

ZOONOSES MONITORING

Belgium

TRENDS AND SOURCES OF ZOONOSES AND ZOONOTIC AGENTS IN FOODSTUFFS, ANIMALS AND FEEDINGSTUFFS

including information on foodborne outbreaks, antimicrobial resistance in zoonotic and indicator bacteria and some pathogenic microbiological agents

IN 2016

PREFACE

This report is submitted to the European Commission in accordance with Article 9 of Council Directive 2003/99/EC*. The information has also been forwarded to the European Food Safety Authority (EFSA).

The report contains information on trends and sources of zoonoses and zoonotic agents in Belgium during the year 2016.

The information covers the occurrence of these diseases and agents in animals, foodstuffs and in some cases also in feedingstuffs. In addition the report includes data on antimicrobial resistance in some zoonotic agents and indicator bacteria as well as information on epidemiological investigations of foodborne outbreaks. Complementary data on susceptible animal populations in the country is also given. The information given covers both zoonoses that are important for the public health in the whole European Union as well as zoonoses, which are relevant on the basis of the national epidemiological situation.

The report describes the monitoring systems in place and the prevention and control strategies applied in the country. For some zoonoses this monitoring is based on legal requirements laid down by the European Union legislation, while for the other zoonoses national approaches are applied.

The report presents the results of the examinations carried out in the reporting year. A national evaluation of the epidemiological situation, with special reference to trends and sources of zoonotic infections, is given. Whenever possible, the relevance of findings in foodstuffs and animals to zoonoses cases in humans is evaluated.

The information covered by this report is used in the annual European Union Summary Reports on zoonoses and antimicrobial resistance that are published each year by EFSA.

^{*} Directive 2003/ 99/ EC of the European Parliament and of the Council of 12 December 2003 on the monitoring of zoonoses and zoonotic agents, amending Decision 90/ 424/ EEC and repealing Council Directive 92/ 117/ EEC, OJ L 325, 17.11.2003, p. 31

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N_A Gallus qallus (fowl) - broilers - Farm - Control and eradication programmes - Official and industry sampling - AMR MON	197
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N.A Salmonella Landon N.A Salmonella London N.A Salmonella London N.A Salmonella London N.A Salmonella Mirantia Meat from pig - carcase - Slaughterhouse - Monitoring - Official sampling - AMR MON N.A Salmonella Mirantia Salmon	
N.A. Salmonella Landon Gallus gallus (fow) - brollers - Farm - Control and eradication programmes - Official and Industry sampling - AMR MON N.A. Salmonella London Meat from pp - carcase - Saughterhouse - Monitoring - Official sampling - AMR MON N.A. Salmonella Mathandaka Gallus gallus (fow) - brollers - Farm - Control and eradication programmes - Official and Industry sampling - AMR MON N.A. Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and Industry sampling - AMR MON N.A. Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and Industry sampling - AMR MON N.A. Gallus gallus (fow) - laying hers - Farm - Control and eradication programmes - Official and Industry sampling - AMR MON N.A. Mate from brollers (Gallus gallus) - carcase - Saughterhouse - Monitoring - EPSA specifications - Official sampling - AMR MON N.A. Salmonella Mateuake Meat from brollers (Gallus gallus) - carcase - Saughterhouse - Monitoring - EPSA specifications - Official sampling - AMR MON N.A. Salmonella Milesuake Meat from brollers (Gallus gallus) - carcase - Saughterhouse - Monitoring - EPSA specifications - Official sampling - AMR MON N.A. Salmonella Milesuake Meat from brollers (Gallus gallus) - carcase - Saughterhouse - Monitoring - EPSA specifications - Official sampling - AMR MON N.A. Salmonella None Gallus gallus (fow) - brollers - Farm - Control and eradication programmes - Official sampling - AMR MON N.A. Salmonella None N.A. Salmonella None Salmonella None Meat from pip - Retall - Monitoring - Official sampling - OTHER AMR MON N.A. Salmonella Parama Frogs leg - Retall - Monitoring - Official sampling - OTHER AMR MON N.A. Salmonella Parama Frogs leg - Retall - Monitoring - Official sampling - OTHER AMR MON N.A. Salmonella Parama Frogs leg - Retall - Monitoring - Official sampling - OTHER AMR MON N.A. Salmonella Parama Meat from brollers (Gallus gallus) - carcase - Saughterhouse - Monitoring - Official sampling -	
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1 ANIMAL POPULATIONS

The relevance of the findings on zoonoses and zoonotic agents has to be related to the size and nature of the animal population in the country

1.1 Populations

1.1.1 Information on susceptible animal population

Sources of information

SANITEL and BELTRACE database of the Federal Agency for the Safety of the Food Chain.

Dates the figures relate to and the content of the figures

Number of animals = number of animals at a certain time point of the year. Number of slaughtered animals = total number of slaughtered animals during the year.

Definitions used for different types of animals, herds, flocks and holdings as well as the types covered by the information

Holding: any establishment, construction or, in the case of an open-air farm, any place in which animals are held, kept or handled. The location of the holding is based on the address and the coordinates of the geographical entity. A geographical entity is a unit of one building or a complex of buildings included grounds and territories where an animal species is or could be hold. Herd: an animal or group of animals kept on a holding as an epidemiological unit; if more than one herd is kept on a holding, each of these herds shall form a distinct unit and shall have the same health status.

National evaluation of the numbers of susceptible population and trends in these figures

Over the last years, there's a continuous decrease in total number of holdings of animal species. The total number of bovine animals remains unchanged what means that the mean total number of animals per holding is increasing. The total numbers of holdings and animals of swine, are decreasing over the last years.

Geographical distribution and size distribution of the herds, flocks and holdings

Belgium can be geographically divided into two regions: the Flemish region situated in the north and the Walloon region situated in the south of the country. There's a very dense animal population of bovines, swine and poultry in the Flemish region. The Walloon region is important for his cattle breeding holdings of the Belgian Blue White race. The number of porcine and poultry holdings in the Walloon region is rather limited.

2 DISEASE STATUS

2.1 TUBERCULOSIS, MYCOBACTERIAL DISEASES

2.1.1 General evaluation of the national situation

2.1.1.1 Mycobacterium - general evaluation

History of the disease and/or infection in the country

Zoonotic tuberculosis (Mycobacterium bovis). Bovine tuberculosis in humans caused by M. bovis is clinically indistinguishable from human tuberculosis caused by M. tuberculosis. In the past, the most important way of transmission of M. bovis to humans was the consumption of raw milk or raw milk products from infected cattle. Industrial heating during production methods or pasteurization of raw milk did stop this way of transmission to humans. Nowadays tuberculosis in humans caused by M. bovis is rare. In regions were M. bovis infections in cattle are largely eliminated, only few residual cases occur among elderly persons as a result of the reactivation of dormant M. bovis within old lesions. Also among migrants from high-prevalence countries or regions, infections with M. bovis are diagnosed. Agricultural workers may acquire infection by M. bovis by inhaling cough aerosols from infected cattle and may subsequently develop typical pulmonary or genito-urinary tuberculosis. Cervical lymphadenopathy, intestinal lesions, chronic skin tuberculosis (lupus vulgaris) and other non-pulmonary forms are also particularly common as clinical symptoms.

Recent actions taken to control the zoonoses

The surveillance program of tuberculosis is based on Directive 64/432/EEC, which is implemented and adapted in National legislation since 1963 and last modified by the Royal Decree of 17 October 2002. The control implies skin testing of animals at the occasion of trade and intensive testing of infected and contact farms in consequence of a confirmation of a bovine TB suspicious case (tracing-on and tracing-back of all contact animals and contact herds). Systematic ante- and post-mortem examination are performed at all slaughterhouses. The Federal Agency for the Safety of the Food chain is informed about any doubtful or positive result of the skin test of bovines and may decide to re-examine (additional tests e.g. comparative tuberculin test, interferon-gamma test) the animals or to kill them for additional analysis (test and slaughter strategy). In case a "TB suspicious" lesion is detected, a tissue sample is sent to the National Reference Laboratory for analysis. Consequently, if Mycobacterium bovis suspicion is confirmed by a positive culture or PCR, all animals in the herd of origin are skin tested and an epidemiological investigation is realized. The total herd is considered as the 'epidemiological unit'. Isolation of M. bovis and biochemical testing is exclusively performed in the National Reference Laboratory where also IFN-gamma, PCR and molecular typing by means of RFLP, spoligotyping or more recently MIRU-VNTR are done to support the epidemiological investigations and to eventually prove the link between different cases.

Suggestions to the European Union for the actions to be taken

In case a holding is infected and if by epidemiological investigation and tracing-back, animals were found to have been exported to another country, the Chief Veterinary Officer of the country of destination has to be informed about the outbreak in the country of origin. This alert can help to rapidly detect an infection in the concerned holding of destination abroad. Monitoring of the type of strains circulating in each country could contribute to the understanding of the temporal-spatial spread of some specific strains between different countries and could possibly bear some epidemiological links between different outbreaks. More attention should be given to intracommunity trade in animals sensitive to bovine tuberculosis (e.g. camelids), especially if those animals have stayed for a time in an endemic region of tuberculosis. Attention should also be given to early detect a possible incurrence of infected wildlife as deer, wildboar and badgers from neighbouring countries.

2.1.2 Mycobacterium in animals

2.1.2.1 Mycobacterium tuberculosis complex (MTC) in animal - Deer - farmed - animal sample

Monitoring system

Sampling strategy

Sampling in case of suspicious TB lesions during post-mortem examinations of "wild" and "farmed" deer at slaughterhouse/ at game handling establishment.

Frequency of the sampling

Depends on the number of hunted/slaughtered animals and the detection of suspicious lesions at post-mortem examination.

Type of specimen taken

Suspicious lesions of lungs, lymph nodes, ... at slaughterhouse or game handling establishment.

Methods of sampling (description of sampling techniques)

TB suspicious tissues: lymph nodes, lungs, ...

Case definition

An animal is positive if Mycobacterium boyis is isolated by culture or confirmed by laboratory analysis.

Diagnostic/analytical methods used

- Ziehl-Neelsen coloration - Culture for isolation - Interferon-gamma - PCR on lesions / organs - PCR on culture

Control program/mechanisms

The control program/strategies in place

Monitoring is done by:- systematic post-mortem examination at the slaughterhouses/game handling establishment- post-mortem examination at autopsy of hunted or killed "wild" deer by accident in the University Center of Liège, Veterinary Medicine Faculty. In case of suspected TB lesions, tissue samples are sent to the National Reference Laboratory for additional analysis to confirm the suspicion.

Recent actions taken to control the zoonoses

Surveillance program in wildlife.

National evaluation of the recent situation, the trends and sources of infection

No Mycobacterium bovis was detected in "wild/hunted" or "farmed" deer for the reporting year 2016.

2.1.2.2 Mycobacterium tuberculosis complex (MTC) in animal - Cattle (bovine animals) - animal sample

Status as officially free of bovine tuberculosis during the reporting year

The entire country free

Belgium is officially free of bovine tuberculosis since the 25th of June 2003 (Commission Decision 2003/467/EC)

Free regions

All regions are officially free of bovine tuberculosis.

Monitoring system

Sampling strategy

Surveillance system. The control of tuberculosis is based on Council Directive 64/432/EEC, which is implemented and adapted in National legislation since 1963 and was last modified by the Royal Decree of 17 October 2002. The surveillance program implies:- skin testing of all animals at purchase by the veterinarian responsible for the epidemiological sanitary situation of the holding (contract between farmer and veterinarian); - in case of a suspected/infected bovine(s) on a holding skin testing of all animals of the holding; - skin testing of all 'contact' animals and herds (tracing-on and tracing-back);- systematic ante- and post-mortem examination of all slaughtered bovines, transmission to the National Reference Laboratory of all "TB suspicious" lesions for further analysis. Isolation of M. bovis and typing is performed at the National Reference Laboratory CODA-CERVA. Also IFN-gamma, PCR and molecular typing by means of RFLP, spoligotyping and more recently MIRU-VNTR are realised at the NRL.

Frequency of the sampling

Frequency of testing depends on:- the introduction of new animals into a herd (mandatory examination at purchase) - the results of tuberculin testing- the detection of suspected bovines- the detection of infected bovines- the epidemiological investigation related to suspected or infected animals or herds (tracing-on and tracing-back)- the follow-up testing of infected and/or eradicated herds during 5 years after partial or total stamping-out. This follow-up testing can be reduced to year 1, year 3 and year 5 if epidemiological investigations indicate a low risk of infection- an 'at random selection' of 200 holdings in the category of holdings with an important number of purchased animals per year: tuberculination of all purchased bovines over the last year.

Type of specimen taken

Organs/tissues: lesions, lymph nodes, lungs, liver, kidneys, spleen, ...Blood

Methods of sampling (description of sampling techniques)

Tuberculin skin testing: single (bovine tuberculin) or comparative (bovine/avian tuberculin) testing. Blood sampling: interferon-gamma tests Laboratory examination of all suspicious lesions by culture: isolation and identification or analysis by PCR. Organs: lymph nodes, lungs, liver, kidneys, ...

Case definition

- A 'bovine' is defined as infected with bovine tuberculosis if the animal is positive by skin testing or if Mycobacterium bovis is isolated by culture or confirmed by laboratory analysis (PCR). - A 'holding' is defined as infected if Mycobacterium bovis was isolated by culture from an animal of the holding.

Diagnostic/analytical methods used

- Simple skin test with bovine tuberculin- Comparative skin test with bovine and avian tuberculin- Ziehl-Neelsen coloration- Culture for isolation- Interferon-gamma assay- PCR on lesions / organs- PCR on culture - RFLP typing- Spoligotyping- MIRU-VNTR

Vaccination policy

Vaccination is prohibited by Royal Decree of 17 October 2002.

Control program/mechanisms

The control program/strategies in place

National surveillance program by the Competent Authority (FASFC) on a mandatory legal base.

Recent actions taken to control the zoonoses

Draw special attention and focus on the post-mortem examination of slaughtered animals; Transmission for further analysis of any lesion that could be 'suspected' of tuberculosis to the National Reference Laboratory; Culture of M. bovis, biochemical testing, PCR are performed on these 'suspicious' lesions; Molecular typing by means of RFLP, Spogilotyping and more recently MIRU-VNTR are realised on all isolates to support the epidemiological investigations and to eventually prove the link between different cases or outbreaks.

Suggestions to the European Union for the actions to be taken

In case of export of bovines, inform the Chief Veterinary Officer of the Member state of destination if tuberculosis has been detected in a holding of the Member State of origin after the date of export. This information can result in an early detection or can avoid a possible further contamination in the Member State of destination.

Measures in case of the positive findings or single cases

If M. bovis is suspected, all animals in the herd of origin are skin tested, the herd is considered as the epidemiological unit. A complete epidemiological investigation is performed. After tracing-back and tracing-on all animals of 'contact' holdings are examined by skin testing. If any doubtful or positive result of the skin test is detected, the FASFC may decide to re-examine the reactor animals (additional tests e.g. comparative skin testing with avian and bovine tuberculin and/or Interferon-gamma testing) or to a direct mandatory slaughter of the reactors (test slaughter) for additional analysis. In case a suspicious lesion is detected at post-mortem examination, a sample is sent to the National reference laboratory for analysis. If in consequence Mycobacterium bovis is isolated, all skin test positive animals during successive testing are mandatory slaughtered. If many bovines are reacting positive to skin testing, the FASFC can decide that all animals of the holding must be mandatory slaughtered (total stamping-out). In most breakdowns a sanitation plan is established taking into account the epidemiological situation. In case of partial stamping-out, only 2 sanitation plans may be realised. After stamping-out, new restocked animals are tested three times during a 5 years period by annual skin testing to prove the TB free status of the holding.

Notification system in place

Animal Health Law of 24 March 1987 Chapter III and Royal Decree of 3 February 2014 (list of all notifiable animal diseases).

Results of the investigation

In 2016, 2 outbreaks of bovine tuberculosis were detected. On the first outbreak (2016-01) a general stamping-out was applied and 48 contact herds had to be followed-up by tuberculination. Hereby one secundary outbreak was detected with a clear epidemiological link to the primary case. On the second outbreak (2016-02) also a general stamping-out was organized. Due to this second outbreak 90 contact herds had to be followed-up by tuberculination. Of outbreak 2016-01 and 2016-02, respectively 1 and 7 isolates where obtained by bacteriological examination.

National evaluation of the recent situation, the trends and sources of infection

Number of infected herds since 2000 = 2000 : 24 / 2001 : 23 / 2002 : 13 / 2003 : 7 / 2004 : 8 / 2005 : 5 / 2006 : 8 / 2007 : 5 / 2008 : 12 / 2009 : 2 / 2010 : 0 / 2011: 1 / 2012: 1 / 2013: 9 / 2014: 0 / 2015: 3 / 2016: 2

2.2 BRUCELLOSIS

2.2.1 Brucella in animals

2.2.1.1 B. suis in animal - All animals - animal sample

Monitoring system

Case definition

An animal is positive if Brucella suis is isolated by culture or typed by additional laboratory methods.

2.2.1.2 B. suis in animal - Pigs - animal sample

Monitoring system

Sampling strategy

Serological screening for Brucella is done for breeding pigs that are gathered (at a fair for example), at artificial insemination centers and in animals intended for trade. The methods used are Rose Bengal test (RBT), Slow Agglutination test (SAT) according to Wright, Complement Fixation test (CFT) and ELISA. Bacteriological examination for Brucella and Yersinia is done in case of positive serology. Regularly, false positive serological reactions are reported. These are due to a Yersinia enterocolitica O9 infection and are confirmed by Yersinia enterocolitica 09 isolation in the absence of Brucella spp. isolation. B. suis biovar 2 may be isolated from wild boars (Sus scrofa). The infection seems to be endemic in wild boar in Belgium. B. suis biovar 2, circulating among wild boars, shows only limited pathogenicity for humans, if pathogenic at all. The domestic pig population is free of brucellosis (last Brucella isolation in domestic pigs in Belgium was in 1969).

Methods of sampling (description of sampling techniques)

Blood sampling Tonsils Spleen

Diagnostic/analytical methods used

Rose Bengal test RBT / Complement fixation test CFT / Indirect ELISA / Bacteriological examination

2.2.1.3 B. abortus in animal - Cattle (bovine animals) - animal sample

Status as officially free of bovine brucellosis during the reporting year

The entire country free

Belgium is officially free from bovine brucellosis since the 25th of June 2003 (Commission Decision 2003/467/EC)

Free regions

Belgium remained officially free of bovine brucellosis during this reporting year.

Additional information

End 2010 a brucellosis breakdown herd was detected after analyzing an abortion. The infected herd was totally depopulated. In March 2012, again a breakdown of brucellosis was detected after analysis of an abortion. No epidemiological link could be found with the breakdown of 2010. Tracing-back and an epidemiological inquiry lead to the detection of 4 other secondary breakdowns linked to the primary case. All these 5 brucellosis breakdown herds were infected with an identical Brucella abortus biovar 3. Another infected herd of brucellosis was detected by analysis of tankmilk and an infection with Brucella suis biovar 2 was confirmed. Finally there was a stamping-out of all the animals of the infected herds. In 2013 a breakdown herd was detected as contact herd of the primary breakdown herd of 2012. The breakdown herd of 2013 was already examined twice by serology in 2012 with negative results. A third follow-up screening by serology indicated some positive results. This positive serology could be confirmed by culture after test and slaughter of the reactors. Finally 6 bovines were infected. There was a stamping-out of all the animals of this infected herd. In 2014, bovine brucellosis was not detected by a serological follow-up surveillance of contact herds of the brucellosis incident. In 2015, no case of brucella infection was detected by a last serological follow-up surveillance of contact herds during the winter campaign of the 2012 - 2013 brucellosis incident. Scientific advice 05-2016 of the Scientific Committee of the FASFC on the re-emergence of bovine brucellosis in Belgium was published in May 2016. End of 2016, in a holding of cattle, one bovine was found infected with B. suis biovar 2 by microbiological examination after mandatory test slaughter due to serological positive reaction. This holding was finally totally depopulated. Probably contact with wildboar could have been the origin of infection with this opportunistic pathogen.

8

Monitoring system

Since Belgium is officially free of bovine brucellosis, the eradication program has been changed in a surveillance program. An animal is legally suspected of brucellosis in case of a positive ELISA. If, according to the epidemiology and the results of the blood test, an animal or herd is found to be at risk, a bacteriological investigation always takes place. Hence, a brucellosis animal is defined as an animal in which Brucella abortus has been isolated, and a cattle holding is considered as an outbreak herd if one of the animals is positive for brucellosis by bacteriological examination. In 2009, a study was realized to evaluate the current national surveillance program of bovine brucellosis. If a Member State has maintained the officially free status of brucellosis for at least 5 consecutive years, the existing surveillance program can be re-evaluated and some modifications on the sampling design are allowed on condition of further proof of freedom of disease (Council Directive 64/432/EEC). The study also clearly indicated that the best approach is to test bovines imported from officially free or nonofficially free Member States of Brucella spp., to test animals at purchase in consequence of national trade as well as to analyze aborting animals in order to early detect infection. Also the mandatory analysis for brucellosis at purchase of new animals changed into a voluntary approach. A new surveillance program has been applied from the end of 2009 on. In 2016, surveillance was focused on following risk categories:- import of non officially free MSs or Third Countries at the moment of trade and follow-up testing during 3 consecutive years during the winterscreening (targeted selection)- at random selection of 450 bovine herds for serological investigation of 40 animals per herd divided in 4 different age categories: 10 animals of 6-12 months of age, 10 animals of 12-24 months of age and 20 animals older then 24 months. - number of analysis of bovines of national trade at purchase. - at random selection of 450 bovine herds of all herds that did not declare any abortion during the past year and did send some lightweight bodies of newborns to the rendering plant. On these herds a maximum of 20 female animals are randomly selected for serological analysis of brucellosis. - abortion protocol: all abortions should be notified and analysed for brucellosis.- a general screening of dairy herds by an ELISA of tankmilk was realised in spring and autumn 2016. Serological follow-up of a positive tankmilk sample with test slaughter of one serological positive reactor animal with microbiological examination proved the infection with Brucella suis biovar 2 of that bovine.

Frequency of the sampling

- import of non officially free MSs or Third Countries at the moment of trade: all imported animals over 12 months of age - import of non officially free MSs or Third Countries follow-up testing during winterscreening for 3 consecutive years of all imported animals over 24 months of age- at random selection of 450 bovine herds: at random selection of maximum 40 female animals in different age categories-bovines of national trade at purchase: at random selection, limited number of analysis - at random selection of 450 bovine herds where no abortion was declared/analyzed during the last year, at random selection of 20 female animals over 24 months of age (goal is to stimulate the notification of abortions)- abortion protocol: examination of abortions for brucellosis and some other diseases which can induce an abortion in bovine animals (IBR, BVD, Neoplasmose, ...).

Type of specimen taken

Blood samples, Tankmilk, individual milk samples

Methods of sampling (description of sampling techniques)

Blood sampling by veterinary practioners Tankmilk samples taken for Milk Quality Assurance Associations at the moment of the collection of the milk by the milk factory

Case definition

An animal is defined as infected if Brucella spp. has been isolated by culture and identified as Brucellosis. A herd is defined as infected if one of its animals is positive by bacteriological examination for Brucellosis.

Diagnostic/analytical methods used

- Micro agglutination test - ELISA on blood or tank milk - Complement Fixation Test- Rose Bengale Test- PCR- Stamp/Ziehl Neelsen coloration- Culture

Vaccination policy

Vaccination is prohibited in Belgium since 1992.

Control program/mechanisms

The control program/strategies in place

National mandatory surveillance program organized by the FASFC.

Measures in case of the positive findings or single cases

In case of a positive result in the micro-agglutination test the same blood sample is tested with an ELISA. If this indirect ELISA is positive, this result has to be confirmed by a blocking (home made) ELISA at the NRL. If this confirmatory test is positive, the animal is considered as infected and is compulsory slaughtered (test and slaughter strategy) for additional analysis to detect a possible Brucella infection by culture.

Notification system in place

Animal Health Law of 24 March 1987 Chapter III, Royal Degree of 3 February 2014 (list of notifiable diseases)

National evaluation of the recent situation, the trends and sources of infection

An intensified bovine brucellosis control program started in Belgium in 1988. In case of active brucellosis, i.e. excretion of Brucella, the plan consisted in the culling of all animals of the infected herd (total depopulation). Culled bovines were compensated for based on the replacement value of the animals. In March 2000, the last case of bovine brucellosis was identified before obtaining the officially brucellosis free status in 2003. In case of positive serological reactors the Federal Agency for the Safety of the Food Chain instruct follow-up testing or 'test and slaughter' for additional analyses. These analyses could not confirm brucellosis. To reduce the number of FPSR (False positive serological reactors) to be slaughtered, the micro-agglutination test has been used as for routine testing whereas the indirect Elisa is accepted as a complementary test by serial or parallel testing. The blocking ELISA of the NRL is considered as the confirmation test. This approach avoids the undeserved mandatory slaughter of false positive reacting animals.In 2016, no infected animals or herds were detected.

2.2.1.4 B. melitensis in animal - Goats - animal sample

Status as officially free of caprine brucellosis during the reporting year

The entire country free

Belgium is officially free of B. melitensis since 29 March 2001 (Commission Decision 2001/292/EC).

Free regions

Belgium is officially free of caprine brucellosis during the reporting year.

Monitoring system

Sampling strategy

Serum samples taken in the framework of a national monitoring program for Maedi-Visna/CAE and at export were examined for Brucella melitensis specific antibodies by means of an ELISA. Sheep and goats were tested for brucellosis by indirect ELISA(iELISA) at the NRL CODA-CERVA. All positive samples in the ELISA were supplementary tested by the Rose Bengal Test (RBT) and Complement Fixation Test (CFT) as confirmatory tests. Animals that where positive in the two confirmatory tests or that could not be analyzed and/or interpreted in RBT and/or CFT were sampled a second time.

Type of specimen taken

Blood

Methods of sampling (description of sampling techniques)

Blood sampling

Case definition

A goat is defined as infected with brucellosis if positive in all three tests: iElisa, Rose Bengal test and Complement Fixation test and isolation of Brucella melitensis by culture after test slaughter.

Diagnostic/analytical methods used

Complement Fixation Test CFT / Rose Bengal Test RBT / Indirect ELISA / Culture for isolation

Notification system in place

Animal Health Law of 24 March 1987 Chapter III and Royal Decree of 3 February 2014 (list of notifiable animal diseases)

Results of the investigation

At the NRL, 6.955 caprine/ovine serum samples were tested. The results confirmed those of previous years, i.e. the absence of any epidemiological or bacteriological evidence of caprine/ovine brucellosis in Belgium.

2.2.1.5 B. melitensis in animal - Sheep - animal sample

Status as officially free of ovine brucellosis during the reporting year

The entire country free

Belgium is officially free from B. melitensis since 29 March 2001 (Commission Decision 2001/292/EC).

Free regions

Belgium is officially free of ovine brucellosis during the reporting year.

Monitoring system

Sampling strategy

Serum samples taken in the framework of a national monitoring program for Visna-Maedi/CAE and at export were examined for Brucella melitensis specific antibodies by means of an iELISA. Positive samples were subsequently tested in Rose Bengal and in complement fixation test. Sheep and goats sera were tested for brucellosis by indirect ELISA (iELISA) at the NRL. All positive samples in the ELISA were than tested by the Rose Bengal Test (RBT) and Complement Fixation Test (CFT) as confirmatory tests. Animals that were positive in the two confirmatory tests or that could not be analyzed and/or interpreted in RBT and/or CFT were sampled a second time.

Type of specimen taken

Blood

Case definition

A sheep is defined as infected with brucellosis if positive in all three tests: the Elisa, the Rose Bengal test and the Complement Fixation test and isolation of Brucella melitensis by culture.

Diagnostic/analytical methods used

- Indirect ELISA - Rose Bengal Test RBT- Complement Fixation Test CFT- Culture for isolation- Brucellin skin test (BST)

Notification system in place

Animal Health Law of 24 March 1987 Chapter III and Royal Decree of 3 February 2014 (list of notifiable animal diseases).

Results of the investigation

At the National Reference Laboratory, 6.955 caprine/ovine serum samples were tested. The results confirmed those of previous years, i.e. the absence of any epidemiological or bacteriological evidence of caprine/ovine brucellosis in Belgium.

3 INFORMATION ON SPECIFIC ZOONOSES AND ZOONOTIC AGENTS

Zoonoses	are dis	eases o	or infections,	which are	e naturally	transmissib	le directly	or indirec	tly between	en anin	nals and	hum	ans.
Foodstuff	s serve	often a	s vehicles of	f zoonotic	infections.	. Zoonotic a	gents cove	er viruses,	bacteria,	fungi, _l	parasites	or o	ther
biological	entities	that are	e likely to ca	use zoono	ses.								

3.1 SALMONELLOSIS

3.1.1 Salmonella in foodstuffs

3.1.1.1 Salmonella in food - All foodstuffs - Unspecified - food sample - Surveillance - Official sampling - Objective sampling

Monitoring system

Sampling strategy

Samples are taken by the Federal Agency for the Safety of the Food Chain. The samples assayed were carcasses, cuts and minced meat from pork, carcasses, cuts and meat preparation from chicken, layer carcasses, beef minced meat and other foodstuffs. Salmonella isolates were serotyped.

3.1.2 Salmonella in animals

3.1.2.1 Salmonella in animal - Gallus gallus (fowl) - broilers - Farm - environmental sample - boot swabs - Control and eradication programmes - Official and industry sampling - Census

Monitoring system

Sampling strategy

Broiler flocks

The official surveillance program for broilers in accordance with Regulations (EC) Nos 2160/2003 and 200/2012 started in 2009. It is compulsory to sample all flocks on farms with a capacity of 200 or more birds as day-old chicks and in the last three weeks before slaughter.

Frequency of the sampling

Broiler flocks: Day-old chicks

Each 'batch' of day-old chicks that enters the farm must be sampled in the hatchery or when arriving on the farm.

Broiler flocks: Before slaughter at farm

Every flock is sampled in the last 3 weeks before slaughter.

Broiler flocks: At slaughter (flock based approach)

Sampling of caeca at slaughter is distributed evenly throughout the year

Type of specimen taken

Broiler flocks: Day-old chicks

For the monitoring of day-old chicks, samples of internal linings of delivery boxes or hatcher basket liners are taken.

Broiler flocks: Before slaughter at farm

In the three weeks before slaughter, boot swab samples are taken.

Broiler flocks: At slaughter (flock based approach)

At slaughter, caeca samples are taken.

Methods of sampling (description of sampling techniques)

Broiler flocks: Day-old chicks

Pieces of inner linings of the delivery boxes are sampled by the owner in the same way as for breeding flocks. The samples have to reach an accredited laboratory within 48 hours of sampling.

Broiler flocks: Before slaughter at farm

All flocks are sampled, by the owner, within 3 weeks before slaughter. The sampling is performed in accordance with Regulation (EU) n 200/2012. Samples have to reach an accredited laboratory within 48 hours.

Broiler flocks: At slaughter (flock based approach)

The intact caeca of 10 birds from the same flock are taken at the slaughterhouse with the aim to determine the load of Salmonella spp. entering the slaughterhouse.

Case definition

Broiler flocks: Day-old chicks

A sample is considered positive if a Salmonella spp. is isolated. A flock is considered positive as soon as one sample is positive.

Broiler flocks: Before slaughter at farm

A sample is considered positive if a Salmonella spp. is isolated. A flock is considered positive as soon as one sample is positive.

Diagnostic/analytical methods used

Broiler flocks: Day-old chicks

The analytical method used is the bacteriological method: ISO 6579:2002 annex D inaccordance with regulation (EU) nr. 200/2012. Al isolates are serotyped by the Kauffmann-White-LeMinor scheme.

Broiler flocks: Before slaughter at farm

The analytical method used is the bacteriological method: ISO 6579:2002 annex D in accordance with regulation (EU) nr. 200/2012. Al isolates are serotyped by the Kauffmann-White-LeMinor scheme.

Broiler flocks: At slaughter (flock based approach)

The analytical method used is the bacteriological method: ISO 6579:2002 annex D. Al isolates are serotyped by the Kauffmann-White-LeMinor scheme.

Vaccination policy

Broiler flocks

There is no vaccination policy for broiler flocks.

Other preventive measures than vaccination in place

Broiler flocks

Minimal requirements are laid down for holdings with at least 200 broilers on infrastructure, management, hygiene and bio-security issues in the framework of the authorization of holdings.

Control program/mechanisms

The control program/strategies in place

Broiler flocks

The minimal requirements in the framework of the authorization of farms with more than 200 birds contains preventive measures (infrastructure, management, hygiene and biosecurity) for the control of Salmonella. Following measures are taken when a flock is positive for Salmonella spp: 1) logistic slaughter of the flock at the end of production; 2) mandatory cleaning and disinfection of the house; 3) hygienogram after disinfection and after the house has dried up; 4) swab control on the presence of Salmonella before restocking the house. If the following flock is positive for the same serotype of Salmonella, the disinfection must be performed by an external company. When the same serotype of Salmonella is found at three consecutive times, the farm must be evaluated on biosecurity and hygiene by the farm veterinarian and necessary measures must be taken. An epidemiological investigation and/or tests are performed to find the source of the infection. It is at all times prohibited to treat for Salmonella with antibiotics.

Measures in case of the positive findings or single cases

Broiler flocks: Day-old chicks

It is prohibited to treat the flock for Salmonella with antibiotics.

Broiler flocks: Before slaughter at farm

See 'the control program/strategies' in place.

Notification system in place

Zoonotic Salmonella is notifiable since the first of January 2004. Notification is done by phone, fax or by e-mail to the Federal Agency for the Safety of the Food Chain. Farmers and laboratories are obliged to notify.

Results of the investigation

5.959 batches of day-old chicks were sampled, 8 were positive for Salmonella spp. of which 2 for S. Typhimurium. 9.846 broiler flocks were sampled in the last 3 weeks of production. 162 flocks were positive for Salmonella spp. of which 25 for S. Typhimurium, 2 for S. Enteritidis and 6 for monophasic S. Typhimurium. The most common other serotypes found was S. Infantis in 72 flocks, followed by S. Gaminara (12 flocks), S. Livingstone (11 flocks) and S. Java (10 flocks). 236 batches were sampled atthe level of the slaughterhouse, 20 were positive for Salmonella spp. of which 2 for S. Typhimurium.

National evaluation of the recent situation, the trends and sources of infection

The prevalence of all serotypes in day old chicks (0,14%) decreased compared to 2015 (0,29%), mainly due to the decrease of the number of S. Mbandaka and S. Enteritidis positive flocks. The prevalence of Salmonella spp. in broiler flocks (1,65%) is comparable with 2015 (1,43%). However, an increase in the number of positive flocks of S. Typhimurium and S. Infantis is seen compared to 2015. The prevalence of Salmonella spp. at the level of the slaughterhouses (8,5%) (caeca) is comparable with 2015 (5%). The prevalence of S. Typhimurium/Enteritidis is less than 1%.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

After 3 years of increase (2.760 in 2013, 2.963 in 2014, 3.119 in 2015) in the total number of reported human Salmonella isolates, a slight decrease was seen in 2016 (3.026). The number of S. Typhimurium isolates decreased from 1.856 in 2015 to 1.412 in 2016 but the number of S. Enteritidis isolates increased from 458 in 2015 to 600 in 2016. An increase in the number of isolates belonging to other serotypes was also seen. Despite the high number of S. Infantis positive flocks, an increase in the number of human cases was not seen (68 in 2015, 66 in 2016).

3.1.2.2 Salmonella in animal - Gallus gallus (fowl) - laying hens - Farm - environmental sample - boot swabs and dust - Control and eradication programmes - Official and industry sampling - Census

Monitoring system

Sampling strategy

Laying hens flocks

All laying hen flocks on farms with at least 200 laying hens are under the Salmonella control programme. Flocks are sampled by the owner at the age of day-old chicks, 16 and 24 weeks, every 15 weeks during production and in the last 3 weeks of production. When a flock has a second production cycle, the sampling continues every 15 weeks.

Frequency of the sampling

Laying hens: Day-old chicks

Every flock of day-old chicks is sampled before entering the house.

Laying hens: Rearing period

Every rearing flock is sampled 2 weeks before entering the production-unit.

Laying hens: Production period

Every flock is sampled every 15 weeks starting at the age of 24 weeks.

Laying hens: Before slaughter at farm

Every flock is sampled within the 3 weeks before slaughter.

Laying hens: At slaughter

Sampling is distributed evenly throughout the year.

Type of specimen taken

Laying hens: Day-old chicks

The sample taken of day-old chicks is a mixed sample of Internal linings of the delivery boxes.

Laying hens: Rearing period

The samples taken during rearing consist of overshoes in accordance with Regulation (EU) N 517/2011.

Laying hens: Production period

The samples taken during production consist of 2 pair of overshoes in accordance with Regulation (EU) N° 517/2011. Offical samples consist of 2 pair of overshoes and one dustsample, also in accordance with Regulation (EU) N 517/2011.

Laying hens: Before slaughter at farm

The samples taken during production consist of 2 pair of overshoes in accordance with Regulation (EU) N° 517/2011.

Laying hens: At slaughter

A mixed sample consisting of 10 caecal samples is taken at slaughter.

Methods of sampling (description of sampling techniques)

Laying hens: Day-old chicks

At the farm, 20 pieces (5 by 5 cm) of the inner linings of delivery boxes are taken of each batch. The samples have to reach an accredited laboratory within 48 hours of sampling.

Laying hens: Rearing period

Samples are taken in accordance with Regulation (EU) N. 517/2011.

Laying hens: Production period

Samples are taken in accordance with Regulation (EU) N. 517/2011.

Laying hens: Before slaughter at farm

Samples are taken in accordance with Regulation (EU) N. 517/2011.

Laying hens: At slaughter

A mixed sample consisting of 10 intact caecal samples is taken at the level of the slaughterline.

Case definition

Laying hens: Day-old chicks

A sample is considered positive if S. Enteritidis or S. Typhimurium is isolated. A flock is considered positive as soon as one sample is positive.

Laying hens: Rearing period

A sample is considered positive if S. Enteritidis or S. Typhimurium is isolated. A flock is considered positive as soon as one sample is positive.

Laying hens: Production period

A sample is considered positive if S. Enteritidis or S. Typhimurium is isolated. A flock is considered positive as soon as one sample is positive.

Laying hens: Before slaughter at farm

A sample is considered positive if S. Enteritidis or S. Typhimurium is isolated. A flock is considered positive as soon as one sample is positive.

Diagnostic/analytical methods used

Laying hens: Day-old chicks

The method used is the bacteriological method: ISO 6579:2002 annex D in accordance with Regulation (EU) N° 517/2011. All isolates are serotyped according to the Kauffmann-White-LeMinor Scheme.

Laying hens: Rearing period

The method used is the bacteriological method: ISO 6579:2002 annex D in accordance with Regulation (EU) N° 517/2011. All isolates are serotyped according to the Kauffmann-White-LeMinor Scheme.

Laying hens: Production period

The method used is the bacteriological method: ISO 6579:2002 annex D in accordance with Regulation (EU) N° 517/2011. All isolates are serotyped according to the Kauffmann-White-LeMinor Scheme.

Laying hens: Before slaughter at farm

The method used is the bacteriological method: ISO 6579:2002 annex D in accordance with Regulation (EU) N° 517/2011. All isolates are serotyped according to the Kauffmann-White-LeMinor Scheme.

Laying hens: At slaughter

The method used is the bacteriological method: ISO 6579:2002 annex D.

Vaccination policy

Laying hens flocks

All laying hen flocks in production must be vaccinated against Salmonella Enteritidis. The vaccination against Salmonella Typhimurium is strongly recommended.

Other preventive measures than vaccination in place

Laying hens flocks

Minimal requirements for infrastructure, management, hygiene and bio-security issues are laid down in the framework of the authorization of holdings.

Control program/mechanisms

The control program/strategies in place

Laying hens flocks

The national control programme for Salmonella in laying hens is based on Regulations (EC) Nos. 2160/2003, 1177/2006 and (EU) No. 517/2011.

Measures in case of the positive findings or single cases

Laying hens flocks

In case of positive findings, following measures are implemented: 1) Pasteurization of eggs before human consumption. 2) Cleaning and disinfection of housing after removal of the positive flock. 3) Swab sampling of housing before entering a new flock. If the result is positive for Salmonella, cleaning, disinfection and swabcontrol has to be repeated before restocking the house.

Notification system in place

Zoonotic Salmonella is notifiable by the farmer and the laboratory since the first of January 2004. Notification is done by phone, fax or electronically to the Federal Agency for the Safety of the Food Chain.

Results of the investigation

253 different batches of day-old chicks were tested. Salmonella was not found. During rearing, 286 flocks were sampled of which 2 were positive for Salmonella spp. (1 S. Idikan, 1 S; Kedougou). During production, 655 flocks were sampled of which 30 were positive for Salmonella spp. (2 for S. Enteritidis).

National evaluation of the recent situation, the trends and sources of infection

During rearing, the prevalence increased from 0,36% in 2013 to 1,35% in 2014 to 1,86% in 2015. In 2016 a decrease was seen in the prevalence of Salmonella spp. to 0,7%. This decrease in prevalence is also noticed during production where a slight decrease from 5,17% in 2015 to 4,57% in 2016 is seen. The prevalence of Salmonella Enteritidis and Salmonella Tyhimurium has consistently decreased from 2% in 2014 to 0,30% in 2016.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

After 3 years of increase (2.760 in 2013, 2.963 in 2014, 3.119 in 2015) in the total number of reported human Salmonella isolates, a slight decrease was seen in 2016 (3.026). The number of S. Typhimurium isolates decreased from 1.856 in 2015 to 1.412 in 2016 but the number of S. Enteritidis isolates increased from 458 in 2015 to 600 in 2016. An increase in the number of isolates belonging to other serotypes was also seen.

3.1.2.3 Salmonella in animal - Gallus gallus (fowl) - breeding flocks, unspecified - Farm - environmental sample - boot swabs - Control and eradication programmes - Official and industry sampling - Census

Monitoring system

Sampling strategy

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

Breeding flocks are sampled as day-old chicks, at the age of 4 and 16 weeks and every 2 weeks during production. An official control takes place at 16 weeks, 22 weeks, 46 weeks and 58 or 62 weeks. A specific Salmonella control is performed 4 times a year in the hatcheries by the owner.

Frequency of the sampling

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

Every flock is sampled.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

During rearing, breeding flocks are sampled as day old chicks and at the age of 4 and 16 weeks.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period

Adult breeding flocks are sampled every 2 weeks.

Type of specimen taken

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

The samples taken from day-old chicks are pieces of internal linings of delivery boxes at time delivery.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

During rearing, boot swab samples are taken in accordance with the sampling method described in Regulation (EU) N° 200/2010.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period

During the production period, sampling is also performed using boot swabs in accordance with the provisions of Regulation (EU) N° 200/2010.

Methods of sampling (description of sampling techniques)

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

At the farm, pieces (5 by 5 cm) of the inner linings of delivery boxes are taken of each flock by the farmer. 2 samples are taken, one for the hen-chicks and one for the cock-chicks. Each sample consists of 20 pieces of interlining. The two samples are analyzed separately. The samples have to be taken the day of delivery, the samples have to reach the lab within 24 hours of sampling. In the hatcheries, pooled samples from dead-in-the-shell chicks and of fluff and meconium, are taken by the owner every 3 months. These are sent to the laboratory of one of the animal health associations.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

Samples are taken by the owner at 4 weeks by the industry and official samples are taken by one of the animal health associations at 16 weeks, both in accordance with regulation (EU) Nr. 200/2010.

Breeding flocks: Production period

All samples are taken in accordance with Regulation (EU) Nr. 200/2010.

Case definition

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

A sample is considered positive if Salmonella Enteritidis, Typhimurium, Hadar, Infantis, Virchow or Paratyphi B var. Java is isolated. A flock is considered positive as soon as one sample is positive.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

A sample is considered positive if Salmonella Enteritidis, Typhimurium, Hadar, Infantis, Virchow or Paratyhpi B var. Java is isolated. A flock is considered positive as soon as one sample is positive. If the farmer requests a confirmation sampling, new samples (5 feces and 2 dust samples for the detection of Salmonella and muscles samples for the detection of the use of antibiotics) are taken by the competent authority or delegated to one of the animal health associations. The flock is considered positive if one of the 6 serotypes of Salmonella is found or if one of the tests on the use of antibiotics is positive.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period

A sample is considered positive if Salmonella Enteritidis, Typhimurium, Hadar, Infantis, Virchow or Paratyhpi B var. Java is isolated. A flock is considered positive as soon as one sample is positive. If the farmer requests a confirmation sampling, new samples (5 feces and 2 dust samples for the detection of Salmonella and muscles samples for the detection of the use of antibiotics) are taken by the competent authority or delegated to one of the animal health associations. The flock is considered positive if one of the 6 serotypes of Salmonella is found or if one of the tests on the use of antibiotics is positive.

