, unspecified |
| 16. Sal | Description of Monitoring/Surveillance/Control programmes system: Imonella in animal - Gallus gallus (fowl) - breeding flocks, unspecified |
| 17. pro | General evaluation: Salmonella in Turkeys - breeding flocks and meat |

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1. Institutions and Laboratories involved in zoonoses monitoring and reporting

Instituto Nacional de Investigação Agrária e Veterinária, I. P. (INIAV)

Laboratório Regional Veterinário (LRV)

Instituto Nacional de Saúde Doutor Ricardo Jorge, I. P. (INSA)

Instituto Português do Mar e da Atmosfera, I.P. (IPMA)

Autoridade de Segurança Alimentar e Económica (ASAE)

Direção Geral de Alimentação e Veterinária (DGAV)

Instituto de Financiamento da Agricultura e Pescas, I.P. (IFAP)

INIAV: laboratory supporting several official control and surveillance programs and plans, with analytical research on zoonotic agents in animal, food and feed samples. Also supports national AMR monitoring plan with analytical performance.

LRV: Autonomous Region of Azores' laboratory supporting several official control and surveillance programs and plans, with analytical research on zoonotic agents in animal, food and feed samples. Also supports national AMR monitoring plan with analytical performance obtaining the isolates that are tested for antimicrobial resistance at INIAV laboratory. INSA: laboratory supporting national health system with investigation and analytical performance on FBO scope and analytical performance on food samples.

IPMA: laboratory supporting official control plans with analytical research on zoonotic agents in food (fishery products and LBM).

ASAE: laboratory performing official analytical control on food, mainly at retail stage.

DGAV: national competent authority with several official control and surveillance programs and plans applicable to animals, food and feed.

IFAP: Institute involved in animal and farms registration.

2. Animal population

1. Sources of information and the date(s) (months, years) the information relates to

Animal farms and bovine animals are legally obliged to be registered. Also the animal slaughter quantities are obliged to be reported.

Data are compiled in the IFAP's database and numbers on animals and farms are given based on the consultation of the database.

The quantities related with slaughtered animals are compiled in another database by meat inspectors. Numbers on slaughtered animals are given based on the consultation of the database.

- 2. Definitions used for different types of animals, herds, flocks and holdings as well as the production types covered
- 3. National changes of the numbers of susceptible population and trends
- 4. Geographical distribution and size distribution of the herds, flocks and holdings
- 5. Additional information

3. General evaluation: B. abortus in animal - Cattle (bovine animals)

1. History of the disease and/or infection in the country

Status as officially free of bovine brucellosis: In the Açores, there are 6 islands (Santa Maria, Faial, Graciosa, Pico, Flores and Corvo) that are Officially Free of Bovine Brucellosis, accordingly with Commission Decisions 2002/588/CE of 11th July 2002 and 2009/600/CE of 5th August. At mainland, the Algarve region was recognised as Officially Free of Bovine Brucellosis accordingly with Commission Decision 2012/204/UE of 19th April.

For more information, in relation to bovine brucellosis, refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff_animal_vet-progs_2017-8_dec-2016-2444-ec bovine-brucellosis prt.pdf

- 2. Evaluation of status, trends and relevance as a source for humans
- 3. Any recent specific action in the Member State or suggested for the European Union
- 4. Additional information

4. Description of Monitoring/Surveillance/Control programmes system: B. abortus in animal - Cattle (bovine animals)

1. Monitoring/Surveillance/Control programmes system

Sampling strategy: The herds are classified and sampled accordingly with Council Directive 64/432/EEC and Decreto-Lei No 244/2000 (Sep. 27th).

Frequency of the sampling: The herds are sampled accordingly with Council Directive 64/432/EEC and Decreto-Lei No 244/2000 (Sep. 27th).

Type of specimen taken: Blood, milk, organs, vaginal mucus, semen, aborted foetus and placenta.

Diagnostic/analytical methods used:

- Serology (live animals): Rose Bengal Test (RBT); Complement Fixation Test (CFT)
- Milk (live animals): ELISA test
- Organs (dead animals): bacteriology (isolation of the agent with differentiation of vaccine and field strains).

For more information, in relation to bovine brucellosis, refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff_animal_vet-progs_2017-8_dec-2016-2444-ec bovine-brucellosis prt.pdf

2. Measures in place

An Eradication Programme for cattle is carried out and supervised by DGAV.

Vaccination is forbidden, but if an exceptional sanitary situation occurs, vaccination can be allowed with specific protocols between the Veterinary Authority and the owner(s) of the cattle.

Other preventive measures than vaccination in place: Pre-movement tests are mandatory accordingly with Council Directive 64/432/EEC.

Measures in case of the positive findings or single cases:

- Suspected Herd:
- Herd under official surveillance;
- Epidemiological survey;
- Animal movements are forbidden from and to the herd:
- Isolation of suspected animals in the herd;
- Sample collection for laboratory diagnosis.

- Positive Herd:
- Herd under official restrictions:
- Compulsory slaughter of all positive animals, under official supervision with sample collection for laboratory diagnosis;
- Animal movements are forbidden from and to the herd;
- Serological control of all remaining animals.
 - Infected Herd:
- All measures mentioned for positive herds;
- Disinfection of all premises, equipment and materials;
- Thermic treatment of the milk.

For more information, in relation to bovine brucellosis, refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff_animal_vet-progs_2017-8_dec-2016-2444-ec_bovine-brucellosis_prt.pdf

3. Notification system in place to the national competent authority

Yes

- 4. Results of investigations and national evaluation of the situation, the trends and sources of infection
- 5. Additional information
 - 5. General evaluation: *Mycobacterium tuberculosis* complex (MTC) in animal Cattle (bovine animals)
- 1. History of the disease and/or infection in the country

Status as officially free of bovine tuberculosis: at mainland, the Algarve region was recognised as Officially Free of Bovine Tuberculosis accordingly with Commission Decision 2012/204/UE of 19th April.

For more information, in relation to bovine tuberculosis, refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff animal vet-progs 2017-8 dec-2016-2444-ec_bovine-tuberculosis_prt.pdf

- 2. Evaluation of status, trends and relevance as a source for humans
- 3. Any recent specific action in the Member State or suggested for the European Union
- 4. Additional information

6. Description of Monitoring/Surveillance/Control programmes system: Mycobacterium tuberculosis complex (MTC) in animal - Cattle (bovine animals)

1. Monitoring/Surveillance/Control programmes system

Sampling strategy: The herds are classified and sampled accordingly with Council Directive 64/432/EEC and National Dec. Lei No 272/2000, November 8th and National Dec. Lei No 79/2011, June 20th.

