European Food Safety Authority

ZOONOSES MONITORING

Greece

TRENDS AND SOURCES OF ZOONOSES AND ZOONOTIC AGENTS IN FOODSTUFFS, ANIMALS AND FEEDINGSTUFFS

including information on foodborne outbreaks, antimicrobial resistance in zoonotic and indicator bacteria and some pathogenic microbiological agents

IN 2015

PREFACE

This report is submitted to the European Commission in accordance with Article 9 of Council Directive 2003/99/ EC*. The information has also been forwarded to the European Food Safety Authority (EFSA).

The report contains information on trends and sources of zoonoses and zoonotic agents in Greece during the year 2015.

The information covers the occurrence of these diseases and agents in animals, foodstuffs and in some cases also in feedingstuffs. In addition the report includes data on antimicrobial resistance in some zoonotic agents and indicator bacteria as well as information on epidemiological investigations of foodborne outbreaks. Complementary data on susceptible animal populations in the country is also given. The information given covers both zoonoses that are important for the public health in the whole European Union as well as zoonoses, which are relevant on the basis of the national epidemiological situation.
The report describes the monitoring systems in place and the prevention and control strategies applied in the country. For some zoonoses this monitoring is based on legal requirements laid down by the European Union legislation, while for the other zoonoses national approaches are applied.

The report presents the results of the examinations carried out in the reporting year. A national evaluation of the epidemiological situation, with special reference to trends and sources of zoonotic infections, is given. Whenever possible, the relevance of findings in foodstuffs and animals to zoonoses cases in humans is evaluated.
The information covered by this report is used in the annual European Union Summary Reports on zoonoses and antimicrobial resistance that are published each year by EFSA.

---

* Directive 2003/ 99/ EC of the European Parliament and of the Council of 12 December 2003 on the monitoring of zoonoses and zoonotic agents, amending Decision 90/ 424/ EEC and repealing Council Directive 92/ 117/ EEC, OJ L 325, 17.11.2003, p. 31

List of Contents

1 ANIMAL POPULATIONS

The relevance of the findings on zoonoses and zoonotic agents has to be related to the size and nature of the animal population in the country

1.1 Populations

1.1.1 Information on susceptible animal population

Sources of information

SUSCEPTIBLE ANIMAL POPULATION: (please advise the relevant electronic summary tables on EFSA Web based zoonoses monitoring system for 2014 Data Collection.)Source of information: Internal Data Base computerized system of Hellenic Ministry of Agriculture (update 2011). These statistics may vary from other national or E.U. sources of animal population records.

2 DISEASE STATUS

2.1 TUBERCULOSIS, MYCOBACTERIAL DISEASES

2.1.1 General evaluation of the national situation

2.1.1.1 Mycobacterium - general evaluation

History of the disease and/or infection in the country

Susceptible population (overall Bovine population size estimate) 730175 animals raised in 40376 holdings. Surveillance systemNational Eradication program for bovine tuberculosis.Method usedRegistration and identification of all bovines.Tuberculin testing of all bovines over the age of 6 weeks.Case definitionInfected animal: Animal positive to tuberculin testing.Infected herd: Herd with one or more animals positive to tuberculin testingVaccination policyVaccination is not permitted.Measures in case of positive findingsSlaughter of positive animals.Ban of animal movement from and within the infected herdRe-examination of the herd and re-establishment of the tuberculosis free health status.

National evaluation of the recent situation, the trends and sources of infection

Epidemiological overview, history and technical evaluationVariations have been recorded on the evolution of bovine Tuberculosis compared to the previous years as the herd prevalence increased from 3,45 in 2011 to 4,33 this year (2014). The herd incidence rate similarly increased from 1,12% in 2011 to 2,66% (2014). In general, the epidemiological indicators are influenced by the number of herds and animals tested in areas with high infection rates.Concerning the overall infection status in the framework of the eradication programme, 213 positive herds with 4082 positive animals were reported at the end of the reporting year 2014. However, following epidemiological data analysis at country level, 10707 herds reported officially free, 3.970 herds reported with suspended health status and 8.700 herds reported as herds of unknown health status.The significant number of herds with unknown health status is mainly due to the livestock structure of the regional unit of Etoloakarnania. This area has a significant number of bovine herds with semi-wild animals of no tuberculosis history that were previously categorized as officially free and from the year 2003 were characterized as herds of unknown health status due to the difficult access in applying animal health programmes at local level. In general, the epidemiological impact of M. bovis situation in 2014 remained steady in endemic areas with observed variations in prevalence and incidence rates in comparison with previous years epidemiological figures. In general, Bovine Tuberculosis infection remains a significant animal health problem in several areas of Greece with endemic characteristics, especially in previous infected herds with adult animals. In addition Control and eradication measures for old and new infected herds should be a major continuous task for the veterinary services at regional and local level. In conclusion, further attempts and actions for investigating the epidemiology of the disease, identifying the source of infection, control the animal movements, tracing the infected farms after identifying TB lesions at slaughterhouse and properly implementing the program respecting the appropriate timetable between the checks will be followed in order to meet the eradication targets of Bovine Tuberculosis for the coming implementation years.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

Relevance as zoonotic disease:In general, bovine Tuberculosis infection remains a significant animal health problem in several areas of Greece.

Additional information

Summary results of the zoonoses monitoring for the year 2014Number of herds under the programme (official controls): 23.927Number of animals under the programme (official controls): 666.863Number of herds tested by tuberculin test: 4.920Number of herds positive: 213Number of new herds positive : 111Number of animals tested by tuberculin test: 175.220Number of animals as positive TB reactors: 4.082Total number of animals slaughtered under the programme: 4.487The results from the eradication programme in 2014 are shown in the relevant EFSA tables.

2.2 BRUCELLOSIS

2.2.1 General evaluation of the national situation

2.2.1.1 Brucella - general evaluation

History of the disease and/or infection in the country

DISEASE: Bovine BrucellosisAFFECTED SPECIES: Animals, BovinesSusceptible population726.221 animals raised in 38.486 holdings Surveillance systemNational Eradication program for bovine brucellosis.Method usedRegistration and identification of all bovinesSerological tests (Rose Bengal and Complement Fixation Test according the Dir. 64/432 as well as Elisa in milk and serum and Serum Agglutination Test) of all bovines over the age of 12 months. Laboratory examination of reported abortions.Case definitionInfected animal: Animal positive to serological tests.Infected herd: Herd with one or more animals positive to serological tests.Vaccination policyVaccination is not permitted.Measures in case of positive findingsSlaughter of positive animals.Ban of animal movement from and into the infected herd.Reexamination of the herd and restoration of the brucellosis free health status.

National evaluation of the recent situation, the trends and sources of infection

Data analysisTables on data for herds and animals investigated during the year 2011 were reported to Commission and EFSA web- based data system alongside the tabulated values of the herd health status according to the epidemiological situation at the end of the year 2011 in the whole country. From 19.786 reported herds at central level under the program, 5.260 herds were tested and 264 herds were found infected (period herd prevalence: 5,02%). From the positive herds, 85 were new cases (incidence: 1,62%). Among 386.867 animals under the program, 220.170 were tested (73.667 tested individually) and 1.885 disease-positive reactors were recorded. Concerning the epidemiological situation at the end of the year, 182 herds were classified as infected herds, 4.459 herds have never been investigated and remained in the unknown health status, 490 herds tested negative and 11.892 herds were reported as officially free. Additionally, in 2.638 herds the health status has been suspended, mainly because the routine serology testing in Blood serum or bulk milk has not been performed during the required by the programme intervals.Further epidemiological investigation of positive herds is necessary to be done as reactors originated from officially free herds based on Reports from Regional and local veterinary authorities.Technical evaluation.Observed variations have been recorded on the evolution of bovine Brucellosis for the year 2011. The 2011 period prevalence rate reported slighter higher (5,02%) compared to the previous year 2010 ((4,62%)). The estimated herd incidence rate decreased from 2% (2010) to 1,62% (2011). The 2011 animal prevalence (0,86%) reported lower in comparison with the previous year 2010 (1,28%). In general, the epidemiological indicators are influenced by the number of herds and animals tested in areas of high infection rate of Bovine Brucellosis.Although the epidemiological situation in 2011 has not significantly improved compared to 2010, Bovine Brucellosis infection still remains a significant animal health problem in several areas of Greece with endemic characteristics, especially in previous infected herds or herds not periodically tested according to the programme requirements. In addition, strict Control and eradication measures for old and new infected herds should be a major task and priority for the veterinary services at regional and local level. In conclusion, further attempts and actions for investigating the epidemiology of the disease, identifying the source of infection and properly implementing the national program shall be urgently undertaken in order to meet the disease eradication targets for the next years.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

Relevance as zoonotic diseaseIn general, bovine Brucellosis infection remains a significant animal health problem in several areas of Greece. The systematic implementation of bovine brucellosis eradication program is associated with the public health relevance of this zoonotic disease.

Recent actions taken to control the zoonoses

RB-51 Vaccination programme in the prefectures of Thessaloniki, Pella and Larisa.As an additional preventive measure in order to rapidly reduce the prevalence of Bovine Brucellosis, a vaccination policy using the RB-51 vaccine (Brucella abortus strain) was initially implemented in the specific high risk area (Thessaloniki) in order to facilitate the progress of the existing Brucellosis eradication programme in Bovine Herds (dairy herds). Since 2013, this vaccination programme was expanded in the prefectures of Pella and Larisa . The vaccination with RB51 vaccine works concurrently with the eradication programme.

Suggestions to the European Union for the actions to be taken

Source of human infection- Causal association.The presence of B. abortus in animals compared to Brucella melitensis in small ruminants, has a lesser public health impact in Humans based on the epidemiology and official records from public health services.

Additional information

Summary Epidemiological and Statistical Data on the evolution of 2014 Bovine Brucellosis Programme are presented in the relevant Reporting Tables of EFSA.Summary results of the zoonoses monitoring in the year 2014Number of herds under the programme (official control): 17.471Number of animals under the programme (official control): 360.371Number of herds tested: 4.664Number of herds positive: 269Number of new herds positive: 142Number of animals tested: 178.920Number of animals tested individually: 73.741Number of animals positive: 3.435Total number of animals slaughtered: 3.854

2.2.2 Brucella in animals

2.2.2.1 B. abortus in animal - Cattle (bovine animals)

Vaccination policy

RB-51 Vaccination programme in Thessaloniki prefectureAs an additional preventive measure in order to rapidly reduce the Bovine Brucellosis Prevalence, a vaccination policy using the RB-51 vaccine (Brucella abortus strain) was implemented in the specific high risk area (Thessalonika) in order to facilitate the progress of the existing Brucellosis eradication programme in Bovine Herds (dairy herds) which is in force and works simultaneously with the vaccination strategy.The evolution of the 2005 vaccination programme is presented in the table below :TABLE 1HERDS UNDER THE PROGRAM800ANIMALS UNDER THE PROGRAM42.445VACCINATED HERDS 141ANIMALS IN VACCINATED HERDS10.295VACCINATED ANIMALS8.203 CUMULATIVE HERD COVERAGE AT THE END OF THE YEAR 200542%ANIMAL COVERAGE IN VACCINATED HERDS80 %CUMULATIVE ANIMAL VACCINATION COVERAGE AT THE END OF THE YEAR 200545%

2.2.2.2 B. melitensis in animal - Sheep

Status as officially free of ovine brucellosis during the reporting year

The entire country free

Non officially free Country

Additional information

Total Susceptible population (Data 2014 / Directorate of Animal Health , MRDF)13.300.622 sheep and Goats raised in 80.520 herds.Surveillance systemThe control program for ovine and caprine brucellosis is in force in the mainland (includes mass vaccination policy in young and adult sheep and goat population) and Brucellosis eradication program runs in islands.Method usedRegistration and identification systems applied in animals.Serological test (test and slaughter policy) in animals raising in the islands.Animal mass vaccination in the mainland.Case definitionInfected animal: Animal positive to serological tests.Infected herd: Flock with one or more animals positive .Vaccination policyVaccination according to the control program. Measures in case of positive findings (according to the eradication program)Slaughter of positive animals.Ban of animal movement from and to the infected herd.Re-examination of the herd and re establishment of the brucellosis free health status.

Vaccination policy

SEMI-WILD BOVINE VACCINATION WITH REV 1 VACCINE As an additional preventive measure under the existing control and eradication brucellosis programme for sheep and goats, the free-ranged (semi-wild) bovines that are sharing common pastures with small ruminants, were vaccinated with REV-1 vaccine in order to reduce the spread of Brucella infection in the field.Number of Bovine herds vaccinated with REV-1 vaccine: 849Number of Bovine animals vaccinated with REV-1 : 10.901

Control program/mechanisms

The control program/strategies in place

EPIDEMIOLOGICAL SITUATION IN THE ISLANDS DATA ANALYSISIn the islands (eradication zone), except Evia, Lesvos and Leros, the 2014 flock incidence and prevalence rates among tested sheep and goats flocks were reported 1% % and 1% % respectively. What is important is that there is a significant reduction in the flock prevalence in comparison to 2011, where the flock prevalence was 5,38% The animal prevalence reported 0,05% in 2014, which is lower in comparison to the same indicator in 2011(0,31 %) . The islands of Lesvos and Leros have been excluded from the eradication policy and belong to the mainland vaccination programme status. Summary results of the zoonoses monitoring in the year 2014 from the eradication zone Number of flocks under the programme (official control): 27.391Number of animals under the programme (official control): 4.144.704Number of flocks tested: 1802Number of flocks positive: 22Number of new flocks positive: 10Number of animals tested individually: 162.350Number of animals positive: 89Total number of animals slaughtered: 84

Notification system in place

Mandatory notification status.