Diagnostic/analytical methods used

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

The bacteriological method: ISO 6579:2002 annex D is used, in accordance with Regulation (EU) Nr. 200/2010 is used. Al isolates are serotyped by the Kauffmann-White-LeMinor scheme.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

The bacteriological method: ISO 6579:2002 annex D is used, in accordance with Regulation (EU) Nr. 200/2010 is used. Al isolates are serotyped by the Kauffmann-White-LeMinor scheme.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period

The bacteriological method: ISO 6579:2002 annex D is used, in accordance with Regulation (EU) Nr. 200/2010. Al isolates are serotyped by the Kauffmann-White-LeMinor scheme.

Vaccination policy

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

Vaccination against Salmonella Enteritidis is compulsory for parent breeding flocks and prohibited for grand parent flocks. Vaccination against Salmonella Typhimurium is strongly recommended for parent breeding flocks and prohibited for grandparent flocks.

Other preventive measures than vaccination in place

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

All holdings with breeding flocks must implement minimum requirement for infrastructure, management, hygiene and biosecurity.

Control program/mechanisms

The control program/strategies in place

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

The Belgian national control programme for Salmonella in breeding flocks is based on Regulations (EG) Nr. 2160/2003 and 1177/2006 and Regulation (EU) nr. 200/2010.

Measures in case of the positive findings or single cases

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

Following measures are implementen in the framework of the Salmonella control programme in breeders: 1) treatment of flock with antimicrobials is forbidden; 2) Incubation of hatching eggs is prohibited; 3) Incubated hatching eggs are removed and destroyed; 4) Not yet incubated hatching eggs may be pasteurized and put on the market for human consumption; 5) Positive breeding flocks are slaughtered within the month; 6) Cleaning and disinfection of housing is mandatory after removal of the breeding flock; 7) After cleaning and disinfection, a hygienogram and the sampling of the house for the detection of Salmonella is performed by one of the animal health associations; 8) A new flock is admitted if Salmonella can not be found after cleaning and disinfection, otherwise the disinfection and swab control is repeated.

Notification system in place

Zoonotic Salmonella is notifiable since the first of January 2004. Notification is done by phone, fax or electronically to the Federal Agency for the Safety of the Food Chain. Laboratories and farmers are submitted to the notification.

Results of the investigation

Salmonella was not found in day old chicks (174 batches) and during rearing (331 flocks). During production, of the 565 flocks, two flocks were positive for S. Enteritidis and 14 flocks were positive for serotypes not included in the programme. In addition, two flock were considered negative for S. Typhimurium and one for S. Infantis after confirmation sampling. These three flocks do not count as positive flocks.

National evaluation of the recent situation, the trends and sources of infection

During production, the number of positive flocks for Salmonella serotypes for which a target is set fluctuates between 0 and 3 in recent years. In 2014, 5 positive flocks were found, in 2015 and 2016 only 2. The number of positive flocks of other serotypes remains about the same as previous years (12 in 2015, 16 in 2014).

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

After 3 years of increase (2.760 in 2013, 2.963 in 2014, 3.119 in 2015) in the total number of reported human Salmonella isolates, a slight decrease was seen in 2016 (3.026). The number of S. Typhimurium isolates decreased from 1.856 in 2015 to 1.412 in 2016 but the number of S. Enteritidis isolates increased from 458 in 2015 to 600 in 2016. An increase in the number of isolates belonging to other serotypes was also seen.

3.1.2.4 Salmonella in Turkeys - breeding flocks and meat production flocks

Monitoring system

Sampling strategy

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

There are no professional breeding turkey flocks in Belgium.

Meat production flocks

All flocks are sampled within three weeks of slaughter.

Frequency of the sampling

Meat production flocks: Before slaughter at farm

Every flock is sampled

Type of specimen taken

Meat production flocks: Before slaughter at farm

All flocks are sampled using boot swabs.

Methods of sampling (description of sampling techniques)

Meat production flocks: Before slaughter at farm

All flocks are sampled, by the owner, within 3 weeks before slaughter conform Regulation (EU) n° 1190/2012.

Case definition

Meat production flocks: Before slaughter at farm

A flock is positive if Salmonella spp. is found.

Diagnostic/analytical methods used

Meat production flocks: Day-old chicks

The analytical method used is the bacteriological method: ISO 6579:2002 annex D.

Meat production flocks: Before slaughter at farm

The analytical method used is the bacteriological method: ISO 6579:2002 annex D as described in Regulation (EU) 1190/2012. Al isolates are serotyped by the Kauffmann-White-LeMinor scheme.

Vaccination policy

Meat production flocks

There is no vaccination policy for meat production flocks.

Other preventive measures than vaccination in place

Meat production flocks

In the framework of the authorization of holdings, infrastructural, management, hygiene and bio-security measures must be implemented on all holdings.

Notification system in place

Zoonotic Salmonella is notifiable since 1 January 2004. Notification is done by phone, fax or e-mail.

Results of the investigation

There are no turkey breeding flocks in Belgium. 194 meat production flocks were tested in 2016. Salmonella could not be found.

National evaluation of the recent situation, the trends and sources of infection

In contrast with previous years, Salmonella could not be found in turkey meat production flocks in 2016.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

Seen the limited number of meatturkey flocks slaughtered in Belgium, there is little to no relevance of the findings in these flocks to human cases.

3.2 CAMPYLOBACTERIOSIS

3.2.1 General evaluation of the national situation

3.2.1.1 Thermophilic Campylobacter spp., unspecified - general evaluation

National evaluation of the recent situation, the trends and sources of infection

3.2.2 Campylobacter in foodstuffs

3.2.2.1 Thermophilic Campylobacter spp., unspecified in food - Meat from broilers (Gallus gallus) - food sample

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Sampling strategy

At slaughterhouse and cutting plant

Campylobacter spp. contamination of broiler meat in Belgian is evaluated in slaughterhouses and cutting plants. Campylobacter is counted on carcasses and cuts of poultry.

Frequency of the sampling

At slaughterhouse and cutting plant

Sampling distributed evenly throughout the year.

At meat processing plant

Sampling distributed evenly throughout the year.

At retail

Sampling distributed evenly throughout the year.

Type of specimen taken

At slaughterhouse and cutting plant

Neck skin samples and cuts of broilers with and without skin.

At meat processing plant

Meat, minced meat, sausages and other.

At retail

Meat, minced meat, meat preparations.

Methods of sampling (description of sampling techniques)

At slaughterhouse and cutting plant

The matrices were carcasses, cuts and meat preparation of broilers. The Campylobacter spp. contamination levels were analyzed in 1g carcasses, 1g cutting meat and 1g meat preparation.

At meat processing plant

The samples contained about 200 g of meat. The amount of Campylobacter has been assessed in 1g of sample.

At retail

The amount of Campylobacter spp. has been assessed in 1g of sample.

Definition of positive finding

At slaughterhouse and cutting plant

A sample is considered positive in case of detection of more than 1.000 cfu/g Campylobacter for carcases and meat with skin and in case of detection of more than 100 cfu/g Campylobacter for meat without skin.

At meat processing plant

A sample is considered positive in case of detection of more than 100 cfu/g Campylobacter in the sample (1.000 cfu for carcases and meat with skin).

At retail

A sample is considered positive in case of detection of more than 100 cfu/g Campylobacter in the sample (1.000 cfu for carcases and meat with skin).

Diagnostic/analytical methods used

At slaughterhouse and cutting plant

detectie: ISO 10272-1:2006 CAMPYLOBACTER; telling: ISO/TS 10272-2:2006 CAMPYLOBACTER Antimicrobial susceptibility testing of C. jejuni isolates in accordance with Decision 2013/652/EU

3.2.2.2 Thermophilic Campylobacter spp., unspecified in food - All foodstuffs - food sample

Monitoring system

Sampling strategy

carcasses and meat are sampled, as well as some RTE foods like raw milk cheeses.

Frequency of the sampling

samples are taken throughout the year.

Type of specimen taken

meat and dairy products

Methods of sampling (description of sampling techniques)

Sampling of bovine carcasses was done by means of swabs (4 areas from the same half carcass constituting 1600 cm2 were putted in the same stomacher bag). The carcass samples of broiler and layer consisted of 10g of neck skin. The other samples were about 200g of meat. 10g to 25g representative of the whole sample were weighted in the laboratory, and the detection of Campylobacter has been assessed in these quantities or dilutions: 25g for pork minced meat, 1600 cm2 (bovine carcasses), 0,01g for chicken carcasses and layer carcasses, 1g for chicken meat preparation, and for chicken cuts, 0,1g and 25g. No pooling has been done.

Definition of positive finding

FASFC has established action limits. For RTE products the limit is less than10 cfu/g. For poultry with skin the limit is 1000 cfu/g. For poulltry without skin the limit is 1000 cfu/g.

Diagnostic/analytical methods used

Detection: Microbiology of food and animal feeding stuffs, Horizontal method for detection and enumeration of Campylobacter spp. Part 1: Detection method Enumeration: Microbiology of food and animal feeding stuffs, Horizontal method for detection and enumeration of Campylobacter spp. Part 2: Colony-count technique.

Measures in case of the positive findings or single cases

Measures to be taken in the case of a non-compliant result:- Notification of the producer or importer- Possibility of a counter analysis- Destruction of the non compliant batch if it is an RTE product. Further investigation: additional sampling, possible recall, RASFF. If it concerns NRTE food: hygiene measures must be taken.

3.2.3 Campylobacter in animals

3.2.3.1 Thermophilic Campylobacter spp., unspecified in animal - Gallus gallus (fowl) - animal sample - caecum

Monitoring system

Frequency of the sampling

At slaughter

Sampling distributed evenly throughout the year.

Type of specimen taken

At slaughter

The intact caeca of 10 birds from the same flock are taken at the slaughterhouse with the aim to determine the load of Campylobacter spp. entering the slaughterhouse.

Methods of sampling (description of sampling techniques)

At slaughter

10 caeca pairs are pooled to one sample. The caeca are emptied at the laboratory. The content is examined for Cambylobacter spp.

Case definition

At slaughter

A sample is positive if Campylobacter spp. is detected.

Measures in case of the positive findings or single cases

Samples are taken for monitoring purposes only. No measures are taken in case of positive findings. The producer gets a warning and hygiene measures must be taken.

3.3 LISTERIOSIS

3.3.1 General evaluation of the national situation

3.3.1.1 Listeria - general evaluation

National evaluation of the recent situation, the trends and sources of infection

The prevalence of Listeria monocytogenes in food has not changed in comparaison to previous years. A trend analysis was performed on the data from 2012 to 2015 in some foodstuffs, namely: milk and products thereof, eggs and products thereof, meat and products thereof. No trend could be observed in these products. However, the number of reported cases of listerioses has raised. In 2016 the highest number of reported liseriose case ever has been observed. Mostly people with cancer, immunodepression, ... get infected. The NRC has made an analysis of the cases from recent years: Diversity of Listeria monocytogenes Strains of Clinical and Food Chain Origins in Belgium between 1985 and 2014 S. Bertrand1*, P. J. Ceyssens1, M. Yde1, K. Dierick2, F. Boyen3, J. Vanderpas4, R. Vanhoof1, W. Mattheus1.

Recent actions taken to control the zoonoses

General food hygiene rules are essential for the prevention of human listeriosis. As some persons are at high risk (pregnant women), they are advised not to eat certain categories of food with proven elevated risk of Listeria monocytogenes contamination, such as unpasteurized milk and butter, soft cheeses and ice cream made from unpasteurized milk, any soft cheese crust, smoked fish, pat, cooked ham, salami, cooked meat in jelly, raw minced meat from beef, pork and poultry, steak tartar, raw fish and shellfish (oysters, mussels, shrimps), fish, meat and surimi salads, insufficiently rinsed raw vegetables, unpeeled fruit. Infection of pregnant women has significantly decreased due to the prevention campaign. The Superior Health Councel and the Scientific Committee of the BFSA have published some dietairy recommendations for people vulnerable for listeriosis: NL: http://www.favv-afsca.fgov.be/wetenschappelijkcomite/adviezen/2016_documents/Advies21-2016_SciCom2016-12Listeriose.pdf, FR: http://www.favv-afsca.fgov.be/comitescientifique/avis/2016/_documents/Avis21-2016_SciCom2016-12_Listeriose_000.pdf

3.3.2 Listeria in foodstuffs

3.3.2.1 L. monocytogenes in food - All foodstuffs - Unspecified - food sample - Surveillance - Official sampling - Objective sampling

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Frequency of the sampling

At retail

Samples are taken according to the national control program or in the framework of RASFF, complaints or suspicion. Samples are taken along the whole food chain.

Type of specimen taken

At retail

Different kind of products susceptible to Listeria monocytogenes are sampled and analysed: soft and semi-hard (soft) cheeses, ice-creams, RTE meals, meat preparations and meat products, ...

Definition of positive finding

At the production plant

A sample is considered to be positive after confirmation of Listeria monocytogenes (detection or enumeration).

At retail

A sample is considered to be positive after confirmation of Listeria monocytogenes (enumeration).

Diagnostic/analytical methods used

At the production plant

ISO 11290-2:1998(Horizontal method for the detection and enumeration of Listeria monocytogenes -- Part 2: Enumeration method) and ISO 11290-2:1998/Amd 1:2004 (Modification of the enumeration medium)or IMMUNOFLUORENSCENCE ASSAY TESTS (IFA)

At retail

ISO 11290-2:1998(Horizontal method for the detection and enumeration of Listeria monocytogenes -- Part 2: Enumeration method) and ISO 11290-2:1998/Amd 1:2004 (Modification of the enumeration medium)

Control program/mechanisms

The control program/strategies in place

see MANCP.

Notification system in place

Notification is mandatory since 1/3/2004 (Ministerial Decree on mandatory notification in the food chain of 22/1/2004). For Listeria monocytogenes, the criterion of 100 cfu/g in ready-to-eat food put on the market may not be exceeded. Laboratories have to inform the Federal Agency for the Safety of the Food Chain in case of a positive sample.

3.4 YERSINIOSIS

3.4.1 General evaluation of the national situation

3.4.1.1 Yersinia - general evaluation

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

Carcasses and minced meat from pigs and other animals like bovine are sampled and analysed. On the data of 2012 to 2015 a trend analysis was performed. There is no trend in the data to be observed. About 10% of the samples are positive, but it especially concerns sero/biotypes not pathogenic for humans.

3.5 TRICHINELLOSIS

3.5.1 General evaluation of the national situation

3.5.1.1 Trichinella - general evaluation

History of the disease and/or infection in the country

Since 1940, the Competent Authority did organize analysis for Trichinella in pigs at the slaughterhouses. The analysis is generalized since 1991. Trichinella has not been detected in carcasses of pigs and horses produced for human consumption in Belgium. One autochthonous human outbreak, involving 4 people belonging to the same family, occurred in 1979. This outbreak was most likely caused by a home raised wild boar.

National evaluation of the recent situation, the trends and sources of infection

Trichinellosis is virtually absent in Belgian domestic livestock. Since systematic controls of pigs and horses are done at slaughter (Regulation EC N 2075/2005) no positive case were found. The last autochtonous outbreak in humans in Belgium occurred in 1979 following the consumption of meat from a home raised wild boar. At the end of 2014, Belgium experienced an outbreak of Trichinellosis, affecting 16 people. this outbreak was most likely caused by the consumption of infected imported Spanish wild boar. Increased monitoring in Belgium, during the last decade, has shown that Trichinella spp. still circulate amongst wildlife, although both the prevalence and the intensity of infections are low. EU Directive requires that also wild boars hunted in the EU for commercial purpose are examined for Trichinella. Each year about 10.000 sport-hunted wild boars are tested. The routine examination of wild boars devoted to the market has proven to be a good measure to protect the consumer against sylvatic trichinellosis. In addition, monitoring of infection through examination of sentinel animals, such as the fox, is recommended to assess the prevalence of trichinellosis and to follow trends in time. Serological examination might be an alternative for muscle digestion in screening programs, but can not be used in safeguarding consumer's health in meat inspection. An extra measure to protect the consumer is to eat meat of wild boar "well done", or to freeze the meat at -20C for 4 weeks. An important measure to avoid spreading of the infection among wildlife is not to leave offal of animal carcasses in the field during hunting.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

The last autochtonous outbreak in humans in Belgium occurred in 1979 following the consumption of meat from wild boar. At the end of 2014, Belgium experienced an outbreak of Trichinellosis, affecting 16 people. This outbreak was most likely caused by the consumption of infected imported Spanish wild boar.

Recent actions taken to control the zoonoses

Monitoring of wildlife. Routine examination of wild boar destined for human consumption. Monitoring of infection through examination of sentinel animals such as the fox was not further realized since 2013. Recommendation to consume wild boar meat only after freezing at -20C for 4 weeks. Recommendation to travellers not to import raw meat products of unknown origin and of susceptible animals, e.g. home-made sausages, and not to consume meat of unknown quality abroad.

Additional information

The status "negligible risk for Trichinella in slaughterpigs kept under industrial housing conditions" was granted by the EC to Belgium end December 2010.

3.5.2 Trichinella in animals

3.5.2.1 Trichinella in animal - Solipeds, domestic - horses - animal sample

Monitoring system

Sampling strategy

Permanent post-mortem surveillance of all slaughtered animals at the slaughterhouses or hunted animals at the game processing plants.

Frequency of the sampling

Sampling of all slaughtered animals at the slaughterhouses and hunted animals at the game processing plants.

Type of specimen taken

Diaphragm, tongue or masseter muscle.

Methods of sampling (description of sampling techniques)

Horses: 5 gram of diaphragm (or tongue, or masseter) for routine diagnosis, analyses on pooled samples; 10 to 25 gram for examination of individual samples.

Case definition

An animal is considered positive in case of detection and identification of Trichinella larvae in the muscle sample.

Diagnostic/analytical methods used

Artificial digestion method of collective or individual samples. The magnetic stirrer method for digestion of pooled samples as described in Commission Implementing Regulation (EC) No 2015/1375 was used on samples of 5 gram of muscles from horses.

Control program/mechanisms

The control program/strategies in place

Commission Implementing Regulation (EC) No 2015/1375 imposes systematic Trichinella examination of all slaughtered pigs, horses and wild boar and other wildlife animals by artificial digestion method of muscle before marketing.

Measures in case of the positive findings or single cases

Carcasses found positive are declared unfit for human consumption.

Notification system in place

Notification to the Federal Agency for the Safety of the Food Chain is compulsory for any positive test result.

Results of the investigation including the origin of the positive animals

Two positive wildboars were detected by the end of 2016 at a hunting party in the province of Luxemburg. Molecular typing indicated that one wildboar was infected with T. spiralis and one wildboar with T. britovi.

National evaluation of the recent situation, the trends and sources of infection

No positive horses were found in 2016.

3.5.2.2 Trichinella in animal - Pigs - animal sample

Officially recognised regions with negligible Trichinella risk

At the end of 2010 Belgium was granted the status of 'negligible Trichinella risk' in pig population by the European Commission

Monitoring system

Sampling strategy

General

Permanent surveillance of all slaughtered fattening and breeding pigs 'raised under controlled housing conditions' at the slaughterhouses in implementation of Commission Implementing Regulation (EC) No 2015/1375. Derogation is foreseen for fattening pigs who do apply for the criteria set in the definition 'Region with negligible risk'.

Frequency of the sampling

General

Systematic Trichinella examinations of all slaughtered fattening and breeding pigs raised under controlled housing conditions, with the exception of some fattening pigs who do apply for the criteria set in the definition 'Region with negligible risk'.

For regions with negligible Trichinella risk

Systematic Trichinella examinations of all slaughtered pigs, with the exception of some fattening pigs who do apply for the criteria set in the definition 'Region with negligible risk'.

Type of specimen taken

General

Diaphragm muscle, 1 gram for fattening pigs, 2 grams for sows and boars.

For regions with negligible Trichinella risk

Diaphragm muscle, 1 gram for fattening pigs, 2 grams for sows and boars. No samples are examined of fattening pigs who do apply to the criteria set in the definition of 'Region with negligible risk'.

Methods of sampling (description of sampling techniques)

General

Fattening pigs: 1 gram of diaphragm muscle to be pooled (up to 100 animals in 1 pool). Sows and boars: 2 grams of diaphragm muscle to be pooled (up to 50 animals in 1 pool).

For regions with negligible Trichinella risk

Still almost all pigs are sampled and tested, because of logistic reasons and possible export outside EU.

Case definition

General

An animal is considered positive in case of detection and identification of Trichinella larvae in a muscle sample by the reference method of detection (magnetic stirrer method for pooled sample digestion). Confirmation of positive results by the digestion method can be done by PCR in the National Reference Laboratory on Trichinellosis.

For regions with negligible Trichinella risk

An animal is considered positive in case of detection and identification of Trichinella larvae in a muscle sample.

Diagnostic/analytical methods used

General

Artificial digestion method of collected samples and Magnetic stirrer method for pooled samples: sedimentation or on filter isolation technique and larvae detection by a latex agglutination test (equivalent method). The analysis is done by artificial digestion: the magnetic stirrer method of pooled 100 gram sample as described in Commission Implementing Regulation (EC) No 2015/1375, reference method, 1 gram per fattening pig, 2 grams per sow and boar, and 5 grams per horse and per wild boar. Serology may be done for epidemiological studies in live pigs and for monitoring of wildlife. Confirmation of positive results by the digestion method can be done by PCR in the National Reference Laboratory on Trichinellosis.

Measures in case of the positive findings or single cases

Carcasses found positive are declared unfit for human consumption.

Notification system in place

Trichinellosis is notifiable for all susceptible animal species. No positive cases were found in 2016 in domestic fattening and breeding pigs. One positive case of Trichinella spiralis and one case of Trichinella britovi were found by the end of 2016 in two wildboars during the hunting party in the province of Luxemburg.

Notification system in place

Notification to the Federal Agency for the Safety of the Food chain is compulsory for any positive test result.

Results of the investigation including description of the positive cases and the verification of the Trichinella species

Fattening pigs raised under controlled housing conditions in integrated production system

One positive case of Trichinella spiralis and one positive case of Trichinella britovi were detected in 2016 on two wildboars during the hunting season. These animals were found positive by the reference method of detection and this positive result was confirmed by PCR at the NRL of Trichinellosis.

Fattening pigs not raised under controlled housing conditions in integrated production system

No positive cases were found in 2016 in slaughtered fattening pigs raised under controlled housing conditions.

Breeding sows and boars

No positive cases were found in 2016 in slaughtered breeding pigs raised under controlled housing conditions.

National evaluation of the recent situation, the trends and sources of infection

Since 1992, when the European Union Council Directive required that wild boars (Sus scrofa) hunted in EU for commercial purpose should be examined for Trichinella, the infection has only been detected a limited number of times in wild boars of Belgium. There is serological evidence of the presence of anti-Trichinella antibodies in wildlife.

3.6 ECHINOCOCCOSIS

3.6.1 General evaluation of the national situation

3.6.1.1 Echinococcus - general evaluation

History of the disease and/or infection in the country

At the slaughterhouses, a small number of carcasses showing lesions of Echinococcus (cysts) are from time to time detected and notified to the Federal Agency for the Safety of the Food Chain. In case of positive findings, depending on the extent of the lesions, carcasses are partially or totally rejected and declared unfit for human consumption. In 2016 only one case was detected on a bovine carcass.

National evaluation of the recent situation, the trends and sources of infection

Echinococcosis is caused either by Echinococcus granulosus or Echinococcus multilocularis. Echinococcus granulosus produces unilocular human hydatidosis. The adult stage is a small tapeworm (6 mm) that lives in the small intestine of domestic and wild canids. Sheep and cattle serve as intermediate hosts for the infection. Humans acquire infection by ingestion of typical taeniid eggs, which are excreted in the faeces of infected dogs: the oncospheres liberated from the eggs migrate via the bloodstream to the liver, lungs and other tissues to develop in hydatid cysts. Indigenous unilocular hydatidosis in man has been reported in Belgium. In 2016 no cysts were found by post-mortem inspection of the carcasses at the slaughterhouses. Echinococcus multilocularis causes alveolar (multilocular) echinococcosis in humans. Foxes and dogs are the definitive hosts of this parasite and small rodents the intermediate hosts. In the liver of rodents the invasive larval stage has a multi-compartimented appearance containing many protoscolices. Ingestion of the eggs by humans can result in the development of invasive cysts in the liver. In Belgium, the percentage of infected foxes varies with the region, with a decreasing rate from the South-East to the North-West: e.g 33% in the Ardennes, 13% in the Condroz region and 1-2% in Flanders Region. The endemic region is situated under the river Meuse, on the heights of the Ardennes in the Walloon Region.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

Post mortem visual examination is performed at the slaughterhouses in the domestic intermediate hosts: cattle, sheep, horses and pigs. Whole carcasses or parts are rejected in case Echinococcus granulosus cysts are found.

Recent actions taken to control the zoonoses

Consumption of berries is discouraged by warning messages, displayed to visitors of Parks and Woodlands.

3.7 RABIES

3.7.1 General evaluation of the national situation

3.7.1.1 Lyssavirus (rabies) - general evaluation

History of the disease and/or infection in the country

Since the last indigenously acquired case of rabies occurred in Belgium in a bovine coming from Bastogne (province of Luxembourg) in July 1999, Belgium obtained the official status of rabies-free country in July 2001 according to the WHO recommendations (1992) and the Office Internationale des Epizooties (OIE) guidelines (1997).

National evaluation of the recent situation, the trends and sources of infection

In October 2007, Belgium lost temporary its official status of rabies free country due to a positive case of rabies in a dog, illegally imported from Morocco. Belgium regained again its official free status of rabies on 28 October 2008.

Recent actions taken to control the zoonoses

Surveillance system and methods used. Domestic animals with nervous symptoms suspected of rabies have to be notified to the Federal Agency for the Safety of the Food chain. Wildlife found dead or shot should also be notified and send for analysis to the Scientific Institute of Public Health, the National Reference laboratory of rabies. Collection of dead-found bats is recommended for rabies surveillance. Live suspected animals are killed and their brain is examined by immunofluorescence and virus cultivation in neuroblasts at the Scientific Institute of Public Health. The high percentage of examinations of cattle is in consequence of the surveillance system for TSE in cattle: all suspected BSE cases were first examined for rabies. Rabies must be considered in the differential diagnosis of BSE, although the clinical course of rabies is usually quicker than the evolution of clinical nervous symptoms in case of BSE. The oral vaccination campaign of foxes with vaccine baits started in 1989 and was stopped by the end of 2003. Regional vaccination in the South of Belgium of dogs and cats is no more compulsory since the 1st March 2016.

Suggestions to the European Union for the actions to be taken

It is highly recommended to report on the rabies virus type detected to be able to differentiate between the classical rabies type (genotype 1) and the European bat Lyssa virus types (unspecified or EBL 1 or EBL 2). Bat rabies is of public health concern. The public should be made aware of the danger of human exposure to bats, especially in case of abnormal behavior of bats. Rabies is transmitted to humans and other animals through saliva, usually by a bite. Any person exposed to bats should be previously vaccinated against rabies. Nobody should handle diseased or dead bats without protection such as gloves. Any person finding a bat behaving abnormally, in an unusual place, or under unusual circumstances, should not attempt to handle or to move the animal but should contact official authority. Education and recommendations should be given to travelers in order to reduce their risk of infection. Although dogs represent a more serious threat in many countries, yet the risk of rabies infection by bat bites also exists. Pre-exposure vaccination should be offered to persons at risk, such as laboratory workers, veterinarians, animal handlers, international travelers. Currently available vaccines are safe and effective against both the classical rabies virus and the bat Lyssa viruses.

3.7.2 Lyssavirus (rabies) in animals

3.7.2.1 Lyssavirus (rabies) in animal - All animals - animal sample

Monitoring system

Sampling strategy

The brain of animals with nervous symptoms suspected of rabies are examined by direct immunofluorescence test and virus cultivation in neuroblasts at the Scientific Institute of Public Health, the National Reference Laboratory for rabies.

Frequency of the sampling

All suspected animals with clinical nervous symptoms are tested.

Type of specimen taken

Brain tissues

Methods of sampling (description of sampling techniques)

Small animals: head / carcass. Huge animals: brain (CNS). Shipping and packaging conditions: brains are transported as soon as possible (refrigerated if possible) in a tightly sealed packet to the National Reference Laboratory. In case of transport of a carcass, an authorization is required. The storage period of samples at the National Reference Laboratory for further analysis is one year.

Case definition

An animal is considered infected in case of a positive direct immunofluorescence test (Antigen detection) confirmed by cell cultivation of the virus or detection by RT-PCR or (rarely performed) by mice inoculation test (clinical observation of rabies symptoms).

Diagnostic/analytical methods used

Direct immunofluorescence for the detection of viral antigen, virus isolation in neuroblastoma cell culture, detection by RT-PCR, mouse inoculation test

Vaccination policy

In the Southern part of the country, below the rivers Sambre and Meuse, vaccination of dogs and cats was compulsory during many years. In addition, all pets staying on any Belgian public camping had to be vaccinated. This mandatory vaccination of dogs and cats was stopped on the 1st of March 2016 as there were no more cases of rabies for many years. Oral vaccination of foxes by baits started in 1989 and was stopped by the end of 2003.

In case of positive findings national legislation has to be applied (Royal Decree of 18 September 2016).

Notification system in place

Royal Decree of 18 September 2016, Animal Health Law of 24 March 1987 Chapter III and Royal Decree of 3 February 2014 (list of all notifiable animal diseases). Notification of all laboratory confirmed cases to the competent Authority is mandatory.

3.8 STAPHYLOCOCCUS AUREUS METICILLIN RESISTANT (MRSA) INFECTION

3.8.1 Staphylococcus in animals

3.8.1.1 S. aureus, meticillin resistant (MRSA) in animal - Pigs - Farm - animal sample - nasal swab - Monitoring - Official sampling - Objective sampling

Monitoring system

Sampling strategy

In 2016, the monitoring of the presence of MRSA and its antimicrobial resistance was performed in fattening and breeding pigs. The number of units to be sampled was based on the results of the same monitoring in 2013.

Frequency of the sampling

The samples are programmed to be taken evenly divided over the year and over the two categories. A mixed sample is taken of at least 10 animals on each farm. The monitoring is repeated every 3 years.

Type of specimen taken

10 nasal swabs were taken on each holding and pooled to one sample. If breeding and fattening pigs were present on the farm, both categories could be sampled. Samples taken from fattening pigs were taken within two months before slaughter.

Methods of sampling (description of sampling techniques)

10 nasal swabs are taken from one nostril from 10 different animals. Each swab is transported in its own transportation tube. The swabs are pooled at the level of the laboratory to one sample. The swabs are stored between 5°C and 25°C.

Case definition

A holding is positive when MRSA is detected and confirmed by PCR.

Diagnostic/analytical methods used

Pooled samples were incubated in Mueller-Hinton (MH) broth (Becton Dickinson) supplemented with NaCl (6.5%) at 37C for 18-24h. One ml of this broth was added to Tryptic Soy Broth (TSB) supplemented with cefoxitin (3.5 mg/l) and aztreonam (75 mg/l) and incubated at 37C for 18-24h. Ten microliter of this enrichment was plated on Brilliance MRSA 2 (Oxoid) and incubated 18-24h at 37C. Presence of MRSA was suspected based on colony morphology. Per sample, one to five suspected colonies were selected from the Brilliance MRSA 2 plate. Presence of MRSA was confirmed using a triplex real-time PCR method. Per sample, one to five suspected colonies were selected from the Brilliance MRSA 2 plate. DNA was extracted as described in SOP/BAC/ANA/18. The PCR allows detecting the Staphylococcal aureus specific gene, nuc, the presence of the mecA gene responsible for methicillin resistance and the variant mecC gene. All MRSA isolates were spatyped by sequencing the repetitive region of the spa gene encoding for the staphylococcal protein A. This method depicts the rapid evolution, since through recombination, the repeats may change fast. The protein A (spa) gene was amplified according to the Ridom StaphType standard protocol (www.ridom.de/staphtype) and the amplification was checked on a 2% agarose gel. Sequencing was performed with CEQ 8000 using standard protocols and sequences were compared with the international Ridom database. CC398 PCR was performed on all MRSA following protocol described by Stegger et al. 2011. This method allows the rapid detection of the S. aureus sequence type ST398.

Measures in case of the positive findings or single cases

There are no measures linked to positive findings. However, farmers are informed of the presence of MRSA on the holding and on possible measures to protect themselves. General hygiene and biosecurity measures are promoted.

National evaluation of the recent situation, the trends and sources of infection

The presence of MRSA was confirmed for 203 strains out of the 330 analyzed samples (61.5%), based on real-time PCR. MRSA was present in both fattening pigs (63.3%) and sows (59.5%). The MRSA prevalence in fattening pigs and sows in 2016 is very similar to the prevalence in 2013 (overall prevalence = 65.6%). Among 175 MRSA strains recovered, 141 (80.6%) were positive for the cc398 PCR and considered as MRSA sequence type ST398. Out of the 199 MRSA strains, 175 were characterized by their genotype (spa-typing and CC398 PCR). Hundred forty-one strains were MRSA ST398. Nine different spa-types were found. The vast majority was however the commonly isolated t011 and all of them were associated with MRSA ST398. Amongst the ST398 strains, 6 different spa types were found. In 2013, still 12 other spa-types were recovered from pigs. A change in spa-types reflects adaptations of MRSA to its host and might indicate that host adaptations are underway. Thirty-four MRSA strains were different from MRSA ST398. Among these MRSA strains the following spa-types were found: t034, t037, t898, t1451, t1456, t1580 and t1985. As expected, all MRSA strains were resistant to cefoxitin and penicillin, except for 9 strains which showed susceptibility to cefotoxin. This should be regarded as a methodological deviation since the presence of the mecA gene was demonstrated.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

MRSA prevalence in pigs is higher than in poultry, bovines for meat and dairy cattle .

3.9 Q-FEVER

3.9.1 Coxiella (Q-fever) in animals

3.9.1.1 Coxiella (Q-fever) in animal - Cattle (bovine animals) - adult cattle over 2 years - Farm - animal sample

Monitoring system

Sampling strategy

There is no monitoring system in place for Coxiella burnetii on cattle farms. In case of abortion, Coxiella burnetii is part of the differential diagnosis and further examination is recommended but not mandatory.

Type of specimen taken

Animals at farm

In case of abortion, different samples are taken: vaginal swabs, foeti, bulk milk, blood, organs and placenta.

Case definition

Animals at farm

The herd is considered positive when the sample of at least one animal is positive on RT-PCR.

Diagnostic/analytical methods used

Animals at farm

Either the RT-PCR or ELISA are used as analytical method.

Measures in case of the positive findings or single cases

The farmer is informed of the positive result and advise on the prevention of the spread of Coxiella burnetii to other animals and visitors/workers on the farm is given.

Notification system in place

It is mandatory for recognized laboratories, veterinarians and farmers to notify positive results to the Federal Agency for the Safety of the Food Chain.

Results of the investigation

In 2016, after abortion, 4.096 samples (vaginal swabs, foeti, placenta and organs) were analyzed by RT-PCR of which 156 were positive (3,8%).

National evaluation of the recent situation, the trends and sources of infection

An increase is seen in the number of positive samples (RT-PCR) in the case of abortion compared to 2015 (2,1%). Contrary to 2015, the monitoring was not mandatory in 2016.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

Q-fever circulates on cattle farms with little to no effect on public health.

3.9.1.2 Coxiella (Q-fever) in animal - Sheep and goats - Farm - animal sample

Monitoring system

Sampling strategy

The monitoring of milkgoats and milkewes consists of bulk milk samples taken every 2 months analyzed by PCR and ELISA for the presence of Coxiella burnetii or antibodies against Coxiella burnetii. In case of abortion, samples must be sent to a laboratory of an animal health association.

Sampling strategy

Animals at farm

For the monitoring (census sampling) of all farms with milkewes and milkgoats, bulk milk samples are taken every 2 months, either by the Federal Agency for the Safety of the Food Chain or by the Milk Control Center. For the passive monitoring of Coxiella on all farms with goats and sheep, samples must be taken when an abortion took place and submitted for further investigation to one of the animal health associations. Analyses in the framework of clinical investigation take place on demand of the farm veterinarian when Coxiella is suspected. This is not a mandatory investigation.

Frequency of the sampling

Animals at farm

Bulk milk samples are taken every 2 months on all farms with milkewes and milkgoats. Sampling in case of abortion is performed after every abortion.

Type of specimen taken

Animals at farm

10 ml of bulk milk is taken at the level of the farm, frozen and sent to the national reference laboratory for further analyses. In case of abortion, a blood sample of the animal that aborted and a sample of the foetus or placenta or a vaginal swab are taken for further analyses. In case of clinical investigation, the sampler decides on the type of sample taken.

Case definition

Animals at farm

The herd is considered positive when the RT-PCR result of a sample of at least one animal is positive.

Diagnostic/analytical methods used

Animals at farm

Bulk milk is analyzed by RT-PCR and ELISA. Samples taken in case of abortion are also analyzed by RT-PCR, the blood sample of the mother animal by ELISA. In case of clinical investigation, the method of analysis is chosen by the sampler.

Vaccination policy

Vaccination is mandatory on farms with milkewes and milkgoats in case of positive RT-PCR. Voluntary vaccination is admitted.

Measures in case of the positive findings or single cases

Following measures are taken when a herd is positive for Coxiella burnetii: 1. mandatory pasteurization of the milk; 2. restricted contacts with the farm and the animals; 3. animals that aborted must be kept in quarantine and can only leave the farm for slaughter for a period of 30 days; 4. the houses where infected animals were present must be cleaned and disinfected after depopulation; 5. when animals are sold, the buyer must be informed about the presence of Coxiella burnetii on the farm.

Notification system in place

It is mandatory for the laboratories recognized by the Federal Agency for the Safety of the Food Chain, veterinarians and farmers to report all positive results. Farmers must notify their veterinarian in case of abortion. The veterinarian notifies an animal health association which on their turn, notify the Federal Agency for the Safety of the Food Chain in case of a positive results. All results are reported by the recognized laboratories on demand of the Federal Agency for the Safety of the Food Chain.

Results of the investigation

In 2016, 118 farms with milkgoats and 21 farms with milkewes were monitored of which respectively 15 and 1 were positive for Coxiella burnetii. In sheep, 47 foeti/placenta/vaginal swabs were examined after abortion of which 2 were positive for Coxiella burnetii. In goats, only 16 samples were taken of which one was positive.

National evaluation of the recent situation, the trends and sources of infection

The situation is stable compared to previous years.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

So far, the link between a human case and a positive herd has not been established. Infected farmers are not incuded.

3.10 CYSTICERCOSIS, TAENIOSIS

3.10.1 General evaluation of the national situation

3.10.1.1 Cysticerci - general evaluation

History of the disease and/or infection in the country

Cattle: Taenia saginata: 2002 in total 3.336 (3.317 lightly, 18 heavily contaminated); 2003 in total 3.886 (3.859 lightly, 25 heavily contaminated); 2004 in total 3.002 (2.981 lightly, 21 heavily contaminated); 2005 in total 2.392 (2.376 lightly, 16 heavily contaminated); 2006 in total 1.824 (1.796 lightly, 28 heavily contaminated); 2007 in total 1.527 (1.517 lightly, 10 heavily contaminated); 2008 in total 2.374 (2.356 lightly, 18 heavily contaminated); 2009 in total 1.820 (1.811 lightly, 9 heavily contaminated); 2010 in total 1.766 (1.756 lightly, 10 heavily contaminated); 2011 in total 1.347 (1.336 lightly, 11 heavily contaminated); 2012 in total 1.214 (1.205 lightly, 9 heavily contaminated); 2013 in total 994 (978 lightly, 16 heavily contaminated); 2014 in total 1.172 (1.154 lightly, 18 heavily contaminated); 2015 in total 1.253 (1,242 lightly, 11 heavily contaminated) and in 2016 in total 2182 (1262 lightly, 20 heavily contaminated carcasses). Pigs The Belgian pig population is free of Cysticercus cellulosae. Taenia solium (and Cysticercus cellulosae) is not autochthonous in Belgium.

National evaluation of the recent situation, the trends and sources of infection

Cysticercus bovis in muscular tissue of cattle is the larval stage of the tapeworm, Taenia saginata, a parasitic cestode of the human gut (taeniasis). Cattle can become infected by ingestion of vegetation contaminated with T. saginata eggs shed in human faeces. Risk factors are access to rivers and flooding of pastures or wetland. Humans contaminate themselves by the ingestion of raw or undercooked beef containing the larval form (cysticerci). Usually pathogenicity for humans is low. The tapeworm eggs contaminate the environment directly or through surface waters. Human carriers should be treated promptly. Strict rules for the hygienic disposal or sanitation of human faeces with a method that inactivates T. saginata eggs should be developed. The spreading of human excrement on land should not be allowed.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

Post-mortem, macroscopic examination of carcasses of adult cattle as well as calves is routinely done in all slaughterhouses. Serological examination is possible and confirmation of the lesions by molecular tests can be done. Lightly contaminated carcasses are treated by freezing at -18C for 10 days before declared fit for human consumption. Heavily contaminated carcasses are unfit for human consumption and are destroyed.

Suggestions to the European Union for the actions to be taken

The introduction of serological analyzes for the detection of cysticerci antigens in the serum of animals (cattle) should be developed. This would allow the detection of more cases of infection than by live and infectious cysts by visual inspection of carcasses at slaughterhouse.

3.11 SARCOCYSTOSIS

3.11.1 General evaluation of the national situation

3.11.1.1 Sarcocystis - general evaluation

History of the disease and/or infection in the country

At the slaughterhouses, a small number of carcasses showing myositis eosinophilica (green colouring spots of the carcass) are detected and notified to the Federal Agency for the Safety of the Food Chain. In case of positive findings, carcasses are totally rejected and declared unfit for human consumption. In 2010, 2011, 2012, 2013, 2014, 2015 and 2016 respectively 37, 44, 60, 75, 94, 107 and 90 cases of sarcosporidiosis in cattle were reported.

National evaluation of the recent situation, the trends and sources of infection

Sarcocystis bovihominis (bovine as intermediate host) and Sarcocystis suihominis (porcine intermediate host) occur sporadically. Domestic carnivores are hosts of the adult stage. Humans can be a definitive host for sarcosporidiosis by ingestion of infected meat or excreted oocysts and develop symptoms like diarrhea, headache, eosinophilia, abortion, congenital disorder. For human sarcosporidiosis there is no immunity development. A majority of grazing animals are inappearent carriers of tissue cysts.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

Carcasses are entirely condemned when myositis eosinophilica lesions are apparent. Myositis eosinophilica is commonly associated with sarcosporidiosis but this has still to be proven!

3.12 TOXOPLASMA

3.12.1 General evaluation of the national situation

3.12.1.1 Toxoplasma - general evaluation

History of the disease and/or infection in the country

The majority of grazing animals seem to be inappearent carriers of tissue cysts.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

Humans are infected with Toxoplasma gondii through ingestion of undercooked infected meat or upon accidental ingestion of sporulated oocysts from the environment. The cat is the final host, man and most warm-blooded animals are intermediate hosts. Most infections with T.gondii are asymptomatic, however mild (flu-like symptoms), moderate (lymphadenopathy, chronic fatigue) to severe disease (disseminated toxoplasmosis, encephalitis) may occur, the latter mainly in immunocompromized hosts. Moreover, when infection occurs in pregnant women, toxoplasmosis may cause abortion and congenital disorders. If a woman acquires primary infection during pregnancy, Toxoplasma can be transmitted through the placenta to the foetus and lead to congenital toxoplasmosis. A percentage of young children (1 to 14-year-old age group) may get post-natal infections with T. gondii and develop symptomatic toxoplasmosis (e.g. ocular disease). A number of cases of the disease in a 15 to 24-year-old age group may be referred to as acquired toxoplasmosis in immunocompetent patients, which may present a wide range of clinical signs, from lymphadenopathy to retinitis and uveitis. Immunocompetent individuals may often develop clinical toxoplasmosis. The majority of adult persons have acquired a degree of immunity to re-infection but can remain carrier.