Frequency of the sampling: The herds are sampled accordingly with Council Directive 64/432/EEC and National Dec. Lei No 272/2000, November 8th and National Dec. Lei No 79/2011, June 20th.

Type of specimen taken: Blood and organs.

Diagnostic/analytical methods used:

- Animal: Intra-dermal comparative test;
- Blood: Gama-interferon:
- Organs: histopathology and bacteriology.

For more information, in relation to bovine tuberculosis, refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff_animal_vet-progs_2017-8_dec-2016-2444-ec_bovine-tuberculosis_prt.pdf

2. Measures in place

An Eradication Programme for Bovine Tuberculosis is carried out and supervised by DGAV. Vaccination is forbidden.

Other preventive measures than vaccination in place: Pre-movement tests are mandatory accordingly with Council Directive 64/432/EEC.

Measures in case of the positive findings or single cases:

- Herd under official restriction;
- Isolation of suspected or infected animals in the herd;
- Positive animals compulsory slaughtered, under official supervision, with sample collection for laboratory diagnosis;
- Animal movements are forbidden from and to the herd;
- Disinfection of all premises, equipment and materials;
- Testing of all remaining animals;
- Thermic treatment of the milk;
- Epidemiological survey.

For more information, in relation to bovine tuberculosis, refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff_animal_vet-progs_2017-8_dec-2016-2444-ec_bovine-tuberculosis_prt.pdf

3. Notification system in place to the national competent authority

Yes

4. Results of investigations and national evaluation of the situation, the trends and sources of infection

5. Additional information

7. General evaluation: Lyssavirus (rabies)

1. History of the disease and/or infection in the country

Portugal is free from Rabies since 1961. In August 1984, the national authorities detected a case of rabies in a 2 months old puppy that came from Maputo (Mozambique) and entered in Portugal on the 10th August 1984. The animal was put in quarantine and euthanized. The disease was confirmed by immunofluorescence on the 31st August. The veterinary authorities maintained the implemented sanitary and prophylactic measures and, since then, no further cases were detected and Portugal maintained its free status.

- 2. Evaluation of status, trends and relevance as a source for humans
- 3. Any recent specific action in the Member State or suggested for the European Union
- 4. Additional information

8. Description of Monitoring/Surveillance/Control programmes system: Lyssavirus (rabies)

1. Monitoring/Surveillance/Control programmes system

Rage is a national notifiable disease since 1953.

Surveillance is based on the investigation of any clinical suspicion and aggression episodes: any dog or cat that bites a human or another animal is considered under suspicion and, therefore, is kept under veterinary surveillance in order to discard any case of rabies.

Laboratorial confirmation: positive result at the direct immunofluorescence test.

2. Measures in place

The control program is defined in the national law (Decreto-Lei No 314/2003, of December 17th) and consists in Vaccination and Surveillance Measures for epidemiological survey with definition of specific rules for owners, for commercial purposes, for exhibits and for the entrance of animals in the country.

The measures are defined in the national and EU legislation.

National legislation (Decreto-Lei No 314/2003, of December the 17th and Portaria No 264/2013, of August the 16th established the obligation of vaccination against rabies in all dogs older than 3 month. Vaccination may be performed either by Municipality Veterinarians in the official campaign or by small animal practitioners in their private clinics. Surveillance is based on the investigation of any clinical suspicion and aggression episodes: any dog or cat that bites a human or another animal is considered under suspicion and, therefore, is kept under veterinary surveillance in order to discard any case of rabies.

In Portugal the annual rabies vaccination of dogs is compulsory since 1925. Vaccination in cats is voluntary.

3. Notification system in place to the national competent authority

Yes

4. Results of investigations and national evaluation of the situation, the trends and sources of infection

5. Additional information

At the rabies vaccination campaign, whenever the animals present show signs of leishmaniosis, dermatophytosis or mange, the municipality veterinarian notifies the respective owner to perform diagnostic tests in the animal and to treat the zoonosis in case of a positive result.

9. General evaluation: B. melitensis in animal - Sheep

1. History of the disease and/or infection in the country

Status as officially free of ovine brucellosis: The Região Autónoma dos Açores is officially free of sheep and goat brucellosis, accordingly with Commission Decision 2003/44/CE of the 17th January 2003.

For more information, in relation to sheep and goat brucellosis, refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff animal vet-progs 2017-8_dec-2016-2444-ec_goat-brucellosis_prt.pdf

- 2. Evaluation of status, trends and relevance as a source for humans
- 3. Any recent specific action in the Member State or suggested for the European Union
- 4. Additional information

10. Description of Monitoring/Surveillance/Control programmes system: *B. melitensis* in animal - Sheep

1. Monitoring/Surveillance/Control programmes system

Sampling strategy: The herds are classified and sampled accordingly with Council Directive 91/68/EEC of 28 January 1991 on animal health conditions governing intra-Community trade in sheep and goat animals and Decreto-Lei No 244/2000 (Sep. 27th).

Frequency of the sampling: The herds are sampled accordingly with Council Directive 91/68/EEC of 28 January 1991 on animal health conditions governing intra-Community trade in sheep and goat animals and Decreto-Lei No 244/2000 (Sep. 27th).

Type of specimen taken: Blood, organs, vaginal mucus, semen, aborted foetus and placenta. Diagnostic/analytical methods used:

- Serology (live animals): Rose Bengal Test (RBT); Complement Fixation Test (CFT)
- Organs (dead animals): bacteriology (isolation of the agent with differentiation of vaccine and field strains).

For more information, in relation to sheep and goat brucellosis, refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff_animal_vet-progs_2017-8_dec-2016-2444-ec_goat-brucellosis_prt.pdf

2. Measures in place

Vaccination of young animals with REV1 is performed in some of the mainland regions: Norte, Centro, Lisboa e Vale do Tejo and Algarve.

Other preventive measures than vaccination in place: Pre-movement tests are mandatory for animals intended for the replacement in depopulated herds.

An Eradication Programme for sheep and goat is carried out and supervised by DGAV.