Results of the investigation

EPIDEMIOLOGICAL SITUATION IN THE MAINLAND DATA ANALYSISSummary results of the official mass vaccination 2014 programme in sheep and goats:Mass vaccination carried out in the Mainland. During 2014, based on vaccination records and reports from the Regional Veterinary Units (at Prefecture level) , 1.036.494 sheep and goats from 28.399 herds were vaccinated with the vaccine REV 1 . Further analysis and detailed statistics (flock and animal vaccination data, follow up and up to date vaccination activities) are available through the central data base files of the Department of Zoonoses (Animal Health Directorate).Number of flocks vaccinated: 28.399Number of animals vaccinated : 1.036.494

National evaluation of the recent situation, the trends and sources of infection

Epidemiological and Technical evaluation The Ovine and Caprine Brucellosis control and eradication programme has been implemented in the mainland and islands of Greece in 2014. The 2014 B.melitensis programme was carried out with co-financing status by the E.U based on the Commission Decision 2013/722/EC. Greek Farmers obtained compensation for positive-infected sheep and goats based on the requirements and provisions of the National Ministerial Decision. The Country is divided in 13 prefectures - Peripheries. For the implementation of brucellosis control and eradication programme, Greece is divided in two programme zones in which different policies and measures are applied, the control strategy in the mainland (mass vaccination of young and adult female small ruminants) and the eradication policy in the islands which is based on test and slaughter of positive reactors receptively.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

Relevance as zoonotic diseaseThe relevance of the disease has a significant impact at Public Health level for the Community and consumers. Source of human infectionMainly from animal contact and consumption of dairy products (especially consumption of dairy products derived from non heated and pasteurized milk or immature types of sheep and goat cheese). In addition, it should be acknowledged the possible risk of obtaining the disease, if various home- made dairy products of unknown origin and hygiene quality are eaten by the consumers.

Additional information

Epidemiological historyOvine and caprine brucellosis due to B. melitensis is a significant disease for both public health and animal production in Greece. During the last years a control and eradication program is running by the veterinary services of the Ministry of Rural Development and Food. The aim of the program is to control the incidence and prevalence of the disease in areas of the country where these estimates are reported high, by vaccination of lambs and kids. At the same time, in the remaining parts of the country, where the prevalence of the disease is reported low among sheep and goat flocks, an eradication program is implemented by test and slaughter policy. Source of human infectionMainly from animal contact and consumption of dairy products of unknown origin and hygiene quality. (Especially, consumption of dairy products prepared from non pasteurized milk or immature types of sheep and goat cheese).

3 INFORMATION ON SPECIFIC ZOONOSES AND ZOONOTIC AGENTS

Zoonoses are diseases or infections, which are naturally transmissible directly or indirectly between animals and humans. Foodstuffs serve often as vehicles of zoonotic infections. Zoonotic agents cover viruses, bacteria, fungi, parasites or other biological entities that are likely to cause zoonoses.

3.1 SALMONELLOSIS

3.1.1 General evaluation of the national situation

3.1.1.1 Salmonella - general evaluation

History of the disease and/or infection in the country

DISEASE/AGENT: Salmonellosis, Salmonella spp.Contaminated materials: Feed materials of animal origin, plant origin and Compound feedingstuffs Surveillance systemThe legal provisions in place and relevant requirements (Zoonoses Directive 2003 and Zoonoses Regulation 2160) had significantly improved the effectiveness of the existing monitoring situation and management practices in the field of Salmonella surveillance. New strategies and schemes for monitoring Salmonella zoonotic agents are in force in accordance with Community Salmonella reduction targets approved. Rapid adaptation and compliance on the new mandatory EU Salmonella control and eradication programmes were observed in all EU member states.Measures in case of positiive findingsAccording to the current EU Directives and Community Legislation. In 2014, 7 Salmonella positive units (S. Chester) form fish meal were reported from 95 sampling units tested, 2 Salmonella positive units (S. Seftenberg) from feed material of animal origin were reported form 25 sampling units tested, 7 Salmonella positive units (2 S. Seftenberg and 2 S. Livingstone) feed of land animal origin were reported form 20 sampling units tested and 2 Salmonella positive units (Salmonella Enteritidis) from compound feedingstuff for poultry were reported form 5 sampling units tested. The rest of the feedingstuffs tested were negative for salmonella spp.In 2010, five (5) Salmonella positive units (from fish meal) were reported from 212 sampling units tested in total under selective sampling and routine monitoring schemes.In 2009, no Salmonella positive units were reported from 232 samples tested in total under selective sampling and routine monitoring schemes.The method ISO 6579 (2002) is used for the detection and isolation of Salmonella serovars.

National evaluation of the recent situation, the trends and sources of infection

DISEASE/AGENT: Salmonellosis, Salmonella SerovarsINFECTED SPECIES: Poultry breeding flocks- Callus gallusSusceptible populationParent breeding stock for egg and meat production line.Surveillance systemFrom the past (Historical data), according to the Annex III of the Dir. 92/117, a Salmonella control program has been carried out since 1998. In 2007, the Salmonella national control programme in breeding flocks of Gallus gallus has been implemented and approved (co- financed) by European Commission. The programme was supervised by the Hellenic central veterinary competent authorities and was in line with the uniform EU guidelines and rules approved by the Commission. The results were collected, analyzed and evaluated by the Commission, EFSA and Member States in accordance with the Community pre- defined targets towards the reduction of Salmonella prevalence in Breeding flocks of Gallus gallus (fowl). Method used:The methods ISO 6579 (2002) and ISO 6579 Amendment 1: Annex D (current version) were used for the detection and isolation of Salmonella serovars.The Salmonella serotyping was conducted in the National Reference Veterinary Laboratory (NRVLS) for Salmonellosis in animals (located in Chalkida Prefecture of Evia) by using the Kauffman-White Le Minor method .Measures in case of positive findingsMeasures comply with part C of Annex II to Regulation (EC) No 2160/2003. Epidemiological and statistical reportIn 2014, in Greece there were 234 adult breeding flocks with more of 250 birds in 85 breeding holdings. All of them were tested for zoonotic salmonella in the context of NSCP. 4 out of the 234 adult flocks were positive for the targeted serovars (2 SE, 1 ST and 1 SI) and 11 were positive for other salmonella serovars (S. Livingstone, S. Bredney, S. Havana, S. Munster, S. Seftenberg and S. Umbilo). Rearing breeding flocks were negative for Salmonella. DISEASE/AGENT: Salmonellsis / Salmonella serovarsINFECTED SPECIES: Laying Hens and Broilers of Gallus, gallus (fowl)Surveillance systemIn 2014, Salmonella control and eradication EU- programmes in Laying hens and broilers have been implemented in the country based on suspected and objective samples submitted into the laboratories under the official investigation in 2014. Under the framework of the programme industry sampling was carried out as well. Method usedThe methods ISO 6579 (2002) and ISO 6579 Amendment 1: Annex D (current version) were used for the detection and isolation of Salmonella serovars in Laying hens and other poultry.The Salmonella serotyping was performed by using the Kauffman-White Le Minor method . In 2014, in Greece there were 603 adult laying flocks in 435 laying hen holdings. 401 of them were tested for zoonotic salmonella in the context of NSCP. 8 out of the 401 adult flocks were positive for the targeted serovars (7 SE, 1 ST) and 30 were positive for other salmonella serovars.Due tothese results, the EU target was achieved in 2014.In 2014, in Greece there were 7551 broiler flocks in 1041 holdings. 7504 of them were tested for zoonotic salmonella in the context of NSCP. The rest were not tested because they were not at the proper age. 19 out of the 7504 tested flocks were positive for Salmonella spp. None of them was positive for the targeted salmonella serovars (SE/ST)Due tothese results, the EU target was achieved in 2014.In 2014, the National Salmonella Control Programme has implemented in flocks of breeding and fattening turkeys and the EU target was achieved for both animal populations.

Recent actions taken to control the zoonoses

EU LEGISLATION FOR CONTROL SALMONELLA PROGRAMMES IN POYLTRYZOONOSES DIRECTIVEHYGIENE PACKAGE

Additional information

DISEASE/AGENT: Salmonella/ Salmonella serovarsTARGET OF MONITORING: Contaminated FoodSurveillance systemRoutine examination and selective official sampling at retail level, processing plan and slaughterhouse carried out based on National and Community legislation.Method usedThe ISO 6579 (2002) is used for the detection of Salmonella in food.The Salmonella serotyping was performed by the Agglutination technique: Antigenic formulas of the Salmonella Serovars (9th edition- 2007- WHO Institute Pasteur) Summary National Report (Reporting Year: 2011). Official and selective sampling Routine monitoring1. Broiler meat and products thereof (all categories)Samples tested: 315Samples positive: 8Reported serovars : S. Enteritidis (n=1), S.Montevideo (n=1) , S. Hadar (n=4) and S. spp unspecified (n=1)2. Pig meat and products thereof (all categories)Samples tested: 356Samples positive: 7Reported serovars : S. Derby (n=2), S. Umbilo (n=5). 3. Bovine meat and products thereof (all categories)Samples tested: 100Samples positive: 04. Meat from other animals and products thereof (all categories)Samples tested: 1Samples positive: 05. Milk and milk products (all categories) Samples tested: 418Samples positive: 06. Eggs and egg products(all categories)Samples tested: 10Samples positive: 07. Fish and fish products (all categories)Samples tested: 53Samples positive: 08. Other Food (all categories)Samples tested: 28Samples positive: 02010 Overall Salmonella reported Food Prevalence (for all food categories) = 3,17 % (75/2367*100)

3.1.2 Salmonella in animals

3.1.2.1 Salmonella in animal - Meat from pig - carcase

Additional information

Meat from pig-carcass (food sample carcase swabs) were not reported for the year 2015, because no isolates deriving from official nor own checks arrived to the Greek NRL for serotyping and AST

3.2 CAMPYLOBACTERIOSIS

3.2.1 General evaluation of the national situation

3.2.1.1 Thermophilic Campylobacter spp., unspecified - general evaluation

History of the disease and/or infection in the country

in 2014, a monitoring programme took place in broiler flocks Gallus gallus according to the requirements of CD 2013/652/EU. The results are presented in the relevant tables.

National evaluation of the recent situation, the trends and sources of infection

DISEASE/AGENT: Thermophilic CampylobacterTARGETS : Animals / Contaminated FoodSurveillance system- HistoryThere is not yet in force an official systematic national Campylobacter control program for animals and food. Sporadic selective samples are collected and examined, especially from sheep (aborted fetus in the field) and broilers (at slaughterhouse).Results of 2009 zoonoses monitoringAnimals: Cattle (n=20), Sheep (n=56) , Goats, Pigs, Horses (n=1) were officially tested and 11 animals (sheep) were found positive to Campylobacter fetus (animal sample: aborted fetus / stomach content). Food : Targeted official sampling of fresh broiler meat at processing plant level revealed 33 positive samples out of 47 tested (Campylobacter spp- unspecified) Data are presented in the relevant tables of EFSA web based electronic system for zoonoses monitoring.

3.3 LISTERIOSIS

3.3.1 General evaluation of the national situation

3.3.1.1 Listeria - general evaluation

National evaluation of the recent situation, the trends and sources of infection

DISEASE/AGENT: ListeriosisAFFECTED SPECIES: Animals and FoodSurveillance systemRoutine and targeted official sampling performed by the national veterinary public health authorities and the Hellenic Food Safety Authority (EFET) respecting the microbiological criteria foreseen by Community Legislation and Hygiene Package.Method usedThe laboratory methods used for Listeria detection and enumeration were : ISO 11290.01 Part 1 (1997), ISO 11290.01/A1 Amendment 1 (2005) and ISO 11290.02 /A1 Part 2 and Amendment 1 (2005) respectively.Summary results of 2014 are presented in the relevant EFSA tables.Summary selected statistical results of 2011 zoonoses monitoringSample CategoriesPercentage % of positive samples among tested units for Listeria monocytogenesAnimals (sheep and Goats) 2,78Other products0Pig Meat 1,66%Pig meat products cooked ready to eat 75%Milk and dairy products0,34%Data analysis are presented in the relevant tables of EFSA web based electronic system for zoonoses monitoring. Summary Statistical ResultsThe overall 2011 reported and calculated percentage of Listeria positive findings (units) in all tested samples was 2,37% (19/800*100) for all food categories examined. This rate is slightly higher compared to the related percentage of positive samples for Listeria in 2010 (1,06 % (5/468*100), which was significantly lower compared to the related percertage of positive samples tested in 2009 and was attributed to Pig meat and products thereof contaminated with Listeria monocytogenes. The overall 2009 reported and calculated percentage of Listeria positive findings (units) in all tested samples was 5,87 % (84/1432*100) for all food categories examined. This rate is significantly higher from the related percertage of positive samples tested in 2008 and was attributed to Pig meat and products thereof contaminated with Listeria monocytogenes.The overall 2008 reported and calculated percentage of Listeria positive findings (units) in all tested samples was 1,53 % (28/1826*100) for all food categories examined.