Recent actions taken to control the zoonoses

Screening for toxoplasmosis during pregnancy is common in Belgium. The seroprevalence in women tested before pregnancy is about 50%. Prevention of congenital toxoplasmosis by specific hygienic measures seems to have limited impact.

3.13 VTEC

3.13.1 Escherichia coli in foodstuffs

3.13.1.1 Verotoxigenic E. coli (VTEC) in food - All foodstuffs - food sample

Monitoring system

Sampling strategy

The Belgan Food Safety Agency foresees sampling of carcasses (bovine) in slaughterhouses, and sampling of meat preparations in cutting plants, meat manufacturing plants, retail and at import. Also carcasses from sheep were sampled at slaughterhouse. Also other products susceptible to STEC were analysed, e.g. raw milk cheeses, sprouted seeds, other dairy products, pre-cut fruits and vegetables and vegetables.

Frequency of the sampling

samples are taken throughout the year, suveillance.

Methods of sampling (description of sampling techniques)

Sampling of beef carcasses was done by means of swabs (4 areas from the same half carcass constituting 1600 cm2 were putted in the same stomacher bag). The detection of STEC has been assessed in 1600 cm2 for beef carcasses and in 25g for beef minced meat and beef cuts, cheese, fruits and vegetables, sprouted seeds, No pooling has been done.

Definition of positive finding

A sample is considered positive after isolation and genetic confirmation of the pathogenicity of the strain in the sample (stx+). Action is taken when as well the eae gene is present in the isolated strain.

Diagnostic/analytical methods used

STEC: Microbiology of food and animal feed -- Real-time polymerase chain reaction (PCR)-based method for the detection of food-borne pathogens -- Horizontal method for the detection of Shiga toxin-producing Escherichia coli (STEC) and the determination of O157. E. coli O104:H4 european protocol (EU-RL)

Control program/mechanisms

Suggestions to the European Union for the actions to be taken

Guidance for hamonised action to be taken in the whole EU.

Measures in case of the positive findings or single cases

Tracing back of the products. Notification of the producer or importer, possibility of a counter analysis, destruction of the non compliant products (or rework), further investigation: additional sampling, possible recall, RASFF, ...

Notification system in place

Notification is mandatory since 1/3/2004 (Ministerial Decree on mandatory notification in the food chain of 22/1/2004). Laboratories have to inform the Federal Agency in case of positive sample.

4 ANTIMICROBIAL RESISTANCE INFORMATION ON SPECIFIC ZOONOSES AND ZOONOTIC AGENTS

4.1 SALMONELLOSIS

4.1.1 Salmonella in foodstuffs

4.1.1.1 Antimicrobial resistance in Salmonella Meat from broilers (Gallus gallus)

Sampling strategy used in monitoring

Frequency of the sampling

Sampling of the programm Salmonella EFSA specifications is distributed evenly throughout the year and designed by the national competent authority (FASFC) Federal Agency for the Safety of the Food Chain

Type of specimen taken

carcasses from broilers (fresh chilled)

Procedures for the selection of isolates for antimicrobial testing

All strains isolated during the zoonosis monitoring program, national monitoring and efsa specific monitoring, were sent to the Institute of Public Health for serotyping and determination of antimicrobial resistance.

Methods used for collecting data

Since 2014, antimicrobia susceptibility testing (AST) was performed in all the serotypes determined

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Minimum Inhibitory Concentrations (MIC) were determined by the broth dilution method using Sensititre EUVSEC and EUVSEC2 panels, as described in the EU-directive of 13 november 2013. The antimicrobials reported as well as the breakpoints for interpretation are listed in the tables below. Quality control was performed by using an Escherichia coli ATCC 25922 strain. First panel EUVSEC Antimicrobials used: Ampicillin, Cefotaxime, Ceftazidime, Meropenem, Nalidixic acid, Ciprofloxacin, Tetracycline, Colistin, Gentamicin, Trimethoprim, Sulfamethoxazole, Chloramphenicol, Azithromycin, Tigecycline. Second panel EUVSEC2 Antimicrobials used: Cefoxitin, Cefepime, Cefotaxime+clavulanic acid, ceftazidime+clavulanic acid, Meropenem, Temocillin, Imipenem, Ertapenem, Cefotaxtime, ceftazidime.

Cut-off values used in testing

Minimum Inhibitory Concentrations (MIC) were determined by the broth dilution method using Sensititre EUVSEC panel, as described in the EU-directive of 13 november 2013. The antimicrobials reported as well as the breakpoints for interpretation are listed in the table below. Quality control was performed by using an Escherichia coli ATCC 25922 strain. First panel EUVSEC Antimicrobials ECOFF (R> mg/l) Ampicillin 8, Cefotaxime 0.5, Ceftazidime 2, Meropenem 0.125, Nalidixic acid 16, Ciprofloxacin 0.064, Tetracycline 8, Colistin 2, Gentamicin 2, Trimethoprim 2, Sulfamthoxazole 256, Chloramphenicol 16, Azithromycin 16, Tigecycline 1. Second panel EUVSEC2 Antimicrobials ECOFF* (R>mg/l) Cefoxitin 8, Cefepime 0.125 Cefotaxime+clavulanic acid 0.5, ceftazidime+clavulanic acid 2, Meropenem 0.125 ,Temocillin 32, Imipenem 1, Ertapeneme 0.06, Cefotaxtime 0.5, ceftazidime 2.

Additional information

Antimicrobial resistance in strains isolated from broiler carcasses (EFSA-specific monitoring). During 2016, 176 Salmonella spp. isolates from broiler carcases (EFSA specification) were performed for their antimicrobial susceptibility testing (AST). Among them, 85 were obtained from the national surveillance plan and 93 were obtained from the food business operators. The most predomiannt serotype was Infantis (40%) followed by Give (15%) and Enteritidis (11 %). The highest resistance reported was to sulfamethoxazole, ciprofloxacine and nalidixic acid (56-55%), followed by Trimethoprim (26%) and Ampicillin (22%). For Azithromicin resistance was mostly linked to the serovar Infantis (8%). For cefotaxime and ceftazidime values ranged between 3.4 to 2.8%. No resistance to meropenem was detected. Among the 176 isolates, 7 were confirmed to be ESBL producers. It belong to the serovars autoagglutinable (n=2), Infantis (n=2) and Paratyphi B var L(+) Tartrate + (n=3). Those isolates display a multidrug resistant phenotype.

4.1.1.2 Antimicrobial resistance in Salmonella Other food

Sampling strategy used in monitoring

Frequency of the sampling

Strains of Salmonella enterica isolated during the zoonosis monitoring program (national and Efsa specific monitoring) were sent to the Scientific Institute of Public Health for serotyping and determination of antimicrobial resistance. Different food matrices were sampled, mainly poultry (carcasses from broilers and spent hens, chicken parts and meat preparations) and pork (carcasses and cut meats). Other matrices where Salmonella was isolated were ready-to-eat meals, meat, meat preparations, frogs legs, crustaceans and fruits. Since 2014, the AMR was performed on all serotypes.

Frequency of the sampling

Sampling of the programm Salmonella Food is distributed evenly throughout the year and designed by the national competent authority (FASFC) Federal Agency for the Safety of the Food Chain

Type of specimen taken

All types of matrices including in the national surveillance programme designed by the national authority

Procedures for the selection of isolates for antimicrobial testing

All strains isolated during the zoonosis monitoring program were sent to the Institute of Public Health for serotyping and determination of antimicrobial resistance. Since 2014, AMR was performed on all serotypes identified.

Laboratory methodology used for identification of the microbial isolates

Minimum Inhibitory Concentrations (MIC) were determined by the broth dilution method using Sensititre, as described in the EU-legislation Official Journal of the European Union, Commission implementing decision of 12 november 2013 on the monitoring and reporting of antimicrobial resistance in zoonotic and commesal bacteria. Interpretation was according to the EU-legislation. Quality control was performed by using an Escherichia coli ATCC 25922 strain.

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Minimum Inhibitory Concentrations (MIC) were determined by the broth dilution method using Sensititre EUVSEC and EUVSEC2 panels, as described in the EU-directive of 13 november 2013. The antimicrobials reported as well as the breakpoints for interpretation are listed in the tables below. Quality control was performed by using an Escherichia coli ATCC 25922 strain. First panel EUVSEC Antimicrobials used: Ampicillin, Cefotaxime, Ceftazidime, Meropenem, Nalidixic acid, Ciprofloxacin, Tetracycline, Colistin, Gentamicin, Trimethoprim, Sulfamethoxazole, Chloramphenicol, Azithromycin, Tigecycline. Second panel EUVSEC2 Antimicrobials used: Cefoxitin, Cefepime, Cefotaxime+clavulanic acid, ceftazidime+clavulanic acid, Meropenem, Temocillin, Imipenem, Ertapenem, Cefotaxtime, ceftazidime.

Cut-off values used in testing

The cut-off values were used as described in the European Decision of 12 november 2013 on the monitoring and reporting of antimicrobial resistance in zoonotic and commensal bacteria. Minimum Inhibitory Concentrations (MIC) were determined by the broth dilution method using Sensititre EUVSEC panel, as described in the EU-directive of 13 november 2013. The antimicrobials reported as well as the breakpoints for interpretation are listed in the table below. Quality control was performed by using an Escherichia coli ATCC 25922 strain. First panel EUVSEC Antimicrobials ECOFF (R> mg/l) Ampicillin 8, Cefotaxime 0.5, Ceftazidime 2, Meropenem 0.125, Nalidixic acid 16, Ciprofloxacin 0.064, Tetracycline 8, Colistin 2, Gentamicin 2, Trimethoprim 2, Sulfamthoxazole 256, Chloramphenicol 16, Azithromycin 16, Tigecycline 1. Second panel EUVSEC2 Antimicrobials ECOFF* (R>mg/l) Cefoxitin 8, Cefepime 0.125, Cefotaxime+clavulanic acid 0.5, ceftazidime+clavulanic acid 2, Meropenem 0.125, Temocillin 32, Imipenem 1, Ertapeneme 0.06, Cefotaxtime 0.5, ceftazidime 2.

Additional information

In total 227 Salmonella strains recovery on 2016 from the food-program, were tested for their antimicrobial susceptibility (AST). This includes mainly isolates from porc carcasses, poultry cut meats but also from other food matrices such as meat preparations, crustaceans and frog legs. Since 2014, all the serovars are subjected to AST determination. The predominant serovars were Enteritidis, Infantis, Derby and Typhimurium. The overall analysis of all the matrices and serovars together, have shown that the resistance to meropenem was 0% however resistance to cefotaxime was 2.2% and to ceftazidime 0.88%. Resistance to other antimicrobials was as follow, sulfamethoxazole (39%), Ampicillin (33%), tetracycline (29%), ciprofloxacine (22%), trimetophrim (20%), acide nalidixique (18%), Tigecycline (7.93%), Chloramphenicol (7.5%), Azithromycin (4.85%) and colistin (3.5%).

4.1.1.3 Antimicrobial resistance in Salmonella spp., unspecified Meat from bovine animals

Description of sampling designs

Carcase samples of bovines under one year of age for the analyses of Salmonella and its antimicrobial resistance are taken at the level of the slaughterhouse in the framework of the national control programme of the Federal Agency for the Safety of the Food Chain by official agents for testing and verification of compliance, in accordance with part G of chapter IX of section IV to Regulation (EC) No 854/2004. Samples of bovines under one year of age are also taken by FBO's in the framework of self-checking in accordance with point 2.1.3 of Chapter 2 of Annex I to Regulation (EC) No 2073/2005.

Stratification procedures per animal populations and food categories

In all slaugherhouses where bovines under one year of age are slaughtered, 5 random samples are taken per week by the FASFC during 10 consecutive weeks to obtain 50 samples. In accordance with Regulation (EC) No 2073/2005, 5 random samples are taken each week by the FBO in the slaughterhouse.

Randomisation procedures per animal populations and food categories

All Salmonella isolates obtained in the framework of the national control programme of the FASFC are selected for antimicrobial resistance testing. Salmonella isolated obtained by FBO's are chosen at random to achieve the minimal required number of isolates.

4.1.1.4 Antimicrobial resistance in Salmonella spp., unspecified Meat from broilers (Gallus gallus)

Description of sampling designs

Broilers carcasses are taken for the analyses of Salmonella and its antimicrobial resistance at the level of the slaughterhouse in the framework of the national control programme of the Federal Agency for the Safety of the Food Chain by official agents for testing and verification of compliance with point 2.1.5 of Chapter 2 of Annex I to Regulation (EC) No 2073/2005. Carcase samples of broilers are also taken by the FBO in the framework of self-checking in accordance with point 2.1.5. of Chapter 2 of Annex I to Regulation (EC) No 2073/2005.

Description of sampling designs

Fresh meat of broilers are taken for the analyses of Salmonella and its antimicrobial resistance at the level of retail in the framework of the national control programme of the Federal Agency for the Safety of the Food Chain by official agents.

Stratification procedures per animal populations and food categories

In all slaugherhouses where > 1.000.000 broilers are slaughtered/year, 5 random samples are taken per week by the FASFC during 10 consecutive weeks to obtain 50 samples. In accordance with Regulation (EC) No 2073/2005, 5 random samples are taken each week by the FBO in the slaughterhouse.

Stratification procedures per animal populations and food categories

All samples (carcases, cutting meat and meat preparations) taken in retail in the framework of the national control programme of FASFC to detect pathogens and hygiene indicators are also used to monitor antimicrobial resistance Salmonella. Sampling is distributed evenly throughout the year and over the whole territory.

Randomisation procedures per animal populations and food categories

All Salmonella isolates obtained in the framework of the national control programme of the FASFC are selected for antimicrobial testing.

Randomisation procedures per animal populations and food categories

All Salmonella isolates obtained in the framework of the national control programme of the FASFC are selected for antimicrobial resistance testing. Salmonella isolated obtained by FBO's are chosen at random to achieve the minimal required number of isolates.

4.1.1.5 Antimicrobial resistance in Salmonella spp., unspecified Meat from pig

Description of sampling designs

Carcase samples of fattening pigs for the analyses of Salmonella and its antimicrobial resistance are taken at the level of the slaughterhouse in the framework of the national control programme of the Federal Agency for the Safety of the Food Chain by official agents for testing and verification of compliance, in accordance with part G of chapter IX of section IV to Regulation (EC) No 853/2004. Carcase samples of fattening pig are also taken by the FBO in the framework of self-checking in accordance with point 2.1.4. of Chapter 2 of Annex I to Regulation (EC) No 2073/2005.

Stratification procedures per animal populations and food categories

In accordance with Regulation (EC) No 854/2004, 49 random samples are taken by the FASFC in all slaughterhouses with more than 10.000 fattening pigs slaughtered per year and 35 in all slaughterhouses with between 1.000 and 10.000 fattening pigs slaughtered per year. In accordance with Regulation (EC) No 2073/2005, 5 random samples are taken each week by the FBO in the slaughterhouses.

Randomisation procedures per animal populations and food categories

All Salmonella isolates obtained in the framework of the national control programme of the FASFC are selected for antimicrobial testing. Salmonella isolated obtained by FBO's are chosen at random to achieve the minimal required number of isolates.

4.1.2 Salmonella in animals

4.1.2.1 Antimicrobial resistance in Salmonella spp., unspecified Gallus gallus (fowl)

Description of sampling designs

Salmonella isolates obtained in the framework of the National Salmonella control programme for broilers are used for the evaluation of the antimicrobial resistance of Salmonella in poultry.

Stratification procedures per animal populations and food categories

In case of Salmonella positive samples, one isolate of every positive sample is serotyped and at least one isolate of every positive flock is tested on AMR according to Decision 2013/652/EU.

Stratification procedures per animal populations and food categories

All laying hen flocks are sampled. In case of Salmonella positive samples, one isolate of every positive sample is serotyped and at least one isolate of every positive flock is tested on AMR according to Decision 2013/652/EU.

Randomisation procedures per animal populations and food categories

Seen the low number of isolates, at least one isolate of each positive flock is tested.

Sampling strategy used in monitoring

Frequency of the sampling

All broiler flocks with at least 200 broilers must be sampled within 3 weeks before slaughter by the food business operator. The FASFC takes official samples of at least one flock on 10% of the holdings with at least 200 broilers.

Frequency of the sampling

All laying hen flocks with at least 200 hens must be sampled as day-old-chicks, within 2 weeks of the movement to the laying unit, every 15 weeks during production and within 3 weeks before slaughter by the food business operator. Yearly, official samples are taken of at least one flock on all holdings with at least 200 adult laying hens.

Type of specimen taken

A sample of one flock consists of 2 pair of overshoes.

Type of specimen taken

The day-old-chicks are sampled by 10 pieces of inner linings of the delivery boxes soiled with feces. The food business operator takes one sample per flock consisting of 2 pair of overshoes. The FASFC takes two samples per flock, one consisting of 2 pair of overshoes and the other of dust.

Methods of sampling (description of sampling techniques)

The sampling is performed according to Regulation (EU) no 200/2012.

Methods of sampling (description of sampling techniques)

The samples are taken according to Regulation (EU) no 517/2011.

Methods used for collecting data

In the framework of the National Salmonella Control Programme, all laboratories involved in the detection of Salmonella gather the requested information concerning the sample and the sampled flock. The information is reported to the FASFC on a monthly basis, together with the results of the Salmonella analyses. All Salmonella isolates are sent to the NRL (CODA-CERVA) for serotyping and if applicable, AMR analyses.

Laboratory methodology used for identification of the microbial isolates

The analytical method used is the bacteriological method: ISO 6579:2002 annex D in accordance with regulation (EU) nr. 517/2011. All isolates are serotyped by the Kauffmann-White-LeMinor scheme.

Laboratory methodology used for identification of the microbial isolates

The analytical method used is the bacteriological method: ISO 6579:2002 annex D in accordance with regulation (EU) nr. 200/2012. Al isolates are serotyped by the Kauffmann-White-LeMinor scheme.

Laboratory used for detection for resistance

Antimicrobials included in monitoring

The test on the antibiomicrobial resistance is performed according to decision 2013/652/EU and includes all antimicrobials requested.

Cut-off values used in testing

The cut-off values described in decision 2013/652/EU were used. Epidemiological cut-offs (ECOFFs), established by the European Committee on Antimicrobial Susceptibility (EUCAST) or as defined by the EU-reference laboratory on antimicrobial resistance (DTU) were used

4.2 CAMPYLOBACTERIOSIS

4.2.1 Campylobacter in foodstuffs

4.2.1.1 Antimicrobial resistance in C. jejuni Meat from poultry, unspecified

Sampling strategy used in monitoring

Procedures for the selection of isolates for antimicrobial testing

In 2016, 370 Campylobacter strains isolated in the zoonoses monitoring programme and originating from poultry, (carcasses of broilers, filets, entrails, meat preparation and carcasses of spent hens) were subjected to antimicrobial susceptibility testing (AST)

Procedures for the selection of isolates for antimicrobial testing

All strains isolated in the zoonosis monitoring program and originating from poultry were sent to the Institute of Public Health for determination of antimicrobial resistance.

Laboratory methodology used for identification of the microbial isolates

Maldi Tof were used for especies indentification

Laboratory methodology used for identification of the microbial isolates

Species identification by Maldi-Tof (coli/jejuni)

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Minimum Inhibitory Concentrations (MIC) were determined by using broth microdilution method (Sensititre EUCAMP2 panel). From 2014, a new European decision on the harmonization of the monitoring and reporting of antimicrobial resistance in zoonotic and commensal bacteria is adopted which specifies new interpretative threshold for resistance for C jejuni and C. coli. Therefore, the antimicrobials tested and the epidemiological cut-off values (ECOFF) used are listed in the table below following the official Journal of the European Union (L303/26 14.11.2013).Campylobacter in meat and meat products: list of antimicrobials tested and breakpoints used.AntimicrobialBreakpoints R > (g / ml) C. jejuni/C. coliTetracycline1/2Nalidixic acid16/16Ciprofloxacin0.5/0.5Erythromycin4/8Gentamicin2/2Streptomycin4/4

Antimicrobials included in monitoring

Minimum Inhibitory Concentrations (MIC) were determined by using broth microdilution method (Sensititre EUCAMP2 panel). From 2014, a new European decision on the harmonization of the monitoring and reporting of antimicrobial resistance in zoonotic and commensal bacteria is adopted which specifies new interpretative threshold for resistance for C jejuni and C. coli. Therefore, the antimicrobials tested and the epidemiological cut-off values (ECOFF) used are listed in the table below following the official Journal of the European Union (L303/26 14.11.2013). In order to compared results in an accurate way, recalculation of resistance to the antibiotics using the new breakpoint established in the European decision was done for the last four years, from 2010 to 2013 included.AntimicrobialBreakpoints R > (g / ml) C. jejuniC. coliTetracycline12Nalidixic

acid1616Ciprofloxacin0.50.5Erythromycin48Gentamicin22Streptomycin44Campylobacter in meat and meat products: list of antimicrobials tested and breakpoints used.

4.2.2 Campylobacter in animals

4.2.2.1 Antimicrobial resistance in Thermophilic Campylobacter spp., unspecified Gallus gallus (fowl)

Description of sampling designs

Samples of broiler flocks for the analyses of Campylobacter and its antimicrobial resistance are taken at the level of the slaughterhouse in the framework of the national control programme of the Federal Agency for the Safety of the Food Chain by official agents. The samples are programmed to be taken evenly spread over the year over all slaughterhouses in the country.

Stratification procedures per animal populations and food categories

The sampling is distributed evenly throughout the year, based on the slaughtercapacity per local control unit.

Randomisation procedures per animal populations and food categories

At the slaughterhouse, the samples are taken of a previously planned flock.

Sampling strategy used in monitoring

Frequency of the sampling

Samples of broiler flocks for the analyses of the antimicrobial resistance of Campylobacter jejuni were taken evenly divided over the period Februari to November 2016.

Type of specimen taken

A sample of one flock consists of 10 pooled caeca samples.

Methods of sampling (description of sampling techniques)

Both caeca of one bird are separated by prudent manual traction from the intestinal packet and placed in a sterile pot. Once all 10 pairs of caeca are collected, the sample is placed in a cooled box or a coolbox containing ice in order to cool down the sample rapidly. The samples are cooled untill arrival at the lab on the same day or at the latest the day after sampling.

Methods used for collecting data

All data concerning the sampling mission (time and place of sampling, the type of sample taken and the identity of the sampler) are registered in the central database 'Foodnet' of the Federal Agency for the Safety of the Food Chain. The results of the internal laboratories are also registered in the same central database. The results of the external laboratories are provided by the external laboratory using specific templates on demand.

Laboratory methodology used for identification of the microbial isolates

Campylobacter isolates were identified by MALDI-TOF following a standard extraction procedure recommended by the manufacturer (BRUKER DALTONICS).

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Following antimicrobials are included in the monitoring: Ciprofloxacin (CIP), Erythromycin (ERY), Gentamicin (GEN), Nalidixic acid (NAL), Streptomycin (STR) and Tetracycline (TET).

Cut-off values used in testing

Followig EUCAST ECOFF's were used in testing: Ciprofloxacin (CIP): > 0.5; Erythromycin (ERY): > 4; Gentamicin (GEN): > 2; Nalidixic acid (NAL): > 16; Streptomycin (STR): > 4 and Tetracycline (TET): > 1.

4.3 ESCHERICHIA COLI, NON-PATHOGENIC

4.3.1 Escherichia coli, non-pathogenic in animals

4.3.1.1 Antimicrobial resistance in E.coli, non-pathogenic, unspecified Cattle (bovine animals)

Description of sampling designs

Samples of young bovines (meat production) for the analyses of the antimicrobial resistance of commensal E. coli are taken at farmlevel in the framework of the national control programme of the Federal Agency for the Safety of the Food Chain by official agents. This yearly monitoring started in 2012. The number of samples taken is calculated based on the detection percentage of commensal E. Coli. The samples are programmed to be taken spread over the year in the whole country.

Description of sampling designs

Samples of veal calves for the analyses of the antimicrobial resistance of commensal E. coli and the specific monitoring of ESBL/AmpC/Carbapenamse producing E. coli are taken at the level of the slaughterhouse in the framework of the national control programme of the Federal Agency for the Safety of the Food Chain by official agents. This yearly monitoring started in 2012. The number of samples taken is calculated based on the detection percentage of commensal E. Coli. The samples are programmed to be taken spread over the year over all slaughterhouses in the country.

Stratification procedures per animal populations and food categories

The sampling programmed is evenly spread over the sampling period (January to December) on a monthly basis in each province. The number of samples to be taken per province is directly related to the number of cattle farms.

Stratification procedures per animal populations and food categories

The sampling programmed is evenly spread over the sampling period (January to December) on a monthly basis in each province. The number of samples to be taken per province is directly related to the number of slaughterhouses.

Randomisation procedures per animal populations and food categories

The sampler chooses on which day of the month the sample will be taken. At the farm, the bovines sampled are also chosen by the sampler.

Randomisation procedures per animal populations and food categories

The sampler chooses on which day of the month the sample will be taken. At the slaughterhouse, the lot sampled is also chosen by the sampler.

Sampling strategy used in monitoring

Frequency of the sampling

Samples of young bovines for the analyses of the antimicrobial resistance of commensal E. coli (177) were taken evenly divided over the period Februari to November 2016.

Frequency of the sampling

Samples of veal calves for the analyses of the antimicrobial resistance of commensal E. coli (210) and the specific monitoring of ESBL/AmpC/Carbapenamse producing E. coli (300) were taken evenly divided over the period Februari to November 2016.

Type of specimen taken

A sample consists of feces of at least 10 bovines of less than 7 month of age taken from the floor in the stable.

Type of specimen taken

A sample of one lot consists of 100 ml feces taken from the colon/rectum.

Methods of sampling (description of sampling techniques)

At least 20 ml of fresh, moist feces are taken from the floor from different places in a box using sterile gloves. If there are less than 10 animals present in a box, several boxes are sampled to assure to have at least feces of 10 animals. Samples must be kept moist during sampling.

Methods of sampling (description of sampling techniques)

100 ml of feces is collected with a sterile glove directly from the colon/rectum. The sample is transported cooled and must arrive at the laboratory within 72 hours.

Methods used for collecting data

All data concerning the sampling mission (time and place of sampling, the type of sample taken and the identity of the sampler) are registered in the central database 'Foodnet' of the Federal Agency for the Safety of the Food Chain. The results of the internal laboratories are also registered in the same central database. The results of the external laboratories are provided by the external laboratories on demand using specific templates.

Laboratory methodology used for identification of the microbial isolates

In Flanders, faecal material was inoculated on McConkey agar and incubated at 37°C for 18 to 24 hours. Suspected colonies (pink, lactose positive) were inoculated on Kligler and indol medium and incubated at 37°C for 18 to 24 hours. When the test outcome was positive for E. coli a colony from the Kligler medium was inoculated on Mac Conckey agar and incubated at 37°C for 18-24 hours. In Wallonie, fecal material was inoculated on Gassner medium and incubated at 37°C for 18 to 24 hours. Suspected colonies were purified on Columbia agar supplemented with 5% sheep blood. Identification was done by the OPNG test, Ureum test and indol test. Finally, before antimicrobial susceptibility testing, all strains were purified on Columbia agar with 5% sheep blood and confirmed to be E. coli by MALDI-TOF.

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Following antimicrobials were included in the monitoring: Ampicillin (AMP); Cefotaxime (FOT); Ceftazidime (TAZ); Meropenem (MERO); Nalidixic acid (NAL); Ciprofloxacin (CIP); Tetracycline (TET); Colistin (COL); Gentamicin (GEN); Trimethoprim (TMP); Sulfamethoxazole (SMX); Chloramphenicol (CHL); Azithromycin (AZI) and Tigecycline (TGC).

Cut-off values used in testing

Following cut-off values were used: AMP > 8; FOT > 0.25; TAZ > 0.5; MERO > 0.125; NAL > 16; CIP > 0.064; TET > 8; COL > 2; GEN > 2; TMP > 2; SMX > 64; CHL > 16; AZI >16 and TGC > 1.

4.3.1.2 Antimicrobial resistance in E.coli, non-pathogenic, unspecified Gallus gallus (fowl)

Description of sampling designs

Samples of broiler flocks for the analyses of the antimicrobial resistance of commensal E. coli and the specific monitoring of ESBL/AmpC/Carbapenamse producing E. coli are taken at the level of the slaughterhouse in the framework of the national control programme of the Federal Agency for the Safety of the Food Chain by official agents. This yearly monitoring started in 2012. The number of samples taken is calculated based on the detection percentage of commensal E. Coli. The samples are programmed to be taken spread over the year over all slaughterhouses in the country.

Stratification procedures per animal populations and food categories

The sampling programmed is evenly spread over the sampling period (January to December) on a monthly basis in each province. The number of samples to be taken per province is directly related to the number of slaughterhouses.

Randomisation procedures per animal populations and food categories

At the slaughterhouse, the samples are taken of a previously planned flock.

Sampling strategy used in monitoring

Frequency of the sampling

Samples of broiler flocks for the analyses of the antimicrobial resistance of commensal E. coli (170) and the specific monitoring of ESBL/AmpC/Carbapenamse producing E. coli (300) were taken evenly divided over the period Februari to November 2016.

Type of specimen taken

A sample of one flock consists of 10 pooled caeca samples.

Methods of sampling (description of sampling techniques)

Both caeca of one bird are separated by prudent manual traction from the intestinal packet and placed in a sterile pot. Once all 10 pairs of caeca are collected, the sample is placed in a cooled box or a coolbox containing ice in order to cool down the sample rapidly. The samples are cooled untill arrival at the lab on the same day or at the latest the day after sampling.

Methods used for collecting data

All data concerning the sampling mission (time and place of sampling, the type of sample taken and the identity of the sampler) are registered in the central database 'Foodnet' of the Federal Agency for the Safety of the Food Chain. The results of the internal laboratories are also registered in the same central database. The results of the external laboratories are provided by the external laboratory using specific templates on demand.

Laboratory methodology used for identification of the microbial isolates

In Flanders, faecal material was inoculated on McConkey agar and incubated at 37°C for 18 to 24 hours. Suspected colonies (pink, lactose positive) were inoculated on Kligler and indol medium and incubated at 37°C for 18 to 24 hours. When the test outcome was positive for E. coli a colony from the Kligler medium was inoculated on Mac Conckey agar and incubated at 37°C for 18-24 hours. In Wallonie, fecal material was inoculated on Gassner medium and incubated at 37°C for 18 to 24 hours. Suspected colonies were purified on Columbia agar supplemented with 5% sheep blood. Identification was done by the OPNG test, Ureum test and indol test. Finally, before antimicrobial susceptibility testing, all strains were purified on Columbia agar with 5% sheep blood and confirmed to be E. coli by MALDI-TOF.

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Following antimicrobials were included in the monitoring: Ampicillin (AMP); Cefotaxime (FOT); Ceftazidime (TAZ); Meropenem (MERO); Nalidixic acid (NAL); Ciprofloxacin (CIP); Tetracycline (TET); Colistin (COL); Gentamicin (GEN); Trimethoprim (TMP); Sulfamethoxazole (SMX); Chloramphenicol (CHL); Azithromycin (AZI) and Tigecycline (TGC).

Cut-off values used in testing

Following cut-off values were used: AMP > 8; FOT > 0.25; TAZ > 0.5; MERO > 0.125; NAL > 16; CIP > 0.064; TET > 8; COL > 2; GEN > 2; TMP > 2; SMX > 64; CHL > 16; AZI > 16 and TGC > 1.

4.3.1.3 Antimicrobial resistance in E.coli, non-pathogenic, unspecified Pigs

Description of sampling designs

Samples of fattening pigs for the analyses of the antimicrobial resistance of commensal E. coli and the specific monitoring of ESBL/AmpC/Carbapenamse producing E. coli are taken at the level of the slaughterhouse in the framework of the national control programme of the Federal Agency for the Safety of the Food Chain by official agents. This yearly monitoring started in 2012. The number of samples taken is calculated based on the detection percentage of commensal E. coli. The samples are programmed to be taken spread over the year over all slaughterhouses in the country.

Stratification procedures per animal populations and food categories

The sampling programmed is evenly spread over the sampling period (January to December) on a monthly basis in each province. The number of samples to be taken per province is directly related to the number of slaughterhouses.

Randomisation procedures per animal populations and food categories

The sampler chooses on which day of the month the sample will be taken. At the slaughterhouse, the lot sampled is also chosen by the sampler.

Sampling strategy used in monitoring

Frequency of the sampling

Samples of fattening pigs for the analyses of the antimicrobial resistance of commensal E. coli (210) and the specific monitoring of ESBL/AmpC/Carbapenamse producing E. coli (300) were taken evenly divided over the period Februari to November 2016.

Type of specimen taken

A sample of one lot consists of 100 ml feces taken from the colon/rectum.

Methods of sampling (description of sampling techniques)

100 ml of feces is collected with a sterile glove directly from the colon/rectum. The sample is transported cooled and must arrive at the laboratory within 72 hours.

Methods used for collecting data

All data concerning the sampling mission (time and place of sampling, the type of sample taken and the identity of the sampler) are registered in the central database 'Foodnet' of the Federal Agency for the Safety of the Food Chain. The results of the internal laboratories are also registered in the same central database. The results of the external laboratories are provided by the external laboratories on demand using specific templates.

Laboratory methodology used for identification of the microbial isolates

In Flanders, faecal material was inoculated on McConkey agar and incubated at 37°C for 18 to 24 hours. Suspected colonies (pink, lactose positive) were inoculated on Kligler and indol medium and incubated at 37°C for 18 to 24 hours. When the test outcome was positive for E. coli a colony from the Kligler medium was inoculated on Mac Conckey agar and incubated at 37°C for 18-24 hours. In Wallonie, fecal material was inoculated on Gassner medium and incubated at 37°C for 18 to 24 hours. Suspected colonies were purified on Columbia agar supplemented with 5% sheep blood. Identification was done by the OPNG test, Ureum test and indol test. Finally, before antimicrobial susceptibility testing, all strains were purified on Columbia agar with 5% sheep blood and confirmed to be E. coli by MALDI-TOF.

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Following antimicrobials were included in the monitoring: Ampicillin (AMP); Cefotaxime (FOT); Ceftazidime (TAZ); Meropenem (MERO); Nalidixic acid (NAL); Ciprofloxacin (CIP); Tetracycline (TET); Colistin (COL); Gentamicin (GEN); Trimethoprim (TMP); Sulfamethoxazole (SMX); Chloramphenicol (CHL); Azithromycin (AZI) and Tigecycline (TGC).

Cut-off values used in testing

Following cut-off values were used: AMP > 8; FOT > 0.25; TAZ > 0.5; MERO > 0.125; NAL > 16; CIP > 0.064; TET > 8; COL > 2; GEN > 2; TMP > 2; SMX > 64; CHL > 16; AZI > 16 and TGC > 1

4.4 STAPHYLOCOCCUS AUREUS METICILLIN RESISTANT (MRSA) INFECTION

4.4.1 Staphylococcus in animals

4.4.1.1 Antimicrobial resistance in S. aureus, meticillin resistant (MRSA) Pigs

Description of sampling designs

In 2016, the monitoring of the presence of MRSA and its antimicrobial resistance was performed in fattening and breeding pigs. The number of units to be sampled was based on the results of the same monitoring in 2013. The monitoring is repeated every 3 years.

Stratification procedures per animal populations and food categories

The samples are programmed to be taken evenly divided over the year and over the different provinces, based on the number of farms per province.

Randomisation procedures per animal populations and food categories

If breeding and fattening pigs were present on the farm, both categories could be sampled. Samples taken from fattening pigs were taken within two months before slaughter.

Sampling strategy used in monitoring

Type of specimen taken

10 nasal swabs are taken from one nostril from 10 different animals.

Methods of sampling (description of sampling techniques)

The nasal swabs are taken from one nostril each from 10 different animals. Each swab is transported in its own transportation tube. The swabs are pooled at the level of the laboratory to one sample. The swabs are stored between 5°C and 25°C.

Methods used for collecting data

All data concerning the sampling mission (time and place of sampling, the type of sample taken and the identity of the sampler) are registered in the central database 'Foodnet' of the Federal Agency for the Safety of the Food Chain. The results of the detection of MRSA (suspected colonies) are also registered in the same central database. The results of the external laboratories are provided by the external laboratories on demand using specific templates.

Laboratory methodology used for identification of the microbial isolates

Pooled samples were incubated in Mueller-Hinton (MH) broth (Becton Dickinson) supplemented with NaCl (6.5%) at 37C for 18-24h. One ml of this broth was added to Tryptic Soy Broth (TSB) supplemented with cefoxitin (3.5 mg/l) and aztreonam (75 mg/l) and incubated at 37C for 18-24h. Ten microliter of this enrichment was plated on Brilliance MRSA 2 (Oxoid) and incubated 18-24h at 37C. Presence of MRSA was suspected based on colony morphology. Per sample, one to five suspected colonies were selected from the Brilliance MRSA 2 plate. Presence of MRSA was confirmed using a triplex real-time PCR method. Per sample, one to five suspected colonies were selected from the Brilliance MRSA 2 plate. DNA was extracted as described in SOP/BAC/ANA/18. The PCR allows detecting the Staphylococcal aureus specific gene, nuc, the presence of the mecA gene responsible for methicillin resistance and the variant mecC gene. All MRSA isolates were spa-typed by sequencing the repetitive region of the spa gene encoding for the staphylococcal protein A. This method depicts the rapid evolution, since through recombination, the repeats may change fast. The protein A (spa) gene was amplified according to the Ridom StaphType standard protocol (www.ridom.de/staphtype) and the amplification was checked on a 2% agarose gel. Sequencing was performed with CEQ 8000 using standard protocols and sequences were compared with the international Ridom database. CC398 PCR was performed on all MRSA following protocol described by Stegger et al. 2011. This method allows the rapid detection of the S. aureus sequence type ST398.

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Following antimicrobials were included in the monitoring: Chloramphenicol (CHL); Ciprofloxacin (CIP); Clindamycine (CLI); Erythromycine (ERY); cefoxitin (FOX); Fusidic acid (FUS); Gentamycin (GEN); Kanamycine (KAN); Linezolid (LZD); Mupirocin (MUP); Penicillin (PEN); Rifampicin (RIF); Sulfamethoxazole (SMX); Streptomycin (STR); Quinupristin/dalfopristin (SYN); Tetracycline (TET); Tiamulin (TIA); Trimethoprim (TMP) and Vancomycin (VAN).

Cut-off values used in testing

Following cut-off values were used: CHL > 16; CIP > 1; CLI > 0,25; ERY > 1; FOX > 4; FUS > 0,5; GEN > 2; KAN > 8; LZD > 4; MUP > 1; PEN > 0,12; RIF > 0,03; SMX > 128; STR > 16; SYN > 1; TET > 1; TIA > 2; TMP > 2 and VAN > 2.

5 INFORMATION ON SPECIFIC MICROBIOLOGICAL AGENTS

5.1 CRONOBACTER

5.1.1 Cronobacter in foodstuffs

5.1.1.1 Cronobacter in food - Infant formula - food sample

Monitoring system

Sampling strategy

Tests for Cronobacter sakazakii were performed in foodstuff intended for special nutritional uses, infant formula. Samples are taken at retail level and at manufactering leven. Also repared powdered milk in bottles for infants and young children is being sampled in hospitals (nursery for infants).

Frequency of the sampling

Samples are taken according to the national control program or in the framework of RASFF, complaints or suspicion. The samples are taken troughout the year.

Methods of sampling (description of sampling techniques)

Samples are taken according to a sampling scheme of n=1.

Definition of positive finding

To determine the conformity of a sample or a batch, the criteria laid down in the Regulation (EC) No 2073/2005 are applied.

Diagnostic/analytical methods used

ISO/TS 22964:2006 (IDF/RM 210: 2006) CRONOBACTERSPP. (ENTEROBACTER SAKAZAKII)

Measures in case of the positive findings or single cases

Measures to be taken in the case of a non-compliant result:- Notification of the producer or importer- Possibility of a counter analysis- Destruction of the non compliant batch - Further investigation: additional sampling, possible recall, RASFF, ...

5.2 HISTAMINE

5.2.1 Histamine in foodstuffs

5.2.1.1 Histamine in food - Fish - Fishery products from fish species associated with a high amount of histidine - not enzyme maturated - food sample

Control program/mechanisms

The control program/strategies in place

The annual control plan forsees controls of histamine in fishery products with high levels of histidine.

6 FOODBORNE OUTBREAKS

Foodborne outbreaks are incidences of two or more human cases of the same disease or infection where the cases are linked or are probably linked to the same food source. Situation, in which the observed human cases exceed the expected number of cases and where a same food source is suspected, is also indicative of a foodborne outbreak.

6.1 Outbreaks

6.1.1 Foodborne outbreaks

System in place for identification, epidemological investigations and reporting of foodborne outbreaks

In Belgium different authorities are dealing with food-borne outbreaks:-The Federal Agency for the Safety of the Food chain FASFC deals with safety of foodstuffs, epidemiological investigation on foodstuffs and animal health issues in case of a food-borne outbreak. -The Communities (Flemish, French and German speaking Community) are dealing with person related matters as human health and can start an epidemiological investigation by Public health medical inspectors in case of a food-borne outbreak. -The Scientific Institute of Public Health WIV-ISP (National Reference Laboratory on Food-borne Outbreaks) analyses all suspected food samples, collects all data on food-borne outbreaks and gives scientific support to the FASFC officers and the Public Health Inspectors. A national "Platform Food-borne outbreaks", approved by the National Conference of Ministers of Public Health, brings together the different competent authorities on food safety, animal health and public health. Furthermore in 2007, for a better communication, a protected web application was made available to exchange outbreak data and laboratory results in real time between the different authorities dealing with FBO. In this web-application a common file is created for each individual outbreak, and the data and laboratory results are shared between food inspectors and human health inspectors. Data in this report come from the Federal Agency for the Safety of the Food Chain, the Public Health Inspection, the sentinel laboratories network for human microbiology, and the Federal Reference Centres for Food-borne outbreaks, for Clostridium botulinum, for Salmonella and Shigella and for Listeria.

Description of the types of outbreaks covered by the reporting:

A food-borne outbreak is defined as an incidence, observed under given circumstances, of two or more human cases of the same disease and/or infection, or a situation in which the observed number of human cases exceeds the expected number and where the cases are linked, or are probably linked, to the same food source (Directive 2003/99/EC, Article 2(d)). Data are collected from FASFC, the Flemish Community, the French community, the Brussels Common Community Committee, the sentinel laboratories network for human clinical microbiology, the National Reference Laboratory for Food-borne outbreaks, and the National Reference Centres for Salmonella and Shigella, Listeria and C. botulinum. The reporting includes both general and household outbreaks. The causative agents covered are Salmonella spp., Shigella spp., Campylobacter spp., Verotoxigenic E.coli, Listeria monocytogenes, Clostridium botulinum, Staphylococcus aureus, Bacillus cereus, Clostridium perfringens, Giardia, Norovirus, enterotoxins of Staphylococcus aureus and Bacillus cereus and histamine.

National evaluation of the reported outbreaks in the country:

Trends in numbers of outbreaks and numbers of human cases involved

During 2016, a total of 377 outbreaks of food-borne infections and intoxications were recorded in Belgium. More than 2154 people were ill, and at least 78 persons were hospitalized. None of the human cases died. The number of reported outbreaks increased in 2011 as compared to former years but remains quite stable since then. The increase was probably due to an adapted Outbreak investigation procedure at the FASFC since 2011 and/or increased sensibility by consumers. The number of human cases involved are similar as in previous years, which is also the case for the number of people hospitalized due to a collective food borne outbreak.