Measures in case of the positive findings or single cases:

• Suspected Herd:

- Herd under official surveillance;
- Epidemiological survey:
- Animal movements are forbidden from and to the herd;
- Isolation of suspected animals in the herd;
- Sample collection for laboratory diagnosis.
 - Positive Herd:
- Herd under official restrictions:
- Epidemiological survey;
- Compulsory slaughter of all positive animals, under official supervision with sample collection for laboratory diagnosis;
- Animal movements are forbidden from and to the herd;
- Serological control of all remaining animals.
 - Infected Herd:
- All measures mentioned for positive herds;
- Disinfection of all premises, equipment and materials;
- Thermic treatment of the milk.

For more information, in relation to sheep and goat brucellosis, refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff animal vet-progs 2017-8_dec-2016-2444-ec_goat-brucellosis_prt.pdf

3. Notification system in place to the national competent authority

Yes

- 4. Results of investigations and national evaluation of the situation, the trends and sources of infection
- 5. Additional information

11. General evaluation: B. melitensis in animal - Goats

1. History of the disease and/or infection in the country

Status as officially free of caprine brucellosis: The Região Autónoma dos Açores is officially free of sheep and goat brucellosis, accordingly with Commission Decision 2003/44/CE of the 17th January 2003.

For more information, in relation to sheep and goat brucellosis, refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff_animal_vet-progs_2017-8 dec-2016-2444-ec goat-brucellosis prt.pdf

- 2. Evaluation of status, trends and relevance as a source for humans
- 3. Any recent specific action in the Member State or suggested for the European Union
- 4. Additional information

12. Description of Monitoring/Surveillance/Control programmes system: B. melitensis in animal - Goats

1. Monitoring/Surveillance/Control programmes system

Sampling strategy: The herds are classified and sampled accordingly with Council Directive 91/68/EEC of 28 January 1991 on animal health conditions governing intra-Community trade in sheep and goat animals and Decreto-Lei No 244/2000 (Sep. 27th).

Frequency of the sampling: The herds are sampled accordingly with Council Directive 91/68/EEC of 28 January 1991 on animal health conditions governing intra-Community trade in sheep and goat animals and Decreto-Lei No 244/2000 (Sep. 27th).

Type of specimen taken: Blood, organs, vaginal mucus, semen, aborted foetus and placenta. Diagnostic/analytical methods used:

- Serology (live animals): Rose Bengal Test (RBT); Complement Fixation Test (CFT)
- Organs (dead animals): bacteriology (isolation of the agent with differentiation of vaccine and field strains).

For more information, in relation to sheep and goat brucellosis, refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff animal vet-progs 2017-8_dec-2016-2444-ec_goat-brucellosis_prt.pdf

2. Measures in place

Vaccination of young animals with REV1 is performed in some of the mainland regions: Norte, Centro, Lisboa e Vale do Tejo and Algarve.

Other preventive measures than vaccination in place: Pre-movement tests are mandatory for animals intended for the replacement in depopulated herds.

An Eradication Programme for sheep and goat is carried out and supervised by DGAV.

Measures in case of the positive findings or single cases:

- Suspected Herd:
- Herd under official surveillance;
- Epidemiological survey;
- Animal movements are forbidden from and to the herd;
- Isolation of suspected animals in the herd;
- Sample collection for laboratory diagnosis.
 - Positive Herd:
- Herd under official restrictions;
- Epidemiological survey;
- Compulsory slaughter of all positive animals, under official supervision with sample collection for laboratory diagnosis;
- Animal movements are forbidden from and to the herd;
- Serological control of all remaining animals.
 - Infected Herd:
- All measures mentioned for positive herds;
- Disinfection of all premises, equipment and materials;
- Thermic treatment of the milk.

For more information, in relation to sheep and goat brucellosis, refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff_animal_vet-progs_2017-8 dec-2016-2444-ec goat-brucellosis prt.pdf

3. Notification system in place to the national competent authority

Yes

4. Results of investigations and national evaluation of the situation, the trends and sources of infection

13. General evaluation: Salmonella in animal - Gallus gallus (fowl) - broilers

1. History of the disease and/or infection in the country

For this information, in relation to broilers – *Gallus gallus* (fowl), refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff_animal_vet-progs_2018-9_salmonella_broiler_gal_prt.pdf

- 2. Evaluation of status, trends and relevance as a source for humans
- 3. Any recent specific action in the Member State or suggested for the European Union
- 4. Additional information

14. Description of Monitoring/Surveillance/Control programmes system: Salmonella in animal - Gallus gallus (fowl) - broilers

1. Monitoring/Surveillance/Control programmes system

Sampling strategy (broiler flocks): Sampling is accomplished by the food business operator and by the competent authority. The sampling is done at the holding. Sampling on the initiative of the food business operator shall take place within three weeks before the birds are moved to the slaughterhouse. Sampling by the competent authority includes each year at least one flock of broilers on 10% of the holdings with more than 5 000 birds. It is done on a risk basis approach and every time that the competent authority considers it necessary.

Frequency of the sampling (broiler flocks): 3 weeks prior to slaughter, at farm.

Type of specimen taken (broiler flocks): Faeces (boot swabs).

Methods of sampling (description of sampling techniques): At least two pairs of boot swabs shall be taken. For free range flocks of broilers, samples shall only be collected in the area inside the house. All boot swabs will be pooled into one sample. In flocks with less than 100 broilers, when the access to the houses is not possible, the boot swabs may be replaced by hand drag swabs and rubbed over surfaces contaminated with fresh faeces, or if not feasible, by other sampling techniques for faeces fit for the intended purpose. It shall be ensured that all sections in a house are represented in the sampling in a proportionated way. Each pair should cover about 50% of the area of the house. On completion of sampling the boot swabs shall be carefully removed so as not to dislodge adherent material. Boot swabs may be inverted to retain material. They shall be placed in a bag or pot and labelled.

Case definition: A flock of broilers is considered positive where the presence of *Salmonella* Enteritidis (other than vaccine strains) and/or *Salmonella* Typhimurium or *Salmonella* typhimurium -Like is detected in the flock at any occasion.

Diagnostic/analytical methods used: Bacteriological method: ISO 6579:2002.

For more information, in relation to broilers – *Gallus gallus* (fowl), refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff_animal_vet-progs_2018-9_salmonella_broiler_gal_prt.pdf

2. Measures in place

The strategy in place is to reinforce surveillance, reinforce biosecurity measures, slaughter the positive flocks and restocking only when environmental samples are negative for *Salmonella*, with birds from flocks or herds that have undergone controls accordingly with the legislation

requirements. The strategy includes also a close cooperation with the associations of producers to implement different means to raise awareness of the producers. The Official Services have developed guidelines for the producer, as a tool in order to guide the implementation of the national programme.