3.4 YERSINIOSIS

3.4.1 General evaluation of the national situation

3.4.1.1 Yersinia - general evaluation

National evaluation of the recent situation, the trends and sources of infection

DISEASE/AGENT: YersiniosisAFFECTED SPECIES: Animals and FoodNo Data were available at central authority level for animal and Food in 2011

3.5 TRICHINELLOSIS

3.5.1 General evaluation of the national situation

3.5.1.1 Trichinella - general evaluation

National evaluation of the recent situation, the trends and sources of infection

DISEASE/AGENT: Trichinellosis , Trichinella spp.AFFECTED SPECIES: AnimalsSusceptible populationAll domestic farmed and wild swine eligible for slaughter.Surveillance systemCompulsory examination for detection of Trichinellosis at Slaughterhouse level.Method usedTwo main diagnostic methods for Trichinella spp in fresh pork meat are used. The first comprises the digestion in artificial gastric juice of muscle tissues from Trichinella pre- determined sites, followed by the microscopic examination of parasitic larvae. The second commonly used in the past covers the examination of tissues from diaphragm in the trichinoscope. New Community legislation (Commission Regulation 2075/2005) which has been adopted by the EU describes diagnostics techniques and sampling methods for target species (swine) expected to be fully implemented on mandatory basis by the national monitoring Trichinella systems in all Member- States. Epidemiological historyData on Trichinella spp. unspecified coming from the meat inspection activities in 2014 are presented in the relevant EFSA tables. 13 positive findings (Trichinella spp. Unspecified) were reported at slaughterhouse level under the meat inspections activities in 2011. The positive samples derived from meat of wild farmed boars raised in Northern Greece. The positive units were sent to Community Reference Laboratory for further diagnostics and parasitic identification. During the reporting year 2011, 1.217.530 pigs were tested for trichinella spp at slaughterhouse level. The targeted animals were examined by the new official reference method of Trichinella detection as forseen and described in the Annex 1 of the Commission Regulation 2075/2005 (Magnetic stirrer method for pooled sample digestion).Results of monitoring13 positive wild farmed boars were found in the framework of zoonosis monitoring. The causative agent was Trichinella spp- unspecified. Data are presented in the relevant table of EFSA web based electronic system for zoonoses monitoring.

3.6 ECHINOCOCCOSIS

3.6.1 General evaluation of the national situation

3.6.1.1 Echinococcus - general evaluation

National evaluation of the recent situation, the trends and sources of infection

DISEASE/AGENT: EchinococcosisAFFECTED SPECIES: Susceptible Animals: Cattle, sheep, Goats, Pigs.Susceptible populationAll animals eligible for slaughter at country level.Surveillance systemInspection of all carcasses at slaughterhouse level.Preventive treatment of all domestic and farm dogs with antiparasitic tablets. Method usedFor farmed animals , meat inspection of carcasses at slaughterhouses. For dogs the arecolin test applied in the past.Epidemiological historyThe infection among the owned dogs has been almost disappeared due to systematic preventive treatment of animals with antiparasitic medication. The infection rate in stray dogs is difficult to be estimated. The overall infection in farmed animals remained stable compared to previous reporting years.Results of 2014 zoonoses monitoring are presented in the relevant EFSA tables.Results of 2011 zoonoses monitoringAnimal species Prevelance (%) at slaughterhouse levelSheep1,23%Goats0,39%Bovine0,71%Pigs 0%Results of 2009 zoonoses monitoringAnimal species Prevelance (%) at slaughterhouse levelSheep1,85Goats0,46Bovines1.01Pigs 0,00Data analysis are presented in the relevant tables of EFSA web based electronic system for zoonoses monitoring. Source of human infectionMainly through the consumption of contaminated raw foodstuffs (i.e vegetables).

3.7 RABIES

3.7.1 General evaluation of the national situation

3.7.1.1 Lyssavirus (rabies) - general evaluation

History of the disease and/or infection in the country

The last rabies case in animals was detected in the Evros prefecture in 1987. Nevertheless the reoccurrence of this infectious disease in Greece in 2012 could not be characterized as unexpected due to several existing observations. First, there was a high prevalence of the disease in neighboring countries such as Turkey, Bulgaria, Albania and FYROM. This, supported concerns for trans-border rabies spread in Greece from any of these countries. Taking into consideration this situation, in 2012, the pre-existing national program for rabies passive surveillance according to Directive 99/2003/EC was enhanced imposing the collection of all dead and suspected for rabies animals from 16 regional units along the northern and eastern borders of the country. In October 2012 rabies was diagnosed in a red fox (Vulpes vulpes) in Kozani Regional Unit. By the end of 2012 nine positive cases were confirmed by the National Reference Laboratory for rabies. The partial sequencing analysis and the subsequent phylogenetic analysis supported the hypothesis of movement of rabies-infected hosts in Western Balkan countries. To address the epidemic and prevent its spread, a Greek National Rabies control and eradication programme was implemented based on the passive surveillance of the disease (-following the identification of the infected fox, the collection of samples from dead animals was extended in 2013 on the entire country with a main aim to achieve a more efficient surveillance of the disease-), the mandatory vaccination of all dogs and cats, the management of all rabies casessuspect animals and the control in animal movements. Until April 2016, in the frame of the passive surveillance of the disease, a total of 48 rabid animals have been confirmed by the National Reference Laboratory for Rabies in animals (NRL), which is the Virology Laboratory in Athens Veterinary Center. In detail, nine (7 foxes, 2 dogs), twenty nine (25 foxes, 1 dog, 1 cat, 2 cattle) and ten (8 foxes, 2 dogs) rabies cases were laboratory confirmed in 2012, 2013 and 2014 respectively, whereas the last rabies case was confirmed in a red fox in Pella Regional Unit in May 2014. Since then no new cases have been recorded.

National evaluation of the recent situation, the trends and sources of infection

Regarding the passive surveillance program, brain samples from 257 animals were collected during the 2015. All these samples were negative for rabies virus. From the total of 257 animals (nervous tissue samples) tested for the detection of rabies virus in 2015, 249 animal samples were indicators (showing clinical signs or abnormal behavior suspect of rabies, were found dead, road kills or were animal involved in human exposure). 8 animals (mice) were tested in the frame of the annual check performed in Democritus lab. Furthermore, an Oral Vaccination project for the immunization of red foxes against rabies was launched in Greece. The first vaccination campaign took place in autumn/winter 2013, the second in autumn/winter 2014 and the third in autumn/winter 2015. Due to fiscal reasons in the country, and besides the efforts of the Ministry so as to proceed to the tender for the vaccination campaigns of the spring of each year, this was not achievable finally until 2015. However the first spring campaign has been implemented in the country (April to May 2016). The second oral rabies vaccination campaign (autumn-winter 2014) in Greece was completed early in 2015 and more precisely on 22.01.2015. The third oral vaccination program (autumn/winter 2015) was implemented as the previous campaigns, in twenty four regional units of the Greek territory during the period November-December of 2015. A communication campaign (posters and leaflets, TV spot, special educational material and courses for children in schools, press rleases) has been implemented with the assistance of the Ministry of Education, the regional veterinary services and the students associations of the two Veterinary Schools, in order to raise the public awareness with regards to rabies and inform the public regarding the philosophy of red fox oral vaccination programme, the time schedule of the vaccination strategy, the areas involved and the necessary prophylactic measures to be adopted in order to avoid potentially harmful exposures to the live attenuated vaccine distributed by air. Vaccine-baits were aerially distributed by fixed-wind aircrafts. Vaccine-baits were aerially distributed in 24 regional units of the country (20 -25 baits per km2). Vaccines were dropped along parallel flight paths 500 m apart in order to optimize aerial missions and achieve homogeneous distribution. The baits were manually dropped one after another following a sound signal (alarm), in order to meet the predetermined density of vaccine baits per square kilometer. A GPS equipment existed in each aircraft registering the coordinates of individual bait position. The whole procedure was supervised by official veterinarians. Monitoring following the vaccinations of autumn 2015 The following monitoring program assessing the effectiveness of oral vaccination campaign was based on the examination of serum samples and canine teeth derived from red foxes collected in the field. The initiation of the program in each regional unit was performed one month following the end of vaccination in this specific area and lasted for all regional units at least two months. The collection of samples was finally completed in the end of March 2016 (just before the new vaccination campaign of spring 2016). The total number of samples required for the monitoring of the effectiveness of the oral vaccination campaign was 2 red foxes/100 km2 as it is recommended by WHO and the target for the total number of collected animals was estimated to be 1.212 in all 24 regional units. For the monitoring following the ORV 2015, samples derived from 115 red foxes were hunted in the frame of active surveillance and are being in process of examination in the NRL for Rabies and the results will be obtained up to the middle of the summer of 2016. All of the brain samples have been already examined with FAT and found negative. The Tetracycline examination as well as the serological testing are still in process.

Recent actions taken to control the zoonoses

Implementation of oral vaccination campaigns with the aerial distribution of vaccine baits. One month after the completion of each campaign, an active monitoring programme is taking place in order to evaluate the effectiveness of the vaccination campaign.

3.7.2 Lyssavirus (rabies) in animals

3.7.2.1 Lyssavirus (rabies) in animal - Dogs

Vaccination policy

All dogs over three months of age are mandatory vaccinated against rabies.

3.8 Q-FEVER

3.8.1 General evaluation of the national situation

3.8.1.1 Coxiella (Q-fever) - general evaluation

National evaluation of the recent situation, the trends and sources of infection

DISEASE/AGENT: Coxiella burnetii (Q fever) in animalsAFFECTED SPECIES: Animals/ sheep and goats mainly Surveillance systemThere is no official National monitoring program in place. Sporadic blood (sera) samples are officially collected and examined following notification of abortion at farm level, especially from sheep and goats.Results of monitoringIn 2010 , 306 sampling units (animals : Cattle n= 11, sheep n =181, goats n=114) were tested for Coxiella burnetii and 56 found positive.Data are presented in the relevant table of EFSA web based electronic system for 2010 zoonoses monitoringIn 2009 , 66 units (animals) were tested for Coxiella burnetii and 13 found positive.Data are presented in the relevant table of EFSA web based electronic system for 2009 zoonoses monitoringEpidemiological historyDuring the period 2001-2006 Coxiella burnettii was detected in 68 small ruminant flocks and 1 bovine herd (Table 1). Animal infection rate in affected flocks ranged from 2.1 % to 31.5%. Findings, along with previously conducted studies indicate the existence of the disease in animals. However, no data for animals or foods exist on a systematic basis. Table 1.: Q fever in small ruminants, 2000-2006.Year 20012002 2003 2004 2005 2006Number of infected flocks28 17 1 8 7 7Source: MRDF

3.8.2 Coxiella (Q-fever) in animals

3.8.2.1 C. burnetii in animal - Sheep and goats - Farm - Clinical investigations

National evaluation of the recent situation, the trends and sources of infection

DISEASE/AGENT: Coxiella burnetii (Q fever) in animalsAFFECTED SPECIES: Animals/ sheep and goats mainly Surveillance systemThere is no official / National program in place. Sporadic blood (sera) samples are collected and examined following notification of abortion at farm level , especially from sheep and goats.Results of monitoringData are presented in the relevant table of 2008 EFSA web based electronic system for zoonoses monitoringEpidemiological historyDuring the period 2001-2006 Coxiella burnettii was detected in 68 small ruminant flocks and 1 bovine herd (Table 1). Animal infection rate in affected flocks ranged from 2.1 % to 31.5%. Findings, along with previously conducted studies (7), indicate the existence of the disease in animals. However, no data for animals or foods exist on a systematic basis. Table 1. Q fever in small ruminants, 2000-2006.Number of infected flocks:Year2001200220032004200520062817877Source: MRDF (Hellenic Ministry of Rural Development and Food)

3.9 ESCHERICHIA COLI, NON-PATHOGENIC

3.9.1 General evaluation of the national situation

3.9.1.1 Escherichia coli, non-pathogenic - general evaluation

Additional information

Results of investigations in the year 2010Only targeted food samples (n= 148) were tested for E.coli spp- non pathogenic in 2010 with negative results.

3.10 TOXOPLASMA

3.10.1 General evaluation of the national situation

3.10.1.1 Toxoplasma - general evaluation

National evaluation of the recent situation, the trends and sources of infection

DISEASE/AGENT: ToxoplasmosisAFFECTED SPECIES: AnimalsIn 2011, 363 blood sera samples were sent to the appropriate laboratory (181 goats and 182 sheep) in the frame of investigating abortions. The tests were not performed due to lack of reagents and therefore only suspected toxoplasmosis can be based on the clinical picture.Toxoplasma gondii is detected in Sheep and Goats tested under national Surveys. The laboratory methods used for 2011 was the IFAT (Indirect Immunofluorescence Antibody test) for detecting Toxoplasma antibodies in blood sera and microscopic examination of the brain of aborted fetuses. The sampling schemes were not random or representative, originated from sheep and goats flocks with reported abortions under clinical investigation practices. Results of 2011 monitoringData are available in the relevant tables of EFSA web based electronic system for zoonoses monitoring. There are no available data on toxoplasmosis for 2011.

3.11 VTEC

3.11.1 General evaluation of the national situation

3.11.1.1 Verotoxigenic E. coli (VTEC) - general evaluation

National evaluation of the recent situation, the trends and sources of infection

DISEASE/AGENT: Verocytotoxic E.coli AFFECTED SPECIES: Animals / FoodSurveillance systemThere is no official National monitoring program in force for detecting VTEC serovars in animals and food. Results of investigations in the year 2011Only severar animal and food samples were tested for E.coli spp in 2011

4 ANTIMICROBIAL RESISTANCE INFORMATION ON SPECIFIC ZOONOSES AND ZOONOTIC AGENTS

4.1 SALMONELLOSIS

4.1.1 Salmonella in animals

4.1.1.1 Antimicrobial resistance in Salmonella Poultry, unspecified

National evaluation of the recent situation, the trends and sources of infection

ANTIMICROBIAL RESISTANCE No official national program is in force. Efforts commenced to develop a systematic reporting system of antimicrobial resistance in various animal species. The results are limited and the only available information is mainly provided from the National Reference Laboratory for Salmonella. Relevant reports for Antimicrobial susceptibility testing in Animals (for both quantative and qualitative data) have been increased year per year at national level, especially for Salmonella agents.