Relevance of the different causative agents, food categories and the agent/food category combinations

In 2016, in total 12 verified food borne outbreaks were reported. In these outbreaks the causative agent was found in the implicated food and/or it was clear by analytical or strong descriptive epidemiology that food was at the origin of disease. All other outbreaks were classified as weak evidence outbreaks and the causative agent was unknown or the agent could be only detected at human level. Norovirus was the most frequently detected causative agent in 7 outbreaks and was responsible for 205 human cases. For 3 of these outbreaks, Norovirus was detected in the suspected food (oysters, swab from a meat cutter and source water) and confirmed in the human cases. The other 4 outbreaks were considered weak evidence outbreaks with Norovirus detection only in human cases but descriptive epidemiological evidence linking to a probable transmission by a food vehicle. The second most reported agent was enterotoxigenic Clostridium perfringens being at the origin of 4 outbreaks and involved 302 human cases. High levels of enterotoxigenic C. perfringens were present in the suspected foods (mainly stew or vol-au-vent) and in stools of human cases. Coagulase positive Staphylococcus (CPS) was at the origin of 2 outbreaks. In total 25 persons became ill and at least 2 were hospitalized. Staphylococcal enterotoxins (SEs) were detected in food leftovers (cheese) resulting in vomiting for 22 young children. For the second outbreak, enterotoxin producing CPS was isolated from cooked ham. Two outbreaks were reported involving Salmonella Enteritidis affecting 4 and 135 human cases, respectively. The first outbreak was due to the consumption of home-made chocolat mousse that contained raw eggs contaminated with Salmonella Enteritidis MLVA type 3-11-5-5-1. Two local outbreaks involving 24 human cases could be linked to the international outbreak of Salmonella Enteritidis 2-9-7-3-2 due to Polish eggs contaminated with this pathogen. Epidemiological investigation and Salmonella MLVA-typing of isolates from human cases revealed 111 more human cases spread over the whole country. E. coli O157:H7 was at the origin of 2 outbreaks involving 14 human cases. For one these, an indistinguishable agent could be identified in hard cheese and the human cases (3) but for the other outbreak, the food source remains unknown. Campylobacter was linked to 3 outbreaks and caused diarrhea in 6 human cases. Consumption of duck meat, chicken meat or bovin meat was probably at the origin of these outbreaks but could not be demonstrated. In 94.7% of the outbreaks (N=357 out of 377) no causative agent could be identified. An important reason for this is the absence of leftovers of the suspected meal in most of those outbreaks and late reporting by the consumer. Only in 23.3% (N=88 out of 377) of the outbreaks, samples (human and/or food) were send for analysis among which 22.7% (N=20) resulted in the detection of a pathogen. Some of the latter outbreaks (N=8) have been categorized as a weak evidence outbreak. Most food-borne outbreaks (49.1%) were due to the consumption of meals composed of different ingredients. Meat and meat based products (bovine, pig, sheep, broiler) were responsible for 17.2 % of the outbreaks. In 11.7% of the outbreaks the implicated food was unknown.

Relevance of the different type of places of food production and preparation in outbreaks

Restaurants and take away or fast food outlets were the most important location of exposure, being the setting of 54.9 % and 22.3 %, respectively, of food-borne outbreaks in Belgium in 2016. Catering at work, institutional catering or temporary mass catering are reported in respectively 1.9 %, 1.3 and 1.3 % of the food-borne outbreaks. 14.3 % of the outbreaks happened at home.

Descriptions of single outbreaks of special interest

In 2016, 115 visitors of a camping place suffered from abdominal pain, fever, diarrhea and vomiting. Norovirus GI and GII could be identified in human cases and in source water that was distributed to the camp visitors for food preparation and consumption. Norovirus was detected during different weeks until chlorination of the water to eliminate further spread of the virus. Two individual outbreaks, involving together 33 human cases, could be linked to the consumption of oysters contaminated with Norovirus GII.P2|GII.2. This genotype was also confirmed in the human cases. The oysters could be traced back to its production area in the Netherlands, where the same Norovirus genotype was confirmed. At the end of September 2016, two butchers associated with a cluster of cases (N=24) of Salmonella Enteritidis reporting purchasing meat in those locations were investigated. Food samples containing raw eggs resulted positive for Salmonella Enteritidis MLVA 2-9-7-3-2 profile. Both local outbreaks could be linked to the international outbreak of Salmonella Enteritidis MLVA 2-9-7-3-2 which was due to contaminated eggs from Poland. WGS analysis was successfully performed on food isolates with the MLVA profile 2-9-7-3-2 and, among these, four isolates belonged to WGS cluster_175 and two isolates belonged to cluster_360. Identical observations were made for the human isolates. Epidemiological investigations and MLVA-typing of human Salmonella Enteritidis isolates demonstrated 135 human cases linked to this outbreak. In July 2016, a 2.5 year old child suffering from HUS was reported. Two other family members also showed symptoms of diarrhea. E. coli O157 stx1 stx2 eae was isolated from the human cases and from hard raw milk cheese that they consumed. All isolates showed an identical PFGE profile and belonged to the IS629- type G profile. Interestingly, a E. coli O157 stx1 eae strain was isolated from a human case but not from the suspected food. Twenty-two children started vomiting shortly after the consumption of fresh cheese at school. Although no enterotoxigenic coagulase positive staphylococcus strain could be isolated, low levels of staphylococcal enterotoxins could be detected in leftovers of the cheese and in a new batch of fresh cheese.

Control measures or other actions taken to improve the situation

Logistic slaughtering is applied for poultry which means that poultry with a Salmonella-free certificate are slaughtered before other poultry. The vaccination of laying hens against salmonellosis, started in 2003 and is mandatory for Salmonella Enteritidis and is strongly recommended for Salmonella Typhimurium.

ANIMAL POPULATION TABLES

Table Susceptible animal population

			Population		
Animal species	Category of animals	holding	animal	slaughter animal (heads)	
Cattle (bovine animals)	Cattle (bovine animals)	28,466	2,672,669	544,952	
	Cattle (bovine animals) - calves (under 1 year) - veal calves			368,793	
Deer	Deer	2,357	8,289		
	Deer - farmed - fallow deer			930	
	Deer - wild - fallow deer			3,034	
	Deer - wild - red deer			10,974	
Ducks	Ducks			45,638	
Gallus gallus (fowl)	Gallus gallus (fowl)	1,547		310,239,434	
	Gallus gallus (fowl) - breeding flocks, unspecified		3,207,764		
	Gallus gallus (fowl) - broilers		32,815,269	281,439,890	
	Gallus gallus (fowl) - laying hens		8,719,228	27,871,407	
Geese	Geese			314	
Goats	Goats	10,117	57,553	12,860	
Guinea fowl	Guinea fowl			13,176	
Partridges	Partridges			8,333	
Pheasants	Pheasants			10,713	
Pigeons	Pigeons			23,668	
Pigs	Pigs	7,609		11,212,479	
	Pigs - breeding animals		5,318,631		
	Pigs - fattening pigs		503,118		
Quails	Quails			351	
Rabbits	Rabbits			2,910,926	
Ratites (ostrich, emu, nandu)	Ratites (ostrich, emu, nandu) - farmed			168	
Sheep	Sheep	26,676	194,687	142,589	
Solipeds, domestic	Solipeds, domestic		303,349	6,086	
Turkeys	Turkeys			783,583	
Wild boars	Wild boars - wild			11,507	

DISEASE STATUS TABLES

Table Bovine brucellosis in countries and regions that do not receive Community co-financing for eradication programme

_					Normalian of													
	Number of				Number of animals													Number of
	animals		Number of	Number of	positive in													animals
		Number of			microbiolog								Number of	Number of				tested by
	y tested	suspended	e animals	positive to	ical testing							Number of	animals or	infected				microbiolog
	under	herds under	under	BST under	under	Number of			Number of	Number of		herds	pools	herds	Number of		Number of	y under
	investigatio	investigatio	investigatio	investigatio	investigatio	herds with			herds	animals		tested	tested	tested	notified	Number of	abortions	investigatio
	ns of	status	Number of	Total	tested	tested	Total	under	under	under	abortions	isolations	due to	ns of				
	suspect	suspect	suspect	suspect	suspect	officially	infected	number of	under	under	number of	surveillance	surveillance	surveillance	whatever	of Brucella	Brucella	suspect
Region	cases	cases	cases	cases	cases	free	herds	animals	surveillance	surveillance	herds	by bulk milk	by bulk milk	by bulk milk	cause	infections	abortus	cases
BELGIUM	15	5 1	7	C) 0	28,466	0	2,672,669	12,178	72,355	28,46	6 7,701	16,370	0	10,209	0	C) 2

Table Ovine or Caprine brucellosis in countries and regions that do not receive Community co-financing for eradication programme

Region	Number of animals serologicall y tested under investigatio ns of suspect cases	suspended herds under	e animals under	Number of animals positive in microbiolog ical testing under investigatio ns of suspect cases	Number of herds with status officially free	Number of infected herds	Total number of animals	Number of animals tested under surveillance	Total number of herds	Number of animals tested by microbiolog y under investigatio ns of suspect cases
BELGIUM	138	0	0	0	36,793	0	252,240	6,955	36,793	85

DISEASE STATUS TABLES

Table Bovine tuberculosis in countries and regions that do not receive Community co-financing for eradication programme

_							Number of animals with		
							suspicious lesions of tuberculosis examined		
					Number of animals	Number of tuberculin	and submitted to histopathological and	Number of animals detected positive in	
Region	Number of herds with status officially free	Number of infected herds	Total number of animals	Interval between routine tuberculin tests	tested with tuberculin		bacteriological examinations	bacteriological examination	Total number of herds
BELGIUM	28,464	2	2,672,669	0	142,383	314,258	129	8	28,466

PREVALENCE TABLES

Table CAMPYLOBACTER in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit		Total units positive	Zoonoses	N of units positive
BELGIUM	Gallus gallus (fowl) - broilers - Slaughterhouse - Not Available - animal sample - caecum - Monitoring - Official sampling - Objective sampling	slaughte	496	236	Campylobacter jejuni	176
		r animal batch			Campylobacter, unspecified sp.	60

Table CAMPYLOBACTER in food

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Cheeses made from cows' milk - fresh - made from raw or low heat-treated milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	59	0	Campylobacter	0
	Cheeses made from cows' milk - fresh - made from raw or low heat-treated milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	24	1	Campylobacter	1
	Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	47	0	Campylobacter	0
	Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	25	0	Campylobacter	0
	Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	110	0	Campylobacter	0
	Live bivalve molluscs - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	80	0	Campylobacter	0
	Meat from bovine animals - carcase - Slaughterhouse - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1600	Square centimetre	386	0	Campylobacter	0
	Meat from bovine animals - meat preparation - intended to be eaten raw - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	24	0	Campylobacter	0
	Meat from bovine animals - meat preparation - intended to be eaten raw - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	34	1	Campylobacter	1
	Meat from bovine animals - minced meat - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	37	0	Campylobacter	0
	Meat from bovine animals and pig - meat preparation - intended to be eaten raw - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	2	0	Campylobacter	0
	Meat from bovine animals and pig - meat preparation - intended to be eaten raw - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	3	0	Campylobacter	0
	Meat from bovine animals and pig - minced meat - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	24	0	Campylobacter	0
	Meat from broilers (Gallus gallus) - carcase - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	91	8	Campylobacter	8
	Meat from broilers (Gallus gallus) - carcase - Slaughterhouse - Not Available - Not Available - Monitoring - Official sampling - Objective sampling	single (food/fee d)	1	Gram	873	174	Campylobacter	174
	Meat from broilers (Gallus gallus) - carcase - spent hens - Slaughterhouse - Not Available - Not Available - Monitoring - Official sampling - Objective sampling	single (food/fee d)	1	Gram	320	11	Campylobacter	11
	Meat from broilers (Gallus gallus) - fresh - skinned - Processing plant - Not Available - Not Available - Monitoring - Official sampling - Objective sampling	single (food/fee d)	1	Gram	307	11	Campylobacter	11

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Meat from broilers (Gallus gallus) - fresh - skinned - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	45	0	Campylobacter	0
	Meat from broilers (Gallus gallus) - fresh - with skin - Processing plant - Not Available - Not Available - Monitoring - Official sampling - Objective sampling	single (food/fee d)	1	Gram	368	14	Campylobacter	14
	Meat from broilers (Gallus gallus) - fresh - with skin - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	46	1	Campylobacter	1
	Meat from other animal species or not specified - minced meat - intended to be eaten cooked - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	45	0	Campylobacter	0
	Meat from other animal species or not specified - minced meat - intended to be eaten raw - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	46	0	Campylobacter	0
	Meat from other poultry species - meat preparation - intended to be eaten cooked - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	46	0	Campylobacter	0
	Meat from pig - meat preparation - intended to be eaten raw - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	24	0	Campylobacter	0
	Meat from pig - meat preparation - intended to be eaten raw - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	9	0	Campylobacter	0
	Meat from pig - minced meat - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	34	0	Campylobacter	0
	Meat from poultry, unspecified - meat preparation - intended to be eaten cooked - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	50	6	Campylobacter	6
	Meat from poultry, unspecified - meat products - cooked, ready-to-eat - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	50	1	Campylobacter	1
	Milk, cows' - raw milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Millilitre	35	0	Campylobacter	0
	Other processed food products and prepared dishes - unspecified - ready-to-eat foods - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1	Gram	111	0	Campylobacter	0

Table COXIELLA in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Total units tested	Total units positive	N of clinical affected herds	Zoonoses	N of units positive
BELGIUM	Cattle (bovine animals) - adult cattle over 2 years - Farm - Not Available - animal sample - blood - Clinical investigations - Industry sampling - Suspect sampling	animal	1148	209		Coxiella burnetii	209
	Cattle (bovine animals) - adult cattle over 2 years - Farm - Not Available - animal sample - faeces - Monitoring - passive - Industry sampling - Suspect sampling	animal	3911	110		Coxiella burnetii	110
	Cattle (bovine animals) - adult cattle over 2 years - Farm - Not Available - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling	animal	50	28		Coxiella burnetii	28
	Cattle (bovine animals) - adult cattle over 2 years - Farm - Not Available - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling	animal	70	63		Coxiella burnetii	63
	Cattle (bovine animals) - adult cattle over 2 years - Farm - Not Available - animal sample - milk - Monitoring - passive - Industry sampling - Suspect sampling	animal	2	2		Coxiella burnetii	2
	Cattle (bovine animals) - adult cattle over 2 years - Farm - Not Available - animal sample - organ/tissue - Monitoring - passive - Industry sampling - Suspect sampling	animal	61	5		Coxiella burnetii	5
	Cattle (bovine animals) - adult cattle over 2 years - Farm - Not Available - animal sample - placental swab - Monitoring - passive - Industry sampling - Suspect sampling	animal	82	27		Coxiella burnetii	27
	Cattle (bovine animals) - adult cattle over 2 years - Farm - Not Available - animal sample - vaginal swab - Monitoring - passive - Industry sampling - Suspect sampling	animal	40	12		Coxiella burnetii	12
	Goats - animals over 1 year - Farm - Not Available - animal sample - blood - Clinical investigations - Industry sampling - Suspect sampling	animal	29	1		Coxiella burnetii	1
	Goats - animals over 1 year - Farm - Not Available - animal sample - blood - Monitoring - passive - Industry sampling - Suspect sampling	animal	3	1		Coxiella burnetii	1
	Goats - animals over 1 year - Farm - Not Available - animal sample - faeces - Monitoring - passive - Industry sampling - Suspect sampling	animal	15	0		Coxiella	0
	Goats - animals over 1 year - Farm - Not Available - animal sample - vaginal swab - Monitoring - passive - Industry sampling - Suspect sampling	animal	1	1		Coxiella burnetii	1
	Goats - milk goats - Farm - Not Available - animal sample - milk - Surveillance - Official sampling - Census	holding	118	15	0	Coxiella burnetii	15
	Goats - milk goats - Farm - Not Available - animal sample - milk - Surveillance - Official sampling - Census	holding	118	37	0	Coxiella burnetii	37
	Sheep - animals over 1 year - Farm - Not Available - animal sample - blood - Clinical investigations - Industry sampling - Suspect sampling	animal	21	0		Coxiella	0
	Sheep - animals over 1 year - Farm - Not Available - animal sample - blood - Monitoring - passive - Industry sampling - Suspect sampling	animal	30	0		Coxiella	0
	Sheep - animals over 1 year - Farm - Not Available - animal sample - faeces - Monitoring - passive - Industry sampling - Suspect sampling	animal	43	2		Coxiella burnetii	2
]	Sheep - animals over 1 year - Farm - Not Available - animal sample - placental swab - Monitoring - passive - Industry sampling - Suspect sampling	animal	1	0		Coxiella	0
	Sheep - animals over 1 year - Farm - Not Available - animal sample - vaginal swab - Monitoring - passive - Industry sampling - Suspect sampling	animal	3	0		Coxiella	0
	Sheep - milk ewes - Farm - Not Available - animal sample - milk - Surveillance - Official sampling - Census	holding	21	1	0	Coxiella burnetii	1
	Sheep - milk ewes - Farm - Not Available - animal sample - milk - Surveillance - Official sampling - Census	holding	21	1	0	Coxiella burnetii	1

Table CRONOBACTER in food

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Foodstuffs intended for special nutritional uses - dried dietary foods for special medical purposes intended for infants below 6 months - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	10	Gram	147	0	Cronobacter	0
	Infant formula - dried - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	10	Gram	6	0	Cronobacter	0
	Infant formula - dried - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	10	Gram	284	0	Cronobacter	0
	Infant formula - ready-to-eat - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	10	Millilitre	103	0	Cronobacter	0

Table CYSTICERCUS in animal

			Total	Total		
		Sampling	units	units		N of units
Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	unit	tested	positive	Zoonoses	positive
Not Available	Cattle (bovine animals) - Slaughterhouse - Not Available - Not Available - Surveillance - Official sampling - Suspect sampling	animal	91374	1282	Cysticercus of Taenia saginata	1,282
			5			

Table ECHINOCOCCUS in animal

		Sampling	Total units	Total units		N of units
Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy				Zoonoses	positive
Not Available	Cattle (bovine animals) - Slaughterhouse - Not Available - Not Available - Surveillance - Official sampling - Suspect sampling	animal	91374 5	0	Echinococcus	0

Table ESCHERICHIA COLI in food

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Cheeses made from cows' milk - fresh - made from raw or low heat-treated milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	54	0	Verocytotoxigenic E. coli (VTEC)	0
	Cheeses made from cows' milk - fresh - made from raw or low heat-treated milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	24	1	VTEC 0157	1
	Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	47	0	Verocytotoxigenic E. coli (VTEC)	0
	Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	25	0	Verocytotoxigenic E. coli (VTEC)	0
	Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	381	0	Verocytotoxigenic E. coli (VTEC)	0
	Cheeses made from goats' milk - unspecified - made from raw or low heat-treated milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	25	0	Verocytotoxigenic E. coli (VTEC)	0
	Cheeses made from goats' milk - unspecified - made from raw or low heat-treated milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	25	1	VTEC O103	1
	Cheeses made from goats' milk - unspecified - made from raw or low heat-treated milk - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	100	0	Verocytotoxigenic E. coli (VTEC)	0
	Cheeses made from sheep's milk - fresh - made from raw or low heat-treated milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	3	0	Verocytotoxigenic E. coli (VTEC)	0
	Cheeses made from sheep's milk - fresh - made from raw or low heat-treated milk - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	25	Gram	71	2	VTEC O103	1
		d)					VTEC, unspecified	1
	Cheeses made from sheep's milk - unspecified - made from raw or low heat-treated milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	2	1	VTEC, unspecified	1
	Dairy products (excluding cheeses) - butter - made from raw or low heat-treated milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	34	1	VTEC, unspecified	1
	Dairy products (excluding cheeses) - butter - made from raw or low heat-treated milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	5	0	Verocytotoxigenic E. coli (VTEC)	0
	Fruits and vegetables - pre-cut - ready-to-eat - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	97	0	Verocytotoxigenic E. coli (VTEC)	0
	Fruits and vegetables - pre-cut - ready-to-eat - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	101	0	Verocytotoxigenic E. coli (VTEC)	0
	Meat from bovine animals - Border inspection activities - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	1	0	Verocytotoxigenic E. coli (VTEC)	0
	Meat from bovine animals - carcase - Slaughterhouse - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	1600	Square centimetre	456	8	VTEC, unspecified	8

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Meat from bovine animals - fresh - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	294	5	VTEC, unspecified	5
	Meat from bovine animals - meat preparation - intended to be eaten cooked - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	17	0	Verocytotoxigenic E. coli (VTEC)	0
	Meat from bovine animals - meat preparation - intended to be eaten raw - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	313	3	VTEC, unspecified	3
	Meat from bovine animals - meat preparation - intended to be eaten raw - Retail - Not Available -	single	25	Gram	203	7	VTEC 0157	1
	Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					VTEC, unspecified	6
	Meat from sheep - Border inspection activities - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	1	0	Verocytotoxigenic E. coli (VTEC)	0
	Milk, cows' - raw milk - Farm - Not Available - Not Available - Surveillance - Official sampling -	single	25	Millilitre	328	4	VTEC O103	1
	Objective sampling	(food/fee d)					VTEC O26	1
		u)					VTEC, unspecified	2
	Seeds, sprouted - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	45	0	Verocytotoxigenic E. coli (VTEC)	0
	Seeds, sprouted - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	87	0	Verocytotoxigenic E. coli (VTEC)	0
	Seeds, sprouted - Wholesale - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	80	0	Verocytotoxigenic E. coli (VTEC)	0
	Spices and herbs - fresh - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	68	0	Verocytotoxigenic E. coli (VTEC)	0
	Spices and herbs - fresh - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	5	0	Verocytotoxigenic E. coli (VTEC)	0
	Spices and herbs - fresh - Wholesale - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	63	0	Verocytotoxigenic E. coli (VTEC)	0
	Vegetables - leaves - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	57	0	Verocytotoxigenic E. coli (VTEC)	0
	Vegetables - leaves - Wholesale - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	57	0	Verocytotoxigenic E. coli (VTEC)	0
	Vegetables - non-pre-cut - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	57	0	Verocytotoxigenic E. coli (VTEC)	0
	Vegetables - non-pre-cut - Wholesale - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	56	0	Verocytotoxigenic E. coli (VTEC)	0

Table FLAVIVIRUS in animal

	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Vaccination status	Total units tested	Total units positive	Zoonoses	N of units positive
BELGIUM	Birds - wild - Natural habitat - Not Available - animal sample - organ/tissue - Monitoring - passive - Official sampling - Suspect sampling	animal	No	85	0	Flavivirus	0

Table HISTAMINE in food

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	•	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Fish - Fishery products from fish species associated with a high amount of histidine - not	batch	1	Gram	45	0	<= 100	Histamine	0	45
	enzyme maturated - Border inspection activities - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100 TO <= 200	Histamine	0	0
							>200	Histamine	0	0
	Fish - Fishery products from fish species associated with a high amount of histidine - not	batch	1	Gram	135	0	<= 100	Histamine	0	135
	enzyme maturated - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100 TO <= 200	Histamine	0	0
							>200	Histamine	0	0
	Fish - Fishery products from fish species associated with a high amount of histidine - not	single	1	Gram	21	0	<= 100	Histamine	0	21
	enzyme maturated - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100 TO <= 200	Histamine	0	0
							>200	Histamine	0	0
е	Fish - Fishery products from fish species associated with a high amount of histidine - not	batch	1	Gram	am 27 0 <u> </u>	<= 100	Histamine	0	27	
	enzyme maturated - Wholesale - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100 TO <= 200	Histamine	0	0
							>200	Histamine	0	0

Table LISTERIA in food

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight		Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Bakery products - desserts - containing raw eggs - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	25	0	<= 100	Listeria monocytogenes	25	0
	Surveillance - Official sampling - Objective sampling	d)					>100	Listeria monocytogenes	25	0
	Bakery products - pastry - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	75	0	<= 100	Listeria monocytogenes	65	0
	Cilicial Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	65	0
	Bakery products - pastry - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	75	0	detection	Listeria monocytogenes	10	0
	Bakery products - pastry - Retail - Not Available - Not Available - Surveillance - Official	single	1	Gram	148	0	<= 100	Listeria monocytogenes	148	0
	sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	148	0
	Cheeses made from cows' milk - fresh - made from pasteurised milk - Farm - Not Available -	single	1	Gram	45	0	<= 100	Listeria monocytogenes	12	0
	Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	12	0
	Cheeses made from cows' milk - fresh - made from pasteurised milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	45	0	detection	Listeria monocytogenes	33	0
-	Cheeses made from cows' milk - fresh - made from pasteurised milk - Processing plant - Not	single	1	Gram	68	0	<= 100	Listeria monocytogenes	24	0
	Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	24	0
	Cheeses made from cows' milk - fresh - made from pasteurised milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	68	0	detection	Listeria monocytogenes	44	0
	Cheeses made from cows' milk - fresh - made from pasteurised milk - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	117	0	<= 100	Listeria monocytogenes	117	0
	Not Available - Surveillance - Official Sampling - Objective Sampling	d)				2	>100	Listeria monocytogenes	117	0
	Cheeses made from cows' milk - fresh - made from raw or low heat-treated milk - Farm - Not single	single (food/fee	1	Gram	77	0	<= 100	Listeria monocytogenes	34	0
	Available - Not Available - Surveillance - Official Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	34	0
	Cheeses made from cows' milk - fresh - made from raw or low heat-treated milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	77	0	detection	Listeria monocytogenes	43	0
	Cheeses made from cows' milk - fresh - made from raw or low heat-treated milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	23	1	<= 100	Listeria monocytogenes	7	0
	plant - Not Avaliable - Not Avaliable - Surveillance - Official Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	7	0
	Cheeses made from cows' milk - fresh - made from raw or low heat-treated milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	23	1	detection	Listeria monocytogenes	16	1
	Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Farm - Not		1	Gram 2 1 <=	<= 100	Listeria monocytogenes	1	0		
	Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	1	0
	Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	2	1	detection	Listeria monocytogenes	1	1
	Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing		1	Gram	135	1	<= 100	Listeria monocytogenes	32	0
	plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	32	0
	Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	135	1	detection	Listeria monocytogenes	103	1

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Retail -	single	1	Gram	327	0	<= 100	Listeria monocytogenes	327	0
	Not Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	327	0
	Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk -	single	1	Gram	69	1	<= 100	Listeria monocytogenes	24	0
	Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	24	0
	Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	69	1	detection	Listeria monocytogenes	45	1
	Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk -	single	1	Gram	25	1	<= 100	Listeria monocytogenes	5	0
	Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	5	0
	Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	25	1	detection	Listeria monocytogenes	20	1
	Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk -	single (food/fee	1	Gram	380	0	<= 100	Listeria monocytogenes	380	0
	Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	d)					>100	Listeria monocytogenes	380	0
	Cheeses made from goats' milk - unspecified - made from pasteurised milk - Farm - Not	single	1	Gram	1	0	<= 100	Listeria monocytogenes	1	0
	Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	1	0
	Cheeses made from goats' milk - unspecified - made from pasteurised milk - Processing plant	single	1	Gram	35	0	<= 100	Listeria monocytogenes	16	0
	- Not Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	16	0
	Cheeses made from goats' milk - unspecified - made from pasteurised milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	35	0	detection	Listeria monocytogenes	19	0
	Cheeses made from goats' milk - unspecified - made from pasteurised milk - Retail - Not	single	1		95	0	<= 100	Listeria monocytogenes	95	0
	Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	95	0
	Cheeses made from goats' milk - unspecified - made from raw or low heat-treated milk - Far	single	1	Gram	33	0	<= 100	Listeria monocytogenes	11	0
	- Not Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	11	0
	Cheeses made from goats' milk - unspecified - made from raw or low heat-treated milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	33	0	detection	Listeria monocytogenes	22	0
	Cheeses made from goats' milk - unspecified - made from raw or low heat-treated milk -	single	1	Gram	30	1	<= 100	Listeria monocytogenes	14	0
	Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	14	1
	Cheeses made from goats' milk - unspecified - made from raw or low heat-treated milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	30	1	detection	Listeria monocytogenes	16	0
	Cheeses made from goats' milk - unspecified - made from raw or low heat-treated milk - Retail			Gram 100 0 <=	<= 100	Listeria monocytogenes	100	0		
	- Not Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	100	0
	Cheeses made from sheep's milk - fresh - made from pasteurised milk - Retail - Not Available	single	1	Gram	74	0	<= 100	Listeria monocytogenes	74	0
	- Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	74	0
	Cheeses made from sheep's milk - fresh - made from raw or low heat-treated milk -	single	1	Gram	3	0	<= 100	Listeria monocytogenes	2	0
	Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	d)	(food/fee d)	>100	Listeria monocytogenes	2	0			
	Cheeses made from sheep's milk - fresh - made from raw or low heat-treated milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	3	0	detection	Listeria monocytogenes	1	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Cheeses made from sheep's milk - fresh - made from raw or low heat-treated milk - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	71	0	<= 100	Listeria monocytogenes	71	0
	Not Available - Not Available - Surveillance - Official Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	71	0
	Cheeses made from sheep's milk - unspecified - made from pasteurised milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	5	0	<= 100	Listeria monocytogenes	1	0
	plant Not/Wallable Not/Wallable Surveillance Similar sampling Specific sampling	d)					>100	Listeria monocytogenes	1	0
	Cheeses made from sheep's milk - unspecified - made from pasteurised milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	5	0	detection	Listeria monocytogenes	4	0
	Cheeses made from sheep's milk - unspecified - made from raw or low heat-treated milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	3	0	<= 100	Listeria monocytogenes	1	0
	- ann - Not Available - Not Available - Surveillance - Official Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	1	0
	Cheeses made from sheep's milk - unspecified - made from raw or low heat-treated milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	3	0	detection	Listeria monocytogenes	2	0
	Dairy products (excluding cheeses) - butter - made from pasteurised milk - Farm - Not	single	1	Gram	7	0	<= 100	Listeria monocytogenes	1	0
	Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	1	0
	Dairy products (excluding cheeses) - butter - made from pasteurised milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	7	0	detection	Listeria monocytogenes	6	0
	Dairy products (excluding cheeses) - butter - made from pasteurised milk - Retail - Not	single	1	Gram	92	0	<= 100	Listeria monocytogenes	92	0
	Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	92	0
	Dairy products (excluding cheeses) - butter - made from raw or low heat-treated milk - Farm - Single Not Available - Not Available - Surveillance - Official sampling - Objective sampling (food/fe d)		1	Gram	n 130	8 0	<= 100	Listeria monocytogenes	6	0
							>100	Listeria monocytogenes	6	0
	Dairy products (excluding cheeses) - butter - made from raw or low heat-treated milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	130	8	detection	Listeria monocytogenes	124	8
	Dairy products (excluding cheeses) - butter - made from raw or low heat-treated milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	30	15	detection	Listeria monocytogenes	30	15
	Dairy products (excluding cheeses) - dairy desserts - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	52	0	<= 100	Listeria monocytogenes	41	0
		d)					>100	Listeria monocytogenes	41	0
	Dairy products (excluding cheeses) - dairy desserts - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	52	0	detection	Listeria monocytogenes	11	0
	Dairy products (excluding cheeses) - dairy desserts - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	50	0	<= 100	Listeria monocytogenes	17	0
	Available - Surveillance - Official Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	17	0
	Dairy products (excluding cheeses) - dairy desserts - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	50	0	detection	Listeria monocytogenes	33	0
	Dairy products (excluding cheeses) - dairy desserts - Retail - Not Available - Not Available -	single	1	Gram	117	0	<= 100	Listeria monocytogenes	117	0
	Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	117	0
	lairy products (excluding cheeses) - fermented dairy products - fermented milk - Processing singulant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling (for		1	Gram	20	0	<= 100	Listeria monocytogenes	20	0
	plant - Not Available - Not Available - Outveillance - Ontolat sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	20	0
	Dairy products (excluding cheeses) - fermented dairy products - fermented milk - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	57	0	<= 100	Listeria monocytogenes	57	0
	Additional Total Manager Control Control Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	57	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Dairy products (excluding cheeses) - ice-cream - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	50	0	<= 100	Listeria monocytogenes	50	0
	Surveillance - Official sampling - Objective sampling	d)					>100	Listeria monocytogenes	50	0
	Dairy products (excluding cheeses) - ice-cream - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	31	0	<= 100	Listeria monocytogenes	29	0
	Available - Surveillance - Official sampling - Objective sampling	d)					>100	Listeria monocytogenes	29	0
	Dairy products (excluding cheeses) - ice-cream - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	31	0	detection	Listeria monocytogenes	2	0
	Dairy products (excluding cheeses) - ice-cream - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	152	0	<= 100	Listeria monocytogenes	152	0
	Surveillance - Official sampling - Objective sampling	d)					>100	Listeria monocytogenes	152	0
	Dairy products (excluding cheeses) - yoghurt - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	46	0	<= 100	Listeria monocytogenes	46	0
	Surveillance - Official sampling - Objective sampling	d)					>100	Listeria monocytogenes	46	0
	Dairy products (excluding cheeses) - yoghurt - Processing plant - Not Available - Not	single	1	Gram	35	0	<= 100	Listeria monocytogenes	35	0
	Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	35	0
	Dairy products (excluding cheeses) - yoghurt - Retail - Not Available - Not Available -	single	1	Gram	70	0	<= 100	Listeria monocytogenes	70	0
	Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	70	0
	Fish - gravad /slightly salted - Processing plant - Not Available - Not Available - Surveillance -	single	1	Gram	10	0	<= 100	Listeria monocytogenes	5	0
	Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	5	0
	Fish - gravad /slightly salted - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	10	0	detection	Listeria monocytogenes	5	0
	Fish - gravad /slightly salted - Retail - Not Available - Not Available - Surveillance - Official	single		Gram	15	0	<= 100	Listeria monocytogenes	15	0
	sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	15	0
	Fish - smoked - Processing plant - Not Available - Not Available - Surveillance - Official	single	1	Gram	65	1	<= 100	Listeria monocytogenes	34	0
	sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	34	0
	Fish - smoked - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	65	1	detection	Listeria monocytogenes	31	1
	Fish - smoked - Retail - Not Available - Not Available - Surveillance - Official sampling -	single	1	Gram	142	0	<= 100	Listeria monocytogenes	142	0
	Objective sampling	(food/fee d)					>100	Listeria monocytogenes	142	0
	Fishery products, unspecified - ready-to-eat - chilled - Processing plant - Not Available - Not	single	1	Gram	199	7	<= 100	Listeria monocytogenes	113	0
	Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	113	0
	Fishery products, unspecified - ready-to-eat - chilled - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	199	7	detection	Listeria monocytogenes	86	7
	Fishery products, unspecified - ready-to-eat - chilled - Retail - Not Available - Not Available -	single	1	Gram	274	0	<= 100	Listeria monocytogenes	274	0
	Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	274	0
	Fishery products, unspecified - ready-to-eat - Retail - Not Available - Not Available -	single	1	Millilitre	227	0	<= 100	Listeria monocytogenes	227	0
	Surveillance - Official sampling - Objective sampling	- Objective sampling (food/fee d)			>100	Listeria monocytogenes	227	0		
	Follow-on formulae - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	8	0	detection	Listeria monocytogenes	8	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Follow-on formulae - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	278	0	detection	Listeria monocytogenes	278	0
	Foodstuffs intended for special nutritional uses - dried dietary foods for special medical purposes intended for infants below 6 months - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	157	1	detection	Listeria monocytogenes	157	1
	Fruits - non-pre-cut - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	51	0	<= 100	Listeria monocytogenes	51	0
	Objective sampling	d)					>100	Listeria monocytogenes	51	0
	Fruits - non-pre-cut - frozen - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	61	0	<= 100	Listeria monocytogenes	61	0
	Official Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	61	0
	Fruits - non-pre-cut - frozen - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	53	0	<= 100	Listeria monocytogenes	53	0
	Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	53	0
	Fruits - non-pre-cut - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	52	0	<= 100	Listeria monocytogenes	52	0
	Objective sampling	d)					>100	Listeria monocytogenes	52	0
	Fruits and vegetables - pre-cut - ready-to-eat - Processing plant - Not Available - Not	single	1	Gram	116	0	<= 100	Listeria monocytogenes	51	0
	Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	51	0
	Fruits and vegetables - pre-cut - ready-to-eat - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	116	0	detection	Listeria monocytogenes	65	0
	Fruits and vegetables - pre-cut - ready-to-eat - Retail - Not Available - Not Available - single 1 Surveillance - Official sampling - Objective sampling d)	1	Gram	113	0	<= 100	Listeria monocytogenes	113	0	
						>100	Listeria monocytogenes	113	0	
	Infant formula - dried - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	6	0	detection	Listeria monocytogenes	6	0
	Infant formula - dried - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	284	0	detection	Listeria monocytogenes	284	0
	Infant formula - ready-to-eat - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Millilitre	103	0	detection	Listeria monocytogenes	103	0
	Juice - fruit juice - pasteurised - Processing plant - Not Available - Not Available - Surveillance		1	Millilitre	5	0	<= 100	Listeria monocytogenes	5	0
	- Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	5	0
	Juice - fruit juice - pasteurised - Retail - Not Available - Not Available - Surveillance - Official	single	1	Millilitre	14	0	<= 100	Listeria monocytogenes	14	0
	sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	14	0
	Juice - fruit juice - unpasteurised - Processing plant - Not Available - Not Available -	single	25	Millilitre	9	0	<= 100	Listeria monocytogenes	9	0
	Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	9	0
	Juice - fruit juice - unpasteurised - Retail - Not Available - Not Available - Surveillance -	· · · · · · · · · · · · · · · · · · ·		143	0	<= 100	Listeria monocytogenes	143	0	
	Official sampling - Objective sampling						>100	Listeria monocytogenes	143	0
	e - vegetable juice - pasteurised - Processing plant - Not Available - Not Available - single 1 Millilitre 5		0	<= 100	Listeria monocytogenes	5	0			
	Surveillance - Official sampling - Objective sampling	Official sampling - Objective sampling (food/fee d)					>100	Listeria monocytogenes	5	0
	Juice - vegetable juice - pasteurised - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single	1	Millilitre 6	6	0	<= 100	Listeria monocytogenes	6	0
	Onicial sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	6	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Juice - vegetable juice - unpasteurised - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	25	Millilitre	1	0	<= 100	Listeria monocytogenes	1	0
	Surveinance - Official Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	1	0
	Juice - vegetable juice - unpasteurised - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Millilitre	15	0	<= 100	Listeria monocytogenes	15	0
	Official Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	15	0
	Meat from bovine animals - meat preparation - intended to be eaten cooked - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	17	0	<= 100	Listeria monocytogenes	17	0
	Available - Not Available - Surveillance - Official Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	17	0
	Meat from bovine animals - meat preparation - intended to be eaten raw - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	338	20	<= 100	Listeria monocytogenes	165	0
	Not Available - Not Available - Surveillance - Official Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	165	0
	Meat from bovine animals - meat preparation - intended to be eaten raw - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	338	20	detection	Listeria monocytogenes	173	20
	Meat from bovine animals - meat preparation - intended to be eaten raw - Retail - Not	single	1	Gram	295	0	<= 100	Listeria monocytogenes	295	0
	Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	295	0
	Meat from bovine animals - minced meat - Processing plant - Not Available - Not Available -	single	1	Gram	47	4	<= 100	Listeria monocytogenes	13	0
	Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	13	0
	Meat from bovine animals - minced meat - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	47	4	detection	Listeria monocytogenes	34	4
	Meat from bovine animals and pig - meat preparation - intended to be eaten raw - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling (food/fee d)	1	Gram	11	1	<= 100	Listeria monocytogenes	7	0	
						>100	Listeria monocytogenes	7	0	
	Meat from bovine animals and pig - meat preparation - intended to be eaten raw - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	11	1	detection	Listeria monocytogenes	4	1
	Meat from bovine animals and pig - meat preparation - intended to be eaten raw - Retail - Not	single	1	Gram	5	0	<= 100	Listeria monocytogenes	5	0
	Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	5	0
	Meat from bovine animals and pig - minced meat - Processing plant - Not Available - Not	single	1	Gram	29	3	<= 100	Listeria monocytogenes	12	0
	Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	12	0
	Meat from bovine animals and pig - minced meat - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	29	3	detection	Listeria monocytogenes	17	3
	Meat from other animal species or not specified - meat products - cooked, ready-to-eat -	single	1	Gram	389	21	<= 100	Listeria monocytogenes	131	0
	Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	131	1
	Meat from other animal species or not specified - meat products - cooked, ready-to-eat - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	389	21	detection	Listeria monocytogenes	258	20
	Meat from other animal species or not specified - meat products - cooked, ready-to-eat - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single	1	Gram	296	1	<= 100	Listeria monocytogenes	296	0
	Retail - Not Available - Not Available - Sul veillance - Official Sampling - Objective Sampling	(food/fee d)					>100	Listeria monocytogenes	296	1
	Meat from other animal species or not specified - meat products - fermented sausages - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective			Gram	113	0	<= 100	Listeria monocytogenes	113	0
	sampling sampling	d)			>100	Listeria monocytogenes	113	0		
	Meat from other animal species or not specified - meat products - fermented sausages - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	114	0	<= 100	Listeria monocytogenes	114	0
	Notali Not Manable - Not Available - Ourveillance - Onicial Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	114	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Meat from other animal species or not specified - meat products - pâté - Processing plant - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	121	3	<= 100	Listeria monocytogenes	27	0
	Not Available - Not Available - Surveillance - Official Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	27	0
	Meat from other animal species or not specified - meat products - pâté - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	121	3	detection	Listeria monocytogenes	94	3
	Meat from other animal species or not specified - meat products - pâté - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	115	0	<= 100	Listeria monocytogenes	115	0
	- Not Available - Surveillance - Official Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	115	0
	Meat from other animal species or not specified - minced meat - intended to be eaten raw - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	112	0	<= 100	Listeria monocytogenes	112	0
	Tretail - Not Available - Not Available - Our veillance - Official Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	112	0
	Meat from other poultry species - meat products - cooked, ready-to-eat - chilled - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	108	1	<= 100	Listeria monocytogenes	32	0
	plant - Not Available - Not Available - Surveillance - Official Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	32	0
	Meat from other poultry species - meat products - cooked, ready-to-eat - chilled - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	108	1	detection	Listeria monocytogenes	76	1
	Meat from pig - meat preparation - intended to be eaten raw - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	52	7	<= 100	Listeria monocytogenes	35	0
	- Not Available - Surveillance - Official Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	35	0
	Meat from pig - meat preparation - intended to be eaten raw - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	52	7	detection	Listeria monocytogenes	17	7
	Available - Surveillance - Official sampling - Objective sampling (food/fee d)		1	Gram	17	0	<= 100	Listeria monocytogenes	17	0
					>100	Listeria monocytogenes	17	0		
	Meat from pig - meat products - cooked ham - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1 Gram 114	114	1	<= 100	Listeria monocytogenes	31	0	
	- Surveillance - Official sampling - Objective sampling	d)					>100	Listeria monocytogenes	31	0
	Meat from pig - meat products - cooked ham - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	114	1	detection	Listeria monocytogenes	83	1
	Meat from pig - meat products - cooked ham - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	113	0	<= 100	Listeria monocytogenes	113	0
	Surveillance - Official Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	113	0
	Meat from pig - meat products - raw ham - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	116	0	<= 100	Listeria monocytogenes	85	0
	Surveillance - Official Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	85	0
	Meat from pig - meat products - raw ham - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	116	0	detection	Listeria monocytogenes	31	0
	Meat from pig - meat products - raw ham - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	114	1	<= 100	Listeria monocytogenes	114	0
	- Onicial Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	114	1
	Meat from pig - minced meat - Processing plant - Not Available - Not Available - Surveillance -	single	1	Gram	50	5	<= 100	Listeria monocytogenes	18	0
	Official sampling - Objective sampling (food/fee d)	>100	Listeria monocytogenes	18	0					
	Meat from pig - minced meat - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	50	5	detection	Listeria monocytogenes	32	5
	Meat from poultry, unspecified - meat products - cooked, ready-to-eat - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	219	0	<= 100	Listeria monocytogenes	219	0
	Not Available - Outvellance - Official sampling - Objective sampling	d)					>100	Listeria monocytogenes	219	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Milk, cows' - raw milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Millilitre	34	1	<= 100	Listeria monocytogenes	34	1
	Objective Sampling	d)					>100	Listeria monocytogenes	34	0
	Other food - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	73	0	<= 100	Listeria monocytogenes	48	0
	- Objective sampling	d)					>100	Listeria monocytogenes	48	0
	Other food - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	73	0	detection	Listeria monocytogenes	25	0
	Other food - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective	single (food/fee	1	Gram	150	0	<= 100	Listeria monocytogenes	150	0
	sampling	d)					>100	Listeria monocytogenes	150	0
	Other processed food products and prepared dishes - unspecified - ready-to-eat foods -	single (food/fee	1	Gram	89	1	<= 100	Listeria monocytogenes	42	0
	Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	(100d/lee d)					>100	Listeria monocytogenes	42	0
	Other processed food products and prepared dishes - unspecified - ready-to-eat foods - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	89	1	detection	Listeria monocytogenes	47	1
	Other processed food products and prepared dishes - unspecified - ready-to-eat foods - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single	1	Gram	1026	1	<= 100	Listeria monocytogenes	1,026	0
	- Not Available - Not Available - Surveillance - Official Sampling - Objective Sampling	(food/fee d)					>100	Listeria monocytogenes	1,026	1
	Other products of animal origin - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	6	0	detection	Listeria monocytogenes	6	0
	Other products of animal origin - Retail - Not Available - Not Available - Surveillance - Official single 1 sampling - Objective sampling (food/fee	1	Gram	31	0	<= 100	Listeria monocytogenes	31	0	
	sampling - Objective sampling	d)	d/fee		>100	Listeria monocytogenes	31	0		
	Seeds, sprouted - Farm - Not Available - Not Available - Surveillance - Official sampling -	single	1	1 Gram 80	80	0	<= 100	Listeria monocytogenes	76	0
	Objective sampling	(food/fee d)					>100	Listeria monocytogenes	76	0
	Seeds, sprouted - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	80	0	detection	Listeria monocytogenes	4	0
	Seeds, sprouted - Processing plant - Not Available - Not Available - Surveillance - Official	single	1	Gram	46	0	<= 100	Listeria monocytogenes	38	0
	sampling - Objective sampling	(food/fee d)					>100	Listeria monocytogenes	38	0
	Seeds, sprouted - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	46	0	detection	Listeria monocytogenes	8	0
	Seeds, sprouted - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	84	0	<= 100	Listeria monocytogenes	84	0
	Objective Sampling	d)					>100	Listeria monocytogenes	84	0
	Surimi - Border inspection activities - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	3	0	<= 100	Listeria monocytogenes	3	0
	sampling - Objective sampling	d)					>100	Listeria monocytogenes	3	0
	Surimi - Processing plant - Not Available - Not Available - Surveillance - Official sampling -	single	1	Gram	10	0	<= 100	Listeria monocytogenes	9	0
	Objective sampling	(food/fee d)			>100	Listeria monocytogenes	9	0		
	Surimi - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	10	0	detection	Listeria monocytogenes	1	0
	Surimi - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective	single (food/fee	1	Gram	31	0	<= 100	Listeria monocytogenes	31	0
	sampling	d)					>100	Listeria monocytogenes	31	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight		Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Not Available	Vegetables - leaves - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	37	0	<= 100	Listeria monocytogenes	37	0
	Objective sampling	d)					>100	Listeria monocytogenes	37	0
	Vegetables - leaves - Retail - Not Available - Not Available - Surveillance - Official sampling -	single	1	Gram	35	2	<= 100	Listeria monocytogenes	35	0
	Objective sampling	(food/fee d)					>100	Listeria monocytogenes	35	2
	Vegetables - non-pre-cut - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	36	0	<= 100	Listeria monocytogenes	36	0
	Sampling - Objective Sampling	d)					>100	Listeria monocytogenes	36	0
	Vegetables - non-pre-cut - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	37	0	<= 100	Listeria monocytogenes	37	0
	sampling - Objective sampling	d)					>100	Listeria monocytogenes	37	0