Measures in case of the positive findings or single cases:

- When there is a positive case in a flock
- Salmonella spp detection;
- Notification of the food business operator;
- Flock under official control (restriction);
- Forcing to keep update records.
 - Whenever the results from serotyping are different from the serotypes relevant to the national programme, than:
- Additional biosecurity measures;
- Free practice;
- The official control measures are withdrawn.
 - When the result is serotype S. Enteritidis and/or S. Typhimurium than the flock will continue under official restriction:
- Flock surveillance (under official control).
- After the slaughter of the positive flock the holding and the environment must be cleaned and disinfected.
- The food business operator must collect environmental samples.
- The restocking of animals must take place from flocks or herds that have undergone controls (with negative results) accordingly with the legislation requirements.

For more information, in relation to broilers – *Gallus gallus* (fowl), refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff_animal_vet-progs_2018-9_salmonella_broiler_gal_prt.pdf

3. Notification system in place to the national competent authority

Yes

4. Results of investigations and national evaluation of the situation, the trends and sources of infection

5. Additional information

15. General evaluation: Salmonella in animal - Gallus gallus (fowl) - breeding flocks, unspecified

1. History of the disease and/or infection in the country

For this information, in relation to breeding flocks – *Gallus gallus* (fowl), refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff_animal_vet-progs_2018-9_salmonella_breeding_gal_prt.pdf

- 2. Evaluation of status, trends and relevance as a source for humans
- 3. Any recent specific action in the Member State or suggested for the European Union
- 4. Additional information

In relation to laying hens, refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff_animal_vet-progs_2018-9 salmonella laying gg prt.pdf

16. Description of Monitoring/Surveillance/Control programmes system: Salmonella in animal - Gallus gallus (fowl) - breeding flocks, unspecified

1. Monitoring/Surveillance/Control programmes system

Sampling strategy (breeding flocks - separate elite, grand parent and parent flocks when necessary): The sampling frame shall cover all adult breeding flocks of *Gallus gallus* comprising at least 250 birds. Sampling is accomplished by the food business operator and by the official authority. Sampling is done at the holding. At the initiative of the food business operator, samples will be taken at day old, 4 weeks old birds, 2 weeks before laying phase and during the laying period, every three weeks. At day-old sampling shall consist of internal linings of delivery boxes and dead chicks. At 4 weeks old and at two weeks before the laying phase, sampling shall consist of pooled faeces made up of separate samples of fresh faeces each weighing no less than 1 g taken at random from a number of sites in the building in which the birds are kept. During the laying phase sampling will consist of 5 boot swabs representative of all parts of the house. In cage breeding flocks, sampling consists of naturally mixed faeces from dropping belts, scrapers or deep pits. 2 samples of at least 150 g will be collected to be tested individually. At the initiative of the official services sampling is done 2 times during the laying phase.

Frequency of the sampling:

- Breeding flocks (separate elite, grand parent and parent flocks when necessary) Day-old chicks: Every flock is sampled.
- Breeding flocks (separate elite, grand parent and parent flocks when necessary) Rearing period: Day-old chicks, at the age of 4 weeks and 2 weeks before moving to the laying phase.
- Breeding flocks (separate elite, grand parent and parent flocks when necessary) -Production period: Every 3 weeks.

Type of specimen taken:

- Breeding flocks (separate elite, grand parent and parent flocks when necessary) Day-old chicks: Internal linings of delivery boxes and dead chicks.
- Breeding flocks (separate elite, grand parent and parent flocks when necessary) Rearing period: Faeces.
- Breeding flocks (separate elite, grand parent and parent flocks when necessary) -Production period: Faeces / boot swabs.

Type of specimen taken:

- Breeding flocks (separate elite, grand parent and parent flocks when necessary) Day-old chicks: Internal linings of delivery boxes and dead chicks.
- Breeding flocks (separate elite, grand parent and parent flocks when necessary) Rearing period: Faeces.
- Breeding flocks (separate elite, grand parent and parent flocks when necessary) Production period: Faeces / boot swabs.

Methods of sampling (description of sampling techniques):

- Breeding flocks (separate elite, grand parent and parent flocks when necessary) Day-old chicks: The sample shall consist of a minimum of one composite sample of visibly soiled hatcher basket liners. The food business operator must sample all dead birds at arrival.
- Breeding flocks (separate elite, grand parent and parent flocks when necessary) Rearing period: At 4 weeks old and 2 weeks before the laying phase the sampling will consist of faecal samples. Pooled faeces made up of separate samples of fresh faeces each weighing no less than 1 g taken at random from a number of sites in the building in which the birds are kept.
- Breeding flocks Production period: During the laying phase 5 pairs of boot swabs walking around to be done in a way which will sample representatively all parts of the sector. In cage breeding flocks, sampling consists of naturally mixed faeces from dropping belts, scrapers or deep pits. 2 samples of at least 150 g will be collected to be tested individually.

Case definition:

• Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks, Rearing period and Production period - At least one positive sample to S. Enteritidis, S. Typhimurium, S. Typhimurium- Like, S. Hadar, S. Virchow and/or S. Infantis.

Diagnostic/analytical methods used:

• Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks, Rearing period and Production period - Bacteriological method: ISO 6579:2002.

For more information, in relation to breeding flocks – *Gallus gallus* (fowl), refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff_animal_vet-progs_2018-9_salmonella_breeding_gal_prt.pdf

2. Measures in place

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Vaccination is voluntary. Compulsive vaccination against *Salmonella* Enteritidis is done in the restocking, after the slaughter of a positive flock.

The strategy in place is to reinforce surveillance, reinforce biosecurity measures, slaughter the positive flocks and restocking only when environmental samples are negative for *Salmonella*, with birds from flocks or herds that have undergone controls accordingly with the legislation requirements, with negative results. All the restocking birds must be vaccinated against *Salmonella*. There is also a focus on biosecurity measures in the holdings. The strategy includes also a close cooperation with the associations of producers to implement different means to raise awareness of the producers. The Official Services have developed guidelines for the producer, as a tool in order to guide the implementation of the national programme.