Additional information

The overall monitoring of antimicrobial resistance , especially for the Salmonella isolates in Poultry was carried out using two laboratory methods.1. Performance Standards for Antmicrobial Disk Susceptibility Tests-Ninth Edition; Approved Standard January 2006 CLSI (M2-A9,Vol.26.No1 and Eighteenth Information Supplement CLSI (M100-S18, Vol.28 No 1)2. Broth Microdilution Method (MIC)- Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria that grow Aerobically; Approved Standard January 2009 CLSI (M07-A8, Vol. 29 No2) and Standard for breakpoint from EFSA suggestions.

4.2 ESCHERICHIA COLI, NON-PATHOGENIC

4.2.1 Escherichia coli, non-pathogenic in foodstuffs

4.2.1.1 Antimicrobial resistance in Escherichia coli, non-pathogenic Meat from bovine animals

Description of sampling designs

Fresh bovine meat samples gathered from the retail were randomly selected from different super markets and butchers shops (originated from 4 different municipality units).

Stratification procedures per animal populations and food categories

The Monitoring program of AMR for 2015 started with delay and was implemented the last 3 month of the year. The samples of bovine meat were randomly selected from the retail without pre selection based on the origin of the food, from 4 municipality units in Greece.

Randomisation procedures per animal populations and food categories

The Monitoring program of AMR for 2015 started with delay and was implemented the last 3 month of the year. The samples of bovine meat were randomly selected from the retail without pre selection based on the origin of the food, from 4 municipality units in Greece.

4.2.1.2 Antimicrobial resistance in Escherichia coli, non-pathogenic Meat from pig

Description of sampling designs

Fresh pig meat samples gathered from the retail were randomly selected from 98 different super markets and butchers shops (originated from 6 different municipality units).

Stratification procedures per animal populations and food categories

The Monitoring program of AMR for 2015 started with delay and was implemented the last 3 month of the year. The samples of pig meat were randomly selected from the retail without pre selection based on the origin of the food, from 6 municipality units in Greece.

Randomisation procedures per animal populations and food categories

The Monitoring program of AMR for 2015 started with delay and was implemented the last 3 month of the year. The samples of pig meat were randomly selected from the retail without pre selection based on the origin of the food, from 6 municipality units in Greece.

4.2.2 Escherichia coli, non-pathogenic in animals

4.2.2.1 Antimicrobial resistance in Escherichia coli, non-pathogenic Pigs

Description of sampling designs

caecal samples gathered at slaughter from fattening pigs were randomly selected from 19 slaughterhouses of large throughput of the country (10 different municipality units). Each sample derived from a different epidemiological unit (different holding)

Stratification procedures per animal populations and food categories

Indicator commensal E.coli & ESBL and AmpC- producing .coli : The Monitoring program of AMR for 2015 started with delay and was implemented the last 3 month of the year (two seasons were covered). A total number of 19 slaughterhouses of large throughput were selected for sampling. One caecal sample deriving from a unique pig carcass was gathered from each epidemiological unit slaughtered. Just one isolate per holding was tested and reported to EFSA.

Randomisation procedures per animal populations and food categories

Indicator commensal E.coli & ESBL and AmpC- producing .coli : The Monitoring program of AMR for 2015 started with delay and was implemented the last 3 month of the year (two seasons were covered). A total number of 19 slaughterhouses of large throughput were selected for sampling. One caecal sample deriving from a unique pig carcass was gathered from each epidemiological unit slaughtered. Just one isolate per holding was tested and reported to EFSA.

5 INFORMATION ON SPECIFIC MICROBIOLOGICAL AGENTS

5.1 HISTAMINE

5.1.1 General evaluation of the national situation

5.1.1.1 Histamine - general evaluation

National evaluation of the recent situation, the trends and sources of infection

DISEASE/AGENT: Histamine in Food Surveillance systemThere is no official monitoring program or systematic scheme applied for Histamine in food. Sporadic samples from fish and fishery products are examined in the designated national veterinary laboratory in Thessalonica- Grrece. Targeted fish species for testing and detecting Histamine are: Scrombridae, Clupeidae, Engraulidae, Coryfenidae, Pomatomidae and Scrombresosidae. Related Legislation: Community Regulation (EC): 1141/2007.Results of monitoringData are presented in the relevant table of EFSA web based electronic system for 2009 zoonoses monitoring.

6 FOODBORNE OUTBREAKS

Foodborne outbreaks are incidences of two or more human cases of the same disease or infection where the cases are linked or are probably linked to the same food source. Situation, in which the observed human cases exceed the expected number of cases and where a same food source is suspected, is also indicative of a foodborne outbreak.

6.1 Outbreaks

6.1.1 Foodborne outbreaks

System in place for identification, epidemological investigations and reporting of foodborne outbreaks

Foodborne outbreaks are included in the Mandatory Notification System (MNS) of the Hellenic Centre for Diseases Control and Prevention (HCDCP). Once a food-borne outbreak is notified, the public health professionals conduct an epidemiological investigation in order to estimate the extent of the outbreak, identify the source and implement control measures. Furthermore, the Public Health Directorate of the competent Prefecture, the National Food Agency, as well as the Ministry of Rural Development and Food are informed, whenever it is needed. The number of reported food-borne outbreaks has been quite stable since 2004. However, data of the MNS should be interpreted with caution due to the probable under-reporting. It should be mentioned that the reporting date was used for the analysis of data. With regard to the severity of illness, 13 (2.5 %) out of the 525 outbreak-related cases, were hospitalised. Finally, 2 outbreak-related deaths due to LIsteria monocytogenes were reported in 2015.

National evaluation of the reported outbreaks in the country:

Trends in numbers of outbreaks and numbers of human cases involved

The increase in the number of human cases can be explained by the fact that large, general waterborne outbreaks were reported in 2015, while outbreaks that had been reported on 2014 were not so extensive and regarded smaller number of cases. Additionally, in 2015, we decided for the first time to report FBOs with less than 10 cases, but with a special public health interest. This change in the reporting method explains the increase in the number of the reported FBO. Thus, this increase cannot be considered as a true one.

ANIMAL POPULATION TABLES

Table Susceptible animal population

		Population			
Animal species	**Category of animals**	**holding**	**animal**	**slaughter animal (heads)**	**herd/flock**
Cattle (bovine animals)	Cattle (bovine animals)		791,220	83,373	21,941
Chinchillas	Chinchillas - farmed	1	800		1
Deer	Deer - farmed		613		17
Ducks	Ducks		12,859		335
Fish	Fish	5	55		
	Fish - farmed	436	95,876		
	Fish - farmed - carp	7	35		
	Fish - farmed - salmon	2	1,500		
Gallus gallus (fowl)	Gallus gallus (fowl)			25,969,676	
	Gallus gallus (fowl) - breeding flocks, unspecified	84	2,513,479		379
	Gallus gallus (fowl) - broilers	1,084	100,121,080		6,872
	Gallus gallus (fowl) - laying hens	528	5,597,927		848
Geese	Geese		4,560		228
Goats	Goats		3,953,655	407,372	7,882
Minks	Minks - farmed		333,000		83
Ostriches	Ostriches - farmed		14		2
Other animals	Other animals	4	220		
	Other animals - unspecified	24,582	1,979,282		
Partridges	Partridges			945	
Pheasants	Pheasants		600	1,044	1
Pigs	Pigs		488,230	626,782	1,745
Quails	Quails		14,500	25,490	2
Rabbits	Rabbits - farmed		32,426	19,210	2,339
Reindeers	Reindeers - farmed		86		2
Sheep	Sheep		9,481,181	1,395,392	24,041
Sheep and goats	Sheep and goats				46,659
Shellfish	Shellfish	590	18,011		
Solipeds, domestic	Solipeds, domestic		25,533		10,209
Turkeys	Turkeys - meat production flocks	42	316,610		77
	Turkeys - parent breeding flocks	5	9,100		6
Wild boars	Wild boars - farmed		3,388	1,714	

DISEASE STATUS TABLES

Table Bovine brucellosis in countries and regions that do not receive Community co-financing for eradication programme

Region	Number of suspended herds under investigations of suspect cases	Number of herds with status officially free	Number of infected herds	Number of herds tested under surveillance	Number of animals tested under surveillance	Total number of herds	Number of infected herds tested under surveillance	Number of herds tested under surveillance by bulk milk	Number of animals or pools tested under surveillance by bulk milk	Number of infected herds tested under surveillance by bulk milk
GREECE (NUTS 2006)	4,234	5,730	199	4,085	68,125	21,941	221	4,085	2,163	45

Table Ovine or Caprine brucellosis - data on animals - Community co-financed eradication programmes

Region	Total number of animals	Number of animals to be tested under the program	Number of animals tested	Number of animals tested individually	Number of positive animals	Number of positive animals slaughtered	Total number of animals slaughtered
GREECE (NUTS 2006)	3,675,388	3,675,388	81,231	81,231	97	82	82

Table Ovine or Caprine brucellosis - data on herds - Community co-financed eradication programmes

Region	Number of new positive herds	Number of depopulated herds	Total number of herds	Number of herds under the program	Number of herds under the program tested/checked	Number of positive herds
GREECE (NUTS 2006)	5	0	16,646	16,646	1,263	5

Table Ovine or Caprine brucellosis - data on status of herds at the end of the period - Community co-financed eradication programmes

Region	Total number of herds under the program, at the end of the period	Total number of animals under the program, at the end of the period	Number of herds with unknown status, at the end of the period	Number of animals with unknown status, at the end of the period	Number of herds with status not free or not officially free and last check positive, at the end of the period	Number of animals with status not free or not officially free and last check positive, at the end of the period	Number of herds with status not free or not officially free and last check negative, at the end of the period	Number of animals with status not free or not officially free and last check negative, at the end of the period	Number of herds with status free or officially free suspended, at the end of the period	Number of animals with status free or officially free suspended, at the end of the period	Number of herds with status officially free, at the end of the period	Number of animals with status officially free, at the end of the period
GREECE (NUTS 2006)	16,646	3,675,388	15,002	3,271,767	21	9,745	1,222	305,508	72	18,398	329	69,970

DISEASE STATUS TABLES

Table Bovine tuberculosis in countries and regions that do not receive Community co-financing for eradication programme

Region	Number of herds with status officially free	Number of infected herds	Total number of animals	Interval between routine tuberculin tests	Number of animals tested with tuberculin routine testing	Number of animals with suspicious lesions of tuberculosis examined and submitted to histopathological and bacteriological examinations	Number of animals detected positive in bacteriological examination	Total number of herds
GREECE (NUTS 2006)	7,320	187	791,220	12	173,123	46	30	21,941

PREVALENCE TABLES

Table COXIELLA in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Total units tested	Total units positive	N of clinical affected herds	Zoonoses	N of units positive
GREECE (NUTS 2006)	Sheep and goats - Farm - Unknown - animal sample - blood - Clinical investigations - Official sampling - Suspect sampling	herd/flock	333	58	27	Coxiella burnetii	58

Table ECHINOCOCCUS in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
GREECE (NUTS 2006)	Cattle (bovine animals) - Slaughterhouse - Greece - animal sample - organ/tissue - Surveillance - Official sampling - Census	animal	100922	849	Echinococcus	849
	Goats - Slaughterhouse - Greece - animal sample - organ/tissue - Surveillance - Official sampling - Census	animal	505787	1661	Echinococcus	1,661
	Pigs - Slaughterhouse - Greece - animal sample - organ/tissue - Surveillance - Official sampling - Census	animal	747208	3	Echinococcus	3
	Sheep - Slaughterhouse - Greece - animal sample - organ/tissue - Surveillance - Official sampling - Census	animal	1872922	9662	Echinococcus	9,662
	Wild boars - farmed - Slaughterhouse - Greece - animal sample - organ/tissue - Surveillance - Official sampling - Census	animal	969	0	Echinococcus	0