Table LYSSAVIRUS in animal

		Sampling		Total units		N of units
Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	unit			Zoonoses	positive
Not Available	Bats - wild - Natural habitat - Not Available - Not Available - Surveillance - Official sampling - Suspect sampling	animal	72	1	European bat lyssavirus 1	1
	Cats - Veterinary clinics - Not Available - Not Available - Surveillance - Official sampling - Suspect sampling	animal	11	0	Lyssavirus	0
	Cattle (bovine animals) - Farm - Not Available - Not Available - Surveillance - Official sampling - Suspect sampling	animal	196	0	Lyssavirus	0
	Dogs - Veterinary clinics - Not Available - Not Available - Surveillance - Official sampling - Suspect sampling	animal	16	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Not Available - Not Available - Surveillance - Official sampling - Suspect sampling	animal	2	0	Lyssavirus	0
	Sheep - Farm - Not Available - Not Available - Surveillance - Official sampling - Suspect sampling	animal	142	0	Lyssavirus	0

Table SALMONELLA in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	N of flocks under contro programme	l Target verification	Total units tested	Total units positive	Zoonoses	N of units positive	
BELGIUM	Gallus gallus (fowl) - broilers - before slaughter - Farm - Not Available - environmental sample - boot swabs -	herd/floc		N	9842	155	Salmonella 4,5,12:i:-	6	
	Control and eradication programmes - Industry sampling - Census	k					Salmonella Agona	2	
							Salmonella Colorado	1	
							Salmonella Derby	4	
							Salmonella Enteritidis	2	
							Salmonella Gaminara	12	
							Salmonella Goldcoast	1	
							Salmonella I 4,12:-:-	1	
							Salmonella Idikan	1	
							Salmonella II, group O:4	2	
							Salmonella Infantis	69	
							Salmonella Java	9	
							Salmonella Kottbus	11	
							Salmonella Livingstone	10	
							Salmonella Llandoff	11	
							Salmonella Mbandaka	3	
							Salmonella Minnesota	1	
							Salmonella Moero	1	
							Salmonella Montevideo	1	
							Salmonella Rissen	3	
								Salmonella Tennessee	1
			hand/flag 00.40				Salmonella Typhimurium	23	
	Gallus gallus (fowl) - broilers - before slaughter - Farm - Not Available - environmental sample - boot swabs -		9846	Υ	9846	162	Salmonella 4,5,12:i:-	6	
	Control and eradication programmes - Official and industry sampling - Census	k					Salmonella Agona	2	
							Salmonella Colorado	1	
							Salmonella Derby	4	
							Salmonella Enteritidis	2	
							Salmonella Gaminara	12	
							Salmonella Goldcoast	1	
							Salmonella I 4,12:-:-	1	
							Salmonella Idikan	1	
							Salmonella II, group O:4	2	
							Salmonella Infantis	72	
							Salmonella Java	10	
							Salmonella Kottbus	1	
							Salmonella Livingstone	11	
							Salmonella Llandoff	1	
							Salmonella Mbandaka	3	
							Salmonella Minnesota	1	
							Salmonella Moero	1	
							Salmonella Montevideo	1	
							Salmonella Rissen	3	

	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	N of flocks under control programme	l Target verification	Total units tested	Total units positive	Zoonoses	N of units positive
	Gallus gallus (fowl) - broilers - before slaughter - Farm - Not Available - environmental sample - boot swabs -	herd/floc	9846	Υ	9846	162	Salmonella Tennessee	1
	Control and eradication programmes - Official and industry sampling - Census	k					Salmonella Typhimurium	25
	Gallus gallus (fowl) - broilers - before slaughter - Farm - Not Available - environmental sample - boot swabs -	herd/floc		N	80	7	Salmonella Infantis	3
	Control and eradication programmes - Official sampling - Objective sampling	k					Salmonella Java	1
							Salmonella Livingstone	1
							Salmonella Typhimurium	2
_	Gallus gallus (fowl) - broilers - day-old chicks - Farm - Not Available - environmental sample - delivery box	herd/floc		N	5837	8	Salmonella Djugu	1
	liner - Control and eradication programmes - Industry sampling - Census	k					Salmonella Infantis	1
							Salmonella Llandoff	2
							Salmonella Mbandaka	2
							Salmonella Typhimurium	2
	Gallus gallus (fowl) - laying hens - adult - Farm - Not Available - environmental sample - boot swabs -	herd/floc		N	637	20	Salmonella Enteritidis	2
	Control and eradication programmes - Industry sampling - Census	k					Salmonella Havana	1
							Salmonella I 6,7:-:-	1
							Salmonella Idikan	2
							Salmonella Infantis	4
							Salmonella Mbandaka	6
							Salmonella Paratyphi B	1
							Salmonella spp., unspecified	1
							Salmonella Tennessee	2
ı	Gallus gallus (fowl) - laying hens - adult - Farm - Not Available - environmental sample - boot swabs -	herd/floc		N	235	4	Salmonella Idikan	3
	Control and eradication programmes - Official sampling - Census	k				•	Salmonella Livingstone	1
-	Gallus gallus (fowl) - laying hens - adult - Farm - Not Available - environmental sample - boot swabs and	herd/floc	655	Υ	655	30	Salmonella Enteritidis	2
	dust - Control and eradication programmes - Official and industry sampling - Census	k		·			Salmonella Havana	1
		K					Salmonella I 6,7:-:-	1
							Salmonella Idikan	5
							Salmonella Infantis	5
							Salmonella Livingstone	4
							Salmonella Mbandaka	7
							Salmonella Paratyphi B	1
							Salmonella spp., unspecified	1
							Salmonella Tennessee	3
-	Gallus gallus (fowl) - laying hens - adult - Farm - Not Available - environmental sample - dust - Control and	herd/floc		N	235	7	Salmonella Idikan	2
	eradication programmes - Official sampling - Census	k		14	200	,	Salmonella Infantis	1
							Salmonella Livingstone	2
							Salmonella Mbandaka	1
							Salmonella Tennessee	1
ī	Gallus gallus (fowl) - laying hens - day-old chicks - Farm - Not Available - environmental sample - delivery	herd/floc		N	253	0	Salmonella	1
	box liner - Control and eradication programmes - Industry sampling - Census	k		IN	200	0	Sairionella	0
	Gallus gallus (fowl) - laying hens - during rearing period - flocks under control programme - Farm - Not Available - environmental sample - boot swabs - Control and eradication programmes - Industry sampling -	herd/floc k		N	286	2	Salmonella Idikan	1
	Census						Salmonella Kedougou	1
-	Gallus gallus (fowl) - parent breeding flocks, unspecified - adult - Farm - Not Available - environmental	herd/floc	565	Υ	565	16	Salmonella 3,19:-:-	2
	sample - boot swabs - Control and eradication programmes - Official and industry sampling - Census	k					Salmonella Enteritidis	2
							Salmonella Idikan	3
							Salmonella Kedougou	1
							Salmonella Llandoff	2

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	N of flocks under control programme	Target verification	Total units tested	Total units positive	Zoonoses	N of units positive
BELGIUM	Gallus gallus (fowl) - parent breeding flocks, unspecified - adult - Farm - Not Available - environmental	herd/floc	565	Υ	565	16	Salmonella Mbandaka	1
	sample - boot swabs - Control and eradication programmes - Official and industry sampling - Census	k					Salmonella Montevideo	1
							Salmonella Senftenberg	4
	Gallus gallus (fowl) - parent breeding flocks, unspecified - day-old chicks - Farm - Not Available - environmental sample - delivery box liner - Control and eradication programmes - Industry sampling - Census	herd/floc k		N	174	0	Salmonella	0
	Gallus gallus (fowl) - parent breeding flocks, unspecified - during rearing period - Farm - Not Available - environmental sample - boot swabs - Control and eradication programmes - Official and industry sampling - Census	herd/floc k		N	331	0	Salmonella	0
	Turkeys - fattening flocks - before slaughter - Farm - Belgium - environmental sample - boot swabs - Control and eradication programmes - Industry sampling - Census	herd/floc k		N	194	0	Salmonella	0
	Turkeys - fattening flocks - before slaughter - Farm - Belgium - environmental sample - boot swabs - Control and eradication programmes - Official and industry sampling - Census	herd/floc k	194	Y	194	0	Salmonella	0
	Turkeys - fattening flocks - before slaughter - Farm - Belgium - environmental sample - boot swabs - Control and eradication programmes - Official sampling - Objective sampling	herd/floc k		N	3	0	Salmonella	0

Table SALMONELLA in food

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Bakery products - desserts - containing raw eggs - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	25	0	Salmonella	0
	Bakery products - pastry - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	30	0	Salmonella	0
	Bakery products - pastry - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	61	0	Salmonella	0
	Cheeses made from cows' milk - fresh - made from pasteurised milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	25	0	Salmonella	0
	Cheeses made from cows' milk - fresh - made from pasteurised milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	35	0	Salmonella	0
	Cheeses made from cows' milk - fresh - made from pasteurised milk - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	31	0	Salmonella	0
	Cheeses made from cows' milk - fresh - made from raw or low heat-treated milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	54	0	Salmonella	0
	Cheeses made from cows' milk - fresh - made from raw or low heat-treated milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	24	0	Salmonella	0
	Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	2	0	Salmonella	0
	Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	77	0	Salmonella	0
	Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	106	0	Salmonella	0
	Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	47	0	Salmonella	0
	Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	24	1	Salmonella spp., unspecified	1
	Cheeses made from cows' milk - soft and semi-soft - made from raw or low heat-treated milk - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	110	0	Salmonella	0
	Cheeses made from goats' milk - unspecified - made from pasteurised milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	1	0	Salmonella	0
	Cheeses made from goats' milk - unspecified - made from pasteurised milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	34	0	Salmonella	0
	Cheeses made from goats' milk - unspecified - made from pasteurised milk - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	56	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Cheeses made from goats' milk - unspecified - made from raw or low heat-treated milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	25	0	Salmonella	0
	Cheeses made from goats' milk - unspecified - made from raw or low heat-treated milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	25	0	Salmonella	0
	Cheeses made from goats' milk - unspecified - made from raw or low heat-treated milk - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	41	0	Salmonella	0
	Cheeses made from goats' milk - Wholesale - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Cheeses made from sheep's milk - fresh - made from pasteurised milk - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	69	0	Salmonella	0
	Cheeses made from sheep's milk - fresh - made from raw or low heat-treated milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	3	0	Salmonella	0
	Cheeses made from sheep's milk - fresh - made from raw or low heat-treated milk - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	85	0	Salmonella	0
	Cheeses made from sheep's milk - unspecified - made from pasteurised milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	5	0	Salmonella	0
	Cheeses made from sheep's milk - unspecified - made from raw or low heat-treated milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	2	0	Salmonella	0
	Chocolate - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	36	0	Salmonella	0
	Chocolate - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	55	0	Salmonella	0
	Crustaceans - Border inspection activities - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	5	0	Salmonella	0
	Crustaceans - prawns - cooked - Wholesale - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	119	0	Salmonella	0
	Crustaceans - unspecified - cooked - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	30	0	Salmonella	0
	Crustaceans - unspecified - cooked - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	41	0	Salmonella	0
	Dairy products (excluding cheeses) - butter - made from pasteurised milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	7	0	Salmonella	0
	Dairy products (excluding cheeses) - butter - made from pasteurised milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	5	0	Salmonella	0
	Dairy products (excluding cheeses) - butter - made from pasteurised milk - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	36	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Dairy products (excluding cheeses) - butter - made from raw or low heat-treated milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	35	0	Salmonella	0
	Dairy products (excluding cheeses) - butter - made from raw or low heat-treated milk - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	5	0	Salmonella	0
	Dairy products (excluding cheeses) - dairy desserts - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	21	0	Salmonella	0
	Dairy products (excluding cheeses) - dairy desserts - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	25	0	Salmonella	0
	Dairy products (excluding cheeses) - dairy desserts - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	41	0	Salmonella	0
	Dairy products (excluding cheeses) - ice-cream - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	20	0	Salmonella	0
	Dairy products (excluding cheeses) - ice-cream - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	20	0	Salmonella	0
	Dairy products (excluding cheeses) - ice-cream - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	46	0	Salmonella	0
	Dairy products (excluding cheeses) - milk powder and whey powder - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	81	0	Salmonella	0
	Egg products - dried - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	9	0	Salmonella	0
	Egg products - dried - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	5	0	Salmonella	0
	Egg products - liquid - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	41	0	Salmonella	0
	Egg products - liquid - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	30	0	Salmonella	0
	Fish - gravad /slightly salted - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	10	0	Salmonella	0
	Fish - gravad /slightly salted - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	10	0	Salmonella	0
	Fish - smoked - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	20	0	Salmonella	0
	Fish - smoked - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	51	0	Salmonella	0
	Fishery products, unspecified - ready-to-eat - chilled - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	66	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Fishery products, unspecified - ready-to-eat - chilled - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	116	0	Salmonella	0
	Fishery products, unspecified - ready-to-eat - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	91	0	Salmonella	0
	Follow-on formulae - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	8	0	Salmonella	0
	Follow-on formulae - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	46	0	Salmonella	0
	Foodstuffs intended for special nutritional uses - dried dietary foods for special medical purposes intended for infants below 6 months - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	59	0	Salmonella	0
	Frogs leg - Border inspection activities - Not Available - Not Available - Surveillance - Official	single	25	Gram	5	5	Salmonella Braenderup	1
	sampling - Objective sampling	(food/fee d)					Salmonella Javiana	1
		u)					Salmonella Panama	1
							Salmonella Saintpaul	1
							Salmonella Wandsworth	1
	Fruits - non-pre-cut - frozen - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	55	0	Salmonella	0
	Fruits - non-pre-cut - frozen - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	36	0	Salmonella	0
	Fruits - non-pre-cut - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	41	0	Salmonella	0
	Fruits - non-pre-cut - Wholesale - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	40	0	Salmonella	0
	Fruits and vegetables - pre-cut - ready-to-eat - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	31	0	Salmonella	0
	Fruits and vegetables - pre-cut - ready-to-eat - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	60	0	Salmonella	0
	Infant formula - dried - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	6	0	Salmonella	0
	Infant formula - dried - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	50	0	Salmonella	0
	Infant formula - ready-to-eat - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Millilitre	72	0	Salmonella	0
	Juice - fruit juice - pasteurised - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	5	0	Salmonella	0
	Juice - fruit juice - pasteurised - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Millilitre	14	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Juice - fruit juice - unpasteurised - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	9	0	Salmonella	0
	Juice - fruit juice - unpasteurised - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Millilitre	143	0	Salmonella	0
	Juice - vegetable juice - pasteurised - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	6	0	Salmonella	0
	Juice - vegetable juice - pasteurised - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Millilitre	6	0	Salmonella	0
	Juice - vegetable juice - unpasteurised - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	1	0	Salmonella	0
	Juice - vegetable juice - unpasteurised - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Millilitre	16	0	Salmonella	0
	Live bivalve molluscs - Retail - Not Available - Not Available - Surveillance - Official sampling -	single	25	Gram	77	3	Salmonella Nottingham	1
	Objective sampling	(food/fee d)					Salmonella Typhimurium	2
	Meat from bovine animals - meat preparation - intended to be eaten cooked - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	10	Gram	8	0	Salmonella	0
	Meat from bovine animals - meat preparation - intended to be eaten cooked - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	10	Gram	16	0	Salmonella	0
	Meat from bovine animals - meat preparation - intended to be eaten raw - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	147	0	Salmonella	0
	Meat from bovine animals - meat preparation - intended to be eaten raw - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	117	0	Salmonella	0
	Meat from bovine animals - minced meat - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	10	Gram	37	0	Salmonella	0
	Meat from bovine animals and pig - meat preparation - intended to be eaten cooked - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	10	Gram	15	0	Salmonella	0
	Meat from bovine animals and pig - meat preparation - intended to be eaten cooked - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	10	Gram	2	0	Salmonella	0
	Meat from bovine animals and pig - meat preparation - intended to be eaten raw - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	2	0	Salmonella	0
	Meat from bovine animals and pig - meat preparation - intended to be eaten raw - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	3	0	Salmonella	0
	Meat from bovine animals and pig - minced meat - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	10	Gram	34	0	Salmonella	0
	Meat from broilers (Gallus gallus) - carcase - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	90	1	Salmonella Infantis	1

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Meat from broilers (Gallus gallus) - carcase - Slaughterhouse - Not Available - Not Available -	single (food/fee	1	Gram	283	6	Salmonella Infantis	4
	Monitoring - Official sampling - Objective sampling	d)					Salmonella Paratyphi B	2
	Meat from broilers (Gallus gallus) - carcase - spent hens - Slaughterhouse - Not Available - Not	single	1	Gram	512	51	Salmonella Agona	1
	Available - Monitoring - Official sampling - Objective sampling	(food/fee d)					Salmonella Enteritidis	38
		u)					Salmonella Give	2
							Salmonella Infantis	6
							Salmonella Mbandaka	3
							Salmonella Typhimurium	1
	Meat from broilers (Gallus gallus) - fresh - skinned - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	39	2	Salmonella Infantis	2
	Meat from broilers (Gallus gallus) - fresh - with skin - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	41	0	Salmonella	0
	Meat from other animal species or not specified - meat products - Border inspection activities - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	7	0	Salmonella	0
	Meat from other animal species or not specified - meat products - cooked, ready-to-eat - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	152	0	Salmonella	0
	Meat from other animal species or not specified - meat products - cooked, ready-to-eat - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	132	0	Salmonella	0
	Meat from other animal species or not specified - meat products - fermented sausages - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	45	0	Salmonella	0
	Meat from other animal species or not specified - meat products - fermented sausages - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	46	0	Salmonella	0
	Meat from other animal species or not specified - meat products - pâté - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	51	0	Salmonella	0
	Meat from other animal species or not specified - meat products - pâté - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	46	0	Salmonella	0
	Meat from other animal species or not specified - mechanically separated meat (MSM) - soft-type - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	10	Gram	20	0	Salmonella	0
	Meat from other animal species or not specified - minced meat - intended to be eaten cooked -	single	10	Gram	47	3	Salmonella 4,5,12:i:-	1
	Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee d)					Salmonella Give	2
	Meat from other animal species or not specified - minced meat - intended to be eaten raw - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	46	0	Salmonella	0
	Meat from other poultry species - meat preparation - intended to be eaten cooked - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	10	Gram	49	2	Salmonella Infantis	2
	Meat from other poultry species - meat products - cooked, ready-to-eat - chilled - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	42	0	Salmonella	0
	Meat from pig - carcase - Slaughterhouse - Not Available - Not Available - Control and eradication	single	600	Square	1066 84	84	Salmonella 4,5,12:i:-	17
	programmes - Official, based on Regulation 854/2004 - Objective sampling	(food/fee d)		centimetre			Salmonella Brandenburg	2
		u)					Salmonella Derby	14

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Meat from pig - carcase - Slaughterhouse - Not Available - Not Available - Control and eradication	single	600	Square	1066	84	Salmonella enterica	2
	programmes - Official, based on Regulation 854/2004 - Objective sampling	(food/fee		centimetre			Salmonella Infantis	7
		d)					Salmonella Livingstone	7
							Salmonella London	2
							Salmonella Rissen	4
							Salmonella Typhimurium	29
	Meat from pig - fresh - Processing plant - Not Available - Not Available - Surveillance - Official	single	25	Gram	229	5	Salmonella 4,5,12:i:-	1
	sampling - Objective sampling	(food/fee					Salmonella Brandenburg	1
		d)					Salmonella Derby	1
							Salmonella Ohio	1
							Salmonella Typhimurium	1
	Meat from pig - meat preparation - intended to be eaten cooked - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	10	Gram	22	0	Salmonella	0
	Meat from pig - meat preparation - intended to be eaten cooked - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	10	Gram	32	1	Salmonella 4,5,12:i:-	1
	Meat from pig - meat preparation - intended to be eaten raw - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	24	0	Salmonella	0
	Meat from pig - meat preparation - intended to be eaten raw - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	9	0	Salmonella	0
	Meat from pig - meat products - cooked ham - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	45	0	Salmonella	0
	Meat from pig - meat products - cooked ham - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	45	0	Salmonella	0
	Meat from pig - meat products - raw ham - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	45	0	Salmonella	0
	Meat from pig - meat products - raw ham - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	46	0	Salmonella	0
	Meat from pig - minced meat - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	10	Gram	34	0	Salmonella	0
	Meat from poultry, unspecified - fresh - skinned - Processing plant - Not Available - Not Available -	single	25	Gram	231	9	Salmonella Bardo	1
	Monitoring - Official sampling - Objective sampling	(food/fee					Salmonella Infantis	4
		d)					Salmonella Kentucky	1
							Salmonella Paratyphi B	1
							Salmonella spp., unspecified	1
							Salmonella Stanley	1
	Meat from poultry, unspecified - fresh - with skin - Processing plant - Not Available - Not Available - Monitoring - Official sampling - Objective sampling	single (food/fee	25	Gram	233	13	Salmonella enterica, subspecies enterica	3
		d)					Salmonella Enteritidis	1
							Salmonella Infantis	4
							Salmonella Paratyphi B	5

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Meat from poultry, unspecified - meat preparation - intended to be eaten cooked - Processing plant	single	25	Gram	61	4	Salmonella Enteritidis	1
	- Not Available - Not Available - Surveillance - Official sampling - Objective sampling	(food/fee					Salmonella Infantis	2
		d)					Salmonella spp., unspecified	1
	Meat from poultry, unspecified - meat products - cooked, ready-to-eat - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	45	0	Salmonella	0
	Meat from poultry, unspecified - meat products - cooked, ready-to-eat - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	92	0	Salmonella	0
	Meat from poultry, unspecified - meat products - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	45	0	Salmonella	0
	Meat from poultry, unspecified - meat products - Retail - Not Available - Not Available - Surveillance	single	25	Gram	48	3	Salmonella Bredeney	1
	- Official sampling - Objective sampling	(food/fee					Salmonella Infantis	1
		d)					Salmonella Paratyphi B	1
	Milk, cows' - raw milk - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Millilitre	35	0	Salmonella	0
	Molluscan shellfish - cooked - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	45	0	Salmonella	0
	Molluscan shellfish - cooked - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	44	0	Salmonella	0
	Other processed food products and prepared dishes - unspecified - ready-to-eat foods - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	31	0	Salmonella	0
	Other processed food products and prepared dishes - unspecified - ready-to-eat foods - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	423	0	Salmonella	0
	Other products of animal origin - Farm - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	7	0	Salmonella	0
	Other products of animal origin - gelatin and collagen - Border inspection activities - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	3	0	Salmonella	0
	Other products of animal origin - gelatin and collagen - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	15	0	Salmonella	0
	Other products of animal origin - gelatin and collagen - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	71	0	Salmonella	0
	Other products of animal origin - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	8	0	Salmonella	0
	Other products of animal origin - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	29	0	Salmonella	0
	Seeds, dried - Border inspection activities - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	36	1	Salmonella Chester	1
	Seeds, sprouted - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	20	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Seeds, sprouted - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	26	0	Salmonella	0
	Seeds, sprouted - Wholesale - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	30	0	Salmonella	0
	Spices and herbs - dried - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	44	1	Salmonella Teddington	1
	Spices and herbs - dried - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	46	0	Salmonella	0
	Spices and herbs - fresh - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	46	0	Salmonella	0
	Spices and herbs - fresh - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	5	0	Salmonella	0
	Spices and herbs - fresh - Wholesale - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	40	0	Salmonella	0
	Surimi - Border inspection activities - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	3	0	Salmonella	0
	Surimi - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	10	0	Salmonella	0
	Surimi - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	31	0	Salmonella	0
	Vegetables - leaves - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	23	0	Salmonella	0
	Vegetables - leaves - Wholesale - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	23	0	Salmonella	0
	Vegetables - non-pre-cut - Retail - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	23	0	Salmonella	0
	Vegetables - non-pre-cut - Wholesale - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee d)	25	Gram	22	0	Salmonella	0

Table SALMONELLA in feed

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	All feedingstuffs - Border inspection activities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	All feedingstuffs - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Compound feedingstuffs for cattle - Catering - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Compound feedingstuffs for cattle - Farm - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	45	0	Salmonella	0
	Compound feedingstuffs for cattle - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	36	0	Salmonella	0
	Compound feedingstuffs for cattle - Unspecified - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	3	0	Salmonella	0
	Compound feedingstuffs for cattle - Wholesale - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	9	0	Salmonella	0
	Compound feedingstuffs for fish - Farm - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	7	0	Salmonella	0
	Compound feedingstuffs for fish - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	7	0	Salmonella	0
	Compound feedingstuffs for fish - Unspecified - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Compound feedingstuffs for fish - Wholesale - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	3	0	Salmonella	0
	Compound feedingstuffs for horses - Farm - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	8	0	Salmonella	0
	Compound feedingstuffs for horses - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	20	0	Salmonella	0
	Compound feedingstuffs for horses - Retail - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Compound feedingstuffs for horses - Unspecified - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Compound feedingstuffs for horses - Wholesale - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	3	0	Salmonella	0
	Compound feedingstuffs for pigs - Farm - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	46	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Compound feedingstuffs for pigs - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	41	0	Salmonella	0
	Compound feedingstuffs for pigs - Retail - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Compound feedingstuffs for pigs - Unspecified - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Compound feedingstuffs for pigs - Wholesale - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	2	0	Salmonella	0
	Compound feedingstuffs for poultry (non specified) - Farm - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	7	0	Salmonella	0
	Compound feedingstuffs for poultry (non specified) - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	3	0	Salmonella	0
	Compound feedingstuffs for poultry (non specified) - Retail - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Compound feedingstuffs for poultry (non specified) - Unspecified - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Compound feedingstuffs for poultry (non specified) - Wholesale - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	2	0	Salmonella	0
	Compound feedingstuffs for poultry, breeders - Farm - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	20	0	Salmonella	0
	Compound feedingstuffs for poultry, breeders - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	17	0	Salmonella	0
	Compound feedingstuffs for poultry, breeders - Hatchery - Not Available - feed sample - Surveillance		25	Gram	1	1	Salmonella	1
	- Official sampling - Objective sampling	(food/fee d)					Salmonella spp., unspecified	1
	Compound feedingstuffs for poultry, breeders - Retail - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Compound feedingstuffs for poultry, breeders - Wholesale - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Compound feedingstuffs for poultry, broilers - Farm - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	19	0	Salmonella	0
	Compound feedingstuffs for poultry, broilers - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	17	0	Salmonella	0
	Compound feedingstuffs for poultry, broilers - Retail - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Compound feedingstuffs for poultry, broilers - Wholesale - Not Available - feed sample - Surveillance	batch	25	Gram	2	1	Salmonella	1
	- Official sampling - Objective sampling	(food/fee d)					Salmonella Typhimurium	1

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Compound feedingstuffs for poultry, laying hens - Farm - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	19	0	Salmonella	0
	Compound feedingstuffs for poultry, laying hens - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	14	0	Salmonella	0
	Compound feedingstuffs for poultry, laying hens - Retail - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Compound feedingstuffs for poultry, laying hens - Unspecified - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Compound feedingstuffs for poultry, laying hens - Wholesale - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	4	0	Salmonella	0
	Compound feedingstuffs for rabbits - Farm - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Compound feedingstuffs for rabbits - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	6	0	Salmonella	0
	Compound feedingstuffs for sheep - final product - Farm - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	9	0	Salmonella	0
	Compound feedingstuffs for sheep - final product - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	9	0	Salmonella	0
	Compound feedingstuffs for sheep - final product - Retail - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Compound feedingstuffs for turkeys - Farm - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	4	0	Salmonella	0
	Compound feedingstuffs for turkeys - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Feed material of cereal grain origin - barley derived - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Feed material of cereal grain origin - Border inspection activities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	3	0	Salmonella	0
	Feed material of cereal grain origin - Conservation facilities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	6	0	Salmonella	0
	Feed material of cereal grain origin - Farm - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	2	0	Salmonella	0
	Feed material of cereal grain origin - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	23	0	Salmonella	0
	Feed material of cereal grain origin - maize derived - Conservation facilities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Feed material of cereal grain origin - maize derived - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	4	0	Salmonella	0
	Feed material of cereal grain origin - maize derived - Wholesale - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Feed material of cereal grain origin - rice derived - Border inspection activities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	3	0	Salmonella	0
	Feed material of cereal grain origin - rice derived - Conservation facilities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	2	0	Salmonella	0
	Feed material of cereal grain origin - rice derived - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Feed material of cereal grain origin - wheat derived - Border inspection activities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	2	0	Salmonella	0
	Feed material of cereal grain origin - wheat derived - Conservation facilities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	4	0	Salmonella	0
	Feed material of cereal grain origin - wheat derived - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	19	0	Salmonella	0
	Feed material of cereal grain origin - wheat derived - Retail - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Feed material of cereal grain origin - wheat derived - Wholesale - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Feed material of cereal grain origin - Wholesale - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	9	0	Salmonella	0
	Feed material of land animal origin - animal fat - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	14	0	Salmonella	0
	Feed material of land animal origin - animal fat - Slaughterhouse - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	3	0	Salmonella	0
	Feed material of land animal origin - animal fat - Wholesale - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Feed material of land animal origin - blood products - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	7	0	Salmonella	0
	Feed material of land animal origin - dairy products - Farm - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	15	0	Salmonella	0
	Feed material of land animal origin - dairy products - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	63	0	Salmonella	0
	Feed material of land animal origin - dairy products - Retail - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	2	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive	
Not Available	Feed material of land animal origin - dairy products - Unspecified - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	5	0	Salmonella	0	
	Feed material of land animal origin - dairy products - Wholesale - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	4	0	Salmonella	0	
	Feed material of land animal origin - egg powder - Border inspection activities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	3	0	Salmonella	0	
	Feed material of land animal origin - egg powder - Conservation facilities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	2	0	Salmonella	0	
	Feed material of land animal origin - egg powder - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	28	0	Salmonella	0	
	Feed material of land animal origin - egg powder - Unspecified - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	6	0	Salmonella	0	
	Feed material of land animal origin - Feed mill - Not Available - feed sample - Surveillance - Official	batch	25	Gram	38	1	Salmonella	1	
	sampling - Objective sampling	(food/fee					Salmonella Infantis	1	
		d)					Salmonella Typhimurium	1	
	Feed material of land animal origin - meat and bone meal - Border inspection activities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	10	0	Salmonella	0	
	Feed material of land animal origin - meat and bone meal - Conservation facilities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	4	0	Salmonella	0	
	Feed material of land animal origin - meat and bone meal - Feed mill - Not Available - feed sample -	batch	25	Gram	129	7	Salmonella	7	
	Surveillance - Official sampling - Objective sampling	(food/fee						Salmonella Brandenburg	7
		u)					Salmonella enterica, subspecies indica	7	
							Salmonella Idikan	7	
							Salmonella Infantis	7	
							Salmonella Isangi	14	
							Salmonella Livingstone	14 7	
							Salmonella Ohio Salmonella Senftenberg	14	
							Salmonella Yoruba	7	
	Feed material of land animal origin - meat and bone meal - Slaughterhouse - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	4	0	Salmonella	0	
	Feed material of land animal origin - meat and bone meal - Unspecified - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0	
	Feed material of land animal origin - meat and bone meal - Wholesale - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	6	0	Salmonella	0	
	Feed material of land animal origin - Slaughterhouse - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	5	0	Salmonella	0	
	Feed material of land animal origin - Unspecified - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	4	0	Salmonella	0	

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Feed material of land animal origin - Wholesale - Not Available - feed sample - Surveillance - Official	batch	25	Gram	7	1	Salmonella	1
	sampling - Objective sampling	(food/fee					Salmonella Infantis	1
		d)					Salmonella spp., unspecified	1
	Feed material of marine animal origin - fish meal - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	8	0	Salmonella	0
	Feed material of marine animal origin - fish meal - Retail - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Feed material of oil seed or fruit origin - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	4	0	Salmonella	0
	Feed material of oil seed or fruit origin - groundnut derived - Conservation facilities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Feed material of oil seed or fruit origin - linseed derived - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	3	0	Salmonella	0
	Feed material of oil seed or fruit origin - linseed derived - Retail - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	3	0	Salmonella	0
	Feed material of oil seed or fruit origin - palm kernel derived - Conservation facilities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Feed material of oil seed or fruit origin - palm kernel derived - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Feed material of oil seed or fruit origin - rape seed derived - Border inspection activities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Feed material of oil seed or fruit origin - rape seed derived - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Feed material of oil seed or fruit origin - soya (bean) derived - Conservation facilities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Feed material of oil seed or fruit origin - soya (bean) derived - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	6	0	Salmonella	0
	Feed material of oil seed or fruit origin - sunflower seed derived - Border inspection activities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	2	0	Salmonella	0
	Feed material of oil seed or fruit origin - sunflower seed derived - Conservation facilities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Feed material of oil seed or fruit origin - sunflower seed derived - Feed mill - Not Available - feed	batch	25	Gram	4	1	Salmonella	1
	sample - Surveillance - Official sampling - Objective sampling	(food/fee d)					Salmonella Cubana	1
	Pet food - Border inspection activities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	7	0	Salmonella	0
	Pet food - Cutting plant - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	2	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Pet food - dog snacks (pig ears, chewing bones) - Border inspection activities - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	13	0	Salmonella	0
	Pet food - dog snacks (pig ears, chewing bones) - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	7	0	Salmonella	0
	Pet food - dog snacks (pig ears, chewing bones) - Retail - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	14	0	Salmonella	0
	Pet food - dog snacks (pig ears, chewing bones) - Unspecified - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	1	0	Salmonella	0
	Pet food - dog snacks (pig ears, chewing bones) - Wholesale - Not Available - feed sample -	batch	25	Gram	3	1	Salmonella	1
	Surveillance - Official sampling - Objective sampling	(food/fee d)					Salmonella spp., unspecified	1
	Pet food - Feed mill - Not Available - feed sample - Surveillance - Official sampling - Objective	batch	25	Gram	59	1	Salmonella	1
	sampling	(food/fee d)					Salmonella Enteritidis	1
		u)					Salmonella Livingstone	1
							Salmonella Paratyphi	1
							Salmonella Typhimurium	1
	Pet food - Retail - Not Available - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/fee d)	25	Gram	2	0	Salmonella	0
	Pet food - Wholesale - Not Available - feed sample - Surveillance - Official sampling - Objective	batch	25	Gram	38	2	Salmonella	2
	sampling	(food/fee					Salmonella Enteritidis	2
		d)					Salmonella Infantis	2
							Salmonella Paratyphi	2

Table STAPHYLOCOCCUS AUREUS METICILLIN RESISTANT (MRSA) in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit		Total units positive	Zoonoses	N of units positive
BELGIUM	Pigs - breeding animals - unspecified - sows and gilts - Farm - Not Available - animal sample - nasal swab - Monitoring - Official sampling - Convenient sampling	herd/floc k	153	74	Methicillin resistant Staphylococcus aureus (MRSA)	74
	Pigs - fattening pigs - Farm - Not Available - animal sample - nasal swab - Monitoring - Official sampling - Convenient sampling	herd/floc k	177	101	Methicillin resistant Staphylococcus aureus (MRSA)	101

Table TRICHINELLA in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit		Total units positive	Zoonoses	N of units positive
Not Available	Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Not Available - Not Available - Surveillance - Official sampling - Census	animal	11212 479	0	Trichinella	0
	Solipeds, domestic - horses - Slaughterhouse - Not Available - Not Available - Surveillance - Official sampling - Census	animal	6086	0	Trichinella	0
	Wild boars - wild - Game handling estabilishment - Not Available - Not Available - Surveillance - Official sampling - Census	animal	11507	2	Trichinella	2

Table YERSINIA in food

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	•	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Not Available	Meat from pig - meat preparation - Processing plant - Not Available - Not Available - Surveillance - Official sampling - Objective sampling	single (food/fee	1	Gram	118	36	Yersinia enterocolitica - biotype 1A (not pathogenic)	34
		d)					Yersinia enterocolitica - biotype 1B	1
							Yersinia enterocolitica unspecified	1
	Meat from pig - meat preparation - Retail - Not Available - Not Available - Surveillance - Official	single	1	Gram	91	17	Biotype 4 - serotype O:3	1
	sampling - Objective sampling	(food/fee d)					Yersinia enterocolitica - biotype 1A (not pathogenic)	16

FOODBORNE OUTBREAKS TABLES

Foodborne Outbreaks: summarized data

	Outbreak strenght		Stro	ng			Wea	ık	
Causative agent	Food vehicle	N outbreaks	N human cases	N hospitalized	N deaths	N outbreaks	N human cases	N hospitalized	N deaths
Campylobacter jejuni	Bovine meat and products thereof					1	2	1	0
Campylobacter, unspecified sp.	Broiler meat (Gallus gallus) and products thereof					1	2	2	0
	Other, mixed or unspecified poultry meat and products thereof					1	2	1	0
Clostridium perfringens	Bovine meat and products thereof	2	246	0	0				
	Broiler meat (Gallus gallus) and products thereof	1	30	0	0				
	Mixed food	1	26	0	0				
Norovirus	Bovine meat and products thereof	1	4	0	0				
	Crustaceans, shellfish, molluscs and products thereof	2	33	1	0				
	Fruit, berries and juices and other products thereof					1	30	0	0
	Tap water, including well water	1	115	5	0				
	Mixed food					2	23	0	0
Salmonella Enteritidis	Eggs and egg products	3	139	41	0				
Staphylococcal enterotoxins	Cheese	1	22	0	0				
	Pig meat and products thereof	1	3	2	0				
Unknown	Milk					2	5	0	0
	Dairy products (other than cheeses)					3	22	3	0
	Cheese					1	5	0	0
	Bovine meat and products thereof					27	86	0	0
	Pig meat and products thereof					3	9	0	0
	Other or mixed red meat and products thereof					15	41	2	0
	Broiler meat (Gallus gallus) and products thereof					11	29	1	0
	Turkey meat and products thereof					1	4	0	0
	Fish and fish products					12	29	0	0
	Crustaceans, shellfish, molluscs and products thereof					20	53	0	0
	Vegetables and juices and other products thereof					8	21	0	0
	Fruit, berries and juices and other products thereof					2	4	0	0
	Drinks, including bottled water					2	5	0	0
	Sweets and chocolate					1	2	0	0
	Bakery products					8	25	1	0
	Mixed food					181	644	8	0
	Buffet meals					16	177	3	0
	Unknown					44	137	0	0
VTEC 0157	Cheese	1	3	1	0				
	Mixed food					1	11	1	0

Strong Foodborne Outbreaks: detailed data

Causative agent	Other Causative Agent	FBO nat. code	Outbreak type	Food vehicle	More food vehicle info	Nature of evidence	Setting	Place of origin of problem	Origin of food vehicle	Contributory factors	Comment	N outbreak	N huma s cases		N N sp. deaths
Clostridiu m perfringen s	unk	1019	General	Mixed food	N_A	Detection of causative agent in food vehicle or its component - Detection of indistinguisha ble causative agent in humans	Camp or picnic	Temporary mass catering (fairs or festivals)	Not Available	NOT AVAILABLE	N_A	1	26	0	0
		1035	General	Broiler meat (Gallus gallus) and products thereof	N_A	Detection of causative agent in food vehicle or its component - Detection of indistinguisha ble causative agent in humans	Resident ial institutio n (nursing home or prison or boarding school)	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Not Available	NOT AVAILABLE	N_A	1	30	0	0
		1062	General	Bovine meat and products thereof	N_A	Descriptive epidemiologic al evidence	Resident ial institutio n (nursing home or prison or boarding school)	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Not Available	NOT AVAILABLE	N_A	1	46	0	0
		917	General	Bovine meat and products thereof	N_A	Detection of causative agent in food vehicle or its component - Detection of indistinguisha ble causative agent in humans	Resident ial institutio n (nursing home or prison or boarding school)	Temporary mass catering (fairs or festivals)	Not Available	NOT AVAILABLE	N_A	1	200	0	0
Norovirus	unk	893	General	Crustaceans, shellfish, molluscs and products thereof	N_A	Detection of causative agent in food vehicle or its component - Detection of indistinguisha ble causative agent in humans	Househ old	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Netherlands	NOT AVAILABLE	N_A	1	30	0	0

Causative agent	Other Causative Agent	FBO nat. code	Outbreak type	Food vehicle	More food vehicle info	Nature of evidence	Setting	Place of origin of problem	Origin of food vehicle	Contributory factors	Comment	N outbreaks	N humar s cases		N p. deaths
Norovirus	unk	897	General	Crustaceans, shellfish, molluscs and products thereof	N_A	Descriptive epidemiologic al evidence	Restaur ant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Netherlands	NOT AVAILABLE	N_A	1	3	1	0
		937	Househol d / domestic kitchen	Bovine meat and products thereof	N_A	Detection of causative agent in food chain or its environment - Symptoms and onset of illness pathognomon ic to causative agent	Restaur ant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Not Available	NOT AVAILABLE	N_A	1	4	0	0
		985 986	General	Tap water, including well water	N_A	Detection of causative agent in food vehicle or its component - Detection of indistinguisha ble causative agent in humans	Camp or picnic	Water source	Not Available	NOT AVAILABLE	N_A	1	115	5	0
Salmonell a Enteritidis	unk	1070 1071	General	Eggs and egg products	N_A	Detection of causative agent in food vehicle or its component - Detection of indistinguisha ble causative agent in humans	Househ old	Retail	Poland	NOT AVAILABLE	the outbreak is part of the multi- country S. Enteritidis outbreak due to Polish eggs	1	24	11	0
		1073	General	Eggs and egg products	N_A	Descriptive epidemiologic al evidence	Househ old	Household	Not Available	NOT AVAILABLE	N_A	1	4	3	0
		1163	General	Eggs and egg products	N_A	Detection of causative agent in food vehicle or its component - Detection of indistinguisha ble causative agent in humans	Unknow n	Unknown	Poland	NOT AVAILABLE	the outbreak is part of the multi- country S. Enteritidis outbreak due to Polish eggs	1	111	27	0
Staphyloc occal enterotoxi ns	unk	1092	General	Cheese	N_A	Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomon ic to causative agent	Resident ial institution (nursing home or prison or boarding school)	Farm (not specified)	Not Available	NOT AVAILABLE	N_A	1	22	0	0