Measures in case of the positive findings or single cases:

- In the case of positive results for *Salmonella* Enteritidis and/or *Salmonella* Typhimurium additional biosecurity measures are implemented, sanitary restriction of the flock and sanitary surveillance of the holding are imposed.
- Destination of birds: The slaughter of the flock will be carried out in an approved slaughterhouse and after the authorisation of Regional Veterinarian Services. Day-oldchicks must be killed and destroyed.
- Destination of eggs: Hatching eggs will be eliminated as animal by-products. Non-incubated eggs from positive flocks must be, at option of the FBO:
 - o eliminate as by-products or

- o forward to egg product units to be heat treated.
- After the slaughter of the positive flock the holding and the environment must be cleaned and disinfected.
- The food business operator must collect environmental samples.
- The restocking of animals must take place from flocks or herds that have undergone controls (with negative results) accordingly with the legislation requirements. All the restocking birds must be vaccinated against *Salmonella* Enteritidis.

For more information, in relation to breeding flocks – *Gallus gallus* (fowl), refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff_animal_vet-progs_2018-9_salmonella_breeding_gal_prt.pdf

3. Notification system in place to the national competent authority

Yes

- 4. Results of investigations and national evaluation of the situation, the trends and sources of infection
- 5. Additional information

17. General evaluation: Salmonella in Turkeys - breeding flocks and meat production flocks

1. History of the disease and/or infection in the country

For this information, in relation to fattening turkeys, refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff_animal_vet-progs_2018-

- 9 salmonella fattening turkeys prt.pdf
- 2. Evaluation of status, trends and relevance as a source for humans
- 3. Any recent specific action in the Member State or suggested for the European Union
- 4. Additional information

18. Description of Monitoring/Surveillance/Control programmes system: Salmonella in Turkeys - breeding flocks and meat production flocks

1. Monitoring/Surveillance/Control programmes system

Sampling strategy (breeding flocks - separate elite, grand parent and parent flocks when necessary): There are no breeding flocks of turkeys in Portugal.

Meat production flocks: Sampling is accomplished by the food business operator and by the competent authority. The sampling is done at the holding. Sampling on the initiative of the food business operator takes place within three weeks before the birds are moved to the slaughterhouse. Sampling by the competent authority includes once a year, all flocks on 10% of the holdings with at least 500 fattening turkeys and all flocks on the holding when one flock tested positive for *Salmonella* Enteritidis or *Salmonella* Typhimurium in samples taken by the food business operator, unless the meat of the turkeys in the flocks is destined for industrial heat

treatment or another treatment to eliminate *Salmonella*, and all flocks on the holding when one flock tested positive for *Salmonella* Enteritidis or *Salmonella* Typhimurium during the previous round in samples taken by the food business operator, and each time the competent authority considers it necessary.

Frequency of the sampling in meat production flocks: 3 weeks prior to slaughter/ 6 weeks prior to slaughter.

Type of specimen taken in meat production flocks: Faeces.

Methods of sampling (description of sampling techniques) in meat production flocks: At least two pairs of boot swabs shall be taken. For free range flocks, samples will only be collected in the area inside the house. All boot swabs must be pooled into one sample. In flocks with less than 100 turkeys, where it is not possible to use boot swabs as access to the houses is not possible, they may be replaced by hand drag swabs, where the boot swabs or socks are wornover, gloved hands and rubbed over surfaces contaminated with fresh faeces, or if not feasible, by other sampling techniques for faeces fit for the intended purpose. It shall be ensured that all sections in a house are represented in the sampling in a proportionate way. Each pair should cover about 50 % of the area of the house. On completion of sampling the boot/sock swabs shall be carefully removed so as not to dislodge adherent material. Boot swabs may be inverted to retain material. They shall be placed in a bag or pot and labelled.

Case definition:

Meat production flocks - Rearing period: A flock of turkeys is considered positive where
the presence of Salmonella Enteritidis and/or Salmonella Typhimurium including
Salmonella Typhimurium -Like (other than vaccine strains) is detected in the flock at any
occasion.

Diagnostic/analytical methods used

Meat production flocks - Rearing period: Bacteriological method: ISO 6579:2002.

For more information, in relation to fattening turkeys, refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff_animal_vet-progs_2018-9_salmonella_fattening_turkeys_prt.pdf

2. Measures in place

Breeding flocks (separate elite, grand parent and parent flocks when necessary):

The strategy in place is to reinforce surveillance, reinforce biosecurity measures, slaughter the positive flocks and restocking only when environmental samples are negative for *Salmonella*, with birds from flocks or herds that have undergone controls (with negative results) accordingly with the legislation requirements. The strategy includes also a close cooperation with the associations of producers to implement different means to raise awareness of the producers. The Official Services have developed guidelines for the producer, as a tool in order to guide the implementation of the national programme.

For more information, in relation to fattening turkeys, refer to the programme approved by the EC: https://ec.europa.eu/food/sites/food/files/safety/docs/cff_animal_vet-progs_2018-

9 salmonella fattening turkeys prt.pdf

3. Notification system in place to the national competent authority

Yes

4. Results of investigations and national evaluation of the situation, the trends and sources of infection

19. General evaluation: Trichinella in animal - Pigs - animal sample

1. History of the disease and/or infection in the country

Disease notifiable since 1953 by national law (Decreto-Lei No 39209, de 14 de Maio). Cases of trichinelosis are not reported since 1960.

- 2. Evaluation of status, trends and relevance as a source for humans
- 3. Any recent specific action in the Member State or suggested for the European Union
- 4. Additional information

20. Description of Monitoring/Surveillance/Control programmes system: *Trichinella* in animal - Pigs - animal sample

1. Monitoring/Surveillance/Control programmes system

Sampling strategy and frequency of the sampling: All slaughtered animals are sampled.

Type of specimen taken: diaphragm pillars.

Methods of sampling (description of sampling techniques): As determined in Commission Implementing Regulation (EU) 2015/1375 of 10 August 2015.

Case definition: It is positive when there is detection of one larvae of Trichinella.

Diagnostic/analytical methods used: Mechanical digestion of pooled samples with magnetic stirrer (Regulation (EU) 2015/1375).

- 2. Measures in place
- 3. Notification system in place to the national competent authority

Yes.

4. Results of investigations and national evaluation of the situation, the trends and sources of infection

Cases of trichinelosis are not reported since 1960.

5. Additional information

Solipeds are also sampled: tongue, diaphragm pillars and masseter.

Special training in *Trichinella* detection on slaughterhouses and game activities is given to the meat inspection team.

21. Description of Monitoring/Surveillance/Control programmes system: *Trichinella* in Wild Boars

1. Monitoring/Surveillance/Control programmes system

National Plan for Sanitary Surveillance of Large Wild Game – Surveillance plan in Large Wild Game that includes testing for several diseases: *Trichinella*, African Swine Fever, Swine Fever, Aujeszky disease, Cysticercosis and skin affections.