Table FLAVIVIRUS in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
GREECE (NUTS 2006)	Solipeds, domestic - Farm - Greece - animal sample - blood - Monitoring - active - Official sampling - Selective sampling	animal	495	0	West Nile virus	0
	Solipeds, domestic - Farm - Greece - animal sample - blood - Unspecified - Official sampling - Not specified	animal	31	0	West Nile virus	0
Ξάνθη (NUTS 2006)	Solipeds, domestic - Farm - Greece - animal sample - blood - Monitoring - active - Official sampling - Selective sampling	animal	69	0	West Nile virus	0
Δράμα (NUTS 2006)	Solipeds, domestic - Farm - Greece - animal sample - blood - Monitoring - active - Official sampling - Selective sampling	animal	58	0	West Nile virus	0
Κιλκίς (NUTS 2006)	Solipeds, domestic - Farm - Greece - animal sample - blood - Monitoring - active - Official sampling - Selective sampling	animal	10	0	West Nile virus	0
Σέρρες (NUTS 2006)	Solipeds, domestic - Farm - Greece - animal sample - blood - Monitoring - active - Official sampling - Selective sampling	animal	78	0	West Nile virus	0
Χαλκιδική (NUTS 2006)	Solipeds, domestic - Farm - Greece - animal sample - blood - Monitoring - active - Official sampling - Selective sampling	animal	74	0	West Nile virus	0
Καστοριά (NUTS 2006)	Solipeds, domestic - Farm - Greece - animal sample - blood - Monitoring - active - Official sampling - Selective sampling	animal	1	0	West Nile virus	0
Φλώρινα (NUTS 2006)	Solipeds, domestic - Farm - Greece - animal sample - blood - Monitoring - active - Official sampling - Selective sampling	animal	8	0	West Nile virus	0
Καρδίτσα (NUTS 2006)	Solipeds, domestic - Farm - Greece - animal sample - blood - Monitoring - active - Official sampling - Selective sampling	animal	3	0	West Nile virus	0
Λάρισα (NUTS 2006)	Solipeds, domestic - Farm - Greece - animal sample - blood - Monitoring - active - Official sampling - Selective sampling	animal	23	0	West Nile virus	0
Ιωάννινα (NUTS 2006)	Solipeds, domestic - Farm - Greece - animal sample - blood - Monitoring - active - Official sampling - Selective sampling	animal	57	0	West Nile virus	0
Κέρκυρα (NUTS 2006)	Solipeds, domestic - Farm - Greece - animal sample - blood - Unspecified - Official sampling - Not specified	animal	1	0	West Nile virus	0
Εύβοια (NUTS 2006)	Solipeds, domestic - Farm - Greece - animal sample - blood - Monitoring - active - Official sampling - Selective sampling	animal	29	0	West Nile virus	0
	Solipeds, domestic - Farm - Greece - animal sample - blood - Unspecified - Official sampling - Not specified	animal	1	0	West Nile virus	0
ΑΤΤΙΚΗ (NUTS 2006)	Solipeds, domestic - Farm - Greece - animal sample - blood - Monitoring - active - Official sampling - Selective sampling	animal	85	0	West Nile virus	0
	Solipeds, domestic - Farm - Greece - animal sample - blood - Unspecified - Official sampling - Not specified	animal	29	0	West Nile virus	0

Table LISTERIA in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
GREECE (NUTS 2006)	Cattle (bovine animals) - dairy cows - Farm - Greece - animal sample - organ/tissue - Clinical investigations - Official sampling - Suspect sampling	animal	1	0	Listeria	0
	Goats - Farm - Greece - animal sample - organ/tissue - Clinical investigations - Official sampling - Suspect sampling	animal	20	13	Listeria monocytogenes	12
					Listeria spp., unspecified	1
	Sheep - Farm - Greece - animal sample - organ/tissue - Clinical investigations - Official sampling - Suspect sampling	animal	11	4	Listeria monocytogenes	4

Table LISTERIA in food

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
GREECE (NUTS 2006)	Cheeses made from cows' milk - hard - made from pasteurised milk - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	5	0	Not Available	Listeria monocytogenes	5	0
	Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	15	0	Not Available	Listeria monocytogenes	15	0
	Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	2	0	Not Available	Listeria monocytogenes	2	0
	Cheeses made from goats' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	25	Gram	8	0	Not Available	Listeria monocytogenes	8	0
		single (food/feed)	25	Gram	9	0	Not Available	Listeria monocytogenes	9	0
	Cheeses made from sheep's milk - hard - made from pasteurised milk - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	40	0	Not Available	Listeria monocytogenes	40	0
	Cheeses made from sheep's milk - soft and semi-soft - made from pasteurised milk - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	25	Gram	10	0	Not Available	Listeria monocytogenes	10	0
		single (food/feed)	25	Gram	20	0	Not Available	Listeria monocytogenes	20	0
	Cheeses made from sheep's milk - soft and semi-soft - made from pasteurised milk - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	24	2	Not Available	Listeria monocytogenes	24	2
	Cheeses, made from mixed milk from cows, sheep and/or goats - hard - made from pasteurised milk - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	35	0	Not Available	Listeria monocytogenes	35	0
	Cheeses, made from mixed milk from cows, sheep and/or goats - hard - made from pasteurised milk - Unspecified - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	15	0	Not Available	Listeria monocytogenes	15	0
	Cheeses, made from mixed milk from cows, sheep and/or goats - soft and semi-soft - made from pasteurised milk - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	35	0	Not Available	Listeria monocytogenes	35	0
	Dairy products (excluding cheeses) - dairy products, not specified - ready-to-eat - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	5	0	Not Available	Listeria monocytogenes	5	0
	Dairy products (excluding cheeses) - ice-cream - Unspecified - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	20	0	Not Available	Listeria monocytogenes	20	0
	Dairy products (excluding cheeses) - yoghurt - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	2	0	Not Available	Listeria monocytogenes	2	0
	Fish - raw - Border inspection activities - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	10	0	Not Available	Listeria monocytogenes	10	0
	Fish - raw - Hospital or medical care facility - Unknown - food sample - Surveillance - HACCP and own check - Objective sampling	single (food/feed)	25	Gram	20	0	Not Available	Listeria monocytogenes	20	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
GREECE (NUTS 2006)	Fish - raw - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	5	2	Not Available	Listeria monocytogenes	5	2
	Fish - smoked - Border inspection activities - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	25	Gram	1	0	Not Available	Listeria monocytogenes	1	0
	Fish - smoked - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	15	0	Not Available	Listeria monocytogenes	15	0
	Fish - smoked - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	25	Gram	30	0	Not Available	Listeria monocytogenes	30	0
	Infant formula - Hospital or medical care facility - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	5	0	Not Available	Listeria monocytogenes	5	0
	Meat from bovine animals - minced meat - intended to be eaten cooked - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	1	0	Not Available	Listeria monocytogenes	1	0
	Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	5	0	Not Available	Listeria monocytogenes	5	0
	Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	6	0	Not Available	Listeria monocytogenes	6	0
	Meat from pig - meat products - cooked, ready-to-eat - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	90	5	<= 100	Listeria monocytogenes	10	3
							>100	Listeria monocytogenes	10	0
	Meat from pig - meat products - cooked, ready-to-eat - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	90	5	Not Available	Listeria monocytogenes	80	2
	Meat from pig - meat products - cooked, ready-to-eat - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	80	8	Not Available	Listeria monocytogenes	80	8
	Milk, cows' - pasteurised milk - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Millilitre	5	0	Not Available	Listeria monocytogenes	5	0
	Milk, cows' - pasteurised milk - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Millilitre	2	0	Not Available	Listeria monocytogenes	2	0
	Snails - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	25	Gram	5	0	Not Available	Listeria monocytogenes	5	0
	Sweets - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	1	0	Not Available	Listeria monocytogenes	1	0
	Vegetables - pre-cut - ready-to-eat - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	20	0	Not Available	Listeria monocytogenes	20	0
	Vegetables - products - Processing plant - Unknown - food sample - Surveillance - HACCP and own check - Objective sampling	single (food/feed)	25	Gram	20	0	Not Available	Listeria monocytogenes	20	0

Table LYSSAVIRUS in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
GREECE (NUTS 2006)	All animals - farmed - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	8	0	Lyssavirus	0
	All animals - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	23	0	Lyssavirus	0
	Bats - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	6	0	Lyssavirus	0
	Cats - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	17	0	Lyssavirus	0
	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	53	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	142	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - Monitoring - active - Official sampling - Selective sampling	animal	87	0	Lyssavirus	0
	Rodents - laboratory animal - Unspecified - Greece - animal sample - brain - Survey - Official sampling - Selective sampling	animal	8	0	Lyssavirus	0
Έβρος (NUTS 2006)	All animals - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	2	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	5	0	Lyssavirus	0
Ξάνθη (NUTS 2006)	All animals - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
	Cats - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	2	0	Lyssavirus	0
	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	6	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	3	0	Lyssavirus	0
Ροδόπη (NUTS 2006)	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
Δράμα (NUTS 2006)	All animals - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	3	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	14	0	Lyssavirus	0
Καβάλα (NUTS 2006)	Cats - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	3	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - Monitoring - active - Official sampling - Selective sampling	animal	2	0	Lyssavirus	0
Ημαθία (NUTS 2006)	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
Θεσσαλονίκη (NUTS 2006)	All animals - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	2	0	Lyssavirus	0
	Bats - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	2	0	Lyssavirus	0
	Cats - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	2	0	Lyssavirus	0
	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	3	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	7	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - Monitoring - active - Official sampling - Selective sampling	animal	2	0	Lyssavirus	0
Κιλκίς (NUTS 2006)	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	2	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	11	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - Monitoring - active - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
Πέλλα (NUTS 2006)	All animals - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
	Cats - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	5	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	4	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - Monitoring - active - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
Πιερία (NUTS 2006)	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
Σέρρες (NUTS 2006)	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	7	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - Monitoring - active - Official sampling - Selective sampling	animal	39	0	Lyssavirus	0
Χαλκιδική (NUTS 2006)	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	3	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	2	0	Lyssavirus	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
Γρεβενά (NUTS 2006)	All animals - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	12	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - Monitoring - active - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
Καστοριά (NUTS 2006)	All animals - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	3	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	21	0	Lyssavirus	0
Κοζάνη (NUTS 2006)	All animals - farmed - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	4	0	Lyssavirus	0
	All animals - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	2	0	Lyssavirus	0
	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	4	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	20	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - Monitoring - active - Official sampling - Selective sampling	animal	3	0	Lyssavirus	0
Φλώρινα (NUTS 2006)	All animals - farmed - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	3	0	Lyssavirus	0
	All animals - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	5	0	Lyssavirus	0
	Cats - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	4	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	8	0	Lyssavirus	0
Καρδίτσα (NUTS 2006)	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	2	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	3	0	Lyssavirus	0
Λάρισα (NUTS 2006)	Cats - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	2	0	Lyssavirus	0
	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
Μαγνησία (NUTS 2006)	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	3	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - Monitoring - active - Official sampling - Selective sampling	animal	14	0	Lyssavirus	0
Τρίκαλα (NUTS 2006)	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	3	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - Monitoring - active - Official sampling - Selective sampling	animal	21	0	Lyssavirus	0
Άρτα (NUTS 2006)	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - Monitoring - active - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
Θεσπρωτία (NUTS 2006)	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
Ιωάννινα (NUTS 2006)	All animals - farmed - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
	All animals - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	5	0	Lyssavirus	0
	Bats - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
	Cats - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	4	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	6	0	Lyssavirus	0
Ζάκυνθος (NUTS 2006)	Bats - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
Λευκάδα (NUTS 2006)	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
Αιτωλοακαρνανία (NUTS 2006)	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - Monitoring - active - Official sampling - Selective sampling	animal	2	0	Lyssavirus	0
Ηλεία (NUTS 2006)	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	3	0	Lyssavirus	0
Βοιωτία (NUTS 2006)	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
Εύβοια (NUTS 2006)	Cats - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
Φωκίδα (NUTS 2006)	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	3	0	Lyssavirus	0
Κορινθία (NUTS 2006)	Cats - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
Λακωνία (NUTS 2006)	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	2	0	Lyssavirus	0
Μεσσηνία (NUTS 2006)	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0
ΑΤΤΙΚΗ (NUTS 2006)	Bats - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	2	0	Lyssavirus	0
	Cats - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	2	0	Lyssavirus	0
	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	3	0	Lyssavirus	0
	Foxes - wild - Natural habitat - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	3	0	Lyssavirus	0
Attiki (NUTS 2006 level 2)	Rodents - laboratory animal - Unspecified - Greece - animal sample - brain - Survey - Official sampling - Selective sampling	animal	8	0	Lyssavirus	0
Λασίθι (NUTS 2006)	Dogs - pet animals - Unspecified - Greece - animal sample - brain - Monitoring - passive - Official sampling - Selective sampling	animal	1	0	Lyssavirus	0