Causative agent	Other Causative Agent	FBO nat. code	Outbreak type	Food vehicle	More food vehicle info	Nature of evidence	Setting	Place of origin of problem	Origin of food vehicle	Contributory factors	Comment	N outbreak	N huma s case		I N sp. deaths
Staphyloc occal enterotoxi ns	unk	987	Househol d / domestic kitchen	Pig meat and products thereof	N_A	Detection of causative agent in food vehicle or its component - Detection of indistinguisha ble causative agent in humans	Househ old	Retail	Not Available	NOT AVAILABLE	N_A	1	3	2	0
VTEC O157	unk	1026	Househol d / domestic kitchen	Cheese	N_A	Detection of causative agent in food vehicle or its component - Detection of indistinguisha ble causative agent in humans	Househ old	Mobile retailer or market/str eet vendor	Not Available	NOT AVAILABLE	N_A	1	3	1	0

Weak Foodborne Outbreaks: detailed data

Causative agent	Other Causative Agent	FBO nat. code	Outbreak type	Food vehicle	More food vehicle info	Nature of evidence	Setting	Place of origin of problem	Origin of foo	d Contributory factors	Comment	N outbreaks	N humai cases		
Campylob acter jejuni	unk	964	Not Available	Bovine meat and products thereof	N_A	Unknown	Househol d	Retail	Not Available	NOT AVAILABLE	N_A	1	2	1	0
Campylob acter, unspecifie d sp.	unk	1030	Not Available	Broiler meat (Gallus gallus) and products thereof	N_A	Unknown	Take- away or fast-food outlet	Take- away or fast-food outlet	Not Available	NOT AVAILABLE	N_A	1	2	2	0
		983	Not Available	Other, mixed or unspecified poultry meat and products thereof	N_A	Unknown	Restauran t or Cafe or Pub or Bar or Hotel or Catering service	Restauran t or Cafe or Pub or Bar or Hotel or Catering service	Not Available	NOT AVAILABLE	N_A	1	2	1	0
Norovirus	unk	1069	Not Available	Fruit, berries and juices and other products thereof	N_A	Unknown	Canteen or workplace catering	Canteen or workplace catering	Not Available	NOT AVAILABLE	N_A	1	30	0	0
		1117	Not Available	Mixed food	N_A	Unknown	Restauran t or Cafe or Pub or Bar or Hotel or Catering service	Restauran t or Cafe or Pub or Bar or Hotel or Catering service	Not Available	NOT AVAILABLE	N_A	1	3	0	0
		899	Not Available	Mixed food	N_A	Unknown	Canteen or workplace catering	Restauran t or Cafe or Pub or Bar or Hotel or Catering service	Not Available	NOT AVAILABLE	N_A	1	20	0	0
Unknown	unk	2016/ 0291	Not Available	Fruit, berries and juices and other products thereof	N_A	Unknown	Unknown	unk	Not Available	NOT AVAILABLE	N_A	1	2	0	0
		2016/ 0746	Not Available	Sweets and chocolate	N_A	Unknown	Househol d	unk	Not Available	NOT AVAILABLE	N_A	1	2	0	0
		936	Not Available	Buffet meals	N_A	Descriptive epidemiological evidence	Unknown	unk	Not Available	NOT AVAILABLE	N_A	10	89	0	0
		N_A	Not Available	Milk	N_A	Unknown	Unknown	unk	Not Available	NOT AVAILABLE	N_A	2	5	0	0

Causative agent	Other Causative Agent	FBO nat. code	Outbreak type	Food vehicle	More food vehicle info	Nature of evidence	Setting	Place of origin of problem	Origin of foo	od Contributory factors	Comment	N outbreaks	N huma cases		N p. deaths
Unknown	unk	N_A	Not Available	Dairy products (other than cheeses)	N_A	Unknown	Unknown	unk	Not Available	NOT AVAILABLE	N_A	3	22	3	0
				Cheese	N_A	Unknown	Unknown	unk	Not Available	NOT AVAILABLE	N_A	1	5	0	0
				Bovine meat and products thereof	N_A	Unknown	Unknown	unk	Not Available	NOT AVAILABLE	N_A	27	86	0	0
				Pig meat and products thereof	N_A	Unknown	Unknown	unk	Not Available	NOT AVAILABLE	N_A	3	9	0	0
				Other or mixed red meat and products thereof	N_A	Unknown	Unknown	unk	Not Available	NOT AVAILABLE	N_A	15	41	2	0
				Broiler meat (Gallus gallus) and products thereof	N_A	Unknown	Unknown	unk	Not Available	NOT AVAILABLE	N_A	11	29	1	0
				Turkey meat and products thereof	N_A	Unknown	Unknown	unk	Not Available	NOT AVAILABLE	N_A	1	4	0	0
				Fish and fish products	N_A	Unknown	Unknown	unk	Not Available	NOT AVAILABLE	N_A	12	29	0	0
				Crustaceans, shellfish, molluscs and products thereof	N_A	Unknown	Unknown	unk	Not Available	NOT AVAILABLE	N_A	20	53	0	0
				Vegetables and juices and other products thereof	N_A	Unknown	Unknown	unk	Not Available	NOT AVAILABLE	N_A	8	21	0	0
				Fruit, berries and juices and other products thereof	N_A	Unknown	Unknown	unk	Not Available	NOT AVAILABLE	N_A	1	2	0	0
				Drinks, including bottled water	N_A	Unknown	Unknown	unk	Not Available	NOT AVAILABLE	N_A	2	5	0	0
_				Bakery products	N_A	Unknown	Unknown	unk	Not Available	NOT AVAILABLE	N_A	8	25	1	0

Causative agent	Other Causative Agent	FBO nat. code	Outbreak type	Food vehicle	More food vehicle info	Nature of evidence	Setting	Place of origin of problem	Origin of foo	d Contributory factors	Comment	N outbreaks	N humar cases		N p. deaths
Unknown	unk	N_A	Not Available	Mixed food	N_A	Unknown	Unknown	unk	Not Available	NOT AVAILABLE	N_A	181	644	8	0
				Buffet meals	N_A	Unknown	Unknown	unk	Not Available	NOT AVAILABLE	N_A	6	88	3	0
				Unknown	N_A	Unknown	Unknown	unk	Not Available	NOT AVAILABLE	N_A	44	137	0	0
VTEC O157	unk	997	Not Available	Mixed food	N_A	Unknown	Residentia I institution (nursing home or prison or boarding school)	Canteen or workplace catering	Not Available	NOT AVAILABLE	N_A	1	11	1	0

ANTIMICROBIAL RESISTANCE TABLES FOR CAMPYLOBACTER

Table Antimicrobial susceptibility testing of Campylobacter jejuni in Meat from broilers (Gallus gallus) - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Netherlands

Sampling details: N_A

	AM substance	Ciprofloxacin	Erythromycin (Erythromycin A)	Gentamicin	Nalidixic acid	Streptomycin	Tetracycline
	ECOFF	0.5	4	2	16	4	1
	Lowest limit	0.12	1	0.12	1	0.25	0.5
	Highest limit	16	128	16	64	16	64
	N of tested isolates	7	7	7	7	7	7
MIC	N of resistant isolates	3	0	0	3	0	2
<=0.12		2		4			
<=0.25						3	
0.25		2		3			
<=0.5							5
0.5						1	
<=1			4				
_1						3	
2			3		1		
4					2		
8					1		
16		3			-		-
64					3		2

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling details: N_A

	AM substance	Ciprofloxacin	Erythromycin (Erythromycin A)	Gentamicin	Nalidixic acid	Streptomycin	Tetracycline
	ECOFF	0.5	4	2	16	4	1
	Lowest limit	0.12	1	0.12	1	0.25	0.5
	Highest limit	16	128	16	64	16	64
	N of tested isolates	173	173	173	173	173	173
MIC	N of resistant isolates	99	2	1	98	3	73
<=0.12		58		40			
<=0.25						5	
0.25		13		102			
<=0.5							81
0.5		3		29		31	
<=1			83				
		1		1		112	19
2			74		10	18	
4			14	1	43	4	1
8		15	1		20		3
16		83			2	3	6
32 64							4
64					98		59
128			1				

Sampling Stage: Slaughterhouse Sampling Type: food sample - neck skin Sampling Context: Monitoring

Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling details: N_A

Sampling details: N_A							
	AM substance	Ciprofloxacin	Erythromycin (Erythromycin A)	Gentamicin	Nalidixic acid	Streptomycin	Tetracycline
	ECOFF	0.5	4	2	16	4	1
	Lowest limit	0.12	1	0.12	1	0.25	0.5
	Highest limit	16	128	16	64	16	64
	N of tested isolates	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0
<=0.12		1					
0.25				1			
<=0.5							1
<=1			1				
1						1	
4					1		

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: France

Sampling details: N_A

	AM substance	Ciprofloxacin	Erythromycin (Erythromycin A)	Gentamicin	Nalidixic acid	Streptomycin	Tetracycline
	ECOFF	0.5	4	2	16	4	1
	Lowest limit	0.12	1	0.12	1	0.25	0.5
	Highest limit	16	128	16	64	16	64
	N of tested isolates	18	18	18	18	18	18
MIC	N of resistant isolates	10	0	0	10	0	9
<=0.12		4		9			
0.25 <=0.5		3		8			
<=0.5							9
0.5		1		1		8	
<=1			10				
1						9	
2			7		2	1	
4			1		4		
8					2		
16		10					1
64					10		8

Sampling Stage: Processing plant

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Netherlands

Sampling details: N_A

	AM substance	Ciprofloxacin	Erythromycin (Erythromycin A)	Gentamicin	Nalidixic acid	Streptomycin	Tetracycline
	ECOFF	0.5	4	2	16	4	1
	Lowest limit	0.12	1	0.12	1	0.25	0.5
	Highest limit	16	128	16	64	16	64
	N of tested isolates	11	11	11	11	11	11
MIC	N of resistant isolates	9	1	0	9	0	7
<=0.12		2		2			
0.25				8			
<=0.5	_						3
0.5				1		3	
<=1			3				
1						7	1
2			6		1	1	2
4			1		1		
8		1	11				
16		8					
64					9		5

Sampling Stage: Processing plant

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling details: N_A

	AM substance	Ciprofloxacin	Erythromycin (Erythromycin A)	Gentamicin	Nalidixic acid	Streptomycin	Tetracycline
	ECOFF	0.5	4	2	16	4	1
	Lowest limit	0.12	1	0.12	1	0.25	0.5
	Highest limit	16	128	16	64	16	64
	N of tested isolates	59	59	59	59	59	59
MIC	N of resistant isolates	40	1	1	39	1	31
<=0.12		11		18			
0.25		6		30			
<=0.5							25
0.5		2		9		13	
<=1			30				
1		1		1		38	3
2			26		5	7	
4			2		7		
8		3	1	1	7		1
16		36			1	1	1
64					39		29

Sampling Stage: Processing plant

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling details: N_A

	AM substance	Ciprofloxacin	Erythromycin (Erythromycin A)	Gentamicin	Nalidixic acid	Streptomycin	Tetracycline
	ECOFF	0.5	4	2	16	4	1
	Lowest limit	0.12	1	0.12	1	0.25	0.5
	Highest limit	16	128	16	64	16	64
	N of tested isolates	12	12	12	12	12	12
MIC	N of resistant isolates	9	1	0	9	0	8
<=0.12		2		2			
0.25		1		7			
<=0.5							4
0.5				3		1	
<=1			7				
1						10	
2			4			1	
4					2		
8		1	1		1		
16		8					
32							1
64					9		7

Sampling Stage: Processing plant

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling details: N_A

AM substance Comparison Co	_							
ECOFF 0.5 4 2 16 4 1 Lowest limit 0.12 1 0.12 1 0.25 0.5 Highest limit 16 128 16 64 16 64 N of tested isolates 4 1 2 2 2 2 2 3 1 2 2 3 1 2 2 2 3 1 2 2 2 3 3 1 3 1 <th< th=""><th></th><th>AM substance</th><th>Ciprofloxacin</th><th>Erythromycin (Erythromycin A)</th><th>Gentamicin</th><th>Nalidixic acid</th><th>Streptomycin</th><th>Fetracycline</th></th<>		AM substance	Ciprofloxacin	Erythromycin (Erythromycin A)	Gentamicin	Nalidixic acid	Streptomycin	Fetracycline
Lowest limit 0.12 1 0.12 1 0.25 0.5 Highest limit 16 128 16 64 16 64 No f tested isolates 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 2 2 2 2 2 5 5 2 2 3 1 2 1 2 3 1 2 1 2 1 2 1 2 2 3 1 2 2 3 1 2 2 3 1 2 2 2 3 1 2 2 2 3 1 2 2 3 1 3 1 3 1 3 1 3 1 3 1 3 1 3 1 3 1 3 1 3 1 3		ECOFF		4			4	1
Highest limit 16 128 16 64 16 64 No f tested isolates 4				1		1	0.25	0.5
MIC No festant isolates 4				128		64		
MIC N of resistant isolates 3 0 0 3 0 2 <=0.12			4	4		4	4	
<=0.12	MIC		3	0		3	0	2
0.25 <=0.5	<=0.12		1		1		-	
<=0.5 1 <=1 2 1 3 1 2 1 1 4 1 1 8 1	0.25				3			
c=1 2 1 3 1 2 2 1 4 1 1 8 1	<=0.5							1
2 1 4 1 1 1 8 1				2				
4 1 1 1 8 1	1						3	1
	2			2			1	
	4		1			1		
16			1					
		<u> </u>	1	<u> </u>	<u> </u>	·	<u> </u>	·
64 3	64					3		2

Sampling Stage: Retail Sampling Type: food sample Sampling Context: Monitoring

Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling details: N_A							
	AM substance	Ciprofloxacin	Erythromycin (Erythromycin A)	Gentamicin	Nalidixic acid	Streptomycin	Tetracycline
	ECOFF	0.5	4	2	16	4	1
	Lowest limit	0.12	1	0.12	1	0.25	0.5
	Highest limit	16	128	16	64	16	64
	N of tested isolates	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0
<=0.12		1					
0.25				1			
<=0.5							1
1						1	
2			1				
8					1		

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling details: N_A

Ciproflox Gentamic Streptom Streptom	Tetra
ECOFF 0.5 4 2 16 4	1
Lowest limit 0.12 1 0.12 1 0.25	0.5
Highest limit 16 128 16 64 16	64
N of tested isolates 9 9 9 9 9	9
MIC N of resistant isolates 4 0 0 0 4 0	4
<=0.12	
0.25 6	
<=0.5	4
0.5	
<=1 5	
1	1
2 4 2	
4	
8 1 1	1
16 3	1
64 4	2

Table Antimicrobial susceptibility testing of Campylobacter jejuni in Gallus gallus (fowl) - broilers

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - active

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling details: N_A

	AM substance	Ciprofloxacin	Erythromycin (Erythromycin A)	Gentamicin	Nalidixic acid	Streptomycin	Tetracycline
	ECOFF	0.5	4	2	16	4	1
	Lowest limit	0.12	1	0.12	1	0.25	0.5
	Highest limit	16	128	16	64	16	64
	N of tested isolates	176	176	176	176	176	176
MIC	N of resistant isolates	94	0	0	88	1	72
<=0.12		68		92			
<=0.25						32	
0.25		10		71			
<=0.5							86
0.5		4		13		108	
<=1			169		11		
1						32	18
2			7		61	3	1
4		32			7		2
8		29			2		11
16		30			7	1	5
>16		3					
32 64					27		5
					49		26
>64					12		22

Sampling Stage: Retail Sampling Type: food sample Sampling Context: Monitoring

Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling details: N_A

Sumpling details: 14_	21						
	AM substance	Ciprofloxacin	Erythromycin (Erythromycin A)	Gentamicin	Validixic acid	Streptomycin	letracycline
	ECOFF	0.5	4	2	16	4	1
	Lowest limit	0.12	1	0.12	1	0.25	0.5
	Highest limit	16	128	16	64	16	64
	N of tested isolates	3	3	3	3	3	3
MIC	N of resistant isolates	1	0	0	1	0	1
<=0.12		2		1			
0.25				1			
<=0.5							2
0.5				1			
<=1			2				
1						2	
2			1		1	1	
4					1		
_16		11					
64					1		1

Sampling Stage: Slaughterhouse Sampling Type: food sample - neck skin Sampling Context: Monitoring Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Luxembourg

Sampling details: N. A.

Sampling details: N_A							
	AM substance	Ciprofloxacin	Erythromycin (Erythromycin A)	Gentamicin	Nalidíxic acid	Streptomycin	Tetracycline
	ECOFF	0.5	4	2	16	4	1
	Lowest limit	0.12	1	0.12	1	0.25	0.5
	Highest limit	16	128	16	64	16	64
	N of tested isolates	2	2	2	2	2	2
MIC	N of resistant isolates	2	0	0	2	0	1
<=0.12				1			
<=0.5							1
0.5				1			
<=1			2				
1						2	
8		1					
16		1					
32							1
64		·	·	·	2	·	·
<u> </u>	·	·	·	·	·	·	·

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Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Netherlands

Sampling details: N_A

	AM substance	Ciprofloxacin	Erythromycin (Erythromycin A)	Gentamicin	Nalidixic acid	Streptomycin	Tetracycline
	ECOFF	0.5	4	2	16	4	1
	Lowest limit	0.12	1	0.12	1	0.25	0.5
	Highest limit	16	128	16	64	16	64
	N of tested isolates	26	26	26	26	26	26
MIC	N of resistant isolates	7	2	0	6	0	8
<=0.12		14		13			
<=0.25						2	
0.25		4		11			
<=0.5							16
0.5		1		2		9	
<=1			17		2		
1						12	2
2			6		2	3	
4			1		10		
8		1	1		5		
16		6			1		1
64					6		7
128			1				

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling details: N_A

	AM substance	Ciprofloxacin	Erythromycin (Erythromycin A)	Gentamicin	Nalidixic acid	Streptomycin	Tetracycline
	ECOFF	0.5	4	2	16	4	1
	Lowest limit	0.12	1	0.12	1	0.25	0.5
	Highest limit	16	128	16	64	16	64
	N of tested isolates	10	10	10	10	10	10
MIC	N of resistant isolates	5	0	1	5	2	4
<=0.12		4		2			
<=0.25						1	
0.25		1		2			
<=0.5							6
0.5				4		1	
<=1			6				
1		1				4	
2			2	1	1	2	
4			2	1	3		
8					1	1	1
16		4				1	
32							1
64					5		2

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Germany

Sampling details: N_A

	AM substance	Ciprofloxacin	Erythromycin (Erythromycin A)	Gentamicin	Nalidixic acid	Streptomycin	Tetracycline
	ECOFF	0.5	4	2	16	4	1
	Lowest limit	0.12	1	0.12	1	0.25	0.5
	Highest limit	16	128	16	64	16	64
	N of tested isolates	11	11	11	11	11	11
MIC	N of resistant isolates	7	1	1	7	1	6
<=0.12		4		1			
0.25				8			
<=0.5							5
0.5				1		1	
<=1			7				
1						8	
2		<u> </u>	3		<u> </u>	1	
4				1	4		
8		1					
16		6				1	1
64					7		5
128			1				
	-	-	-	-	-	-	-

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: France

Sampling details: N_A

	AM substance	Ciprofloxacin	Erythromycin (Erythromycin A)	Gentamicin	Nalidixic acid	Streptomycin	Tetracycline
	ECOFF	0.5	4	2	16	4	1
	Lowest limit	0.12	1	0.12	1	0.25	0.5
	Highest limit	16	128	16	64	16	64
	N of tested isolates	23	23	23	23	23	23
MIC	N of resistant isolates	10	1	0	10	1	16
<=0.12		8		8			
<=0.25						3	
0.25		3		11			
<=0.5							7
0.5		2		3		6	
<=1			17				
1				1		10	
2			4		2	2	1
4			1		8	1	
8		3			2		
16		7			11	1	1
32			1		1		
64					9		14

ANTIMICROBIAL RESISTANCE TABLES FOR SALMONELLA

Table Antimicrobial susceptibility testing of Salmonella 1,4,[5],12:i:- in Meat from pig - carcase

Sampling Stage: Slaughterhouse Sampling Type: food sample - carcase swabs Sampling Context: Monitoring

Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceffazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	18	18	18	18	18	18	18	18	18	18	18	18	18	18
MIC	N of resistant isolates	16	1	0	0	1	2	0	0	0	1	17	12	0	5
<=0.03										16					
0.03							13								
0.064							3			2					
<=0.25				18										4	10
0.25							1								
<=0.5					17				15						
0.5							1							9	3
<=1		2			<u> </u>			17							
1					1				3					5	
<=2								1					6		
<=4								<u> </u>			13				
4			3								13				
<=8			<u> </u>			16									
8			11			10					3				
16			3			1					1	1			
32			1												5
64		16											12		
128						1					1				

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	18	18	18	18	18	18	18	18	18	18	18	18	18	18
MIC	N of resistant isolates	16	1	0	0	1	2	0	0	0	1	17	12	0	5
512												1			
1024												16			

Table Antimicrobial susceptibility testing of Salmonella 1,4,[5],12:i:- in Meat from pig - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1											
<=0.5					1				1						
0.5														1	1
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									
8			1												
64												1			

Table Antimicrobial susceptibility testing of Salmonella 1,4,[5],12:i:- in Meat from other animal species or not specified

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	0	0	0	0	0	0	0	0	1	1	0	0
<=0.03										1					
0.03							1								
<=0.25				1										1	1
<=0.5					1				1						
<=1								1							
<=4											1				
<=8						1									
8			1												
64		1											1		
1024												1			

Table Antimicrobial susceptibility testing of Salmonella 1,4,[5],12:i:- in Meat, mixed meat - meat preparation

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	0	0	0	0	0	0	0	0	1	1	0	0
<=0.015							1								
0.064										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1								1							
<=4											1				
4			1												
<=8						1							<u> </u>		
64		1										4	1		
1024												1			

Table Antimicrobial susceptibility testing of Salmonella 1,4,[5],12:i:- in Meat from goat - fresh - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	1	0	0
<=0.03										1					
0.03							1								
<=0.25				1											1
<=0.5					1				1						
<=1								1							
1														1	
2		1													
<=4			1								1				
4			1			4									
<=8 16						1						1			
64												1	1		
04													1		

Table Antimicrobial susceptibility testing of Salmonella 1,4,[5],12:i:- in Other food

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	0	0	0	0	0	0	0	0	1	1	0	1
<=0.03										1					
0.064							1								
<=0.25				1										1	
<=0.5					1				1						
2								1							
<=8						1									
8			1								1				
32															1
64		1											1		
1024												1			

Table Antimicrobial susceptibility testing of Salmonella 1,4,[5],12:i:- in Meat from pig

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	0	0	0	0	0	0	0	0	1	1	0	0
<=0.03										1					
0.03							1								
<=0.25				1										1	1
<=0.5					1				1						
<=1								1							
<=4											1				
4			1												
<=8						1									
64		1											1		
1024												1			

Table Antimicrobial susceptibility testing of Salmonella 1,4,[5],12:i:- in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
МІС	N of resistant isolates	3	0	0	0	0	2	0	0	0	2	3	2	0	0
<=0.03										3					
0.03							1								
<=0.25				3										2	2
<=0.5					3				2						
0.5							2							1	1
<=1								3	<u> </u>						
1									1						
<= <u>2</u>													1		
4			2								<u>'</u>				
<=8			2			3									
16			1			<u> </u>									
32			•								2				
64		3											2		
1024												3			

Table Antimicrobial susceptibility testing of Salmonella 4,5,12:i:- in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	6	6	6	6	6	6	6	6	6	6	6	6	6	6
MIC	N of resistant isolates	6	0	0	0	0	0	0	0	0	0	6	6	0	1
<=0.015							2								
<=0.03										5					
0.03							4								
0.064										1					
<=0.25				6										6	5
<=0.5					5				6						
<=1								6							
<u>1</u> <=4					1										
4			3								6				
<=8			ა			6									
8			3			0									
>32			3												1
>64		6											6		'
>1024		- 0										6			

Table Antimicrobial susceptibility testing of Salmonella 6,7:z29 in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
						1								
									1					
			1										1	
				1				1						
														11
	1						1							
												1		
										1				
					1									
		1												
											1			
	ECOFF Lowest limit Highest limit N of tested isolates N of resistant	ECOFF 8 Lowest limit 1 Highest limit 64 N of tested isolates 1 N of resistant	ECOFF 8 16 Lowest limit 1 2 Highest limit 64 64 N of tested isolates 1 1 N of resistant	ECOFF 8 16 0.5	ECOFF 8 16 0.5 2	ECOFF 8 16 0.5 2 16	ECOFF 8 16 0.5 2 16 0.064	ECOFF 8 16 0.5 2 16 0.064 2	ECOFF 8 16 0.5 2 16 0.064 2 2	ECOFF 8 16 0.5 2 16 0.064 2 2 0.125	Fig. Fig.	Fig. Fig.	Fig. Fig.	Fig. Fig.

Table Antimicrobial susceptibility testing of Salmonella Agona in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	1	1	0	0
<=0.015							1								
<=0.03										1					
0.03							1								
0.064										1					
<=0.25				2											
<=0.5					2				2						
0.5														2	2
<=1								2					1		
<=2		2											1		
<=4											2				
<=8						2									
8			2												
64												1			
>64													1		
>1024												1			

Table Antimicrobial susceptibility testing of Salmonella Agona in Compound feedingstuffs, not specified - final product - non-pelleted/meal

Sampling Stage: Farm

Sampling Type: feed sample

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	
<=0.5									1						
0.5															1
<=1		1						1							
					1										
<=2											<u> </u>		1		
<=4											1				
4			1			4									
<=8						1						4			
64												1			

Table Antimicrobial susceptibility testing of Salmonella Agona in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	5	5	5	5	5	5	5	5	5	5	5	5	5	5
МІС	N of resistant isolates	1	0	0	0	0	1	0	0	0	0	1	1	0	1
<=0.015							1								
<=0.03										5					
0.03							3								
<=0.25				5										2	3
0.25							1								
<=0.5					5				4						
0.5														2	1
<=1		4						5	<u> </u>					<u> </u>	
1									1					1	
<=2													4		
<=4											3				
<=8			4			4									
8 16			4			1					2		1		
32						-									1
64		1										2			'
256												2			
1024												1			
1021															

Table Antimicrobial susceptibility testing of Salmonella Agona in Meat from broilers (Gallus gallus) - carcase - spent hens

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Netherlands

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.03										1					
0.03							1								
<=0.25				1											
<=0.5					1				1						
0.5															1
<=1		1						1							
1														11	
<=2													1		
<=4											1				
<=8						1									
8			1												
32												1			

Table Antimicrobial susceptibility testing of Salmonella Bardo in Meat from broilers (Gallus gallus) - meat preparation

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Poland

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	1	0	0	0	1	1	1	0	0
<=0.03										1					
<=0.25				1											
<=0.5					1				1						
0.5							1								
<=1								1							
1														1	1
2		1													
<=8						1									
16			1												
64													1		
128											1				
1024												1			

Table Antimicrobial susceptibility testing of Salmonella Brandenburg in Meat from pig - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										2					
0.03							1								
<=0.25				2										1	2
<=0.5					2				2						
0.5								<u> </u>						1	
<=1		1						2							
<=2		1											2		
2 <=4		<u> </u>									2				
<=8						2					2				
8			1												
16			1												
32			'									1			
64												1			

Table Antimicrobial susceptibility testing of Salmonella Brandenburg in Meat from pig

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	0	0	1	0	0	0	0	0	1	1	1	1
<=0.03										1					
0.064							1								
<=0.25				1											
<=0.5					1										
<=1								1							
1									1						
2											4			1	
<=4 16			1								1				
32			ı												1
64		1											1		,
128		·				1									
1024												1			

Table Antimicrobial susceptibility testing of Salmonella Brandenburg in Frogs leg

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Vietnam

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	1	0	0	0	0	0	0	0	0
<=0.03										1					
<=0.25				1											
<=0.5					1										
0.5							1							1	1
<=1		1						1							
1									1						
<=2													1		
<=8						1									
8			1												
16											1				
64												1			

Table Antimicrobial susceptibility testing of Salmonella Bredeney in Meat, mixed meat - meat preparation

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	1	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1											1
<=0.5					1				1						
0.5														1	
<=1								11							
2		1													
<=4						<u> </u>					1				
<=8						1									
8			1									1			
16												11	1		
32													1		

Table Antimicrobial susceptibility testing of Salmonella Bredeney in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.03										2					
0.03							2								
<=0.25				2											11
<=0.5					2				2						
0.5														1	
<=1								2							
1														1	1
<=2													2		
2		2													
<=8			<u> </u>			2					-				
8			2								2				
32												2			

Table Antimicrobial susceptibility testing of Salmonella Chester in Nuts and nut products

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: India

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	1	0	0	0	0	0	0
<=0.03										1					
0.03							1								
<=0.25				1										1	1
<=0.5					1										
<=1		1						1							
<=2													1		
<=4											1				
4									1						
<=8						1									
16			1												
128												1			

Table Antimicrobial susceptibility testing of Salmonella Colorado in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.03										1					
0.03							1								
<=0.25				1										1	
<=0.5					1				1						
0.5															11
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									
8			1												
64												1			

Table Antimicrobial susceptibility testing of Salmonella Derby in Meat from pig - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	14	14	14	14	14	14	14	14	14	14	14	14	14	14
MIC	N of resistant isolates	2	0	0	0	0	1	0	0	0	1	4	4	1	3
<=0.015							4								
<=0.03										14					
0.03							9								
<=0.25				14										5	4
<=0.5					13				13						
0.5							1							6	7
<=1		10						14							
1					1				1					2	
<=2		-											9		
2		2													
<=4			•								13			4	
4			3										1	1	
<=8			0			14									
16			3									2			
32			3									1			3
64		2										5	4		3
128											1	2			
1024												4			
1024															

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Table Antimicrobial susceptibility testing of Salmonella Derby in Meat from pig - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	9	9	9	9	9	9	9	9	9	9	9	9	9	9
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							4								
<=0.03										9					
0.03							5								
<=0.25				9										4	3
<=0.5					9				9						
0.5														4	6
<=1		9						9							
1														1	
<= <u>2</u>											9		8		
4			6								9				
<=8			0			9									
8			3			9							1		
32			<u> </u>									5	<u>'</u>		
64												4			
												7			

Table Antimicrobial susceptibility testing of Salmonella Derby in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
MIC	N of resistant isolates	3	0	0	0	0	0	0	0	0	0	3	0	0	3
<=0.015							3								
<=0.03										3					
<=0.25				3										3	
<=0.5					3				3						
<=1								3							
<=2													3		
<=4											3				
4			2												
<=8			4			3									
8			1												
>32		2													3
>64		3										3			
>1024												ა			

Table Antimicrobial susceptibility testing of Salmonella Derby in Meat from pig

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	0	0	0	0	0	0	0	0	1	1	0	1
<=0.015							1								
<=0.03										1					
<=0.25				1											
<=0.5					1				1						
<=1								1							
1														1	
<=4											1				
<=8						11									
8			1												
32															1
64		1											1		
1024												1			

Table Antimicrobial susceptibility testing of Salmonella enterica, subspecies enterica in Meat from pig - carcase

Sampling Stage: Slaughterhouse Sampling Type: food sample - carcase swabs Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance Cefotaxime synergy test	e E e E e E e D O C e E e D O O O O O O O O O O O O O O O O O O	Cefotax Cefotax Not Available	Cefotaxime + Clavulanic acid	Cefoxitin Cefoxitin Not Available	Ceftazidia Ovailable	Ceftazidime + Clavulanic acid	E L L L L L L L L L L L L L L L L L L L	Eə eə d <u>ir.</u> Not Available	Merope Merope Merope Mot Available	LE L
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.5	0.5	8	2	2	0.06	1	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	1	1	0	1	1	0	0	0	0
<=0.015								1			
0.064										1	
0.5									1		
1		1									
4			1		1						
8				1							
16						1	1				
32											

Table Antimicrobial susceptibility testing of Salmonella enterica, subspecies enterica in Meat from pig - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	1	1	1	1	1	1	0	1	0	1	1	0	0	0
<=0.03										2					
0.03							1								
<=0.25				1										2	2
<=0.5					1										
0.5							1								
<=1		1						2							
1									1						
<=2													2		
<=4											1				
4				1											
<=8						1									
8 16			1		1										
									1						
32			11									1			
64		1													
128						1					1				
1024												1			

Table Antimicrobial susceptibility testing of Salmonella enterica, subspecies enterica in Complementary feedingstuffs

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.03										1					
0.03							1								
<=0.25				1										1	
<=0.5					1										
0.5															1
<=1		1						1							
<=2													1		
2									1						
<=4											1				
<=8						1									
8			1												
128												1			

Table Antimicrobial susceptibility testing of Salmonella enterica, subspecies enterica in Complementary feedingstuffs

Sampling Stage: Retail

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: France

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<=0.03										1					
0.03							1								
<=0.25				1										1	
<=0.5					1				1						
0.5															1
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									
8			1												
1024												1			

Table Antimicrobial susceptibility testing of Salmonella enterica, subspecies enterica in Meat from broilers (Gallus gallus) - meat preparation

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.03										2					
0.064							2								
<=0.25				1											
<=0.5									1						
0.5				1										1	2
<=1								2							
					2				1					1	
<=2													1		
2		1													
4		1											1		
8											2				
16			2			2									
32												1			
64												1			

Table Antimicrobial susceptibility testing of Salmonella enterica, subspecies enterica in Meat from broilers (Gallus gallus) - meat preparation

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	1	0	0	0	1	0	0	0	0	0	0	0	0
<=0.03										1					
0.12							1								
<=0.25				1										1	
<=0.5					1										
0.5															1
<=1								1							
1									1						
<=2		1											1		
2		1									1				
8 16						1					1				
32			1									1			
32			<u> </u>									'			

Table Antimicrobial susceptibility testing of Salmonella enterica, subspecies enterica in Meat from goat - fresh - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015			·				1	·							
<=0.03										1					
<=0.25				1										1	
<=0.5									1						
0.5															1
<=1		1						1							
1					1										
<=4											1				
4			1										1		
<=8						1									
64												1			

Table Antimicrobial susceptibility testing of Salmonella enterica, subspecies enterica in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	1	0	0	0	0	0	0	0	0	0	1	1	0	0
<=0.03										2					
0.03							1								
0.064							1								
<=0.25				2										1	1
<=0.5					1				1						
0.5															1
<=1		11						2							
1					1								<u> </u>	1	
<=2													1		
2									1						
<=4											1				
<=8						1									
8 16			0			1					1				
32			2			1						1			
64		1										<u> </u>	1		
1024		1										1	ı		
1024												ı			

Table Antimicrobial susceptibility testing of Salmonella enterica, unspecified O:9 in Gallus gallus (fowl) - laying hens - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

Ampic Cefota Cefota Colist Chlora Genta Sulfan Tigec	
ECOFF 8 16 0.5 2 16 0.064 2 2 0.125 16 256 8 1	2
Lowest limit 1 2 0.25 0.5 8 0.015 1 0.5 0.03 4 8 2 0.25	0.25
Highest limit 64 64 4 8 128 8 16 32 16 128 1024 64 8	32
N of tested isolates 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	1
N of resistant MIC isolates 0 0 0 0 0 0 0 0 0 0 0 0	0
<=0.015	
<=0.03	
<=0.25 1	11
<=0.5	
<=1 1	
<=2 1	
2	
<=4	
<=8 1	
128	

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Meat from broilers (Gallus gallus) - meat preparation

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Netherlands

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	1	0	0	0	0	0	1	0
0.03							1								
0.064										1					
<=0.25				1											
<=0.5					1				1						
<=1		1													
1															1
2														1	
<=4								1			1		1		
4								1					1		
<=8						11									
8			1									4			_
128												1			

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - laying hens - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

Nalidixic acid	Sulfamethoxazol	Tigecycline	Trimethoprim
16 2	256 8	1	2
4	8 2	0.25	0.25
128 1	1024 64	4 8	32
2	2 2	2	2
0	0 0	0	0
		2	2
	2		
2			
	2		
1	16 4 128 2	### ##################################	16 256 8 1 4 8 2 0.25 128 1024 64 8 2 2 2 2 0 0 0 0 0 0

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	1	0	0	0	0	0	0	0
<=0.03										1					
0.03							1								
<=0.25				1											
<=0.5					1				1						
0.5														1	1
<=2													1		
2		1													
<=4								<u> </u>			1				
4								1							
<=8						1									
8			1									4			
64												1			

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - dust

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	1	0	0	0	0	0	0	0
<=0.03										1					
0.03							1								
<=0.25				1											
<=0.5					1				1						
0.5														1	1
<=2													1		
2		1													
<=4											1				
4								1							
<=8						1									
8			11									4			
64												1			

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Meat from poultry, unspecified - meat preparation - intended to be eaten cooked

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	1	0	0	0	1	0	0	0	0
<=0.03										1					
<=0.25				1										1	1
0.25							1								
<=0.5					1										
<=1		1						1							
1									1						
<=2													1		
<=8						1									
16			1												
32												1			
128											1				

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Egg products

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	1	0	0	0	1	0	0	0	0
<=0.03										1					
0.12							1								
<=0.25				1											
<=0.5					1				1						
0.5														1	1
<=1		1											<u> </u>		
<=2													1		
2								1							
<=8			4			1									
8			1									1			
64 128											1	<u> </u>			
120											1				

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	8	8	8	8	8	8	8	8	8	8	8	8	8	8
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							6								
<=0.03										8					
0.03							2								
<=0.25				8										8	8
<=0.5					8				8						
<=1		8						7							
<=2													8		
2								1							
<=4											8				
4			7												
<=8						8									
8			1												
64												8			

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - dust

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	5	5	5	5	5	5	5	5	5	5	5	5	5	5
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							4								
<=0.03										5					
0.03							1								
<=0.25				5										5	5
<=0.5					5				5						
<=1		3						4							
<=2													5		
2		2						1							
<=4											5				
4			5												
<=8						5									
32												2			
64												3			

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										2					
0.03							1								
<=0.25				2										2	2
<=0.5					2				2						
<=1		1													
<=2													2		
2		1						2							
<=4											2				
4			2												
<=8						2									,
32												1			
64												1			

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	20	20	20	20	20	20	20	20	20	20	20	20	20	20
MIC	N of resistant isolates	0	0	0	0	0	7	2	0	0	7	1	0	0	0
<=0.03										20					
0.03							13								
<=0.25				20										15	5
<=0.5					20				16						
0.5							7							5	15
<=1		10						14							
1									4						
<=2													20		
2		10						4							
<=4											12				
4			3					2							
<=8						20									
8			11								1				
16			6												
32												14			
64												5			
128											7				
1024												1			

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Netherlands

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	27	27	27	27	27	27	27	27	27	27	27	27	27	27
МІС	N of resistant isolates	1	0	0	0	0	2	4	0	0	1	1	0	1	0
<=0.015							4								
<=0.03										27					
0.03							21								
0.12							1								
<=0.25				27										16	11
<=0.5					27				25						
0.5							1							8	15
<=1		16						12							
1									2					2	1
<=2													26		
2		10						11							
<=4											25				
4			13					4					1	1	
<=8						27									
8			12								1				
16			2									2			
32		4									1	15			
64		1										9			
1024												1			

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	6	6	6	6	6	6	6	6	6	6	6	6	6	6
MIC	N of resistant isolates	0	0	0	0	0	0	2	0	0	0	0	0	0	0
<=0.03										5					
0.03							6								
0.064										1					
<=0.25				6										2	2
<=0.5					6				6						
0.5														4	4
<=1		1						1							
<=2		_											6		
2		5						3							
<=4 4			2								6				
<=8			3			6		2							
8			3			0									
32			<u> </u>									1			
64												4			
128												1			

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Germany

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<=0.03										3					
0.03							3								
<=0.25				3											2
<=0.5					3				3						
0.5														2	1
<=1		1						2							
<=2													2		
2		2						1							
<=4											2				
4													1	1	
<=8						3									
8			3								1	<u> </u>			
32												1			
64												2			

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: France

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.03										2					
0.03							2								
<=0.25				2										1	1
<=0.5					2				1						
0.5														11	11
<=1								1							
1									1						
<=2													2		
2		2						1							
<=4			<u> </u>								2				
4			1			•									
<=8			4			2									
8 16			1									1			
32												1			
- 52												ı			

Table Antimicrobial susceptibility testing of Salmonella Gaminara in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	14	14	14	14	14	14	14	14	14	14	14	14	14	14
MIC	N of resistant isolates	0	0	0	0	0	0	0	1	0	0	2	0	0	0
<=0.015							14								
<=0.03										13					
0.064										1					
<=0.25				14										12	13
<=0.5					14				13						
0.5														2	1
<=1		4						14							
<=2													14		
2		10													
<=4											14				
4			7			10									
<=8						13									
8 16			7			1			4						
32						1			1			2			
64												<u>3</u>			
128												1			
>1024												2			
71024															

Table Antimicrobial susceptibility testing of Salmonella Give in Other food

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										2					
0.03							1								
<=0.25				2										2	2
<=0.5					2				1						
<=1		2						2							
1									11						
<=2													2		
<=4						2					2				
<=8						2									
32			2									1			
64												1			
- U 1												<u> </u>			

Table Antimicrobial susceptibility testing of Salmonella Give in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	27	27	27	27	27	27	27	27	27	27	27	27	27	27
МІС	N of resistant isolates	0	0	0	0	0	2	0	0	0	1	1	0	0	1
<=0.015							1								
<=0.03										25					
0.03							23								
0.064							1			1					
0.12							2			1					
<=0.25				24										14	25
<=0.5					24				22						
0.5				3										12	1
<=1		24						24							
1					2				5					1	
<= <u>2</u>													26		
2		2			1			3							
<=4											26				
4		1	18										1		
<=8						27									
8			6												
16			3									5			1
32												11			
64											1	6			
128												4			
1024												1			

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.03										2					
0.03							2								
<=0.25				2										1	2
<=0.5					2				1						
0.5														1	
<=1		2						2							
1									1						
<=2													2		
<=4			4								2				
<u>4</u> <=8			1												
16			1			2									
32			ı									2			
<u> </u>															

Table Antimicrobial susceptibility testing of Salmonella Idikan in Gallus gallus (fowl) - laying hens - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
4			1												
<=8						1						1			
64												I			

Table Antimicrobial susceptibility testing of Salmonella Idikan in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		11						1							
<=2													1		
<=4											1				
4			1												
<=8						1									
128												1			

Table Antimicrobial susceptibility testing of Salmonella Idikan in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
0.03							1								
0.064										1					
<=0.25				2										1	2
<=0.5					2				2						
0.5														1	
<=1		2						2							
<=2													2		
<=4											2				
4			2												
<=8						2									
64												1			
128												1			

Table Antimicrobial susceptibility testing of Salmonella Idikan in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - dust

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		11						1							
<=2													1		
<=4											1				
4			1												
<=8						1									
128												1			

Table Antimicrobial susceptibility testing of Salmonella Indiana in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		11						1							
<=2													1		
<=4											1				
4			1			<u> </u>									
<=8						1						4			
64												1			

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	4	4	4	4	4	4	4	4	4	4	4	4	4	4
MIC	N of resistant isolates	1	0	0	0	0	4	0	0	0	4	4	0	0	0
<=0.03										4					
<=0.25				4										1	3
<=0.5					4				4						
0.5							3							3	1
<=1		2						4							
1							1								
<=2													3		
2		1											1		
4						0							1		
<=8						3									
8 16			3			1									
64		1	<u> </u>			<u>'</u>									
128		ı									4				
1024											4	4			
-1021												<u>.</u>			