Sampling strategy and frequency of the sampling: All slaughtered animals are sampled.

Type of specimen taken: tongue, diaphragm pillars, masseter.

Methods of sampling (description of sampling techniques): As determined in Commission

Implementing Regulation (EU) 2015/1375 of 10 August 2015.

Case definition: It is positive when there is detection of one larvae of *Trichinella*.

Diagnostic/analytical methods used: Mechanical digestion of pooled samples with magnetic stirrer (Regulation (EU) 2015/1375).

2. Measures in place

Trichinella official testing on Wild Boars from selected hunting events. The testing is performed by the NRL for *Trichinella*.

3. Notification system in place to the national competent authority

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4. Results of investigations and national evaluation of the situation, the trends and sources of infection

In the *Trichinella* testing carried out on Wild Boars from hunting under the National Plan for Sanitary Vigilance of Large Wild Game, at the end of 2017, there were 3 positive cases in animals from the region of Trás-os-Montes, near the border with Spain. The species identified was *Trichinella britovi*. The meat of these animals was collected and destroyed.

These were the first *Trichinella* positive cases in Wild Boars in the last 10 years of official sampling.

5. Additional information

Special training in *Trichinella* detection on slaughterhouses and game activities is given to the meat inspection team.

22. Institutions and laboratories involved in antimicrobial resistance monitoring and reporting

Instituto Nacional de Investigação Agrária e Veterinária, I. P. (INIAV) Laboratório Regional Veterinário (LRV)

INIAV: laboratory supporting several official control and surveillance programs and plans, with analytical research on zoonotic agents in animal, food and feed samples. Also supports national AMR monitoring plan with analytical performance.

LRV: Autonomous Region of Azores' laboratory supporting several official control and surveillance programs and plans, with analytical research on zoonotic agents in animal, food and feed samples. Also supports national AMR monitoring plan with analytical performance obtaining the isolates that are tested for antimicrobial resistance at INIAV laboratory.

23. General Antimicrobial Resistance Evaluation

- 1. Situation and epidemiological evolution (trends and sources) regarding AMR to critically important antimicrobials (CIAs) over time until recent situation
- 2. Public health relevance of the findings on food-borne AMR in animals and foodstuffs
- 3. Recent actions taken to control AMR in food producing animals and food

Preparation of a joint action for the Reduction of collistin consumption on treatment of infectious diseases in swine producing farms in Portugal. Several partners were involved considering data from the selling of collistin in Portugal and considering also the known data for the surveillance, accordingly with the surveillance Programme undertaken in Portugal since 2014.

4. Any specific action decided in the Member State or suggestions to the European Union for actions to be taken against food-borne AMR threat

Whenever an antimicrobial (considering the B1 list of substances searched in the National Residue Monitoring Plan – NRMP) is found, the competent authority undergoes an extra official control to the farm in order to verify compliance of Legal Requirements regarding Veterinary Medicine administration in farm animals, including records, Food Chain Information and prescription.

5. Additional information

Accordingly with recent strategies adopted not only by the EU, but also by the WHO, OIE, FAO a National Plan was developed in Portugal, during the year of 2017, compromising three Ministries (Health, Agriculture and Environment) towards the fight against Antimicrobial Resistance. The National Strategic Plan was developed and will be in place for the period 2018-2020.

The strategic goals are:

- 1. Improve and pursue the goals and strategy behind the One Health concept.
- 2. Improve awareness and understanding of antimicrobial resistance.
- 3. Strengthen the knowledge and evidence base through surveillance, monitoring and research.
- 4. Reduce the incidence of infection.
- 5. Optimise the use of antimicrobial medicines in human and animal health.
- 6. Develop the economic case for sustainable investments that takes account of the needs for new medicines, diagnostic tools, vaccines and other relevant interventions.
- 7. Better coordination and implementation of EU coordination and implementation of EU rules to tackle AMR

24. General Description of Antimicrobial Resistance Monitoring: Isolates of *Escherichia coli*, non-pathogenic in fattening pigs

1. General description of sampling design and strategy

The isolates were collected under Decision 2013/652/EU with the objective of testing for antimicrobial susceptibility.

2. Stratification procedure per animal population and food category

The sampling plan was designed to obtain 170 isolates of Indicator commensal *E. coli* and 210 isolates of ESBL-or AmpC-or carbapenemase-producing *E. coli*, from 300 representative caecal samples from fattening pigs to test for antimicrobial susceptibility.

For the sample design it was used a prospective sampling strategy to cover the all year.

The first step was to identify the slaughterhouses where the fattening pigs are slaughtered and determine the number of samples to be collected in each one in function of the data on their capacity of slaughtering in 2015.

It was only used one isolate per epidemiological unit (herd), as it is determined by the legislation.

The sampling in pigs was executed in 8 slaughterhouses because they represent 64.5% of the all slaughtering.

The sampling plan was stratified per slaughterhouses by allocating the number of samples to be collected per slaughterhouse proportionally to the annual throughput of fattening pigs in each slaughterhouse in the previous year.

The number of isolates planned to be taken in each abattoir per year, was proportional allocated to each one, accordingly with the respective throughput.

After select the 8 abattoirs, we have considered the sum of their capacity and then calculate the allocation proportion (a.p.) of each one. The result of this calculation was the follow:

Abattoir A, a.p. 0,2013, 60 samples; Abattoir B, a.p. 0,1800, 54 samples; Abattoir C, a.p. 0,1586, 48 samples; Abattoir D, a.p. 0,1511, 45 samples; Abattoir E, a.p. 0,1124, 34 samples; Abattoir F, a.p. 0,0969, 29 samples; Abattoir G, a.p. 0,0499, 15 samples and Abattoir H, a.p. 0,0498, 15 samples.

The sampling has started in August and due to logistical reasons it was not possible to achieve the 300 samples collection (it was possible to collect 254 samples).

The number of isolates planned for monitoring AMR in Indicator commensal *E. coli* was achieved, except for one abattoir, where the number of isolates was lower than the allocated number to be analysed (less 30 in total); the absent 30 isolates were offset by the remainder abattoirs.

For ESBL-or AmpC-or Carbapenemase-producing *E. coli*, only in 3 abattoirs achieved it. In the other 5 abattoirs the number of available isolates was lower than the allocated number to be analysed (less 50 in total) and the total number of available isolates was lower than the required. The other abattoirs offset with 9 isolates.

The number of samples planned to collect in each slaughterhouse were divided by weeks in the different slaughterhouses, to facilitate the work of the laboratory.