Table SALMONELLA in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	N of flocks under control programme	Target verification	Total units tested	Total units positive	Zoonoses	N of units positive
GREECE (NUTS 2006)	Cattle (bovine animals) - adult cattle over 2 years - Farm - Greece - animal sample - organ/tissue - Clinical investigations - Official sampling - Suspect sampling	animal		N_A	1	0	Salmonella	0
	Cattle (bovine animals) - adult cattle over 2 years - Farm - Greece - food sample - milk - Clinical investigations - Official sampling - Suspect sampling	animal		N_A	1	1	Salmonella Dublin	1
	Cattle (bovine animals) - calves (under 1 year) - Farm - Greece - animal sample - organ/tissue - Clinical investigations - Official sampling - Suspect sampling	animal		N_A	3	0	Salmonella	0
	Cattle (bovine animals) - Slaughterhouse - Unknown - animal sample - Surveillance - Official sampling - Objective sampling	animal		N_A	15	0	Salmonella	0
	Gallus gallus (fowl) - breeding flocks, unspecified - adult - Farm - Greece - animal sample - Control and eradication programmes - Official and industry sampling - Census	herd/flock	281	Y	280	4	Salmonella Bredeney	1
							Salmonella Enteritidis	1
							Salmonella Typhimurium	1
							Salmonella Winston	1
	Gallus gallus (fowl) - breeding flocks, unspecified - day-old chicks - Farm - Greece - animal sample - Control and eradication programmes - Industry sampling - Census	herd/flock	145	N_A	143	0	Salmonella	0
	Gallus gallus (fowl) - breeding flocks, unspecified - during rearing period - Farm - Greece - animal sample - Control and eradication programmes - Industry sampling - Census	herd/flock	191	N_A	190	0	Salmonella	0
	Gallus gallus (fowl) - broilers - before slaughter - Farm - Greece - animal sample - Control and eradication programmes - Official and industry sampling - Census	herd/flock	6871	Y	6824	27	Salmonella Enteritidis	1
							Salmonella Haardt	2
							Salmonella Hadar	1
							Salmonella Infantis	2
							Salmonella Kedougou	2
							Salmonella Livingstone	11
							Salmonella Muenster	1
							Salmonella Szentes	1
							Salmonella Tennessee	3
							Salmonella Thompson	2
							Salmonella Typhimurium	1
	Gallus gallus (fowl) - Farm - Greece - animal sample - organ/tissue - Clinical investigations - Official sampling - Suspect sampling	animal		N_A	19	1	Salmonella Gallinarum	1
	Gallus gallus (fowl) - laying hens - adult - Farm - Greece - animal sample - Control and eradication programmes - Official and industry sampling - Census	herd/flock	723	Y	476	18	Salmonella Braenderup	2
							Salmonella Dabou	1
							Salmonella Enteritidis	1
							Salmonella Hadar	3
							Salmonella Infantis	2
							Salmonella Montevideo	1
							Salmonella Rissen	1
							Salmonella Szentes	1
							Salmonella Thompson	3
							Salmonella Typhimurium	1
							Salmonella Umbilo	1
							Salmonella Yoruba	1
	Gallus gallus (fowl) - laying hens - day-old chicks - Farm - Greece - animal sample - Control and eradication programmes - Industry sampling - Census	herd/flock	56	N_A	51	0	Salmonella	0
	Gallus gallus (fowl) - laying hens - during rearing period - Farm - Greece - animal sample - Control and eradication programmes - Industry sampling - Census	herd/flock	148	N_A	75	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	N of flocks under control programme	Target verification	Total units tested	Total units positive	Zoonoses	N of units positive
GREECE (NUTS 2006)	Goats - Farm - Greece - animal sample - organ/tissue - Clinical investigations - Official sampling - Suspect sampling	animal		N_A	12	0	Salmonella	0
	Pigs - breeding animals - unspecified - Farm - Greece - animal sample - organ/tissue - Clinical investigations - Official sampling - Suspect sampling	animal		N_A	3	0	Salmonella	0
	Pigs - fattening pigs - Farm - Greece - animal sample - organ/tissue - Clinical investigations - Official sampling - Suspect sampling	animal		N_A	3	0	Salmonella	0
	Pigs - Slaughterhouse - Unknown - animal sample - Surveillance - Official sampling - Objective sampling	animal		N_A	15	0	Salmonella	0
	Quails - Farm - Greece - animal sample - organ/tissue - Clinical investigations - Official sampling - Suspect sampling	animal		N_A	2	0	Salmonella	0
	Sheep - Farm - Greece - animal sample - organ/tissue - Clinical investigations - Official sampling - Suspect sampling	animal		N_A	4	0	Salmonella	0
	Sheep - Slaughterhouse - Unknown - animal sample - Surveillance - Official sampling - Objective sampling	animal		N_A	25	0	Salmonella	0
	Turkeys - fattening flocks - before slaughter - Farm - Greece - animal sample - Control and eradication programmes - Official and industry sampling - Census	herd/flock	77	Y	73	2	Salmonella Typhimurium	2
	Turkeys - parent breeding flocks - adult - Farm - Greece - animal sample - Control and eradication programmes - Official and industry sampling - Census	herd/flock	4	Y	3	0	Salmonella	0
	Turkeys - parent breeding flocks - day-old chicks - Farm - Greece - animal sample - Control and eradication programmes - Industry sampling - Census	herd/flock	2	N_A	1	0	Salmonella	0
	Turkeys - parent breeding flocks - during rearing period - Farm - Greece - animal sample - Control and eradication programmes - Industry sampling - Census	herd/flock	2	N_A	1	0	Salmonella	0

Table SALMONELLA in food

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
GREECE (NUTS 2006)	Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	15	0	Salmonella	0
	Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	2	0	Salmonella	0
	Cheeses made from goats' milk - soft and semi-soft - made from pasteurised milk - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	20	0	Salmonella	0
	Cheeses made from goats' milk - soft and semi-soft - made from pasteurised milk - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	5	0	Salmonella	0
	Cheeses made from sheep's milk - soft and semi-soft - made from pasteurised milk - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	80	0	Salmonella	0
	Cheeses made from sheep's milk - soft and semi-soft - made from pasteurised milk - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	18	0	Salmonella	0
	Coconut - Border inspection activities - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	25	Gram	2	0	Salmonella	0
	Crustaceans - unspecified - raw - Border inspection activities - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	25	0	Salmonella	0
	Dairy products (excluding cheeses) - ice-cream - Border inspection activities - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	10	0	Salmonella	0
	Dairy products (excluding cheeses) - ice-cream - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	20	0	Salmonella	0
	Dairy products (excluding cheeses) - yoghurt - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	25	0	Salmonella	0
	Dairy products (excluding cheeses) - yoghurt - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	1	0	Salmonella	0
	Dairy products, unspecified - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	15	0	Salmonella	0
	Egg products - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	5	0	Salmonella	0
	Eggs - table eggs - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	25	Gram	2	0	Salmonella	0
	Fish - marinated - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	5	0	Salmonella	0
	Fish - raw - Border inspection activities - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	1	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
GREECE (NUTS 2006)	Fish - raw - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	35	0	Salmonella	0
	Fish - raw - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	4	0	Salmonella	0
	Fish (food) - Unspecified - Unknown - food sample - Surveillance - Official sampling - Suspect sampling	single (food/feed)	25	Gram	1	0	Salmonella	0
	Fruits and vegetables - pre-cut - ready-to-eat - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	25	Gram	20	0	Salmonella	0
	Infant formula - Hospital or medical care facility - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Millilitre	5	0	Salmonella	0
	Live bivalve molluscs - Farm - Greece - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	703	1	Salmonella Infantis	1
	Live bivalve molluscs - Retail - Greece - food sample - Unspecified - Official sampling - Suspect sampling	batch (food/feed)	25	Gram	1	0	Salmonella	0
	Meat from bovine animals - fresh - Hospital or medical care facility - Unknown - food sample - Surveillance - HACCP and own check - Objective sampling	single (food/feed)	25	Gram	22	0	Salmonella	0
	Meat from bovine animals - fresh - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	25	Gram	5	0	Salmonella	0
		single (food/feed)	25	Gram	1	0	Salmonella	0
	Meat from bovine animals - meat preparation - intended to be eaten cooked - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	10	Gram	10	1	Salmonella	1
	Meat from bovine animals - minced meat - intended to be eaten cooked - Hospital or medical care facility - Unknown - food sample - Surveillance - HACCP and own check - Objective sampling	single (food/feed)	25	Gram	21	0	Salmonella	0
	Meat from bovine animals - minced meat - intended to be eaten cooked - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	10	Gram	20	0	Salmonella	0
	Meat from bovine animals - minced meat - intended to be eaten cooked - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	10	Gram	15	0	Salmonella	0
		single (food/feed)	10	Gram	7	0	Salmonella	0
	Meat from broilers (Gallus gallus) - carcase - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	9	2	Salmonella	2
					115	3	Salmonella Enteritidis	2
							Salmonella Livingstone	1
	Meat from broilers (Gallus gallus) - carcase - Slaughterhouse - Greece - food sample - neck skin - Surveillance - Official sampling - Selective sampling	single (food/feed)	25	Gram	99	31	Salmonella Corvallis	2
							Salmonella Enteritidis	3
							Salmonella Livingstone	25
							Salmonella Not typeable	1
	Meat from broilers (Gallus gallus) - carcase - Slaughterhouse - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	10	1	Salmonella Livingstone	1

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
GREECE (NUTS 2006)	Meat from broilers (Gallus gallus) - fresh - frozen - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	20	7	Salmonella Infantis	3
							Salmonella Typhimurium	4
	Meat from broilers (Gallus gallus) - fresh - Hospital or medical care facility - Unknown - food sample - Surveillance - HACCP and own check - Objective sampling	single (food/feed)	25	Gram	35	0	Salmonella	0
	Meat from broilers (Gallus gallus) - fresh - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	10	0	Salmonella	0
	Meat from broilers (Gallus gallus) - fresh - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	25	Gram	10	0	Salmonella	0
	Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - Border inspection activities - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	40	0	Salmonella	0
	Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	55	4	Salmonella Enteritidis	2
							Salmonella Infantis	2
	Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	240	15	Salmonella Infantis	6
							Salmonella spp., unspecified	9
	Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	5	0	Salmonella	0
	Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	10	0	Salmonella	0
	Meat from broilers (Gallus gallus) - meat products - raw but intended to be eaten cooked - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	60	0	Salmonella	0
	Meat from broilers (Gallus gallus) - meat products - raw but intended to be eaten cooked - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	3	0	Salmonella	0
	Meat from broilers (Gallus gallus) - mechanically separated meat (MSM) - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	10	0	Salmonella	0
	Meat from broilers (Gallus gallus) - minced meat - intended to be eaten cooked - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	5	5	Salmonella Thompson	5
	Meat from broilers (Gallus gallus) - minced meat - intended to be eaten cooked - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	10	Gram	5	0	Salmonella	0
	Meat from pig - fresh - Hospital or medical care facility - Unknown - food sample - Surveillance - HACCP and own check - Objective sampling	single (food/feed)	25	Gram	39	0	Salmonella	0
	Meat from pig - fresh - Slaughterhouse - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	10	Gram	45	0	Salmonella	0
	Meat from pig - meat preparation - intended to be eaten cooked - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	10	Gram	5	0	Salmonella	0
			25	Gram	5	0	Salmonella	0
	Meat from pig - meat preparation - intended to be eaten cooked - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	10	Gram	63	0	Salmonella	0

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
GREECE (NUTS 2006)	Meat from pig - meat products - cooked, ready-to-eat - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	10	Gram	75	0	Salmonella	0
			25	Gram	20	0	Salmonella	0
	Meat from pig - meat products - cooked, ready-to-eat - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	86	0	Salmonella	0
	Meat from pig - meat products - raw but intended to be eaten cooked - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	10	Gram	55	0	Salmonella	0
	Meat from pig - meat products - raw but intended to be eaten cooked - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	6	0	Salmonella	0
	Meat from pig - minced meat - intended to be eaten cooked - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	10	Gram	25	0	Salmonella	0
	Meat from pig - minced meat - intended to be eaten cooked - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	10	Gram	15	0	Salmonella	0
	Meat from turkey - fresh - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	25	Gram	5	0	Salmonella	0
	Meat from turkey - meat preparation - intended to be eaten cooked - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	10	Gram	5	0	Salmonella	0
	Meat from turkey - meat products - cooked, ready-to-eat - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	10	0	Salmonella	0
	Meat, mixed meat - minced meat - intended to be eaten cooked - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	10	Gram	5	0	Salmonella	0
	Milk, cows' - pasteurised milk - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Millilitre	20	0	Salmonella	0
	Milk, cows' - pasteurised milk - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Millilitre	2	0	Salmonella	0
	Milk, goats' - raw milk for manufacture - intended for manufacture of pasteurised/UHT products - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Millilitre	1	0	Salmonella	0
	Molluscan shellfish - cooked - Border inspection activities - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	10	0	Salmonella	0
	Other processed food products and prepared dishes - unspecified - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	1	0	Salmonella	0
	Seeds, dried - Border inspection activities - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	25	Gram	93	19	Salmonella Agona	2
							Salmonella Amsterdam	1
							Salmonella Bergen	1
							Salmonella Bredeney	1
							Salmonella Dallgow	2
							Salmonella Ekotedo	1
							Salmonella enterica, subspecies enterica	1
							Salmonella Hongkong	1

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
GREECE (NUTS 2006)	Seeds, dried - Border inspection activities - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	25	Gram	93	19	Salmonella Kristianstad	1
							Salmonella Mbandaka	2
							Salmonella Montevideo	2
							Salmonella Orion	3
							Salmonella Ruiru	1
							Salmonella Senftenberg	2
							Salmonella spp., unspecified	1
							Salmonella Tennessee	1
	Spices and herbs - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	25	Gram	2	0	Salmonella	0
	Vegetables - Processing plant - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	20	0	Salmonella	0

Table SALMONELLA in feed

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
GREECE (NUTS 2006)	Compound feedingstuffs for cattle - final product - Border inspection activities - Unknown - feed sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	6	0	Salmonella	0
	Compound feedingstuffs for fish - Unspecified - Unknown - feed sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	20	0	Salmonella	0
	Feed material of land animal origin - poultry offal meal - Processing plant - Unknown - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	25	Gram	25	1	Salmonella Thompson	1
		single (food/feed)	25	Gram	30	0	Salmonella	0
	Feed material of marine animal origin - fish meal - Processing plant - Unknown - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	25	Gram	100	6	Salmonella Mbandaka	3
							Salmonella Muenster	1
							Salmonella spp., unspecified	2
	Feed material of marine animal origin - fish oil - Processing plant - Unknown - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	25	Gram	40	0	Salmonella	0
	Pet food - dog snacks (pig ears, chewing bones) - Processing plant - Unknown - feed sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	250	0	Salmonella	0
	Pet food - Processing plant - Unknown - feed sample - Surveillance - Official sampling - Objective sampling	batch (food/feed)	25	Gram	45	3	Salmonella Thompson	3

Table STAPHYLOCOCCAL ENTEROTOXINS in food

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
GREECE (NUTS 2006)	Cheeses made from cows' milk - soft and semi-soft - made from pasteurised milk - Retail - Unknown - food sample - Surveillance - Official sampling - Objective sampling	single (food/feed)	25	Gram	10	0	Staphylococcal enterotoxins	0

Table TOXOPLASMA in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
GREECE (NUTS 2006)	Goats - Farm - Greece - animal sample - blood - Clinical investigations - Official sampling - Suspect sampling	animal	58	31	Toxoplasma gondii	31
	Sheep - Farm - Greece - animal sample - blood - Clinical investigations - Official sampling - Suspect sampling	animal	46	9	Toxoplasma gondii	9
	Wild boars - Farm - Greece - animal sample - blood - Clinical investigations - Official sampling - Suspect sampling	animal	49	27	Toxoplasma gondii	27