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from pig - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	7	7	7	7	7	7	7	7	7	7	7	7	7	7
MIC	N of resistant isolates	0	1	0	0	0	1	0	0	0	1	1	0	0	0
<=0.03										6					
0.03							6								
0.064										1					
<=0.25				7										2	4
<=0.5					7				5						
0.5														4	3
<=1		4						7							
1							1		2					1	
<=2													7		
2		3													
<=4											6				
<=8						7									
8			6												
32			1									4			
64												2			
128											1				
1024												1			

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from broilers (Gallus gallus) - meat preparation

Sampling Stage: Retail Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Netherlands

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	1	1	0	0	0	1	1	1	1	0
<=0.03										1					
<=0.25				1											
<=0.5					1				1						
0.5															1
<=1								1							
1							1							4	
2														1	
16		1	1												
32			·			1									
64						•							1		
128											1				
1024												1			

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from broilers (Gallus gallus) - meat preparation

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

ECOFF 8 16 0.5 2 16 0.064 2 2 0.125 16 256 8 1 2 Lowest limit 1 2 0.25 0.5 8 0.015 1 0.5 0.03 4 8 2 0.25 0.25 Notested isolates 5		AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
Highest limit		ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
Nof tested		Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
Solition Solition		Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
MIC isolates 0 1 0 0 4 4 1 2 1 <=0.015		N of tested isolates	5	5	5	5	5	5	5	5	5	5	5	5	5	5
\$\sigma \cdot 0.25\$ 5 2 3 0.25 1 \$\sigma \cdot 0.5\$ 4 0.5 1 1 1 \$ 1 5 1 1 2 1 1 \$ 2 1 1 \$ 2 1 1 \$ 2 2 2 \$ 1 1 1 \$ 8 1 1 1 \$ 1 1 1 1 \$ 1 1 1 1 \$ 1 1 1 1 \$ 1 1 1 1 \$ 1 1 1 1 \$ 1 1 1 1 \$ 1 1 1 1 \$ 1 1 1 1 \$ 1 1 1 1 \$ 1 1 1 1 \$ 1 1 1 1 1 \$ 1 1 1 1 1 \$ 1 1 1 1 1 1 1		N of resistant isolates	0	1	0	0	0	4	0	0	0	4	4	1	2	11
<=0.25								1								
0.25 1 <0.5											5					
<=0.5					5										2	3
0.5 1 1 <=1								1								
<=1						5				4						
1 2 1 1 <=2								1								1
<=2			2					2	5	1						
2 2 <=4															ı	
<=4			2											4	2	
4 1 1 <=8												1				
<=8			1	1								·				
8 1 16 2 32 1 64 1 128 4			•				5									
16 2 32 1 64 1 128 4	8			1												
64 128 4	16			2									1			
128	32			1												1
														1		
1024												4				
	1024												4			

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from broilers (Gallus gallus) - meat preparation

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	4	4	4	4	4	4	4	4	4	4	4	4	4	4
MIC	N of resistant isolates	1	2	0	0	0	4	0	0	0	4	4	3	0	2
<=0.03										4					
<=0.25				4										1	1
0.25							3								
<=0.5					4				3						
0.5														1	1
<=1		1						4							
1							1		1					2	
<=2													1		
2		1													
<u>4</u> <=8		1				4									
8			2			4									
32			2												2
64		1											3		
128											4				
1024												4			
-															

Table Antimicrobial susceptibility testing of Salmonella Infantis in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	51	51	51	51	51	51	51	51	51	51	51	51	51	51
MIC	N of resistant isolates	15	0	0	0	0	47	0	0	0	47	47	23	0	16
<=0.015							4								
<=0.03										51					
0.12							4								
<=0.25				49										6	20
0.25							13								
<=0.5					46				51						
0.5				2			22							21	14
<=1		6						51							
1					5		7							24	1
<=2			1										23		
2		21					1								
<=4											4				
4		9	3										5		
<=8						32									
8			34												
16			13			19									
32												3			10
>32															16
64		45										1			
>64 128		15									10		23		
>128											12 35				
>120									107		33				

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	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	51	51	51	51	51	51	51	51	51	51	51	51	51	51
MIC	N of resistant isolates	15	0	0	0	0	47	0	0	0	47	47	23	0	16
>1024												47			

Table Antimicrobial susceptibility testing of Salmonella Infantis in Gallus gallus (fowl) - broilers

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	1	0	0	0	1	1	1	0	0
<=0.03										1					
<=0.25				1											
<=0.5					1				1						
0.5							1								1
<=1								1							
1		4												1	
4 <=8		1				1									
8			1												
>64			'										1		
>128											1		•		
>1024												1			

Table Antimicrobial susceptibility testing of Salmonella Infantis in Gallus gallus (fowl) - broilers

Sampling Stage: Slaughterhouse

Sampling Type: animal sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	0	0	0	0	0	1	0	0	0	1	1	1	0	0
<=0.03										1					
<=0.25				1											
<=0.5									1						
0.5															1
<=1								1							
1					1		1							1	
4		1													
16			1			1									
64													1		
128											1				
1024												1			

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from poultry, unspecified - meat preparation - intended to be eaten cooked

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	0	1	0	0	0	2	0	0	0	2	2	1	1	1
<=0.03										2					
<=0.25				2											1
0.25							1								
<=0.5					2				1						
0.5														1	
<=1		1						2							
1							1		1						
<=2		1											1		
2 <=8		1				2								1	
8			1			2									
32			1												1
64													1		1
128											2		'		
1024												2			

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat, mixed meat - meat preparation

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	1	0	0	0	1	1	1	0	1
<=0.03										1					
<=0.25				1											
0.25							1								
<=0.5					1				1						
<=1								1							
1														1	
4		1				4									
<=8 16			1			1									
32			ı												1
64													1		
128											1		•		
1024												1			

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from poultry, unspecified - carcase

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	0	0	0	0	0	1	0	0	0	1	1	0	0	0
<=0.03										1					
<=0.25				1											
<=0.5					1				1						
0.5							1							1	1
<=1								1							
<=2													1		
2		1													
8			1												
16						1									
128											1				
1024												1			

Table Antimicrobial susceptibility testing of Salmonella Infantis in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes Programme Code: AMR MON

Sampler: Official and industry sampling

Analytical Method: Dilution - sensititre

Sampling Strategy: Census

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							3								
<=0.03										3					
<=0.25				3										3	3
<=0.5					3				3						
<=1		2						3							
<=2													3		
2		1													
<=4											3				
4			3												
<=8						3									
32												1			
64												1			
128												1			

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Table Antimicrobial susceptibility testing of Salmonella Infantis in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - dust

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
4			1												
<=8						1						4			
128												1			

Sampling Stage: Slaughterhouse Sampl

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance Cefotaxime	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Synergy test									Not Available	
	ECOFF	0.125	0.5	0.5	8	2	2	0.06	1	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	2	2	2	2	2	2	2	2	2	2
	N of resistant				_		_	_	_	_	
MIC	isolates	1	0	0	2	0	0	0	0	0	0
<=0.015								2		0	
<=0.03 0.12		1								2	
<=0.25		ı	2								
0.25		1	2	2					2		
0.5		•				2	2				
8											1
16					2						1

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	71	71	71	71	71	71	71	71	71	71	71	71	71	71
MIC	N of resistant isolates	16	11	1	0	0	67	1	0	0	67	67	24	2	18
<=0.015							3								
<=0.03							-			67					
0.03							1								
0.064										1					
0.12							1			3					
<=0.25				70										10	35
0.25							19								
<=0.5					68				66						
0.5							29							38	17
<=1		16						70							
1					2		17		5					21	
<=2													43		
2		33			1		1							2	1
<=4											4				
4		6		1				1					4		
<=8						61									
8			47												
16			13			10									
32		1	9									3			18
64		15	2								1		24		
128											66	1			
1024												67			

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Germany

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	4	4	4	4	4	4	4	4	4	4	4	4	4	4
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										4					
0.03							3								
<=0.25				4										1	4
<=0.5					4				4						
0.5														3	
<=1								4							
<=2													4		
2		4													
<=4			<u> </u>								4				
4			1												
<=8						4									
8			3									2			
32 64												3			
04												<u> </u>			

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: France

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	1	0	0	0	1	1	0	0	0
<=0.03										1					
<=0.25				1											
<=0.5					1				1						
0.5							1							1	1
<=1								1							
2		1													
4													1		
<=8						1									
8			1												
128											1				
1024												1			

Table Antimicrobial susceptibility testing of Salmonella Javiana in Frogs leg

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Indonesia

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.03										1					
0.03							1								
<=0.25				1											1
<=0.5					1				1						
0.5														1	
<=1		1						1							
<=2													1		
<=4											1				
4			1												
<=8						1									
32												1			

Table Antimicrobial susceptibility testing of Salmonella Kasenyi in Other food

Sampling Stage: Slaughterhouse

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2			1										1		
<=4											1				
<=8						1									
64												1			

Table Antimicrobial susceptibility testing of Salmonella Kedougou in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.03										1					
0.03							1								
<=0.25				1										1	
<=0.5					1				1						
0.5															1
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									
8			1												
128												1			

Table Antimicrobial susceptibility testing of Salmonella Kentucky in Meat from broilers (Gallus gallus) - meat preparation

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Poland

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	0	0	0	1	0	0	0	1	0	0	0	0
<=0.03										1					
<=0.25				1											1
<=0.5									1						
0.5														1	
<=1								1							
1					1										
<=2			1										1		
<u>4</u> <=8			1			1									
8						'	1								
32							'					1			
64		1													
128											1				

Table Antimicrobial susceptibility testing of Salmonella Kentucky in Other food

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	0	0	0	1	0	0	0	1	1	1	0	0
<=0.03										1					
<=0.25				1											
<=0.5									1						
0.5														1	1
<=1								1							
1					1										
<=8						1									
8			1				1								
64		1											1		
128											1				
1024												1			

Table Antimicrobial susceptibility testing of Salmonella Kottbus in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1						1			
8			1												

Table Antimicrobial susceptibility testing of Salmonella Livingstone in Meat from pig - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	7	7	7	7	7	7	7	7	7	7	7	7	7	7
MIC	N of resistant isolates	0	1	0	0	0	0	0	0	0	0	0	1	2	0
<=0.015							2								
<=0.03										6					
0.03							4								
0.064							1			1					
<=0.25				7										5	2
<=0.5					7				6						
0.5															3
<=1		4						6							
1									1						2
<=2		_											4		
2		3						1						2	
<=4											7				
4			2										2		
<=8						5									
8			4			0						2			
32						2						2			
64			1									3	4		
04												2	1		

Table Antimicrobial susceptibility testing of Salmonella Livingstone in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	5	5	5	5	5	5	5	5	5	5	5	5	5	5
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							4								
<=0.03										5					
0.03							1								
<=0.25				5										5	1
<=0.5					5				5						
0.5															4
<=1		5						5							
<=2													5		
<=4											5				_
4			2												
<=8						5									
8			3												
32												1			
64												4			

Table Antimicrobial susceptibility testing of Salmonella Livingstone in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	
<=0.5					1				1						
0.5															11
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									
8			1												
64												1			

Table Antimicrobial susceptibility testing of Salmonella Livingstone in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - dust

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	11
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									
8			11												
64												1			

Table Antimicrobial susceptibility testing of Salmonella Livingstone in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							3								
<=0.03										3					
<=0.25				3										3	3
<=0.5					3				3						
<=1		3						3							
<=2													3		
<=4											3				
4			1												
<=8						3									
8			2												
64												3			

Table Antimicrobial susceptibility testing of Salmonella Llandoff in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8			<u> </u>			1									
8			11												
32												1			

Table Antimicrobial susceptibility testing of Salmonella London in Meat from pig - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	1	0	0	0	0	0	0	0	0	0	1	0	0	1
<=0.015							1								
<=0.03										1					
0.064							1			1					
<=0.25				2										2	1
<=0.5					2				2						
<=1		1						2							
<=2			1										1		
<=4			<u> </u>								2				
4			1			4									
<=8						1						1	1		
8						4							1		
16 32						ı									1
64		1													ı
1024												1			

Table Antimicrobial susceptibility testing of Salmonella Mbandaka in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
МІС	N of resistant isolates	0	0	0	0	0	1	0	2	0	1	2	1	0	0
<=0.015							2								
<=0.03										3					
<=0.25				3										2	2
<=0.5					3				1						
0.5							1							1	1
<=1		2	<u>.</u>					3							
<=2			1										2		
2		1									2				
<=4 <=8						3					2	1			
8			2			3						ı			
16									1						
32									1						
>64									'				1		
>128											1				
>1024											· ·	2			

Table Antimicrobial susceptibility testing of Salmonella Mbandaka in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							2								
<=0.03										2					
<=0.25				2										2	
<=0.5					2				2						
0.5															2
<=1		2						2							
<=2													2		
<=4											2				
<=8						2									
8			2												
64												1			
128												1			

Table Antimicrobial susceptibility testing of Salmonella Mbandaka in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - dust

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							2								
<=0.03										2					
<=0.25				2										2	2
<=0.5					2				1						
<=1								2							
1									1						
<=2													2		
2		2													
<=4						•					2				
<=8						2									
8			2									1			
64												1			
128												<u> </u>			

Table Antimicrobial susceptibility testing of Salmonella Mbandaka in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	1	0	0	0	0	0	0	0	0	0	1	0	0	0
<=0.015							2								
<=0.03										2					
<=0.25				2										1	
<=0.5					2				2						
0.5														1	2
<=1		1						2					<u> </u>		
<=2													2		
<=4											2				
<=8						2									
8 64			2												
128		ı										1			
1024												1			
1024												· · ·			

Table Antimicrobial susceptibility testing of Salmonella Mbandaka in Meat from broilers (Gallus gallus) - carcase - spent hens

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: France

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.03										3					
0.03							3								
<=0.25				3										1	
<=0.5					3				3						
0.5														1	3
<=1		2						3						<u> </u>	
1													•	1	
<=2 2		1											2		
<=4		<u> </u>									3				
4											ა		1		
<=8						2							·		
8			3			2									
16						1									
64						•						1			
128												1			
256												1			

Table Antimicrobial susceptibility testing of Salmonella Milwaukee in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	6	6	6	6	6	6	6	6	6	6	6	6	6	6
MIC	N of resistant isolates	0	0	0	0	0	1	0	0	0	0	0	0	2	0
<=0.03										6					
0.03							5								
0.12							1								
<=0.25				6											6
<=0.5					6				6						
0.5														2	
<=1								6							
1														2	
<=2		-											2		
2		6									_			2	
<=4											5				
4			4								1		4		
8 16			1 5			6					1				
32			5			б						2			
64												3			

Table Antimicrobial susceptibility testing of Salmonella Moero in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	
<=0.5					1				1						
0.5															1
<=1		1						1							
<=2											1		1		
<=4 <=8						1					1				
8			1			ı									
16												1			

Table Antimicrobial susceptibility testing of Salmonella Newport in Compound feedingstuffs, not specified - final product - non-pelleted/meal

Sampling Stage: Farm

Sampling Type: feed sample

Sampling Context: Control and eradication

Sampler: Official sampling

Sampling Strategy: Objective sampling

programmes Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							3								
<=0.03										3					
<=0.25				3										3	3
<=0.5					3				3						
<=1		3						3							
<=2													3		
<=4											3				
4			3												
<=8						3									
64												3			

Table Antimicrobial susceptibility testing of Salmonella Ohio in Meat from pig

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.03										1					
0.064							1								
<=0.25				1											
<=0.5					1				1						
0.5														1	11
<=1								1							
2		1													
<=4											1				
4													1		
16			1			1									
32												1			

Table Antimicrobial susceptibility testing of Salmonella Panama in Frogs leg

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Indonesia

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.03										1					
0.03							1								
<=0.25				1										1	1
<=0.5					1										
<=1		1						1							
1									1						
<=2													1		
<=4											1				
<=8			4			1						4			
16			1									1			

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Netherlands

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	1	0	0	1
<=0.015							1								
<=0.03										1					
<=0.25				1										1	
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
4			1												
<=8						1									
32															1
1024												1			

Sampling Stage: Slaughterhouse Sampling Type: food sample - carcase swabs Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance Cefotaxime synergy test	e elebime Celebime Not Available	Cefotaxi n Oot Available	Cefotaxime + Clavulanic acid	Cefoxitin Cot Available	Ceftazidim Not Available	DOM Ceftazidime + Clavulanic acid	E-table with the Not Available	E e e od E E Not Available	E e u e d o o o o e u e o o o o o o o o o o o o o	⊑ E E E E E E E E E E E E E E E E E E E
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.5	0.5	8	2	2	0.06	1	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	1	1	0	1	0	0	0	0	0	0
<=0.015								1			
<=0.03										1	
0.25				1					1		
0.5							1				
2						1					
4		1	1								
16					1						1

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	1	0	0	1	0	0	0	1	0	0	0	1
<=0.03										1					
<=0.5									1						
0.5							1								
<=1								1							
														1	
2				1	1										
4													1		
<=8						11									
8			1												
16												1			
32															1
64		1													
128											1				

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	0	0	1	1	0	0	0	1	1	1	0	1
<=0.03										1					
<=0.25				1											
0.25							1								
<=0.5					1				1						
0.5														1	
<=1								1							
8			1												
32													1		1
64		1													
128						1					1				
1024												1			

Sampling Stage: Retail Sampling Type: food sample Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

		AM substance Cefotaxime synergy test Ceftazidime									E e e e e e e e e e e e e e	
Lowest limit 0.064 0.25 0.064 0.5 0.25 0.12 0.015 0.12 0.03 0.5			0.125	0.5	0.5	8	2	2	0.06	1	0.125	32
Highest limit 32 64 64 64 128 128 2 16 16 64 N of tested isolates 1 1 1 1 1 1 1 1 1										<u> </u>		0.5
N of tested												64
MIC isolates 1 1 0 0 1 0		N of tested	1	1	1		1	1	1	1	1	1
<=0.015		N of resistant										
0.064 1 0.25 1 1 0.5 1		isolates	1	1	0	0	1	0	0	0	0	0
0.25 1 1 0.5 1									1			
0.5											1	
					1					1		
4 1 1								1				
			1				1					
8 1 1				1		1						
16	16											1

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	4	4	4	4	4	4	4	4	4	4	4	4	4	4
MIC	N of resistant isolates	4	0	1	0	0	4	0	0	0	4	0	0	0	4
<=0.03										4					
0.12							1								
<=0.25				3										2	
0.25							1								
<=0.5					3				4						
0.5							2							1	
<=1								3							
<u>1</u> <=2			1										4	1	
2			'	1	1			1					4		
4			1	ı											
<=8			<u>'</u>			4									
8			2			7									
16												2			
32												 1			4
64		4													
128											4	1			

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
МІС	N of resistant isolates	2	0	0	0	0	2	0	0	0	2	2	0	1	2
<=0.03										2					
<=0.25				2											
<=0.5					2				2						
<=1								2							
_1							1							1	
2							1								
4													1	1	
<=8						1									
8			1										1		
16			1			1									
32		-													2
64		2													
128											2	-			
1024												2			

Table Antimicrobial susceptibility testing of Salmonella Paratyphi B in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	3	0	0	3
<=0.015							3								
<=0.03										3					
<=0.25				3										3	
<=0.5					3				3						
<=1		3						3							
<=2													3		
<=4											3				
4			3												
<=8						3									
>32															3
>1024												3			

Table Antimicrobial susceptibility testing of Salmonella Paratyphi B in Gallus gallus (fowl) - broilers

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	0	0	0	1	0	0	0	1	1	0	0	1
<=0.03										1					
<=0.25				1											
<=0.5					1				1						
<=1								1							
_1							1							1	
<=2			11												
4													1		
16						1									
>32															1
>64		1									4				
>128											11	4			
>1024												1			

Table Antimicrobial susceptibility testing of Salmonella Paratyphi B in Meat, mixed meat - meat preparation

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	1	0	0	1
<=0.03										1					
0.03							1								
<=0.25				1											
<=0.5					1				1						
0.5														1	
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									
8			11												
32															1
1024												1			

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance Cefotaxime synergy test Ceftazidime				يا کې کې O Not Available ا						
	synergy test										
	ECOFF	0.125	0.5	0.5	8	2	2	0.06	11	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	3	3	3	3	3	3	3	3	3	3
MIC	N of resistant isolates	2	2	1	3	1	1	4	0	0	0
<=0.015	isolates			<u>'</u>	<u>_</u>	<u> </u>	<u> </u>	2	<u> </u>	<u> </u>	
<=0.015										2	
0.064										1	
0.004		1		1				1		1	
0.12		ı		1				ı	1		
0.5		1	1	<u>'</u>		1	2		1		
1		'					L		1		
2		1	1			1			•		
8						•					1
16				1	2		1				2
32			1			1					
64					1						

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	19	19	19	19	19	19	19	19	19	19	19	19	19	19
MIC	N of resistant isolates	9	2	3	1	1	8	0	2	0	8	15	1	0	18
<=0.015							2								
<=0.03										19					
0.03							9								
<=0.25				16										8	1
0.25							4								
<=0.5					16				13						
0.5							1							9	
<=1		10						18							
1				1	1		3		4					2	
<=2			1										14		
2				1	1			1							
<=4											11				
4			5	1									4		
<=8						18									
8			8		1				1						
16			3						1						
32			2									2	1		18
64		9										1			
128						1					8	1			
1024												15			

Table Antimicrobial susceptibility testing of Salmonella Rissen in Meat from pig - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	4	4	4	4	4	4	4	4	4	4	4	4	4	4
MIC	N of resistant isolates	0	0	0	0	0	4	0	0	0	0	0	4	0	0
<=0.03										4					
<=0.25				4											2
<=0.5					4				3						
0.5							4							2	2
<=1		4						4							
1									1					2	
<=8						4									
8			2												
16			2								4				
32												4			
64													4		

Table Antimicrobial susceptibility testing of Salmonella Rissen in Other feed material

Sampling Stage: Slaughterhouse

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										2					
0.03							1								
<=0.25				2										2	
<=0.5					2				1						
0.5															2
<=1		2						2							
1									1						
<=2													2		
<=4											2				
<=8			0			2									
8 128			2									2			
120												2			

Table Antimicrobial susceptibility testing of Salmonella Rissen in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	11
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
4			1												
<=8						1									
64												1			

Table Antimicrobial susceptibility testing of Salmonella Saintpaul in Frogs leg

Sampling Stage: Retail Sampling Type: food sample Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Vietnam

Sampling Details: N_A

	AM substance Cefotaxime synergy test Ceftazidime					Ceffazidim B aldaliavA vo					
	synergy test					Not Available I					
	ECOFF	0.125	0.5	0.5	8	2	2	0.06	1	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Lowest limit Highest limit	0.064 32	0.25 64	0.064 64	0.5 64	0.25 128	0.12 128	0.015	0.12 16	16	64
MIC	Highest limit N of tested	32 1	64	64	64	128	128	2	16		
MIC <=0.03	Highest limit N of tested isolates N of resistant	1	1	1	1	128	128	1	16	16	1
	Highest limit N of tested isolates N of resistant	1	1	1	1	128	128	1	16	16 1 0	1
<=0.03	Highest limit N of tested isolates N of resistant	1	1	1	1	128	128	1 0	16	16 1 0	1
<=0.03 0.03	Highest limit N of tested isolates N of resistant	1	1	1	1	128	128	1 0	16 1 0	16 1 0	1
<=0.03 0.03 0.25	Highest limit N of tested isolates N of resistant	1	1	1 0	1	128	128	1 0	16 1 0	16 1 0	1
<=0.03 0.03 0.25 0.5	Highest limit N of tested isolates N of resistant	1	1	1 0	1	128	128	1 0	16 1 0	16 1 0	1
<=0.03 0.03 0.25 0.5	Highest limit N of tested isolates N of resistant	1	1	1 0	1 1	128	128	1 0	16 1 0	16 1 0	1

Table Antimicrobial susceptibility testing of Salmonella Saintpaul in Frogs leg

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Vietnam

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	1	1	1	1	1	1	1	0	1	1	1	1	1
<=0.03										1					
2														1	
4				1				1							
8					1		1								
32									1						1
64		1	1										1		
128						1					1				
1024												1			

Table Antimicrobial susceptibility testing of Salmonella Senftenberg in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	1	0	0	0	1	0	0	0	0
<=0.03										1					
0.12							1								
<=0.25				1										1	
<=0.5					1				1						
0.5															1
<=1								1							
<=2													1		
2		1													
4			1			4									
<=8						1									
32 128											1	!			
120															

Table Antimicrobial susceptibility testing of Salmonella Senftenberg in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1								1							
<=2													1		
2		1													
<=4											1				
4			1												
<=8						1									
128												1			

Table Antimicrobial susceptibility testing of Salmonella Senftenberg in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<=0.03										3					
0.03							3								
<=0.25				3										1	
<=0.5					3				2						
0.5														2	3
<=1		3						3							
1									1						
<=2													3		
<=4											3				
4			3												
<=8						3									
128												1			
256												1			_
1024												1			

Table Antimicrobial susceptibility testing of Salmonella spp., unspecified in Meat from pig - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	8	8	8	8	8	8	8	8	8	8	8	8	8	8
MIC	N of resistant isolates	6	0	0	0	1	1	0	0	0	0	5	6	2	3
<=0.015							1								
<=0.03										7					
0.03							4								
0.064							2			1					
<=0.25				8										1	3
0.25							1								
<=0.5					6				8						
0.5														3	1
<=1								8							
1					1									2	1
<=2													2		
2		2			1						<u> </u>			11	
<=4											6				
4														1	
<=8						5					•				
8			4			-					2	4			
16			4			2						1	4		0
32 64		-										4	1		3
128		6				1						1	5		
1024												5			
1024												<u> </u>			

Table Antimicrobial susceptibility testing of Salmonella spp., unspecified in Meat from broilers (Gallus gallus) - meat preparation

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Netherlands

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	0	0	0	0	0	0	0	0	1	1	0	1
<=0.03										1					
0.03							1								
<=0.25				1											
<=0.5					1				1						
0.5														1	
<=1								1							
<=4											1				
<=8						1									
8			1												
32															1
64		1										4	1		
1024												1			

Table Antimicrobial susceptibility testing of Salmonella spp., unspecified in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
0.064										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1								1							
<=2													1		
2		1													
<=4											1				
<=8						1									
8			1												
32												1			

Table Antimicrobial susceptibility testing of Salmonella spp., unspecified in Meat from poultry, unspecified - meat preparation - intended to be eaten cooked

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	1	0	0	1
<=0.03										1					
0.03							1								
<=0.25				1											
<=0.5					1				1						
0.5														1	
<=1		1						1							
<=2													1		
<=4											1				
4			1												
<=8						1									•
32															1
1024												1			

Table Antimicrobial susceptibility testing of Salmonella spp., unspecified in Compound feedingstuffs, not specified - final product - non-pelleted/meal

Sampling Stage: Farm

Sampling Type: feed sample

Sampling Context: Control and eradication

Sampler: Official sampling

Sampling Strategy: Objective sampling

programmes Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
4			1												
<=8						1						4			
256												1			

Table Antimicrobial susceptibility testing of Salmonella spp., unspecified in Meat from goat - fresh - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	0	0	0	1	0	0	0	1	1	1	0	1
<=0.03										1					
<=0.25				1											
0.25							1								
<=0.5					1				1						
0.5														1	
<=1								1							
<=8						1									
16			1												
32															1
64		1											1		
128											1				
1024												1			

Table Antimicrobial susceptibility testing of Salmonella spp., unspecified in Cheeses, made from unspecified milk or other animal milk

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	1	0	0	0	1	0	0	0	1	0	0	0	0
<=0.03										1					
<=0.25				1										1	
<=0.5					1										
0.5							1								1
<=1		1						1							
1									1						
<=2													1		
<=8						1									
32			11									1			
128											1				

Table Antimicrobial susceptibility testing of Salmonella spp., unspecified in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceffazidim	Ceffazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
synergy test										
synergy test	Not Available			Not Available		Not Available		Not Available		
ECOFF	0.125	0.5	0.5	8	2	2	0.06	1	0.125	32
Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
Highest limit	32	64	64	64	128	128	2	16	16	64
N of tested isolates	2	2	2	2	2	2	2	2	2	2
isolates	1	2	1	1	2	1	0	0	0	0
							1			
							1		2	
	1		1							
						1				
								1		
								1		
	1	1								
				1	1					
										1
		1	1			1				1
				1	1					
	Cefotaxime synergy test Ceftazidime synergy test ECOFF Lowest limit Highest limit N of tested isolates	Cefotaxime synergy test Ceftazidime synergy test ECOFF 0.125 Lowest limit 0.064 Highest limit 32 N of tested isolates 2 N of resistant isolates 1	Cefotaxime synergy test Ceftazidime synergy test ECOFF 0.125 0.5 Lowest limit 0.064 0.25 Highest limit 32 64 N of tested isolates 2 2 N of resistant isolates 1 1	AM substance Cefotaxime synergy test Ceftazidime synergy test ECOFF 0.125 0.5 0.5 Lowest limit 0.064 0.25 0.064 Highest limit 32 64 64 N of tested isolates 2 2 2 2 N of resistant isolates 1 2 1	AM substance	AM substance	AM substance Ceftotaxime synergy test Ceftazidime synergy test BCOFF 0.125 0.05 0.064 0.064 0.064 0.064 0.064 0.064 0.064	AM substance	AM	AM substance

Table Antimicrobial susceptibility testing of Salmonella spp., unspecified in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	12	12	12	12	12	12	12	12	12	12	12	12	12	12
МІС	N of resistant isolates	8	0	2	2	1	10	0	0	0	10	6	0	0	9
<=0.03										11					
0.03							2								
0.064										1					
<=0.25				10										5	1
0.25							3								
<=0.5					10				12						
0.5							6							5	2
<=1		1						12							
_1														2	
<=2													10		
2		3		1			1								
<=4											2				
4			4	1	1								2		
<=8						9									
8			5		1										
16			3			2									
32												3			9
64		8										2			
128						1					10	1			
1024												6			

Table Antimicrobial susceptibility testing of Salmonella Stanley in Meat from broilers (Gallus gallus) - meat preparation

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Hungary

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	0	0	0	0	0	1	0	0	0	0	0	0	0	0
<=0.03										1					
<=0.25				1										1	1
0.25							1								
<=0.5					1				1						
<=1		1						1							
<=2													1		
4			1												
<=8						1					<u> </u>				
16											1	,			
128												11			

Table Antimicrobial susceptibility testing of Salmonella Teddington in Other processed food products and prepared dishes - vegetable based dishes

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.03										1					
0.03							1								
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
4			1												
<=8						11									
64												11			

Table Antimicrobial susceptibility testing of Salmonella Tennessee in Other feed material

Sampling Stage: Retail

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	
<=0.5					1				1						
0.5															11
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									
8			1												
64												1			

Table Antimicrobial susceptibility testing of Salmonella Tennessee in Gallus gallus (fowl) - laying hens - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

Ampicillin Ampicillin Ceftazidim Cefotaxim Colistin Gentamicin Meropenem Malidixic acid Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
ECOFF 8 16 0.5 2 16 0.064 2 2 0.125 16 256	8	1	2
Lowest limit 1 2 0.25 0.5 8 0.015 1 0.5 0.03 4 8	2	0.25	0.25
Highest limit 64 64 4 8 128 8 16 32 16 128 1024	64	8	32
N of tested isolates 2 2 2 2 2 2 2 2 2 2 2 2	2	2	2
N of resistant MIC isolates 0 0 0 0 0 0 0 0 0 0 0	0	0	0
<=0.015			
<=0.03			
<=0.25 2		2	
<=0.5			
0.5			2
<=1 2			
<=2	2		
<=4			
4 1			
<=8 2			
8 1			
64			

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from pig - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	28	28	28	28	28	28	28	28	28	28	28	28	28	28
MIC	N of resistant isolates	25	0	0	0	9	1	0	0	0	2	16	15	1	12
<=0.015							8								
<=0.03										27					
0.03							15								
0.064							4			1					
<=0.25				27										15	13
0.25							1								
<=0.5					28				27						
0.5				1										11	3
<=1		2						27							
1									1					1	
<=2			1										12		
2		1						1						1	
<=4											23				
4			12			40									
<=8			4.4			18						1			
8 16			14 1			4					3		1		
32			1			1						2	7		10
64		25									1	8 1	7		12
128		25				9					1	ı			
1024						9					- 1	16			
1024												10			

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from bovine animals - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	1	1	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=4											11				
4			1												
<=8						1							<u> </u>		
64												,	1		
1024												11			

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	20	20	20	20	20	20	20	20	20	20	20	20	20	20
MIC	N of resistant isolates	16	0	0	0	2	5	0	0	0	5	11	9	0	11
<=0.015							8								
<=0.03										20					
0.03							7								
<=0.25				20										16	9
0.25							3								
<=0.5					20				19						
0.5														3	
<=1		2						20							
							2							1	
<=2													9		
2		1							1						
<=4											15				
4		1	17										2		
<=8						17									
8 16			1			4									
32			2			1									
>32												5			11
64												4	1		- 11
>64		16										<u> </u>	8		
>128		10				2					5		0		
>1024											<u> </u>	11			
7 1024												- 11			

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Gallus gallus (fowl) - laying hens - day-old chicks

Sampling Stage: Farm

Sampling Type: environmental sample - delivery box liner

Sampling Context: Monitoring - active

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	1	0	0	0	1	0	0	0	0	0	1	1	0	0
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1								1							
<=4											1				
4			1												
64													1		
>64		1													
>128						1									
>1024												1			

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Gallus gallus (fowl) - broilers

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication

Sampler: Official and industry sampling

Sampling Strategy: Census

programmes Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
МІС	N of resistant isolates	1	0	0	0	2	1	0	0	0	1	2	2	0	1
<=0.015							1								
<=0.03										2					
<=0.25				2										2	1
0.25							1								
<=0.5					2				2						
<=1		1						2			<u> </u>				
<=4											1				
4			2												
>32													4		1
64 >64		1											1		
>128		ı				2					1		1		
>128						2					ļ	2			
7 1024															

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Compound feedingstuffs, not specified - final product - non-pelleted/meal

Sampling Stage: Farm

Sampling Type: feed sample

Sampling Context: Control and eradication

Sampler: Official sampling

Sampling Strategy: Objective sampling

programmes Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.03										1					
0.03							1								
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
4			1												
<=8						1									
16												1			

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat, mixed meat - meat preparation

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	0	0	0	0	0	0	0	0	1	1	0	0
<=0.03										1					
0.03							1								
<=0.25				1											1
<=0.5					1				1						
0.5														1	
<=1								1							
<=8						1									
8			1								1				
64		1											1		
1024												1			

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Crustaceans

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.03										1					
0.03							2								
0.064										1					
<=0.25				2											2
<=0.5					2				1						
0.5														2	
<=1		2						2							
1									1						
<=2													2		
<=4											2				
4			1												
<=8						2									
8			1												
64												2			

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from pig

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	0	0	1	0	0	0	0	0	1	1	0	0
<=0.03										1					
0.03							1								
<=0.25				1											1
<=0.5					1				1						
0.5														1	
<=1								1							
<=4											1				
8			1												
64		1											1		
128						1									
1024												1			

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from bovine animals and pig - meat preparation

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	0	0	0	0	0	0	0	0	1	1	0	1
<=0.03										1					
0.03							1								
<=0.25				1											
<=0.5					1				1						
0.5														1	
<=1								1							
<=4											1				
<=8						1									
8			1												
32															1
64		1											1		
1024												1			

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from broilers (Gallus gallus) - carcase - chilled

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
MIC	N of resistant isolates	1	1	0	0	1	0	0	0	0	0	3	1	0	1
<=0.015							1								
<=0.03										3					
0.03							2								
<=0.25				3										2	1
<=0.5					3				2						
0.5														1	1
<=1 <=2		2						3					2		
2									1				2		
<=4									<u>'</u>		2				
4			1												
<=8			•			2									
16			1								1		1		
32			1												1
64		1													
128						1									
1024												3			
	•						•								

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from broilers (Gallus gallus) - carcase - spent hens

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.03										1					
0.03							1								
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
4			1												
<=8						1						1			

Table Antimicrobial susceptibility testing of Salmonella Wandsworth in Frogs leg

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Indonesia

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<=0.03										1					
0.03							1								
<=0.25				1											
<=0.5					1				1						
0.5														1	1
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									
8			1												
1024												1			

Table Antimicrobial susceptibility testing of Salmonella Weltevreden in Other food

Sampling Stage: Retail

Sampling Type: food sample

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.5	2	16	0.064	2	2	0.125	16	256	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<=0.03										1					
0.03							1								
<=0.25				1										1	
<=0.5					1				1						
0.5															1
<=1		1						1							
<=2													1		
<=4											1				
4			1												
<=8						1									
1024												1			

ANTIMICROBIAL RESISTANCE TABLES FOR INDICATOR ESCHERICHIA COLI

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - carcase

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceffazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	1	0	0	0	0	0	0	0	0
<=0.015								1			
<=0.03										1	
<=0.064				1							
<=0.12									1		
0.25							1				
0.5		11				1					
1			1		1						4
8					1						I

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - carcase

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	1	0	1	0	0	0	0	0	1	1	0	1
<=0.015							1								
<=0.03										1					
<=0.25														1	
<=0.5					1										
<=1								1							
1				1					1						
<=4											1				
4			1												
32															1
64		1				1							1		
1024												1			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring - EFSA

Sampler: Official sampling

Sampling Strategy: Objective sampling

specifications Programme Code: ESBL MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Bangladesh

Sampl	ing Details: N_A										
	AM substance	O	Ε	Cefotaxime + Clavulanic acid		E	Ceftazidime + Clavulanic acid	E	F	ш	<u>.</u>
		Cefepime	Cefotaxim	Cefotaxii	Cefoxitin	Ceftazidim	Ceftazidi	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	0	1	0	0	1	0	0	0	0	0
<=0.015	isolates		•					1			
<=0.03										1	
<=0.064				1						•	
<=0.12							1		1		
0.12		1									
1			1								
4					1						
8											1
16						1					

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring - EFSA

Sampler: Official sampling

Sampling Strategy: Objective sampling

specifications Programme Code: ESBL MON

Analytical Method: Dilution - sensititre

Country of Origin: Bangladesh

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	1	1	0	1	0	0	0	1	1	1	0	1
<=0.03										1					
<=0.25														1	
0.25							1								
<=0.5									1						
<=1								1							
2				1											
<=8						1									
8			1		1										1
32		,													1
64		11									4		11		
128											1	4			
1024												1			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	lmipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available I	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available I	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	154	154	154	154	154	154	154	154	154	154
MIC	N of resistant isolates	138	154	28	36	143	30	4	1	0	0
<=0.015								109			
<=0.03										144	
0.03								37			
<=0.064		4		106							
0.064								4		9	
<=0.12							100		67		
0.12		12		19				3		1	
0.25		28		1			23		83		
0.5		27	3			11	1	1	3		
1		12	14	4	1	23	1				_
2		16	21	1	8	21	6				8
4		31	25	15	53	20	15				77
8		21	33	6	56	38	8				61
16		2	30	2	10	23			1		7

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceffazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	154	154	154	154	154	154	154	154	154	154
МІС	N of resistant isolates	138	154	28	36	143	30	4	1	0	0
32		1	21		11	13					1
64			7		15	4					
128						1					

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	154	154	154	154	154	154	154	154	154	154	154	154	154	154
MIC	N of resistant isolates	154	7	154	141	47	108	0	18	0	104	130	86	7	94
<=0.015							34								
<=0.03										151					
0.03							11								
0.064							1			3					
0.12							4								
<=0.25														104	25
0.25							29								
<=0.5					13				94						
0.5				3			19							32	25
<=1								149							
1				12	29		7		38					11	7
<=2			10				_	_					62		
2 <=4				22	16		5	5	4		4.4			4	3
4			74	117	19		7				44		5	2	
<=8			74	117	19	105						11	5	3	
8			56		77	105	37		3		3	- 11	1		
16			7		11	2	31		3		3	5	2		
32		1	4			10			12		2	6	6		94
64		153	3			15			12		5	2	78		J-1
128		100				22					97		, 5		
											<u> </u>				

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	154	154	154	154	154	154	154	154	154	154	154	154	154	154
MIC	N of resistant isolates	154	7	154	141	47	108	0	18	0	104	130	86	7	94
1024												130			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring - EFSA

Sampler: Official sampling

Sampling Strategy: Objective sampling

specifications Programme Code: ESBL MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	4	4	4	4	4	4	4	4	4	4
МІС	N of resistant isolates	3	4	0	0	4	0	0	0	0	0
<=0.015								4			
<=0.03										4	
<=0.064		1		3							
<=0.12							3		3		
0.12				1							
0.25							1		1		
0.5		1	1			4					
2			ı			<u>1</u> 1					
4		1			2	1					2
8		1	1		2	ı					1
16		•	1			1					1
64			1								·
			· · · · · · · · · · · · · · · · · · ·								

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring - EFSA

Sampler: Official sampling

Sampling Strategy: Objective sampling

specifications Programme Code: ESBL MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	4	4	4	4	4	4	4	4	4	4	4	4	4	4
МІС	N of resistant isolates	4	0	4	4	2	1	0	1	0	1	3	3	0	2
<=0.015							3								
<=0.03										4					
<=0.25														4	2
0.25							1								
<=0.5									3						
<=1								4							
1 <=2				1	1								1		
2					1								ı		
<=4											3				
4			2	3											
<=8						2									
8			2		2				1						
16			_		_							1			
32						2					-				2
64		4											3		
128											1				
1024												3			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from turkey - meat preparation - intended to be eaten cooked

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	1	1	0	0	1	0	0	0	0	0
<=0.015								1			
<=0.03										1	
<=0.064				1							
<=0.12							1				
0.25									1		
0.5		1									
			1								
1											
2						1					
					1	1					1

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from turkey - meat preparation - intended to be eaten cooked

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	1	1	0	0	0	0	0	0	1	1	0	0
<=0.03										1					
0.03							1								
<=0.25														1	1
<=0.5									1						
<=1								1							
1					1										
2				1											
<=4						<u> </u>					1				
<=8						1									
8			1												
64		1											1		
1024												1			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from bovine animals - meat preparation

Sampling Stage: Retail Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

Samp	ling Details: N_A										
	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	тірепет	Мегорепет	Temocillin
	Cefotaxime synergy test		Not Available						_		
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	3	3	3	3	3	3	3	3	3	3
MIC	N of resistant isolates	3	3	2	2	3	2	0	0	0	0
<=0.015								1			
<=0.03										3	
0.03								2			
<=0.064				1							
<=0.12									3		
0.25							1				
				1		1					
2				1			2				
4		3				2					1
8			1		1						2
16			2		2						
64					2						

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from bovine animals - meat preparation

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
MIC	N of resistant isolates	3	1	3	3	0	2	0	0	0	2	3	2	0	2
<=0.015							1								
<=0.03										3					
<=0.25														3	
0.25							2								
<=0.5									3						
0.5															1
<=1								3							
1					1										
<=2			2	3	2								1		
<u>4</u> <=8			2	ა		3									
8						<u> </u>					1				
32													1		2
64		3	1								1		1		
128			•								1				
1024												3			

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sam	pling Details: N_A										
	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	1	1	1	1	1	1	0	0	0	0
<=0.03										1	
0.064								1			
0.25		1							1		
8			1	1		1	1				1
64					1						

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	1	1	0	1	0	0	0	1	1	0	0	0
<=0.03										1					
<=0.25														1	
<=0.5									1						
0.5							1								1
<=1								1							
<=2													1		
4			1	1											
<=8						1									
8					1										
64		1									<u> </u>				
128											1				
1024												1			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Cattle (bovine animals) - calves (under 1 year)

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - active

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	167	167	167	167	167	167	167	167	167	167	167	167	167	167
MIC	N of resistant isolates	90	5	0	0	41	31	3	7	0	29	98	116	0	67
<=0.015							124								
<=0.03										167					
0.03							11								
0.064							1								
0.12							1								
<=0.25				167										160	36
0.25							10								
<=0.5					167				138						
0.5							5							6	57
<=1		2						164							
1							1		19					1	5
<=2			14										49		
2		36							3						2
<=4											134				
4		39	88					2					2		
<=8						119		<u>.</u>				15			
8			50				7	1	1		3				
>8			10				7					0.1			
16 32			10			7			1		1	21	2		1
>32			4			10			3 2			20	2		1 66
>32															00