All samples were collected in the first three days of the week (Monday to Wednesday) because of the reception days in the laboratory.

Accordingly, in all slaughterhouses we tested all new herds presented for slaughter in each selected day.

Failure to apply randomness criteria relates to the following factors:

- Small variability of the herds in each abattoir;
- Logistics of sampling in slaughterhouses and availability of transportation to the laboratories;
- Capacity and availability of laboratories to receive samples every day.

3. Randomisation procedure per animal population and food category

Please, see answer to point 2.

4. Analytical method used for detection and confirmation

All protocols followed in the isolation, identification and antimicrobial susceptibility testing are those recommended by the EURL-AR.

In the case of the voluntary specific monitoring on carbapenemase-producers, CARBA_SMART, a selective culture media from Biomérieux was used.

5. Laboratory methodology used for detection of antimicrobial resistance

Microdilution method using commercially available microplates from TREK (EUVSEC, EUVSEC2 and EUCAMP2).

Epidemiological cut-offs from EUCAST were used.

6. Results of investigation

7. Additional information

25. General Description of Antimicrobial Resistance Monitoring: Isolates of Escherichia coli, non-pathogenic in bovines under one year of age

1. General description of sampling design and strategy

The isolates were collected under Decision 2013/652/EU with the objective of testing for antimicrobial susceptibility.

2. Stratification procedure per animal population and food category

The sampling plan was designed to obtain 170 isolates of Indicator commensal *E. coli* and 210 isolates of ESBL-or AmpC-or carbapenemase-producing E. coli, from 300 representative caecal samples from bovines under one year of age to test for antimicrobial susceptibility.

For the sample design it was used a prospective sampling strategy to cover the all year.

The first step was to identify the slaughterhouses where the bovines under one year of age are slaughtered and determine the number of samples to be collected in each one in function of the data on their capacity of slaughtering in 2016.

It was only used one isolate per epidemiological unit (herd), as it is determined by the legislation.

The sampling in bovines was executed in 7 slaughterhouses, representing 60,3% of the all slaughtering.

The sampling plan was stratified per slaughterhouses by allocating the number of samples to be collected per slaughterhouse proportionally to the annual throughput of bovines under one year of age in each slaughterhouse in the previous year.

The number of isolates planned to be taken in each abattoir per year, was proportional allocated to each one, accordingly with the respective throughput.

After select the 7 abattoirs, we have considered the sum of their capacity and then calculate the allocation proportion (a.p.) of each one. The result of this calculation was the follow:

Abattoir A, a.p. 0,2944, 88 samples; Abattoir B, a.p. 0,1916, 58 samples; Abattoir C, a.p. 0,1202, 36 samples; Abattoir D, a.p. 0,1141, 34 samples; Abattoir E, a.p. 0,1071, 32 samples; Abattoir F, a.p. 0,0898, 27 samples and Abattoir G, a.p. 0,0827, 25 samples.

The sampling has started in August and due to logistical reasons it was not possible to achieve the 300 samples collection (it was possible to collect 297 samples).

The number of isolates planned for monitoring AMR in Indicator commensal *E. coli* was achieved but for ESBL-or AmpC-or Carbapenemase-producing *E. coli*, only in 1 abattoir achieved it. In the other 6 abattoirs the number of available isolates was lower than the allocated number to be analysed (less 103 in total) and the total number of available isolates was lower than the required.

The number of samples planned to collect in each slaughterhouse were divided by weeks in the different slaughterhouses, to facilitate the work of the laboratory.

All samples were collected in the first three days of the week (Monday to Wednesday) because of the reception days in the laboratory.

Accordingly, in each selected day the sampling was done in animals coming from herds that were not sampled before.

Failure to apply randomness criteria relates to the following factors:

- Small variability of the herds in each abattoir;

- Logistics of sampling in slaughterhouses and availability of transportation to the laboratories;
- Capacity and availability of laboratories to receive samples every day.
- 3. Randomisation procedure per animal population and food category

Please, see answer to point 2.

4. Analytical method used for detection and confirmation

All protocols followed in the isolation, identification and antimicrobial susceptibility testing are those recommended by the EURL-AR.

In the case of the voluntary specific monitoring on carbapenemase-producers, CARBA_SMART, a selective culture media from Biomérieux was used.

5. Laboratory methodology used for detection of antimicrobial resistance

Microdilution method using commercially available microplates from TREK (EUVSEC, EUVSEC2 and EUCAMP2).

Epidemiological cut-offs from EUCAST were used.

- 6. Results of investigation
- 7. Additional information

26. General Description of Antimicrobial Resistance Monitoring: Isolates of Salmonella spp in pig carcasses

1. General description of sampling design and strategy

The isolates were collected under Decision 2013/652/EU with the objective of testing for antimicrobial susceptibility.

2. Stratification procedure per animal population and food category

The isolates of *Salmonella* used for the monitoring of AMR under Decision 2013/652/EU were obtained from the samples of the food business operators, collected in order to verify compliance with process hygiene criteria set out in points 2.1.4 of Chapter 2 of Annex I of Regulation (EC) No 2073/2005.

It was not necessary to select representative isolates because we have submitted to AST all the isolates available during the year.

Only 38 isolates from pig carcasses swabs were received from the food business operators for AST testing.

3. Randomisation procedure per animal population and food category

Please, see answer to point 2.

4. Analytical method used for detection and confirmation

All protocols followed in the isolation, identification and antimicrobial susceptibility testing are those recommended by the EURL-AR.

5. Laboratory methodology used for detection of antimicrobial resistance

Epidemiological cut-offs from EUCAST were used.

6. Results of investigation

7. Additional information

All the isolates of *Salmonella* and *Campylobacter* coming from the Food Zoonoses Monitoring Plan are tested for antimicrobial resistance.

27. General Description of Antimicrobial Resistance Monitoring: Isolates of Salmonella spp in bovine carcasses

1. General description of sampling design and strategy

The isolates were collected under Decision 2013/652/EU with the objective of testing for antimicrobial susceptibility.

2. Stratification procedure per animal population and food category

The isolates of Salmonella used for the monitoring of AMR under Decision 2013/652/EU were obtained from the samples of the food business operators, collected in order to verify compliance with process hygiene criteria set out in points 2.1.3. of Chapter 2 of Annex I of Regulation (EC) No 2073/2005.

It was not necessary to select representative isolates because we have submitted to AST all the isolates available during the year.

Only 8 isolates from bovine carcasses swabs were received from the food business operators for AST testing.