Table TRICHINELLA in animal

Area of Sampling	Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
GREECE (NUTS 2006)	Other animals - Slaughterhouse - Greece - animal sample - organ/tissue - Surveillance - Official sampling - Census	animal	815	0	Trichinella	0
	Pigs - breeding animals - raised under controlled housing conditions - boars - Slaughterhouse - Greece - animal sample - organ/tissue - Surveillance - Official sampling - Census	animal	467	0	Trichinella	0
	Pigs - breeding animals - raised under controlled housing conditions - sows - Slaughterhouse - Greece - animal sample - organ/tissue - Surveillance - Official sampling - Census	animal	23111	0	Trichinella	0
	Pigs - fattening pigs - not raised under controlled housing conditions - Slaughterhouse - Greece - animal sample - organ/tissue - Surveillance - Official sampling - Census	animal	4685	0	Trichinella	0
	Pigs - fattening pigs - raised under controlled housing conditions - Slaughterhouse - Greece - animal sample - organ/tissue - Surveillance - Official sampling - Census	animal	1089787	0	Trichinella	0
	Wild boars - farmed - Slaughterhouse - Greece - animal sample - organ/tissue - Surveillance - Official sampling - Census	animal	1619	0	Trichinella	0
	Wild boars - wild - Slaughterhouse - Greece - animal sample - organ/tissue - Surveillance - Official sampling - Census	animal	95	0	Trichinella	0

FOODBORNE OUTBREAKS TABLES

Foodborne Outbreaks: summarized data

		Outbreak strenght	Strong			Weak			
Causative agent	Food vehicle	N outbreaks	N human cases	N hospitalized	N deaths	N outbreaks	N human cases	N hospitalized	N deaths
Hepatitis virus	Crustaceans, shellfish, molluscs and products thereof					1	3	3	0
Listeria monocytogenes - serovar 1/2a	Unknown					1	2	2	2
Norovirus	Tap water, including well water	1	230	0	0	1	23	0	0
Salmonella Enteritidis	Cheese					1	3	2	0
	Other, mixed or unspecified poultry meat and products thereof	1	5	3	0				
Unknown	Tap water, including well water	1	213	0	0				
	Other foods					1	16	3	0
	Buffet meals	1	30	0	0				

Strong Foodborne Outbreaks: detailed data

Causative agent	Other Causative Agent	FBO nat. code	Outbreak type	Food vehicle	More food vehicle info	Nature of evidence	Setting	Place of origin of problem	Origin of food vehicle	Contributory factors	Comment	N outbreaks	N human cases	N hosp.	N deaths
Norovirus	unknown	N_A	General	Tap water, including well water	N_A	Analytical epidemiological evidence	Others	Water distribution system	Not Available	Other contributory factor	Community waterborne outbreak	1	230	0	0
Salmonella Enteritidis	unknown	N_A	Not Available	Other, mixed or unspecified poultry meat and products thereof	Chicken used in club sandwich and caesar salad	Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Not Available	Infected food handler	One case was food handler	1	5	3	0
Unknown	unknown	N_A	Not Available	Tap water, including well water	Water supply from a private well	Descriptive epidemiological evidence	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Water source	Not Available	Other contributory factor	N_A	1	213	0	0
			General	Buffet meals	N_A	Analytical epidemiological evidence	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Not Available	Unknown	N_A	1	30	0	0

Weak Foodborne Outbreaks: detailed data

Causative agent	Other Causative Agent	FBO nat. code	Outbreak type	Food vehicle	More food vehicle info	Nature of evidence	Setting	Place of origin of problem	Origin of food vehicle	Contributory factors	Comment	N outbreaks	N human cases	N hosp.	N deaths
Hepatitis virus	unknown	N_A	Not Available	Crustaceans, shellfish, molluscs and products thereof	N_A	Descriptive epidemiological evidence	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Not Available	Unknown	N_A	1	3	3	0
Listeria monocytogenes - serovar 1/2a	unknown	N_A	Not Available	Unknown	N_A	Descriptive epidemiological evidence	Not Available	Hospital or medical care facility	Not Available	NOT AVAILABLE	N_A	1	2	2	2
Norovirus	unknown	N_A	General	Tap water, including well water	N_A	Descriptive epidemiological evidence	Others	Water distribution system	Not Available	Untreated drinking water	Community waterborne oubreak	1	23	0	0
Salmonella Enteritidis	unknown	N_A	Not Available	Cheese	N_A	Descriptive epidemiological evidence	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Not Available	Unknown	N_A	1	3	2	0
Unknown	unknown	N_A	Not Available	Other foods	Spaghetti traditionally cooked	Descriptive epidemiological evidence	Others	Others	Not Available	Unknown	N_A	1	16	3	0

ANTIMICROBIAL RESISTANCE TABLES FOR CAMPYLOBACTER

ANTIMICROBIAL RESISTANCE TABLES FOR SALMONELLA

Table Antimicrobial susceptibility testing of Salmonella 6,7:-:- in Meat from broilers (Gallus gallus) - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - passive

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.5							1								
8			1												
16											1				
64												1			
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella 6,7:-:- in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
8			1												
64												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella 6,7:-:- in Meat from pig - meat preparation

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**1**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**1**	**1**	**1**	**1**	**1**
0.25							1								
2														1	
8			1												
>32															1
>64		1											1		
>128											1				
>1024												1			
<=0.03										1					
<=0.25				1											
<=0.5					1				1						
<=1								1							
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Braenderup in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
8			1												
128												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Braenderup in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: environmental sample - dust

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**
0.12							1								
1									1						
4			1												
8											1				
64												1			
>64													1		
<=0.03										1					
<=0.25				1										1	1
<=0.5					1										
<=1		1						1							
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Bredeney in Geese - breeding flocks, unspecified - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**1**	**1**	**1**	**0**	**0**
0.5							1								
16			1												
32											1				
>64													1		
>1024												1			
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Corvallis in Meat from broilers (Gallus gallus) - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - passive

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
4			1												
256												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Dabou in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
256												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2			1										1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Meat from broilers (Gallus gallus) - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - active

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
8			1												
128												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.03							1								
8			1												
256												1			
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Meat from broilers (Gallus gallus) - meat preparation

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
8			1												
128												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
64												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2			1										1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Geese - breeding flocks, unspecified - adult

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
8			1												
256												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: environmental sample - dust

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**
0.03							1								
0.25							1								
4			2												
8			1												
64												1			
128												2			
>128											1				
<=0.015							1								
<=0.03										3					
<=0.25				3										3	3
<=0.5					3				3						
<=1		3						3							
<=2													3		
<=4											2				
<=8						3									

Table Antimicrobial susceptibility testing of Salmonella Enteritidis in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: environmental sample - dust

Sampling Context: Control and eradication programmes

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**
0.5							1								
128												1			
>128											1				
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2			1										1		
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Haardt in Meat from broilers (Gallus gallus) - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - passive

Sampler: HACCP and own check

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.03							1								
0.5															1
64			1												
256												1			
<=0.03										1					
<=0.25				1										1	
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Haardt in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.03							1								
64			1									1			
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Haardt in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
32												1			
64			1												
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Hadar in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: environmental sample - dust

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**2**	**0**	**0**	**0**	**2**	**0**	**2**	**0**	**0**
0.12							1								
0.25							1								
32												1			
64												1	2		
>128											2				
<=0.03										2					
<=0.25				2										2	2
<=0.5					2				2						
<=1		2						2							
<=2			2												
<=8						2									

Table Antimicrobial susceptibility testing of Salmonella Hadar in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**1**	**0**	**1**	**0**	**0**
0.25							1								
32												1	1		
128											1				
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2			1												
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella II 42:b:e,n,x,z15 in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
8			1												
32												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Cutting plant

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: HACCP and own check

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**1**	**1**	**1**	**0**	**0**
1							1							1	
2		1													
16			1												
>64													1		
>128											1				
>1024												1			
<=0.03										1					
<=0.25				1											1
<=0.5					1				1						
<=1								1							
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**1**	**1**	**1**	**0**	**0**
1							1								
2		1													
16			1												
>64													1		
>128											1				
>1024												1			
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1								1							
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from broilers (Gallus gallus) - meat preparation

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: HACCP and own check

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**1**	**1**	**1**	**0**	**1**
0.5				1											
1					1		1							1	
2		1													
16			1												
>32															1
>64													1		
>128											1				
>1024												1			
<=0.03										1					
<=0.5									1						
<=1								1							
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from broilers (Gallus gallus) - meat preparation

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**1**	**1**	**1**	**0**	**1**
0.5							1								
8			1												
>32															1
>64													1		
>128											1				
>1024												1			
<=0.03										1					
<=0.25				1										1	
<=0.5					1				1						
<=1		1						1							
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Infantis in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**2**	**0**	**0**	**0**	**2**	**2**	**2**	**0**	**0**
0.5				1			1								
1					1		1							1	
4		1													
16			2												
>64													2		
>128											2				
1024												1			
>1024												1			
<=0.03										2					
<=0.25				1										1	2
<=0.5					1				2						
<=1		1						2							
<=8						2									

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from broilers (Gallus gallus) - meat preparation - intended to be eaten cooked

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Netherlands

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**1**	**1**	**1**	**0**	**0**
1							1								
8			1												
>64													1		
>128											1				
>1024												1			
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from broilers (Gallus gallus) - carcase - frozen

Sampling Stage: Unspecified

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: HACCP and own check

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**1**	**0**	**0**	**0**	**1**	**1**	**0**	**0**	**0**	**1**	**1**	**1**	**0**	**0**
0.5							1								
64						1									
>64		1											1		
>128											1				
>1024												1			
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1								1							
<=2			1												

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from broilers (Gallus gallus) - carcase - frozen

Sampling Stage: Conservation facilities

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Romania

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**2**	**0**	**0**	**0**	**2**	**2**	**2**	**0**	**2**
1							2							2	
2		2													
16			2												
>32															2
>64													2		
>128											2				
>1024												2			
<=0.03										2					
<=0.25				2											
<=0.5					2				2						
<=1								2							
<=8						2									

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from broilers (Gallus gallus) - carcase - frozen

Sampling Stage: Conservation facilities

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**1**	**1**	**1**	**0**	**1**
1							1							1	
2		1													
16			1												
>32															1
>64													1		
>128											1				
>1024												1			
<=0.03										1					
<=0.25				1											
<=0.5					1				1						
<=1								1							
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Infantis in Meat from bovine animals and pig - meat preparation

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**1**	**0**	**0**	**0**	**1**	**1**	**0**	**0**	**0**	**1**	**1**	**1**	**0**	**1**
1							1							1	
16			1												
>32															1
>64		1											1		
>128						1					1				
>1024												1			
<=0.03										1					
<=0.25				1											
<=0.5					1				1						
<=1								1							

Table Antimicrobial susceptibility testing of Salmonella Infantis in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: environmental sample - dust

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
8			1												
32												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Infantis in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.03							1								
1									1						
8			1												
128												1			
<=0.03										1					
<=0.25				1										1	1
<=0.5					1										
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Kedougou in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.03							1								
4			1												
8			1												
16												1			
64												1			
<=0.015							1								
<=0.03										2					
<=0.25				2										2	2
<=0.5					2				2						
<=1		2						2							
<=2													2		
<=4											2				
<=8						2									

Table Antimicrobial susceptibility testing of Salmonella Livingstone in Meat from broilers (Gallus gallus) - carcase

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - passive

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**7**	**0**	**0**	**0**	**2**	**0**	**0**	**0**	**0**
0.5							7								
8			7												
16											5				
32											2	3			
64												4			
<=0.03										7					
<=0.25				7										7	7
<=0.5					7				7						
<=1		7						7							
<=2													7		
<=8						7									

Table Antimicrobial susceptibility testing of Salmonella Livingstone in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: HACCP and own check

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.5							1								
8			1												
16											1				
64												1			
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Livingstone in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**3**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.5							3								
8			3												
16											3				
32												1			
64												2			
<=0.03										3					
<=0.25				3										3	3
<=0.5					3				3						
<=1		3						3							
<=2													3		
<=8						3									

Table Antimicrobial susceptibility testing of Salmonella Livingstone in Meat from broilers (Gallus gallus) - meat preparation

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.5							1								
8			1												
16											1				
32												1			
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Livingstone in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**5**	**5**	**5**	**5**	**5**	**5**	**5**	**5**	**5**	**5**	**5**	**5**	**5**	**5**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**5**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.5							5								
8			4												
16			1								4				
64												4			
128												1			
<=0.03										5					
<=0.25				5										5	5
<=0.5					5				5						
<=1		5						5							
<=2													5		
<=4											1				
<=8						5									

Table Antimicrobial susceptibility testing of Salmonella Montevideo in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.5							1								
8			1												
16											1	1			
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Muenster in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
2		1													
64												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1								1							
<=2			1										1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Ohio in Meat from bovine animals - minced meat

Sampling Stage: Conservation facilities

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
4			1												
128												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Rissen in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**1**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**
>64		1													
128						1									
>1024												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1								1							
<=2			1										1		
<=4											1				

Table Antimicrobial susceptibility testing of Salmonella Szentes in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
8			1												
64												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Tennessee in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**	**3**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.06							2								
0.5															1
2		2													
4			1										1		
8											1				
16			2												
32												1			
128												2			
<=0.015							1								
<=0.03										3					
<=0.25				3										3	2
<=0.5					3				3						
<=1		1						3							
<=2													2		
<=4											2				
<=8						3									