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	167	167	167	167	167	167	167	167	167	167	167	167	167	167
MIC	N of resistant isolates	90	5	0	0	41	31	3	7	0	29	98	116	0	67
64		1	1			4					3	13	31		
>64		89											83		
128						9					5	5			
>128						18					21				
256												2			
1024					·							1			
>1024												90			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Cattle (bovine animals) - calves (under 1 year)

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - active

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

	AM substance	90	<u>E</u>		ime + Clavulanic acid	5	uii			Œ	Ē	nem	<u>=</u>
		Cefepime	Cefotaxim		Cefotaxim	Cefoxitin	Ceftazidim	: :		Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Positive/Pres ent	Negative/Abs ent	Not Available			ailable			Not Available	
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available F	Positive/Pres ent	Negative/Ab ent	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	0.25	8	0.5	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.064	0.5	0.25	0.12	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	64	128	128	128	2	16	16	128
	N of tested isolates	181	181	181	181	181	181	181	181	181	181	181	181
MIC	N of resistant isolates	i 171	181	16	16	22	168	15	15	4	0	0	0
<=0.015										145			
<=0.03												180	
0.03										22			
<=0.064		8		155								<u> </u>	
0.064								400	10	10	440	1	
<=0.12 0.12		2		6	1			126	13	4	143		
<=0.25				0	'		1			4			
0.25		4		2	1		ı	15	10		37		
0.5			1	_	6		12	1	1		1		
1		8	7	1	4		50		6				
2		21	7		2	12	27	1	2				10
4		76	2		3	71	32		2				101
8		45	11			76	44		3				62
									200				

	AM substance	Cefepime	Cefotaxim		Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim		Cenazionne + Ciavulanic acio	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Positive/Pre ent	s Negative/Ab ent	S Not Available			ailable			Not Available	
	Ceftazidime synergy test	Not Available	Not Available	Not Availabl	e Not Availabl	e Not Available	Not Available	Positive/Pres ent	Negative/Ab ent	S Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	0.25	8	0.5	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.064	0.5	0.25	0.12	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	64	128	128	128	2	16	16	128
	N of tested isolates	181	181	181	181	181	181	181	181	181	181	181	181
міс	N of resistant isolates	171	181	16	16	22	168	15	15	4	0	0	0
16		15	30			8	11		1				7
32		1	38			7	2						1
>32		1							·				
64			50			7	1						
>64			35										
>128							1						

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Cattle (bovine animals) - calves (under 1 year)

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - active

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	181	181	181	181	181	181	181	181	181	181	181	181	181	181
MIC	N of resistant isolates	181	29	181	166	88	108	3	31	0	80	149	159	0	105
<=0.015							63								
<=0.03										181					
0.03							9								
0.064							1								
0.12							1								
<=0.25														176	28
0.25							16								
<=0.5					15				122						
0.5				3			20							5	42
<=1							<u> </u>	177							
1			10	7	41		4		23				2.1		5
<=2			10										21		
2				5	33		2	1	5		70				1
<=4			73	1	22		2		7		78		1		1
4 >4			73	165	32		3	2					<u>'</u>		
<=8				100		85						7			
8			59		43	00	13	1	6		16				
>8			59		17		49	ı	U		10				
16			10		17	8	73		4		7	10			
32			5			10			8		2	12			
- 52			<u> </u>			10			<u> </u>			14			

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	181	181	181	181	181	181	181	181	181	181	181	181	181	181
MIC	N of resistant isolates	181	29	181	166	88	108	3	31	0	80	149	159	0	105
>32									6						104
64			6			15					1	3	41		
>64		181	18										118		
128						29					6	3			
>128						34					71				
512												1			
1024												6			
>1024												139			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Cattle (bovine animals) - meat production animals - calves (under 1 year)

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Monitoring - active

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Samp	oling Details: N_A											
	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim		Ceffazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Positive/Pres ent	Not Available	Not Available		vailable			Not Available	
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available F	Positive/Pres ent	Negative/Al ent	os Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	128	2	16	16	128
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	2	2	0	0	1	0	0	0	0	0	0
<=0.015									2			
<=0.03											2	
<=0.064				2								
<=0.12							1	1		1		
<=0.25						1						
0.25										1		
2		1			1							11
4		1			1	1						1
16			1									
64			1									

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Cattle (bovine animals) - meat production animals - calves (under 1 year)

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Monitoring - active

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	176	176	176	176	176	176	176	176	176	176	176	176	176	176
MIC	N of resistant isolates	27	2	2	1	18	10	1	7	0	9	47	34	0	21
<=0.015							159								
<=0.03										175					
0.03							7								
0.12							1			1					
<=0.25				174										176	69
0.25							3								
<=0.5					175				138						
0.5															72
<=1		13						174							
1									27						13
<=2			26										139		
2		73						1	4						1
<=4											166				
4		61	95	-				1	2				3		
>4				2											
<=8						155						18			
8		2	53		1		3		2		1				
>8						<u> </u>	3								
16						3						45			
32			2			2			3			31	1		
>32															21

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	176	176	176	176	176	176	176	176	176	176	176	176	176	176
MIC	N of resistant isolates	27	2	2	1	18	10	1	7	0	9	47	34	0	21
64											1	35	12		
>64		27											21		
128						8					1	6			
>128						8					7				
256												2			
512												1			
1024												7			
>1024									·			31			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Pigs - fattening pigs

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - active

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

ECOFF 8	32
Highest limit	32 169
N of tested	169
Solates 169	
MIC isolates 77 1 0 0 42 8 1 1 0 3 85 80 0 <=0.015	76
<=0.03	
0.03 12 0.064 1 0.12 2 <=0.25	
0.064 1 0.12 2 <=0.25	
0.12 2 <=0.25	
<=0.25 169	
0.25	37
<=0.5 0.5 138 4	50
0.5 4 4 168	50
1 30	6
<=2 20 80	
2 54	
<=4	
4 32 102 1 5	1
<=8 122 28	
8 3 43 4	
>8	
16 1 3 5 29	
32 1 1 1 16 19 2	
>32	1 74

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	169	169	169	169	169	169	169	169	169	169	169	169	169	169
MIC	N of resistant isolates	77	1	0	0	42	8	1	1	0	3	85	80	0	76
64		2				14					2	8	34		
>64		73											44		
128						6						1			
>128						6					1				
256												1			
512	·		<u> </u>			<u> </u>					<u> </u>	1			
1024												1			
>1024												81			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Pigs - fattening pigs

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - active

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

	AM substance	Cefepime	Cefotaxim		Cerotaxime + Clavulanic acid	Cefoxitin	Ceftazidim		Cenazidime + Ciavuianic acid	Ertapenem	mipenem	Meropenem	Temocillin
		Cefe	Cefo		0 0	Cefo	Ceff	č	61	Erta	Ē	Mero	Гет
	Cefotaxime synergy test				N (* /A)		Not Available		railable		_	Not Available	
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available P	ositive/Pres ent	Negative/Ab ent	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	0.25	8	0.5	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.064	0.5	0.25	0.12	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	64	128	128	128	2	16	16	128
	N of tested isolates	176	176	176	176	176	176	176	176	176	176	176	176
МІС	N of resistant isolates	165	176	16	16	17	149	16	16	0	0	0	0
<=0.015										157			
<=0.03												175	
0.03										16			
<=0.064		3		157								4	
0.064 <=0.12								112	27	3	139	1	
0.12		8		3				112	21		139		
<=0.25							6						
0.25		5					-	9	12		37		
0.5		13					21						
1		21	4		10		51						
2		43	12		3	20	28		7				22
4		59	27		1	112	31		6				96
8		18	19		2	27	30		1				50
									207				

	AM substance	Cefepime	Cefotaxim		Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	; ;	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Positive/Pre-	s Negative/Abs ent	Not Available	Not Available	Not A	vailable	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	e Not Available	Not Available	Not Available	Positive/Pres ent	Negative/Ab	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	0.25	8	0.5	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.064	0.5	0.25	0.12	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	64	128	128	128	2	16	16	128
	N of tested isolates	176	176	176	176	176	176	176	176	176	176	176	176
10	N of resistant		470	46	46	47	440	46	46	•	^	^	^
IIC	isolates	165	176	16	16	17	149	16	16	0	0	0	0
16		5	54			3	8		2				6
32		1	26			7	1						2
64			26			3							
>64			8			4							

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Pigs - fattening pigs

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - active

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	176	176	176	176	176	176	176	176	176	176	176	176	176	176
MIC	N of resistant isolates	176	19	176	159	41	56	3	10	0	29	128	85	0	112
<=0.015							115								
<=0.03										176					
0.03							4								
0.064							1								
0.12							2								
<=0.25														173	41
0.25							24								
<=0.5					17				138						
0.5							14							3	20
<=1								173							
1				3	66				25						3
<=2			15										86		
2				12	25				3						
<=4											127				
4			90	23	34			3	1				4		
>4				138											
<=8						130	_					19			
8			46		27		7		1		12		1		
>8			•		7		9				•	10			
16			6			5			1		8	13			
32			5			11			4		11	9	3		

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	176	176	176	176	176	176	176	176	176	176	176	176	176	176
MIC	N of resistant isolates	176	19	176	159	41	56	3	10	0	29	128	85	0	112
>32									3						112
64		1	5			16					2	7	29		
>64		175	9										53		
128						7					5	1			
>128						7					21				
512		•										1			
>1024												126			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - meat preparation

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	e Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	e Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	4	4	4	4	4	4	4	4	4	4
МІС	N of resistant isolates	3	4	1	1	4	1	0	0	0	0
<=0.015								3			
<=0.03										4	
0.03								1			
<=0.064				3					<u> </u>		
<=0.12									1		
0.12		1					2		3		
0.25							1		<u> </u>		
1						2					
2			1	1		1					
4		3			1	1	1				2
8					2						2
16			2								
32			1								
						· · · · · · · · · · · · · · · · · · ·			244		

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	4	4	4	4	4	4	4	4	4	4
IIC	N of resistant isolates	3	4	1	1	4	1	0	0	0	0
64	-				1						

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - meat preparation

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

ECOFF 8 16 0.25 0.5 16 0.064 2 2 0.125 16 64 8 1 2 Lowest limit 1 2 0.25 0.5 8 0.015 1 0.5 0.03 4 8 2 0.25 0.25 0.25 18 Highest limit 64 64 4 8 128 8 16 32 16 128 1024 64 8 32 Nof tested Isolates 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4		AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
Highest limit		ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
Nof fested isolates		Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
Solates		Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
MIC isolates 4 0 4 4 1 3 0 0 0 0 3 4 4 0 0 0 <			4	4	4	4	4	4	4	4	4	4	4	4	4	4
\$\begin{array}{c} \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \	MIC		4	0	4	4	1	3	0	0	0	3	4	4	0	0
<=0.25	<=0.015							1								
0.25 <=0.5	<=0.03										4					
<=0.5 0.5 <=1 4 1 2 <=4 1 3 <=8 3 1 1 8 3 1 1 64 4 4 128 1 3															1	1
0.5 <=1								2								
<=1										2						
1 1 2 2 1 2 <=4															3	3
2 1 2 <=4	<=1								4							
<=4	1					<u> </u>				2						
4 1 3 <=8					1	2						1				
<=8				1	2							1				
8 3 1 1 64 4 4 128 1 3				1			3									
64 4 128 1 3				3		1	0	1								
128 1 3			4											4		
							1					3				
	1024												4			

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

Sampli	ng Details: N_A										
	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	1	1	0	0	0	0	0	0	0	0
<=0.015								1			
<=0.03										1	
<=0.064				1							
<=0.12							1		1		
0.5						1					
2		1									1
4					1						
32			1								

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	1	0	0	1	0	0	0	1	0	0	0	1
<=0.03										1					
<=0.25														1	
0.25							1								
<=0.5					1				1						
<=1								1							
<=2			1										1		
4				1		4									
<=8						1						1			
32 64		1									1				1
04		- 1													

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	13	13	13	13	13	13	13	13	13	13
МІС	N of resistant isolates	13	13	5	7	13	5	0	0	0	0
<=0.015								6			
<=0.03										13	
0.03								7			
<=0.064				5							
<=0.12				2			5		6		
0.12		6		<u>2</u> 1			3		6		
0.25		- 6		ı			<u> </u>		<u>6</u>		
1						3					
2		1	1	1		2					
4		2		4	5	<u>-</u>	3				4
8		1	6		1	6	2				8
16		3	1		2	1					1
32			2		2	1					
									24.6		

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	13	13	13	13	13	13	13	13	13	13
MIC	N of resistant isolates	13	13	5	7	13	5	0	0	0	0
64			3		3						

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	13	13	13	13	13	13	13	13	13	13	13	13	13	13
МІС	N of resistant isolates	13	0	13	13	3	10	0	1	0	10	13	9	0	9
<=0.015							2								
<=0.03										13					
0.03							1								
<=0.25														9	1
<=0.5									8						
0.5							4	10						4	3
<=1					3			13	4						
<=2					<u> </u>				4				4		
2				1	2								-		
<=4				<u> </u>							3				
4			6	12											
<=8						10									
8			7		8		6								
16									1						
32						1									9
64		13											9		
128						2					10				
1024												13			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	e Not Available I	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	e Not Available I	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	4	4	4	4	4	4	4	4	4	4
МІС	N of resistant isolates	: 4	4	1	1	4	1	0	0	0	0
<=0.015								2			
<=0.03										4	
0.03								1			
<=0.064				3							
0.064								1			
<=0.12							3		1		
0.25		2							3		
0.5		1									
1		1	•								
2			2	4	4	4					4
4			1	1	1	1	4				1
8			1		2	2 1	1				3
16 64					1	ı					
204 Relaium - 201	6								310		

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	4	4	4	4	4	4	4	4	4	4	4	4	4	4
MIC	N of resistant isolates	4	0	4	4	2	2	0	1	0	2	4	3	0	2
<=0.015							1								
<=0.03										4					
0.03							1								
<=0.25														1	2
<=0.5									3						
0.5														3	
<=1								4							
<=2													1		
2				1											
<=4											2				
4			2	3	1										
<=8					<u> </u>	1									
8			2		3		2								
16						1			1						_
32													•		2
64		4				1							3		
128						1					2				
1024												4			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - active

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Positive/Pres ent	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Positive/Pres ent	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	128
	N of tested isolates	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	1	0	0	1	0	0	0	0	0
<=0.015								1			
<=0.03										1	
<=0.064				1							
<=0.12							1		1		
2						1					
4		1			1						
8											1
16			1								

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - active

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	151	151	151	151	151	151	151	151	151	151	151	151	151	151
MIC	N of resistant isolates	124	1	1	1	34	85	0	6	0	72	100	75	0	85
<=0.015							59								
<=0.03										151					
0.03							7								
0.12							13								
<=0.25				150										144	27
0.25							32								
<=0.5					150				118						
0.5							18							7	30
<=1		1						151							
1					1		7		24						7
<=2			10										75		
2		11					1		3						2
<=4											66				
4		15	66				2								
>4				1											
<=8						113						11			
8			64				7		1		5		1		
>8							5								
16 32			10			4			3		8	16			
32			1			10			2		1	17	1		
>32															85

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	151	151	151	151	151	151	151	151	151	151	151	151	151	151
MIC	N of resistant isolates	124	1	1	1	34	85	0	6	0	72	100	75	0	85
64		1				6					18	7	24		
>64		123											50		
128						4					20	1			
>128						14					33				
256												1			
512												1			
1024												2			
>1024												95			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - active

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance				e + Clavulanic acid				e + Clavulanic acid			5	
		Cefepime	Cefotaxim		Cefotaxime	Cefoxitin	Ceftazidim	: :	Certazionne	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Positive/Pres ent	Negative/Abs ent	Not Available			ailable			Not Available	
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available F	ositive/Pres ent	Negative/Al ent	os Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	0.25	8	0.5	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.064	0.5	0.25	0.12	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	64	128	128	128	2	16	16	128
	N of tested isolates	190	190	190	190	190	190	190	190	190	190	190	190
MIC	N of resistant isolates	1 183	190	19	19	27	173	19	19	0	0	0	0
<=0.015										166			
<=0.03												190	
0.03										15			
<=0.064				166									
0.064								4.40	40	9	457		
<=0.12 0.12		7		5				142	18		157		
<=0.25		,		j			3						
0.25		37					3	8	3		30		
0.5		30	2				14				3		
1		26	15				33						
2		38	23		2	14	25		1				27
4		40	34		12	87	16		13				85
8		12	45		5	62	56		5				74
								•	22.4				

	AM substance	Cefepime	Cefotaxim		Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	;	Ceffazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Positive/Pres	s Negative/Abs ent	Not Available	Not Available		vailable			Not Available	
	Ceftazidime synergy test	Not Available	Not Available	Not Available	e Not Available	Not Available	Not Available	Positive/Pres ent	Negative/Ab ent	S Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	0.25	8	0.5	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.064	0.5	0.25	0.12	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	64	128	128	128	2	16	16	128
	N of tested isolates	190	190	190	190	190	190	190	190	190	190	190	190
міс	N of resistant isolates	183	190	19	19	27	173	19	19	0	0	0	0
16			26			8	29						3
32			23			3	13						1
64			15			11	1						
>64			7			5							

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - active

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	190	190	190	190	190	190	190	190	190	190	190	190	190	190
MIC	N of resistant isolates	190	13	190	168	65	130	0	13	0	116	161	117	0	109
<=0.015							49								
<=0.03										190					
0.03							9								
0.064							2								
0.12							9								
<=0.25														185	38
0.25							45								
<=0.5					22				147						
0.5				1			15	100						5	32
<=1				4.4	34		40	190	26						40
1 <=2			30	14	34		12		26				67		10
2			30	29	16		3		4				67		1
<=4					10				4		60				ı
4			104	32	19		5						6		
>4			104	114	10										
<=8				117		117						6			
8			35		51		26				10				
>8					48		15								
16			8		-	8			6		4	10			
32			8			21			3		5	8	5		

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	190	190	190	190	190	190	190	190	190	190	190	190	190	190
МІС	N of resistant isolates	190	13	190	168	65	130	0	13	0	116	161	117	0	109
>32									4						109
64			4			20					14	5	49		
>64		190	1										63		
128						8					23	1			
>128						16					74				
256												1			
512												2			
1024	·		·						·			5			
>1024												152			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from other animal species or not specified

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampl	ing Details: N_A										
	AM substance			Cefotaxime + Clavulanic acid			Ceftazidime + Clavulanic acid			F	
		Cefepime	Cefotaxim	Cefotaximo	Cefoxitin	Ceftazidim	Ceftazidim	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	1	0	0	0	0	0	0	0	0
<=0.015	isolates	<u> </u>	<u> </u>		<u> </u>		<u> </u>	1	<u> </u>	<u> </u>	
<=0.015								ı		1	
<=0.064				1						'	
<=0.12				•			1				
<=0.25						1					
0.25									1		
0.5		1									
2			1								
4					1						1

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from other animal species or not specified

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	1	1	0	0	0	0	0	0	0	0	0	0
<=0.015							1								
<=0.03										1					
<=0.25														1	1
<=0.5									1						
<=1								1							
<=2													1		
2				1											
<=4											1				
<=8			1			1									
8			1		1										
16		1										1			
64		1													

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Crustaceans - unspecified

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampli	ing Details: N_A										
	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	2	2	2	2	2	2	2	2	2	2
МІС	N of resistant isolates	2	2	0	0	2	0	0	0	0	0
<=0.015								2			
<=0.03										2	
<=0.064				2							
<=0.12							2		1		
0.25									1		
4		2			1	2					1
8					1						1
16			1								
64			1								

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Crustaceans - unspecified

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	2	0	2	2	0	2	0	0	0	1	1	1	0	0
<=0.03										2					
<=0.25														1	1
0.25							11								
<=0.5									1						
0.5														1	
<=1								2							
1									1				1		1
<=2 4				2	1								1		
<=8						2									
8			2		1		1				1				
16												1			
64		2											1		
128											1				
1024												1			

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Bangladesh

Sampling Details: N_A

Samp	ing Details: N_/										
	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	mipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	1	1	1	1	1	1	0	0	0	0
<=0.03										1	
0.03								1			
<=0.12									1		
4		1		1							
8			1								
16											1
32					1	1	1				

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Bangladesh

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	1	1	1	1	0	1	0	1	1	1	0	1
<=0.03										1					
<=0.25														1	
<=1								1							
4				1											
8			1		1		1								
16									1						
32		4											4		1
64		1				4					4		1		
128						11					<u> </u>	1			
1024												1			

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

Certazidime synergy test Not Available		AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Ітірепет	Meropenem	Temocillin
Synergy test Syne		Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available I	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
Lowest limit 0.064 0.25 0.064 0.5 0.25 0.12 0.015 0.12 0.03 0.5 0.5 0.12 0.015 0.12 0.03 0.5			Not Available	Not Available	Not Available	Not Available I	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
Highest limit 32 64 64 64 128 128 2 16 16 64 N of tested isolates 25 25 25 25 25 25 25 2		ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
N of tested		Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
Solution Solution		Highest limit	32	64	64	64	128	128	2	16	16	64
MIC isolates 23 25 6 5 25 7 0 0 0 0 <=0.015 17 <=0.03 6 <=0.04 17 2 2 2 <=0.12 12 12 12 12 0.12 2 2 2 2 2 0.25 1 6 12 1 1 1 2 2 6 1 2 2 6 1 2 2 6 1 4 6 3 1 4 1 4 5 6 1 5 4 5 6 1 7 6 1 7 6 1 7 6 1 7 6 1 7 6 1 7 6 1 7 7 9 9 1 1 7 9 9 1 1 7			25	25	25	25	25	25	25	25	25	25
\$\sigma \cdot	MIC			25	6	5	25	7	0	0	0	0
0.03 6 <=0.064									17			
<=0.064 17 0.064 2 <=0.12											23	
0.064 2 2 <=0.12 12 12 0.12 2 2 0.25 1 6 12 0.5 1 1 1 2 2 6 1 2 3 3 1 4 1 4 6 3 1 10 5 4 5 8 8 2 9 9 1 17									6			
<=0.12 12 12 0.12 2 2 0.25 1 6 12 0.5 1 1 1 2 2 6 1 2 3 3 1 4 1 4 6 3 1 10 5 4 5 8 8 2 9 9 1 17					17							
0.12 2 0.25 1 0.5 1 1 2 2 6 1 2 3 3 3 1 4 4 6 8 2 9 9 1 2 5 4 6 7 8 1 8 1 8 1 9 1 17								12		10		
0.25 1 0.5 1 1 2 2 6 1 2 3 3 3 4 6 8 8 2 9 9 9 1 1 2 1 4 6 3 1 4 1 5 1 8 2 9 9 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 2 3 4 4 5 4 6 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 <td< td=""><td></td><td></td><td>2</td><td></td><td>2</td><td></td><td></td><td>12</td><td></td><td>IZ</td><td></td><td></td></td<>			2		2			12		IZ		
0.5 1 1 2 2 6 1 2 3 3 1 4 1 4 6 3 1 10 5 4 5 8 8 2 9 9 1 17								6		12		
2 3 3 3 1 4 1 4 6 3 1 10 5 4 5 8 8 2 9 9 1 17								-				
4 6 3 1 10 5 4 5 8 8 2 9 9 1 17	1		2		2		6	1				
8 8 2 9 9 1 17	2		3	3	3	1	4	1				
	4		6	3	11	10	5	4				5
16 2 7 1						9		1				
	16		2	7			1					3

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	25	25	25	25	25	25	25	25	25	25
NIC	N of resistant isolates	23	25	6	5	25	7	0	0	0	0
32			3		1						
64			7		4						

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	25	25	25	25	25	25	25	25	25	25	25	25	25	25
MIC	N of resistant isolates	25	4	25	23	6	12	0	4	0	9	21	14	1	15
<=0.015							7								
<=0.03										25					
0.03							6								
<=0.25														24	2
0.25							8								
<=0.5					2				14						
0.5							1								5
<=1								25							
1					6		1		5						3
<=2			2										10		
2				1					2						
<=4											13				
4			7	24	7								1	1	
<=8						19						3			
8			11		10		2		1		1				
16			1						1		2	1			
32			1			2			2				1		15
64		25	3			1					1		13		
128						3					8				
256												1			
1024												20			

Sampling Stage: Retail Sampling Type: food sample - meat

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Sampling Context: Monitoring

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

Ceftazidime synergy test Ceftazidime synergy test		AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	lmipenem	Meropenem	Temocillin
Ceftazidime synergy test		Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
Lowest limit 0.064 0.25 0.064 0.5 0.25 0.12 0.015 0.12 0.03 0.5 Highest limit 32 64 64 64 128 128 2 16 16 64 N of tested		Ceftazidime	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
Highest limit		ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
Nof tested		Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
Nof tested		Highest limit	32	64	64	64	128	128	2	16	16	64
MIC N of resistant isolates 3 3 1 1 3 1 0 0 0 0 <=0.015												
MIC isolates 3 1 1 3 1 0 0 0 0 <=0.015		isolates	3	3	3	3	3	3	3	3	3	3
<=0.03	МІС			3	1	1	3	1	0	0	0	0
0.03 1 <=0.064	<=0.015								2			
<=0.064											3	
<=0.12									1			
0.25 2 1 1 2 1 4 1 1 1 1 8 1 1 3 16 1 1 32 1					2							
1 1 2 1 4 1 1 1 1 8 1 1 1 3 16 1 1 1 32 1 1 1								2				
2 1 4 1 1 1 1 8 1 1 3 16 1 1 32 1										2		
4 1 1 1 1 8 1 1 3 16 1 1 32 1			1		1							
8 1 1 3 16 1 32 1												
16 1 32 1			· ·	1			1	1				
32 1			1			1						3
							11					
64 1 1												
	64			1		1						

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
MIC	N of resistant isolates	3	0	3	3	0	1	0	0	0	1	2	2	0	3
<=0.015							1								
<=0.03										3					
0.064							1								
<=0.25														3	
0.25							1								
<=0.5									3						
<=1								3							
1					1								<u> </u>		
<=2			•										1		
4			2	3	1										1
<=8						3						1			
8			1		1						1				
16 32											1				2
64 128		3									4		2		
1024											1	2			
1024															

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus)

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	2	2	2	2	2	2	2	2	2	2
МІС	N of resistant isolates	2	2	1	2	2	1	0	0	0	0
<=0.03										2	
0.03								1			
0.064								1			
<=0.12									1		
0.25		1		1					1		
0.5							1				
2				11			11				
4			1			4					<u>1</u> 1
8 16		1			1	1					<u> </u>
32		I			1						
64			1		ı						
J 1			· ·								

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus)

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	2	0	2	2	1	2	0	0	0	2	2	1	0	2
<=0.03										2					
<=0.25														2	
<=0.5									2						
<=1								2							
<=2													1		
4				2	1		1								
<=8						1									
8			2		1		1								
32						1									2
64		2											1		
128											2				
1024												2			

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Ітірепет	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	13	13	13	13	13	13	13	13	13	13
МІС	N of resistant isolates	10	13	6	6	11	6	0	0	0	0
<=0.015								10			
<=0.03										12	
0.03								1			
<=0.064				6							
0.064							-	2	-	1	
<=0.12 0.12		2		1			7		2		
0.12		3		11					10		
0.25		3		1		2			10		
1		1	1	3		1	1		'		
2			3			2	3				
4		5	2		4	4	1				7
8		1		2	3		1				6
16			5		1	3					
	_								244		

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	lmipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	13	13	13	13	13	13	13	13	13	13
1IC	N of resistant isolates	10	13	6	6	11	6	0	0	0	0
32			2		3						
64					2	1					

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

Lowest limit		AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
Highest limit		ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
Not fested					0.25			0.015							0.25	0.25
No fresistant Solates 13			64	64	4	8	128	8	16	32	16	128	1024	64	8	32
MIC isolates 13 2 13 12 7 10 0 5 0 10 13 11 1 9 <=0.015		N of tested isolates	13	13	13	13	13	13	13	13	13	13	13	13	13	13
\$<-0.03	МІС		13	2	13	12	7	10	0	5	0	10	13	11	1	9
\$ \$ <td><=0.015</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td>3</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td>	<=0.015							3								
0.25 1 <=0.5	<=0.03										13					
\$ 1 4 0.5 3 2 \$ 13 2 4 3 \$ 2 4 3 \$ 2 2 1 \$ 2 3 2 1 \$ 4 5 9 4 \$ 8 5 4 6 \$ 1 1 1 \$ 1 4 9 \$ 1 4 9 \$ 1 1 1 \$ 1 4 1 \$ 1 4 1 \$ 1 4 1 \$ 1 4 1 \$ 1 4 1 \$ 1 4 1 \$ 1 4 1 \$ 1 1 1 \$ 1 4 1 \$ 1 1 1 \$ 1 1 1 \$ 1 1 1 \$ 1 1 1 \$ 1 1 1 \$ 1 1 1	<=0.25														6	2
0.5 3 2 <=1	0.25							1								
<=1						1				4						
1 1 2 4 3 <=2								3							3	2
<=2 3 2 <=4 3 4 5 9 4 <=8 6 8 5 4 6 16 1 1 1 32 1 4 9 64 13 2 2 11 128 4 10									13							
2 3 2 <=4	-				1	2				4					3	
<=4														2		
4 5 9 4 <=8					3	2									1	
<=8 5 4 6 16 1 1 32 1 4 9 64 13 2 2 11 128 4 10				-	0	4						3				
8 5 4 6 16 1 1 32 1 4 9 64 13 2 2 11 128 4 10				5	9	4	6									
16 1 32 1 64 13 128 10				5		4	0	6								
32 1 4 9 64 13 2 2 11 128 4 10						7		U		1						
64 13 2 2 11 128 4 10				<u> </u>			1									9
128 4 10			13	2			2			•				11		
												10				
1024	1024												13			

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Vietnam

Samp	oling Details: N_A										
	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Availa
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Availa
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	1	1	1	1	1	1	1	1	1	1
С	N of resistant isolates	0	1	1	1	1	1	0	0	0	0
=0.015								1			
=0.03										1	
.12		1									
.25									1		
				1							
			1								
						1	1				
											1
2					1						

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Vietnam

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	1	1	0	0	0	0	0	0	1	1	0	0
<=0.015							1								
<=0.03										1					
<=0.5									1						
0.5														1	
<=1								1							
1															1
2				1											
<=4						<u> </u>					1				
<=8						1									
8			1		1								1		
64		1										4	1		
1024												1			

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

AM substance	Sampl	ing Details: N_A										
Synergy test Ceftazidime Not Available			Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	+	Ertapenem	Imipenem	Meropenem	Temocillin
Synergy test Not Available Not Not Available Not Available Not Available Not Available Not Avail			Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
Lowest limit 0.064 0.25 0.064 0.5 0.25 0.12 0.015 0.12 0.03 0.5 Highest limit 32 64 64 64 128 128 2 16 16 64 Nof tested isolates 1 1 1 1 1 1 1 1 1		Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
Highest limit 32 64 64 64 128 128 2 16 16 64 N of tested		ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
N of tested		Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
Solates 1		Highest limit	32	64	64	64	128	128	2	16	16	64
MIC isolates 1 1 0			1	1	1	1	1	1	1	1	1	1
<=0.03	МІС			1	0	0	1	0	0	0	0	0
<=0.064	<=0.015								1			
<=0.12	<=0.03										1	
0.25 1 2 1 4 1 1 8 1	<=0.064				1							
2 1 4 1 1 1 8 1	<=0.12							1				
4 1 1 8 1	0.25									1		
8 1	2						1					
						1						1
32 1			1									
	32			1								

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	1	1	1	1	0	1	0	1	1	1	0	1
<=0.03										1					
<=0.25														1	
<=1								1							
2					1										
4				11											
8			1				1								
32		1							1				1		1
64 128		I				1					1		l l		
1024						ı					ı	1			
1024															

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Fish - raw - chilled

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampl	ing Details: N_A										
	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	1	1	1	1	1	1	1	1	1	1
МІС	N of resistant isolates	1	1	0	1	0	0	0	0	0	0
<=0.03										1	
0.03								1			
<=0.25						1					
0.25				1							
0.5							1		1		
1											1
4		1									
32			1								
64					1						

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Fish - raw - chilled

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	1	1	0	1	1	0	0	0	1	1	0	1
<=0.03										1					
<=0.5									1						
0.5							1							1	
1					1										
<=4											1				
4				1											
<=8						1									
16			1					1							
32															1
64		1											1		
1024												1			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - frozen

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

mpling Dotails: N

Samp	oling Details: N_A										
	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	mipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	2	2	2	2	2	2	2	2	2	2
	10014100	2	2								
MIC	N of resistant isolates		2	0	0	2	0	0	0	0	0
<=0.015	N of resistant	:				2	0	0	0		0
<=0.015 <=0.03	N of resistant	:				2	0		0	2	0
<=0.015 <=0.03 <=0.12	N of resistant	:		0		2	0		1		0
<=0.015 <=0.03 <=0.12	N of resistant	:				2					0
<=0.015 <=0.03 <=0.12 0.12 0.25	N of resistant	:		0			2				0
<=0.015 <=0.03 <=0.12 0.12 0.25 2	N of resistant	:		0	0	1			1		0
<=0.015 <=0.03 <=0.12 0.12 0.25 2	N of resistant	1	2	0	1				1		0
<=0.015 <=0.03 <=0.12 0.12 0.25 2 4	N of resistant	2	1	0	0				1		1
<=0.015 <=0.03 <=0.12 0.12 0.25 2	N of resistant	1	2	0	1				1		

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Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - frozen

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Unknown

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	2	0	2	2	1	1	0	0	0	0	2	2	0	2
<=0.03										2					
0.03							1								
<=0.5									1						
0.5							1							2	
<=1								2							
1					1				1						
<=4											1				
4				2											
<=8						1									
8			2		1						1				
32															2
64		2											2		
128						1									
1024												2			

Sampling Stage: Retail Sampling Type: food sample - meat Sampling Context: Monitoring

Sampler: Official sampling Sampling Sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	13	13	13	13	13	13	13	13	13	13
МІС	N of resistant isolates	13	13	3	2	13	3	1	0	0	0
<=0.015								9			
<=0.03										13	
0.03				_				3			
<=0.064				7							
<=0.12				2			6	4	2		
0.12		1		3			4	1	11		
0.25		2		1			4		- 11		
1				2		3	1				
2		1	1	_		7	2				
4		3	2		6		_				6
8		4			5	1					4
16		1	5		2	2					3
32		1	3								
1 : 204									252		

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceffazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.064	0.25	0.064	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	64
	N of tested isolates	13	13	13	13	13	13	13	13	13	13
МІС	N of resistant isolates	1 13	13	3	2	13	3	1	0	0	0
64			2								

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

Sampling Details: N_A

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	13	13	13	13	13	13	13	13	13	13	13	13	13	13
MIC	N of resistant isolates	13	2	13	13	3	4	0	0	0	4	13	10	0	9
<=0.015							6								
<=0.03										12					
0.03							3								
0.064										1					
0.12							1								
<=0.25														12	3
0.25							1								
<=0.5									11						
0.5														1	
<=1								13							
1				1	4				2				•		1
<=2				1	4								3		
2 <=4				1	4						8				
4			7	11	2		1				0				
<=8				,,		10	·								
8			4		3	10	1								
16			7		J		<u>'</u>				1		1		
32			1			1					1		•		9
64		13	1								2		9		
128						2					1	1			

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	13	13	13	13	13	13	13	13	13	13	13	13	13	13
MIC	N of resistant isolates	13	2	13	13	3	4	0	0	0	4	13	10	0	9
1024												12			

OTHER ANTIMICROBIAL RESISTANCE TABLES

Table Antimicrobial susceptibility testing of Methicillin resistant Staphylococcus aureus (MRSA) in Pigs - fattening pigs

Sampling Stage: Farm Sampler: Official sampling

Sampling Type: animal sample - nasal swab
Sampling Context: Monitoring - active
Sampling Strategy: Objective sampling
Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country Of Origin:Belgium

Sampling Details:N_A

M Substance	Cefox	citin	Chloram	phenicol	Ciprof	oxacin	Clind	amycin	Erythromycin (E	rythromycin A)	Fusid	ic acid	Gent	amicin	Kana	mycin	Line	ezolid	Mup	irocin	Pen	icillin	Quinupristin	/Dalfopristin	Rifan	picin	Strep	ptomyci
erformed CC IRSA haracterisatio	* *	ž	\$ e	ž	, se	ž	, se	ž	, se	ž	* *	2	Xes.	2	\$ \$	2	×e×	2	Xes.	2	% *	2	s ÷	2	×es.	2	Yes	
erformed LST MRSA saracterisatio	ş	ů	Š	8	8	Š	Š	ů	Š	ş	ů	N _o	Š	8	Š	Š	N _o	Š	Š	8	8	8	Š	ž	ž	8	ş	
COFF	4	4	16	16	1	1	0.25	0.25	1	1	0.5	0.5	2	2	8	8	4	4	1	1	0.12	0.12	1	1	0.03	0.03	16	
owest limit	0.5	0.5	4	4	0.25	0.25	0.12	0.12	0.25	0.25	0.5	0.5	1	1	4	4	1	1	0.5	0.5	0.12	0.12	0.5	0.5	0.016	0.016	4	_
hest limit	16	16	64	64	8	8	4	4	8	8	4	4	16	16	64	64	8	8	256	256	2	2	4	4	0.5	0.5	32	
																									76	24		
							31	10																				_
																									1			
					24	4	- 4		14	1																		_
							4				71	22							75	22			56	15			$\overline{}$	_
					8	4	2		30	9		20							75	20	1		50	10				_
													57	14			28	8										
					12		1				3								1				10	5				_
						1	1				2		2				49	16	1	1	3		6	2				
																					73	24						
			3	1											59	15											26	
					2	2					1	11	5	3									11	2				_
	23			04	04		38	14							-								4				05	_
	23	-	99	21	10	5			33	14			3			3												_
	44	17	12	2	10				33	14			3	2	1												- 6	_
	10	3											7	4													_	
																											3	_
																											7	
			3												1													

Belgium - 2016 356 Table Antimicrobial susceptibility testing of Methicillin resistant Staphylococcus aureus (MRSA) in Pigs - fattening pigs - CONTINUED

	Sampling Stage:	Farm		Sa	ampling Type: ani	mal sample - nas	al swab	Sampling Context: Monitoring - active							
	Sampler: Official	sampling		Sa	ampling Strategy:	Objective sampli	ng	Program	me Code: OTHE	R AMR MON					
	Analytical Method	d: Dilution -	sensititre												
	Country of Origin	: Belgium													
	AM substance		S uf famethox azole		Tetracycii ne		uinue:	Tries office and		Vancomy cin					
	Performed CC MRSA characterisatio n	Yes	ž	, se	ž	, se	2	× ×	ž	× × ×	2				
	Performed MLST MRSA characterisatio n	Š.	ž	ĝ	ž	ž	ĝ	ž	ž	ž	ž				
	ECOFF	128	128	1	1	2	2	2	2	2	2				
	Lowest limit	64	64	0.5	0.5	0.5	0.5	2	2	1	1				
	Highest limit	512	512	16	16	4	4	32	32	16	16				
<=0.5				1		55	18								
<=1						2	1			77	24				
<=2								3							
>4						20	5								
>16				76	24										
>32								74	24						
<=64		67	23												
128		4													
256 >512		6	1												
- 312															

Table Antimicrobial susceptibility testing of Methicillin resistant Staphylococcus aureus (MRSA) in Pigs - breeding animals - unspecified - sows and gilts

Sampling Stage: Farm Sampler: Official sampling Sampling Type: animal sample - nasal swab Sampling Strategy: Objective sampling Sampling Context: Monitoring - active Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country Of Origin:Belgium

Sampling Details:N_A

AM Substance		Cefoxiti	n	Chloram	phenicol	Ciprof	floxacin	Clind	amycin	Erythromycin (I	Erythromycin A)	Fusid	fic acid	Gent	amicin	Kana	mycin	Line	zolid	Mupir	ocin	Pen	nicillin	Quinupristin	/Dalfopristin	Rifan	npicin	Strepton	mycin
Performed CC MRSA characterisation			ê	Yes	ž	Yes	ĝ	Yes	ž	Yes	ž	Yes	ĝ	Yes	ž	Yes	Ŷ	Yes	Ŷ	Yes	ž	Yes	ž	Yes	Ŷ	Yes	Ŷ	Yes	ŝ
Performed MLST MRSA characterisation	Š		ê	ž	ž	ĝ	ž	ž	ê	ž	ž	ž	ž	ž	ž	ž	ŝ	ŝ	ŝ	ž	ž	ž	ž	ŝ	ŝ	ž	ŝ	ž	ž
ECOFF	4		4	16	16	1	1	0.25	0.25	1	1	0.5	0.5	2	2	8	8	4	4	1	1	0.12	0.12	1	1	0.03	0.03	16	16
Lowest limit	0.5	5	0.5	4	4	0.25	0.25	0.12	0.12	0.25	0.25	0.5	0.5	1	1	4	4	1	1	0.5	0.5	0.12	0.12	0.5	0.5	0.016	0.016	4	4
MIC Highest limit	16		16	64	64	8	8	4	4	8	8	4	4	16	16	64	64	8	8	256	256	2	2	4	4	0.5	0.5	32	32
<=0.016																										61	10		
0.064																										2			
<=0.12 <=0.25								35	4																				
						19				16																			
0.25 <=0.5								5																			1		
<=0.5						- 5				29	5	60	10							62	10			51	8				
0.5						ь	1	1		29	ь			45	10			20	2										
1						6	- 1				1	- 1		40	10			20	2	1	1			4					
2							1				<u> </u>	2		1				A1	8	<u> </u>		4	- 1		- 1			$\overline{}$	$\overline{}$
>2							· ·															59	10						
<=4				3												45	9											20	1
4						4							1	3					1					2	2				
>4								22	7															1					
8	20		5	55	5	20	5								1	1	1	2										26	5
>8						9	3			18	5																		
16	36		4	2	4									7		1												7	3
>16	7		2											7															
32																												6	
>32																												4	2
64				1	2											4	1												
>64				2												12													

Table Antimicrobial susceptibility testing of Methicillin resistant Staphylococcus aureus (MRSA) in Pigs - breeding animals - unspecified - sows and gilts - CONTINUED

	Sampling Stage: I	Farm		Sa	mpling Type: ani	mal sample - nas	al swab	Samplin	Sampling Context: Monitoring - active					
	Sampler: Official	sampling		Sa	mpling Strategy:	Objective sampli	ng	Program	me Code: OTHER	R AMR MON				
	Analytical Method	: Dilution -	sensititre											
	Country of Origin	: Belgium												
	AM substance		S uf famethox azole	i	Tetracycli ne			Tries otherwise		Vancomy cin				
	Performed CC MRSA characterisatio	Yes	ž	Yes	2	Yes	2	× × ×	ž	Yes	2			
	Performed MLST MRSA characterisatio n	ů.	ž	ž	ž	ž	ž	ž	ž	ž	2			
	ECOFF	128	128	1	1	2	2	2	2	2	2			
	Lowest limit	64	64	0.5	0.5	0.5	0.5	2	2	1	1			
	Highest limit	512	512	16	16	4	4	32	32	16	16			
<=0.5				2		44	7							
<=1										63	11			
1 <=2						8	1	3						
4						1		3						
>4						10	3							
>16				61	11		-							
>32								60	11					
<=64		51	9											
128		2												
256		1												
>512	>512 9 2													

Specific monitoring of ESBL-/AmpC-/carbapenemase-producing bacteria and specific monitoring of carbapenemase-producing bacteria, in the absence of isolate detected

Programme Code	Matrix Detailed	Zoonotic Agent Detailed	Sampling Strategy	Sampling Stage	Sampling Details	Sampling Context	Sampler	Sample Type	Sampling Unit Type	Sample Origin	Comment	Units Tested	Units Positive
CARBA MON	Gallus gallus (fowl) - broilers	Escherichia coli, non- pathogenic, unspecified	Objective sampling	Slaughte rhouse	N_A	Monitorin g - active	Official samplin g	animal sample - caecum	slaughter animal batch	Belgium	N_A	301	0
	Meat from broilers (Gallus gallus) - fresh	Escherichia coli, non- pathogenic, unspecified	Objective sampling	Retail	N_A	Monitorin g - EFSA specificat ions	Official samplin g	food sample	single (food/feed)	Unknown	N_A	234	0



Latest Transmission set

Last submitted

Table Name	dataset transmission date
Antimicrobial Resistance	19-Jan-2018
Animal Population	12-May-2017
Disease Status	12-May-2017
Food Borne Outbreaks	21-Apr-2017
Prevalence	19-Jan-2018
Text Forms	07-Jul-2017