3. Randomisation procedure per animal population and food category

Please, see answer to point 2.

4. Analytical method used for detection and confirmation

All protocols followed in the isolation, identification and antimicrobial susceptibility testing are those recommended by the EURL-AR.

5. Laboratory methodology used for detection of antimicrobial resistance

Epidemiological cut-offs from EUCAST were used.

6. Results of investigation

7. Additional information

All the isolates of *Salmonella* and *Campylobacter* coming from the Food Zoonoses Monitoring Plan are tested for antimicrobial resistance.

28. General Description of Antimicrobial Resistance Monitoring: Isolates of Escherichia coli, non-pathogenic in fresh pig meat

1. General description of sampling design and strategy

The isolates were collected under Decision 2013/652/EU with the objective of testing for antimicrobial susceptibility.

2. Stratification procedure per animal population and food category

The sampling plan was designed to obtain 210 isolates of ESBL-or AmpC-or carbapenemase-producing *E. coli*, from 300 representative samples of fresh pig meat to test for antimicrobial susceptibility.

The samples were collected at retail level, i.e., supermarkets and butchers.

For the sample design it was used a prospective sampling strategy to cover the all year.

The first step was to identify the NUTS-3 areas that represent 80% of the national population and allocate the samples in proportion to the size of the human population in each NUTS-3 areas.

It was collected only one sample per epidemiological unit which means one sample per lot of fresh meat.

The sampling plan was stratified per NUTS-3 by allocating the number of samples to be collected per NUTS-3 proportionally to the number of inhabitants in each NUTS-3 region.

We have selected 13 areas that represents 81,15% of the population. After, we have considered the sum of their capacity and then calculate the allocation proportion (a.p.) of each one. The result of this calculation was the follow:

Área Metropolitana de Lisboa - a.p. 0,3292, 99 samples; Área Metropolitana do Porto - a.p. 0,2053, 62 samples; Região de Coimbra - a.p. 0,0537, 16 samples; Algarve - a.p. 0,0526, 16 samples; Tâmega e Sousa - a.p. 0,0505, 15 samples; Ave - a.p. 0,0496, 15 samples; Cávado - a.p. 0,0479, 14 samples; Região de Aveiro - a.p. 0,0432, 13 samples; Oeste - a.p. 0,0423, 13 samples; Região de Leiria - a.p. 0,0344, 10 samples; Região Autónoma da Madeira - a.p. 0,0312, 9 samples; Viseu Dão Lafões - a.p. 0,0312, 9 samples and Lezíria do Tejo - a.p. 0,0289, 9 samples.

Due to logistical reasons only 224 pig meat samples were collected instead of the targeted 300.

The samples planned to collect in each NUTS-3 area, were distributed for the retailers starting by the biggest category of outlets existing in each area.

In almost all the regions the main category are the big and medium supermarkets. That's why our sampling was majority done in this kind of shops.

In each shop the maximum number of samples collected was 5 samples.

3. Randomisation procedure per animal population and food category

Please, see answer to point 2.

4. Analytical method used for detection and confirmation

All protocols followed in the isolation, identification and antimicrobial susceptibility testing are those recommended by the EURL-AR.

In the case of the voluntary specific monitoring on carbapenemase-producers, CARBA_SMART, a selective culture media from Biomérieux was used.

5. Laboratory methodology used for detection of antimicrobial resistance

Microdilution method using commercially available microplates from TREK (EUVSEC, EUVSEC2 and EUCAMP2).

Epidemiological cut-offs from EUCAST were used.

6. Results of investigation

7. Additional information

29. General Description of Antimicrobial Resistance Monitoring: Isolates of *Escherichia coli*, non-pathogenic in fresh bovine meat

1. General description of sampling design and strategy

The isolates were collected under Decision 2013/652/EU with the objective of testing for antimicrobial susceptibility.

2. Stratification procedure per animal population and food category

The sampling plan was designed to obtain 210 isolates of ESBL-or AmpC-or carbapenemase-producing *E. coli*, from 300 representative samples of fresh bovine meat to test for antimicrobial susceptibility.

The samples were collected at retail level, i.e., supermarkets and butchers.

For the sample design it was used a prospective sampling strategy to cover the all year.

The first step was to identify the NUTS-3 areas that represent 80% of the national population and allocate the samples in proportion to the size of the human population in each NUTS-3 areas.

It was collected only one sample per epidemiological unit which means one sample per lot of fresh meat.

The sampling plan was stratified per NUTS-3 by allocating the number of samples to be collected per NUTS-3 proportionally to the number of inhabitants in each NUTS-3 region.

We have selected 13 areas that represents 81,15% of the population. After, we have considered the sum of their capacity and then calculate the allocation proportion (a.p.) of each one. The result of this calculation was the follow:

Área Metropolitana de Lisboa - a.p. 0,3292, 99 samples; Área Metropolitana do Porto - a.p. 0,2053,

62 samples; Região de Coimbra - a.p. 0,0537, 16 samples; Algarve - a.p. 0,0526, 16 samples; Tâmega e Sousa - a.p. 0,0505, 15 samples; Ave - a.p. 0,0496, 15 samples; Cávado - a.p. 0,0479, 14 samples; Região de Aveiro - a.p. 0,0432, 13 samples; Oeste - a.p. 0,0423, 13 samples; Região de Leiria - a.p. 0,0344, 10 samples; Região Autónoma da Madeira - a.p. 0,0312, 9 samples; Viseu Dão Lafões - a.p. 0,0312, 9 samples and Lezíria do Tejo - a.p. 0,0289, 9 samples.

Due to logistical reasons only 224 pig meat samples were collected instead of the targeted 300.

The samples planned to be collect in each NUTS-3 area, were distributed for the retailers starting by the biggest category of outlets existing in each area.

In almost all the regions the main category are the big and medium supermarkets. That's why our sampling was majority done in this kind of shops.

In each shop the maximum number collected was 5 samples.

3. Randomisation procedure per animal population and food category

Please, see answer to point 2.

4. Analytical method used for detection and confirmation

All protocols followed in the isolation, identification and antimicrobial susceptibility testing are those recommended by the EURL-AR.

In the case of the voluntary specific monitoring on carbapenemase-producers, CARBA_SMART, a selective culture media from Biomérieux was used.

5. Laboratory methodology used for detection of antimicrobial resistance

Microdilution method using commercially available microplates from TREK (EUVSEC, EUVSEC2 and EUCAMP2).

Epidemiological cut-offs from EUCAST were used.

6. Results of investigation

7. Additional information