Table Antimicrobial susceptibility testing of Salmonella Thompson in Meat from broilers (Gallus gallus) - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.5							1								
8			1												
16											1				
64												1			
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Thompson in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.5							1								
8			1												
16			1								1				
64												1			
128												1			
<=0.015							1								
<=0.03										2					
<=0.25				2										2	2
<=0.5					2				2						
<=1		2						2							
<=2													2		
<=4											1				
<=8						2									

Table Antimicrobial susceptibility testing of Salmonella Thompson in Meat from broilers (Gallus gallus) - minced meat

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.5							1								
8			1												
16											1				
256												1			
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Thompson in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: environmental sample - dust

Sampling Context: Control and eradication programmes

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**	**2**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.03							2								
4			2												
32												2			
<=0.03										2					
<=0.25				2										2	2
<=0.5					2				2						
<=1		2						2							
<=2													2		
<=4											2				
<=8						2									

Table Antimicrobial susceptibility testing of Salmonella Thompson in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: HACCP and own check

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.03							1								
8			1												
64												1			
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Thompson in Meat from broilers (Gallus gallus) - offal - liver

Sampling Stage: Processing plant

Sampling Type: food sample

Sampling Context: Surveillance

Sampler: HACCP and own check

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**
0.5															1
1							1							1	
2		1													
16			1												
32											1				
128												1			
<=0.03										1					
<=0.25				1											
<=0.5					1				1						
<=1								1							
<=2													1		
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Thompson in Meat from broilers (Gallus gallus) - meat products - cooked, ready-to-eat - chilled

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.5							1								
8			1												
16											1				
64												1			
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from bovine animals and pig - meat products

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**1**	**0**	**0**
0.03							1								
8			1												
64												1			
>64													1		
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**1**	**0**	**0**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**1**	**1**	**0**	**0**
4			1												
>64		1											1		
>128						1									
>1024												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1								1							
<=4											1				

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from broilers (Gallus gallus) - carcase - frozen

Sampling Stage: Conservation facilities

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Netherlands

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**1**	**0**	**0**	**0**	**1**	**1**	**0**	**0**	**0**	**1**	**1**	**1**	**0**	**1**
0.5							1								
1														1	
8			1												
>32															1
64						1									
>64		1											1		
>128											1				
>1024												1			
<=0.03										1					
<=0.25				1											
<=0.5					1				1						
<=1								1							

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Meat from pig - meat preparation

Sampling Stage: Conservation facilities

Sampling Type: food sample - meat

Sampling Context: Surveillance

Sampler: Official sampling

Sampling Strategy: Census

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**1**	**1**	**0**	**0**	**1**	**0**	**0**	**0**	**0**	**0**	**1**	**1**	**0**	**1**
0.06							1								
1														1	
8											1				
>32															1
64			1												
>64		1											1		
>128						1									
>1024												1			
<=0.03										1					
<=0.25				1											
<=0.5					1				1						
<=1								1							

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Geese - breeding flocks, unspecified - adult

Sampling Stage: Farm

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
1									1						
4			1												
64												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1										
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Typhimurium in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: environmental sample - dust

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
4			1												
32												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Umbilo in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
4			1												
64												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Winston in Geese - breeding flocks, unspecified - adult

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
0.03							1								
8			1												
64												1			
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

Table Antimicrobial susceptibility testing of Salmonella Yoruba in Gallus gallus (fowl) - laying hens

Sampling Stage: Farm

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.5**	**2**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**256**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**	**1**
MIC	**N of resistant isolates**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**	**0**
4			1												
16												1			
<=0.015							1								
<=0.03										1					
<=0.25				1										1	1
<=0.5					1				1						
<=1		1						1							
<=2													1		
<=4											1				
<=8						1									

ANTIMICROBIAL RESISTANCE TABLES FOR INDICATOR ESCHERICHIA COLI

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from bovine animals - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Denmark

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.06	0.25	0.06	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	128
	N of tested isolates	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	1	1	1	1	1	0	0	0	0
0.03								1			
0.25		1							1		
4				1							
8			1			1	1				1
64					1						
<=0.03										1	

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from bovine animals - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method: Dilution - sensititre

Country of Origin: Denmark

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
MIC	N of resistant isolates	1	0	1	1	0	1	0	1	0	1	1	1	0	0
0.5							1								
>4				1											
8			1		1										
16									1						
>64		1											1		
128											1				
>1024												1			
<=0.03										1					
<=0.25														1	1
<=1								1							
<=8						1									

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from bovine animals - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid		Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid		Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Not Available	Positive/Present	Not Available	Not Available	Not Available		Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Positive/Present	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	0.25	8	0.5	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.06	0.25	0.06	0.06	0.5	0.25	0.12	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	64	128	128	128	2	16	16	128
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	1	2	1	1	1	2	1	1	0	0	0	0
0.03										2			
0.12		1											
2				1			1						
4			1				1	1					
8						1							2
>32		1											
>64			1			1							
<=0.03												2	
<=0.06					1								
<=0.12									1		2		

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from bovine animals - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	2	0	2	2	0	2	0	0	0	2	2	2	0	1
0.25							1								
2					1				1						
4				1											
>4				1											
8			1		1										
>8							1								
>32															1
>64		2											2		
128											1				
>128											1				
>1024												2			
<=0.03										2					
<=0.25														2	1
<=0.5									1						
<=1								2							
<=2			1												
<=8						2									

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Pigs - fattening pigs

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Positive/Present	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Positive/Present	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.06	0.25	0.06	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	128
	N of tested isolates	6	6	6	6	6	6	6	6	6	6
	N of resistant isolates	6	6	0	0	6	0	0	0	0	0
MIC											
1						1					
4					3						2
8					3	5					3
16		2									1
32		1									
>32		3									
64			1								
>64			5								
<=0.015								6			
<=0.03										6	
<=0.06				6							
<=0.12							6		6		

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Pigs - fattening pigs

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	116	116	116	116	116	116	116	116	116	116	116	116	116	116
MIC	N of resistant isolates	67	3	6	6	42	14	0	4	0	10	75	92	0	66
0.03							6								
0.06							1								
0.12							1								
0.25							6								
0.5							4								3
1					1				44						1
2		34							5						
4		10	15		1										
>4				6											
8			55		4		1				4		1		
>8							2								
16			13								1	9			
32						4			1			6	1		
>32									3						66
64		3	1			14					1	1	15		
>64		64	2										76		
128						9					4	1			
>128						15					5				
1024												2			
>1024												72			
<=0.015							95								
<=0.03										116					

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.25**	**0.5**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**64**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**116**	**116**	**116**	**116**	**116**	**116**	**116**	**116**	**116**	**116**	**116**	**116**	**116**	**116**
MIC	**N of resistant isolates**	**67**	**3**	**6**	**6**	**42**	**14**	**0**	**4**	**0**	**10**	**75**	**92**	**0**	**66**
<=0.25				110										116	46
<=0.5					110				63						
<=1		5						116							
<=2			30										23		
<=4											101				
<=8						74						25			

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Pigs - fattening pigs

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Positive/Present	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Positive/Present	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.06	0.25	0.06	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	128
	N of tested isolates	38	38	38	38	38	38	38	38	38	38
MIC	N of resistant isolates	38	38	0	0	38	0	0	0	0	0
0.03								1			
0.25							1				
1					1	7					
2		1			1	9					2
4		4			20	4					7
8		4			16	15					27
16		3				3					2
32		9	5								
>32		17									
64			8								
>64			25								
<=0.015								37			
<=0.03										38	
<=0.06				38							
<=0.12							37		38		

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Pigs - fattening pigs

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.25**	**0.5**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**64**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**
MIC	**N of resistant isolates**	**38**	**3**	**38**	**38**	**24**	**15**	**0**	**3**	**0**	**10**	**31**	**35**	**0**	**28**
0.03							2								
0.06							1								
0.25							9								
0.5							6							1	
1					7				17						
2					9				1						
4			4		3				1						
>4				38											
8			16		13						3				
>8					6										
16			6								1	4			
32						3			1			1	1		1
>32									1						27
64			1			5					2	2	11		
>64		38	2										23		
128						2					6				
>128						14					2				
>1024												31			
<=0.015							20								
<=0.03										38					
<=0.25														37	10
<=0.5									17						

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.25**	**0.5**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**64**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**	**38**
MIC	**N of resistant isolates**	**38**	**3**	**38**	**38**	**24**	**15**	**0**	**3**	**0**	**10**	**31**	**35**	**0**	**28**
<=1								38							
<=2			9										3		
<=4											24				
<=8						14									

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from pig - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Netherlands

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	Not Available	Not Available	Positive/Present	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available	Not Available
	Ceftazidime synergy test	Not Available	Not Available	Not Available	Not Available	Not Available	Positive/Present	Not Available	Not Available	Not Available	Not Available
	ECOFF	0.125	0.25	0.25	8	0.5	0.5	0.06	0.5	0.125	32
	Lowest limit	0.06	0.25	0.06	0.5	0.25	0.12	0.015	0.12	0.03	0.5
	Highest limit	32	64	64	64	128	128	2	16	16	128
	N of tested isolates	2	2	2	2	2	2	2	2	2	2
	N of resistant isolates	2	2	0	0	2	0	0	0	0	0
MIC											
2						1					
4					1	1					
8					1						1
16											1
32		1									
>32		1									
>64			2								
<=0.015								2			
<=0.03										2	
<=0.06				2							
<=0.12							2		2		

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from pig - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method: Dilution - sensititre

Country of Origin: Netherlands

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MIC	N of resistant isolates	2	0	2	2	1	2	0	0	0	1	2	1	0	1
0.5							1								
1							1								
2					1										
4					1										
>4				2											
8			2								1				
>32															1
64											1				
>64		2											1		
>128						1									
>1024												2			
<=0.03										2					
<=0.25														2	1
<=0.5									2						
<=1								2							
<=2													1		
<=8						1									

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from pig - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON pnl2

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Cefepime	Cefotaxim	Cefotaxime + Clavulanic acid	Cefoxitin	Ceftazidim	Ceftazidime + Clavulanic acid	Ertapenem	Imipenem	Meropenem	Temocillin
	Cefotaxime synergy test	**Not Available**	**Not Available**	**Positive/Present**	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Not Available**
	Ceftazidime synergy test	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Not Available**	**Positive/Present**	**Not Available**	**Not Available**	**Not Available**	**Not Available**
	ECOFF	**0.125**	**0.25**	**0.25**	**8**	**0.5**	**0.5**	**0.06**	**0.5**	**0.125**	**32**
	Lowest limit	**0.06**	**0.25**	**0.06**	**0.5**	**0.25**	**0.12**	**0.015**	**0.12**	**0.03**	**0.5**
	Highest limit	**32**	**64**	**64**	**64**	**128**	**128**	**2**	**16**	**16**	**128**
	N of tested isolates	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**
MIC	**N of resistant isolates**	**7**	**7**	**0**	**0**	**7**	**0**	**0**	**0**	**0**	**0**
2						2					
4					4	1					2
8		1			3	3					4
16						1					1
32		3									
>32		3									
64			1								
>64			6								
<=0.015								7			
<=0.03										7	
<=0.06				7							
<=0.12							7		7		

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic, unspecified in Meat from pig - fresh

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: ESBL MON

Analytical Method: Dilution - sensititre

Country of Origin: Greece

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	8	16	0.25	0.5	16	0.064	2	2	0.125	16	64	8	1	2
	Lowest limit	1	2	0.25	0.5	8	0.015	1	0.5	0.03	4	8	2	0.25	0.25
	Highest limit	64	64	4	8	128	8	16	32	16	128	1024	64	8	32
	N of tested isolates	7	7	7	7	7	7	7	7	7	7	7	7	7	7
MIC	N of resistant isolates	7	0	7	7	5	5	0	1	0	3	7	6	0	7
0.25							2								
0.5							1								
1									3						
2					2										
4			1		1										
>4				7											
8			4												
>8					4		2								
16			1												
32									1						
>32															7
64													2		
>64		7											4		
128						2									
>128						3					3				
1024												1			
>1024												6			
<=0.015							2								
<=0.03										7					
<=0.25														7	
<=0.5									3						
<=1								7							

	AM substance	Ampicillin	Azithromycin	Cefotaxim	Ceftazidim	Chloramphenicol	Ciprofloxacin	Colistin	Gentamicin	Meropenem	Nalidixic acid	Sulfamethoxazole	Tetracycline	Tigecycline	Trimethoprim
	ECOFF	**8**	**16**	**0.25**	**0.5**	**16**	**0.064**	**2**	**2**	**0.125**	**16**	**64**	**8**	**1**	**2**
	Lowest limit	**1**	**2**	**0.25**	**0.5**	**8**	**0.015**	**1**	**0.5**	**0.03**	**4**	**8**	**2**	**0.25**	**0.25**
	Highest limit	**64**	**64**	**4**	**8**	**128**	**8**	**16**	**32**	**16**	**128**	**1024**	**64**	**8**	**32**
	N of tested isolates	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**	**7**
MIC	**N of resistant isolates**	**7**	**0**	**7**	**7**	**5**	**5**	**0**	**1**	**0**	**3**	**7**	**6**	**0**	**7**
<=2			1										1		
<=4											4				
<=8						2									

OTHER ANTIMICROBIAL RESISTANCE TABLES

Specific monitoring of ESBL-/AmpC-/carbapenemase-producing bacteria and specific monitoring of carbapenemase-producing bacteria, in the absence of isolate detected

No data returned for this view. This might be because the applied filter excludes all data.